

2 **Effect of vibration frequency on agonist and antagonist arm**  
3 **muscle activity**

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9 **Abstract**

10 *Purpose* This study aimed to assess the effect of vibration  
11 frequency ( $f_{out}$ ) on the electromyographic (EMG) activity  
12 of the biceps brachii (BB) and triceps brachii (TB) muscles  
13 when acting as agonist and antagonist during static exer-  
14 cises with different loads.  
15 *Methods* Fourteen healthy men were asked to hold a  
16 vibratory bar as steadily as possible for 10 s during lying  
17 row (pulling) and bench press (pushing) exercise at  $f_{out}$  of  
18 0 (non-vibration condition), 18, 31 and 42 Hz with loads of  
19 20, 50, and 80 % of the maximum sustainable load (MSL).  
20 The root mean square of the EMG activity ( $EMG_{RMS}$ ) of  
21 the BB and TB muscles was expressed as a function of the  
22 maximal  $EMG_{RMS}$  for respective muscles to characterize  
23 agonist activation and antagonist coactivation.  
24 *Results* We found that (1) agonist activation was greater  
25 during vibration (42 Hz) compared to non-vibration exer-  
26 cise for the TB but not for the BB muscle ( $p < 0.05$ );  
27 (2) antagonist activation was greater during vibration

compared to non-vibration exercise for both BB ( $p < 0.01$ ) 28  
and TB ( $p < 0.05$ ) muscles; (3) the vibration-induced 29  
increase in antagonist coactivation was proportional to 30  
vibration  $f_{out}$  in the range 18–42 Hz and (4) the vibration- 31  
induced increase in TB agonist activation and antagonist 32  
coactivation occurred at all loading conditions in the range 33  
20–80 % MSL. 34

*Conclusion* The use of high vibration frequencies within 35  
the range of 18–42 Hz can maximize TB agonist activation 36  
and antagonist activation of both BB and TB muscles dur- 37  
ing upper limb vibration exercise. 38

**Keywords** Electromyography · EMG activity · 39  
Coactivation · Vibration exercise · Vibratory bar 40

**Abbreviations** 41

BB Biceps brachii 42  
EMG Electromyography 43  
 $f_{out}$  Vibration frequency 44  
MSL Maximum sustainable load 45  
RMS Root mean square 46  
TB Triceps brachii 47  
WBV Whole body vibration 48

**Introduction** 49

Vibration exercise is an attractive complement to traditional 50  
forms of resistance exercise for athletes, elderly people, 51  
and health-compromised individuals (Rittweger 2010). 52  
Whole body vibration (WBV) has become the most popu- 53  
lar modality of vibration exercise for the lower limbs (Rit- 54  
tweger 2010; Roelants et al. 2006), while several tools like 55  
vibratory dumbbells (Bosco et al. 1999), vibrating pulley- 56  
like devices (Issurin and Tenenbaum 1999; Mischi and 57

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Cardinale 2009) and more recently vibratory bars (Moras et al. 2010; Poston et al. 2007; Rodríguez-Jiménez et al. 2014) have also been specifically designed for upper limb vibration exercise. One of the main features of the vibratory stimulus is that it has been shown to acutely improve specific aspects of neuromuscular performance like maximal power output (Bosco et al. 1999; Cochrane et al. 2008; Issurin and Tenenbaum 1999; Poston et al. 2007), maximal strength (Liebermann and Issurin 1997) and muscle activity, as evaluated with surface electromyography (EMG) (Bosco et al. 1999; Mischi and Cardinale 2009; Moras et al. 2010).

