1	Multiscale control of bacterial production by phytoplankton
2	dynamics and sea ice along the western Antarctic Peninsula:
3	A regional and decadal investigation.
4	
5	Authors:
6	Hugh W. Ducklow ^a (Corresponding Author)
7	Oscar Schofield ^b
8	Maria Vernet ^c
9	Sharon Stammerjohn ^d
10	Matthew Erickson ^a .
11	
12	
13	
14	
15	
16	Addresses:
17 18	a. The Ecosystems Center, Marine Biological Laboratory, 7 MBL St. Woods Hole, MA 02543 USA. Email: <u>hducklow@mbl.edu</u> . Phone 508-289-7193
19 20	b. Institute of Marine and Coastal Sciences, Rutgers University, 71 Dudley Road, New Brunswick, NJ 08901-8525 USA. Email: <u>oscar@marine.rutgers.edu</u> .
21 22	c. Scripps Institution of Oceanography, 9500 Gilman Drive, La Jolla, CA 92093-0210 USA. Email: <u>mvernet@ucsd.edu</u> .
23 24	d. Institute of Arctic and Alpine Research, University of Colorado, Campus Box 450, Boulder, CO 80309-0450 USA. Email: sharon.stammerjohn@colorado.edu .
25	
26	
27	
28	Revised ms resubmitted March 07 2012

1 Abstract

2 We present results on phytoplankton and bacterial production and related hydrographic 3 properties collected on nine annual summer cruises along the western Antarctic Peninsula. This 4 region is strongly influenced by interannual variations in the duration and extent of sea ice cover, 5 necessitating a decade-scale study. Our study area transitions from a nearshore region influenced 6 by summer runoff from glaciers to an offshore, slope region dominated by the Antarctic 7 Circumpolar Current. The summer bacterial assemblage is the product of seasonal warming and 8 freshening following spring sea ice retreat and the plankton succession occurring in that evolving water mass. Bacterial production rates averaged 20 mgC $m^{-2} d^{-1}$ and were a low (5%) fraction of 9 10 the primary production (PP). There was significant variation in BP between regions and years, 11 reflecting the variability in sea ice, Chlorophyll and PP. Leucine incorporation was significantly correlated (r² ranging 0.2-0.7, p<0.001) with both chlorophyll and PP across depths, regions and 12 13 years indicating strong phytoplankton-bacteria coupling. Relationships with temperature were variable, including positive, negative and insignificant relationships ($r^2 < 0.2$ for regressions with 14 15 p<0.05). Bacterial production is regulated indirectly by variations in sea ice cover within regions 16 and over years, setting the levels of phytoplankton biomass accumulation and PP rates; these in 17 turn fuel BP, to which PP is coupled via direct release from phytoplankton or other less direct 18 pathways.

19

20

1 Keywords:

2 Bacteria, Antarctica, Bacterial Production, Primary Production, Sea Ice

3 Highlights:

- Bacterial & primary production measured at >400 stations on the West Antarctic Peninsula,
- 5 2003-11
- 6 Bacteria and phytoplankton are strongly coupled over depth, regions and years
- 7 Sea ice duration influences bacterial variability via mixed layer depth and primary
- 8 production
- 9

1 1.0 Introduction

2 Antarctic coastal and shelf waters exhibit high rates of primary productivity (Smith Jr. et 3 al., 2000) that support large stocks of upper level consumers including seabirds, seals and whales 4 (Smetacek and Nicol, 2005) and possibly constitute an important sink for atmospheric CO_2 5 (Arrigo *et al.*, 2008). As in other marine ecosystems, net primary production (PP) flows both to a 6 microbial foodweb and to higher trophic levels (Clarke et al., 2007). The relative allocation of 7 organic matter flow between these two pathways is governed by phytoplankton cell size, 8 dissolved organic matter (DOM) release and other variables (Legendre and Rassoulzadegan, 9 1996). The rich Antarctic production system might be expected to deliver large amounts of 10 organic matter for bacterial utilization; yet paradoxically, bacterial production (BP) rates in the 11 Ross Sea, Antarctica (Ducklow et al., 2001a) and in the Arctic Ocean (Kirchman et al., 2009a) 12 are significantly lower on average, than in other, lower latitude ocean ecosystems. Moreover BP 13 rates are low relative to the local PP, as well as in an absolute sense. That is, a smaller fraction of 14 the PP is incorporated into bacterial biomass than in other, mostly better-studied ecosystems. 15 Two contrasting explanations for the low bacterial to primary production ratio (BP:PP) 16 are: 1) the flux of labile organic matter to bacteria is low; or 2) low temperature inhibits bacterial 17 activity. There is a longstanding debate about the role of low temperature as an explanation for 18 low microbial rates in cold water (Pomeroy and Wiebe, 2001). Kirchman et al. (2009b) provided 19 a critical review of the factors potentially influencing BP rates in polar seas. Their review found 20 that the latest analyses do not indicate that cold temperatures per se, are the principal factor 21 regulating bacterial growth. There are few direct measurements of the availability or flux of

22 labile dissolved organic matter. There are some indications that top-down effects suppress

bacterial stocks, and thus BP rates (Bird and Karl, 1999), but there is no *a priori* reason to
 suggest that bacterivory or viral lysis is more intense than in other systems.

3 Antarctic coastal and shelf waters remain sparsely sampled due to the remote location 4 and difficulty of access even under ice-free conditions in summer. The western Antarctic 5 Peninsula (WAP) region is experiencing rapid climate warming, resulting in a gradient of sea ice 6 cover, ranging from a region of decreasing sea ice duration in the north to a now ice-free summer 7 season in the south (Montes-Hugo et al., 2009; Stammerjohn et al., 2008b). The Palmer 8 Antarctica Long Term Ecological Research program initiated a new series of measurements of 9 bacterial abundance and production rates in 2002-03 to investigate the biogeochemical 10 importance of bacteria in the regional carbon cycle (Daniels *et al.*, 2006; Ducklow *et al.*, 2006b) 11 and to test hypotheses about the regulation of bacterial processes in polar seas. Here we examine 12 the resulting large, regional-scale, multiyear dataset of bacterial production measurements in 13 relation to bacterial and phytoplankton stocks and primary production rates, with associated 14 physical and sea ice properties collected each summer from 2003 to 2011.

15 The overall goal of this paper is to investigate the regional-scale distribution and 16 interannual variability of BP in the west Antarctic Peninsula area in the context of local 17 hydrography and coupling to PP by phytoplankton. We begin by describing the hydrography of 18 the study area and placing our regional-scale, summertime observations in a seasonal context. 19 Next we present vertical profiles of phytoplankton and bacterial properties in the different 20 hydrographically-defined regions of the study area. Then we analyze the bacterial distributions at 21 a range of spatial and temporal scales (local/annual to regional/decadal) primarily through 22 regressions with phytoplankton (PP, Chl) properties. Finally we examine long-term 23 (climatological) distributions and decadal records of phytoplankton and bacterial properties, and

1 relate them to variations in sea ice and climate forcing. We conclude that accumulation of 2 phytoplankton biomass regulates BP in the Antarctic Peninsula region. Phytoplankton biomass 3 has increased in the south and decreased in the north in response to climate change and sea ice 4 decline (Montes-Hugo *et al.*, 2009); however as yet we are unable to say if these changes in the 5 ecosystem have impacted other microbial processes. BP is greater in the south region, where 6 phytoplankton biomass is also high, and where there is now an ice-free summer season. But, 7 BP:PP is greater in the north region, indicating a complex and dynamic picture of bacteria-8 phytoplankton relationships.

9 **2.0 Materials and Methods.**

10 2.1 Study area and sampling. The PAL study region encompasses a roughly 200 x 700 11 km area along the western Antarctic Peninsula, extending from the coast in the east across the 12 continental shelf to the offshore, continental slope region, and from Anvers Island in the north to 13 Charcot Island in the south (Figure 1). The study area is divided into coastal, shelf and offshore 14 (slope) regions on the basis of bathymetry, hydrographic properties and ecology (Martinson et al., 2008). The mean depth of the shelf is 430 m. Annually-occupied hydrographic stations were 15 16 spaced 20 km apart along cross-shelf lines 100 km apart (Waters and Smith, 1992). Marguerite 17 Bay, immediately south and east of Adelaide Island, experiences the largest phytoplankton 18 bloom in the region. Stations south of Marguerite Bay typically have sea ice cover in January, in 19 contrast to the northern region. Thus, we differentiated Marguerite Bay and the southern part of 20 the study area (lines 100, 000 and -100) from the northern stations on the 200-600 lines (Fig. 1). All stations on the 200 to 600 lines, including stations in Marguerite Bay were sampled in 2003-21 22 08 resulting in comprehensive sampling of the shelf region. Occasional stations were sampled in 23 the coastal region (triangles in Fig. 1). Sampling on the Southern (-100, 000 and 100) lines

started in 2009. Stations sampled after 2009 are given in Supplementary Table S1. The full
 study region is described in detail elsewhere (Ducklow, 2008; Ducklow et al., 2006a).

3 The regional-scale datasets reported here were obtained during annual summer cruises 4 aboard ARSV Laurence M Gould in 2003-2011, occurring roughly between 01 January and 10 5 February each year (Table S1). At each station, sampling consisted of one or more hydrocasts 6 with a Seabird CTD-rosette system and 24 Niskin-type 12-liter bottles fitted with Vicor TM 7 silicone springs. In general, two bottles were closed at each of 12 depths, extending from the 8 surface to the bottom, irrespective of bottom depth, and with sampling concentrated in the upper 9 50-100 m. Usually 4-6 samples were obtained in the upper 50 m. One of the two Niskin bottles at 10 each depth was subsampled into 5% HCl-washed, deionized and seawater-rinsed polycarbonate 11 bottles for bacterial and biogeochemical assays. The other bottle was dedicated to phytoplankton 12 measurements.

Seasonal time-series sampling was performed at Station E, 5 km from Palmer Station
(Fig. 1; 64.48 deg S, 66.04 deg W) approximately every 4 days between late October and late
March in 2002-06 and 2008-11. Sampling and analyses were similar to the cruise-based
sampling but using Zodiac boats as sampling platforms, and Go-Flo bottles hung individually at
preselected depths on the hydrowire.

