

Supporting Information

Acetogenin and Prenylated Flavonoids from *Helminthostachys zeylanica* with Inhibitory Activity on Superoxide Generation and Elastase Release by Neutrophils

**Yaun-Chao Huang¹, Tsong-Long Hwang², Yu-Liang Yang³, Shih-Hsiung Wu³,
Mei-Hua Hsu¹, Jih-Pyang Wang⁴, Sheng-Chih Chen¹, Li-Jiau Huang¹,
Chih-Chuang Liaw^{1,5}**

Affiliation

¹ Graduate Institute of Pharmaceutical Chemistry, College of Pharmacy, China
Medical University, Taichung 404, Taiwan

² Graduate Institute of Natural Products, College of Medicine, Chang Gung University,
Taoyuan 333, Taiwan

³ Institute of Biological Chemistry, Academia Sinica, Taipei 115, Taiwan

⁴ Department of Education and Research, Taichung Veterans General Hospital,
Taichung 407, Taiwan

⁵ Department of Life Sciences, National Chung-Hsing University, Taichung 402,
Taiwan

Correspondence

Dr. Chih-Chuang Liaw

Graduate Institute of Pharmaceutical Chemistry

College of Pharmacy

China Medical University

91 Hsueh-Shih Road

Taichung 40402

Taiwan

Tel.: +886/4/2205/3366 ext. 5611

Fax: +886/4/2207/8083

ccliaw@mail.cmu.edu.tw

Measurement of rat mast cell degranulation and neutrophil degranulation [1-5]

A compound stock solution (30 µg/mL in DMSO) was prepared and stored at -25 °C.

It was then diluted with DMSO to 1-30 µg/mL range at room temperature before the

experiment. The final percentage of DMSO in the reaction mixture was less than 0.5%

(v/v). Rat (Sprague Dawley, 250-300 g) peritoneal mast cells and peripheral blood

neutrophils were isolated and incubated with test compounds for 5 min at 37 °C

before stimulation with 10 µg/mL of the compound 48/80 for another 15 min or with

1 µM FMLP/CB for another 45 min, respectively. The degranulation of mast cells and

neutrophils was assessed by the determination of histamine and β-glucuronidase, as

well as β -glucuronidase and lysozyme in the supernatant, respectively. The total content of lysozyme and β -glucuronidase was measured from the *Micrococcus lysodeikticus*-treated cells and phosphoglucuronide.

1 Wang JP, Raung SL, Kuo YH and Teng CM. Daphnoretin-induced respiratory burst in rat neutrophil is probably mainly through protein kinase C activation. Eur J Pharmacol 1995; 288: 341-348

2 Barrett AJ. Lysosomes. In: Dingle JT, editor. A laboratory handbook. Amsterdam: Elsevier; 1972: 118-120

3 Absolom DR. Basic methods for the study of phagocytosis. Methods Enzymol 1986; 132: 92-179

4 Wang JP, Hsu MF, Ouyang C, Teng CM. Edematous response caused by [Thi^{5,8}-D-Phe]bradykinin, $\alpha\beta_2$ receptor antagonist, is due to mast cell degranulation. Eur J Pharmacol 1989; 161: 143-149

5 Hakanson R, Ronnberg AL. Improved fluorometric assay of histamine. Analyt Biochem 1974; 60: 560-567

Table 1S UV spectral properties of compounds **2-5**

Compounds	$\lambda_{\max}^{\text{MeOH}}$ (nm)		$\lambda_{\max}^{\text{MeOH}+\text{AlCl}_3}$ (nm)		$\lambda_{\max}^{\text{MeOH}+\text{AlCl}_3+\text{HCl}}$ (nm)	
	Bd II	Bd I	Bd II	Bd I	Bd II	Bd I
2	271	349	275	333, 425	283	361
3	271	331	274	359, 465	273	340, 415
4	292	344	292	418	296,	350
5	251	300, 351	264	418	264	334, 393

