

# **CHEMISTRY**

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## **A EUROPEAN JOURNAL**

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### Supporting Information

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#### **Weakly Associated TFPB Anions Are Superior to PF<sub>6</sub> Anions When Preparing (Pseudo)Rotaxanes from Crown Ethers and Secondary Dialkylammonium Ions\*\***

**Nai-Chia Chen,<sup>[a]</sup> Chun-Ju Chuang,<sup>[a]</sup> Liang-Yun Wang,<sup>[a]</sup> Chien-Chen Lai,<sup>[b]</sup> and Sheng-Hsien Chiu<sup>\*[a]</sup>**

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Sheng-Hsien Chiu\*

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Threadlike salt **1-H·TFPB**: **1-H·PF<sub>6</sub>** (114 mg, 0.247 mmol) and NaTFPB (222 mg, 0.25 mmol) were mixed in MeOH (10 mL) and then the organic solvent was evaporated under reduced pressure. The residue was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and H<sub>2</sub>O (10 mL). The organic layer was collected, washed with H<sub>2</sub>O (4 × 10 mL), dried (MgSO<sub>4</sub>), and concentrated to give the desired product as a pale yellow liquid (275 mg, 93%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.30 (s, 9H), 4.07 (s, 4H), 4.37 (s, 2H), 7.14 (d, *J* = 8 Hz, 2H), 7.20 (d, *J* = 8 Hz, 2H), 7.36 (d, *J* = 8 Hz, 2H), 7.46 (d, *J* = 8 Hz, 2H), 7.49–7.52 (br, 4H), 7.65–7.72 (br, 8H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 30.9, 35.0, 51.8, 52.3, 53.8, 117.5, 124.5 (q, <sup>1</sup>*J*<sub>CF</sub> = 272 Hz), 124.9, 127.1, 128.1, 128.8 (q, <sup>2</sup>*J*<sub>CF</sub> = 31 Hz), 128.8, 129.5, 134.7, 138.8, 154.9, 161.5 (q, <sup>1</sup>*J*<sub>CB</sub> = 50 Hz) (one signal is missing, possibly because of signal overlap); MS (ESI): C<sub>19</sub>H<sub>25</sub>N<sub>4</sub><sup>+</sup> ([**1-H**]<sup>+</sup>) calcd *m/z* 309.2079; found *m/z* 309.2073.

[2]Rotaxane **2-H·TFPB**: Triethyl phosphite (80 μL, 0.47 mmol) was added dropwise to a solution of **1-H·TFPB** (266 mg, 0.227 mmol) and DA24C8 (243 mg, 0.458 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2.3 mL). The mixture was stirred at ambient temperature overnight. The solvent was evaporated and the residue purified chromatographically (SiO<sub>2</sub>; CH<sub>3</sub>CN/CH<sub>2</sub>Cl<sub>2</sub>, 5:95). The product was isolated as a pale yellow oil (284 mg, 69%). <sup>1</sup>H NMR (800 MHz, CDCl<sub>3</sub>): δ = 1.27–1.34 (m, 15H), 2.21 (s, 6H), 2.82–2.95 (br, 8H), 3.22–3.34 (br, 8H), 3.49–3.60 (br, 16H), 4.02–4.13 (m, 4H), 4.13–4.20 (m, 2H), 4.51–4.59 (br, 2H), 4.59–4.66 (br, 2H), 6.38 (d, *J* = 6 Hz, 4H), 6.92 (d, *J* = 8 Hz, 4H), 7.42 (d, *J* = 8 Hz, 2H), 7.45–7.48 (br, 4H), 7.48–7.51 (br, 4H), 7.56 (d, *J* = 7 Hz, 2H), 7.66–7.70 (br, 8H), 7.91–7.99 (br, 2H); <sup>13</sup>C NMR (200 MHz, CDCl<sub>3</sub>): δ = 16.1 (d, <sup>3</sup>*J*<sub>CP</sub> = 7 Hz), 20.3, 31.1, 34.9, 44.9, 52.3, 52.8, 53.1, 62.7, 69.9, 71.0, 71.1, 117.2, 117.4, 124.5 (q, <sup>1</sup>*J*<sub>CF</sub> = 273 Hz), 126.1, 127.7, 127.9, 128.9 (q, <sup>2</sup>*J*<sub>CF</sub> = 31 Hz), 130.0, 130.3, 130.5, 130.8, 134.8, 142.2, 144.6, 154.1, 161.7 (q, <sup>1</sup>*J*<sub>CB</sub> = 50 Hz) (one signal is missing, possibly because of signal overlap); MS (ESI): C<sub>53</sub>H<sub>82</sub>N<sub>4</sub>O<sub>9</sub>P<sup>+</sup> ([**2-H**]<sup>+</sup>) calcd *m/z* 949.5819; found *m/z* 949.5779.

[2]Rotaxane **2-H·PF<sub>6</sub>**: Amberlite® IRA-402 resin (0.21 g) was added to a solution of the [2]rotaxane **2-H·TFPB** (77 mg, 43 μmol) in MeOH (10 mL). The suspension was stirred at room temperature for 5 min and then filtered. The same resin addition/filtration process was applied to the filtrate for another nine cycles. The organic solution was then treated with 1 N HCl<sub>(aq)</sub> (0.1 mL, 0.1 mmol) and saturated NH<sub>4</sub>PF<sub>6(aq)</sub> (20 mL). The organic solvent was evaporated under reduced pressure and the residue was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and H<sub>2</sub>O (10 mL). The organic layer was separated, washed with H<sub>2</sub>O (4 × 10 mL), dried (MgSO<sub>4</sub>), and concentrated; the residue was purified chromatographically (SiO<sub>2</sub>; CH<sub>3</sub>CN/CH<sub>2</sub>Cl<sub>2</sub>,

1:9) to give the product as a white solid (43 mg, 92%). M.p. = 182–183 °C; <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN): δ = 1.23 (t, *J* = 7 Hz, 6H), 1.33 (s, 9H), 2.19 (s, 6H), 2.89–2.98 (br, 8H), 3.25–3.34 (m, 8H), 3.59–3.66 (m, 16H), 3.92–4.00 (m, 4H), 4.09–4.13 (m, 2H), 4.59–4.67 (m, 4H), 6.47 (d, *J* = 8 Hz, 4H), 6.93 (d, *J* = 8 Hz, 4H), 7.44 (d, *J* = 8 Hz, 2H), 7.52 (d, *J* = 8 Hz, 2H), 7.57 (d, *J* = 8 Hz, 2H), 7.63 (d, *J* = 8 Hz, 2H), 7.95–8.10 (br, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 16.7 (<sup>3</sup>*J*<sub>CP</sub> = 7 Hz), 20.4, 31.6, 35.5, 45.5, 53.2, 53.4, 53.5, 63.0 (<sup>2</sup>*J*<sub>CP</sub> = 5 Hz), 70.7, 72.0, 72.1, 117.4, 127.0, 128.9, 129.6, 129.7, 130.8, 131.5, 131.9, 146.6 (<sup>3</sup>*J*<sub>CP</sub> = 5 Hz), 154.3; MS (ESI): C<sub>53</sub>H<sub>82</sub>N<sub>4</sub>O<sub>9</sub>P<sup>+</sup> ([2-H]<sup>+</sup>) calcd *m/z* 949.5819; found *m/z* 949.5788.

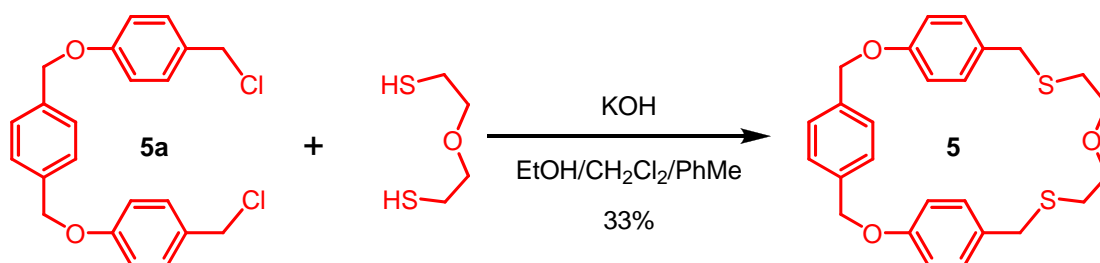
*N*-Benzyl-1-(3,5-di-*tert*-butylphenyl)methanamine (**4**): A mixture of benzylamine (0.5 mL, 4.58 mmol), 3,5-di-*tert*-butylbenzaldehyde (1.0 g, 4.58 mmol), and molecular sieves (1.0 g) in dry MeOH (23 mL) was stirred at room temperature for 16 h before being cooled to 0 °C and treated with NaBH<sub>4</sub> (0.348 g, 9.16 mmol) in portions. The resulting mixture was stirred at room temperature for 4 h and then the organic solvent was evaporated under reduced pressure. The residue was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (100 mL) and H<sub>2</sub>O (100 mL). The organic phase was collected, washed with H<sub>2</sub>O (100 mL), dried (MgSO<sub>4</sub>), and concentrated; the residue was purified chromatographically (SiO<sub>2</sub>; MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 2:98) to give the product as a colorless oil (0.73 g, 51%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.32 (s, 18H), 3.79 (s, 2H), 3.82 (s, 2H), 7.15 (d, *J* = 2 Hz, 2H), 7.29–7.34 (m, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 31.6, 34.8, 53.3, 53.7, 120.7, 122.1, 126.6, 128.0, 128.1, 139.1, 140.2, 150.4; MS (ESI): C<sub>22</sub>H<sub>32</sub>N<sup>+</sup> ([4-H]<sup>+</sup>) calcd *m/z* 310.2535; found *m/z* 310.2567.

**4-H**·PF<sub>6</sub>: A solution of the amine **4** (216 mg, 0.704 mmol) in MeCN (10 mL) was treated with 1 N HCl<sub>(aq)</sub> (5 mL) and saturated NH<sub>4</sub>PF<sub>6(aq)</sub> (10 mL). After evaporating the organic solvent under reduced pressure, the white precipitate was filtered off, washed with water (5 mL) and ether (2 mL), and then dried to give a white solid (232 mg, 72%). M.p. = 173–178 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.31 (s, 18H), 4.12–4.20 (br, 4H), 7.14 (d, *J* = 2 Hz, 2H), 7.29–7.36 (br, 2H), 7.38–7.44 (br, 3H), 7.45–7.49 (br, 1H); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>CN): δ = 31.6, 35.6, 52.4, 52.8, 124.4, 125.2, 129.8, 130.5, 130.5, 131.0, 131.2, 152.5; MS (ESI): C<sub>22</sub>H<sub>32</sub>N<sup>+</sup> ([4-H]<sup>+</sup>) calcd *m/z* 310.2535; found *m/z* 310.2562.

**4-H**·TFPB: A solution of the amine **4** (290 mg, 0.945 mmol) in MeOH (10 mL) was treated with 1 N HCl<sub>(aq)</sub> (10 mL). The organic solvent was evaporated under reduced pressure and then the precipitate was filtered off, dissolved in MeOH (10 mL), and treated with NaTFPB (256 mg, 0.289 mmol). After evaporating the organic solvent

under reduced pressure, the residue was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and H<sub>2</sub>O (50 mL). The organic layer was collected, washed with H<sub>2</sub>O (4 × 50 mL), dried (MgSO<sub>4</sub>), and concentrated to give a pale green liquid (194 mg, 57%). M.p. > 66 °C (dec.); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.29 (s, 18H), 4.08–4.16 (m, 4H), 7.06 (d, *J* = 2 Hz, 2H), 7.17 (d, *J* = 7 Hz, 2H), 7.39 (t, *J* = 7 Hz, 2H), 7.46 (d, *J* = 7 Hz, 1H), 7.48–7.52 (br, 4H), 7.56 (t, *J* = 2 Hz, 1H), 7.65–7.72 (br, 8H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 31.2, 35.0, 52.2, 53.0, 117.4, 124.5 (q, <sup>1</sup>*J*<sub>CF</sub> = 272 Hz), 123.2, 125.1, 127.5, 128.1, 128.8 (q, <sup>2</sup>*J*<sub>CF</sub> = 32 Hz), 129.0, 129.9, 130.9, 134.7, 153.1, 161.5 (q, <sup>1</sup>*J*<sub>CB</sub> = 50 Hz); MS (ESI): C<sub>22</sub>H<sub>32</sub>N<sup>+</sup> ([**4-H**]<sup>+</sup>) calcd *m/z* 310.2535; found *m/z* 310.2525.

### Macrocycle **5**



A solution of the dichloride **5a** (3.12 g, 8 mmol) and bis(2-mercaptoethyl) ether (1 mL, 8 mmol) in a mixture of CH<sub>2</sub>Cl<sub>2</sub> (100 mL) and toluene (700 mL) was added slowly (addition funnel) to a solution of KOH (896 mg, 16.0 mmol) in EtOH (2.4 L). The mixture was then stirred at room temperature for 7 days. The organic solvents were evaporated under reduced pressure and then the residue was purified chromatographically (SiO<sub>2</sub>; EtOAc/hexanes, 2:8) to give a white solid (1.2 g, 33%). M.p. = 129–130 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 2.42 (t, *J* = 7 Hz, 4H), 3.47 (t, *J* = 7 Hz, 4H), 3.63 (s, 4H), 5.17 (s, 4H), 6.67 (d, *J* = 8 Hz, 4H), 7.04 (d, *J* = 8 Hz, 4H), 7.30 (s, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 29.6, 35.5, 69.5, 70.5, 115.5, 127.0, 129.6, 130.0, 136.9, 156.6; MS (ESI): C<sub>26</sub>H<sub>28</sub>NaO<sub>3</sub>S<sub>2</sub><sup>+</sup> ([**5** + Na]<sup>+</sup>) calcd *m/z* 475.1378; found *m/z* 475.1343.

Threadlike salt **6-H**·TFPB: A solution of **6-H**·PF<sub>6</sub> (300 mg, 0.676 mmol) and NaTFPB (599 mg, 0.676 mmol) in MeOH (20 mL) was treated with H<sub>2</sub>O (10 mL). The mixture was stirred at room temperature for 5 min and then the organic solvent was evaporated under reduced pressure. The solution was partitioned between CH<sub>2</sub>Cl<sub>2</sub> (50 mL) and H<sub>2</sub>O (150 mL). The organic layer was collected, washed with H<sub>2</sub>O (4 × 150 mL), dried (MgSO<sub>4</sub>), and concentrated to give a deep green oil (707 mg, 90%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 0.60 (q, *J* = 6 Hz, 1H), 1.13–1.22 (m, 2H),

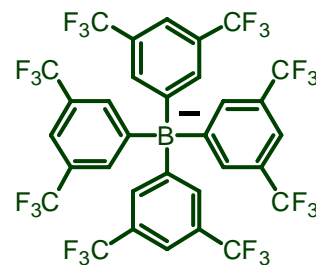
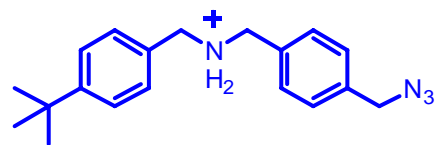
1.29 (s, 9H), 1.66–1.82 (m, 4H), 2.16–2.34 (m, 2H), 2.91–3.07 (m, 2H), 4.00 (t,  $J = 6$  Hz, 2H), 4.94 (dd,  $J = 2, 10$  Hz, 1H), 5.08 (dd,  $J = 1, 17$  Hz, 1H), 5.16–5.67 (m, 2H), 6.73–6.93 (br, 2H), 7.18 (d,  $J = 8$  Hz, 2H), 7.45 (d,  $J = 8$  Hz, 2H), 7.50–7.53 (br, 4H), 7.67–7.76 (br, 8H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 15.1, 17.1, 22.9, 24.3, 25.1, 31.0, 34.9, 48.1, 52.2, 115.1, 117.4, 124.4$  (q,  $^1J_{\text{CF}} = 272$  Hz), 125.1, 126.7, 127.0, 128.8 (q,  $^2J_{\text{CF}} = 27$  Hz), 132.7, 134.6, 137.4, 154.4, 161.5 (q,  $^1J_{\text{CB}} = 50$  Hz); MS (ESI):  $\text{C}_{21}\text{H}_{32}\text{N}^+$  ( $[\mathbf{6}\text{-H}]^+$ ) calcd  $m/z$  298.2535; found  $m/z$  298.2482.

[2]Rotaxane **7**-H·TFPB and dumbbell-shaped salt **8**-H·TFPB: A solution of the threadlike salt **6**-H·TFPB (466 mg, 0.401 mmol) and the macrocycle **5** (363 mg, 0.802 mmol) in  $\text{CDCl}_3$  (2.0 mL) was heated at 50 °C with stirring for 48 h. After evaporating the organic solvent under reduced pressure, the residue was purified chromatographically ( $\text{SiO}_2$ ;  $\text{CH}_2\text{Cl}_2$ /hexanes, 8:2) to give the [2]rotaxane **7**-H·TFPB, which solidified in hexane to give a white powder (193 mg, 30%). The corresponding dumbbell-shaped salt **8**-H·TFPB was isolated as a colorless liquid (106 mg, 23%) after two sequential chromatographic purification processes ( $\text{SiO}_2$ ;  $\text{CH}_2\text{Cl}_2$ /MeOH, 9:1 and then  $\text{CH}_2\text{Cl}_2$ /MeOH 98:2). Data for **7**-H·TFPB: m.p. = 125–127 °C;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta = 0.25\text{--}0.75$  (m, 6H), 1.37 (s, 9H), 1.95–2.26 (m, 3H), 2.65–2.85 (m, 5H), 2.92–3.09 (m, 3H), 3.36–3.62 (m, 8H), 5.08–5.25 (m, 4H), 5.38 (dd,  $J = 5, 11$  Hz, 1H), 5.64–5.79 (m, 3H), 6.54–6.86 (several overlapping broad peaks, 10H), 6.99 (d,  $J = 8$  Hz, 2H), 7.43 (s, 4H), 7.48–7.52 (br, 4H), 7.56 (d,  $J = 8$  Hz, 2H), 7.65–7.73 (br, 8H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta = 24.2, 29.0, 31.1, 32.1, 32.4, 33.8, 34.9, 36.3, 37.0, 47.1, 51.6, 67.7, 69.0, 117.4, 124.6$  (q,  $^1J_{\text{CF}} = 272$  Hz), 125.7, 126.4, 128.5, 128.6, 128.7, 128.9 (q,  $^2J_{\text{CF}} = 31$  Hz), 129.2, 129.4, 131.3, 134.0, 137.3, 137.4, 154.7, 156.9, 161.7 (q,  $^1J_{\text{CB}} = 50$  Hz) (two signals missing, possibly because of signal overlap); MS (ESI):  $\text{C}_{47}\text{H}_{60}\text{NO}_3\text{S}_2^+$  ( $[\mathbf{7}\text{-H}]^+$ ) calcd  $m/z$  750.4014; found  $m/z$  750.3990. Data for **8**-H·TFPB:  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 1.26\text{--}1.40$  (m, 11H), 1.50–1.65 (m, 2H), 1.99–2.10 (m, 1H), 2.13–2.24 (m, 1H) 2.35–2.48 (br, 1H), 2.62–2.73 (m, 1H), 2.80 (t,  $J = 8$  Hz, 2H), 2.86–2.98 (m, 1H), 3.65–3.80 (br, 2H), 3.83 (s, 2H), 5.39–5.47 (m, 1H), 5.55–5.70 (m, 3H), 7.08 (d,  $J = 8$  Hz, 2H), 7.43 (d,  $J = 8$  Hz, 2H), 7.52 (s, 4H), 7.69 (s, 8H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 24.7, 28.6, 31.1, 32.6, 32.6, 34.9, 36.7, 48.4, 52.4, 117.4, 124.4$  (q,  $^1J_{\text{CF}} = 271$  Hz), 126.8, 127.1, 128.5, 128.6, 128.8 (q,  $^2J_{\text{CF}} = 32$  Hz), 129.1, 133.9, 134.7, 154.1, 161.5 (q,  $^1J_{\text{CB}} = 50$  Hz) (one signal missing, possibly because of signal overlap); MS (ESI):  $\text{C}_{21}\text{H}_{32}\text{N}^+$  ( $[\mathbf{8}\text{-H}]^+$ ) calcd  $m/z$  298.2535; found  $m/z$  298.2518.