18 2.2 Analytical methods. Chlorophyll (Chl) and PP rates and corresponding bacterial 19 abundance and BP rates were determined at every hydrostation. Water samples from this region 20 include both bacterial and archaeal cells in varying proportions. Autofluorescent picoplankton 21 are <1% of the total count. In the upper 100 m in summer, >80% of the total picoplankton count 22 is bacterial (Church *et al.*, 2003), and the counts are termed bacterial for simplicity. As with 23 abundance, we term the leucine incorporation data (see below) as indicating heterotrophic

bacterial production (BP) rates, recognizing that some small and variable fraction might be
 attributed to other, nonbacterial, organisms.

3 Chl and PP measurements were conducted on samples from the euphotic zone (0.1-1%) of 4 surface irradiance) as determined from PAR measurements made prior to each CTD cast, as 5 described in Vernet et al. (2008). PP was measured by C14 bicarbonate incorporation in 24-hour 6 deck incubations. Chl was assayed fluorometrically on acetone extracts. Bacterial abundance was 7 determined at all depths sampled from surface to bottom. Samples for abundance determinations 8 were preserved in 2% formaldehyde, kept frozen at -80°C and returned to the home laboratory 9 for flow cytometric analysis using SYBR-Green (Invitrogen, Carlsbad, CA). Sample analyses 10 took place 3-6 months after each cruise. Flow cytometer samples from 2003-2007 were assayed 11 on a Beckman-Coulter EPICS Altra at Virginia Institute of Marine Science. Samples for 2007-11 12 were assayed using a Becton-Dickinson FacsCalibur at the Marine Biological Laboratory in 13 Woods Hole. Results for the two instruments were compared on the entire 2007 sample set and did not differ significantly. The analytical protocols of Gasol and del Giorgio (2000) were 14 15 followed throughout.

BP rates were derived from rates of ³H-leucine incorporation measured on samples 16 17 extending over the upper 50-100 m. The leucine assays followed a procedure modified from the 18 protocol originally proposed by Smith and Azam (1992). Briefly, triplicate 1.5 ml samples were incubated in the dark for ~3 h with ³H-leucine (MP Biomedical, Santa Ana, CA; >100 Ci/mmol, 19 20 20-25 nM final concentration) in 2.0 ml microcentrifuge tubes (Axygen SCT-200, Union City, 21 CA). Incubations were maintained within 0.5°C of the in situ temperature in refrigerated 22 circulator baths and terminated by the addition of 0.1 ml of 100% trichloroacetic acid (TCA). 23 Samples were concentrated by centrifugation, rinsed with 5% TCA and 70% ethanol and air-

dried overnight prior to radioassay by liquid scintillation counting in Ultima Gold cocktail
 (Perkin-Elmer, Waltham, MA). Blank values of TCA-killed samples were subtracted from the
 average of the triplicates for each discrete depth sample.

4 2.3 Data analysis. The coefficient of variation of triplicate flow cytometric counts was 5 5%. The coefficient of variation of triplicate leucine assays was 6%. The mean blank value was 6 90 dpm. These estimates include the analytical precision and sample pipetting and processing 7 errors. The limit of detection for leucine incorporation rates (ten times the background for a 3 h incubation) is $<1 \text{ pmol } l^{-1} h^{-1}$ (~0.05 mgC m⁻³ d⁻¹). Bacterial abundance was converted to carbon 8 biomass using 10 fgC cell⁻¹ (Fukuda et al., 1998). Leucine incorporation rates were converted to 9 bacterial carbon production using 1.5 kgC mol⁻¹ (Ducklow *et al.*, 2000; Kirchman *et al.*, 2009b). 10 11 Chl, PP, bacterial abundance and leucine incorporation rate data exhibited relatively little 12 variability below 50 meters (see below). Bacterial properties were therefore integrated to 50 m to 13 provide a consistent comparison across regions and years. This depth range encompassed the euphotic zone in all but a few cases. Integration depths were obtained by linear interpolation 14 15 between sample depths if necessary. Statistical analyses were performed with Systat (ver. 13, 16 Systat Inc. Chicago IL). Regression analyses of discrete depth values were confined to the upper 17 20 meters (approximating the summer mixed layer) to remove the effect of depth-related 18 variability.

19 All data considered in this paper are available at the Palmer LTER DataZoo:

20 <u>http://oceaninformatics.ucsd.edu/datazoo/data/pallter/datasets</u>.

21 **3.0 Results**

3.1 Seasonality and hydrography. In this paper, we present observations obtained at the
 regional scale (Fig. 1) over nine years, 2003-11. Each year's cruise provides a snapshot of

midsummer (January) conditions in the marginal sea ice zone to the west of the Antarctic 1 2 Peninsula, (usually) following the annual sea ice retreat. These midsummer observations can be 3 placed in the seasonal context by comparison with twice-weekly time series sampling undertaken 4 at Palmer Station between October and March each field season. This comparison assumes that 5 the seasonal cycle is similar in the nearshore and offshore regions. Water column (0-50 m) 6 integrated leucine incorporation rates are low in the Austral spring (Figure 2), around 200 nmol $m^{-2} hr^{-1}$, corresponding to a mean volumetric rate of ~5 pmol l⁻¹ hr⁻¹, or a bacterial production 7 rate of about 5-10 mg C m⁻² d⁻¹. Peak annual rates (~1000 nmol m⁻² hr⁻¹) occur in mid-January to 8 9 early February, coincident with the summer cruise period. There are few BP measurements from winter (i.e., April to September), when rates are typically $<5 \text{ pmol } l^{-1} hr^{-1}$ (Grzymski *et al.*, 10 11 submitted).

12 We are principally concerned with the ecological and biogeochemical character of the 13 upper 50 meters of the water column in summer, the illuminated zone and season in which most 14 biological activity occurs. In summer Antarctic Surface Water (AASW) is a freshened and 15 warmed version of the sea ice-produced Winter Water (WW). It is freshened by spring sea ice 16 melt and subsequently warmed by exposure to solar radiation (Figure 3). The AASW typically 17 overlies a remnant amount of the colder, saltier WW. In summer the water in the upper 50 m can 18 be warmed and/or freshened considerably, depending on location and the time since sea ice 19 retreat. Vertical profiles of temperature and salinity (Figure 4) show that warming and 20 freshening were on average restricted to the upper 50 m, with most change occurring in the upper 21 20 m encompassing the summer mixed layer. Different regions also exhibited varying 22 characteristics: for example, in summer, waters in the Marguerite Bay and South regions were 23 fresher and colder (<0°C) due to more recent or ongoing ice melt (including sea ice, brash ice

1 and glacier melt), whereas shelf and oceanic waters were typically warmer and saltier due to 2 longer exposure to solar warming and relatively less in situ sea ice melt and/or longer exposure 3 to wind mixing with the underlying saltier WW. This mosaic of characteristics reflects regional 4 variations in the seasonal modification of the summer mixed layer with the underlying WW, 5 setting the stage for plankton succession, including development of phytoplankton and bacterial 6 properties following ice retreat and the spring phytoplankton bloom.

7

3.2 Phytoplankton stocks and production. We observed considerable interannual and 8 regional variability in midsummer Chl and PP. For example, in 2009, an early ice retreat year, 9 both rates and stocks were very low, with few discrete values even reaching the 2003-11 means 10 (Figure 5 C,D). In years with later ice retreat (e.g., 2005, Fig. 5 A,B), peak Chl and PP exceeded 10 mg m⁻³ and 100 mgC m⁻³ d⁻¹, respectively. Maximum Chl concentrations were nearly always 11 12 in the surface mixed layer and declined with depth. Local hydrography showed variable 13 relationships with Chl and PP (Supplemental Figure S1), whereas regional differences were 14 more distinct. For example, PP tended to be highest in Marguerite Bay, and lowest in the 15 offshore slope region, irrespective of salinity or temperature. In the coastal region, PP was 16 weakly but significantly (p<0.001) inversely related to both salinity and temperature, with the 17 highest surface PP values occurring in cold, fresh nearshore waters (Suppl. Fig. S1 C,D; 18 Supplementary Table S2).

19 3.3 Bacterial distributions and relationships. Vertical profiles of bacterial abundance 20 and leucine incorporation rates (Figure 6; Supplemental Figures S2,S3) were similar to the 21 profiles of phytoplankton properties. In general the highest values were observed at the surface 22 and declined with depth, mirroring the phytoplankton distributions. There was little year to year 23 variation in the abundance profiles, with the exception of 2007 (up to five times higher than

average at all depths) and 2010 (10-50% of mean values). Over all years there were no
systematic differences between regions (see below). Peak surface values seldom exceeded 10⁹
cells l⁻¹. Leucine profiles were more variable, with greater surface (upper 20 m) enhancements
and rates exceeding 60 pmol l⁻¹ hr⁻¹ in 6 of 9 years. The highest leucine rates tended to occur in
years of late ice retreat (e.g., 2005, Figure 6A), mirroring PP rates and Chl.

6 There were no systematic (i.e., across regions and/or years) relationships between leucine 7 incorporation and hydrography, when <20 m samples were pooled across regions 8 (Supplemental Figure S4), but there were some within-region relationships. For example, 9 leucine rates were inversely related to salinity in the Shelf (Fig. S4C) and Southern regions, and 10 directly related to temperature in Marguerite Bay (Fig S4A and Table S2). However it is notable 11 there was no universal influence of temperature across regions: equally high rates were found in 12 the Southern region at low temperature (-1°C) and in Marguerite Bay at high temperatures 13 (>3°C; Fig. S4B). Temperature effects on leucine incorporation are considered further below.

14

3.4 Chlorophyll and PP relationships with leucine incorporation and bacterial

15 *production.* Chlorophyll was the most reliable indicator of leucine incorporation rates within and across depths, years and regions. Coefficients of determination (R²) for chlorophyll-leucine 16 17 regressions (discrete depth samples) over all depths and regions ranged from 28 to 72% for 18 individual years 2003-11 (Table 1). Over all regions and all nine years combined, chlorophyll 19 explained 52% of the discrete-depth variation in leucine incorporation. These Chl-Leucine 20 relationships reflect both within- and between-regional components. Within the Shelf region 21 alone, Chl explained at least 25% of the leucine variability in five of nine years (Supplementary 22 Table S3). The pooled data sets in Figure 7 show that the overall relationships were partly 23 determined by gradients in Chl and Leu increasing from the Slope through the Shelf regions to

Marguerite Bay. Relationships within other regions were not significant in most years, possibly
because of lower sample sizes. Regulation of leucine incorporation rates by Chl was also
manifested at larger, cross-regional and decadal scales. Water column integrated leucine
incorporation rates were significantly related to water column integrated chlorophyll within the
Shelf, Ocean and Marguerite Bay regions over all years, but not in the Coastal and Southern
regions (Supplementary Table S4). The slopes of the regressions varied between years and
regions, indicating different responses of leucine incorporation to Chl accumulation.