The majority of the studies that focused on the characteristics of the vibration exercise protocol such as vibration frequency, vibration amplitude, body position and load condition (with the objective to define adequate training doses) were conducted on lower limb muscles. In general, WBV exercise resulted in greater EMG activity compared to non-WBV exercise (Cardinale and Lim 2003; Roelants et al. 2006). These studies have used vibration frequencies ranging from 20 to 50 Hz and submaximal (Hazell et al. 2010; Ritzmann et al. 2013) or maximal loads (Rønnestad et al. 2012). For WBV, there is some evidence suggesting that EMG activity would increase linearly as a function of the vibration frequency within a range of 5–30 Hz (Berschlin and Sommer 2004; Pollock et al. 2010; Ritzmann et al. 2013), while inconsistent results were obtained within the 25–50 Hz range (Cardinale and Lim 2003; Hazell et al. 2007, 2010). Similarly to WBV, upper limb vibration exercise in the frequency range 23–31 Hz has resulted in greater EMG activity of upper limb muscles compared to non-vibration exercise (Bosco et al. 1999; Mischi and Cardinale 2009; Moras et al. 2010). In a recent study evaluating the effects of vibration on arm muscle activity during isometric elbow flexion and extension (Mischi and Cardinale 2009), the vibration-induced (28 Hz) increase in agonist muscle activity was more evident when higher levels of muscular tension were exerted, while the vibration-induced increase in antagonist coactivation was greater at lower levels of tension. However, until now, no study has systematically compared the impact of different vibration frequencies, including frequencies clearly above 30 Hz, on agonist activation and antagonist coactivation during upper limb vibration exercise with different loads.

Therefore, the aim of the present study was to assess the effect of different vibration frequencies on the EMG activity of the biceps brachii (BB) and triceps brachii (TB) muscles when acting as agonist and antagonist during pulling and pushing static exercises with different loads. The EMG data reported in the present study were collected in our previous work (Rodríguez-Jiménez et al. 2014). Based on the existing literature, we formulated the following hypotheses: (1) vibration exercise would result in greater agonist

activation and antagonist coactivation than non-vibration exercise; (2) the expected vibration-induced increase in agonist activation and antagonist coactivation would be proportional to the vibration frequency and (3) would occur regardless of the applied load.

