



Title	Detection of the meq gene in the T cell subsets from chickens infected with Marek's disease virus serotype 1
Author(s)	Chang, Kyung-Soo; Ohashi, Kazuhiko; Lee, Sung-II; Takagi, Michihiro; Onuma, Misao
Citation	Japanese Journal of Veterinary Research, 53(1-2), 3-11
Issue Date	2005-08
DOI	10.14943/jjvr.53.1-2.3
Doc URL	http://hdl.handle.net/2115/879
Type	bulletin (article)
File Information	3-11.pdf



[Instructions for use](#)

Detection of the *meq* gene in the T cell subsets from chickens infected with Marek's disease virus serotype 1

Kyung-Soo Chang¹⁾, Kazuhiko Ohashi¹⁾, Sung-Il Lee¹⁾, Michihiro Takagi²⁾
and Misao Onuma¹⁾

(Accepted for publication : June 15, 2005)

Abstract

The *meq* gene was thought to be only detected in Marek's disease virus serotype 1 (MDV 1) including a very virulent strain, Md5, while L-*meq*, in which a 180-bp sequence is inserted into the *meq* open reading frame, is found in other strains of MDV 1, such as CVI 988/R6. However, both *meq* and L-*meq* were previously detected by PCR in chickens infected with MDV 1, suggesting that MDV 1 may consist of at least two subpopulations, one with *meq*, the other with L-*meq*. To further analyze these subpopulations, we analyzed the time course changes in distribution of these subpopulations among T cell subsets from chickens infected with MDV1. Both *meq* and L-*meq* were detected in CD4⁺ and CD8⁺ T cells infected with strain Md5 or CVI 988/R6. The shift in MDV subpopulations from one displaying *meq* to the other displaying L-*meq* and/or the conversion from *meq* to L-*meq* occurred mainly in the CD8⁺ T cell subset from Md5-infected chickens. PCR products corresponding to L-*meq* rather than *meq* were frequently amplified from the CD8⁺ T cell subset from CVI 988/R6-infected chickens. These results suggest that a dominant subpopulation of MDV 1 changes depending on the T cell subsets, and that L-*meq* is dominantly present in the CD8⁺ T cells which play a role in the clearance of pathogenic agents.

KEY WORDS : Marek's disease virus, *meq*, L-*meq*, CD4⁺ T cell, CD8⁺ T cell

¹⁾Department of Disease Control, Graduate School of Veterinary Medicine, Hokkaido University, Sapporo 060-0818, Japan

²⁾Department of Microbiology and Immunology, Faculty of Agriculture, Kobe University, Kobe 657-8501, Japan

Corresponding author : Kazuhiko Ohashi, Department of Disease Control, Graduate School of Veterinary Medicine, Hokkaido University, Sapporo 060-0818, Japan at address a.

Tel. : 011-706-5274

Fax. : 011-706-5217

E-mail address : okazu@vetmed.hokudai.ac.jp

Introduction

Marek's disease virus (MDV) is the causative agent of Marek's disease (MD), which is characterized by CD4⁺ T cell lymphoma and nerve enlargement caused by infiltration of lymphocytes and/or lymphoma cells^{1,6}. The pathogenesis of MD can be chronologically divided into three phases: early cytolitic infection; latent infection; and secondary cytolitic infection with immunosuppression and tumor development^{1,14,15}. Early cytolitic infection occurs mainly in B cells, and latently infected lymphocytes are mainly activated T cells¹⁹. In addition, target cells for transformation by MDV are mainly CD4⁺ T cells, suggesting that latent infection in this T cell subset could

be intimately related to the subsequent transformation by MDV¹⁸.

MDV strains are classified into three serotypes based on their pathogenicity, and only strains of MDV serotype 1 (MDV 1) are oncogenic. An MDV 1-specific gene, *meq*, has been identified, and its gene product, Meq, a basic leucine zipper (bZIP) oncoprotein, has been detected in all tumor samples and established MDV-transformed T-cell lines examined thus far, suggesting that Meq protein plays an important role in the transformation by MDV^{9,10,11,16}. However, the role of the Meq protein in MDV transformation remains to be determined. In the genome of strains of vvMDV 1, the *meq* open reading frame (ORF) encoding a 339-amino-acid bZIP protein, is present,

