

Title	Rice ragged stunt Oryzavirus genome segment 9 encodes a 38 600 Mr structural protein
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Citation	Journal of General Virology, 76, 975-978
Issue Date	1995-05
Doc URL	http://hdl.handle.net/2115/6064
Туре	article
File Information	JGV76.pdf



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Rice ragged stunt *Oryzavirus* genome segment 9 encodes a 38600 $M_{\rm r}$ structural protein

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The complete nucleotide sequence of rice ragged stunt virus genome segment 9 (S9) was determined. The S9 segment is 1132 nucleotides long and has a long open reading frame starting from the first AUG codon at nucleotide position 14–16 and terminating at a UAG codon located at 1028–1030, which could encode a polypeptide with an M_r of 38600 (P9). The encoded polypeptide has no sequence homology to polypeptides of any other plant reoviruses published previously. An

Introduction

Rice ragged stunt virus (RRSV) is reported to occur in several south-east and east Asian countries and is an economically important disease of rice (Milne et al., 1982). The virus is transmitted by the planthopper Nilaparvata lugens and belongs to the new genus Oryzavirus of the family Reoviridae (Francki et al., 1991; Mayo & Martelli, 1993). The genome consists of 10 segmented double-stranded RNAs (dsRNAs) with estimated $M_{\rm r}$ values ranging from 0.78×10^6 – 2.58×10^6 (Kawano et al., 1984; Uyeda et al., 1990). Five major structural polypeptides were detected when the purified virus particles were subjected to SDS-PAGE (Hagiwara et al., 1990; Chen et al., 1989). Hagiwara et al. (1990) reported that a high concentration of $MgCl_2$ removed M_r 47000 and 37000 structural proteins, suggesting that these two proteins are components of the spike. Neither of these has been assigned to a particular genomic segment. Molecular characterization of the genome has not been reported except for analyses of the nucleotide sequences of the terminal portions (Yan et al., 1992). In this paper, we have determined the complete nucleotide

immunological study demonstrated that P9 was the smallest of the structural proteins. The P9 polypeptide was expressed as a fusion protein with maltose binding protein in *Escherichia coli*. Antisera to purified virions and to the fusion protein reacted with both the bacterially expressed polypeptide and the smallest polypeptide of the purified virus in immunoblotting analyses.

sequence of the genome segment 9 (S9) and demonstrated that it encodes an M_r 38000 structural protein (P9).

The viral transcripts were synthesized from purified virions (Lee et al., 1987 a) and a poly(A) tail was added by poly(A) polymerase and used for cDNA synthesis by the method of Gubler & Hoffman (1983) as previously described (Lee et al., 1987b). cDNA clones that reacted with the electrophoretically separated S9 probe in a dotblot hybridization (Lee et al., 1987b) were selected for nucleotide sequencing. A cDNA clone that covered a complete 3'-terminal portion was selected by a colony hybridization using a synthetic 17-mer oligonucleotide that was complementary to the 3' terminus as a probe. Four cDNA clones were sequenced by the dideoxy chain termination method of Sanger et al. (1977) and were found to cover the entire genomic sequence, except for the 5'-terminal 19 nucleotides. One clone had a sequence identical to that of the 3'-terminal oligonucleotides determined directly from the genome dsRNA followed by a poly(d/A) tail (Yan et al., 1992). More than 96% of the segment was sequenced in both directions. Since the cDNA clones did not cover the entire 5'-terminal region, its sequence was determined by reverse transcription of the viral transcript using the dideoxy termination method. A synthetic 16-mer that was complementary to nucleotide positions 49-64 from the 5' terminus was used as a primer (Yan et al., 1992). The entire nucleotide sequence of S9 is presented in Fig. 1. It is 1132 nucleotides long and has one long open reading frame (ORF)