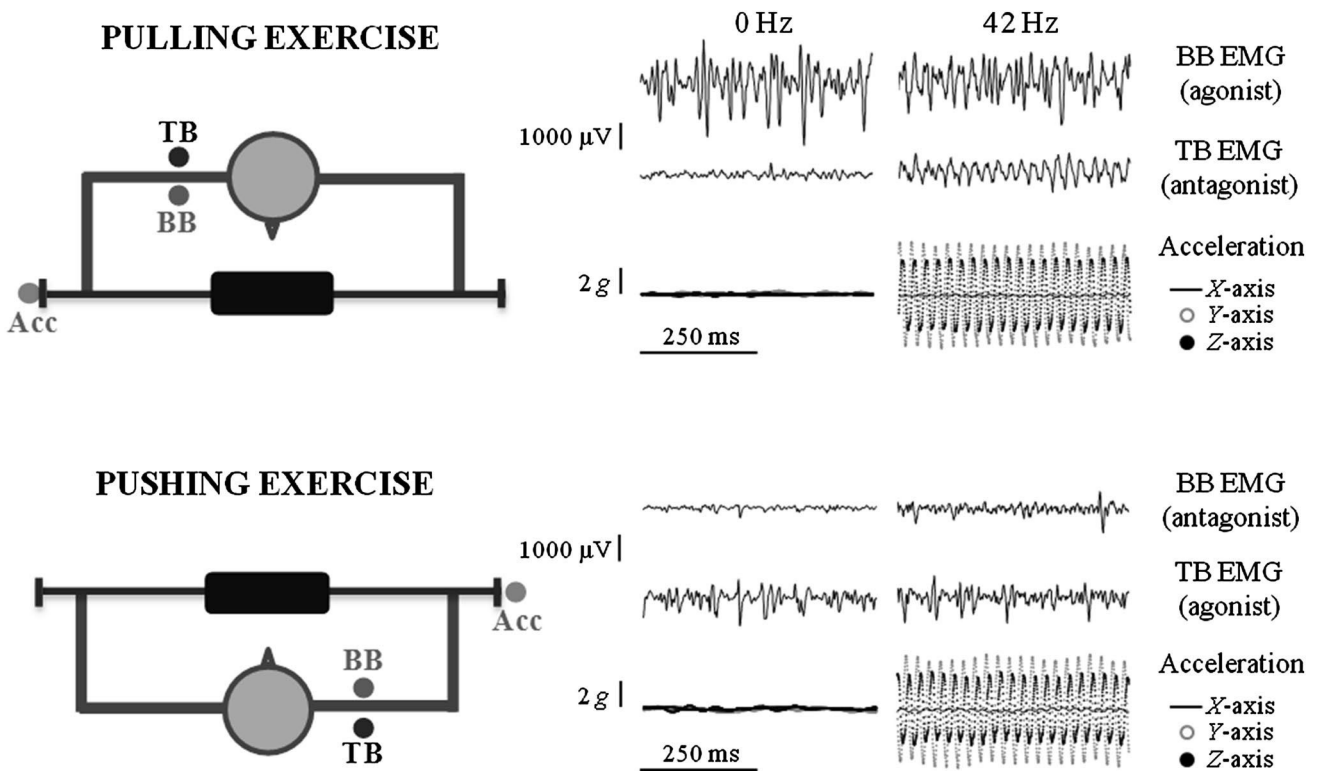
## Methods

### Subjects and study design

Fourteen healthy men (mean  $\pm$  SD: age  $25 \pm 5$  years; height  $179 \pm 5$  cm; body weight  $74 \pm 6$  kg) volunteered to participate in the study. They had previous experience in supervised resistance training (at least 2 sessions/week during the 2 years preceding the study), but not in vibration exercise. A repeated-measures (single-session) design consisting of 24 randomly-presented measurements was used to analyze the influence of four different vibration frequencies ( $f_{out}$ ) (0 [non-vibration], 18, 31 and 42 Hz) on the EMG activity of the BB and TB muscles during pulling and pushing static exercises with three different loading conditions (20, 50 and 80 % of the maximum sustainable load [MSL]). For each of the 24 measurements ( $4 f_{out} \times 2$  exercises  $\times 3$  loads), subjects were asked to hold the vibratory bar as steadily as possible for 10 s while keeping the elbow joint angle at approximately  $90^\circ$ . The study was conducted in accordance with the Declaration of Helsinki and was approved by the “Comitè d’ètica d’investigacions clíniques de l’Administració esportiva de Catalunya” (01/2011/CEICEGC). Written informed consent was obtained from all participants before inclusion.

### Experimental setup

For measurements during the lying row (pulling) exercise, subjects lay prone on a horizontal bench elevated over another horizontal bench. For measurements during the bench press (pushing) exercise, subjects lay supine on the lower horizontal bench with their knees flexed approximately  $90^\circ$ . The vibratory bar entails a three-phase electric vibrating motor (Italvibras M3/45-S02, 50 Hz, Fiorano Modenese, Italy) fixed in a cylindrical central body with two bars welded to its lateral sheets. The length and mass of the vibratory bar were 1.1 m and 8.8 kg, respectively. An-inverter (Omron Sysdrive 3G3JV, single phase 230 V, 0.55 kW) was used to reduce the speed of the motor (3,000 rpm) according to the required frequency. The two extremities of the vibratory bar were attached to the guide rails of a pneumatic resistance system (Keiser Half Rack, Fresno, CA) by means of carabineers, which allowed pneumatic pulleys to roll seamlessly with the movement of the bar (Rodríguez-Jiménez et al. 2014).



**Fig. 1** EMG data of a representative subject with  $f_{out}$  of 0 Hz (non-vibration) and 42 Hz and load of 50 % MSL during pulling and pushing exercises. For each exercise, the top shows the EMG data of the biceps brachii (BB) and triceps brachii (TB) muscles. The curves at the bottom illustrate the three-dimensional acceleration of the vibratory bar

158 Assessment of MSL

159 Subjects were asked to maintain the exercise position as  
 160 steadily as possible for 5 s, while the 90°-elbow joint angle  
 161 was verified with an electronic goniometer (SG110, Biome-  
 162 trics Ltd, Newport, United Kingdom). They were asked to  
 163 hold the vibratory bar with a pronated grip and not to mod-  
 164 ify the wrist position during the assessments. Previously  
 165 documented training experience was used as guidance for  
 166 selecting the initial test weight. The load was progressively  
 167 increased by 10 kg after each successful attempt until the  
 168 subject was unable to maintain the isometric position for  
 169 at least 5 s. Then, the load was decreased by 5 kg and a  
 170 last attempt was made. The load corresponding to the last  
 171 successful attempt was considered as the MSL (pulling:  
 172 86 ± 8 kg; pushing: 79 ± 14 kg).