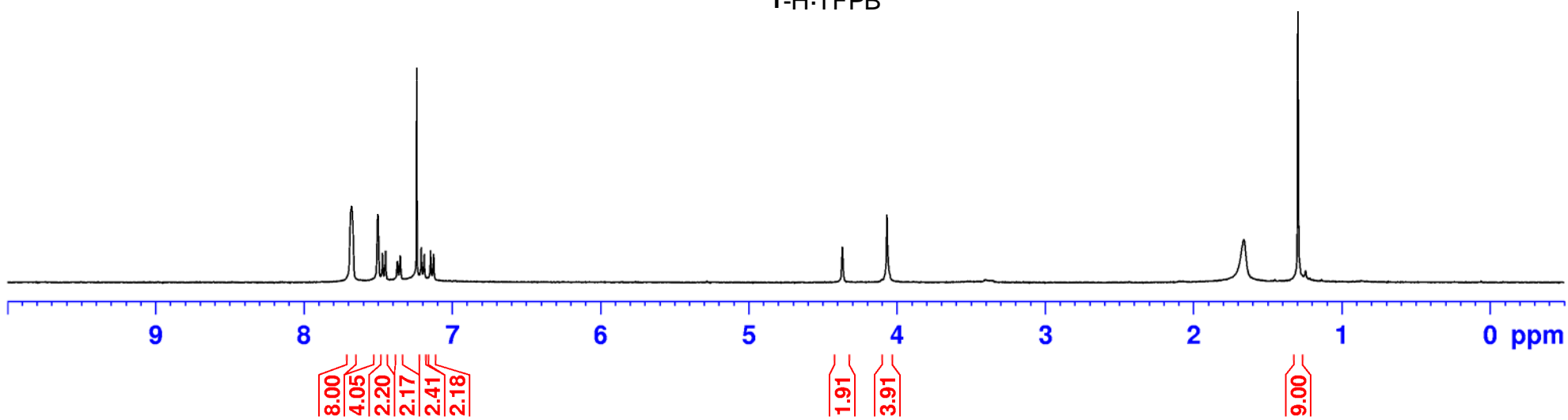
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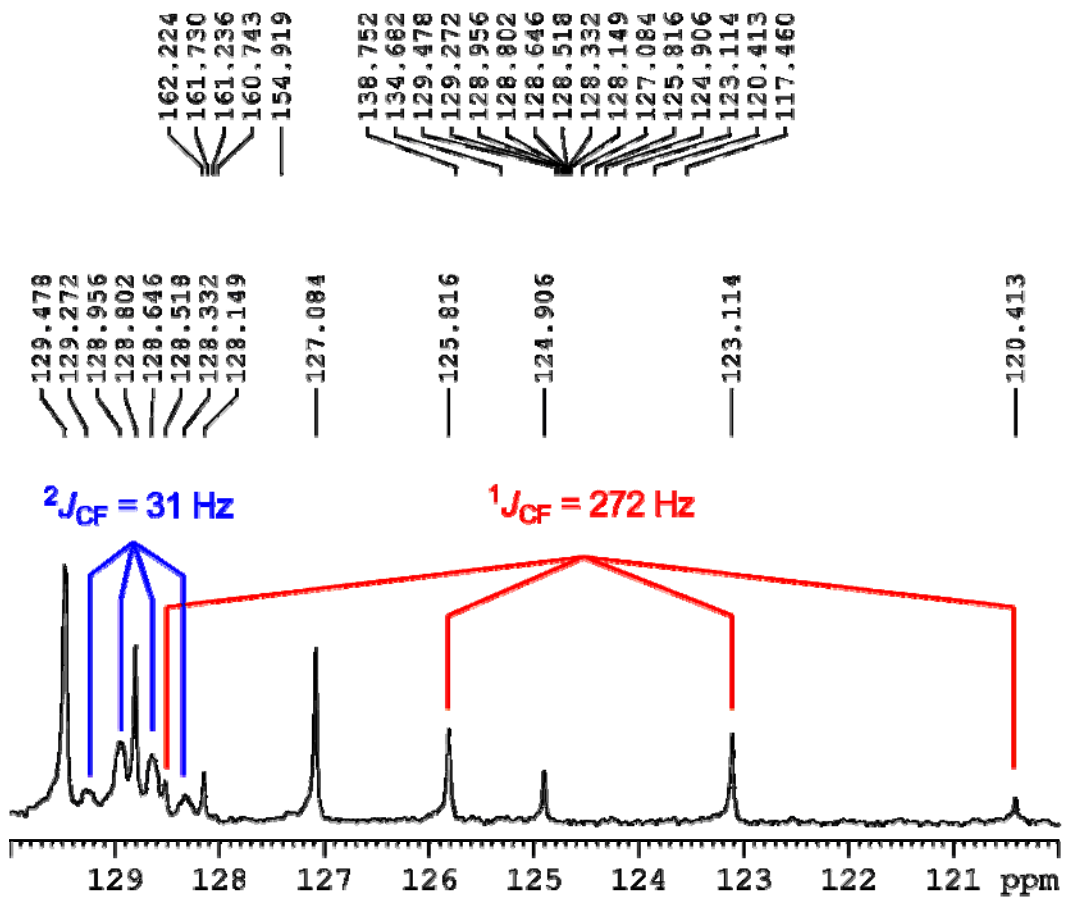
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1.662  
1.297



1-H·TFPB





162.224  
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161.236  
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134.682  
129.478  
129.272  
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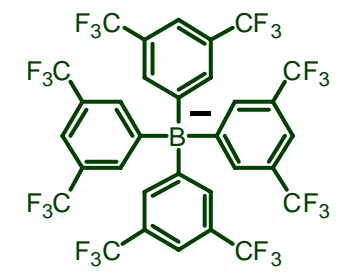
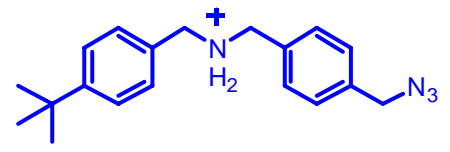
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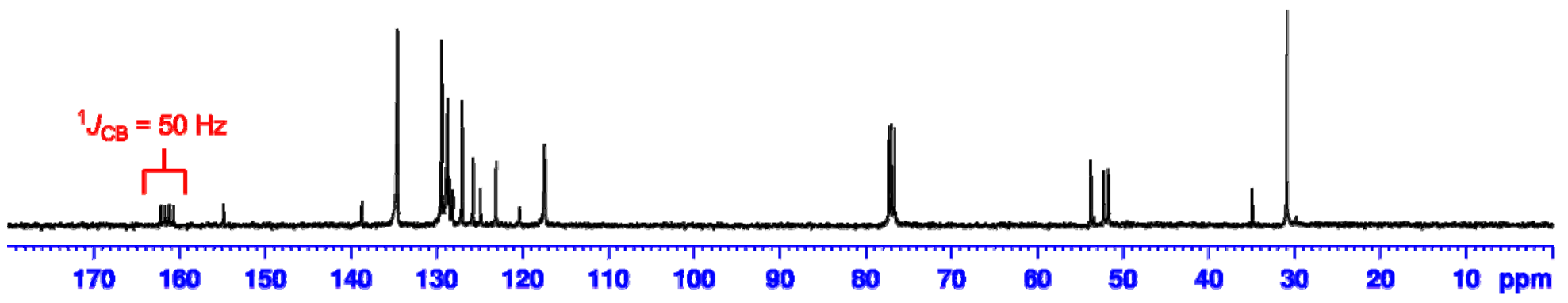
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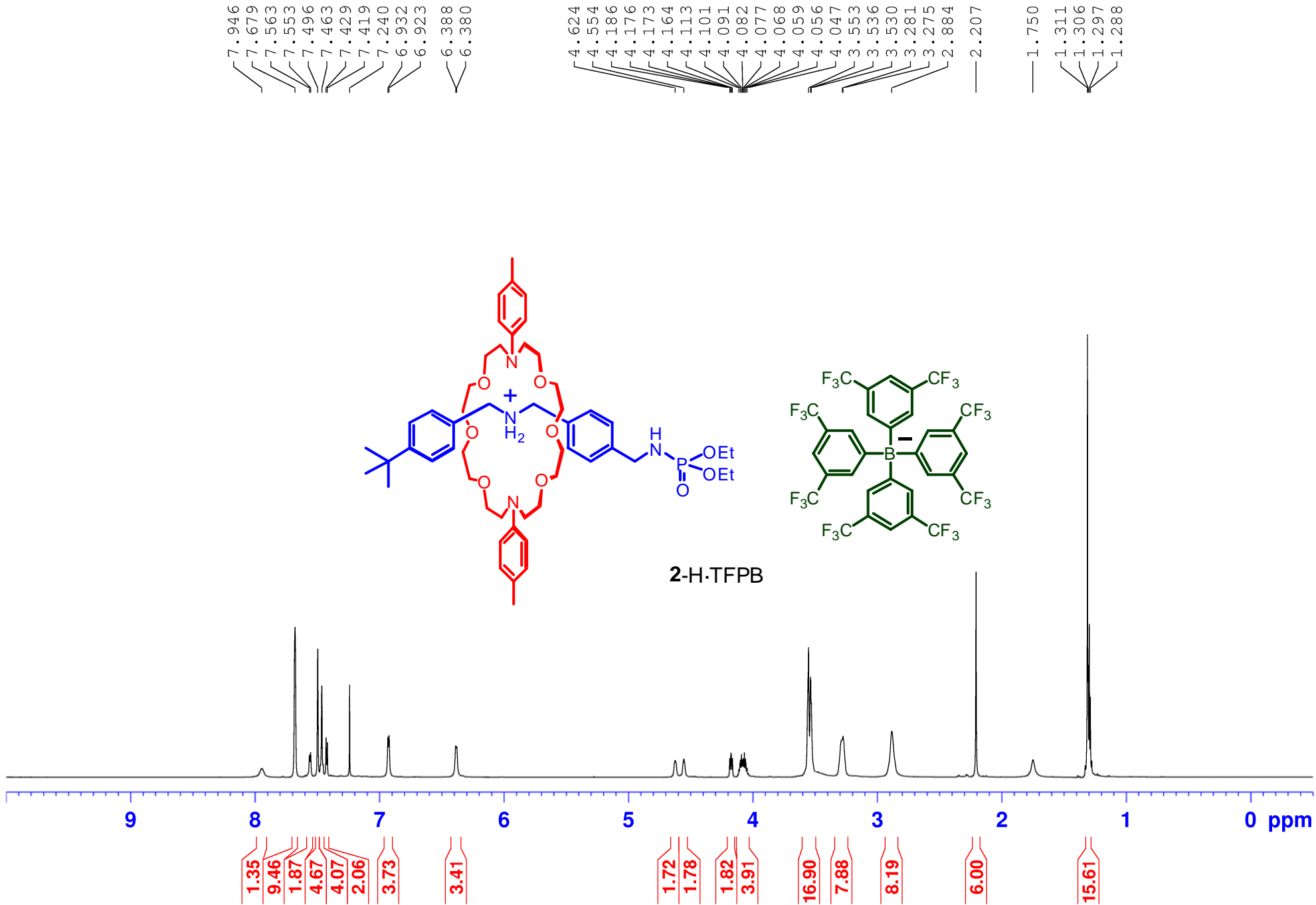