Table 2S Inhibitory effects of crude extracts from *H. zeylanica* on β -glucuronidase and histamine by rat mast cell in response to compound 48/80

Drugs ($\mu\text{g/mL}$)	Percent Release								
	β -glucuronides ($\mu\text{g/mL}$) or (Inh %)				histamine ($\mu\text{g/mL}$) or (Inh %)				
Control		14.2 \pm 1.6				83.1 \pm 2.2			
Methanol extract	10	14.8 \pm 1.0	(3.8 \pm 8.9)	85.8 \pm 7.9	(-3.0 \pm 9.4)				
	30	11.6 \pm 0.9	(24.5 \pm 6.3) *	76.3 \pm 3.9	(8.3 \pm 4.6)				
<i>n</i> -Hexane extract	10	12.5 \pm 1.3	(15.3 \pm 4.4) **	92.3 \pm 3.3	(-10.8 \pm 4.1)				
	30	13.3 \pm 0.7	(12.9 \pm 8.8)	93.2 \pm 3.6	(-11.9 \pm 4.6)				
Chloroform extract	10	12.5 \pm 0.5	(-6.1 \pm 9.5)	81.6 \pm 5.8	(-1.6 \pm 6.6)				
	30	7.8 \pm 0.3	(33.4 \pm 6.4) *	53.4 \pm 0.3	(33.2 \pm 3.0) **				
Water extract	10	13.0 \pm 1.7	(16.7 \pm 6.9)	89.1 \pm 1.5	(-7.0 \pm 1.9)				
	30	8.9 \pm 1.5	(43.5 \pm 6.6) **	79.3 \pm 2.3	(4.8 \pm 2.1)				
Mepacrine	10 μM	12.1 \pm 0.4	(18.1 \pm 2.5) *	64.1 \pm 2.3	(21.3 \pm 2.5) *				
	30 μM	7.2 \pm 0.2	(46.3 \pm 3.2) **	34.8 \pm 1.1	(53.7 \pm 1.3) **				
	100 μM	3.1 \pm 0.2	(81.9 \pm 1.4) **	14.2 \pm 1.3	(85.2 \pm 1.7) **				
	IC ₅₀	27 \pm 9.2		26.2 \pm 1.6					

Mepacrine was used as positive control. Results are presented as mean \pm S.E.M. (n = 3). * p < 0.05, ** p < 0.01, compared with the control value.

Table 3S Inhibitory effects of crude extracts from *H. zeylanica* on β -glucuronidase and lysozyme by rat neutrophils in response to FMLP/C

Drugs	β -glucuronides	lysozyme
	IC ₅₀ (μ g/mL)	IC ₅₀ (μ g/mL)
Control	30.0 \pm 0.7	29.3 \pm 1.4
Methanol extract	1.1 \pm 0.1	N
<i>n</i> -Hexane extract	2.0 \pm 0.7	9.0 \pm 2.9
Chloroform extract	1.3 N 0.1	N
Trifluoperazine	9.8 \pm 0.4	9.6 \pm 0.4

Trifluoperazine (TFP) was used as positive control. Results are presented as mean \pm S.E.M. (n = 3). * p < 0.05, ** p < 0.01, compared with the control value. Water extract was inactive.

Table 4S Cytotoxicity activities of crude extracts from *H. zeylanica*

Compounds	HL-60 ^a	U937 ^a
	IC ₅₀ (µg/mL) ^a or (Inh %) ^b	IC ₅₀ (µg/mL) ^a or (Inh %) ^b
Methanol extract	19.9	21.0
<i>n</i> -Hexane extract	5.1	5.3
Chloroform extract	19.3	14.6
Ethyl acetate extract	7.8	5.9
Retinoic acid (0.03 µg/mL)	(34.2 ± 3.3)	(31.3 ± 1.3)

Retinoic acid (0.03 µg/mL) was used as positive control. Results are presented as mean ± S.E.M. (n = 3).

