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1 The Minimum information about a marker gene sequence (MIMARKS) and minimum 2 information about any (x) sequence (MIxS) specifications 3 Pelin Yilmaz^{1,2}, Renzo Kottmann¹, Dawn Field³, Rob Knight^{4,5}, James R. Cole^{6,7}, Linda 4 Amaral-Zettler⁸, Jack A. Gilbert^{9,10,11}, Ilene Karsch-Mizrachi¹², Anjanette Johnston¹², 5 Guy Cochrane¹³, Robert Vaughan¹³, Christopher Hunter¹³, Joonhong Park¹⁴, Norman 6 Morrison^{3,15}, Philippe Rocca-Serra¹⁶, Peter Sterk³, Manimozhiyan Arumugam¹⁷, Mark 7 Bailey³, Laura Baumgartner¹⁸, Bruce W. Birren¹⁹, Martin J. Blaser²⁰, Vivien Bonazzi²¹, 8 Tim Booth³, Peer Bork¹⁷, Frederic D. Bushman²², Pier Luigi Buttigieg^{1,2}, Patrick S. G. 9 Chain^{7,23,24}, Emily Charlson²², Elizabeth K. Costello⁴, Heather Huot-Creasy²⁵, Peter 10 Dawyndt²⁶, Todd DeSantis²⁷, Noah Fierer²⁸, Jed A. Fuhrman³⁰, Rachel E. Gallery³¹, Dirk 11 Gevers¹⁹, Richard A. Gibbs^{32,33}, Inigo San Gil³⁴, Antonio Gonzalez³⁵, Jeffrey I. Gordon³⁶, 12 Robert Guralnick^{28,29}, Wolfgang Hankeln^{1,2}, Sarah Highlander^{32,37}, Philip Hugenholtz³⁸, 13 Janet Jansson^{23,39}, Andrew L. Kau³⁶, Scott T. Kelley⁴⁰, Jerry Kennedy⁴, Dan Knights³⁵, 14 Omry Koren⁴¹, Justin Kuczynski¹⁸, Nikos Kyrpides²³, Robert Larsen⁴, Christian L. 15 Lauber⁴², Teresa Legg²⁸, Ruth E. Ley⁴¹, Catherine A. Lozupone⁴, Wolfgang Ludwig⁴³, 16 Donna Lyons⁴², Eamonn Maguire¹⁶, Barbara A. Methé⁴⁴, Folker Meyer¹⁰, Brian 17 Muegge³⁶, Sara Nakielny⁴, Karen E. Nelson⁴⁴, Diana Nemergut⁴⁵, Josh D. Neufeld⁴⁶, 18 Lindsay K. Newbold³, Anna E. Oliver³, Norman R. Pace¹⁸, Giriprakash Palanisamv⁴⁷. 19 Jörg Peplies⁴⁸, Joseph Petrosino^{32,37}, Lita Proctor²¹, Elmar Pruesse^{1,2}, Christian Quast¹, 20 Jeroen Raes⁴⁹, Sujeevan Ratnasingham⁵⁰, Jacques Ravel²⁵, David A. Relman^{51,52}, Susanna 21 Assunta-Sansone¹⁶, Patrick D. Schloss⁵³, Lynn Schriml²⁵, Rohini Sinha²², Michelle I. 22 Smith³⁶, Erica Sodergren⁵⁴, Aymé Spor⁴¹, Jesse Stombaugh⁴, James M. Tiedje⁷, Doyle V. 23

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<u> </u>	maia ,	George MI.	Weinstock	, Doug	wenuer,	Owen	winte	, marcw	v interey	,

- 25 Andreas Wilke¹⁰, Jennifer R. Wortman²⁵, Tanya Yatsunenko³⁶, Frank Oliver Glöckner^{1,2}
- 26
- 27
- 28 1 Microbial Genomics and Bioinformatics Group, Max Planck Institute for Marine
- 29 Microbiology, D-28359 Bremen, Germany
- 30 2 Jacobs University Bremen gGmbH, D-28759 Bremen, Germany
- 31 3 Natural Environment Research Council Environmental Bioinformatics Centre,
- 32 Wallington CEH, Oxford OX10 8BB, UK
- 33 4 Department of Chemistry and Biochemistry, University of Colorado, Boulder, Colorado
- 34 80309, USA
- 35 5 Howard Hughes Medical Institute, San Francisco, California 94143-2208, USA
- 36 6 Ribosomal Database Project, Michigan State University, East Lansing, Michigan
- 37 48824-4320, USA
- 38 7 Center for Microbial Ecology, Michigan State University, East Lansing, Michigan
- 39 48824-1325, USA
- 40 8 The Josephine Bay Paul Center for Comparative Molecular Biology and Evolution,
- 41 Marine Biological Laboratory, Woods Hole, Massachusetts, USA
- 42 9 Plymouth Marine Laboratory, Plymouth PL1 3DH, UK
- 43 10 Mathematics and Computer Science Division, Argonne National Laboratory,
- 44 Argonne, Illinois 60439, USA
- 45 11 Department of Ecology and Evolution, University of Chicago, Chicago, Illinois
- 46 60637, USA
- 47 12 National Center for Biotechnology Information (NCBI), National Library of