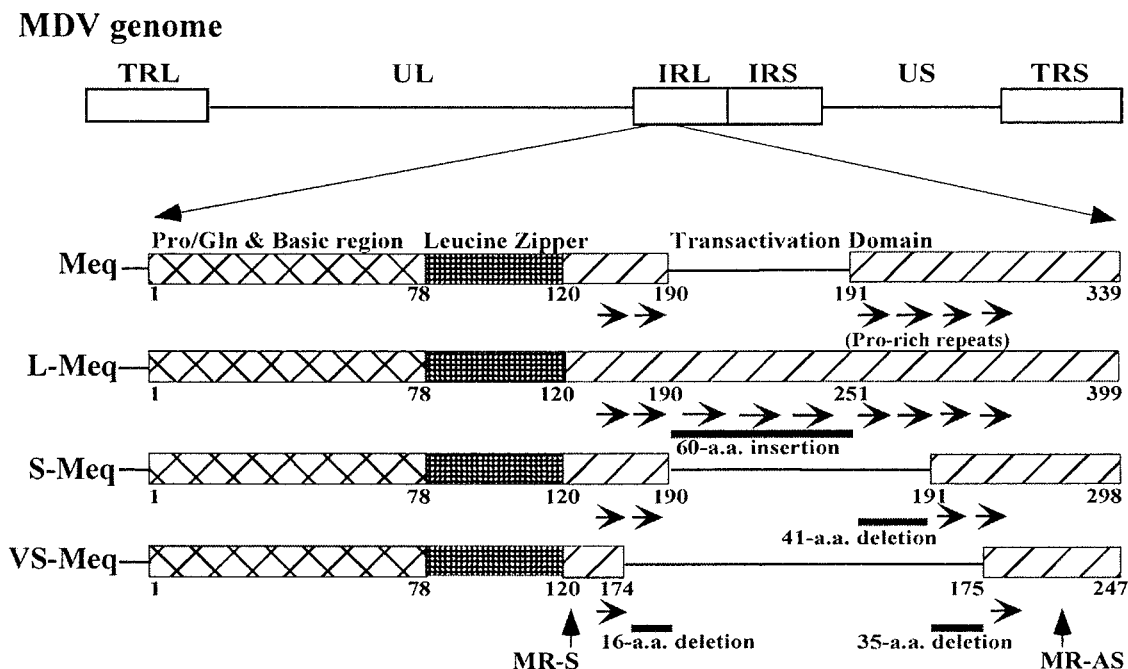


Fig. 1. Structures of the *meq* ORF in MDV1. The *meq* ORF of oncogenic MDV 1, Md5, encodes a 339 amino-acid protein consisting of an N-terminal proline-glutamine-rich region (Pro/Gln), the basic region, and the leucine zipper, as well as the transactivation domain. In the case of L-*meq* found in the MDV tumor cell lines, MSB 1 and MTB 1, and JM attenuated by serial passage, 60 amino acids (a 180-bp sequence) are inserted into the transactivation domain. In the case of S-Meq and VS-Meq, 92 amino acids (a 276-bp sequence) and 41 (a 123-bp sequence) are deleted in the transactivation domain of the Meq protein, respectively. The proline-rich repeat region is composed of 6 copies of the repeats in Meq, 9 copies in L-Meq, 4 copies in S-Meq, and 2 copies in VS-Meq.

while a slightly longer *meq* ORF, termed L-*meq*, in which a 180-bp sequence is inserted into the *meq* ORF, is found in other strains of MDV 1, such as JM and CVI988/R6. Furthermore, short *meq* (S-*meq*) and very short *meq* (VS-*meq*), in which 123 and 275-bp sequences are deleted in the *meq* ORF, have been also detected, respectively. This diversity in *meq* is due to the difference in the copy number of the proline-rich repeat region (Fig. 1)^{5,7}. When the *meq* gene was monitored in chickens infected with vvMDV 1, both the *meq* and L-*meq* genes were detected, suggesting that MDV 1 consists of some subpopulations though the possibility of conversion from *meq* to L-*meq* can not be ruled out³. Since an MDV subpopulation displaying the L-*meq* gene is detected at the latent phase, the L-*meq* and its gene product, if any, might contribute to the maintenance of the MDV latency³. Though the shift in subpopulations and/or conversion of the *meq* genes have been observed in chickens infected with MDV 1, these changes in the T cell subsets have not been elucidated. Thus, in this study, to characterize the shift in subpopulations and/or conversion of the *meq* genes in T cell subsets, especially CD4⁺ T cells, targets for transformation by MDV 1, we have analyzed the time course changes in the *meq* genes detected in T cell subsets from chickens infected with MDV 1. In addition, we have also compared this shifts/conversion in the T cell subsets from both MDV-sensitive and -resistant lines of SPF chickens.

Materials and methods

Chickens

Fertilized eggs of commercial chickens were purchased from Hokuren Co. Ltd. (Sapporo, Japan) and two lines of SPF chickens (MHC B¹⁹ B¹⁹ sensitive to MD: B19 and B²¹ B²¹ resistant to MD: B21)¹³ were obtained from

Nisseiken Co. Ltd. (Oume, Japan). These eggs were hatched in our laboratory, and chickens were raised in isolators.