^a HL-60 and U937 cells (2 x 10⁴ cells) were incubated with different concentration in 96 hours.

^b Percentage of inhibition (Inh %) at 0.03 µg/mL concentration.

Figure 1S Selected HMBC correlations of compounds **1-5**.

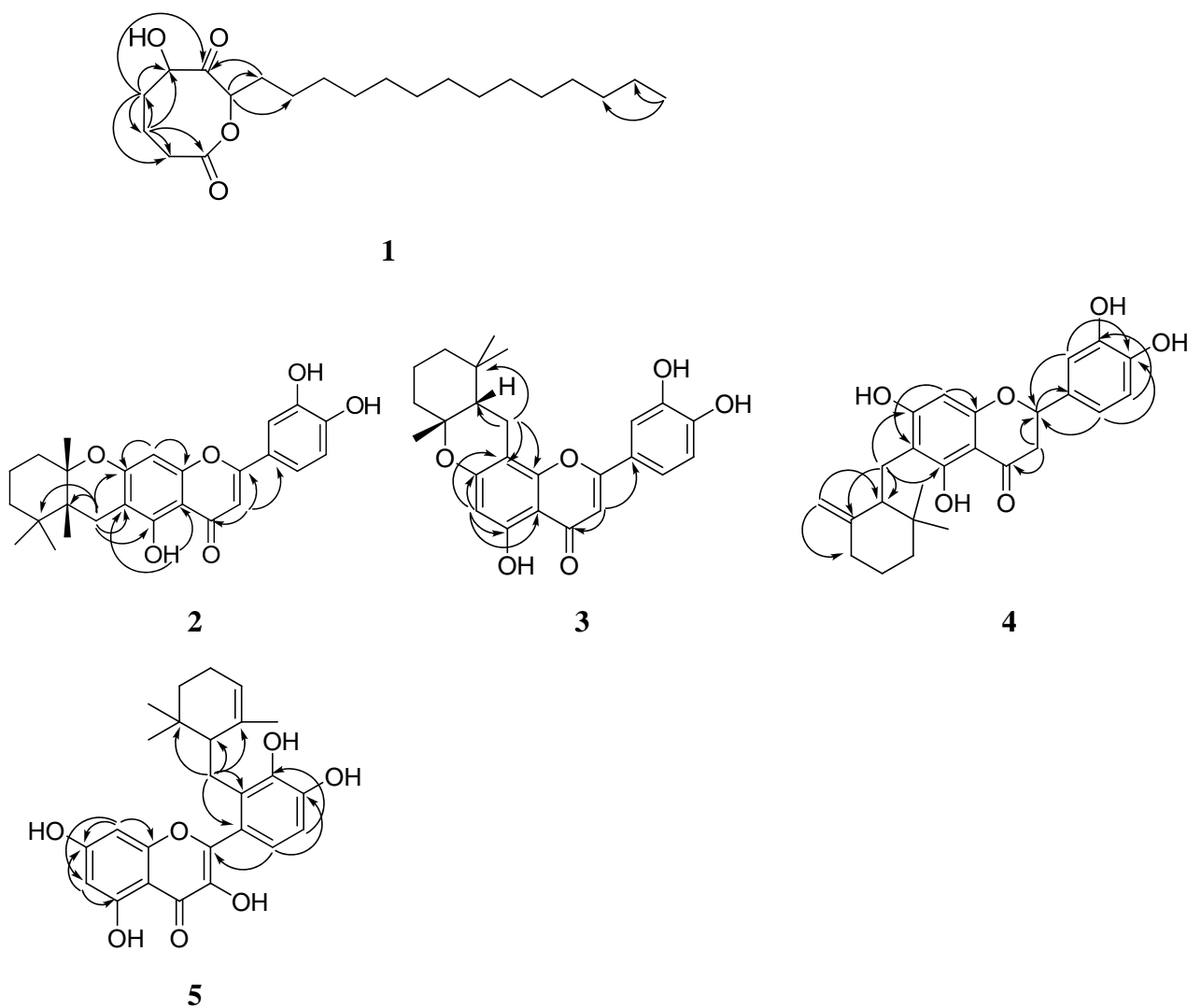


Figure 2S The ^1H -NMR spectrums of ugonin L and compounds **2** and **3**.