- 48 Medicine, National Institutes of Health, Bethesda, Maryland 20894, USA
- 49 13 European Molecular Biology Laboratory (EMBL) Outstation, European
- 50 Bioinformatics Institute (EBI), Wellcome Trust Genome Campus, Hinxton, Cambridge
- 51 CB10 1SD, UK
- 52 14 School of Civil and Environmental Engineering, Yonsei University, Seoul 120-749,
- 53 Republic of Korea
- 54 15 School of Computer Science, University of Manchester, Manchester M13 9PL, UK
- 55 16 Oxford e-Research Centre, University of Oxford, Oxford OX1 3QG, UK
- 56 17 Structural and Computational Biology Unit, European Molecular Biology Laboratory,
- 57 D-69117 Heidelberg, Germany
- 58 18 Department of Molecular, Cellular and Developmental Biology, University of
- 59 Colorado, Boulder, Colorado 80309, USA
- 60 19 Broad Institute of Massachusetts Institute of Technology and Harvard University,
- 61 Cambridge, Massachusetts 02142, USA
- 62 20 Department of Medicine and the Department of Microbiology, New York University
- 63 Langone Medical Center, New York 10017, USA
- 64 21 National Human Genome Research Institute, National Institutes of Health, Bethesda,
- 65 Maryland 20892, USA
- 66 22 Department of Microbiology, University of Pennsylvania School of Medicine,
- 67 Philadelphia 19104, USA
- 68 23 DOE Joint Genome Institute, Walnut Creek, California 94598, USA
- 69 24 Los Alamos National Laboratory, Bioscience Division, Los Alamos, New Mexico,
- 70 USA

- 71 25 Institute for Genome Sciences, University of Maryland School of Medicine,
- 72 Baltimore, Maryland 21201, USA
- 73 26 Department of Applied Mathematics and Computer Science, Ghent University, 9000
- 74 Ghent, Belgium
- 75 27 Center for Environmental Biotechnology, Lawrence Berkeley National Laboratory,
- 76 Berkeley, California, USA
- 28 Department of Ecology and Evolutionary Biology, University of Colorado, Boulder,
- 78 Colorado 80309, USA
- 79 29 University of Colorado Museum of Natural History, University of Colorado, Boulder,
- 80 Colorado 80309, USA
- 81 30 Department of Biological Sciences, University of Southern California, Los Angeles,
- 82 California 90089, USA
- 83 31 National Ecological Observatory Network (NEON), Boulder, Colorado 80301, USA
- 84 32 Human Genome Sequencing Center, Baylor College of Medicine, Houston, Texas
- 85 77030, USA
- 86 33 Department of Molecular and Human Genetics, Baylor College of Medicine, Houston,
- 87 Texas 77030, USA
- 88 34 Department of Biology, University of New Mexico, LTER Network Office,
- 89 Albuquerque, New Mexico 87131, USA
- 90 35 Department of Computer Science, University of Colorado, Boulder, Colorado 80309,
- 91 USA
- 92 36 Center for Genome Sciences and Systems Biology, Washington University School of
- 93 Medicine, St. Louis, Missouri 63108, USA

- 94 37 Department of Molecular Virology and Microbiology, Baylor College of Medicine,
- 95 Houston, Texas 77030, USA
- 96 38 Australian Centre for Ecogenomics, School of Chemistry and Molecular Biosciences,
- 97 The University of Queensland, Brisbane QLD 4072, Australia
- 98 39 Earth Science Division, Lawrence Berkeley National Laboratory, Berkeley,
- 99 California, USA
- 100 40 Department of Biology, San Diego State University, San Diego, California 92182-
- 101 4614, USA
- 102 41 Department of Microbiology, Cornell University, Ithaca, New York 14853, USA
- 103 42 Cooperative Institute for Research in Environmental Sciences, University of Colorado,
- 104 Boulder, Colorado 80302, USA
- 105 43 Lehrstuhl für Mikrobiologie, Technische Universität München, D-853530 Freising,
- 106 Germany
- 107 44 J. Craig Venter Institute, Rockville, Maryland 20850-3213, USA
- 108 45 Department of Environmental Sciences, University of Colorado, Boulder, Colorado
- 109 80309, USA
- 110 46 Department of Biology, University of Waterloo, Ontario, N2L 3G1, Canada
- 111 47 Environmental Sciences Division, Oak Ridge National Laboratory, Oak Ridge,
- 112 Tennessee 37830-8020, USA
- 113 48 Ribocon GmbH, D-28359 Bremen, Germany
- 114 49 VIB Vrije Universiteit Brussel, 1050 Brussels, Belgium
- 115 50 Canadian Centre for DNA Barcoding, Biodiversity Institute of Ontario, University of
- 116 Guelph, Guelph, Ontario N1G 2W1, Canada