173 Vibration exercise

174 Subjects were asked to hold the vibratory bar as steadily as  
 175 possible for 10 s with an elbow joint angle of 90°, and with  
 176  $f_{out}$  of 0 (non-vibration condition), 18, 31 and 42 Hz and  
 177 loads of 20, 50 and 80 % MSL. The order in which the dif-  
 178 ferent frequencies and loads were assigned was randomized

between subjects. The duration of recovery phases was  
 self-selected on an individual basis, with a minimum of  
 3 min between trials and 10 min between the two exercise  
 types (pushing and pulling).

EMG recordings

Surface EMG activity of the right TB (long head) and BB  
 muscles (Fig. 1) was recorded during all maximal (MSL  
 assessment) and submaximal trials. For each muscle, two  
 sensors (inter-electrode distance: 25 mm; Blue Sensor,  
 Medicotest, Ølstykke, Denmark) were placed longitudi-  
 nally to the orientation of muscle fibers, approximately  
 halfway from the motor point area to the distal part of the  
 muscle (Hermens et al. 2000). The reference electrode was  
 placed 5–6 cm away from the active sensors, as per man-  
 ufacturer’s specifications. Prior to electrode placement,  
 the skin was shaved, abraded and cleaned with isopropyl  
 alcohol to reduce inter-electrode impedance. The cables  
 were carefully taped to the skin to avoid motion artifacts.  
 EMG electrodes were connected to a 14-bit AD converter  
 (ME6000 Biomonitor, Mega Electronics, Kuopio, Fin-  
 land; sensitivity: 1 μV, CMRR: 110 dB) by pre-amplified  
 cables. Raw EMG signals were pre-amplified (gain: 305,

**Table 1** Raw EMG<sub>RMS</sub> data ( $\mu\text{V}$ ) of the biceps brachii (BB) and triceps brachii (TB) by frequency ( $f_{\text{out}}$ ) and load conditions

Exercise	Muscle	Load	Vibration $f_{\text{out}}$			
			0 Hz	18 Hz	31 Hz	42 Hz
Pulling	BB	20 % MSL	214 $\pm$ 149	210 $\pm$ 109	223 $\pm$ 114	250 $\pm$ 137
		50 % MSL	487 $\pm$ 289	488 $\pm$ 308	457 $\pm$ 275	549 $\pm$ 314
		80 % MSL	863 $\pm$ 436	784 $\pm$ 449	789 $\pm$ 453	777 $\pm$ 426
	TB	20 % MSL	84 $\pm$ 30	88 $\pm$ 31	108 $\pm$ 36	129 $\pm$ 41
		50 % MSL	143 $\pm$ 54	143 $\pm$ 53	180 $\pm$ 76	224 $\pm$ 62
		80 % MSL	217 $\pm$ 94	223 $\pm$ 91	266 $\pm$ 118	305 $\pm$ 114
Pushing	BB	20 % MSL	34 $\pm$ 15	87 $\pm$ 36	115 $\pm$ 62	134 $\pm$ 75
		50 % MSL	76 $\pm$ 50	112 $\pm$ 55	123 $\pm$ 58	172 $\pm$ 91
		80 % MSL	92 $\pm$ 46	124 $\pm$ 43	178 $\pm$ 124	226 $\pm$ 123
	TB	20 % MSL	89 $\pm$ 49	103 $\pm$ 39	116 $\pm$ 54	140 $\pm$ 48
		50 % MSL	238 $\pm$ 107	242 $\pm$ 101	267 $\pm$ 112	270 $\pm$ 115
		80 % MSL	431 $\pm$ 119	403 $\pm$ 137	414 $\pm$ 168	443 $\pm$ 125

Mean data  $\pm$  SD. MSL maximum sustainable load; 0 Hz = non-vibration condition

bandwidth: 8–500 Hz) and sampled at 2,000 Hz before being stored on a personal computer. No additional filters were used to avoid significant alterations of the EMG signals, as recently suggested by Ritzmann et al. (2010).

The middle 3-s portion of each maximal contraction (MSL assessment) and the middle 5-s portion of each sub-maximal contraction were considered for further analysis. EMG data were processed using custom-written Matlab routines (The MathWorks Version 7.11.0.584, Natick, MA). The root mean square of the EMG signal (EMG<sub>RMS</sub>) was calculated by applying the Parseval's theorem (Brandt 2011): first, the power spectrum was estimated using the periodogram; second, the power spectrum was integrated from 10 to 500 Hz; finally, the RMS value was obtained by taking the square root of the obtained power. To calculate agonist activation and antagonist coactivation, sub-maximal EMG<sub>RMS</sub> values of BB and TB muscles when acting as agonist (pulling for BB and pushing for TB) and antagonist (pushing for BB and pulling for TB) were consistently normalized to the maximal EMG<sub>RMS</sub> values (obtained at 100 % MSL) of respective muscles when acting as agonist. All EMG<sub>RMS</sub> data were normalized to the values obtained at 100 % MSL on an individual basis to ensure comparable conditions between the different subjects.