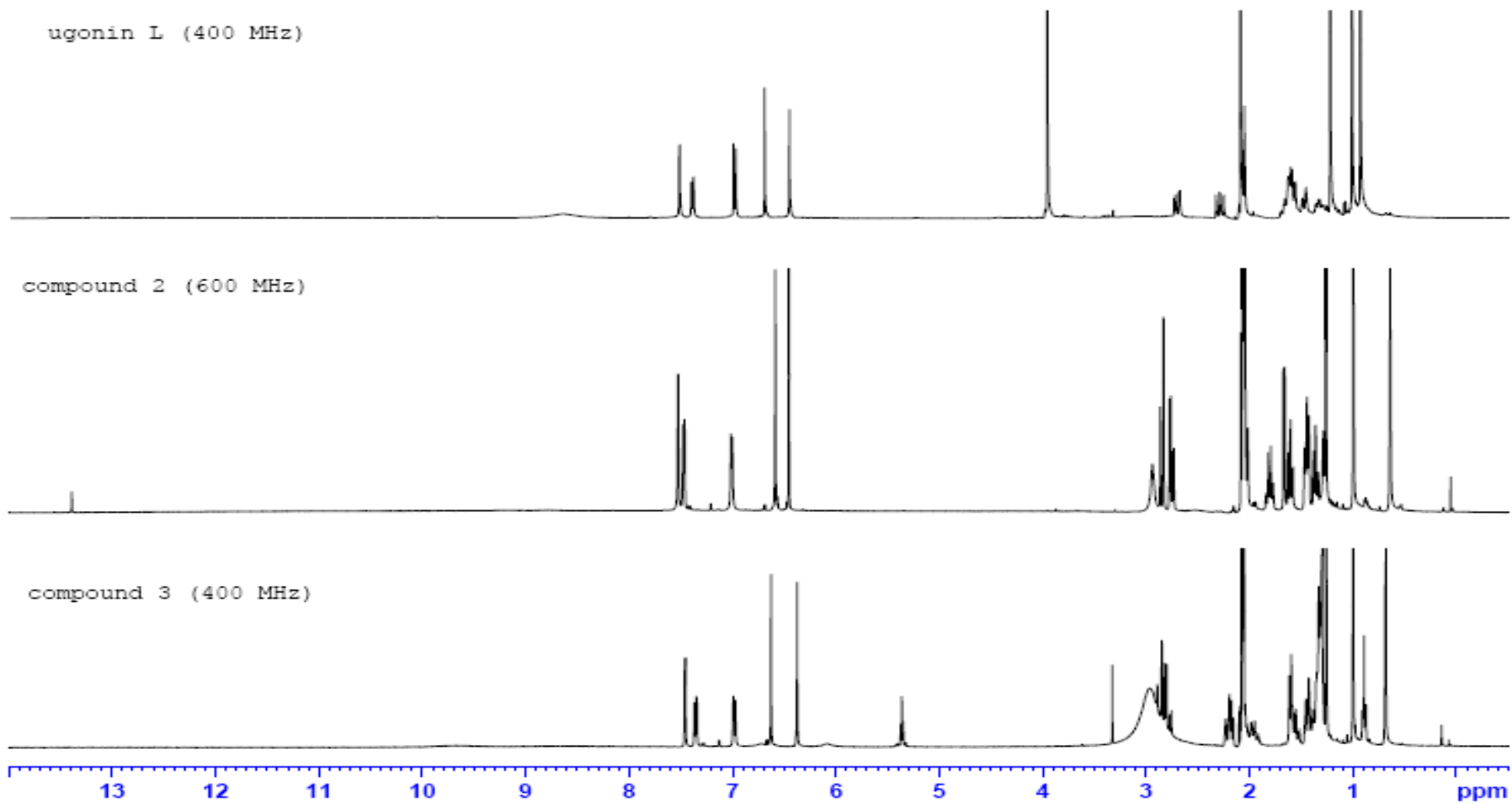