117	51 Departments of Microbiology and Immunology and of Medicine, Stanford University
118	School of Medicine, Stanford, California 94305, USA
119	52 Veterans Affairs Palo Alto Health Care System, Palo Alto, California 94304, USA
120	53 Department of Microbiology and Immunology, Ann Arbor, Michigan 48109-5620,
121	USA
122	54 The Genome Center, Department of Genetics, Washington University in St. Louis
123	School of Medicine, St. Louis, Missouri 63108, USA
124	
125	
126	Here we present a standard developed by the Genomic Standards Consortium
127	(GSC) to describe marker gene sequences—the minimum information about a
128	marker gene sequence (MIMARKS). We also introduce a system for describing the
129	environment from which a biological sample originates. The "environmental
130	packages" apply to any sequence whose origin is known and can therefore be used
131	in combination with MIMARKS or other GSC checklists. Finally, to establish a
132	unified standard for describing sequence data and to provide a single point of entry
133	for the scientific community to access and learn about GSC checklists, we establish
134	the minimum information about any (x) sequence (MIxS). Adoption of MIxS will
135	enhance our ability to analyze natural genetic diversity across the Tree of Life as it
136	is currently being documented by massive DNA sequencing efforts from myriad
137	ecosystems in our ever-changing biosphere.

139	CBOL: Consortium for the Barcode of Life
140	COI: cytochrome c oxidase I
141	DDBJ: DNA DataBank of Japan
142	DOI: Digital Object Identifier
143	DRA: DDBJ Sequence Read Archive
144	ENA: European Nucleotide Archive
145	EnvO: Environment Ontology
146	GAZ: Gazetteer
147	GCDML: Genomic Contextual Data Markup Language
148	GSC: Genomic Standards Consortium
149	ICoMM: International Census of Marine Microbes
150	INSDC: International Nucleotide Sequence Database Collaboration
151	ISA: Investigation/Study/Assay Infrastructure
152	ISO: International Organization for Standardization
153	MICROBIS: The Microbial Oceanic Biogeographic Information System
154	MIMARKS: Minimum Information about a MARKer Gene Sequence
155	MIGS/MIMS: Minimum Information about a Genome/Metagenome Sequence
156	MIRADA-LTERs: Microbial Inventory Research Across Diverse Aquatic Long Term
157	Ecological Research Sites

- 158 OBO: Open Biological and Biomedical Ontologies
- 159 PMID: Pubmed ID

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Abbreviations

- 160 RDP: Ribosomal Database Project
- 161 *rRNA*: ribosomal RNA
- 162 SI: International System of Units
- 163 SRA: Sequence Read Archive

- 164 SSU: small subunit
- 165 URL: Uniform Resource Locator
- 166 WGS84: World Geodetic System 84
- 167 XML Schema: Extensible Markup Language Schema
- 168

169 Without specific guidelines, most genomic, metagenomic and marker gene 170 sequences in databases are sparsely annotated with the information required to guide data 171 integration, comparative studies and knowledge generation. Even with complex keyword 172 searches, it is currently impossible to reliably retrieve sequences that have originated 173 from certain environments or particular locations on Earth-for example, all sequences 174 from "soil" or "freshwater lakes" in a certain region of the world. Since public databases 175 of the International Nucleotide Sequence Database Collaboration (INSDC; comprising 176 DNA Data Bank of Japan (DDBJ), European Nucleotide Archive (EBI-ENA) and 177 GenBank (http://www.insdc.org)) depend on author-submitted information to enrich the 178 value of sequence datasets, we argue that the only way to change the current practice is to 179 establish a standard of reporting that requires contextual data to be deposited at the time 180 of sequence submission. The adoption of such a standard would elevate the quality, 181 accessibility, and utility of information that can be collected from INSDC and the eco-182 system of other biological resources. 183 The GSC has previously proposed standards for describing genomic sequences, 184 the "minimum information about a genome sequence" (MIGS), and metagenomic 185 sequences, the "minimum information about a metagenome sequence" (MIMS)¹. Here 186 we introduce an extension of these standards for capturing information about marker 187 genes, MIMARKS. Additionally, we introduce "environmental packages" that 188 standardize sets of measurements and observations describing particular habitats that are 189 applicable across all GSC checklists and beyond². We define "environment" as any 190 location in which a sample or organism is found, e.g., soil, air, water, human-associated, 191 plant-associated, or laboratory. The original MIGS/MIMS checklists included contextual

data about the location from which a sample was isolated and how the sequence data was
produced. However, standard descriptions for a more comprehensive range of
environmental parameters, which would help to better contextualize a sample, were not
included. The environmental packages presented here are relevant to any genome
sequence of known origin, and would usefully be combined with many projects described
by MIGS, MIMS or MIMARKS.
To create a single entry point to all minimum information checklists from the

199 GSC and to the environmental packages, we propose an overarching framework, the

200 MIxS standard [AU: ADD URL]. MIxS is a new standard that includes the technology-

201 specific checklists from the previous MIGS and MIMS standards, provides a way of

202 introducing additional checklists such as MIMARKS, and also allows annotation of

203 sample data using environmental packages. A schematic overview of MIxS along with

the MIxS environmental packages is shown in **Figure 1**.