Discrete-depth leucine incorporation rates were also significantly related to PP (e.g., Fig.
7) across regions and within years (**Table 2**). Euphotic zone-integrated relationships were
significant within the Shelf, Ocean and Marguerite Bay regions (Table S4). The slopes of the *linear* regressions of BP on PP averaged about 1%, suggesting very low ratios of bacterial to
primary production rates. The integrated water column values are somewhat higher (~5%).
BP:PP ratios are further addressed below.

14 3.5 Regional relationships and trends. In spite of interannual variations, there were 15 significant differences in phytoplankton and bacterial stocks and production rates across years 16 and between regions, suggesting clear differences in the regulation of these variables by regional 17 hydrography and ecology. In general, most variables were highest in the Marguerite Bay and 18 Southern regions of the study area (where surface waters were relatively fresher and often colder 19 due to recent or ongoing ice melt), and lowest in the offshore, oceanic region (Figure 8). 20 Phytoplankton and bacterial properties in the Shelf region were intermediate, greater than the offshore, but less than the Marguerite Bay and Southern areas. There was little variation in total 21 bacterial abundance across regions (see also Fig. S2), with about 5 x 10^8 cells 1^{-1} in the upper 50 22 23 meters. The mean regional leucine incorporation rates closely mirrored Chl and PP (Fig. 8). The

BP:PP ratio did not differ among regions (ANOVA, p>0.05; mean 4%). Time series 1 2 observations for the well-sampled (over 200 stations) Shelf region also showed similar patterns 3 for the phytoplankton and bacterial properties (Figure 9). All four properties increased steadily 4 over 2003-06, then were lower, without a discernable trend in 2007-09, and then high again in 5 2010 (PP) or 2011 (Chl, leucine). The similarity of phytoplankton and bacterial variability in 6 these plots illustrates coupling at cross-regional and decadal scales (Figure 10).

7

3.6. Role of temperature. Temperature appears to influence BP in some regions or years, 8 but not others. Within the shelf region, there was no relationship in four of the nine years or in all years pooled together (Figure 11, Table 3). There were significant (p < 0.01) but weak ($R^2 =$ 9 10 0.002-0.20) relationships in five years, but the slopes were negative in three years (i.e., rates 11 were higher in colder waters) and positive in two other years (suggesting suppression in colder 12 water). Clearly there was no universal suppression of leucine incorporation rate by cold 13 temperature.

14 4.0 Discussion.

15 Discerning the large-scale controls on bacterial variability in marine systems requires 16 sampling over a broad range of time and space scales that is difficult to satisfy. Time series 17 approaches have been undertaken successfully in a number of locations, but typically only one or 18 a few fixed stations are observed repeatedly over longer time spans (e.g., Fuhrman et al. 2006; 19 Morris et al. 2005; Li, 2009). In contrast more geographically extensive studies tend to be 20 limited to a few seasons over one or two years (e.g., Garrison et al., 2000; Ducklow et al., 21 2001a; Straza et al. 2009). The Long Term Ecological Research program enables an 22 unprecedented examination, in terms of geographic and temporal coverage, of the 23 interrelationships among climate, sea ice, hydrography, plankton ecology, biogeochemical

1 processes and microbial dynamics (Ducklow et al., 2006a; Karl et al., 1996). Here we employ 2 these extended and extensive observations to address the question of bacterial coupling to 3 phytoplankton and sea ice in the marginal sea ice zone of the Antarctic Peninsula. The marked 4 seasonality of Antarctic coastal seas, the intensity of the phytoplankton bloom and the absence of 5 complicating terrestrial inputs of organic matter all make this a uniquely valuable system in 6 which to examine this fundamental question in microbial oceanography. In addition, the 7 Antarctic Peninsula region is warming rapidly (Meredith and King, 2005; Vaughan et al., 2003), 8 driving a range of biological responses (Montes-Hugo et al., 2009; Schofield et al., 2010). The 9 observations reported here establish a baseline for analyzing possible microbial responses in this 10 rapidly-changing ecosystem.

11 4.1 Phytoplankton dynamics and coupling to bacteria. We examine the working 12 hypothesis that large-scale climate forcing of sea ice extent and duration modulates the timing 13 and distribution of phytoplankton blooms (Vernet *et al.*, 2008). In turn, we hypothesize that 14 regional- and interannual differences in this physical-biological coupling produce a supply of 15 labile organic matter whose variability generates geographic patterns and year-to-year variability 16 in BP. We begin with a brief review of the larger-scale regulation of the phytoplankton bloom by 17 the annual cycle and duration of regional sea ice coverage. Extreme seasonality in Antarctica sets 18 the stage for the spring phytoplankton bloom, historically dominated by diatom production in 19 response to increasing solar irradiance and mixed layer shoaling triggered by melting sea ice 20 (Smith Jr. and Nelson, 1985; Smith Jr. and Nelson, 1986). Across the region, interannual 21 variations in the magnitude of the bloom are regulated by variations in sea ice extent and the 22 timing of sea ice retreat. In years with a late sea ice retreat, enhanced stratification from melting 23 sea ice leads to shallower mixed layers and higher PP. Vernet et al. (2008) illustrate this

1	relationship with a decade-long analysis of climate variability, sea ice retreat and PP for the PAL
2	LTER study region (200-600 lines), 1995-2006. Updated sea ice retreat anomalies for this region
3	are shown for ice years 2003-2011 in Figure 12. Sea ice retreat in ice years 2004, 2005 and 2009
4	(corresponding to Chl and PP in January 2005, 2006 and 2010, respectively) was anomalously
5	late over the shelf and inshore, and Chl and PP in those years were also high (Figs. 9,10). In
6	contrast, in years with early sea ice retreat (e.g., ice years 2006-08, Fig. 12), Chl and PP are
7	inhibited by high spring winds maintaining deeper mixed layers as indicated by low Chl and PP
8	in January 2007-09 (Fig. 9). At the regional scale, areas with later sea ice retreat (e.g.,
9	Marguerite Bay and the South) have higher Chl and PP (Fig. 8).
10	These patterns do not hold in all years. Sea ice retreat was anomalously early in 2010, but
11	Chl and PP were disproportionately high in January, 2011. Smith et al. (2008) showed that the
12	seasonal progression of the phytoplankton bloom did not necessarily follow the retreating ice
13	edge as the simple model would predict. In particular, they observed that early sea ice retreat
14	offshore resulted in enhanced blooms in the vicinity of the Southern Antarctic Circumpolar Front
15	Zone, counter to the sea ice/bloom relationship just stated for the shelf region. This offshore
16	bloom was succeeded by inshore blooms in years when the early sea ice retreat offshore was
17	followed by a late retreat inshore of the ACC front. In addition to these effects, the responses of
18	PP to sea ice retreat may be changing. Phytoplankton have responded differently to sea ice
19	decline in different areas of the WAP area since 1978, decreasing in response to sea ice decline
20	in the north, while increasing in response to similar declines in the south (Montes-Hugo et al.,
21	2009). Thus as mentioned above, the early sea ice retreat in 2010 resulted in relatively high PP in
22	January, 2011, particularly in the south, which was once perennially ice covered. Sea ice decline
23	opens new areas of ocean surface to increased irradiance and increasing PP (Peck et al., 2010).

1	These large-scale processes provide a foundation for exploring regional and interannual
2	variability in regional bacterial processes as well. Inputs of terrestrial organic matter to Antarctic
3	waters are minimal, and the immediate nearshore zone has DOC concentrations typical of the
4	open ocean (40-70 µM, http://oceaninformatics.ucsd.edu/datazoo/data/pallter/datasets; datasets
5	69,70). Therefore, heterotrophic bacteria in Antarctic shelf waters must ultimately depend on in
6	situ PP for organic matter. The question of coupling between phytoplankton and bacterial
7	activity has been studied at several locations around the Antarctic continent (Billen and
8	Becquevort, 1991; Leakey et al., 1996) including the WAP region. In the RACER (Research on
9	Coastal Antarctic Ecosystem Rates) Project, Karl and colleagues (Bird and Karl, 1991; Bird and
10	Karl, 1999; Karl et al., 1991) investigated microbial processes in waters of the northern Antarctic
11	Peninsula and Drake Passage in summer 1987 and spring 1989. Bacterial activity was not
12	correlated with Chl (as in some of our individual regional or yearly datasets) and did not
13	immediately respond to the spring phytoplankton bloom in the Gerlache Strait. Bacterial biomass
14	was <2% of the total plankton biomass and BP was ~3% of the co-occurring PP. Bird and Karl
15	concluded that at least in their study area and during the spring bloom period, the microbial loop
16	was uncoupled from primary producers, but they added that the uncoupling was not necessarily
17	widespread in space and time, and may be expressed more strongly in other seasons.
18	Morán and colleagues investigated phytoplankton-bacteria coupling experimentally in the
19	same region (Morán and Estrada, 2002; Morán et al., 2001). They identified the flow of recently-
20	synthesized DOC from active phytoplankton (14% of total particulate plus dissolved PP) and its

- 21 immediate (within hours) incorporation into bacteria as the mechanism of coupling. They
- showed that the released DOC met the metabolic requirements of bacteria in the same region
- 23 studied in RACER and concluded that bacteria and phytoplankton were directly coupled. They

also determined that BP was a very low fraction (mean 1.5 ± 0.4%) of the total particulate plus
dissolved PP, but termed the coupling "strong" nonetheless. That is, the strength of the coupling
is indicated by the covariation of bacterial and phytoplankton properties, not the ratio of the
production rates.