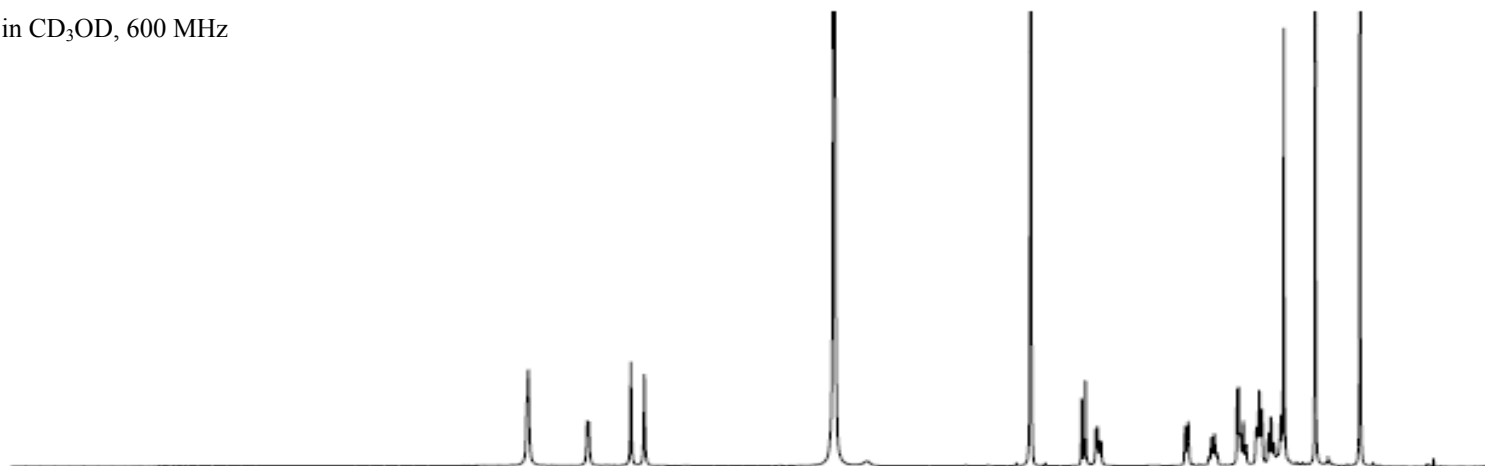