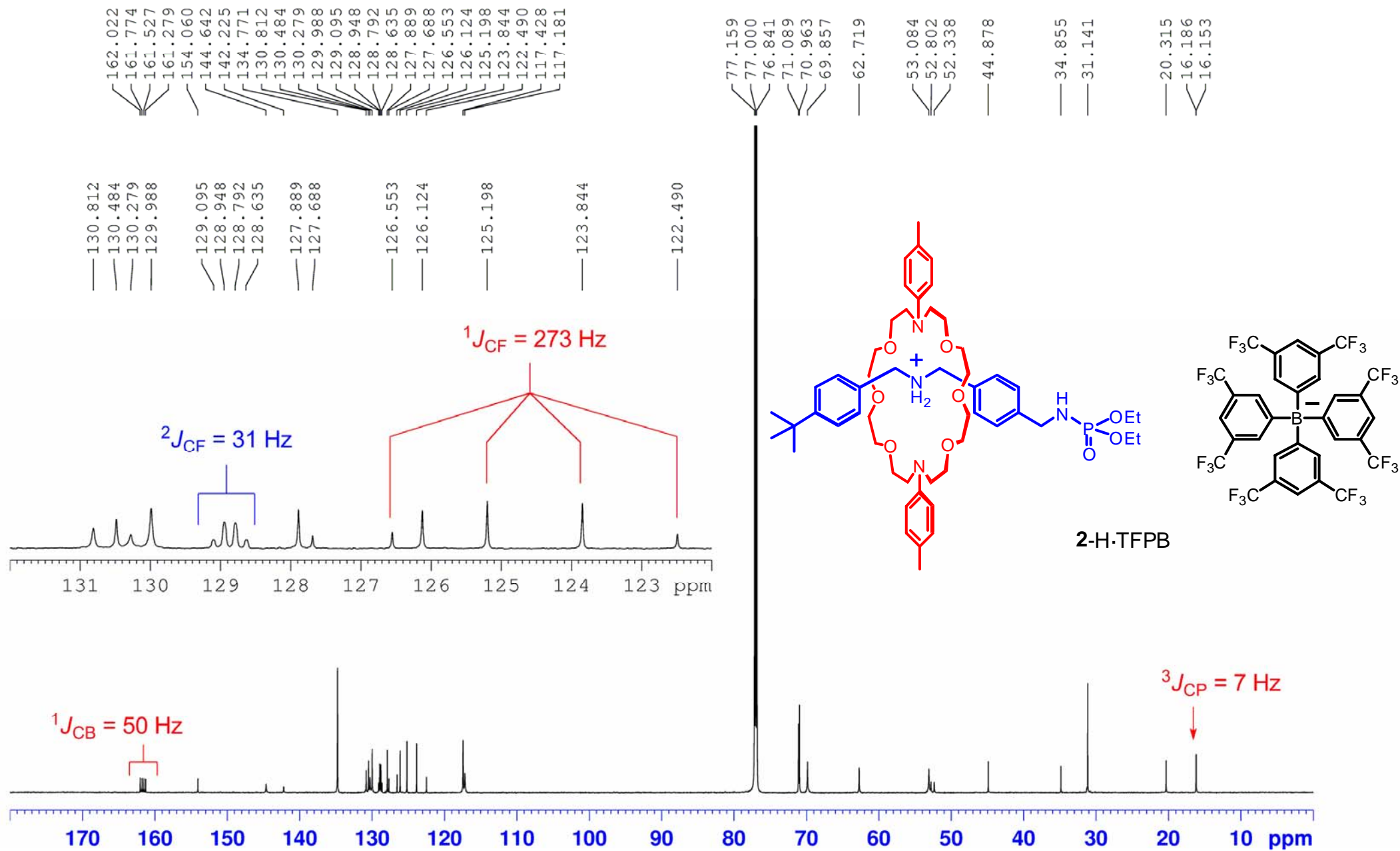
1-H.TFPB

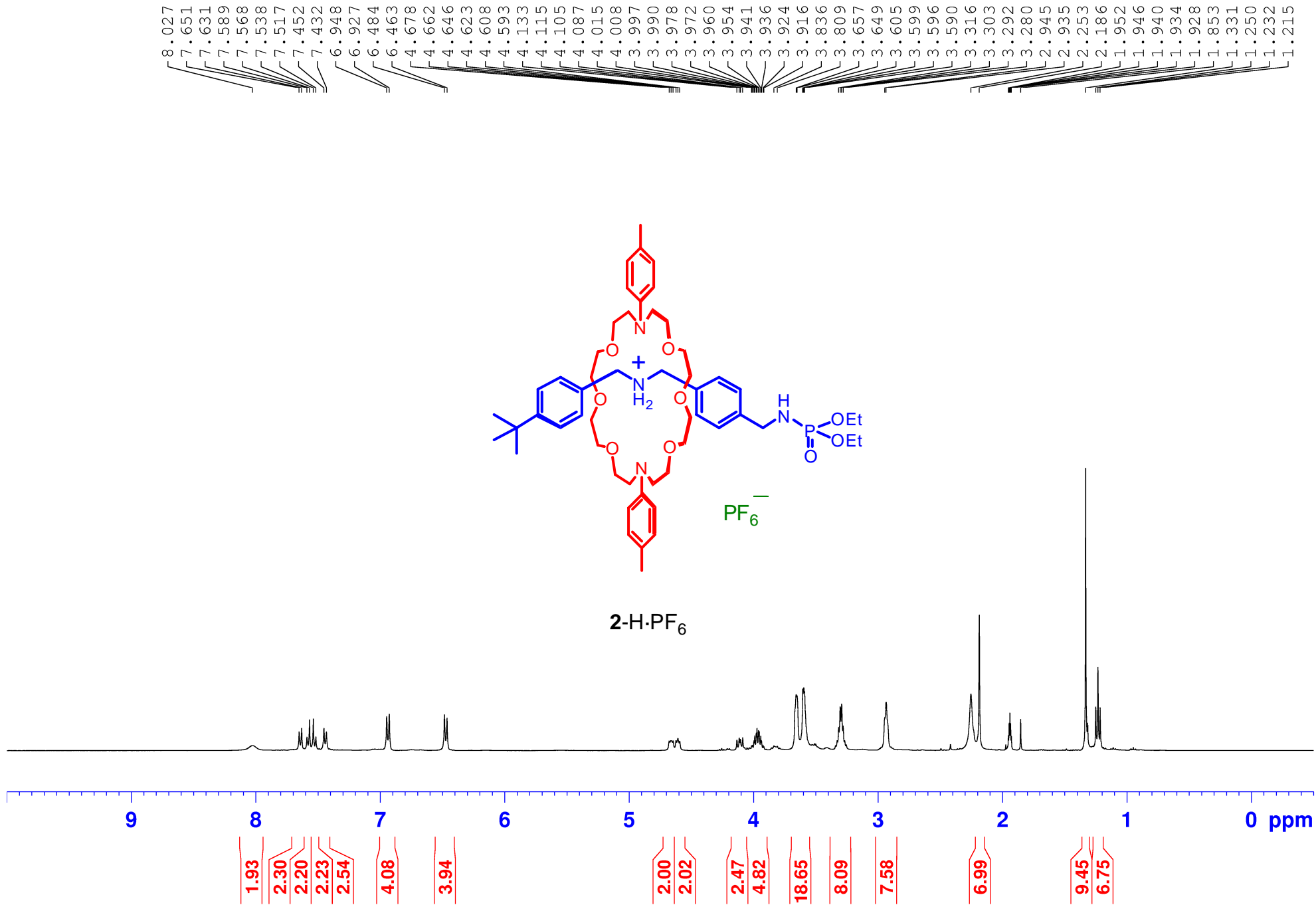


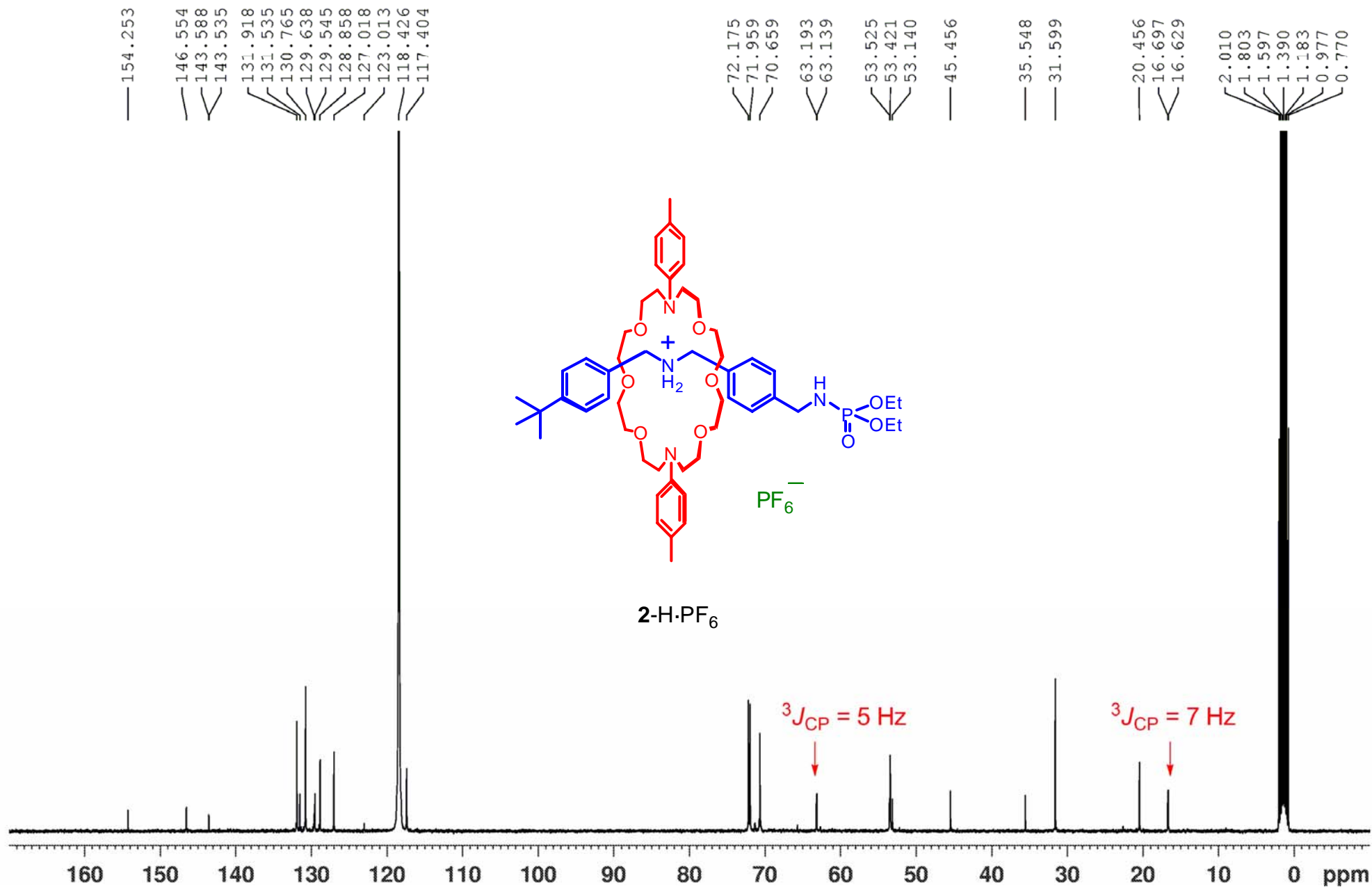
170  
180  
150  
140  
130  
120  
110  
100  
90  
80  
70  
60  
50  
40  
30  
20  
10 ppm

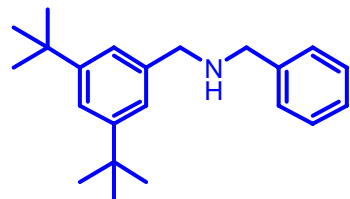




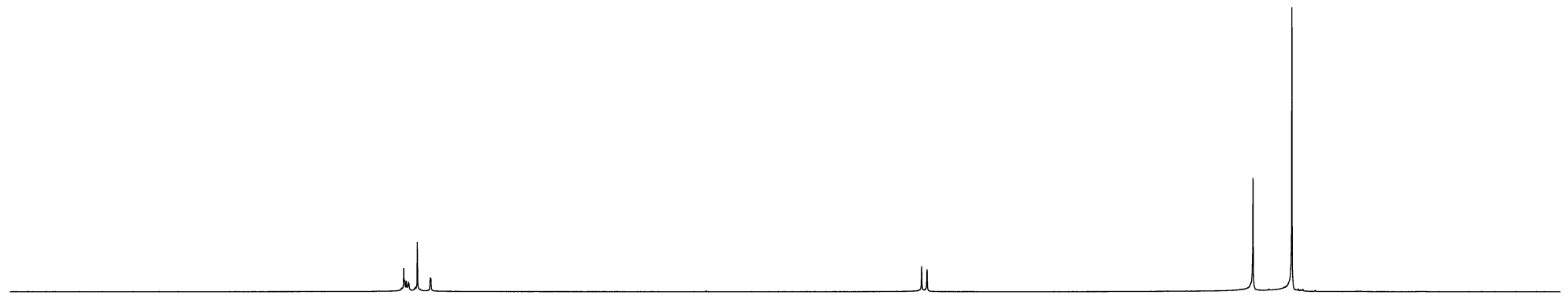








4



7.332  
7.327  
7.315  
7.305  
7.301  
7.296  
7.240  
7.153  
7.149

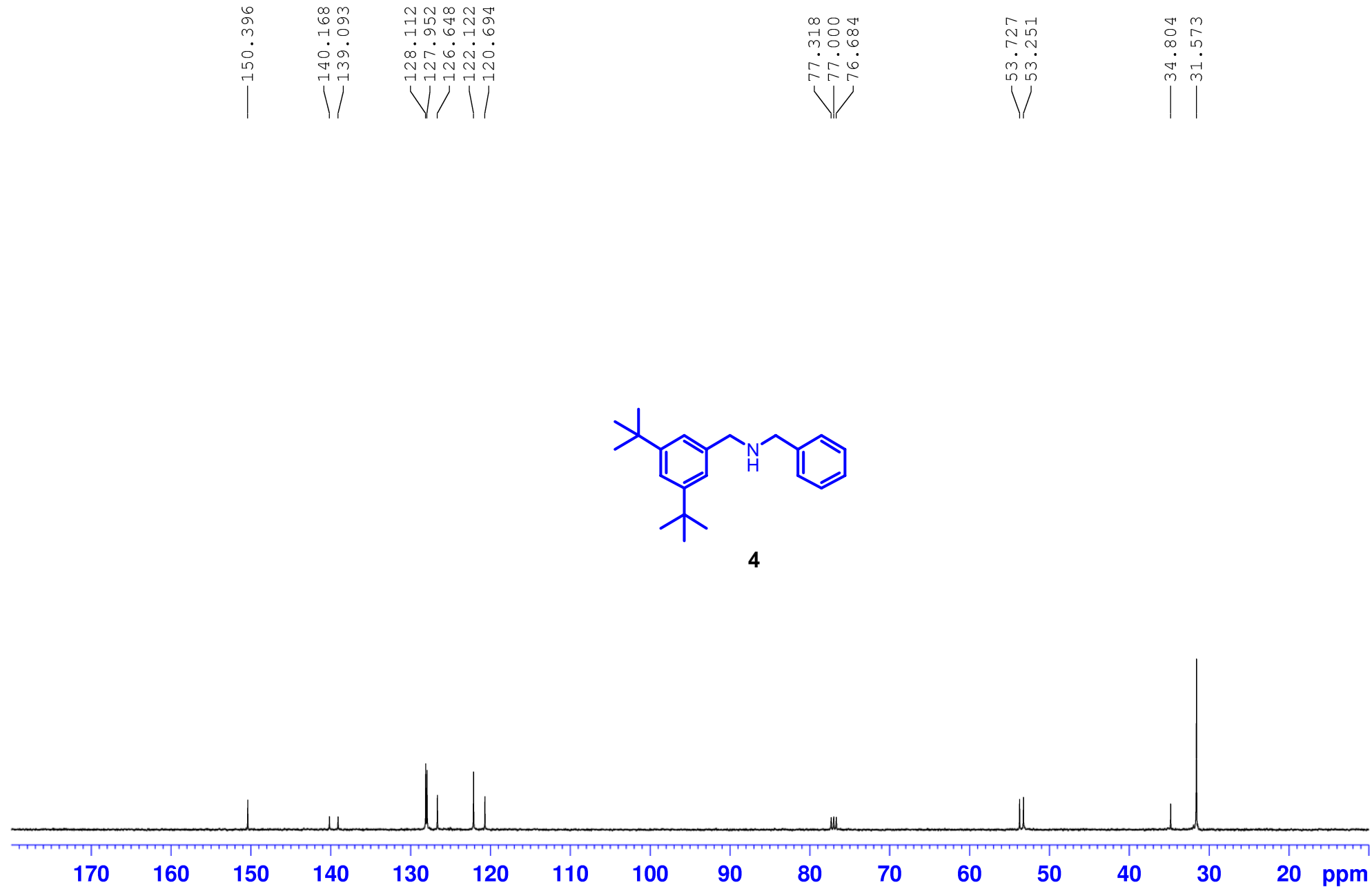
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3.788

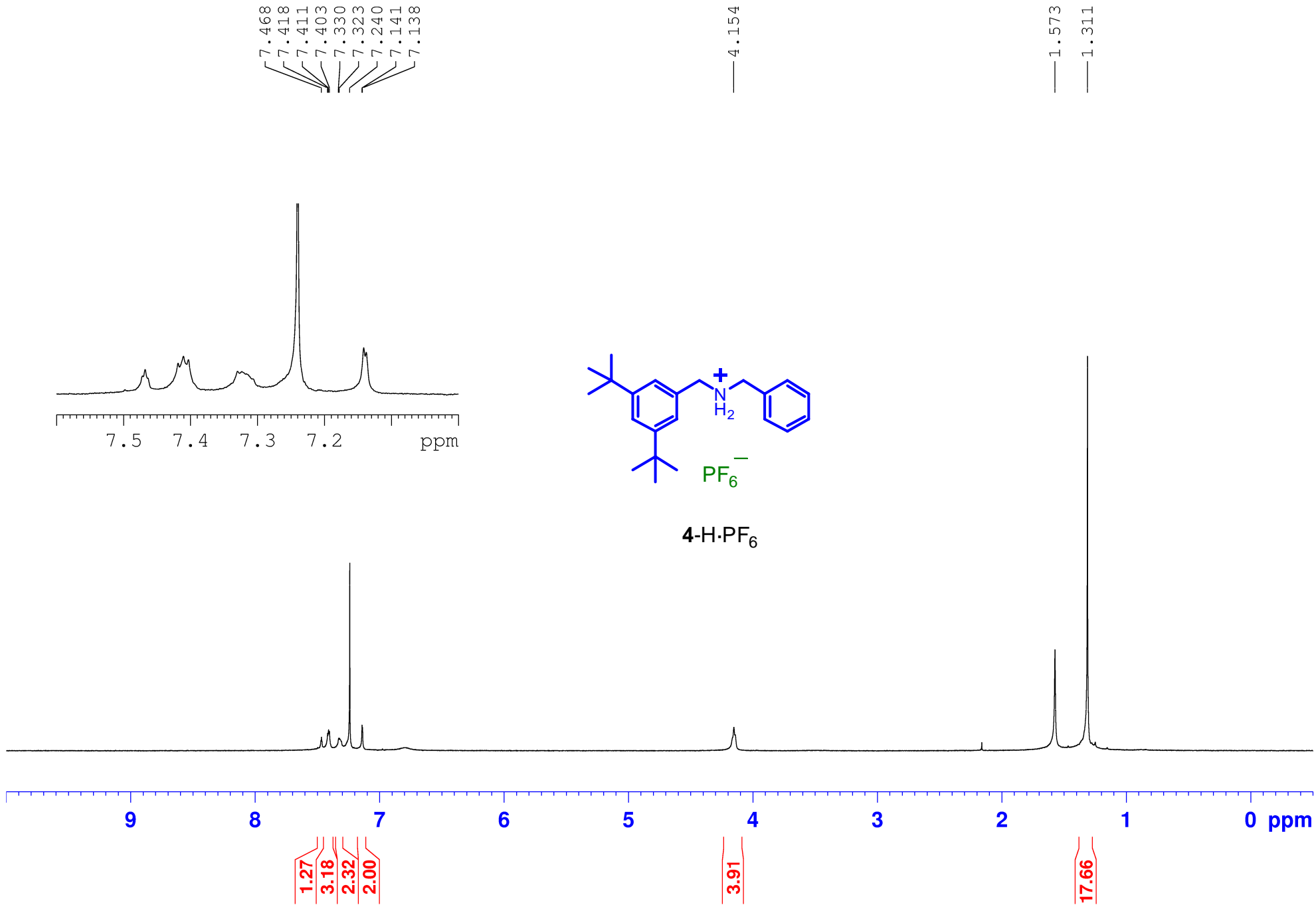
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1.319

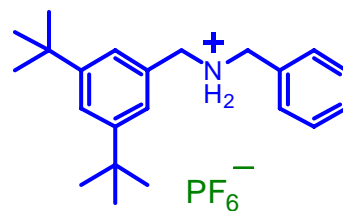
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1.99

2.05  
2.01

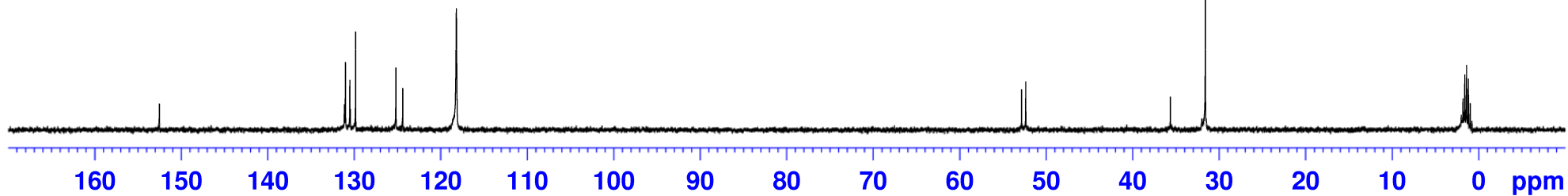
18.49







4-H·PF<sub>6</sub>



— 152.519

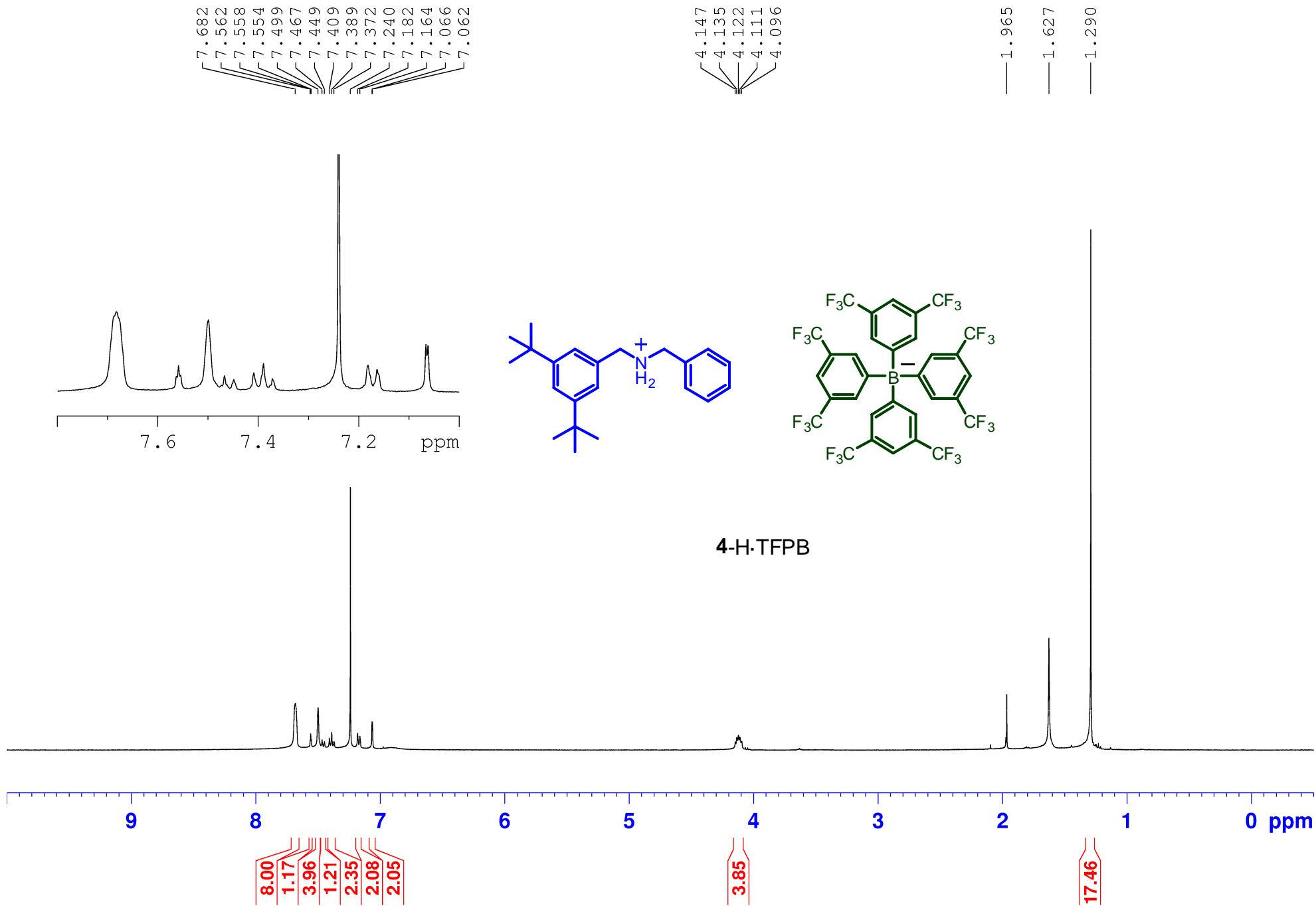
131.152  
131.031  
130.517  
130.471  
129.848  
125.187  
124.404  
— 118.199