5 It follows from these results that leucine incorporation rates should be correlated with PP 6 and Chl, if bacteria are dependent on the recent products of photosynthesis, sensu Morán and 7 colleagues. Bacteria may be related to Chl, but not necessarily to simultaneous PP, if organic 8 matter is supplied by other processes such as zooplankton grazing activity (Cole et al., 1982; 9 Ducklow and Carlson, 1992). If the coupling is more remote as suggested by Karl and 10 colleagues, for example if bacteria depend on accumulated semilabile DOC (Ducklow, 2003), 11 BP-Chl relationships might be absent altogether. In our study, leucine incorporation was 12 correlated with both PP and Chl across years, regions and depths. Relationships with Chl were somewhat stronger than with PP. For example, R^2 values for Chl-Leucine regressions were 13 14 usually greater than values for PP-Leucine (Tables 1,2). Relationships between dissolved 15 primary production (not measured in our study) might have been stronger than particulate 16 primary production (Morán et al., 2009). We interpret the stronger relationships with Chl as 17 indicating greater dependence of BP on DOM supplied from a variety of trophic pathways 18 besides direct release of photosynthetic products from phytoplankton alone. Thus, in regions or 19 years when sea ice retreat triggers large phytoplankton blooms, BP is higher, and vice-versa. As with other ecosystem components, sea ice regulates variations in BP via its effects on trophic 20 21 coupling to food supply.

4.2 Multiscale regulation of bacterial production. Here we demonstrate widespread,
 though variable, phytoplankton-bacterial coupling by relating bacterial leucine incorporation

1 (BP) rates to chlorophyll stocks and PP over a range of time and space scales. Such relationships 2 are well-known in aquatic microbial ecology. Typically the strongest relationships are expressed 3 at the largest scales. That is, bacterial and phytoplankton properties tend to be related most 4 strongly (highest r values) when many studies from different regions and years are pooled 5 together, illustrating the ultimate reliance of bacteria on primary producers in a wide range of 6 aquatic systems (Bird and Kalff, 1984; Cole et al., 1988; Ducklow and Carlson, 1992; Kirchman 7 et al., 2009b; Li et al., 2004). Strong relationships are also commonly observed over full annual 8 cycles, especially in the highly seasonal polar seas, encompassing wide dynamic ranges of 9 bacterial and phytoplankton properties encountered between winter and summer (e.g., Pearce et 10 al., 2007; Garneau et al., 2008).

11 Relationships at smaller scales, e.g., within seasons and regions or depth intervals, are 12 indicative of mechanistic, process-level couplings and can be more complicated. For example, in 13 the Antarctic Polar Front Zone, Simon et al. (2004) demonstrated a significant correlation 14 between euphotic zone (0-100 meter) Chl and integrated leucine incorporation in the underlying 15 mesopelagic zone (100-1000 m), but not with euphotic zone (0-100 m) incorporation rates. 16 However, there was a correlation between BP and PP within the upper layer. These varying 17 mechanisms and degrees of coupling were manifested through relationships among Chl, leucine 18 incorporation and concentrations of dissolved amino acids and carbohydrates (Simon and 19 Rosenstock, 2007). They observed significant relationships on a summer cruise but not in the fall 20 three years later. Their comparisons potentially include competing seasonal, regional and 21 interannual sources of variability. Similarly Kirchman et al. (2009a) established correlations 22 between BP and Chl in the western Arctic Ocean (Chuckchi Sea and Canada Basin) in spring-23 summer 2004 but not in 2002.

1	In our study, leucine incorporation rates in summer within the shelf region were
2	significantly correlated with Chl in all years except 2011 (Table S3), both in the surface layer
3	(discrete depths 0-20 m) and in the euphotic zone (integrated 0-50 m). Bacterial abundances
4	varied little between years and regions (Figs. 8, S3), and seldom exceeded 1 x 10^9 cells l ⁻¹ . Bird
5	and Karl (1999) suggested that bacterial biomass in the Peninsula region may have been
6	suppressed by heterotrophic nanoplankton grazers. Some support for this idea comes from
7	comparison with the Ross Sea where a large bacterial bloom (up to 3×10^9 cells l ⁻¹) was
8	observed in 1996-97 (Ducklow et al., 2001a) and where the ratio of bacteriovores to bacteria is
9	lower than in the Peninsula region (Ducklow et al., 2006a). In our study, bacterial abundance
10	was relatively stable, suggesting possible grazer control.
11	The summertime bacterial assemblage in WAP shelf waters results from selective growth
12	of relatively few heterotrophic populations, transforming a high-diversity winter community
13	dominated by chemolithoautotrophs into a lower-diversity, mostly heterotrophic assemblage
14	(Grzymski et al., submitted). Straza et al. (2010) showed that substrate utilization by the summer
15	community was dominated by relatively few taxa of Gammaproteobacteria, Sphingobacteria-
16	Flavobacteria, and Alphaproteobacteria. This pattern was consistent across the shelf and
17	extended throughout the study region.
18	4.3 Role of temperature. The role of cold temperature as a possible suppressant of
19	microbial activity is a classic problem in polar oceanography (Karl, 1993; Karl et al., 1996;
20	Pomeroy and Deibel, 1986). Indeed, on average, both primary and bacterial production rates

21 were lower in the Ross Sea and Western Arctic Ocean than in other warmer, lower latitude

- 22 regions reviewed by Kirchman et al (2009b). Kirchman et al (2009a) found significant
- 23 relationships between BP and temperature both within and across seasons and years (spring-

summer, 2002, 2004) in the western Arctic Ocean, but Garneau et al. (2008), working in nearby
Franklin Bay, did not. Simon et al (1999) demonstrated that temperature regulated bacterial
activity differently in different water masses in the Southern Ocean. The ratio of bacterial to
primary production (BP:PP) was also lower (~0.05) in the Ross Sea and Western Arctic Ocean,
and exhibited a significant correlation with temperatures below 4°C. In our study BP:PP also
averaged about 0.05.

7 Rivkin et al. (1996) claimed that bacterial production and growth rates were not 8 intrinsically lower in cold oceans. They argued that empirically-derived conversion factors 9 specific to time and location resulted in BP estimates that were the same as in warmer waters. 10 The validity of this argument hinges on conversion factors (CF) having higher values in cold 11 waters; i.e., a fundamentally different relationship between leucine or thymidine incorporation 12 and cellular production at low temperatures. This argument has seldom been critically evaluated. 13 In the Ross Sea Ducklow et al (1999) found *lower* thymidine CF than the canonical value of 2 x 10¹⁸ cells mole⁻¹ (Fuhrman and Azam, 1980). The empirically-determined leucine CF was 1.5 14 kgC mole⁻¹, identical to the original value proposed by Simon and Azam (1989). This value was 15 16 used in Kirchman et al. (2009) and in the present treatment. Although we used a constant factor, 17 we recognize the possibility that systematic CF variability could change our results (Alonso-Sáez 18 et al., 2008; Kirchman et al., 1982; Morán et al., 2009).

We did not find a consistent relationship between temperature and leucine incorporation (Fig. 11). Although the average leucine incorporation rates tended to be lower in our study area than in warmer locations, incorporation rates can attain higher levels comparable to lower latitude systems. Thus in our dataset, the 50-meter integrated BP:PP was 0.1 or greater at 38 of 390 stations. Leucine incorporation rates exceeded 50 pmol l⁻¹ hr⁻¹ in most years (Fig. S3), even

1	in the coldest water (Fig. S4B). These higher rates are no different than peak rates in the
2	Equatorial Pacific Ocean and Arabian Sea, where surface water temperatures exceed 25°C
3	(Ducklow et al., 2001b; Kirchman et al., 1995).
4	In their analysis Kirchman et al (2009b) concluded that low bacterial production in cold
5	polar waters was a consequence of bottom-up factors, mainly the supply of dissolved organic
6	matter (DOM) to bacterial consumers. We found no specific evidence to support the Pomeroy
7	Hypothesis that bacteria are more sensitive to temperature when organic matter concentrations
8	were low (Table 3, Chl< 1-2 μ g L ⁻¹). Kirchman et al. (2009b) hypothesized a fundamental
9	difference in how carbon flows to bacteria in polar ecosystems. The result is limitation of BP by
10	labile organic matter availability. Our individual-year regressions for the shelf region (Table S3)

11

support this view.

12 4.4 Importance of bacterial production and long-term trends. Using a mean bacterial 13 conversion efficiency of 0.15 (Carlson et al., 1999; del Giorgio and Cole, 1998), a BP:PP value 14 of 0.05 implies that about 33% of the primary production flows through bacteria in the 15 Peninsular shelf region. Although this level of PP utilization is lower than other, mostly warmer 16 ocean regions, it still represents a substantial carbon flux in a system once believed to be 17 dominated by larger organisms such as diatoms and krill (Hart, 1934). In more recent times the 18 importance of microbial interactions has been recognized in Antarctic waters, overturning the old 19 diatom-krill-penguin/seal/whale paradigm that once governed most thinking about the region 20 (El-Sayed, 1988; Hewes, 1985; Karl et al., 1996).

Climate change appears to be progressively altering foodwebs from north to south along
 the Antarctic Peninsula, transforming them from diatom-krill-dominated to microbe-dominated
 systems (Sailley et al., 2011). The original support for this idea comes from observations of rapid

sea ice loss (Stammerjohn et al., 2008a; Stammerjohn et al., 2008c), declines of Adélie penguin
populations in the north (Ducklow et al., 2006a; Fraser and Ainley, 1986; Smith et al., 1999),
shifts in the dominant phytoplankton species from diatoms to cryptophytes (Moline *et al.*, 2004;
Moline *et al.*, 2008), and decadal-scale declines in phytoplankton and krill stocks in northern
regions (Atkinson *et al.*, 2004; Montes-Hugo *et al.*, 2009).

6 Sailley et al (2011) tested this climate migration hypothesis by incorporating twelve years 7 of Palmer LTER data (1995-2006) into an inverse foodweb model that satisfied various criteria 8 including observational constraints such as primary production, krill biomass and export levels 9 (Stukel and Landry, 2010). The model was used to seek solutions to the complete flow structure 10 of exchanges among organisms each year in northern and southern foodwebs. Their model 11 results estimated that even though BP:PP was lower (1.5-2.8%) than the observations reported 12 here, bacterial DOM utilization and respiration rates were equivalent to krill ingestion and 13 respiration rates across years and throughout the region. Furthermore, by evaluating the trophic indices of Legendre and Rassoulzadegan (1996) with model-generated data, they suggested that 14 15 the foodwebs of the Antarctic Peninsula region are changing from herbivore-dominated systems 16 with krill and diatoms toward microbial foodwebs dominated by smaller phytoplankton, bacteria 17 and microzooplankton. The apparent trend of increasing bacterial production over time (Fig. 9) is provocative but not significant ($R^2=0.41$, p=0.06, N=9). But further warming, sea ice loss and 18 19 ecosystem change seem inevitable in this region. The effects of climate change on bacterial 20 processes are poorly understood, but rapidly-evolving polar systems seem like the best places to 21 try and understand these important interactions.