Viruses and cells

A strains of very virulent MDV 1, Md5,²⁰ was obtained from chicken kidney cell culture from experimentally infected chickens. A vaccine strain of MDV 1, CVI 988/R6, was purchased from Geln Corp. Inc. (Gifu, Japan). These viruses were propagated in chicken embryo fibroblasts (CEF). When cytopathic effects were confluent, the infected cells were harvested, and virus titers were measured by plaque assay. These infected CEFs were used for viral inoculation of chickens.

Inoculation of MDVs and sample collection from infected chickens

Groups of 3-day-old chickens were inoculated intraperitoneally with 2,000 plaque forming unit (PFU) /0.2ml of Md5, or 10,000 PFU/0.2ml of CVI 988/R6. We used B19 and B21 chickens for the inoculation of Md5 or CVI988/R6. The spleens were collected from 4 chickens from each group infected with MDV every week pi. for 10 weeks.

Isolation of CD4⁺ and CD8⁺ T cells

Peripheral blood mononuclear cells (PBMC) were obtained by homogenizing the spleens between two frosted glass slides and subsequent centrifugation over Ficoll-Conray gradients to remove dead cells and red blood cells. PBMC (5×10^7) were incubated with 1 ml of monoclonal antibodies (mAbs), anti-chicken CD4⁺ mAb (CT4⁺)²¹ or anti-chicken CD8⁺ mAb (11-39)¹² (culture supernatant), for 30min at 4 °C. After incubation, cells were washed twice with DMEM/F-12 (GIBCO BRL, Grand Island, NY) containing 10% heat-inactivated fetal calf serum (FCS: Filtron, Brooklyn, Australia), 5×10^5

M 2-mercaptoethanol (2-ME), 100 IU/ml penicillin, and 100 µg/ml streptomycin (complete DMEM/F-12). These cells were then resuspended in 500 µl of complete DMEM/F-12 containing 2×10^7 of sheep anti-mouse IgG-magnetic beads (Dynabeads M-450, Dynal Inc., Oslo, Norway), and incubated at 4 °C for 45 min with gentle rotation. After incubation, cells attached to the beads were trapped by a magnetic field. To separate the cells from the beads, the cells binding to the beads were cultured in RPMI 1640 (GIBCO BRL, Grand Island, NY) containing 10% heat-inactivated FCS, 5×10^{-5} M 2-ME, 100 IU/ml penicillin, and 100 µg/ml streptomycin, at 40 °C for 3.5 hours in a humidified atmosphere containing 5% CO₂. Flowcytometric analysis revealed that the purity of each T cell subset separated by this procedure ranged from 92 to 98%. The isolated CD4⁺ and CD8⁺ T cells were used for the extraction of total cellular DNA.

Extraction of DNA from T cell subsets

Total cellular DNA was extracted from CD4⁺ and CD8⁺ T cell subsets using SepaGene (Sankoujunyaku Co., Japan). The DNA was treated with RNase and re-extracted with phenol-chloroform-isoamyl alcohol (25 : 24 : 1), precipitated with ethanol, and dissolved in distilled water.

Nested polymerase chain reaction (PCR)

Total cellular DNA samples extracted from each of T cell subsets of infected chickens were used as a template for nested PCR amplification to detect the *meq* gene. The first round of PCR was performed with a primer set, M-S and M-AS, to amplify the 1.0-kb *meq* or 1.2-kb L-*meq* gene fragments as described by Lee *et al.*⁸⁾. After the amplification, 1 µl of the reaction was used for the second round of PCR. The second round of PCR was performed in the 20 µl reaction mixture contain-

ing 1.5 mM MgCl₂, and a primer set, 10 pM of MR-S (5'-AGTTGGCTTGTCATGAGCCAG-3') and MR-AS (5'-TGTTCCGGGATCCTCGGTAA GA-3'), to amplify a 0.6-kb *meq* or 0.8-kb L-*meq* fragment. Amplification was carried out over 35 cycles of 94 °C for 45 sec, 55 °C for 45 sec, and 72 °C for 1.5 min. The amplified fragments were separated on an agarose gel (1.5%) and visualized by staining with ethidium bromide.