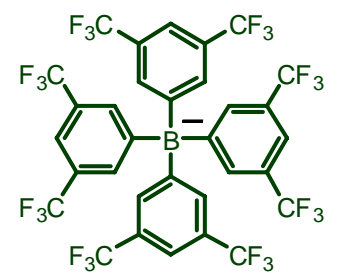
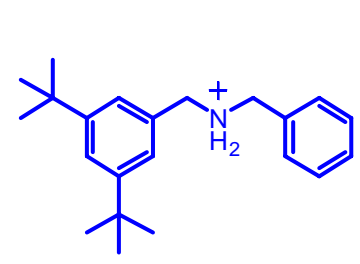
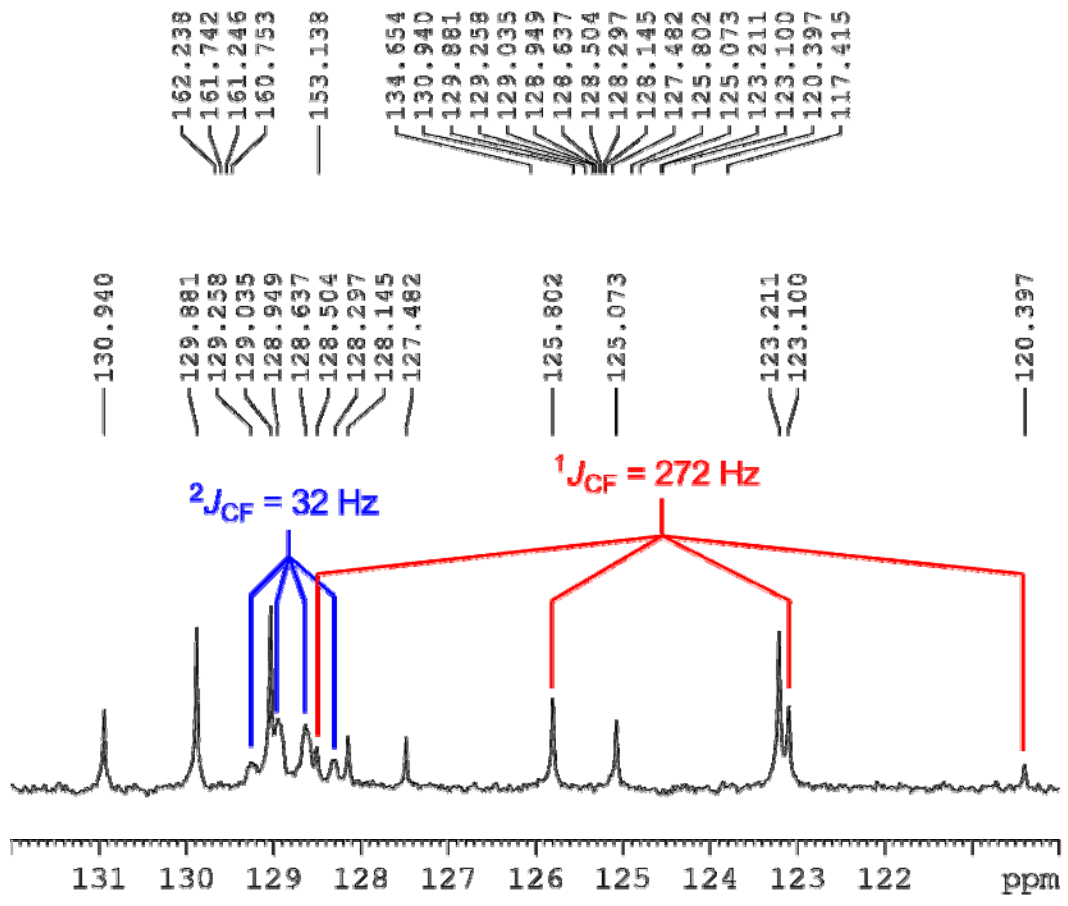
52.846  
52.363

— 35.647  
— 31.595

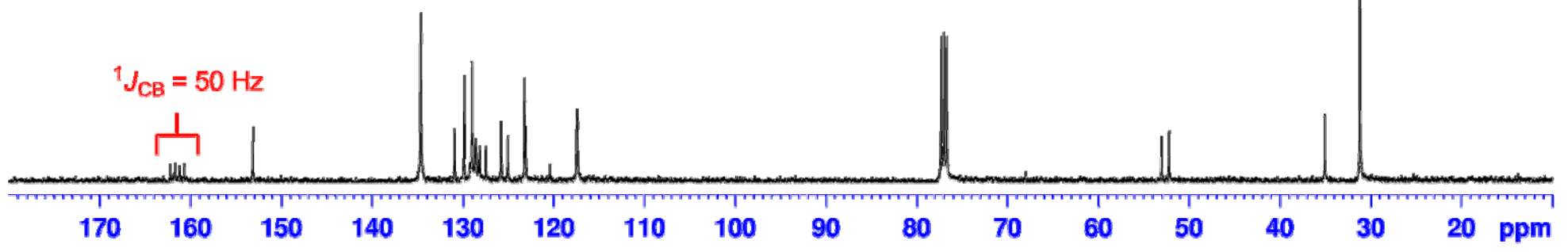
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1.802  
1.593  
1.390  
1.184  
0.976  
0.774

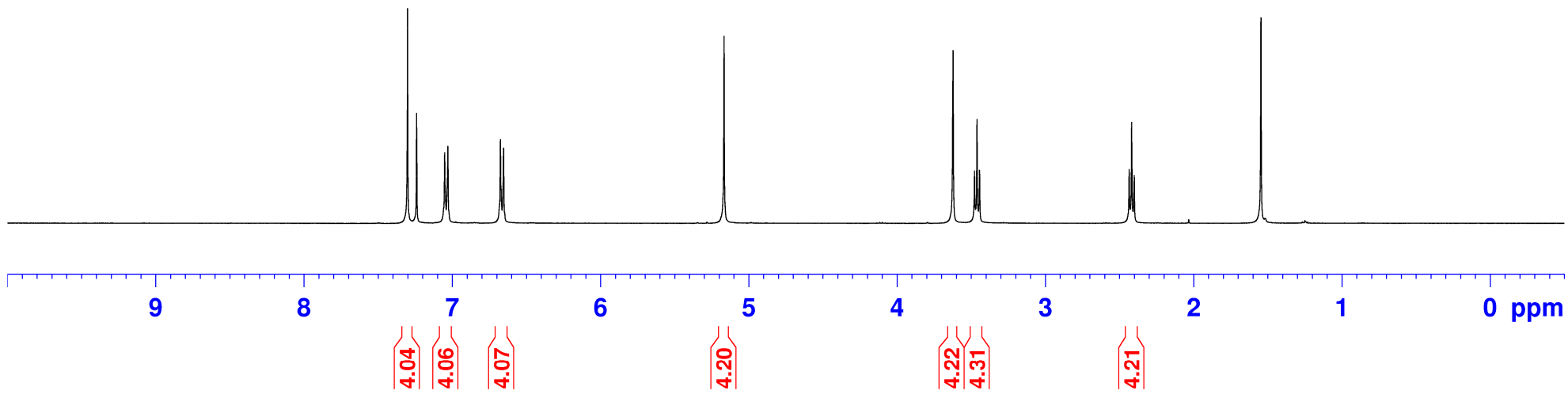
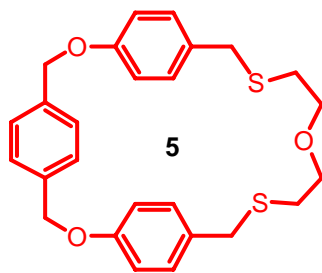






4-H-TFPB





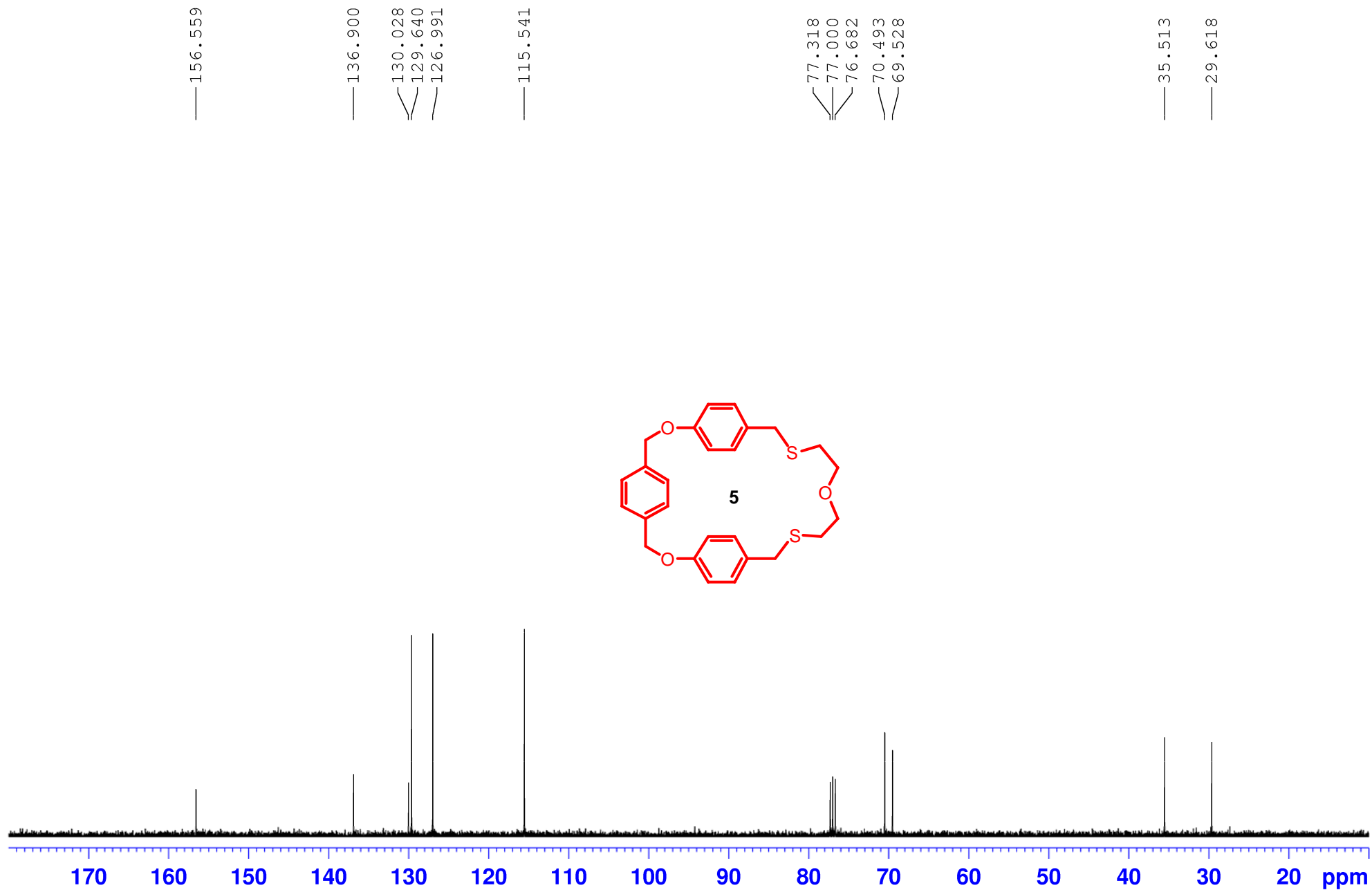
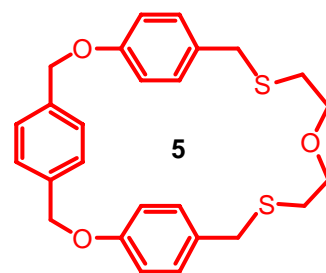
7.301  
7.240  
7.051  
7.030  
6.675  
6.654

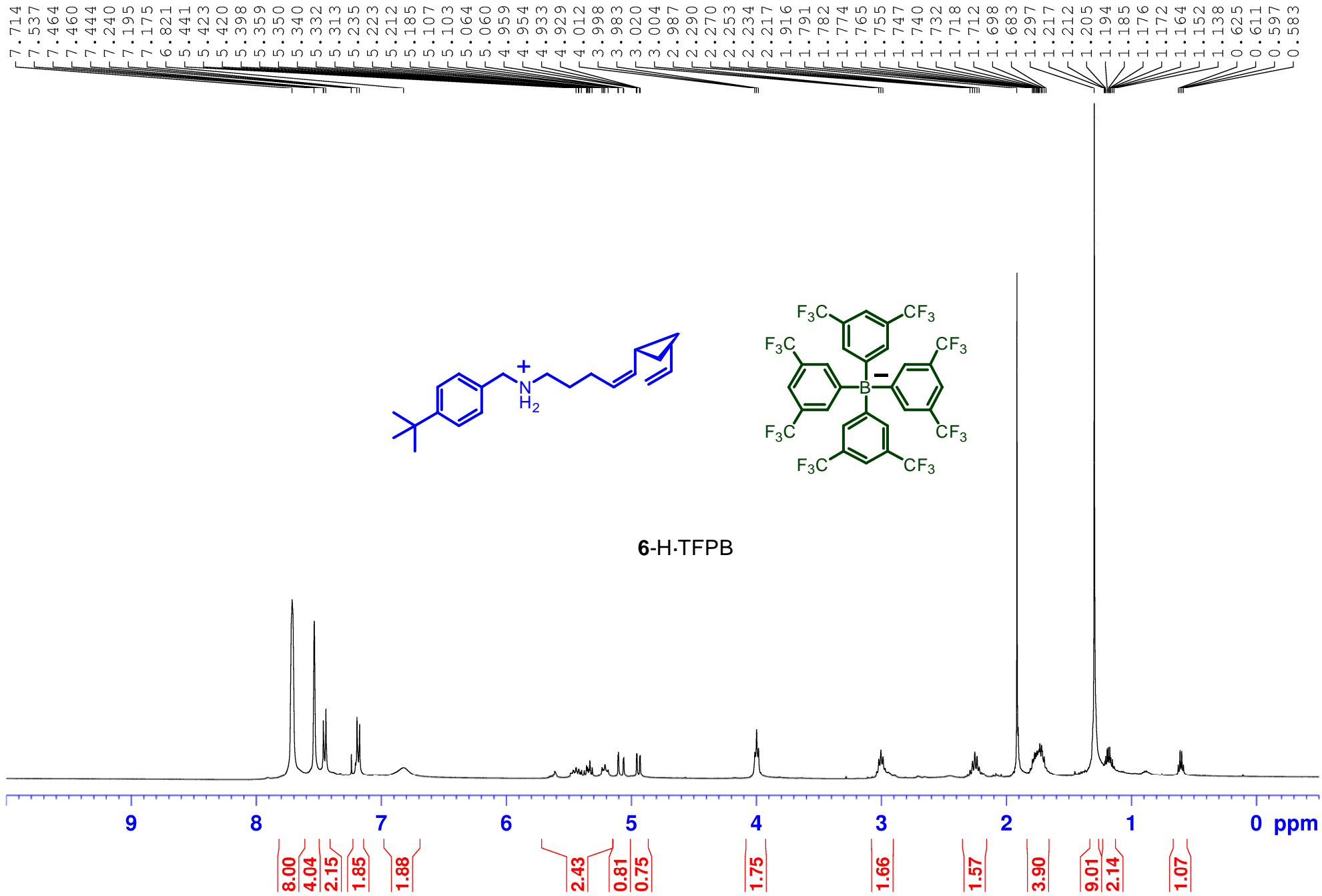
5.167

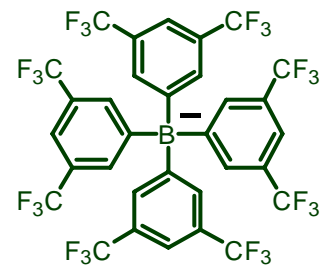
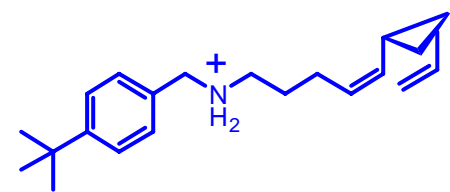
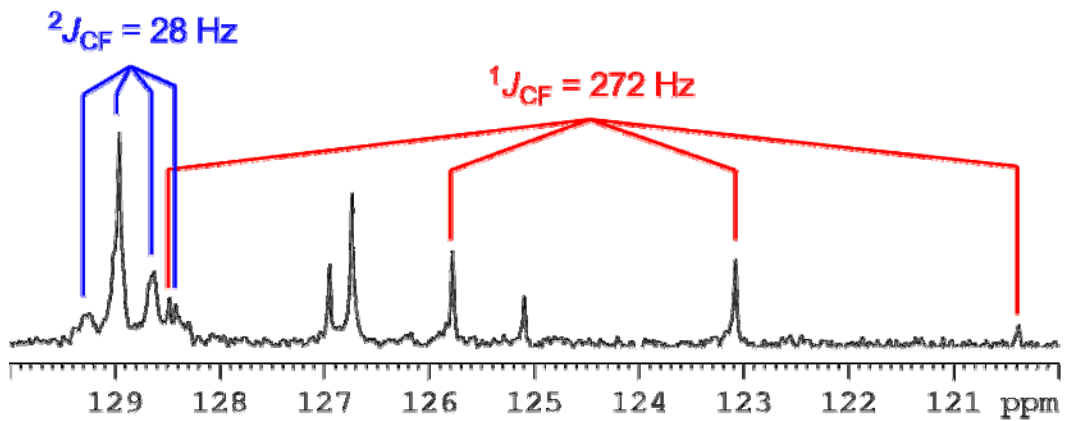
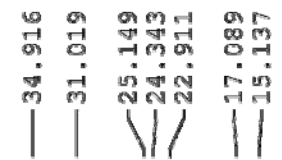
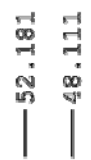
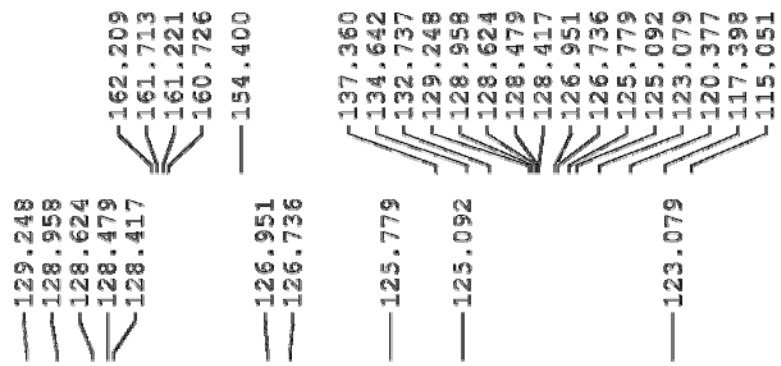
3.624  
3.479  
3.462  
3.445