22

1 **5.0 Conclusions.**

2 Our results show that as with other trophic levels, variations in bacterial production rates 3 reflect interannual and regional differences in sea ice retreat. Variability in rates of 3H-leucine 4 incorporation were best explained by chlorophyll, and to a lesser extent by PP, suggesting that 5 biomass accumulation and trophic exchange processes such as grazing, detritus turnover and 6 zooplankton excretion all contribute dissolved organic matter to supplement the direct release 7 from phytoplankton. These processes are dependent on the magnitude of phytoplankton blooms 8 which are regulated by the timing of sea ice retreat. These conclusions point out the dependence 9 of bacterial processes on large-scale climate variations and related physical-biological couplings, 10 factors not generally considered in microbial oceanography. 11 12 13 Acknowledgements. This research was supported by NSF Grants OPP-0217282 and 0823101 14 from the Antarctic Organisms and Ecosystems Program to HWD. We gratefully thank science 15 and logistics support personnel from Raytheon Polar Services and AGUNSA, and the Edison-16 Chouest Offshore officers and crews of ARSV Laurence M Gould for their skilled and dedicated

17 assistance. This research also benefitted from students and volunteers too numerous to name

18 here.

References.

- Alonso-Sáez, L. et al., 2008. Factors Controlling the Year-Round Variability in Carbon Flux Through Bacteria in a Coastal Marine System. Ecosystems, 11: 397-409.
- Arrigo, K.R., van Dijken, G. and Long, M., 2008. Coastal Southern Ocean: A strong anthropogenic CO2 sink. Geophysical Research Letters, 35(21): 6.
- Atkinson, A., Siegel, V., Pakhomov, E. and Rothery, P., 2004. Long-term decline in krill stock and increase in salps within the Southern Ocean. Nature, 432(7013): 100-103.
- Billen, G. and Becquevort, S., 1991. Phytoplankton-bacteria relationship in the Antarctic marine ecosystem. Polar Research, 10: 245-253.
- Bird, D.F. and Kalff, J., 1984. Empirical Relationships between Bacterial Abundance and Chlorophyll Concentration in Fresh and Marine Waters. Canadian Journal of Fisheries and Aquatic Sciences, 41(7): 1015-1023.
- Bird, D.F. and Karl, D.M., 1991. Spatial patterns of glutamate and thymidine assimilation in the Bransfield Strait, Antarctica, during and following the austral spring bloom. Deep-Sea Research, 38(8/9): 1057-1075.
- Bird, D.F. and Karl, D.M., 1999. Uncoupling of bacteria and phytoplankton during the austral spring bloom in Gerlache Strait, Antarctic Peninsula. Aquatic Microbial Ecology, 19(1): 13-27.
- Carlson, C.A., Bates, N.R., Ducklow, H.W. and Hansell, D.A., 1999. Estimation of bacterial respiration and growth efficiency in the Ross Sea, Antarctica. Aquatic Microbial Ecology, 19: 229-244.
- Church, M.J. et al., 2003. Abundance and distribution of planktonic Archaea and Bacteria in the waters west of the Antarctic Peninsula. Limnology and Oceanography, 48: 1893-1902.
- Clarke, A. et al., 2007. Climate change and the marine ecosystem of the western Antarctic Peninsula. Philosophical Transactions of the Royal Society B: Biological Sciences, 362(1477): 149-166.
- Cole, J.J., Findlay, S. and Pace, M.L., 1988. Bacterial production in fresh and saltwater ecosystems: a cross-system overview. Marine Ecology Progress Series, 43: 1-10.
- Cole, J.J., Likens, G.E. and Stryer, D.L., 1982. Photosynthetically produced dissolved organic carbon: an important carbon source for planktonic bacteria. Limnol. Oceanogr., 27: 1080-1090.
- Daniels, R.M., Ducklow, H.W. and Richardson, T.L., 2006. Food web structure and biogeochemical processes during oceanic phytoplankton blooms: An inverse model analysis. Deep-Sea Research II, 53: 532-54.
- del Giorgio, P.A. and Cole, J.J., 1998. Bacterial Growth Efficiency in Natural Aquatic Systems. Annual Review of Ecology and Systematics, 29: 503-541.
- Ducklow, H.W., 2003. Seasonal production and bacterial utilization of DOC in the Ross Sea, Antarctica. In: G. DiTullio (Editor), Biogeochemical Cycles in The Ross Sea. Antarctic Research Series. American Geophysical Union., Washington, DC, pp. 143-158.

- Ducklow, H.W., 2008. Long-term studies of the marine ecosystem along the west Antarctic Peninsula. Deep Sea Research II, 55(18-19): 1945-1948.
- Ducklow, H.W. et al., 2006a. Marine ecosystems: The West Antarctic Peninsula. Philosophical Transactions of the Royal Society of London B, 362: 67-94.
- Ducklow, H.W. et al., 2001a. The seasonal development of the bacterioplankton bloom in the Ross Sea, Antarctica, 1994-1997. Deep-Sea Research II, 48: 4199-4221.
- Ducklow, H.W. and Carlson, C.A., 1992. Oceanic bacteria production. In: K.C. Marshall (Editor), Advances in Microbial Ecology. Plenum Press, New York, pp. 113 181.
- Ducklow, H.W., Carlson, C.A. and Smith, W.O., Jr., 1999. Bacterial growth in experimental plankton assemblages and seawater cultures from the *Phaeocystis antarctica* bloom in the Ross Sea, Antarctica. Aquatic Microbial Ecology, 19: 215-227.
- Ducklow, H.W. et al., 2000. Constraining bacterial production, conversion efficiency and respiration in the Ross Sea, Antarctica, January-February, 1997. Deep-Sea Research II, 47(15-16): 3227-3247.
- Ducklow, H.W. et al., 2006b. Water column processes in the West Antarctic Peninsula and the Ross Sea: foodweb structure and interannual variability. Deep-Sea Research II, 53: 834-52.
- Ducklow, H.W. et al., 2001b. Heterotrophic bacterioplankton in the Arabian Sea: Basinwide response to year-round high primary productivity. Deep-Sea Research II, 48(6-7): 1303-1323.
- El-Sayed, S.Z., 1988. Productivity of the Southern Ocean: A Closer look. Comparative Biochemistry and Physiology, 90B: 489-98.
- Fraser, W.R. and Ainley, D.G., 1986. Ice edges and seabird occurrence in Antarctica. BioScience, 36: 258-263.
- Fuhrman, J.A. and Azam, F., 1980. Bacterioplankton secondary production estimates for coastal waters of British Columbia, Antarctica, and California. Applied and Environmental Microbiology, 39(6): 1085-1095.
- Fuhrman, J.A. et al., 2006. Annually reoccurring bacterial communities are predictable from ocean conditions. PNAS, 103(35): 13104-13109.
- Fukuda, R., Ogawa, H., Nagata, T. and Koike, I., 1998. Direct determination of carbon and nitrogen contents of natural bacterial assemblages in marine environments. Applied and Environmental Microbiology [Appl. Environ. Microbiol.]. 64(9): 3352-3358.
- Garneau, M.-E., S. Roy, C. Lovejoy, Y. Gratton, a. and Vincent, W.F., 2008. Seasonal dynamics of bacterial biomass and production in a coastal arctic ecosystem: Franklin Bay, western Canadian Arctic. Journal of Geophysical Research, 113(C07S91): doi:10.1029/2007JC004281.
- Garrison, D.L. et al., 2000. Microbial food web structure in the Arabian Sea: A U. S. JGOFS Study. Deep-Sea Research II, 47: 1387-1422.

- Gasol, J.M. and del Giorgio, P.A., 2000. Using flow cytrometry for counting natural planktonic bacteria and understanding the structure of planktonic bacterial communities. Scientia Marina, 64: 197-224.
- Grzymski, J.J. et al., submitted. Chemolithoautotrophy in Antarctic winter bacterioplankton. ISME Journal.
- Hart, T.J., 1934. On the phytoplankton of the south-west Atlantic and the Bellingshausen Sea, 1929-31. Discovery Reports, 8: 1-268.
- Hewes, C., D. O. Holm-Hansen, and E. Saukshaug, 1985. Alternate carbon pathways at lower trophic levels in the Antarctic food web. In: W.R. Siegfried, P.R.Condy and R.M. Laws (Editors), Antarctic Nutrient Cycles and Food Webs. Springer Verlag, pp. 277-283.
- Karl, D.M., 1993. Microbial processes in the southern oceans. In: I.E. Friedman (Editor), Antarctic Microbiology. Wiley-Liss, Inc, pp. 1-63.
- Karl, D.M., Christian, J.R., Dore, J.E. and Letelier, R.M., 1996. Microbiological oceanography in the region west of the Antarctic Peninsula: Microbial dynamics, nitrogen cycle and carbon flux. In: R.M. Ross, E.E. Hofmann and L.B. Quetin (Editors), Foundations for Ecological Research West of the Antarctic Peninsula. AGU Antarctic Research Series. American Geophysical Union, Washington, DC, pp. 303-332.
- Karl, D.M., Holm-Hansen, O., Taylor, G.T., Tien, G. and Bird, D.F., 1991. Microbial biomass and productivity in the western Bransfield Strait, Antarctica during the 1986-87 austral summer. Deep-Sea Research, 38(8/9): 1029-1055.
- Kirchman, D.L., Ducklow, H.W. and Mitchell, R., 1982. Estimates of bacterial growth from changes in uptake rates and biomass. Applied and Environmental Microbiology, 44: 1296-1307.
- Kirchman, D.L. et al., 2009a. Standing stocks, production, and respiration of phytoplankton and heterotrophic bacteria in the western Arctic Ocean. Deep Sea Research II, 56: 1237-1248.
- Kirchman, D.L., Moran, X.A.G. and Ducklow, H., 2009b. Microbial growth in the polar oceans role of temperature and potential impact of climate change. Nature Reviews of Microbiology, 7(6): 451-459.
- Kirchman, D.L., Rich, J.H. and Barber, R.T., 1995. Biomass and biomass production of heterotrophic bacteria along 140 degree W in the Equatorial Pacific: Effect of temperature on the microbial loop. Deep-Sea Research II, 42(2-3): 603-619.
- Leakey, R.J.G., Archer, S.D. and Grey, J., 1996. Microbial dynamics in coastal waters of East Antarctica: bacterial production and nanoflagellate bacterivory. Marine Ecology Progress Series, 142: 3-17.
- Legendre, L. and Rassoulzadegan, F., 1996. Food-web mediated export of biogenic carbon in oceans: hydrodynamic control. Marine Ecology Progress Series, 145: 179-93.
- Li, W.K.W., 2009. From cytometry to macroecology: a quarter century quest in microbial oceanography. Aquatic Microbial Ecology, 57(3): 239-251.