Reverse transcriptase-polymerase chain reaction (RT-PCR)

Total cellular RNAs were extracted from CD4⁺ and CD8⁺ T cell subsets using the Trizol reagent (Invitrogen, CA). Lymphocytes separated by immuno-magnetic beads were resuspended in 1 ml of Trizol reagent, and 200 µl of chloroform was added. After centrifugation at 12,000 × g for 10 min at 4 °C, 500 µl of isopropanol was added to the resultant supernatants. Total cellular RNA was precipitated by centrifugation at 12,000 µg for 10 min at 4 °C after incubation at room temperature for 10 min. The resultant RNA was dissolved in diethylpyrocarbonate-treated water, and used as template to synthesize cDNA by using RAV 2 reverse transcriptase (Takara, Japan). The cDNA samples were used to amplify the *meq* transcripts by PCR using the M-S and M-AS primer set and MR-S and MR-AS primer set as described above.

Results

Detection of the meq gene in the T cell subsets from chickens infected with vvMDV 1, Md5

When the *meq* gene was monitored in the T cell subsets from chickens infected with Md 5, the *meq* gene was detected in CD4⁺ and CD8⁺ T cells from the infected chickens throughout the 10 weeks though the intensities of the bands corresponding to the *meq* gene detected were variable (Figs. 2-4). The

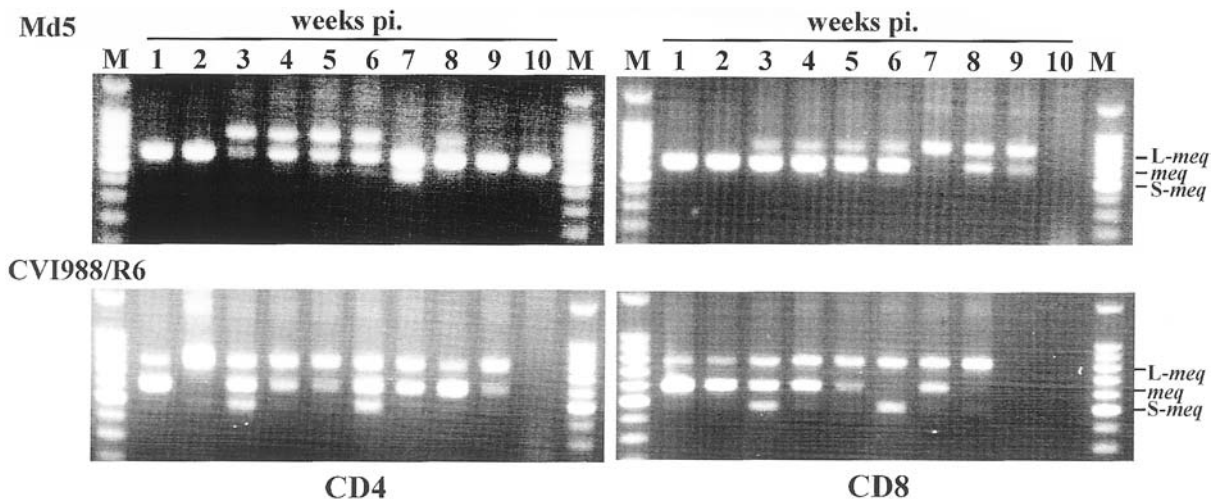


Fig. 2 . Nested PCR amplification of the *meq* genes from T cell subsets of chickens infected with 2,000 PFU/ chicken of strain Md5, or 10,000 PFU/chicken of strain CVI988/R6. DNA samples were collected at every week for 10 weeks pi. M : molecular size marker (100-bp DNA ladder).