Figure 3S The ^1H -NMR spectrums of compound **2** measured in different *d*-solvents.

(1) Compound **2** measured in CD_3OD , 600 MHz



(2) Compound **2** measured in $\text{MeCO-}d_6$, 600 MHz

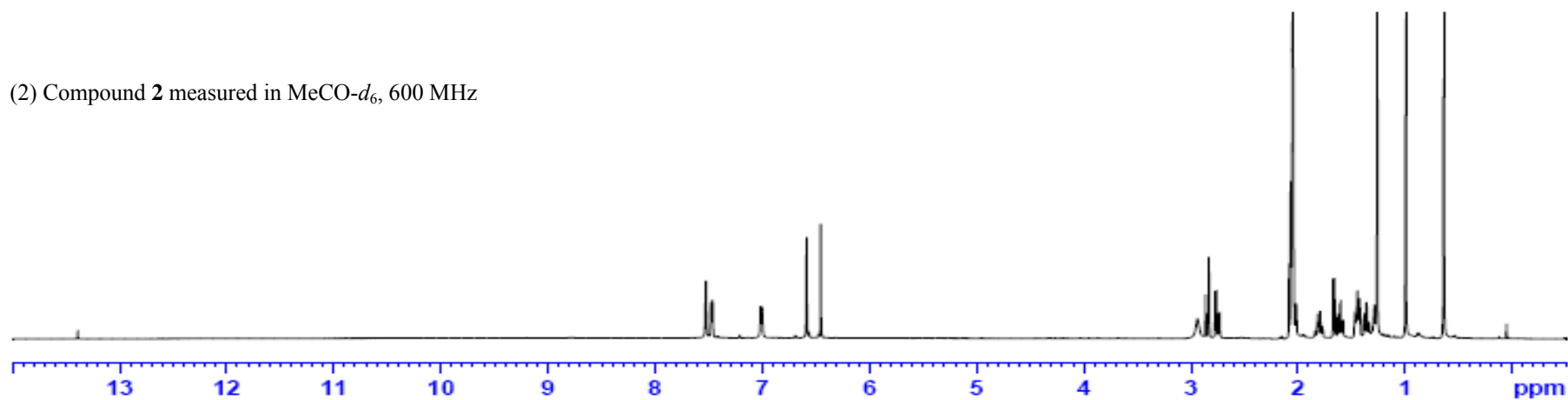
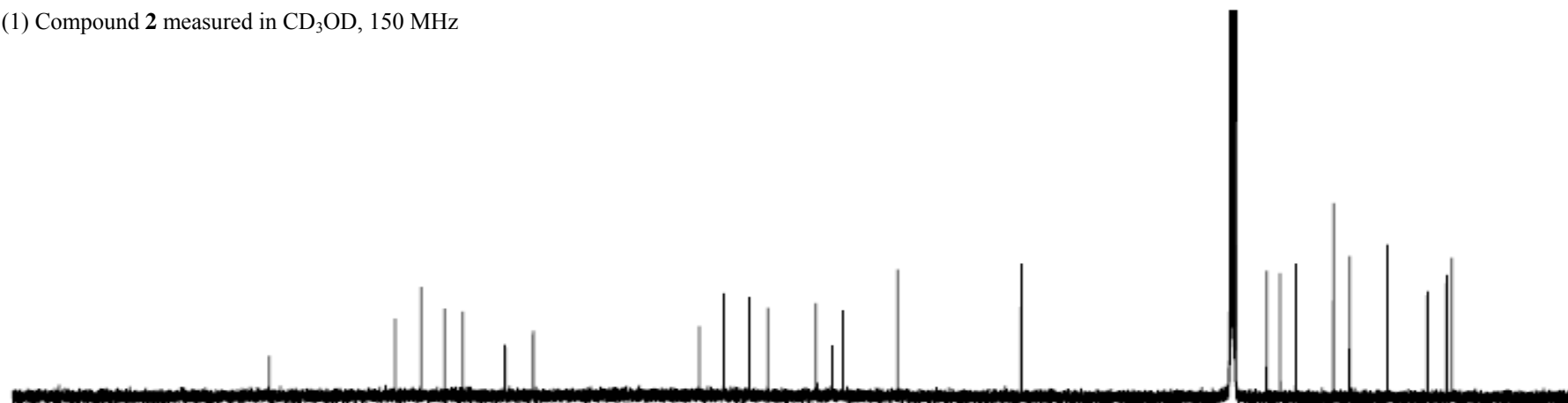


Figure 4S The ^{13}C -NMR spectrums of compound **2** measured in different *d*-solvents.