- Li, W.K.W., Head, E.J.H. and Glen Harrison, W., 2004. Macroecological limits of heterotrophic bacterial abundance in the ocean. Deep Sea Research Part I: Oceanographic Research Papers, 51(11): 1529-1540.
- Martinson, D.G., Stammerjohn, S.E., Iannuzzi, R.A., Smith, R.C. and Vernet, M., 2008. Western Antarctic Peninsula physical oceanography and spatio-temporal variability. Deep Sea Research II, 55(18-19): 1964-1987.
- Meredith, M.P. and King, J.C., 2005. Rapid climate change in the ocean west of the Antarctic Peninsula during the second half of the 20th century. Geophysical Research Letters, 32: doi:10.1029/2005GL024042.
- Moline, M.A., Claustre, H., Frazer, T.K., Schofield, O. and Vernet, M., 2004. Alteration of the food web along the Antarctic Peninsula in response to a regional warming trend. Global Change Biology, 10(12): 1973-1980.
- Moline, M.A. et al., 2008. High Latitude Changes in Ice Dynamics and Their Impact on Polar Marine Ecosystems. Annals of the New York Academy of Sciences, 1134: 267-319.
- Montes-Hugo, M. et al., 2009. Recent Changes in Phytoplankton Communities Associated with Rapid Regional Climate Change Along the Western Antarctic Peninsula. Science, 323(5920): 1470-1473.
- Morán, X.A.G., Calvo-Diaz, A. and Ducklow, H.W., 2009. Total and phytoplankton mediated bottom-up control of bacterioplankton change with temperature in NE Atlantic shelf waters. Aquatic Microbial Ecology, 58: 229-239.
- Morán, X.A.G. and Estrada, M., 2002. Phytoplanktonic DOC and POC production in the Bransfield and Gerlache Straits as derived from kinetic experiments of 14C incorporation. Deep Sea Research II, 49(4-5): 769-786.
- Morán, X.A.G., Gasol, J.M., Pédros-Alió, C. and Estrada, M., 2001. Dissolved and particulate primary production and bacterial production in offshore Antarctic waters during Austral summer: coupled or uncoupled? Marine Ecology Progress Series, 222: 25-39.
- Morris, R.M. et al., 2005. Temporal and Spatial Response of Bacterioplankton Lineages to Annual Convective Overturn at the Bermuda Atlantic Time-Series Study Site. Limnology and Oceanography, 50(5): 1687-1696.
- Pearce, I., Davidson, A.T., Bell, E.M. and Wright, S., 2007. Seasonal changes in the concentration and metabolic activity of bacteria and viruses at an Antarctic coastal site. AQUATIC MICROBIAL ECOLOGY, 47(1): 11-23.
- Peck, L.S., Barnes, D.K.A., Cook, A.J., Fleming, A.H. and Clarke, A., 2010. Negative feedback in the cold: ice retreat produces new carbon sinks in Antarctica. Global Change Biology, 16(9): 2614-2623.
- Pomeroy, L.R. and Deibel, D., 1986. Temperature regulation of bacterial activity during the spring bloom in Newfoundland coastal waters. Science (Washington), 233(4761): 359-361.
- Pomeroy, L.R. and Wiebe, W.J., 2001. Temperature and substrates as interactive limiting factors for marine heterotrophic bacteria. Aquatic Microbial Ecology, 23: 187-204.

- Rivkin, R.B., Anderson, M.R. and Lajzerowicz, C., 1996. Microbial processes in cold oceans. 1. Relationship between temperature and bacterial growth rate. Aquatic Microbial Ecology, 10: 243-254.
- Sailley, S. et al., 2011. Carbon fluxes and pelagic ecosystem dynamics around two Western Antarctic Peninsula Adélie penguin colonies: an inverse model approach. Limnology and Oceanography, Manuscript in review.
- Schofield, O. et al., 2010. How Do Polar Marine Ecosystems Respond to Rapid Climate Change? Science, 328: 1520-1523.
- Simon, M. and Azam, F., 1989. Protein content and protein synthesis rates of planktonic marine bacteria. Marine Ecology Progress Series, 51: 201-213.
- Simon, M., Glockner, F.O. and Amann, R., 1999. Different community structure and temperature optima of heterotrophic picoplankton in various regions of the Southern Ocean. Aquatic Microbial Ecology, 18(3): 275-284.
- Simon, M. and Rosenstock, B., 2007. Different coupling of dissolved amino acid, protein, and carbohydrate turnover to heterotrophic picoplankton production in the Southern Ocean in austral summer and fall. Limnology and Oceanography, 52(1): 85-95.
- Simon, M., Rosenstock, B. and Zwisler, W., 2004. Coupling of Epipelagic and Mesopelagic Heterotrophic Picoplankton Production to Phytoplankton Biomass in the Antarctic Polar Frontal Region. Limnology and Oceanography, 49(4): 1035-1043.
- Smetacek, V. and Nicol, S., 2005. Polar ocean ecosystems in a changing world. Nature (London), 437(7057): 362-368.
- Smith, D.C. and Azam, F., 1992. A simple, economical method for measuring bacterial protein synthesis rates in seawater using 3H-leucine. Marine Microbial Foodwebs, 6(2): 107-114.
- Smith Jr., W.O., Marra, J., Hiscock, M.R. and Barber, R.T., 2000. The seasonal cycle of phytoplankton biomass and primary productivity in the Ross Sea, Antarctica. Deep Sea Research II, 47(15-16): 3119-3140.
- Smith Jr., W.O. and Nelson, D.M., 1985. Phytoplankton bloom produced by a receding ice edge in the Ross Sea: spatial coherence with the density field. Science, 227: 163-166.
- Smith Jr., W.O. and Nelson, D.M., 1986. Importance of ice edge phytoplankton production in the Southern Ocean. Bioscience, 36(4): 251-257.
- Smith, R.C. et al., 1999. Marine Ecosystems sensitivity to historical climate change: Antarctic Peninsula. BioScience, 49(5): 393-404.
- Smith, R.C., Martinson, D.G., Stammerjohn, S.E., Iannuzzi, R.A. and Ireson, K., 2008. Bellingshausen and western Antarctic Peninsula region: Pigment biomass and sea-ice spatial/temporal distributions and interannual variability. Deep Sea Research II, 55(18-19): 1949-1963.
- Stammerjohn, S.E., Martinson, D.G., Smith, R.C. and Iannuzzi, R.A., 2008a. Sea ice in the western Antarctic Peninsula region: Spatio-temporal variability from ecological and climate change perspectives. Deep Sea Research II, 55(18-19): 2041-2058.

- Stammerjohn, S.E., Martinson, D.G., Smith, R.C. and Iannuzzi, S.A., 2008b. Sea ice in the western Antarctic Peninsula region: Spatio-temporal variability from ecological and climate change perspectives. Deep Sea Research II, 55: 2041-2058.
- Stammerjohn, S.E., Martinson, D.G., Smith, R.C., Yuan, X. and Rind, D., 2008c. Trends in Antarctic Annual Sea Ice Retreat and Advance and their Relation to ENSO and Southern Annular Mode Variability. Journal of Geophysical Research, 113(C03S90): doi: 10.1029/2007JC004269.
- Straza, T.R.A., Cottrell, M.T., Ducklow, H.W. and Kirchman, D.L., 2009. Geographic and phylogenetic variation in bacterial biovolume using protein and nucleic acid staining. Appl. Environ. Microbiol.: AEM.00183-09.
- Stukel, M.R. and Landry, M.R., 2010. Contribution of picophytoplankton to carbon export in the equatorial Pacific: A re-assessment of food-web flux inferences from inverse models. Limnology and Oceanography, 55: 2669-2685.
- Vaughan, D.G. et al., 2003. Recent rapid regional climate warming on the Antarctic Peninsula. Climatic Change, 60: 243-274.
- Vernet, M. et al., 2008. Primary production within the sea-ice zone west of the Antarctic Peninsula: I--Sea ice, summer mixed layer, and irradiance. Deep Sea Research II, 55: 2068-2085.
- Waters, K.J. and Smith, R.C., 1992. Palmer LTER: A sampling grid for the Palmer LTER program. Antarctic Journal of the United States, 27(5): 236-239.

Table 1. Regression statistics for yearly volumetric (discrete depth) chlorophyll-leucine (Chl-Leu) incorporation relationships. All regions are pooled for the annual data sets. Regression equations are Log (Leu) = mLog (Chl) + b. All regressions are significant at p<0.001 unless noted. The significance of the regression is given in the R² column if greater than 0.001. Regressions were calculated for the full water column (e.g., Fig. 7) and for the upper 50 meters alone.