205

206 The development of MIMARKS and the environmental packages

207 Over the past three decades, the 16S rRNA, 18S rRNA and internal transcribed 208 spacer gene sequences (ITS) from Bacteria, Archaea, and microbial Eukaryotes have provided deep insights into the topology of the tree of life^{3, 4} and the composition of 209 210 communities of organisms that live in diverse environments, which range from deep sea hydrothermal vents to ice sheets in the Arctic⁵⁻¹⁶. Numerous other phylogenetic marker 211 212 genes have also proven useful, including RNA polymerase subunits (*rpoB*), DNA gyrases (gyrB), DNA recombination and repair proteins (*recA*) and heat shock proteins (*HSP70*)³. 213 214 Marker genes can also reveal key metabolic functions rather than phylogeny; examples

include nitrogen cycling (amoA, nifH, ntcA)^{17, 18}, sulfate reduction $(dsrAB)^{19}$ or 215 phosphorus metabolism (*phnA*, *phnI*, *phnJ*)^{20, 21}. In this paper we collectively define all of 216 217 these different phylogenetic and functional genes (or gene fragments) as "marker genes" 218 as they are used to profile natural genetic diversity across the Tree of Life, and argue that 219 a small amount of additional effort invested in describing them with specific guidelines in 220 our public databases will revolutionize the study types that can be performed with these 221 large data resources. This effort is timely, given the need to determine how climate 222 change and various other anthropogenic perturbations of our biosphere are affecting 223 biodiversity, and how marked changes in our cultural traditions and lifestyles are 224 affecting human microbial ecology, and, ultimately, human health.

MIMARKS (Table 1) complements the MIGS/MIMS checklists for genomes and
metagenomes by adding two new checklists, a MIMARKS-survey, for uncultured
diversity marker gene surveys, and a MIMARKS-specimen, for marker gene sequences
obtained from any material identifiable via specimens. The MIMARKS extension adopts
and incorporates the standards being developed by the Consortium for the Barcode of
Life (CBOL)

231 (http://www.barcodeoflife.org/sites/default/files/legacy/pdf/DWG_data_standards-

Final.pdf). Therefore, the checklist can be universally applied to any marker gene, from
SSU rRNA to COI, to all taxa, and to studies ranging from single individuals to complex
communities.

Both MIMARKS and the environmental packages were developed by collating

information from several sources and evaluating it in the framework of the existing

237 MIGS/MIMS checklists. These include four independent community-led surveys,

examination of the parameters reported in published studies, and examination of

compliance with optional features in INSDC documents. The overall goal of these

activities was to design the backbone of the MIMARKS checklist, which describes the

241 most important aspects of marker gene contextual data.

242 Results of community-led surveys

243 To date, four online surveys about descriptors for marker genes have been conducted to

244 determine researcher preferences for core descriptors. The Department of Energy Joint

245 Genome Institute and SILVA²² surveys focused on general descriptor contextual data for

a marker gene, whereas the Ribosomal Database Project (RDP)²³ focused on prevalent

habitats for rRNA gene surveys, and the Terragenome Consortium²⁴ focused on soil

248 metagenome project contextual data (supplementary information 1). The above

recommendations were joined by an extensive set of contextual data items suggested by

an International Census of Marine Microbes (ICoMM) working group that met in 2005.

251 These collective resources provided valuable insights into community requests for

252 contextual data items to be included in the MIMARKS checklist and the main habitats

253 constituting the environmental packages.

254 Survey of published parameters

255 We reviewed published rRNA gene studies, retrieved via SILVA and the ICoMM

256 database MICROBIS (The Microbial Oceanic Biogeographic Information System)

257 (<u>http://icomm.mbl.edu/microbis</u>) to further supplement contextual data items that are

258 included in the respective environmental packages. In total, 39 publications from SILVA

and >40 ICoMM projects were scanned for contextual data items to constitute the core of

the environmental package sub-tables (supplementary information 1).

261 Survey of INSDC source feature qualifiers

262 In a final analysis step, we surveyed usage statistics of INSDC source feature key

263 qualifier values of rRNA gene sequences contained in SILVA (supplementary

information 1). Notably, less than 10% of the 1.2 million 16S rRNA gene sequences

265 (SILVA release 100) were associated with even basic information such as

266 latitude/longitude, collection date or PCR primers.

267 The MIMARKS checklist

268 The MIMARKS checklist provides users with an "electronic laboratory notebook"

269 containing core contextual data items required for consistent reporting of marker gene

270 investigations. MIMARKS uses the MIGS/MIMS checklists with respect to the nucleic

acid sequence source and sequencing contextual data, but extends them with further

272 experimental contextual data such as PCR primers and conditions, or target gene name.

273 For clarity and ease of use, all items within the MIMARKS checklist are presented with a

value syntax description, as well as a clear definition of the item. Whenever terms from a

specific ontology are required as the value of an item, these terms can be readily found in

the respective ontology browsers linked by URLs in the item definition. Although this

277 version of the MIMARKS checklist does not contain unit specifications, we recommend

all units to be chosen from and follow the International System of Units (SI)

279 recommendations. In addition, we strongly urge the community to provide feedback

280 regarding the best unit recommendations for given parameters. To facilitate comparative

studies, unit standardization across data sets will be vital in future. An Excel[®] version of

the MIMARKS checklist is provided to the community on the GSC web site at:

283 <u>http://gensc.org/gc_wiki/index.php/MIMARKS</u>.