(1) Compound **2** measured in CD_3OD , 150 MHz



(2) Compound **2** measured in $\text{MeCO-}d_6$, 150 MHz

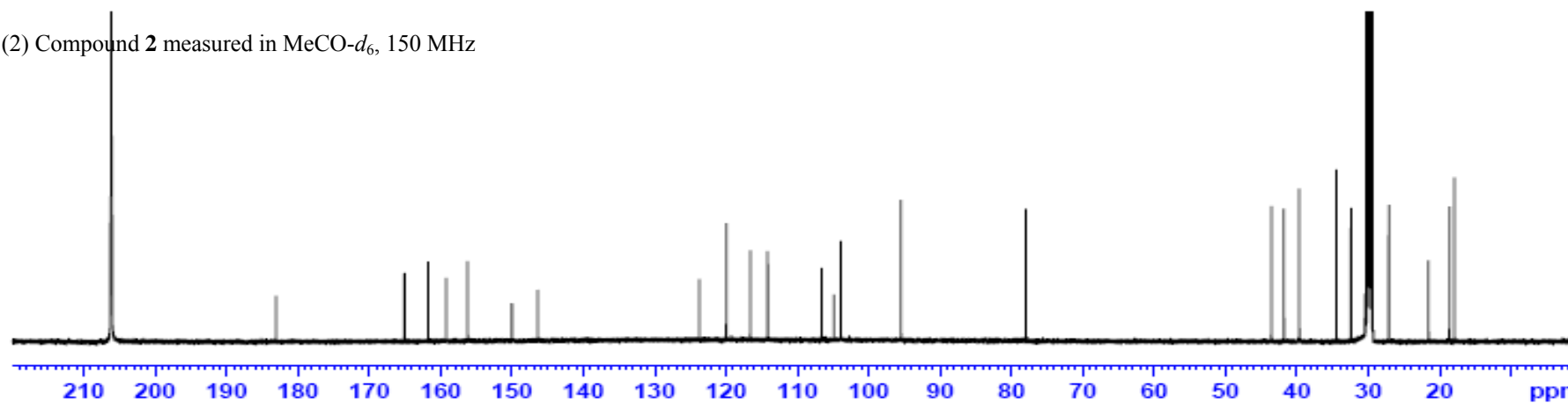
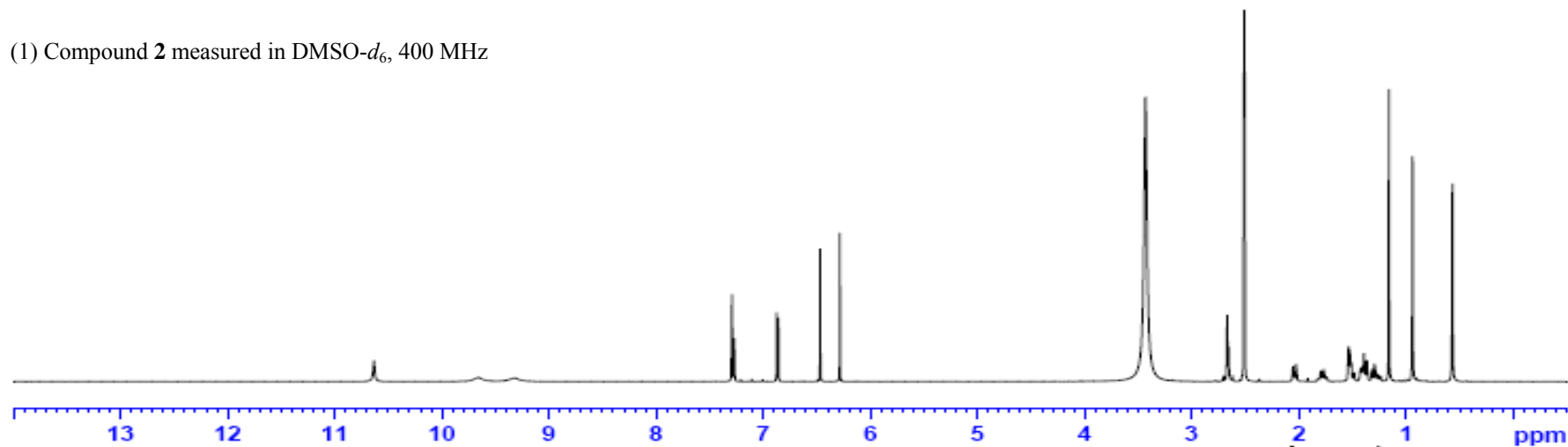


Figure 5S The ^1H -NMR spectrums of compound **3** measured in different *d*-solvents.