2.436  
2.419  
2.402

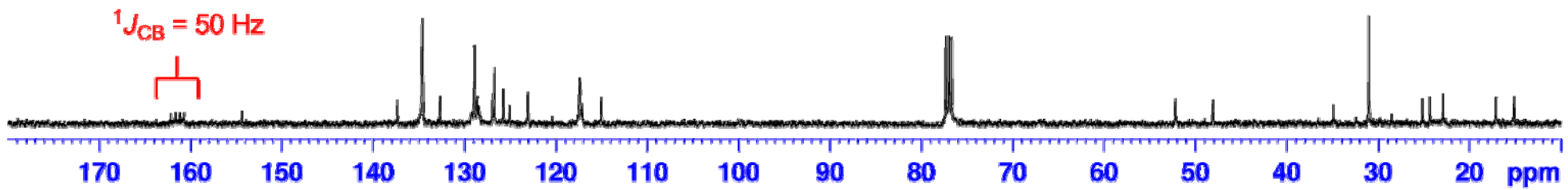
1.547



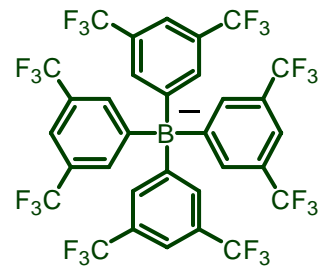
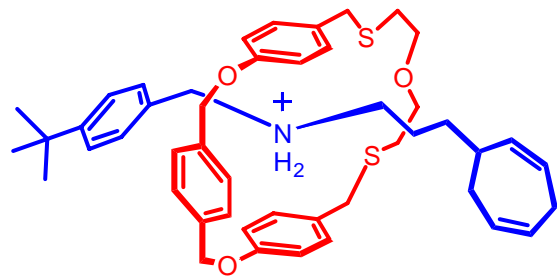




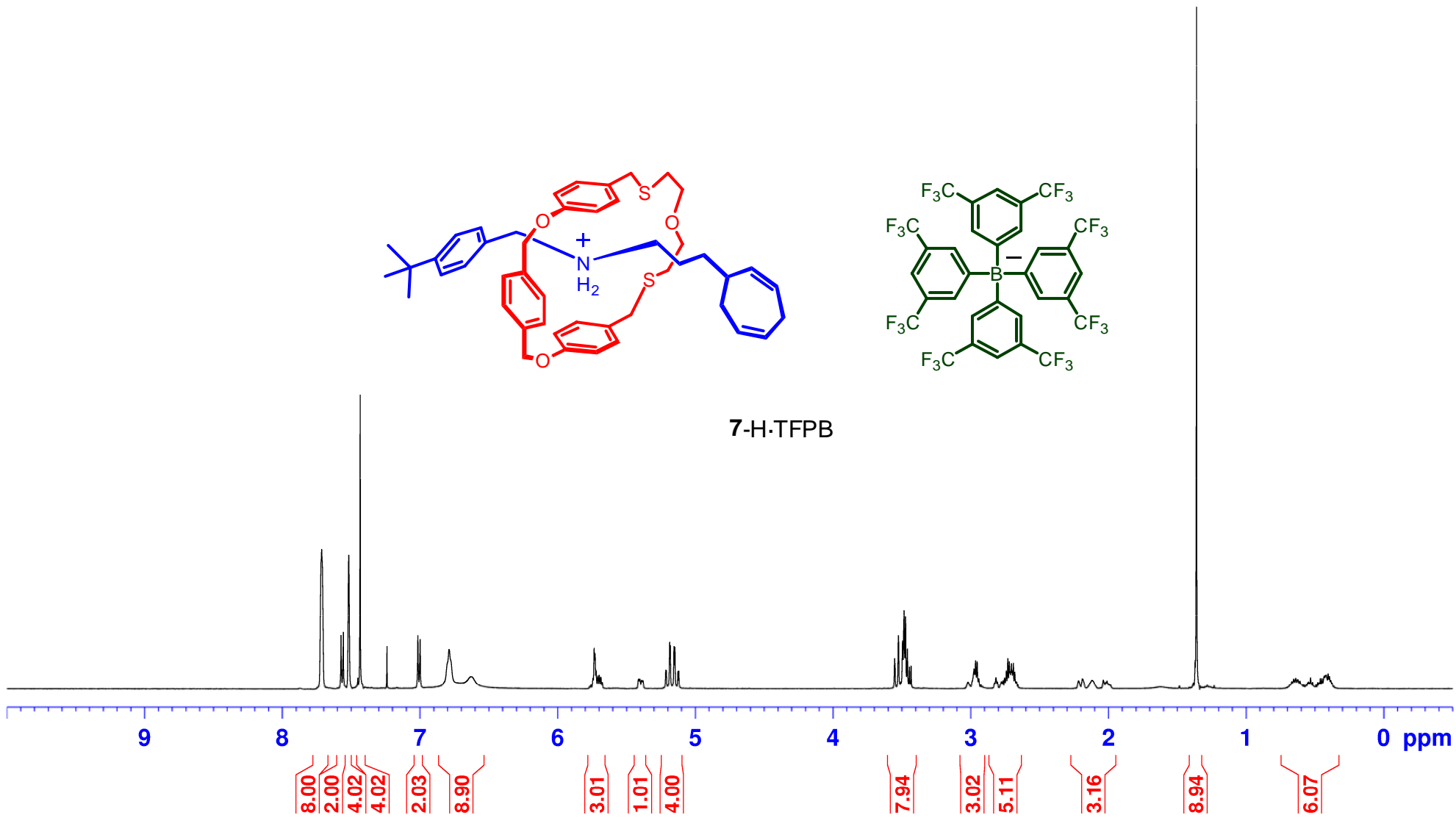
6-H-TFPB

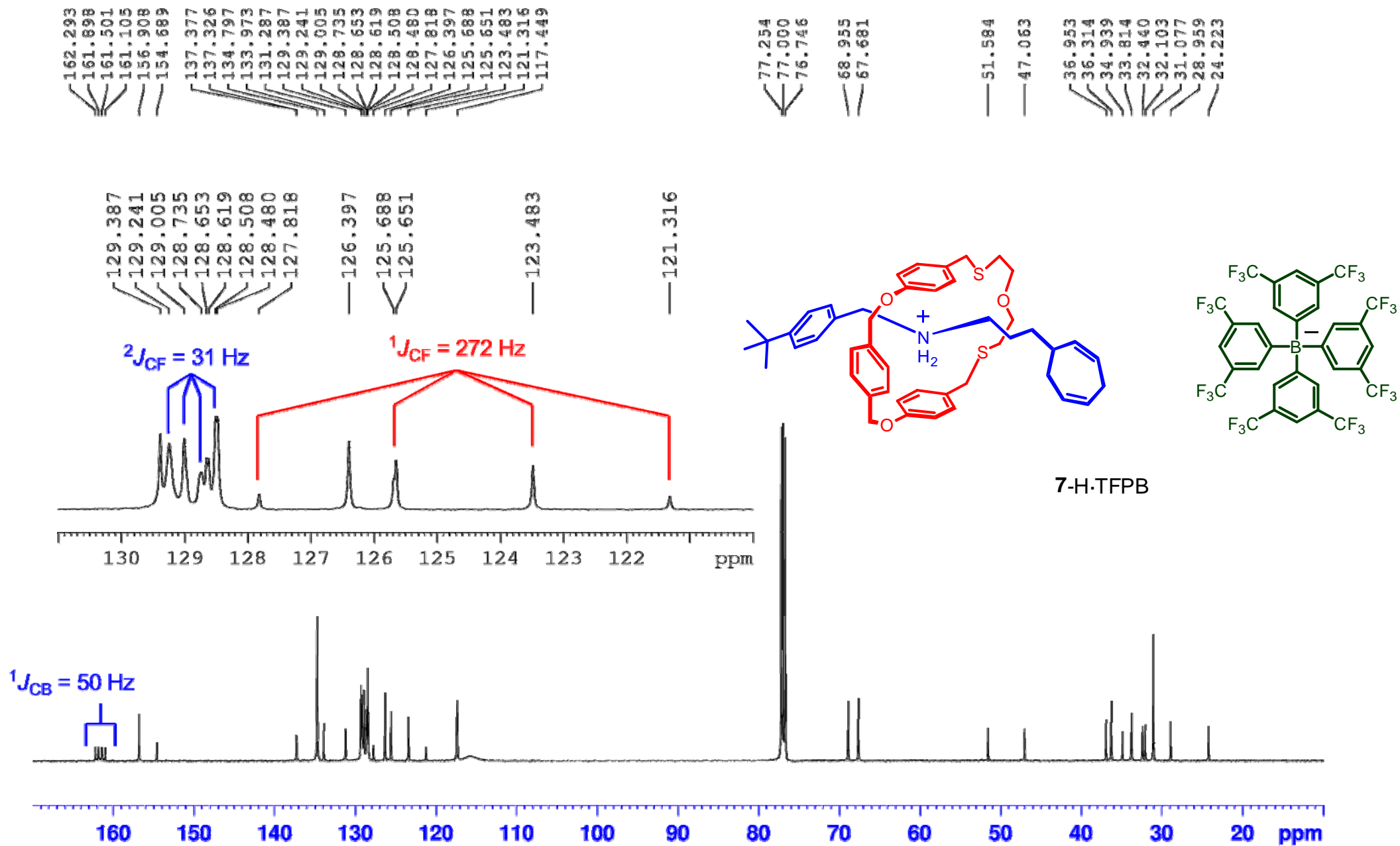


7.715  
7.574  
7.557  
7.518  
7.435  
7.240  
7.016  
6.999  
6.788  
6.627  
5.743  
5.735  
5.729  
5.721  
5.708  
5.698  
5.686  
5.413  
5.404  
5.390  
5.215  
5.211  
5.186  
5.182  
5.155  
5.149  
5.127  
5.120  
3.553  
3.550  
3.526  
3.523  
3.495  
3.484  
3.474  
3.460  
3.445  
3.433  
2.976  
2.964  
2.953  
2.941  
2.817  
2.755  
2.741  
2.730  
2.720  
2.714  
2.708  
2.704  
2.700  
2.690  
2.680  
2.189  
2.185  
2.118  
2.037  
1.359  
0.653  
0.639  
0.624  
0.531  
0.460  
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0.431  
0.417  
0.404  
0.394  
0.385