284 The MIxS environmental packages

285 Fourteen environmental packages provide a wealth of environmental and epidemiological 286 contextual data fields for a complete description of sampling environments. Furthermore, 287 the environmental packages can be combined with any of the GSC checklists (figure 1 288 and supplementary information 2). Researchers within The Human Microbiome Project²⁵ 289 contributed the host-associated and all human packages. The Terragenome Consortium 290 contributed sediment and soil packages. Finally, ICoMM, Microbial Inventory Research 291 Across Diverse Aquatic Long Term Ecological Research Sites (MIRADA-LTERs), and 292 the Max Planck Institute for Marine Microbiology contributed the water package. The 293 MIMARKS working group developed the remaining packages (air, microbial 294 mat/biofilm, miscellaneous natural or artificial environment, plant-associated, and 295 wastewater/sludge). The package names describe high-level habitat terms in order to be 296 exhaustive. The miscellaneous natural or artificial environment package contains a 297 generic set of parameters, and is included for any other habitat that does not fall into the 298 other thirteen categories. Whenever needed, multiple packages may be used for the 299 description of the environment.

300 Examples of MIMARKS-compliant datasets

301 Several MIMARKS-compliant reports are included in Supplementary Information 3.

302 These include a 16S rRNA gene survey from samples obtained in the North Atlantic, a

303 18S pyrosequencing tag study of anaerobic protists in a permanently anoxic basin of the

304 North Sea, a *pmoA* survey from Negev Desert soils, a *dsrAB* survey of Gulf of Mexico

305 sediments, and a 16S pyrosequencing tag study of bacterial diversity in the Western

306 English Channel (accessible via SRA study accession number SRP001108).

307 Adoption by major database and informatics resources

Support for adoption of MIMARKS and the MIxS standard has spread rapidly. Authors of this paper include representatives from genome sequencing centers, maintainers of major resources, principal investigators of large- and small-scale sequencing projects, and individual investigators who have provided compliant datasets, showing the breadth of support for the standard within the community.

In the past, the INSDC has issued a reserved "BARCODE" keyword for the CBOL²⁶. Following this model, the INSDC has recently recognized the GSC as an authority for the MIxS standard and issued it with official keywords within INSDC nucleotide sequence records²⁷. This greatly facilitates automatic validation of the submitted contextual data and provides support for datasets compliant with previous versions by including the checklist version as a keyword.

319 GenBank accepts MIxS metadata in tabular format using the sequin and tbl2asn 320 submission tools, validates MIxS compliance, and reports the fields in the structured 321 comment block. The EBI-ENA Webin submission system provides prepared web forms 322 for the submission of MIxS compliant data; it presents all of the appropriate fields with 323 descriptions, explanations, and examples, and validates the data entered. One tool that can aid submitting contextual data is MetaBar²⁸, a spreadsheet and web-based software, 324 325 designed to assist users in the consistent acquisition, electronic storage and submission of 326 contextual data associated with their samples in compliance with the MIxS standard. The 327 online tool CDinFusion (http://www.megx.net/cdinfusion) was created to facilitate the 328 combination of contextual data with sequence data, and generation of submission-ready 329 files.

330 The next-generation Sequence Read Archive (SRA) collects and displays MIxS-331 compliant metadata in sample and experiment objects. There are several tools that are 332 already available or under development to assist users in SRA submissions. The myRDP 333 SRA PrepKit allows users to prepare and edit their submissions of reads generated from 334 ultra-high-throughput sequencing technologies. A set of suggested attributes in the data 335 forms assist researchers in providing metadata conforming to checklists such as 336 MIMARKS. The Quantitative Insights Into Microbial Ecology ("QIIME") web 337 application (http://www.microbio.me/qiime) allows users to generate and validate 338 MIMARKS-compliant templates. These templates can be viewed and completed in the users' spreadsheet editor of choice (e.g. Microsoft Excel[®]). The QIIME web-platform also 339 340 offers an ontology lookup and geo-referencing tool to aid users when completing the 341 MIMARKS templates. The Investigation/Study/Assay (ISA) is a software suite that 342 assists in the curation, reporting, and local management of experimental metadata from 343 studies employing one or a combination of technologies, including high-throughput sequencing²⁹. Specific ISA configurations (available from http://isa-tools.org/tools.html) 344 345 have been developed to ensure MIxS compliance by providing templates and validation 346 capability. Another tool, ISAconverter, produces SRA.xml documents, facilitating 347 submission to the SRA repository. 348 Further detailed guidance for submission processes can be found under the 349 respective wiki pages (http://gensc.org/gc_wiki/index.php/MIGS/MIMS/MIMARKS) of

the standard.