Year / depths	Slope (m)	Intercept (b)	\mathbf{R}^2	n
2003 all	0.82	0.75	0.61	252
2003 upper 50	0.63	0.73	0.36	195
2004 all	0.62	1.20	0.37	216
2004 upper 50	0.54	1.21	0.34	193
2005 all	0.36	1.11	0.18	212
2005 upper 50	0.38	1.18	0.28	190
2006 all	0.56	1.05	0.43	212
2006 upper 50	0.48	1.10	0.32	195
2007 all	0.57	1.22	0.63	350
2007 upper 50	0.37	1.29	0.46	251
2008 all	0.64	1.02	0.50	323
2008 upper 50	0.64	1.10	0.49	236
2009 all	0.78	1.34	0.72	220
2009 upper 50	0.46	1.33	0.39	139
2010 all	0.73	0.88	0.45	212
2010 upper 50	0.15	1.02	0.02 (ns, p=0.054)	142
2011 all	0.57	1.21	0.58	251
2011 upper 50	0.45	1.32	0.40	195
All years & depths	0.67	1.08	0.52	2248
All years, upper 50	0.54	1.13	0.40	1737

Table 2. Regression statistics for yearly volumetric (discrete depth) primary production-leucine (PP-Leu) incorporation and PP-BP relationships. All regions are pooled for the annual data sets. Regression equations for PP-Leu and PP-BP are Log (Leu) = mLog (PP) + b; and BP = mPP + b, respectively. The slope of the *linear* BP regression indicates BP as a fraction of PP. All regressions are significant at p<0.001 unless noted.

Year	Slope (m)	Intercept (b)	\mathbf{R}^2	n
2003 PP-Leu	0.39	0.14	0.29	180
2003 PP-BP	0.007	0.07	0.30	180
2004 PP-Leu	0.41	0.72	0.32	190
2004 PP-BP	0.03	0.25	0.22	190
2005 PP-Leu	0.43	0.68	0.48	205
2005 PP-BP	0.01	0.41	0.61	205
2006 PP-Leu	0.37	0.75	0.55	201
2006 PP-BP	0.005	0.56	0.27	201
2007 PP-Leu	0.46	0.80	0.68	258
2007 PP-BP	0.006	0.61	0.39	258
2008 PP-Leu	0.48	0.64	0.49	243
2008 PP-BP	0.017	0.28	0.33	243
2009 PP-Leu	0.30	0.87	0.26	117
2009 PP-BP	0.010	0.46	0.17	117
2010 PP-Leu	0.30	0.81	0.30	131
2010 PP-BP	0.008	0.37	0.03, (p=0.026)	131
2011 PP-Leu	0.28	1.13	0.23	179
2011 PP-BP	0.001	1.43	0.01, (ns, p=0.077)	179
All years PP-Leu	0.43	0.69	0.39	1704
All years PP-BP	0.004	0.57	0.13	1704

Table 3. Temperature-leucine regression statistics for Shelf region, 2003-11. Discrete-depth (volumetric) relationships for <20 m samples. Regression equations are Log (Int Leu) = m (Temperature) + b. All regressions are significant at p<0.001 unless noted (p-value or ns). See also Figure 11.

Year / property	Slope	Intercept (b)	\mathbf{R}^2	n
2003 Discrete depth (<20)	-0.19	0.82	0.14 (p=0.002) ¹	56
2004 Discrete depth (<20)			p=0.71, ns	68
2005 Discrete depth (<20)	-0.13	1.25	0.08 (p=0.008)	72
2006 Discrete depth (<20)			p=0.39, ns	91
2007 Discrete depth (<20)			p=0.41, ns	75
2008 Discrete depth (<20)	0.26	0.72	0.07 (p=0.005)	85
2009 Discrete depth (<20)			p=0.72, ns	30
2010 Discrete depth (<20)	-0.33	1.15	0.18 (p=0.004)	40
2011 Discrete depth (<20)	0.22	1.40	0.20 (p=0.003)	39
All years			p=0.36, ns	556
All years, Chl<2			p=0.06, ns	383
All years, Chl <1			p=0.9, ns	239

Notes:

1. two measurements with Leucine < 0.1 excluded..

Figure Legends

- Figure 1. Map of Palmer LTER study region along the Antarctic Peninsula (white). Palmer Station E is 5 km south of Anvers Island, (green dot). Hydrographic lines are 100 km apart, north to south. The standard on to offshore hydrographic stations (yellow dots) are 20 km apart,. All stations >1000 m deep are Slope stations. Stations on the 200-600 lines < 1000 m deep are termed Shelf stations. The triangles and white circles show Coastal and Marguerite Bay stations, respectively. Stations <1000 m deep south of the 200 line and Marguerite Bay are called South stations (the 100, 000 and -100 lines), and were not occupied prior to 2008.</p>
- Figure 2. Climatological (mean) seasonal cycle of the water column (0-50 m) ³H-leucine incorporation rate at nearshore Station E, Palmer Station, Antarctica. Observations from February 2003 March 2011 (excluding the 2006-07 and 2007-08 field seasons) were binned into 15-day intervals for this plot. The grey-shaded box indicates the period of the annual summer regional survey cruise (ca. Jan 01 Feb 06). Error bars are standard errors of the mean.
- Figure 3. Temperature salinity (T-S) plot for the Shelf region of the Palmer LTER study area along the western Antarctic Peninsula (see Figure 1 for location), summer 2003-11. Each symbol is one discrete depth from a CTD cast at a sampling station. The upper and lower blue boxes are Upper Circumpolar Deep Water (UCDW) and Winter Water (WW), respectively. The dotted line is the freezing point of seawater.
- **Figure 4.** Vertical distribution of temperature (A,C, degrees C) and salinity (B,D) in the Palmer LTER study region in 2005 (late sea ice retreat) and 2009 (early retreat), showing vertical, interannual and regional variability. Regions as in Figure 1. The lines indicate the 2003-11 mean and 95% confidence intervals.
- **Figure 5.** Vertical distribution of chlorophyll (A, C; mg m⁻³) and primary production rates (B,D; mgC m⁻³ d⁻¹) in the Palmer LTER study region in 2005 (late sea ice retreat) and 2009 (early retreat), showing vertical, interannual and regional variability. Regions as in Figure 1. The lines indicate the 2003-11 mean and 95% confidence intervals.
- **Figure 6.** Vertical profiles of ³H-leucine incorporation rates (A,C; pmol l⁻¹ hr⁻¹) and bacterial abundance (B,D; cells l⁻¹) in the Palmer LTER study region in 2005 (late sea ice retreat) and 2009 (early retreat), showing vertical, interannual and regional variability. Regions as in Figure 1. The lines indicate the 2003-11 mean and 95% confidence intervals.
- **Figure 7.** Relationships between discrete depth chlorophyll concentrations (A,C) or PP rates (B,D) with ³H-leucine incorporation rates in 2005 (late sea ice retreat) and 2009 (early retreat). Regions as in Figure 1. See Tables 1,2 for regression statistics.
- Figure 8. Water column integral chlorophyll and primary production rates (both through euphotic zone to 1% of surface irradiance), bacterial abundance and leucine incorporation rates (both to 50 meters). Years 2003-11 pooled for regional comparisons. Regions sharing letters are not significantly different (Tukey Post hoc HSD tests; p<0.05). Number of stations is given inside bars.</p>

Figure Legends, continued.

- **Figure 9.** Time series of phytoplankton and bacterial stocks and production rates for the shelf region, 2003-11. See Table S1 for stations sampled in each year. Years sharing letters are not significantly different (Tukey Post hoc HSD tests; p<0.05). Number of stations is given inside bars.
- Figure 10. Regional (A,B) and decadal (C,D) scale relationships among chlorophyll, primary and bacterial production rates. See Figures 8.9 for source data. All years 2003-11 are pooled in each region in panels A, B. Panels C,D are for Shelf region only. Dashed lines and coefficients of determination are given for significant regressions (p<0.05). Regions in A,B as in previous figures (SL-slope; Sh-shelf; C-coastal; MB-Marguerite Bay; S-south). Numbers on panels C,D are years.
- **Figure 11.** Relationships between discrete depth temperature and ³H-leucine incorporation rates in the Shelf region for the upper 20 meters. Regressions are significant (p<0.05) where lines are depicted. See Table 3 for regression statistics.
- Figure 12. Maps of annual anomalies of the date of sea ice retreat in the PAL study region, 2003-2011 (200-600 lines). Each map shows the anomaly in days relative to the mean date of ice retreat shown in the map at the bottom right corner. Negative anomalies are earlier retreats, positive anomalies are late retreats. In these plots, the year is defined from the March-July ice advance to the following year's Sept-January retreat period. Thus ice year 2002 includes the January 2003 bacterial and other observations, and so on. See Stammerjohn et al. (2008) for details.

Figure 1 Click here to download high resolution image







Figure 4 Click here to download high resolution image



Figure 5 Click here to download high resolution image



Figure 6 Click here to download high resolution image



Figure 7 Click here to download high resolution image



Fig 8





Fig 9

Figure 10 Click here to download high resolution image



Figure 11 Click here to download high resolution image



Figure 12 Click here to download high resolution image



Region or Station	2003	2004	2005	2006	2007	2008	2009	2010	2011	Total
Shelf		1 1 1			1 1 1	1 1 1			1 1 1	
600.040	1	nd	1	1	1	2	1	1	2	10
600.060	2	1	nd	1	1	1				6
600.080	nd	1	1	1	1	1				5
600.100	1	1	nd	1	1	1		1	1	7
600.120	1	1	1	1	1	1			, , ,	6
600.140	1	1	1	1	1	1				6
600.160	nd	1	1	1	1	1				5
585.131					j		1			1
585.135		, ,	, , ,	, , ,	, , ,		, , ,	4	, , ,	4
590.130			, ,				2			2
500.060	1	1	nd	1	1	1		, , , ,	1	6
500.080	1	1	1	1	1	1				6
500.100	1	nd	1	1	1	1		1	 	6
500.120	1	nd	1	1	1	1				5
500.140	1	1	1	1	1	1			¦	6
500.160	1	1	1	1	1	1	1			7
500.180	nd	1	1	1	1	1				5
400.040	nd	1	nd	1	1	1	1	1	1	7
400.060	1	1	1	1	1	1				6
400.080	1	1	1	1	1	1		, , ,	; ;	6
400.100	1	1	1	1	1	1	1	1	1	9
400.120	1	1	1	1	1	1				6
400.140	1	1	1	1	1	1				6
400.160	1	1	1	1	nd	1	1	1	¦ {	7
460.046							1			1
300.040	1	1	nd	1	1	1	1	1	1	8
300.060	1	1	1	1	1	1				6
300.080	1	1	1	1	1	1			<u> </u>	6
300.100	1	1	1	1	1	1	1	1	1	9
300.120	nd	nd	1	1	1	1	1	1		6

Supplementary Table S1. Number of hydrostations sampled for bacterial properties in PAL-LTER study region 2003-2011. A blank entry means the station was not occupied in that year. Nd indicates bacterial samples were not collected. See Figure 1 for station locations. Sampling regions are defined in Methods section