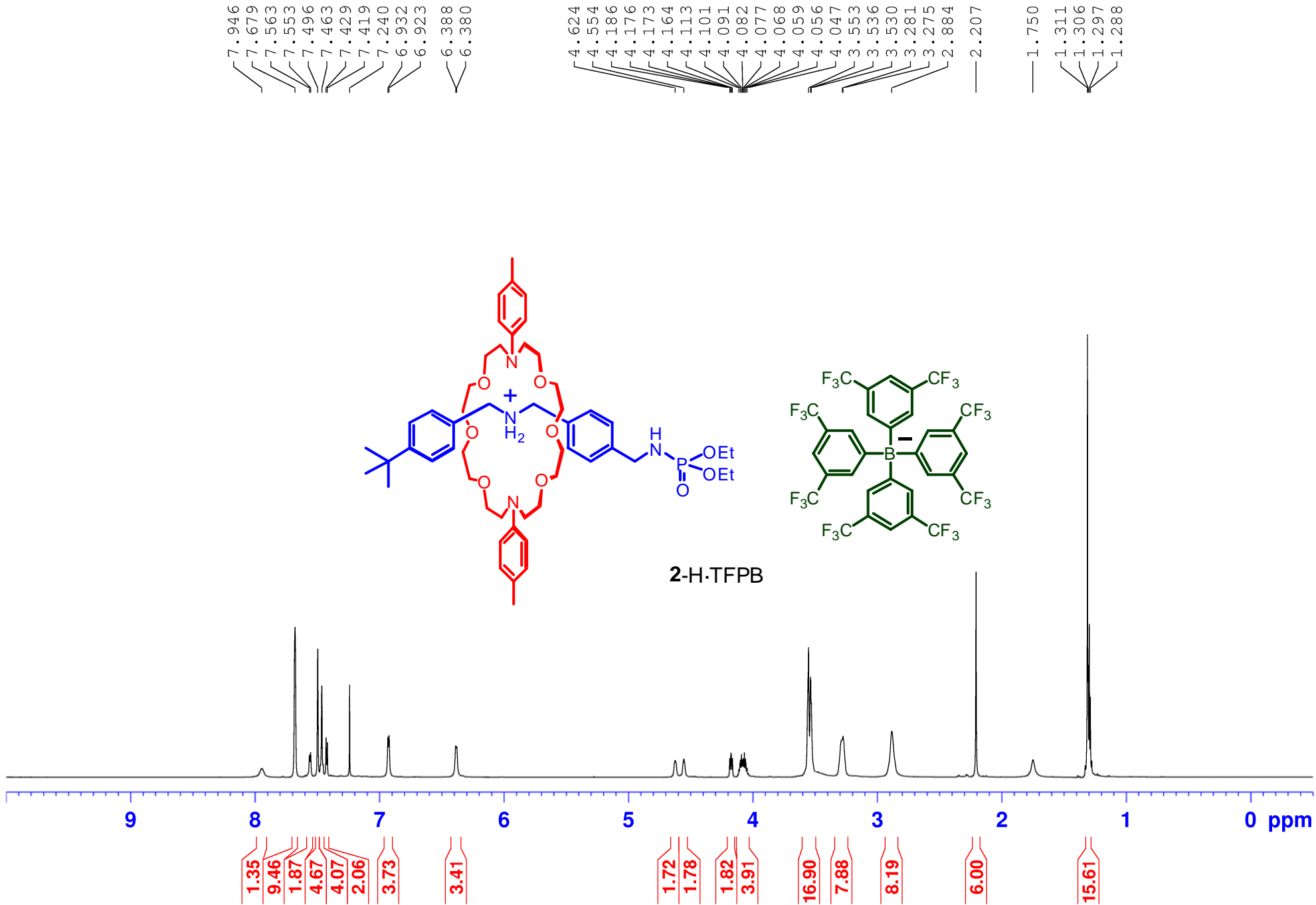


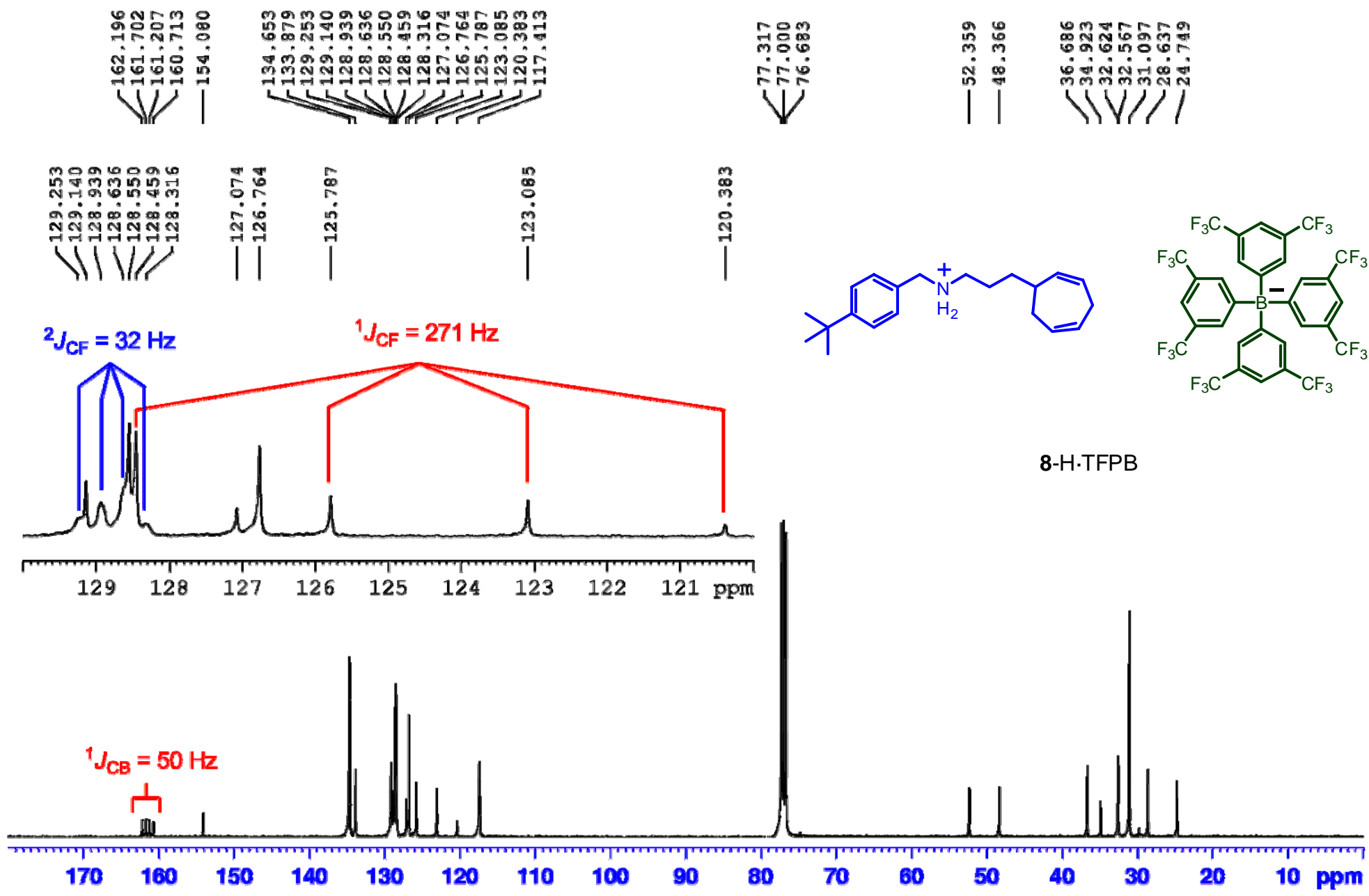
7-H·TFPB



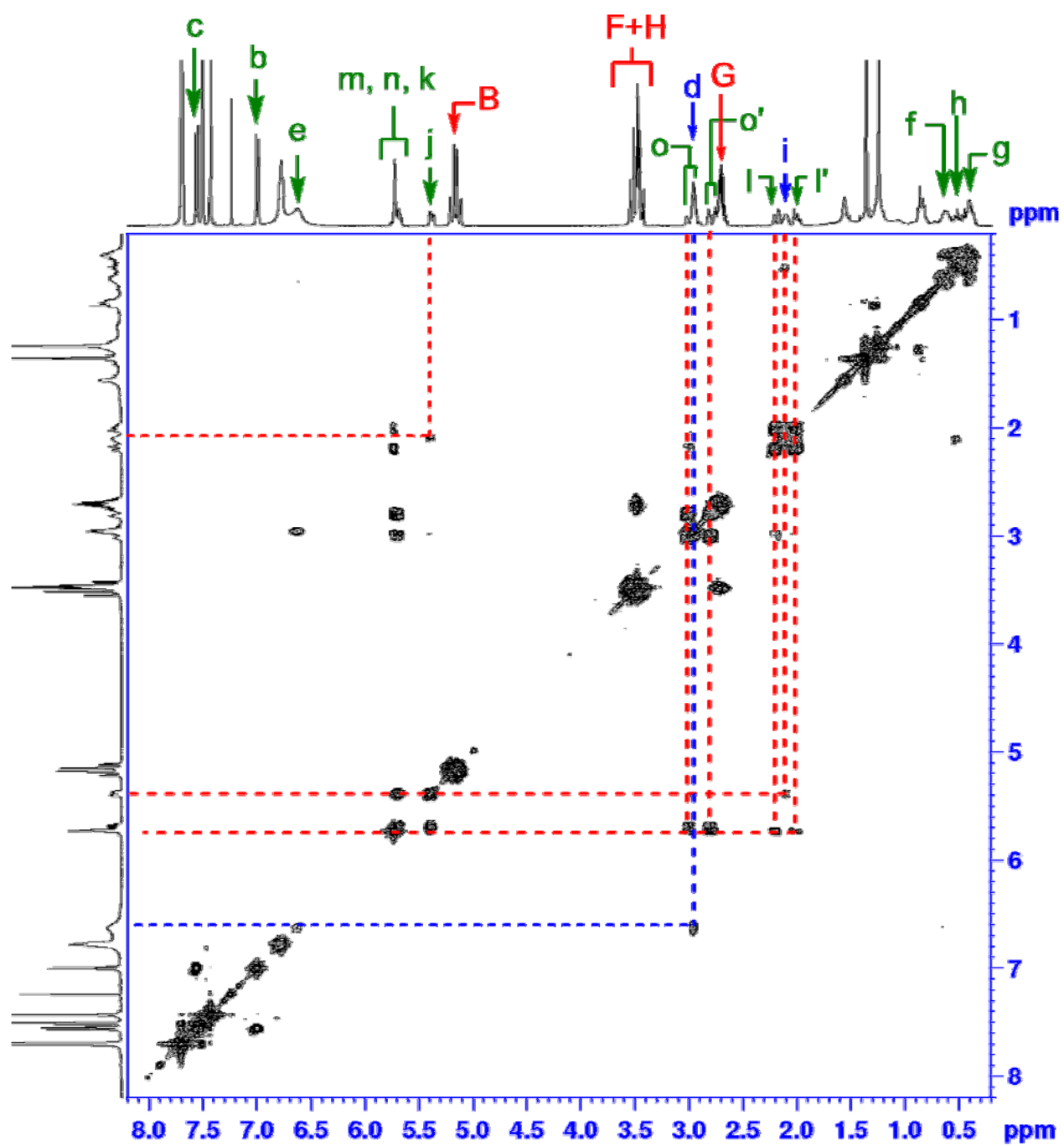
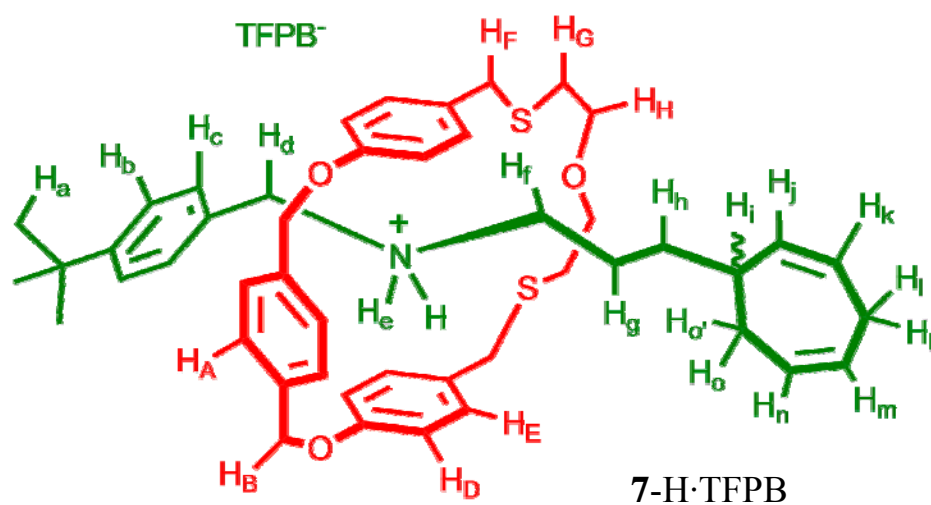




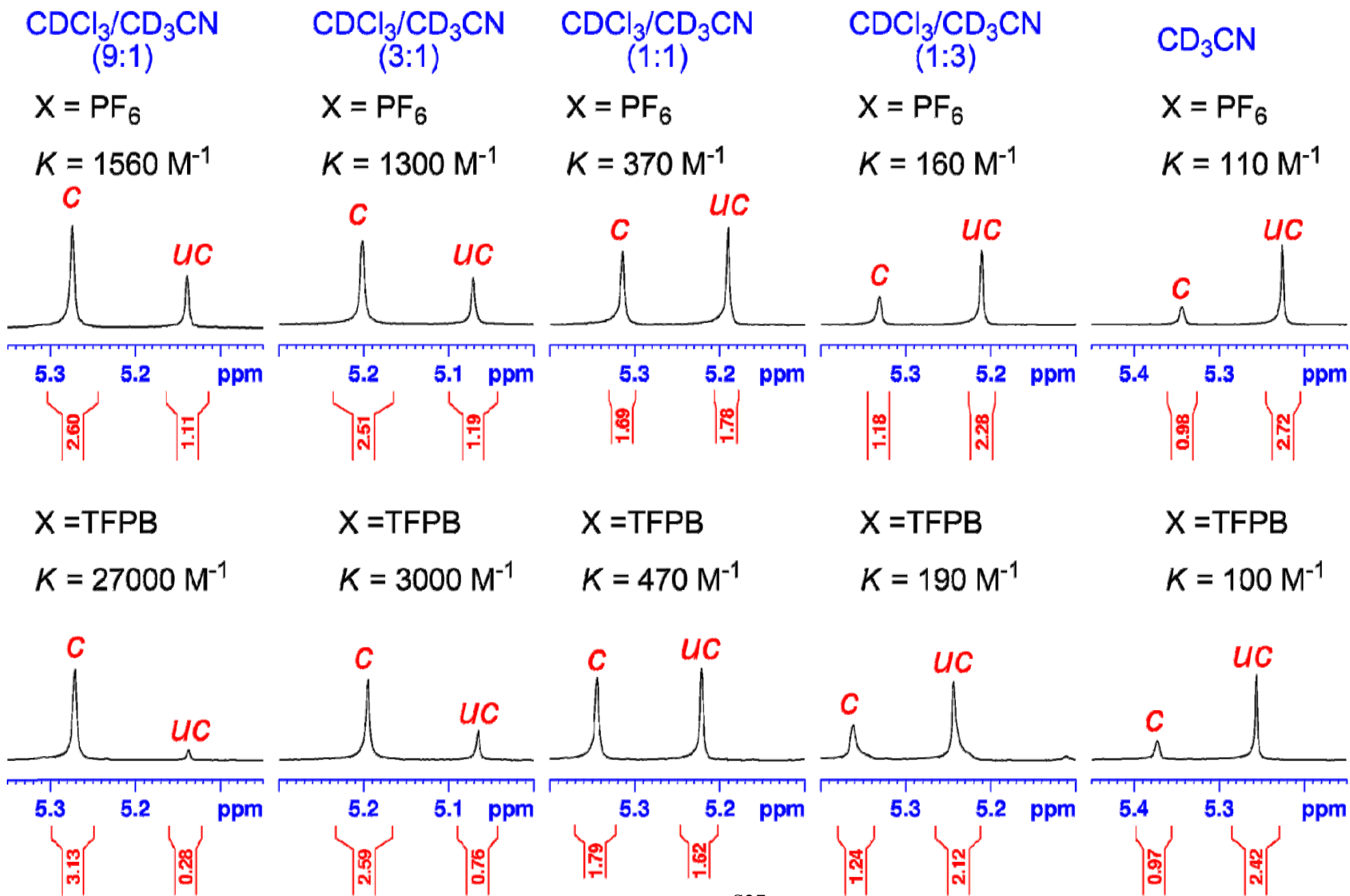




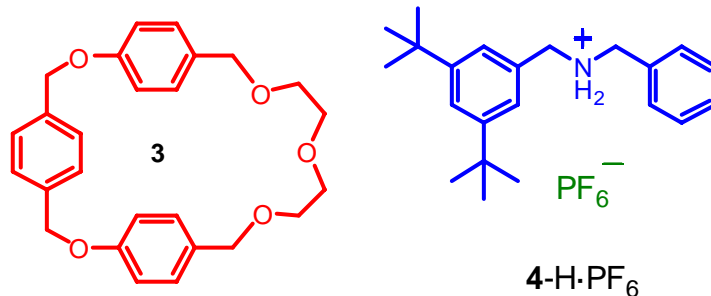
400 MHz NMR COSY CDCl<sub>3</sub> 298 K



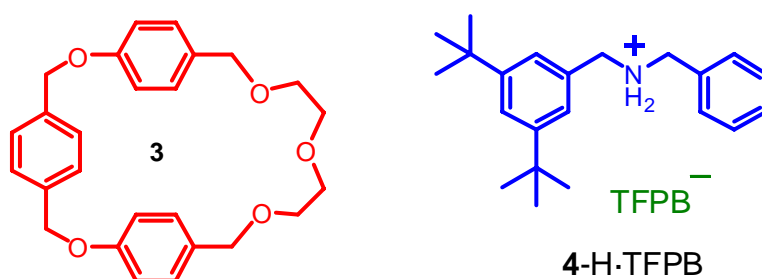
Partial  $^1\text{H}$  NMR spectra for the complexation of macrocycle **3** to threadlike salts  $[\mathbf{4}\text{-H}]^+$  in Various Solvents (400 MHz, 298K, 5mM)



ITC measurements were performed using a Microcal MCS calorimeter interfaced with a microcomputer. All sample solutions were carefully degassed prior to titration using the equipment provided with the instrument. The host molecule in solution ( $\text{CHCl}_3$ ) was titrated into the guest solution ( $\text{CHCl}_3$ ) using a 280- $\mu\text{L}$  syringe. Each titration consisted of a preliminary 3- $\mu\text{L}$  injection followed by 27 subsequent additions of 10  $\mu\text{L}$ . The entropy of complexation was determined by subtracting the heat of dilution for each titration from the enthalpy of the titration. All experiments were performed at 25 °C. Microcal LLC software was used to compute the thermodynamic parameters of the titration based on the one-site binding model or competitive binding model.

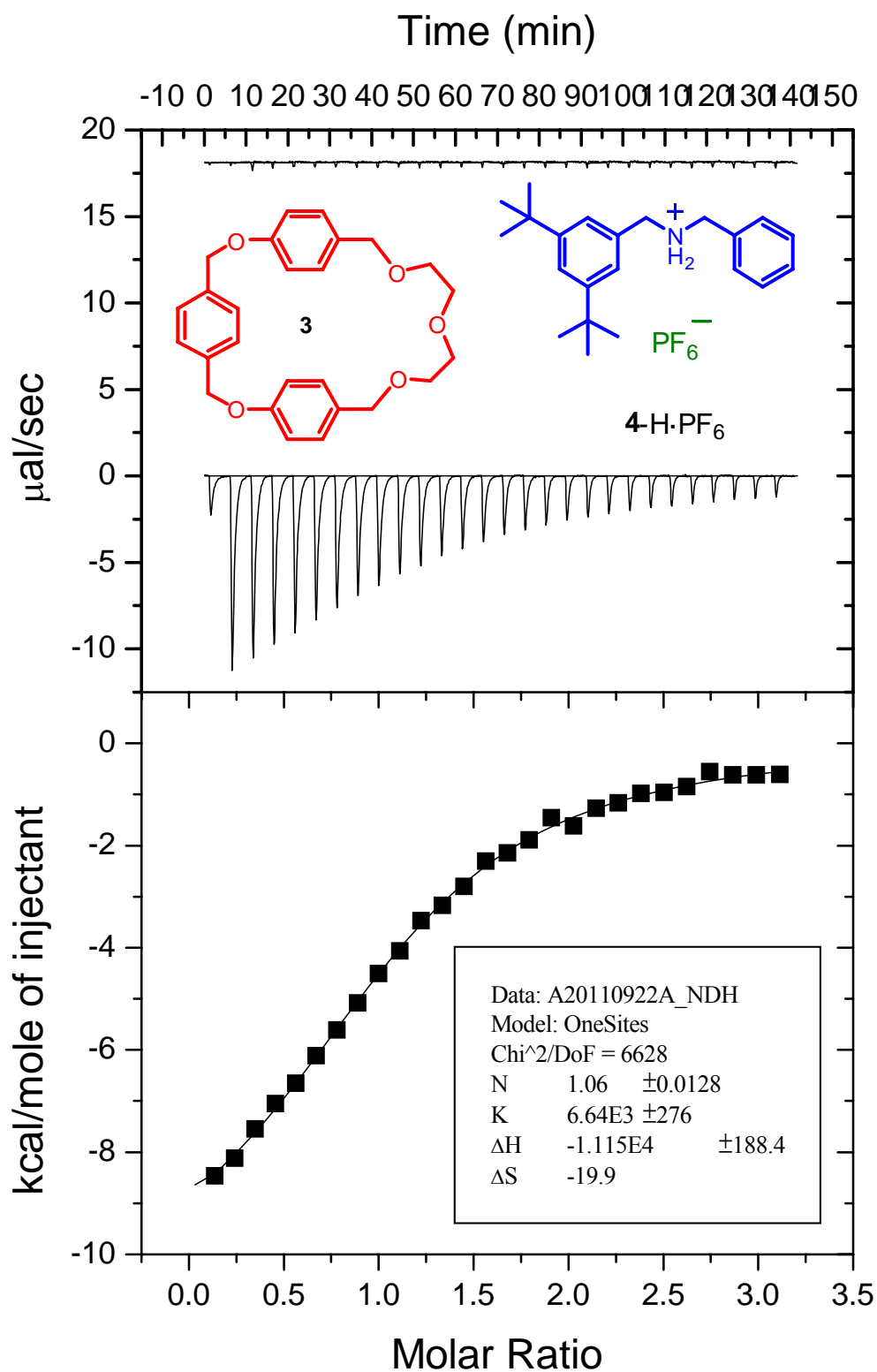


entry	N		K		$\Delta H$		$\Delta S$
1	1.060	$\pm$ 0.0128	6.64E+03	$\pm$ 276	-11150	$\pm$ 188.4	-19.9
2	1.040	$\pm$ 0.0141	7.17E+03	$\pm$ 344	-11690	$\pm$ 220.7	-21.6
average	1.050	$\pm$ 0.0141	6.91E+03	$\pm$ 375	-11420	$\pm$ 382	-20.8 $\pm$ 1.2

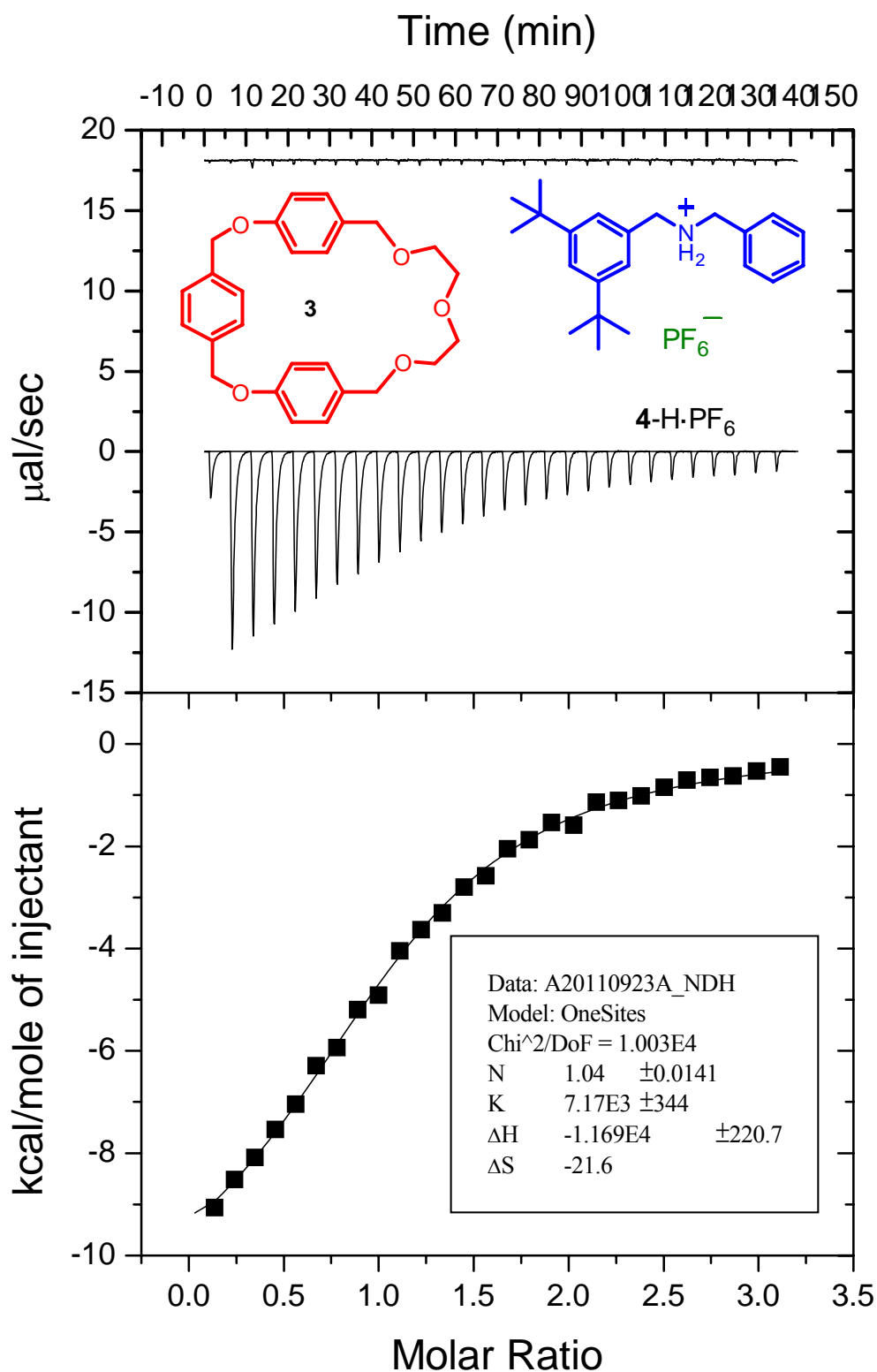


entry	N		K		$\Delta H$		$\Delta S$
1	1.050	$\pm$ 0.00188	7.36E+06	$\pm$ 1.27E+06	-10010	$\pm$ 44.81	-2.15
2	0.998	$\pm$ 0.00261	5.21E+06	$\pm$ 1.59E+06	-10790	$\pm$ 75.36	-5.45
average	1.024	$\pm$ 0.03677	6.29E+06	$\pm$ 1.52E+06	-10400	$\pm$ 552	-3.80 $\pm$ 2.33