351 Maintenance of the MIxS standard

352 To allow further developments, extensions, and enhancements of MIxS, we set up a

353 public issue tracking system to track changes and accomplish feature requests 354 (http://mixs.gensc.org/). New versions will be released annually. Technically, the MIxS 355 standard, including MIMARKS and the environmental packages, is maintained in a 356 relational database system at the Max Planck Institute for Marine Microbiology Bremen 357 on behalf of the GSC. This provides a secure and stable mechanism for updating the 358 checklist suite and versioning. In future, we plan to develop programmatic access to this 359 database in order to allow automatic retrieval of the latest version of each checklist for 360 INSDC databases and for GSC community resources. Moreover, the Genomic Contextual 361 Data Markup Language (GCDML) is a reference implementation of the GSC checklists 362 by the GSC and now implements the full range of MIxS standards. It is based on XML 363 Schema technology and thus serves as an interoperable data exchange format for Web Service based infrastructures³⁰. 364

365

366 **Conclusions and call for action**

367 The GSC is an international body with a stated mission of working towards richer 368 descriptions of the complete collection of genomes and metagenomes through the MIxS 369 standard. The present report extends the scope of GSC guidelines to marker gene 370 sequences and environmental packages and establishes a single portal where 371 experimentalists can gain access to and learn how to use GSC guidelines. The GSC is an 372 open initiative that welcomes the participation of the wider community. This includes an 373 open call to contribute to refinements of the MIxS standards and their implementations. 374 The adoption of the GSC standards by major data providers and organizations, as well as 375 the INSDC, underlines and seconds the efforts to contextually enrich our sequence data

376 collection, and complements the recent efforts to enrich other (meta) omics data. The 377 MIxS standard, including MIMARKS, has been developed to the point that it is ready for 378 use in the publication of sequences. A defined procedure for requesting new features and 379 stable release cycles will facilitate implementation of the standard across the community. 380 Compliance among authors, adoption by journals and use by informatics resources will 381 vastly improve our collective ability to mine and integrate invaluable sequence data 382 collections for knowledge- and application-driven research. In particular, the ability to 383 combine microbial community samples collected from any source, using the universal 384 Tree of Life as a measure to compare even the most diverse communities, should provide 385 new insights into the dynamic spatiotemporal distribution of microbial life on our planet 386 and in/on the human body.

388	Figur	e Legend	
389	Figure	e 1: Schematic overview about the GSC MIxS standard (brown), including	
390	combination with specific environmental packages (blue). Shared descriptors apply to all		
391	MIxS checklists, however each checklist has its own specific descriptors as well.		
392	Enviro	nmental packages can be applied to any of the checklists. (EU: Eukarya, BA:	
393	Bacteria/Archaea, PL: Plasmid, VI: Virus, ORG: Organelle).		
394			
395	1.	Field, D. et al. The minimum information about a genome sequence (MIGS)	
396		specification. Nat. Biotechnol. 26, 541-547 (2008).	
397	2.	Taylor, C.F. et al. Promoting coherent minimum reporting guidelines for	
398		biological and biomedical investigations: the MIBBI project. Nat. Biotechnol. 26,	
399		889-896 (2008).	
400	3.	Ludwig, W. & Schleifer, K.H. in Microbial phylogeny and evolution, concepts	
401		and controversies. (ed. J. Sapp) 70-98 (Oxford university press, New York, USA,	
402		2005).	
403	4.	Ludwig, W. et al. Bacterial phylogeny based on comparative sequence analysis.	
404		Electrophoresis 19, 554-568 (1998).	
405	5.	Giovannoni, S.J., Britschgi, T.B., Moyer, C.L. & Field, K.G. Genetic diversity in	
406		Sargasso Sea bacterioplankton. Nature 345, 60-63 (1990).	
407	6.	Stahl, D.A. Analysis of hydrothermal vent associated symbionts by ribosomal	
408		RNA sequences. Science 224, 409-411 (1984).	
409	7.	Ward, D.M., Weller, R. & Bateson, M.M. 16S rRNA sequences reveal numerous	
410		uncultured microorganisms in a natural community. Nature 345, 63-65 (1990).	

- 411 8. DeLong, E.F. Archaea in coastal marine environments. *Proc. Nat. Acad. Sci. USA*412 89, 5685-5689 (1992).
- 413 9. Diez, B., Pedros-Alio, C. & Massana, R. Study of genetic diversity of eukaryotic
 414 picoplankton in different oceanic regions by small-subunit rRNA gene cloning
 415 and sequencing. *Appl. Environ. Microbiol.* 67, 2932-2941 (2001).
- 416 10. Fuhrman, J.A., McCallum, K. & Davis, A.A. Novel major archaebacterial group
 417 from marine plankton. *Nature* 356, 148-149 (1992).
- 418 11. Hewson, I. & Fuhrman, J.A. Richness and diversity of bacterioplankton species
- 419 along an estuarine gradient in Moreton Bay, Australia. *Appl. Environ. Microbiol.*
- **420 70**, 3425-3433 (2004).
- 421 12. Huber, J.A., Butterfield, D.A. & Baross, J.A. Temporal changes in archaeal
- 422 diversity and chemistry in a mid-ocean ridge subseafloor habitat. *Appl. Environ.*423 *Microbiol.* 68, 1585-1594 (2002).
- 424 13. Lopez-Garcia, P., Rodriguez-Valera, F., Pedros-Alio, C. & Moreira, D.
- 425 Unexpected diversity of small eukaryotes in deep-sea Antarctic plankton. *Nature*426 409, 603-607 (2001).
- 427 14. Moon-van der Staay, S.Y., De Wachter, R. & Vaulot, D. Oceanic 18S rDNA

428 sequences from picoplankton reveal unsuspected eukaryotic diversity. *Nature*429 409, 607-610 (2001).