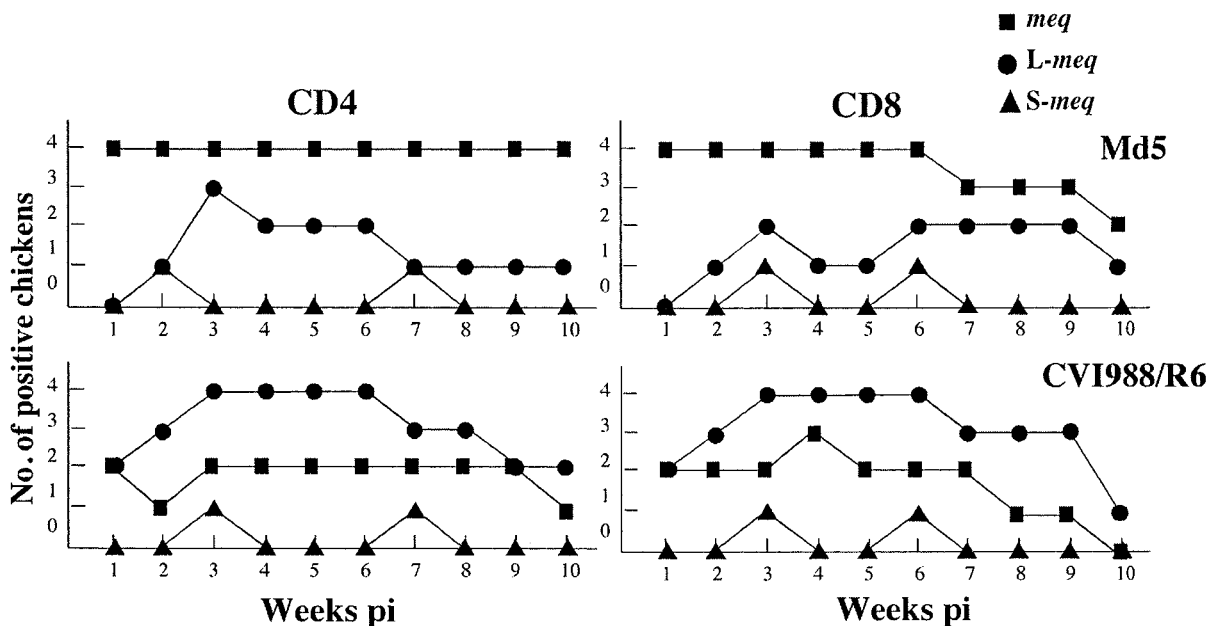


Fig. 3 . Detection of the *meq* genes in the T cell subsets from the B19 chickens infected with MDV 1 . DNA samples were collected from 4 chickens/group at each week pi.

L-meq gene was also detected in both the CD4⁺ and CD8⁺ T cells after 3 weeks pi. Interestingly, the frequency of the chickens from which the *L-meq* gene was transiently increased in the CD4⁺ T cell from Md5-infected chickens at 3 to 6 weeks pi, presumably the latent phase of the MDV infection, and then re-

duced after 7 weeks pi (Figs. 3- 4). However, this frequency was lower in the case of CD8⁺ T cells at 3 to 6 weeks pi, while higher after 7 weeks pi. In addition, 0.5kb-short *meq* (*S-meq*) was occasionally detected at 2 to 3 and 6 to 7 weeks pi. These changes were observed both in the B19 and B21 chickens, but the de-

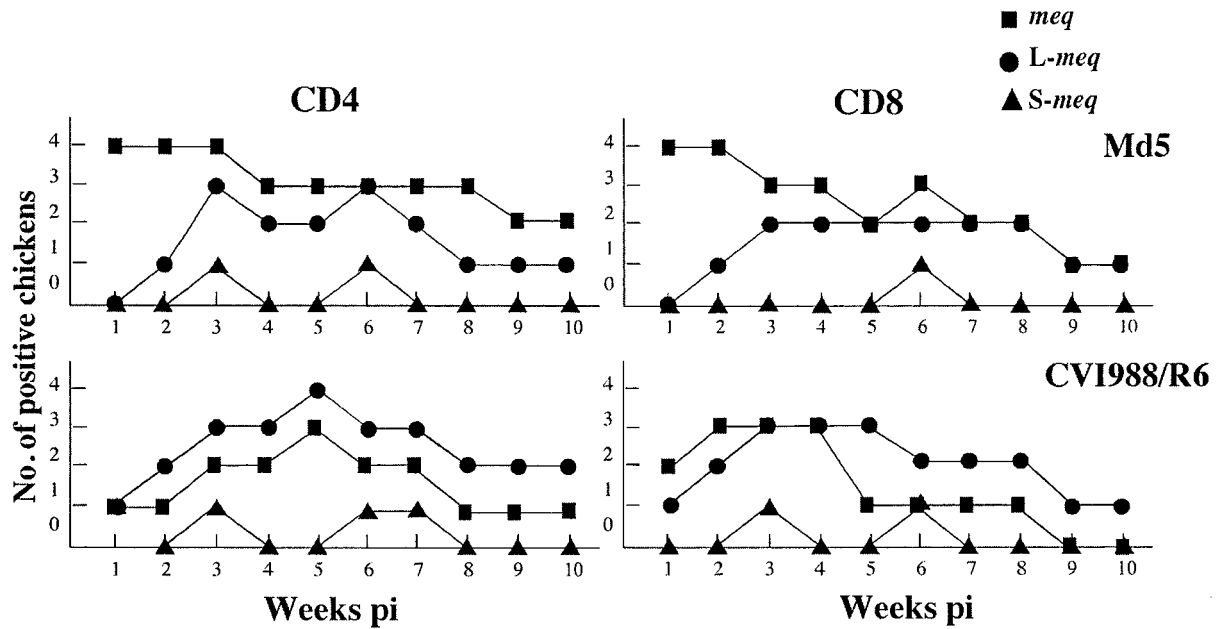


Fig. 4 . Detection of the *meq* genes in the T cell subsets from the B21 chickens infected with MDV1. DNA samples were collected from 4 chickens/group at each week pi.

tection frequency of the *meq* genes was decreased faster in MD-resistant B21 chickens than in MD-sensitive B19 chickens (Figs. 3-4).