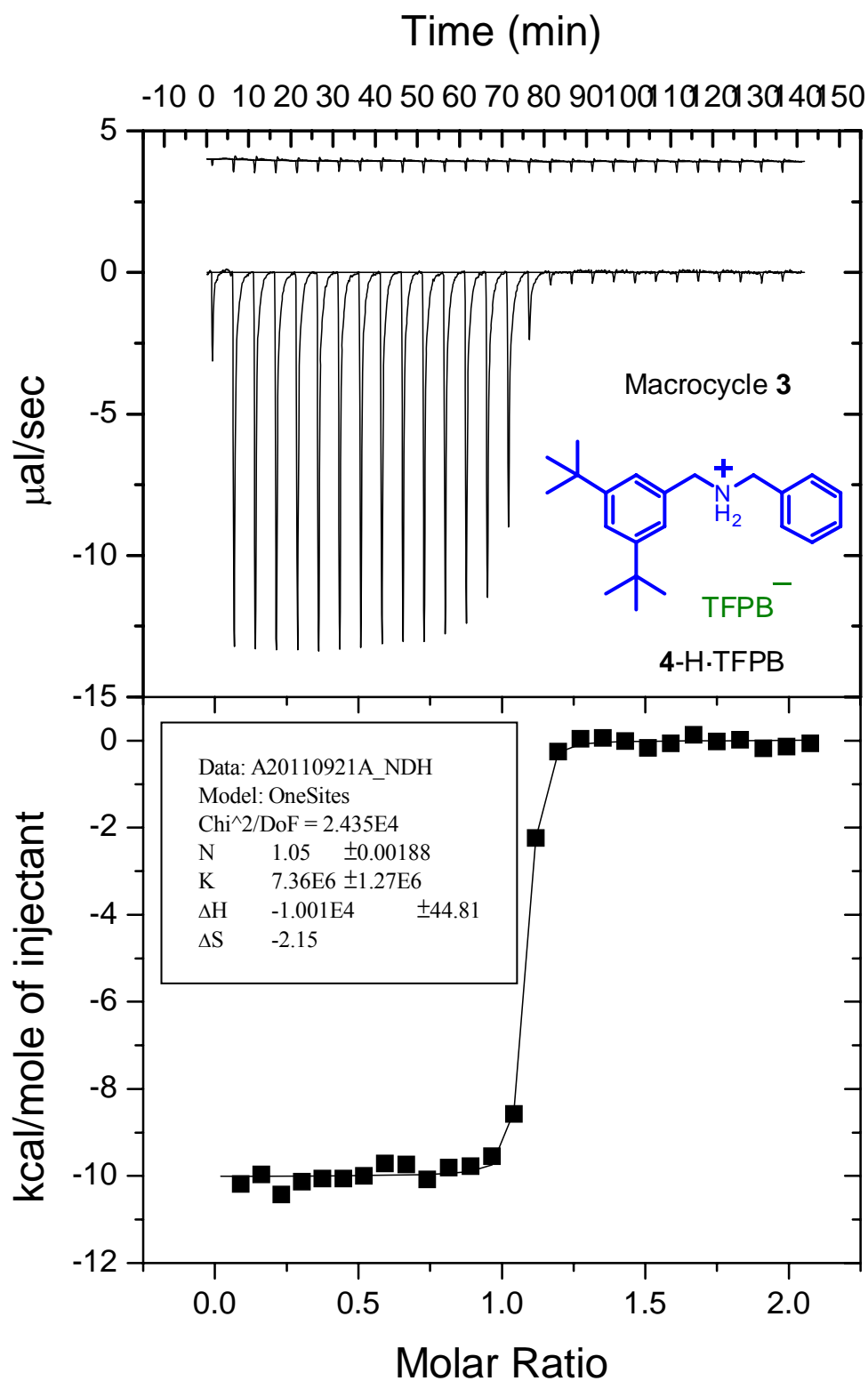
Using a Microcal VP-ITC titration microcalorimeter, aliquots (10  $\mu\text{L}$ , 7.5 mM) of degassed  $\text{CHCl}_3$  solution of macrocycle **3** were titrated into stirring  $\text{CHCl}_3$  solution of **4-H** $\cdot\text{PF}_6$  (0.5 mM) at 298 K. The entropy of complexation was determined by subtracting the heat of dilution for each titration from the enthalpy of the titration. Microcal LLC software was used to compute the thermodynamic parameters of the titration based on the one-site binding model.



Using a Microcal VP-ITC titration microcalorimeter, aliquots (10  $\mu\text{L}$ , 7.5 mM) of degassed  $\text{CHCl}_3$  solution of macrocycle **3** were titrated into stirring  $\text{CHCl}_3$  solution of **4-H** $\cdot\text{PF}_6$  (0.5 mM) at 298 K. The entropy of complexation was determined by subtracting the heat of dilution for each titration from the enthalpy of the titration. Microcal LLC software was used to compute the thermodynamic parameters of the titration based on the one-site binding model.

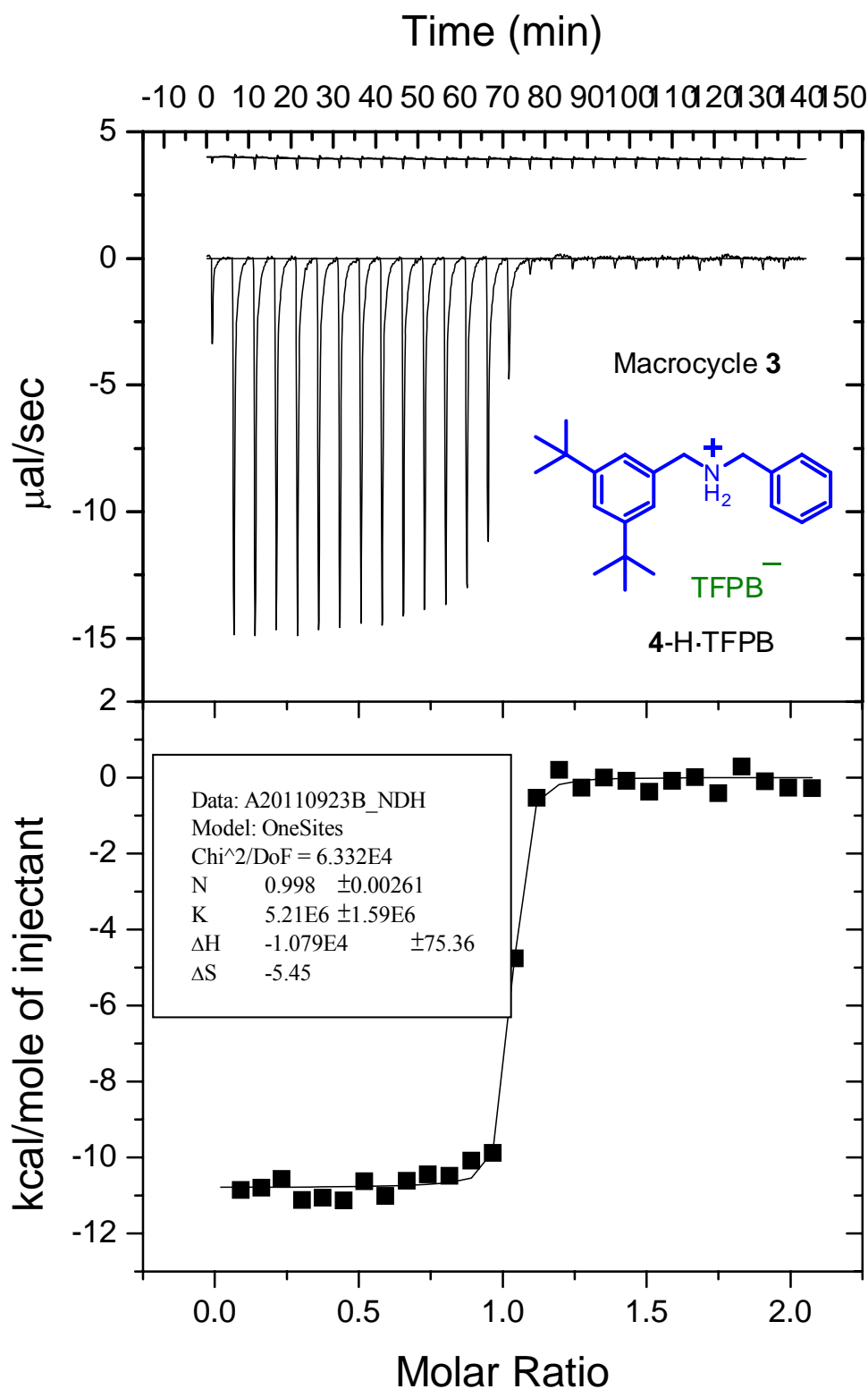


Using a Microcal VP-ITC titration microcalorimeter, aliquots (10  $\mu\text{L}$ , 5.0 mM) of degassed  $\text{CHCl}_3$  solution of macrocycle **3** were titrated into stirring  $\text{CHCl}_3$  solution of **4-H·TFPB** (0.5 mM) at 298 K. The entropy of complexation was determined by subtracting the heat of dilution for each titration from the enthalpy of the titration. Microcal LLC software was used to compute the thermodynamic parameters of the titration based on the one-site binding model.





Using a Microcal VP-ITC titration microcalorimeter, aliquots (10  $\mu\text{L}$ , 5.0 mM) of degassed  $\text{CHCl}_3$  solution of macrocycle **3** were titrated into stirring  $\text{CHCl}_3$  solution of **4-H·TFPB** (0.5 mM) at 298 K. The entropy of complexation was determined by subtracting the heat of dilution for each titration from the enthalpy of the titration. Microcal LLC software was used to compute the thermodynamic parameters of the titration based on the one-site binding model.



## Kinetic Data for the Dissociation of 7-H·TFPB into Its Components

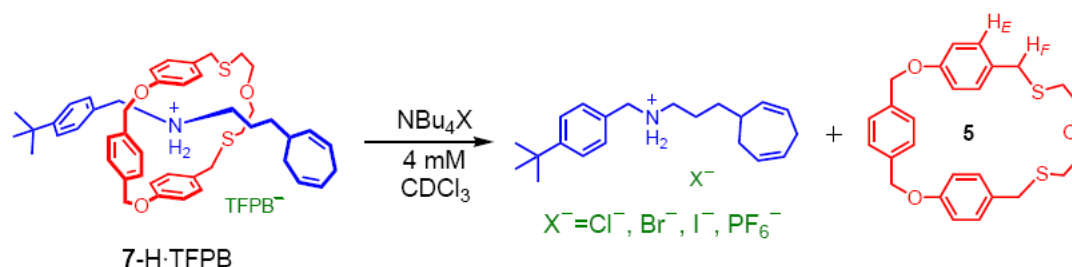


Figure S1. Dethreading of the macrocycle **5** from the dumbbell-shaped component after adding a tetra-*n*-butylammonium salt (NBu<sub>4</sub>X; X was chloride, bromide, iodide, or hexafluorophosphate) to a CDCl<sub>3</sub> solution of the [2]rotaxane 7-H·TFPB.

Experiments were performed in CDCl<sub>3</sub> with an initial concentration of 7-H·TFPB of 4 mM, followed by the addition of 1 equiv of NBu<sub>4</sub>X [X = Cl (Part A), Br (Part B), or I (Part C)] or 6 equiv of NBu<sub>4</sub>PF<sub>6</sub> (Part D). The values of  $k_d$  (s<sup>-1</sup>) were obtained from the slopes of the straight line in the plots of  $\ln([A_0]/[A_t])$  against  $t$  (s) at five temperatures. The values of  $[A_t]$  were determined based on the standard signal at  $\delta$  7.69 (br, 8H; the aromatic protons of TFPB) and by integration of the signals of the macrocycle **5** (H<sub>E</sub> or H<sub>F</sub>) over a period of time. The values of  $\Delta G^\ddagger$  (kcal mol<sup>-1</sup>) were calculated using the relationship  $\Delta G^\ddagger = -RT \ln(k_d h / k_B T)$ , where  $R$ ,  $h$ , and  $k_B$  correspond to the gas, Planck, and Boltzmann constants, respectively. The values of  $\Delta H^\ddagger$  (kcal mol<sup>-1</sup>) and  $\Delta S^\ddagger$  (cal mol<sup>-1</sup> K<sup>-1</sup>) were obtained from the intercepts and slopes, respectively, of the straight lines in the plots of  $\Delta G^\ddagger$  against  $T$ , using the relationship  $\Delta G^\ddagger = \Delta H^\ddagger - T\Delta S^\ddagger$ .

## Part A

In the chloride experiments, the  $k_d$  ( $s^{-1}$ ) were obtained at 258 K, 268 K, 278 K, 288 K and 298 K.  $[A]_t$  were determined by integration of the signals of macrocycle **5** at  $\delta$  6.66 ( $H_E, d, J = 9$  Hz, 4H).

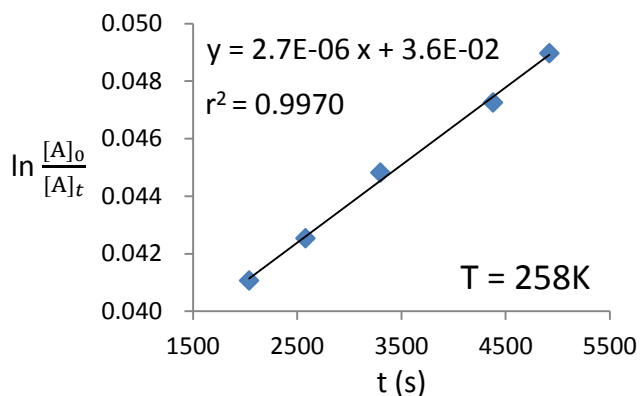


Figure S2.

$k_d = \text{Slope} = 2.7 \times 10^{-6} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 21.6 \text{ (kcal/mol)}$

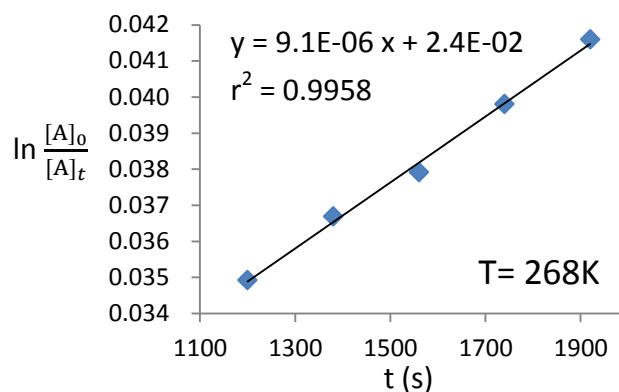


Figure S3.

$k_d = \text{Slope} = 9.1 \times 10^{-6} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 21.8 \text{ (kcal/mol)}$

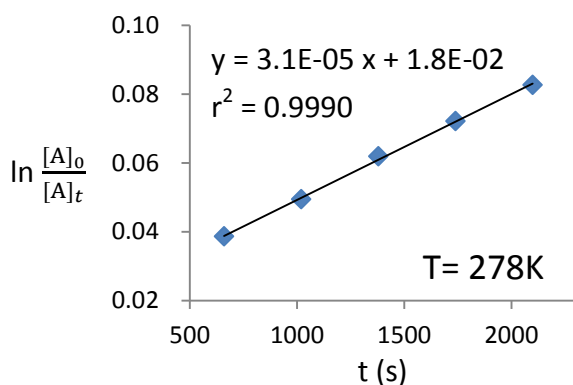


Figure S4.

$k_d = \text{Slope} = 3.1 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.0 \text{ (kcal/mol)}$

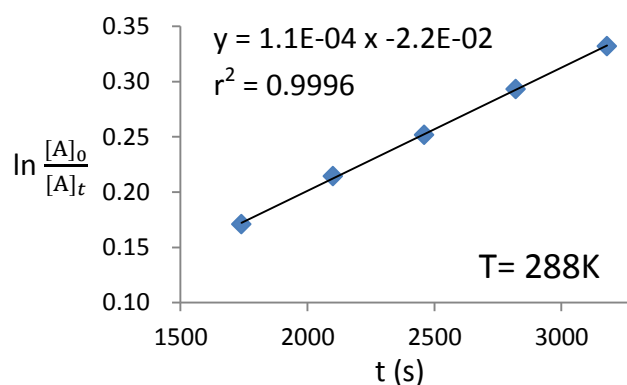


Figure S5.