- 430 15. Pace, N.R. A molecular view of microbial diversity and the biosphere. *Science*431 **276**, 734-740 (1997).
- 432 16. Rappe, M.S. & Giovannoni, S.J. The uncultured microbial majority. *Annu. Rev.*433 *Microbiol.* 57, 369-394 (2003).

434	17.	Francis, C.A., Beman, J.M. & Kuypers, M.M.M. New processes and players in
435		the nitrogen cycle: the microbial ecology of anaerobic and archaeal ammonia
436		oxidation. ISME. J. 1, 19-27 (2007).
437	18.	Zehr, J.P., Mellon, M.T. & Zani, S. New nitrogen-fixing microorganisms detected
438		in oligotrophic oceans by amplification of nitrogenase (nifH) genes. Appl.
439		Environ. Microbiol. 64, 3444-3450 (1998).
440	19.	Minz, D. et al. Diversity of sulfate-reducing bacteria in oxic and anoxic regions of
441		a microbial mat characterized by comparative analysis of dissimilatory sulfite
442		reductase genes. Appl. Environ. Microbiol. 65, 4666-4671 (1999).
443	20.	Gilbert, J., A. et al. The seasonal structure of microbial communities in the
444		Western English Channel. Environ. Microbiol. 11, 3132-3139 (2009).
445	21.	Martinez, A., W. Tyson, G. & DeLong, E., F. Widespread known and novel
446		phosphonate utilization pathways in marine bacteria revealed by functional
447		screening and metagenomic analyses. Environ. Microbiol. 12, 222-238 (2009).
448	22.	Pruesse, E. et al. SILVA: a comprehensive online resource for quality checked
449		and aligned ribosomal RNA sequence data compatible with ARB. Nucleic Acids
450		<i>Res.</i> 35 , 7188-7196 (2007).
451	23.	Cole, J.R. et al. The Ribosomal Database Project: improved alignments and new
452		tools for rRNA analysis. Nucleic Acids Res. 37, D141-145 (2009).
453	24.	Vogel, T.M. et al. TerraGenome: a consortium for the sequencing of a soil
454		metagenome. Nat. Rev. Microbiol. 7, 252-252 (2009).
455	25.	Turnbaugh, P.J. et al. The Human Microbiome Project. Nature 449, 804-810

456 (2007).

457	26.	Benson, D.A. et al. GenBank. Nucl. Acids Res. 36, D25-30 (2008).
458	27.	Hirschman, L. et al. Meeting report: Metagenomics, Metadata and Meta-analysis"
459		(M3) Workshop at the Pacific Symposium on Biocomputing 2010. SIGS 2, 357-
460		360 (2010).
461	28.	Hankeln, W. et al. MetaBar - a tool for consistent contextual data acquisition and
462		standards compliant submission. BMC Bioinformatics 11, 358 (2010).
463	29.	Rocca-Serra, P. et al. ISA infrastructure: supporting standards-compliant
464		experimental reporting and enabling curation at the community level.
465		Bioinformatics 26, 2354-2356 (2010).
466	30.	Kottmann, R. et al. A standard MIGS/MIMS compliant XML schema: Toward
467		the development of the Genomic Contextual Data Markup Language (GCDML).
468		<i>OMICS</i> 12 , 115-121 (2008).

Specification projects	MIGS	MIMS	MIMARKS	New checklists	
Checklists	EU BA PL VI ORG	metagenomes	survey specimen	e.g. pan-genomes	
Shared descriptors	collection date, environmental package, environment (biome), environment (feature), environment (material), geographic location (country and/or sea, region), geographic location (latitude and longitude), investigation type, project name, sequencing method, submitted to INSDC				
Checklist specific descriptors	assembly, estimated size, finishing strategy, isolation and growth condition, number of replicons, ploidy, propagation, reference for biomaterial		target gene		
Applicable environmental packages (measurements and observations)	Air Host-associated Human-associated Human-oral Human-gut Human-skin Human-vaginal	Misce	Microbial mat/biofilm cellaneous natural or artificial environment Plant-associated Sediment Soil Wastewater/sludge Water		