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1	GATA	AAT	СТС	GCC	CATO M	GAA K	GA	CTG r	CC1 A	FTT F	GCC A	AGA R	AGA' D	rcc P	TTI F	TA	CAC T	CTO A	CCT(P	A A	T	GG. G	60
	TACO T	TAC Y	GGA G	ACC T	CATA I	ATA Y	TG	CCT A	CC/ S	AGA R	TCA S	TTA L	ACC P	CCG R	ACT I	СТ	CTA S	TC' I	ГСТ/ S	AAG1 K	rtc' F	ΓT L	120
	GGAA E	IGAI D	GCA	AAT N	rcc. P	AGA E	AA	TTI I	Y Y	GAA E	CTA L	TC S	GAG R	ATA Y	TG/ F	AAG E	CGC A	CTĆ(L	GGA. G	ACCA T	AAC. N	AG R	180
	ACCO	CAGO	CTCA	AGG(G	GAA K	GCG R	AG	CAA A	ATG M	CAA Q	CCA P	AGC A	CGT V	CAG S	TA/	AAC C	CAC P	GCT A	TTA L	CTG L	GAA E	AC T	240
	CGTA	TT(CACA	ACT	CGA	TAT	AT	GG1 W	FAC Y	AGA R	AG(R	GAC T	GAA N	CAA N		AGA Q	ACA N	ATC I	GGC G	AAT' N	TTA L	AG R	300
	AGA:	r DTTC	GGT	ATC	TCG	ATI	TC	TA	rct	GAI	GA'	rcg R	AGT V	AAC	GAG.	AA(E	GCG A	GTG V	ATG M	GTG V	CGA R	CT L	360
	D AGA'	S FTT	V AGA	CAT	т TGT	TGI	rcc	AG	CTA	AAA	IGA.	ATA	CTC	GTI	ſĠĂ	TA(T	GTT. V	AAA K	GAC D	AAA K	GAA E	.GC A	420
	D CCA	L GAC	D TTT	1 CGC	v GGA	.cco	GA1	Q TG	ь GCC	TTI	E GA	CTC	GCA	TCI	TAT	TT(GTC	AAT	CGC	GGC	GAA E	AA N	480
	Q CGC	T GAA	F TTA	A TGA	D TCT	TG:	R FAA	L	A CAG	F	U TTE	CAI	TCC	CGA	GTG	AT	GCT	TTI	CTI	'AAG	GAI	AA N	540
	A CTT	N TAA	Y GAC	D GGA	L AGC	AC	V FTA	T AAG	Q AAC	T STT(F GTT	1 GCI	AA(GTG	rcc	D AA	A AAC	r CAI	CACO	GGA	CTA	AGA	600
	F	K	T	E	AGO	TG.	L AT/	K AGT	K TCC	L FAA	L AGC	I GAC	, s	ACA.	V ACA	Q .TT	N GGC	H CTA	T AGGA	G	L	тст	660
	A	G	L	Q			D	S	S	K	A	ן דרי	r GGI	Y I	N TAC	I CAA	G CAA		G GGC	Q TTT1	Y rgc(L GGA	720
	CGA E	GGA D	TGA E	A A	I		L.	Y	R	Q	G	1		A T	L	Q	Q	M	A CGA	F TCT4	A AAG	E CAA	700
	GTT L	GGA E	ATT L	AGC		GAG २	GA) G	GCA A	IGA0 E	JAA. K	AGA E		A .	F	P	S	T	F	D	L	S	N	840
	TAC R	ACC P	GGC A	TTC C	GCA C 1	ACT V	TG. L	ATC I	L CT L	ГАА К	GCC F		CGT F	GTA C	AG1 K	W	Y	Q	Q	T	F	K	0.00
	GGA I	TGA E	GGA E	GAG I	GAA R I	AGG K	AA E	TTI F	GC A	GAA K	GAC	GTT' G	ГGТ L	GGG W	TG(V	GAT D	GAC D	CTT' F	rgc A	CGA E	A GGC	N	900
	TTC V	GAA I N	ACAC I I	CAG(GTA. G 1	ATT N	TG L	TCI S	TTT F	TGG G	CTT I	TTT ?	CAA S	CCA T	CA' T	TTA L	AA' N	rgt. V	AAT I	TGA E	GAG R	ATG W	960
	GC(H	TTT 1	rga(GAT R	TTI F	Y Y	GTI V	ГСА Н	TAT M	GT/	ACT Y	CAT S	CTC S	TG V	CA3 H	TAT. I	ATA Y	TTC S	TCA Q	GAA K	GGC A	1020
	CTO	CGG(S (GGT/ G ?	AGG. *	AAA	CGC	CTG	GGG	GAT	CAA	GG	rga	ATI	`GAA	AT	GAC	GGA	СТТ	СТС	ACC	CTC	ATG	1080
	CA	<u>ጉ ለ ጥ</u> ′	rce	GCC	тст	TCA	ACG	¦GG'	TGG	CAC	CCC	CGA	AAA	CGC	GCG	AG	ACC	ATG	GTG	С			1132