Detection of the *meq* gene in the T cell subsets from chickens infected with CVI 988/R 6

When the *meq* genes were monitored in chickens infected with CVI 988/R 6 in which the *L-meq* gene was already identified (11, 18), the *L-meq* gene was consistently detected in both $CD4^+$ and $CD8^+$ T cells from the infected chickens throughout the experimental period. The *meq* gene was also detected in the $CD4^+$ and $CD8^+$ T cells, though the frequency of the chickens from which the *meq* gene was detected was lower than that from which the *L-meq* gene was detected (Figs. 2-4). These frequencies were gradually decreased after 7 weeks pi. In addition, 0.5kb-*S-meq* was detected temporarily at 2 to 3 or 6 to 8 weeks pi. These changes were observed both in B19 and in B21 chickens. The detection frequency of the *meq* genes was lower in B21 than B19

chickens throughout the experimental period (Figs. 3-4).

Expression of the *meq* genes in the T cell subsets from chickens infected with MDV 1

To determine whether the *meq* genes present in T cell subsets were transcribed, RT-PCR analysis was performed. As shown in Fig. 5, when RNA samples were analyzed from infected chickens at 6 weeks pi, the *meq* gene was abundantly transcribed in the $CD4^+$ T cells from Md5-infected chickens from which both the *meq* and *L-meq* genes were detected. On the other hand, both the *meq* and *L-meq* genes were transcribed in the $CD8^+$ T cells from these chickens. Only the *meq* transcript was detected in $CD4^+$ and $CD8^+$ T cells from chickens from which the *meq* gene was detected (data not shown).

In the case of CVI 988/R 6-infected chickens, the *L-meq* gene was transcribed in both the $CD4^+$ and $CD8^+$ T cells though the *meq* transcript was also detected in the $CD4^+$ T cells. Only the *L-meq* transcript was detected

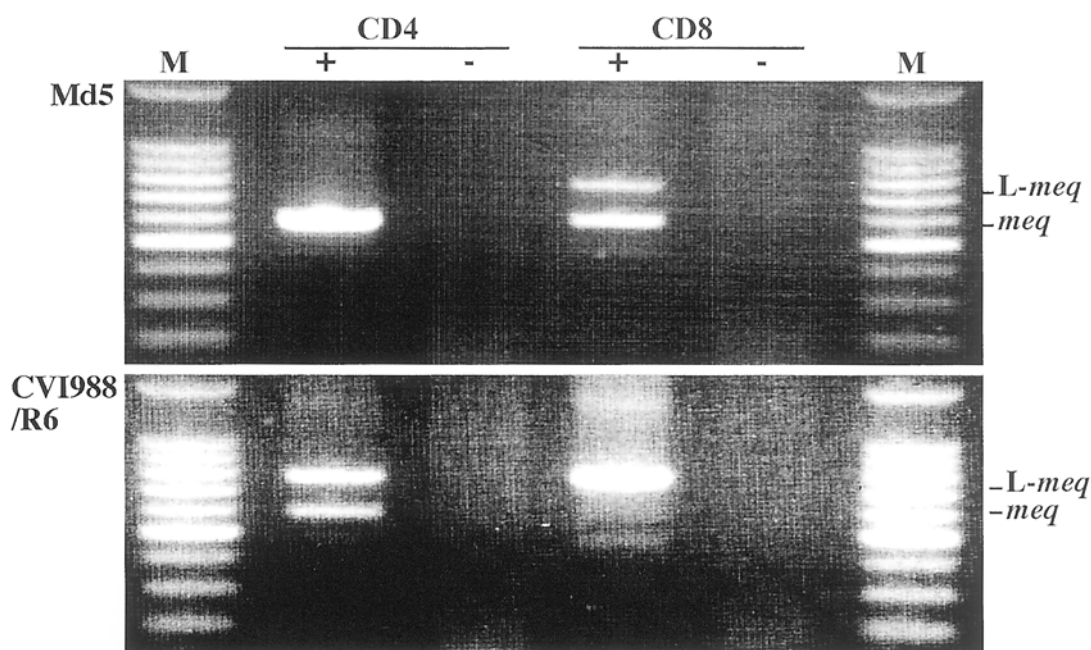


Fig. 5. RT-PCR detection of the *meq* transcripts from T cell subsets of chickens infected with 2,000 PFU/chicken of strain Md5 or 10,000 PFU/chicken of strain CVI988/R6. RNA samples, collected from chickens from which both the *meq* and *L-meq* genes were detected at 6 weeks pi., were used for RT-PCR (lane +), or directly used for PCR to examine the contamination of genomic DNA (lane -). M: molecular size marker (100-bp DNA ladder, Promega).

in $CD4^+$ and $CD8^+$ T cells from chickens from which the *L-meq* gene was detected (data not shown).