#### Statistical analysis

All data were initially evaluated for normality using the Kolgomorov–Smirnov test. The dependent variables were agonist activation and antagonist coactivation of BB and TB muscles. The independent variables were  $f_{\text{out}}$  and load. Two-way ANOVAs for repeated measures followed by Tukey post hoc comparisons were used to investigate the effect of  $f_{\text{out}}$  (0 [non-vibration], 18, 31 and 42 Hz) and load

(20, 50 and 80 % MSL) on dependent variables. Effect size statistic,  $\eta^2$ , was analyzed to determine the magnitude of the effect independent of the sample size. The level of significance was set at 0.05 for all the analyses. Data were analyzed using Statistical 7.0 (StataSoft Inc., Tulsa, OK) and are presented as means  $\pm$  standard deviations (SD).

#### Results

Raw EMG<sub>RMS</sub> values of BB and TB muscles by  $f_{\text{out}}$  and load conditions are presented in Table 1, for both pulling and pushing exercise.

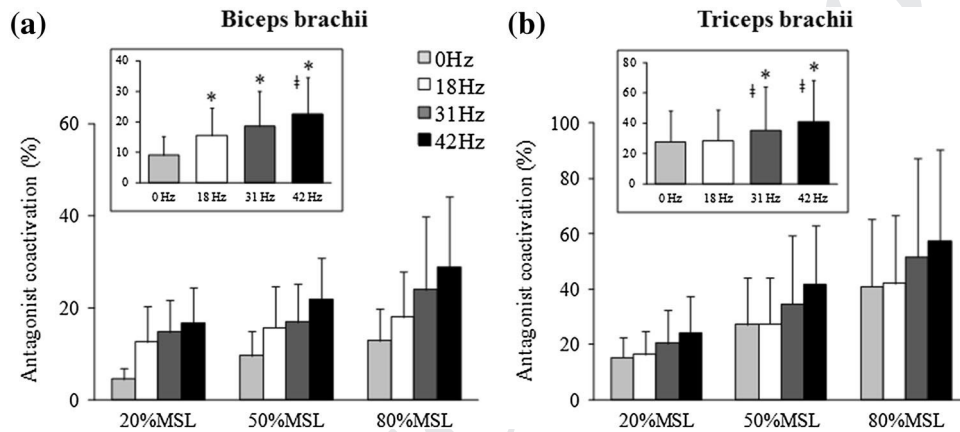
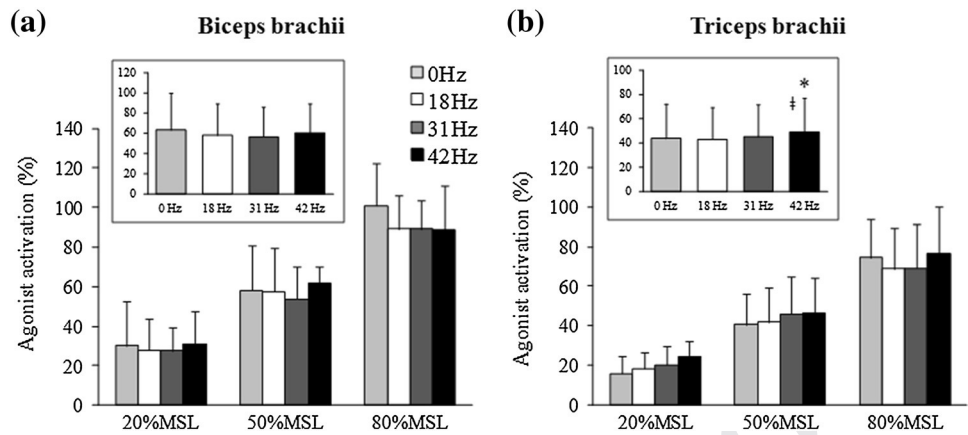
A significant load effect was consistently observed for all the EMG variables ( $p < 0.001$ ) with a progressive increase in BB and TB agonist activation (see also Fig. 2) and antagonist coactivation (see also Fig. 3) with increasing load.