Supplementary Table S1, continued										
Region or Station	2003	2004	2005	2006	2007	2008	2009	2010	2011	Total
Shelf, cont'd					<u></u>					
300.140	nd	1	1	1	1	1				5
300.160	1	nd	1	1	1	1	1	1		7
200.040	1	1	1	1	1	1		1	1	8
200.060	1	1	nd	1	1	1	1		; ;	6
200.080	1	1	1	1	1	1	<u> </u>			6
200.100	1	1	1	1	1	1		1	1	8
200.120	1	1	1	1	1	1	}		 	6
200.140	1	1	1	1	1	1				6
Total	29	29	28	34	33	35	14	16	10	228
Oceanic					<u> </u>		<u> </u>			
600.180	1	nd	1	1	1	1	1			6
600.200	1	1	1	1	1	1		1	1	8
600.220	nd	1	1	1	1	1			; ;	5
600.240	1	1	1		<u> </u>	1	}		<u> </u>	4
600.260	1	1	1		1					4
500.200	1	1	1	1	1	1	1	1	1	9
500.220	1	nd	1	1	1					4
500.240	1									1
500.260										0
400.180	1	nd	1	1	1	1]			5
400.200	1	nd	1	1	1	1				5
300.180	1	1	1	1	1	1]			6
300.200	1	nd	1	1	1	1				5
200.160	1	1	1	1	1	1	1	1	1	9
200.180	1	1	1	1	1	1				6
200.200	1	1	1	1	1	1]			6
200.220				nd	1	1				2
200.240				nd	1	1				2
200.260				1	1	1			,	3
000.120								1		1
000.180							 		1	1

Supplementary Tabl	e S1, contir	nued								
Region or Station	2003	2004	2005	2006	2007	2008	2009	2010	2011	Total
Ocean, cont'd		ļ	 	ļ	<u> </u>	ļ	<u> </u>	l 	 	
-100.160							1			1
-100.180									1	1
Total	14	9	14	13	16	15	4	4	5	94
Marguerite Bay										
200.020	1	1	1	1	1	1				6
200.000	1	1	1	1	1	1	1		1	8
200010					¦		 		1	1
200020	1	1	nd	1	1	1			1	6
200040	nd	1	1	nd	1	1	 	1	1	6
200060					1					1
200080			1		<u></u>				<u></u>	1
167033		, , ,	, , ,	, ,			6		, , ,	6
168.030	, ,		; ;					4	; ;	4
191.019			: ; ;		: ; ;	; 	; ;	;	1	1
203.031									1	1
208.015			 		¦		¦	ļ	1	1
212020									1	1
213001				ļ		ļ		ļ	1	1
222029									1	1
Total	3	4	4	3	5	4	7	5	10	45
Southern										
-100.000			<u></u>		<u></u>		5	1	1	7
-100.060	, , ,	, ,	, , ,						1	1
-114024	; ; ;		, ,						1	1
-120024			¦ 		¦		¦	1	¦ 	1
-144021									1	1
-158034			 					ļ 	1	1
000.000								1		1
000.040			¦		¦	[¦	1	¦	1
000.120								1		1
100060						1				1

Supplementary Table S1, continued										
Region or Station	2003	2004	2005	2006	2007	2008	2009	2010	2011	Total
South, cont'd							j			
100040			¦		¦		<u> </u>	<u> </u>	1	1
100.000			; 		1		1	1	<u>.</u>	3
100.020					1					1
100.040			, ,		1			1	1	3
100.060					1		1			2
100.100			, , ,		, , , ,		1	1	1	3
116047						1				1
Total	0	0	0	0	4	2	8	8	8	30
Coastal]					
550.000	1		1	1	1					4
550.020		1								1
585.010	1	1				1	}		 	3
Total	2	2	1	1	1	1	0	0	0	8
Total, all regions	48	44	47	51	59	57	33	33	33	405

Supplementary Table S2. Regression statistics for regional-scale relationships among volumetric properties (discrete depth samples). All years are pooled for each region unless noted. Regression equations are log-log except unless noted*. The slope of the PP-BP regression indicates BP as a fraction of PP. All regressions are significant at p<0.001 unless noted. Superscripts refer to depth range for each regression (1-all depths, 2-euphotic zone, 3-upper 50 m).

Region	Slope	Intercept (b)	\mathbf{R}^2	n
Shelf Chl-Leu ¹	0.53	1.13	0.29	925
Shelf Leu-PP ²	0.39	0.72	0.36	935
*Shelf BP-PP ²	0.007	0.48	0.19	935
Ocean Chl-Leu ¹	0.72	1.17	0.39	324
Ocean Leu-PP ²	0.44	0.56	0.33	449
*Ocean BP-PP ²	0.016	0.20	0.25	449
Marg Bay Chl-Leu ¹	0.37	1.20	0.28	249
Marg Bay Leu-PP ²	0.23	1.06	0.20	192
*Marg Bay BP-PP ²	0.001	1.03	0.05 (P=0.0011)	192
Coast Chl-Leu ¹	0.98	1.14	0.48	30
Coast Leu-PP ²	0.57	0.56	0.52	18
*Coast BP-PP ²	0.018	0.47	0.28 (p=0.014)	18
South Chl-Leu ¹	0.46	1.27	0.27	149
South Leu-PP ²	0.47	0.80	0.41	110
*South BP-PP ²	0.012	0.88	0.41	110
*Coastal Chl-Temp ³	1.011	-0.144	0.28	96
*Coastal PP-Temp ³	-45.3	103	0.18	83
*Coastal PP-Salt ³	-95	3210	0.40	83
*Marg Bay Leu-Temp ³	15.5	30.9	0.20	169
*South Leu-Salt ³	-40.7	1403	0.36	89
*Shelf Leu-Salt ³	-30.3	1041	0.21	558
*2011 Leu-Temp	24.6	26.8	0.36	108

Supplementary Table S3. Chlorophyll-leucine regression statistics for Shelf region, 2003-11. Discrete-depth (volumetric) relationships for <20 m samples and integrated (upper 50 m). Regression equations are Log (Leu) = mLog (Chl) + b or Log (Int Leu) = mLog (Int Chl) + b. All regressions are significant at p<0.001 unless noted (p-value or ns).

Year / property	Slope	Intercept (b)	\mathbf{R}^2	n
2003 Discrete depth (<20)	0.55	0.72	0.21	58
2003 Integral (0-50)	1.26	0.39	0.25 (p=0.007)	28
2004 Discrete depth (<20)	0.59	1.22	0.32	62
2004 Integral (0-50)	0.62	1.80	0.26 (p=0.004)	30
2005 Discrete depth (<20)	0.27	1.15	0.07 (p=0.020)	62
2005 Integral (0-50)	0.37	2.15	0.18 (p=0.027)	27
2006 Discrete depth (<20)	0.18	1.39	0.05 (p=0.044)	86
2006 Integral (0-50)			p=0.81 (ns)	34
2007 Discrete depth (<20)	0.33	1.36	0.31	73
2007 Integral (0-50)			p=0.19 (ns)	33
2008 Discrete depth (<20)	0.3	1.09	0.40	80
2008 Integral (0-50)	0.69	1.60	0.25 (p=0.002)	35
2009 Discrete depth (<20)	0.32	1.35	0.15 (p=0.041)	28
2009 Integral (0-50)			p=0.50 (ns)	14
2010 Discrete depth (<20)	0.40	1.04	0.32	41
2010 Integral (0-50)			p=0.40 (ns)	16
2011 Discrete depth (<20)			0.03 (p=0.15, ns)	31
2011 Integral (0-50)			p=0.42 (ns)	11
All years, Discrete (<20)	0.55	1.15	0.34	521
All years, Integral (0-50)	0.51	1.92	0.22	221

Supplementary Table S4. Regression statistics for regional integrated (upper 50 m or euphotic zone) chlorophyll-leucine (Chl-Leu) incorporation and PP-leucine relationships. All years are pooled for these decadal-scale analyses. Regression equations are Log (Int Leu) = mLog (Int Chl or PP) + b. All regressions are significant at p<0.001 unless noted (ns).

Region / property	Slope	Intercept (b)	\mathbf{R}^2	n
Shelf PP-LEU	0.20	2.22	0.08	225
Shelf Chl-LEU	0.57	1.83	0.26	224
Ocean PP-LEU	0.39	1.57	0.17	93
Ocean Chl-LEU	0.86	1.32	0.31	93
Marg Bay PP-LEU	0.21	2.28	0.08	41
Marg Bay Chl-LEU	0.29	2.41	0.14	46
Coast PP-LEU			(ns)	5
Coast Chl-LEU			(ns)	5
South PP-LEU			(ns)	25
South Chl-LEU			(ns)	28
Shelf, ocean, MB PP-Leu	0.32	1.85	0.21	359
Shelf, ocean, MB Chl-LEU	0.60	1.75	0.36	363
All regions, PP-LEU	0.05	703	0.05	389
All regions, Chl-LEU	0.59	1.78	0.34	396



Supplemental Figure S1. Relationships among temperature, salinity and primary production rates. A, B: all regions. C,D: coastal region only. All years are merged within each region indicated. These plots only include depths less than 20 meters, to exclude the influence of depth-related factors. Symbols as in Figure 4. The relationships shown by dashed lines in the coastal region (C,D) are significant at p<0.05. See Table S2 for regression statistics.



Supplemental Figure S2. Vertical profiles of picoplankton (i.e., "bacterial") abundance in the Palmer LTER study region, 2003-11, showing vertical, interannual and regional variability. Regions as in Figure 1. The lines indicate the 2003-11 mean and 95% confidence intervals.



Supplemental Figure S3. Vertical profiles of ³H-leucine incorporation rates in the Palmer LTER study region, 2003-11, showing vertical, interannual and regional variability. Regions as in Figure 1. The lines indicate the 2003-11 mean and 95% confidence intervals.



Supplemental Figure S4. Relationships between leucine incorporation, temperature and salinity. A. Marguerite Bay, all years. C. Shelf, all years. B, D. All years and regions. These plots only include depths less than 20 meters, to exclude the influence of depth-related factors. Symbols as in previous plots. The relationships shown by dashed lines in A, C are significant at p<0.05. See Supplementary Table 2 for regression statistics.