Discussion

There is evidence that the *meq* gene and its gene product, Meq, play important roles in transformation of $CD4^+$ T cells by MDV¹⁹. The Meq protein is consistently expressed in the vast majority of MD-transformed cell lines and $CD4^+$ T cells obtained from lymphomas¹⁷. MDV 1 mutants lacking a functional *meq* failed to cause tumors in chickens¹⁶. Polymorphism of the *meq* genes in the attenuated MDV 1, MDV-tumor cell lines and chickens infected with MDV 1 has been observed⁵. This polymorphism suggested that MDV 1 consists of at least 3 subpopulations, virus displaying the *meq*, *L-meq* and *S-meq* genes, respectively. During the MDV 1 infection, a dynamic shift

from one subpopulation of MDV 1 to another may occur due to some selective pressures by host chickens. In addition, the *L-meq* gene product suppressed the transactivation activity of the Meq protein of oncogenic MDV 1, and replication of vvMDV 1 carrying the *meq* gene was partially suppressed in cells transfected with the *L-meq* gene⁴.

In this study, we monitored the *meq* gene in the T cell subsets from chickens infected with strains of oncogenic and nononcogenic MDV 1 for 10 weeks pi. Surprisingly, both the *meq* and *L-meq* genes were detected in the T cell subsets from infected chickens during experimental periods. In addition, *S-meq* has also been detected temporarily though biological significance of the *S-meq* gene and its gene product remains unknown. It is of interest that, in chickens infected with vvMDV 1, Md5, the *L-meq* gene was detected during the

latent phase (3 - 6 weeks pi.) in CD4⁺ T cells which are the target cells for latent infection and subsequent transformation by oncogenic MDV 1, though the *meq* gene was consistently detected throughout the experimental period. In the case of chickens infected with CVI 988/R6, the L-*meq* but not *meq* gene was consistently detected in both the CD4⁺ and CD8⁺ T cells. The detection frequency of the *meq* genes in the chickens was decreased more rapidly in the CD8⁺ than in the CD4⁺ T cells. These results suggest that a major subpopulation displaying the *meq* gene in the CD4⁺ T cells would be important for the transformation by Md5, and that the presence of a subpopulation displaying the L-*meq* gene may contribute to the maintenance of MDV latency. Furthermore, the presence of the major subpopulation with the L-*meq* gene in the CD8⁺ T cells, as detected in CVI988/R 6 (vaccine) -inoculated chickens, suggest that the L-*meq* gene product recognized by CD8⁺ T cells could play a role in the cell-mediated immunity against MDV infection. It should be noted that the L-*meq* rather than *meq* gene is dominantly detected in MD tumor cell lines⁵⁾.

The shift in subpopulations and/or conversion of the *meq* genes mentioned above was observed in both the B19 chickens sensitive to MD, and the B21 chickens resistant to MD. Interestingly, detection frequency of the *meq* gene was decreased faster in both the CD4⁺ and CD8⁺ T cells from B21 chickens than from B19 chickens. The reason for this observation remains unknown, but these differences observed between two lines of chickens may be intimately related to the resistance to tumor formation by MDV 1.

Thus, MDV 1 could potentially express not only the *meq* but also L-*meq* gene in each of the T cell subsets from the infected chickens dependent upon the phase of MDV 1 infection. In the future, it will be necessary to ana-

lyze the roles of the L-*meq* and S-*meq* gene and/or their gene products in viral transcription and transformation and immune responses to MDV infection.