Agonist activation of the BB muscle was not significantly affected by  $f_{\text{out}}$  ( $p = 0.18$ ;  $\eta^2 = 0.007$ ; Fig. 2a). On the other hand, a significant main effect of  $f_{\text{out}}$  ( $p < 0.05$ ;  $\eta^2 = 0.009$ ) was observed for agonist activation of the TB muscle (Fig. 2b). The highest  $f_{\text{out}}$  (42 Hz) elicited a greater EMG<sub>RMS</sub> response than 0 and 18 Hz (21  $\pm$  19 %;  $p < 0.05$ ).

Antagonist coactivation was significantly affected by  $f_{\text{out}}$  for both BB ( $p < 0.001$ ;  $\eta^2 = 0.278$ ; Fig. 3a) and TB ( $p < 0.001$ ;  $\eta^2 = 0.058$ ; Fig. 3b) muscles. For the BB muscle, coactivation was significantly greater at all vibration  $f_{\text{out}}$  compared to 0 Hz (18 Hz: 93  $\pm$  74 %; 31 Hz: 128  $\pm$  84 %; 42 Hz: 172  $\pm$  84 %). BB antagonist coactivation was also significantly greater at 42 Hz than at 18 Hz (44  $\pm$  15 %). For the TB muscle, coactivation was significantly greater at 31 and 42 Hz compared to both 0 Hz (29  $\pm$  4 and 50  $\pm$  9 %, respectively) and 18 Hz (24  $\pm$  2 and 45  $\pm$  8 %, respectively).



**Fig. 2** Changes in agonist activation of biceps (a) and triceps brachii (b) muscles by loading (20, 50 and 80 % MSL) and  $f_{out}$  conditions. For each muscle, the top small panel shows the post hoc comparisons for  $f_{out}$  main effect. Values are mean  $\pm$  SD,  $n = 14$ . \*Significantly higher than 0 Hz ( $p < 0.05$ ); †Significantly higher than 18 Hz ( $p < 0.05$ )



**Fig. 3** Changes in antagonist coactivation of biceps (a) and triceps brachii (b) muscles by loading (20, 50 and 80 % MSL) and  $f_{out}$  conditions. For each muscle, the top small panel shows the post hoc comparisons for  $f_{out}$  main effect. Note that the higher the  $f_{out}$  the higher

the antagonist coactivation in both muscles. Values are mean  $\pm$  SD,  $n = 14$ . **a** \*Significantly higher than 0 Hz ( $p < 0.01$ ); †Significantly higher than 18 Hz ( $p < 0.001$ ); **b** \*Significantly higher than 0 Hz ( $p < 0.05$ ); †Significantly higher than 18 Hz ( $p < 0.05$ )

266 **Discussion**

267 The main findings of this study were that: (1) agonist  
 268 activation was greater during vibration exercise at 42 Hz  
 269 compared to non-vibration exercise for the TB but not for  
 270 the BB muscle; (2) antagonist activation was greater dur-  
 271 ing vibration compared to non-vibration exercise for both  
 272 BB and TB muscles; (3) the vibration-induced increase in  
 273 antagonist coactivation was proportional to vibration  $f_{out}$   
 274 and (4) the vibration-induced increase in TB agonist acti-  
 275 vation and TB and BB antagonist coactivation was independ-  
 276 ent from the applied load.

277 Our first hypothesis was not fully confirmed as vibra-  
 278 tion exercise at the highest  $f_{out}$  resulted in higher agonist  
 279 activation than non-vibration exercise for the TB (+21 %)  
 280 but not for the BB muscle. A potential explanation for this  
 281 unexpected discordance between muscles may refer to bio-  
 282 mechanical differences between pulling and pushing the  
 283 vibratory bar. Pulling exercise was mainly characterized by