(1) Compound **2** measured in $\text{DMSO-}d_6$, 400 MHz



(2) Compound **3** measured in $\text{MeCO-}d_6$, 400 MHz

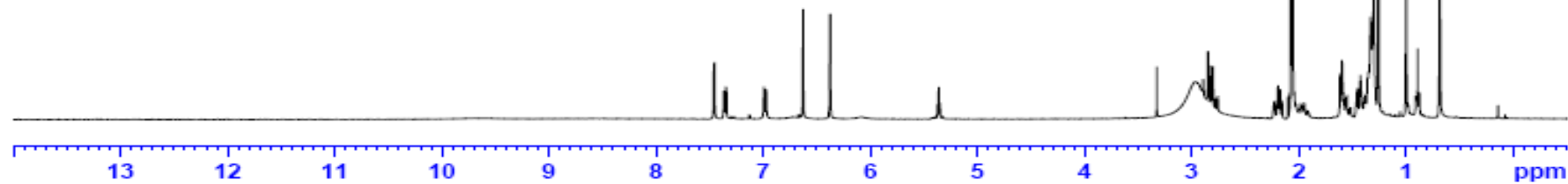
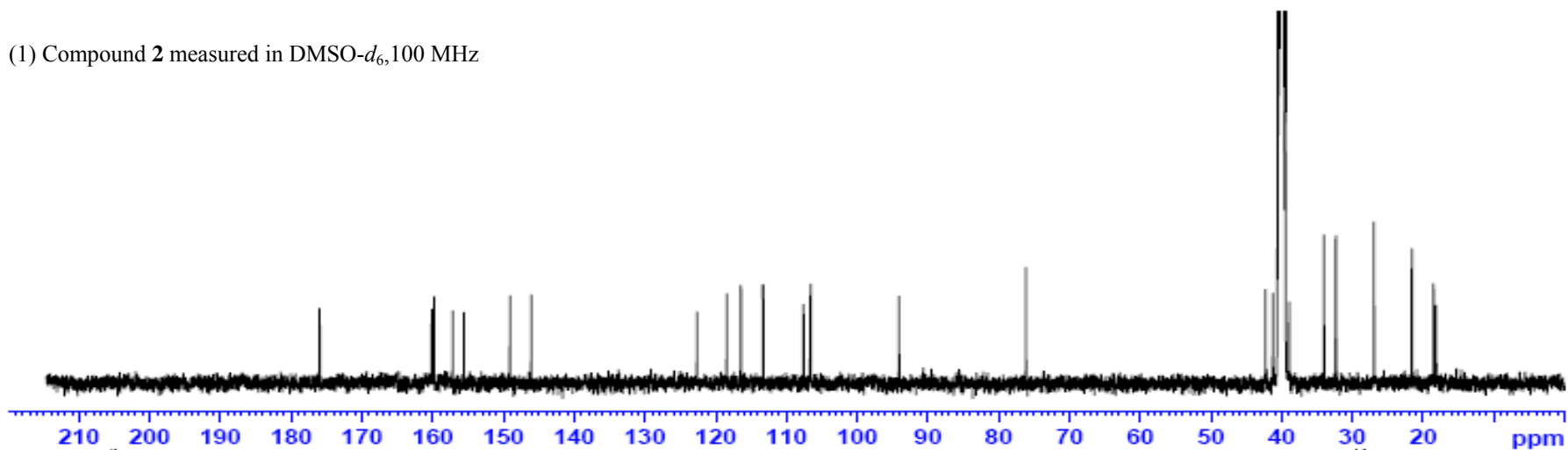


Figure 6S The ^{13}C -NMR spectrums of compound **3** measured in different *d*-solvents.

(1) Compound **2** measured in $\text{DMSO-}d_6$, 100 MHz



(2) Compound **3** measured in $\text{MeCO-}d_6$, 100 MHz