References

- 1) Calnek, B. W. 1986. Marek's disease-a model for herpesvirus oncology. *Crit. Rev. Microbiol.*, 12 : 293-320.
- 2) Chan, M.M., Chen, C.L.H., Ager, L.L. and Cooper, M.D. 1988. Identification of the avian homologues of mammalian CD4 and CD8 antigens. *J. Immunol.*, 140 : 2133-2138.
- 3) Chang, K.S., Lee S.I., Ohashi, K., Ibrahim A. and Onuma, M. 2002. The detection of the *meq* gene in chicken infected with Marek's disease virus serotype 1. *J. Vet. Med. Sci.*, 64 : 413-417.
- 4) Chang, K.S., Lee, S.I., Ohashi, K. and Onuma, M. 2002. Suppression of transcription activity of the MEQ protein of oncogenic Marek's disease virus serotype 1 (MDV 1) by L-MEQ of non-oncogenic MDV1. *J. Vet. Med. Sci.*, 64 : 1091-1095.
- 5) Chang, K.S., Ohashi, K. and Onuma, M. 2002. Diversity (Polymorphism) of the *meq* gene in the attenuated Marek's disease virus (MDV) serotype 1 and MDV-transformed cell lines. *J. Vet. Med. Sci.*, 64 : 1097-1101.
- 6) Lee, S.I., Ohashi, K., Morrimura, T., Sugimoto, C. and Onuma, M. 1999. Re-isolation of Marek's disease virus from T cell subsets of vaccinated and non-vaccinated chickens. *Arch. Virol.*, 144 : 45-54.
- 7) Lee, S.I., Ohashi, K., Takagi, M., Chang, K. S., Sugimoto, C. and Onuma, M. 2001. Insertion of a 178-bp sequence into the *meq* gene of Marek's disease virus serotype 1. *In* : Schat K.A., Morgan R.M., Parcells M. S., Spencer J.L. (eds.) Current research

- on Marek's disease. American Association of Avian Pathologists, Kennett Square, pp 289-294.
- 8) Lee, S.I., Takagi, M., Ohashi, K., Sugimoto, C. and Onuma, M. 2000. Difference in the *meq* gene between oncogenic and attenuated strains of Marek's disease virus serotype 1. *J. Vet. Med. Sci.*, 62 : 287-292.
 - 9) Liu, J.L. and Kung, H.J. 2000. Marek's disease herpesvirus transforming protein MEQ : a c-Jun analogue with an alternative life style. *Virus Genes*, 21 : 51-64.
 - 10) Liu, J.L., Lin, S.F., Xia, L., Brunovskis, P., Li, D., Davidson, I., Lee, L. - F. and Kung, H. - J. 1999. MEQ and v-IL 8 : cellular genes in disguise? *Acta. Virol.*, 43 : 94-101.
 - 11) Liu, J.L., Ye, Y., Lee, L.F. and Kung, H. J. 1998. Transforming potential of the herpesvirus oncoprotein *MEQ* : morphological transformation, serum-independent growth, and inhibition of apoptosis. *J. Virol.*, 72 : 388-395.
 - 12) Luhtala, M., Salmonsén, J., Hirota, Y., Onodera, T., Toivanen, P. and Vainio O. 1993. Analysis of chicken CD4 by monoclonal antibodies indicates evolutionary conservation between avian and mammalian species. *Hybridoma*, 12 : 633-646.
 - 13) Mizutani, M. 1995. *Okayama Lab. Anim. Res. Group Info.*, 12 (suppl.) : 26-29 (in Japanese).
 - 14) Morimura, T., Hattori, M., Ohashi, K., Sugimoto, C. and Onuma, M. 1995. Immunomodulation of peripheral T cells in chickens infected with Marek's disease virus : involvement in immunosuppression. *J. Gen. Virol.*, 76 : 2979-2985.
 - 15) Morimura, T., Ohashi, K., Kon, Y., Hattori M., Sugimoto, C. and Onuma, M. 1996. Apoptosis and CD8 - down-regulation in the thymus of chickens infected with Marek's disease virus. *Arch. Virol.*, 141 : 2243-2249.
 - 16) Ross, N.L.J. 1999. T-cell transformation by Marek's disease virus. *Trends Microbiol.*, 7 : 22-29.
 - 17) Ross, N., O'Sullivan, G., Rothwell, C., Smith, G., Burgess, S.C., Rennie, M., Lee, L.F. and Davison, T.F. 1997. Marek's disease virus *EcoRI-Q* gene (*meq*) and a small RNA antisense to *ICP4* are abundantly expressed in CD4⁺ cells and cells carrying a novel lymphoid marker, AV37, in Marek's disease lymphomas. *J. Gen. Virol.*, 78 : 2191-2198.
 - 18) Schat, K.A., Chen, C.-L.H., Calnek, B.W. and Char, D. 1991. Transformation of T-lymphocyte subsets by Marek's disease herpesvirus. *J. Virol.*, 65 : 1408-1413.
 - 19) Shek, W.R., Calnek, B.W., Schat, K.A. and Chen, C.-L.H. 1983. Characterization of Marek's disease virus-infected lymphocytes : discrimination between cytolytically and latently infected cells. *J. Natl. Cancer Inst.*, 70 : 485-491.
 - 20) Witter, R.L., Sharma, J.M. and Fadly, A. M. 1980. Pathogenicity of variant Marek's disease virus isolates in vaccinated and unvaccinated chickens. *Avian Dis.*, 24 : 210-232.