combined grip and pull-only coupling action with the ten- 284  
 sion acting mainly on the fingers. In contrast, pushing exer- 285  
 cise was characterized by combined grip and push with the 286  
 tension acting mainly at the palm. These differences could 287  
 have affected in a different way the biodynamic response of 288  
 the finger-hand-arm system during vibration exposure and 289  
 the vibration transmissibility to the target muscles (Aldien 290  
 et al. 2006; Dong et al. 2004, 2005). This leads us to con- 291  
 jecture that the higher effective palm force while pushing 292  
 the vibratory bar might have favored a larger vibration 293  
 power transmission to the upper extremity (Dong et al. 294  
 2005) compared with pulling, which in turn resulted in 295  
 greater agonist activation of the TB. However, the observed 296  
 vibration-induced increase in TB agonist activation was 297  
 only significant at the higher  $f_{out}$  of 42 Hz. It has been dem- 298  
 onstrated that vibration exercise can elicit a stretch reflex 299  
 response similar to the tonic vibration reflex (Pollock et al. 300  
 2012; Ritzmann et al. 2010), which can result in increased 301  
 motor unit recruitment and/or firing rate (Pollock et al. 302

2012; Ritzmann et al. 2010). In the same way, it has also been suggested that during vibration exercise the muscle tendon units are stretched in every vibration cycle (Cochrane et al. 2009), which could induce a frequency-dependent activation of the muscles spindles that elicit the stretch reflex responses detectable in the EMG signal (Ritzmann et al. 2010). If it is the case, it can be speculated that the observed vibration-induced increase in TB agonist activation at the highest  $f_{out}$  was caused, at least in part, by a greater number of stretch reflex responses compared to lower  $f_{out}$ . Previous studies have found greater agonist activation for the TB with vibration frequencies of 23 and 31 Hz (Moras et al. 2010), for both TB and BB with 28 Hz (Mischi and Cardinale 2009), and for the BB with 30 Hz (Bosco et al. 1999) and 50 Hz (García-Gutiérrez et al. 2014) when compared to non-vibration exercise. The discordances between these previous studies and our present work may be due to differences in several set up aspects like exercise posture, handle coupling actions, elbow joint angle, vibration device, vibration direction and magnitude of vibration acceleration, which could have influenced the level of agonist activation.

Antagonist activation was greater during vibration compared to non-vibration exercise for both BB and TB muscles. These findings are in line with previous WBV and upper limb vibration studies (Mischi and Cardinale 2009; Ritzmann et al. 2013). Interestingly, the fact that vibration moderately increased (for TB) or did not affect (for BB) agonist activation, but strongly increased both BB and TB antagonist coactivation leads us to conjecture that one or more of the following mechanisms could have played a role (Rothmuller and Cafarelli 1995): first, driving  $\alpha$ -motoneurons during vibration may increase excitation of Renshaw cells, which would inhibit Ia inhibitory interneurons, and increase coactivation (Brooks 1986); second, the complexity of the task for controlling force production during vibration could have resulted in greater coactivation since this latter is needed to control precise movements (Smith 1981). In the upper extremity, muscle coactivation has been demonstrated to produce greater movement accuracy and reduced phase lag to external perturbations (Humphrey and Reed 1983). It is also important to note that the vibration-induced increase in antagonist coactivation observed in the present study was greater for the BB than for the TB (~130 vs. ~40 %). The above-discussed differences in the hand-handle coupling actions between pulling and pushing exercises could have favored the higher BB coactivation while pushing the vibratory bar. Moreover, and according to the findings of Mischi and Cardinale (2009), both BB and TB seem to be more sensitive to the vibration exposure during static exercises involving elbow extension (such as the present pushing exercise) as compared to elbow flexion. This could occur because our

subjects had more difficulties in controlling joint rotation and upper extremity position during pushing exercise in general, hence adopting a neural strategy favoring increased antagonist EMG activity. The difficulty in controlling this exercise position was also accentuated by the fact that the hand-arm system acted like an inverted pendulum during the pushing exercise. That is, whereas a normal pendulum (pulling lying row exercise) is stable when the bar is hanging downwards, an inverted pendulum (pushing bench press exercise) is inherently unstable (kumar et al. 2013). It has been suggested that antagonist coactivation is necessary to aid the ligaments in maintaining joint stability, equalizing the articular surface pressure distribution, and regulating the mechanical impedance of the joint (Baratta et al. 1988). The observed vibration induced-increase of coactivation that was proportional to vibration  $f_{out}$  in the range of 18–42 Hz is considered to have a positive effect on joint protection and stabilization associated with postural control strategies during vibration exercise (Berschin and Sommer 2004; Mischi and Cardinale 2009).