Fig. 1. Nucleotide sequence of cDNA to rice ragged stunt virus genome segment 9 (upper) and amino acid sequence (lower) of the encoded polypeptide.

encoding 338 amino acids (P9). The predicted polypeptide has no significant sequence homologies to any of those polypeptides encoded by plant reoviruses sequenced so far (Uyeda *et al.*, 1994; Nuss & Dale, 1990).

In order to assign a polypeptide encoded by S9, the longest ORF was expressed in E. coli as a fusion with the maltose binding protein as described in Masuta *et al.* (1992). A cDNA clone was made by reverse transcription

coupled with PCR of genomic dsRNAs (Murao *et al.*, 1994) using a set of primers complementary to the 5' translational start site and the 3'-terminal 17 nucleotides. *Eco*RI and *Pst*I recognition sites were added to their 5' termini for subsequent cloning into bacterial plasmid pUC119. The longest ORF was then cloned in the *Eco*RI and *Pst*I sites of pMAL C2 (New England Biolabs). The *E. coli* strain JM83 was transformed with the plasmid and



Fig. 2. Identification of P9 as the smallest structural protein. The polypeptide P9 was expressed as a fusion protein with maltose binding protein in *E. coli*, subjected to electrophoresis in a 12.5% polyacrylamide gel (Laemmli, 1970) and analysed by immunoblotting. Lane 1, fusion protein purified on an amylose resin column to which the maltose binding protein binds; lane 2, fusion protein digested with proteinase factor Xa; lane 3, purified virions. The blotted proteins were probed with antisera raised against purified virions (*a*) and the fusion protein (*b*) and detected with a Konica Immunostaining HRD kit IS-50B. The large and small arrows at the right indicate the positions of the smallest and other structural proteins, respectively. Arrows at left indicate position of size markers.

the fusion protein was expressed as described by Masuta et al. (1992). The bacterial cells expressing the fusion protein produced a band with an M_r of 81000 in SDS-PAGE (Laemmli, 1970). Cleavage of the fusion protein by the proteinase factor Xa yielded a band with an apparent M_r of 36200, which is close to the M_r (38600) of P9 expected from the nucleotide sequence. This band reacted in immunoblotting analyses with antiserum raised against purified virions, suggesting that P9 was the smallest structural protein (Fig. 2a), previously reported as the M_r 37000 structural protein by Hagiwara et al. (1990). The mobility of the cleavage product was slightly slower than the smallest structural protein, probably because four amino acid residues derived from the plasmid vector sequence had been added to the N terminus of the protein. Antisera were raised against the fusion protein in rabbits. When the purified virus was electrophoresed, blotted to an Immobilon-P membrane (Millipore), and tested with the antiserum, only the smallest structural protein specifically reacted (Fig. 2b). These experiments prove that P9 is the smallest structural protein with an M_r of 38600.

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This study was supported in part by Grant-in-Aid for Scientific Research no. 62480042 and 01560045, and Grant-in-Aid for Cooperative Research no. 6304013 from The Ministry of Education, Science and Culture, and a grant from Suhara Foundation. We thank Dr C. Masuta, Japan Tobacco Inc., for suggestions on handling pMAL expression vectors.

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(Received 25 July 1994; Accepted 16 November 1994)