			type	
		MIMARKS-	MIMARKS	
		survey	-specimen	
Submitted to INSDC ^[boolean]	Depending on the study (large-scale e.g. done with next generation sequencing technology, or small-scale) sequences have to be submitted to SRA (Sequence Read Archives), DRA (DDBJ Sequence Read Archive) or via the classical Webin/Sequin systems to Genbank, ENA and DDBJ	М	М	
Investigation type ^[mimarks-survey or mimarks-specimen]	Nucleic Acid Sequence Report is the root element of all MIMARKS compliant reports as standardized by Genomic Standards Consortium (GSC). This field is either MIMARKS survey or MIMARKS specimen	М	М	
Project name	Name of the project within which the sequencing was organized	М	М	
	Environment			
Geographic location (latitude and longitude ^[float, point, transect and region])	The geographical origin of the sample as defined by latitude and longitude. The values should be reported in decimal degrees and in WGS84 system	М	М	
Geographic location (depth [integer, point, interval, unit])	Please refer to the definitions of depth in the environmental packages	Е	Е	
Geographic location (elevation of site ^[integer, unit] ; altitude of sample ^[integer, unit])	Please refer to the definitions of either altitude or elevation in the environmental packages	E	Е	
Geographic location (country and/or sea ^[INSDC or GAZ] ; region ^[GAZ])	The geographical origin of the sample as defined by the country or sea name. Country, sea, or region names should be chosen from the INSDC list (http://insdc.org/country.html), or the GAZ (Gazetteer, v1.446) ontology (http://bioportal.bioontology.org/visualize/406 51)	М	М	
Collection date ^[ISO8601]	The time of sampling, either as an instance (single point in time) or interval. In case no exact time is available, the date/time can be right truncated i.e. all of these are valid times: 2008-01-23T19:23:10+00:00; 2008-01- 23T19:23:10; 2008-01-23; 2008-01; 2008; Except: 2008-01; 2008 all are ISO6801 compliant	М	М	

Environment (biome ^[EnvO])	In environmental biome level are the major classes of ecologically similar communities of plants, animals, and other organisms. Biomes are defined based on factors such as plant structures, leaf types, plant spacing, and other factors like climate. Examples include: desert, taiga, deciduous woodland, or coral reef. Environment Ontology (EnvO) (v1.53) terms listed under environmental biome can be found from the link: http://bioportal.bioontology.org/visualize/4440 5/?conceptid=ENVO%3A00000428	М	М				
Environment (feature ^[EnvO])	Environmental feature level includes geographic environmental features. Examples include: harbor, cliff, or lake. EnvO (v1.53) terms listed under environmental feature can be found from the link: http://bioportal.bioontology.org/visualize/4440 5/?conceptid=ENVO%3A00002297	М	М				
Environment (material ^[EnvO])	The environmental material level refers to the matter that was displaced by the sample, prior to the sampling event. Environmental matter terms are generally mass nouns. Examples include: air, soil, or water. EnvO (v1.53) terms listed under environmental matter can be found from the link: http://bioportal.bioontology.org/visualize/4440 5/?conceptid=ENVO%3A00010483	М	М				
	MIGS/MIMS/MIMARKS Extension						
Environmental package ^[air, host-associated, human-associated, human-skin, human-oral, human-gut, human-vaginal, microbial mat/biofilm, miscellaneous natural or artificial environment, plant-associated, sediment, soil, wastewater/sludge, water]	MIGS/MIMS/MIMARKS extension for reporting of measurements and observations obtained from one or more of the environments where the sample was obtained. All environmental packages listed here are further defined in separate subtables. By giving the name of the environmental package, a selection of fields can be made from the subtables and can be reported	М	М				
Nucleic acid sequence source							
Isolation and growth conditions [PMID, DOI, or URL]	Publication reference in the form of pubmed ID (PMID), digital object identifier (DOI), or URL for Isolation and growth condition specifications of the organism/material	-	М				
	Sequencing						
Target gene or locus (e.g. 16S rRNA, 18S rRNA, nif, amoA, rpo)	Targeted gene or locus name for marker gene study	М	М				
Sequencing method (e.g. dideoxysequencing, pyrosequencing, polony)	Sequencing method used; e.g. Sanger, pyrosequencing, ABI-solid.	М	М				

Table 1. Items for the MIMARKS specification and their mandatory (M), conditionally mandatory (C) (the item is mandatory only when applicable to the study) or recommended (X) status for both MIMARKS-survey and MIMARKS-specimen checklists. Furthermore, "-" denotes that an item is not applicable for a given checklist. "E" denotes that a field has environment-specific requirements. For example, while "depth" is mandatory for environments water, sediment or soil; it is optional for human-associated environments. **MIMARKS-survey** is applicable to contextual data for marker gene sequences, obtained directly from the environment, without culturing or identification of the organisms. **MIMARKS-specimen**, on the other hand, applies to the contextual data for marker gene sequences from cultured or voucher-identifiable specimens. Both MIMARKS-survey and specimen checklists can be used for any type of marker gene sequence data, ranging from 16S, 18S, 23S, 28S rRNA to COI, hence the checklists are universal for all three domains of life.

Item names are followed by a short description of the value of the item in parentheses and/or value type in brackets as a superscript. Whenever applicable, value types are chosen from a controlled vocabulary (CV), or an ontology from the Open Biological and Biomedical Ontologies (OBO) foundry (http://www.obofoundry.org). This table only presents the very core of MIMARKS checklists, i.e. only mandatory items for each checklist. Supplementary information 2 in spreadsheet format contains all MIMARKS items, the tables for environmental packages in the MIGS/MIMS/MIMARKS extension, and GenBank structured comment name that should be used for submitting MIMARKS data to GenBank. In case of submitting to EBI/ENA the full names can be used.