As expected, the progressive increase of load from 20 to 80 % of the MSL was associated to a proportional increase in agonist and antagonist activity for both muscles. The present results confirm the previously-reported load-dependent enhancement of EMG activity during WBV (Hazell et al. 2010; Ritzmann et al. 2013) and upper limb vibration exercise (Mischi and Cardinale 2009). Accordingly, this suggests that external loading is a prerequisite for maximizing agonist and antagonist muscle recruitment during upper limb vibration exercise. More interestingly, the effect of vibration on TB agonist activation and antagonist coactivation was consistent whatever the load in the range of 20–80 % MSL. This implies that such modulation is not muscle-tension dependent, and that, whatever the applied load, the superimposed vibration can modify the pattern of motor unit recruitment with respect to the same exercise with no vibration. However, our present results are contrary to those reported by Mischi and Cardinale (2009). Again, this could be partly explained by differences between vibrating devices in stimulus transmission, resulting in a different vibration damping effect across loads. The acceleration and peak-to-peak displacement of our vibratory bar were hardly influenced by overloading partly because the vibration source was mounted directly onto the bar (Rodríguez-Jiménez et al. 2014). In contrast, the vibration transmissibility of the device used by Mischi and Cardinale (2009) is clearly tension-dependent because the sinusoidal vibrations are produced by an electromagnetic generator and mechanically transmitted to the hands through belts; an excessive damping effect onto the belts during the lower levels of muscular tension could partly explain the absence of vibration effects they observed. On the other hand, our current results are in line with the findings of Moras et al.

(2010) who observed similar vibration-induced increases in TB (agonist activation), deltoid and pectoralis EMG activity with low loads (vibratory bar of 15 kg), which is close to the lighter intensity (20 % MSL) used in the present study.

For the first time, we attempted to understand the impact of different vibration frequencies, including frequencies clearly above 30 Hz, on agonist and antagonist muscle activity during upper limb vibration exercise with different loads. The observed effects suggest specific vibration-related benefits as an alternative to resistance exercise with no vibration for both strength training and rehabilitation purposes. Moreover, the observed load-independent increases in agonist and antagonist muscle activity mediated by the vibratory stimulus suggest that such approach could be particularly suitable for rehabilitation programs where only low-force contractions could be produced. Based on the present findings, we propose the following recommendations for upper limb vibration exercise: (1) if the aim is to maximize agonist activation, this is possible for the TB (not for the BB) using a frequency of ~42 Hz and whatever load in the range of 20–80 % MSL; (2) if the aim is to maximize antagonist coactivation, this is possible for both TB and BB muscles using frequencies higher than 30 Hz and whatever load in the range 20–80 % MSL. Further research is required to demonstrate how (and if) the acute increases in agonist and antagonist EMG activity mediated by the vibratory stimulus could result into chronic neuromuscular and functional adaptations. Additional studies are also required to explore the mechanisms underlying the differences in muscle activity between vibration and non-vibration upper limb exercise, particularly for the antagonist muscles.

A major limitation of the present study is that we did not demonstrate the chronic effects of vibration exercise. However, the observed acute adjustments could have a potential long-term impact on specific neuromuscular features, provided the stimulus is repeated and adequately modulated over time. Another limitation of our study is that the EMG activity was only recorded from two upper limb muscles, while many other muscles of the shoulder, elbow and wrist contributed to the pulling and pushing exercises. Finally, because all measurements were conducted during static exercise, it is unclear whether similar results can also occur during dynamic actions (concentric or eccentric) which are actually more common in strength training.

## Conclusions

This study has demonstrated that upper limb vibration exercise by use of a vibratory bar did not modify BB agonist activation during lying row (pulling) static

exercise compared to non-vibration exercise. On the other hand, a vibration frequency of 42 Hz was able to increase TB agonist activation during bench press (pushing) static exercise. The vibration-induced increase in antagonist coactivation was proportional to the vibration frequency, and was greater for the BB than for the TB muscle. Therefore, the use of high vibration frequencies (within the range of 18–42 Hz) regardless of the level of muscular tension is supposed to be the most important prerequisite for optimizing upper limb vibration exercise.

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**Conflict of interest** None.

**Ethical standards** The experiments comply with the current laws of the country in which they were performed.

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