$k_d = \text{Slope} = 1.1 \times 10^{-4} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.0 \text{ (kcal/mol)}$

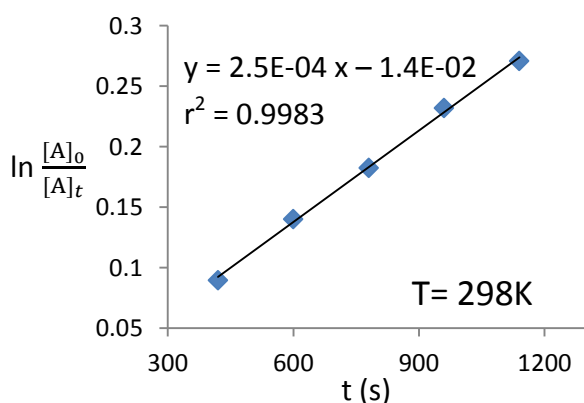


Figure S6.

$k_d = \text{Slope} = 2.5 \times 10^{-4} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.3 \text{ (kcal/mol)}$

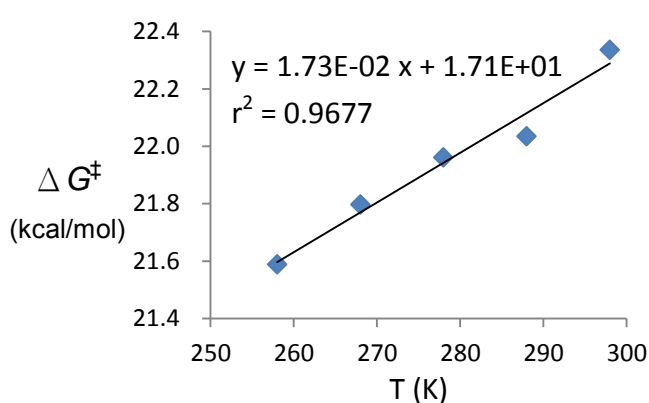


Figure S7.

$\Delta H^\ddagger = \text{intercept} = 17.1 \text{ (kcal/mol)},$

$\Delta S^\ddagger = - \text{slope} = - 17.3 \text{ (cal/mol)}$

## Part B

In the bromide experiments, the  $k_d$  ( $s^{-1}$ ) were obtained at 268 K, 278 K, 288 K, 298 K and 308 K.  $[A]_t$  were determined by integration of the signals of macrocycle 5 at  $\delta$  6.66 ( $H_E$ , d,  $J = 9$  Hz, 4H).

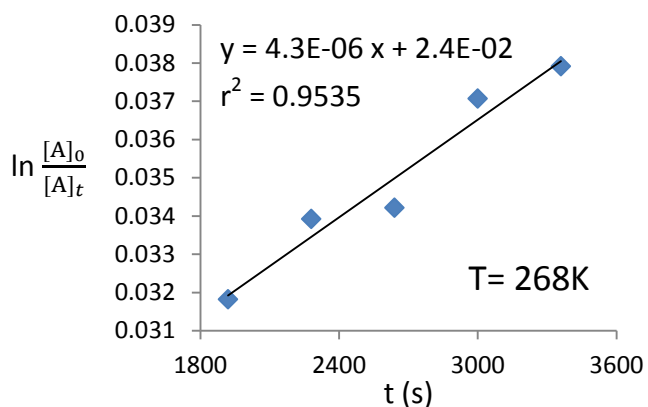


Figure S8.

$$k_d = \text{Slope} = 4.3 \times 10^{-6} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.2 \text{ (kcal/mol)}$$

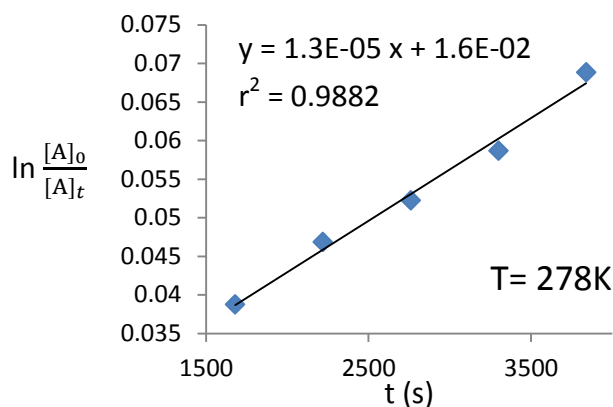


Figure S9.

$$k_d = \text{Slope} = 1.3 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.4 \text{ (kcal/mol)}$$

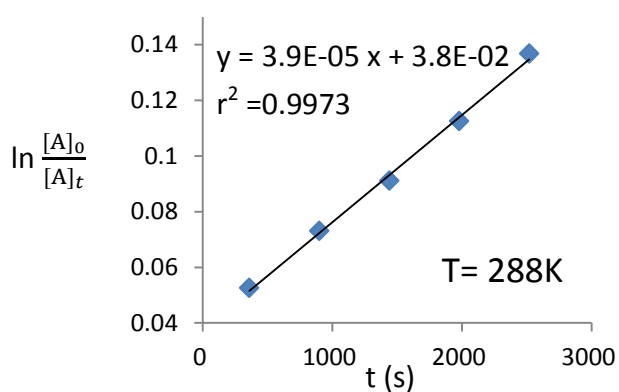


Figure S10.

$$k_d = \text{Slope} = 3.9 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.6 \text{ (kcal/mol)}$$

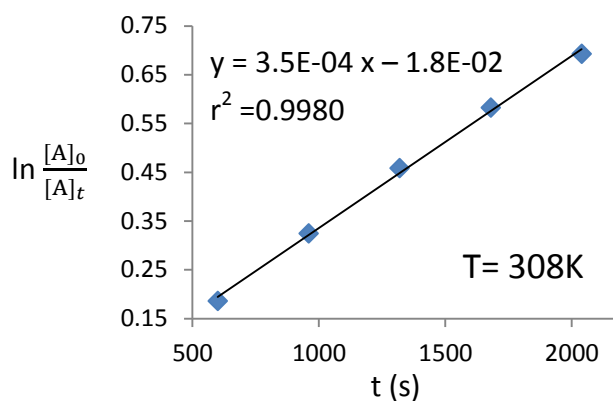


Figure S11.

$$k_d = \text{Slope} = 3.5 \times 10^{-4} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.9 \text{ (kcal/mol)}$$

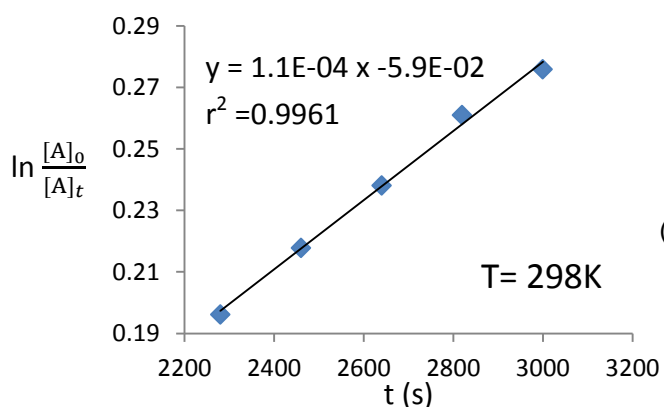


Figure S12.

$$k_d = \text{Slope} = 1.1 \times 10^{-4} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 22.8 \text{ (kcal/mol)}$$

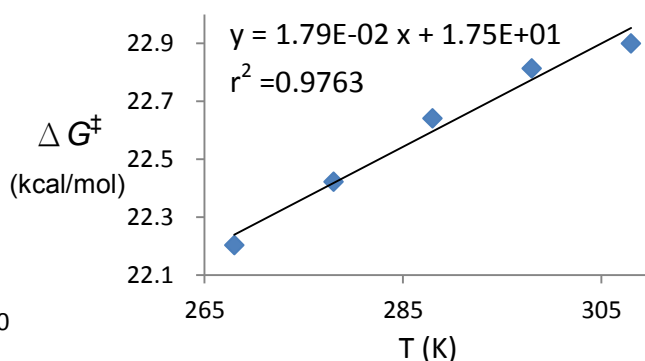


Figure S13.

$$\Delta H^\ddagger = \text{intercept} = 17.5 \text{ (kcal/mol)},$$

$$\Delta S^\ddagger = -\text{slope} = -17.9 \text{ (cal/mol)}$$

### Part C

In the iodide experiments, the  $k_d$  ( $s^{-1}$ ) were obtained at 288 K, 293 K, 298 K, 303 K and 308 K.  $[A]_t$  were determined by integration of the signals of macrocycle **5** at  $\delta$  6.66 ( $H_E, d, J = 9$  Hz, 4H).

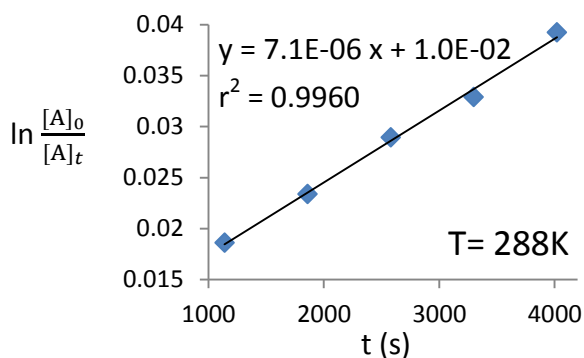


Figure S14.

$k_d = \text{Slope} = 7.1 \times 10^{-6} (s^{-1})$ ,  $\Delta G^\ddagger = 23.6$  (kcal/mol)

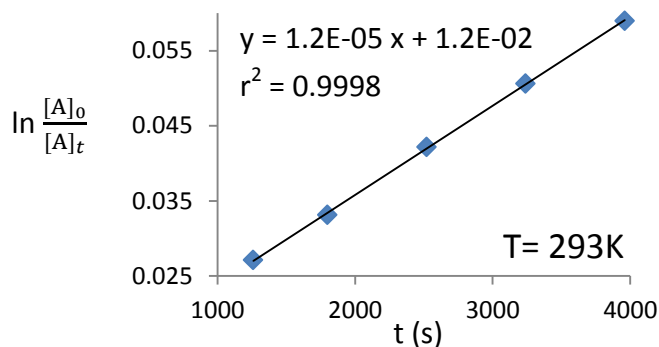


Figure S15.

$k_d = \text{Slope} = 1.2 \times 10^{-5} (s^{-1})$ ,  $\Delta G^\ddagger = 23.7$  (kcal/mol)

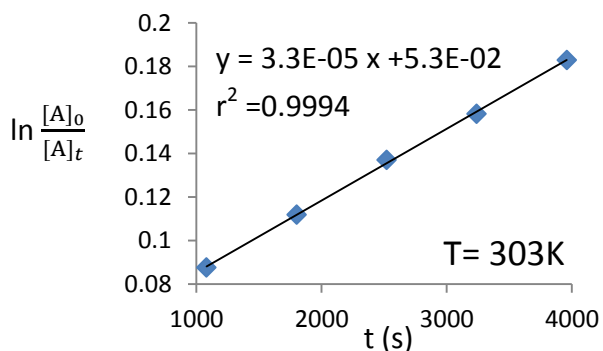


Figure S16.

$k_d = \text{Slope} = 3.3 \times 10^{-5} (s^{-1})$ ,  $\Delta G^\ddagger = 23.9$  (kcal/mol)

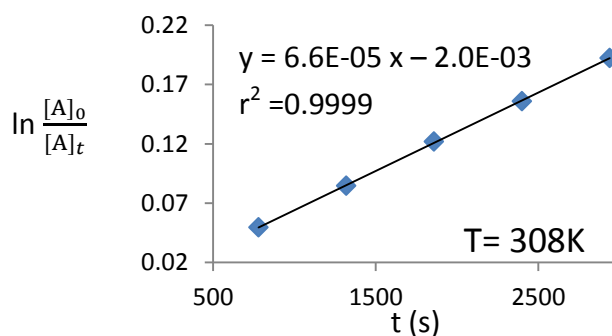


Figure S17.

$k_d = \text{Slope} = 6.6 \times 10^{-5} (s^{-1})$ ,  $\Delta G^\ddagger = 23.9$  (kcal/mol)

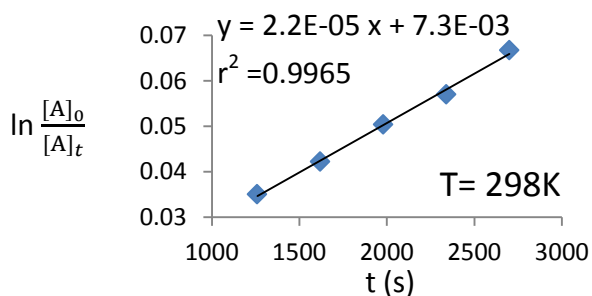


Figure S18.

$k_d = \text{Slope} = 2.2 \times 10^{-5} (s^{-1})$ ,  $\Delta G^\ddagger = 23.8$  (kcal/mol)

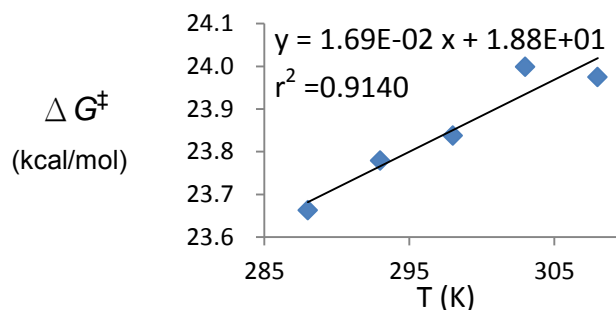


Figure S19.

$\Delta H^\ddagger = \text{intercept} = 18.8$  (kcal/mol),

$\Delta S^\ddagger = -\text{slope} = -16.9$  (cal/mol)

### Part D

In the hexafluorophosphate experiments, the  $k_d$  ( $s^{-1}$ ) were obtained at 298 K, 303 K, 308 K, 313 K and 318 K.  $[A]_t$  were determined by integration of the signals of macrocycle 5 at  $\delta$  3.62 ( $H_F$ , s, 4H).

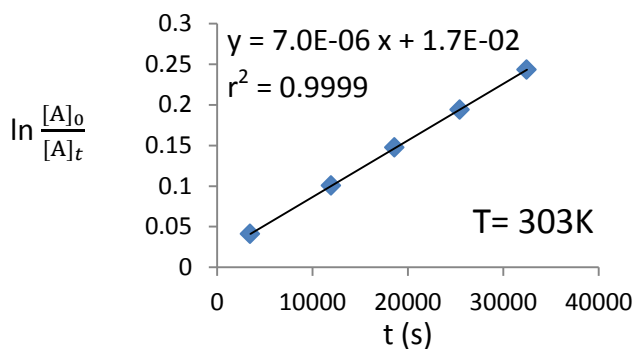


Figure S20.

$$k_d = \text{Slope} = 7.0 \times 10^{-6} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 24.9 \text{ (kcal/mol)}$$

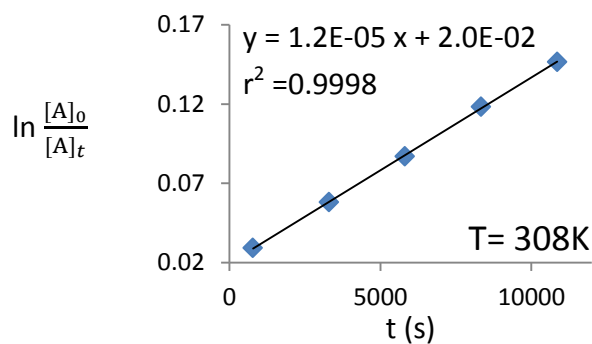


Figure S21.

$$k_d = \text{Slope} = 1.2 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 25.0 \text{ (kcal/mol)}$$

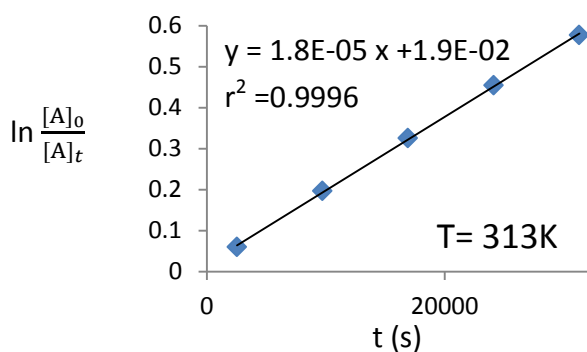


Figure S22.

$$k_d = \text{Slope} = 1.8 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 25.1 \text{ (kcal/mol)}$$

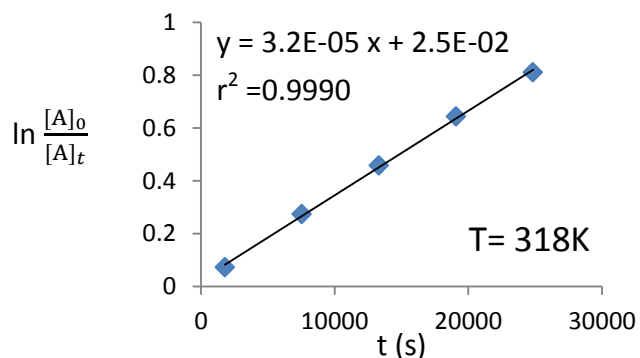


Figure S23.

$$k_d = \text{Slope} = 3.2 \times 10^{-5} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 25.2 \text{ (kcal/mol)}$$

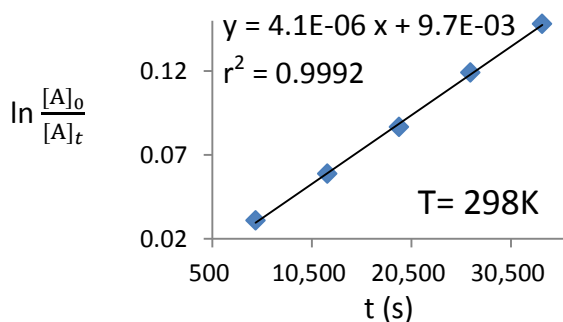


Figure S24.

$$k_d = \text{Slope} = 4.1 \times 10^{-6} \text{ (s}^{-1}\text{)}, \Delta G^\ddagger = 24.8 \text{ (kcal/mol)}$$

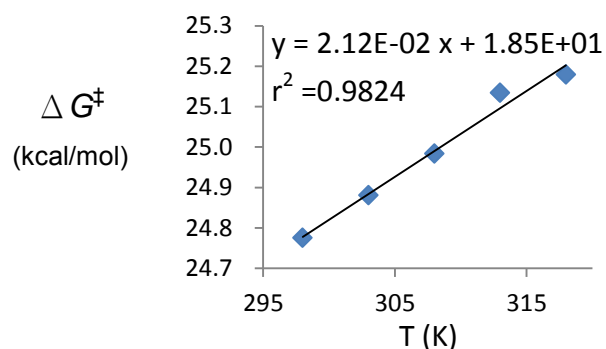


Figure S25.

$$\Delta H^\ddagger = \text{intercept} = 18.5 \text{ (kcal/mol)}$$

$$\Delta S^\ddagger = -\text{slope} = -21.2 \text{ (cal/mol)}$$

7-H• TPFB + Bu<sub>4</sub>NCl

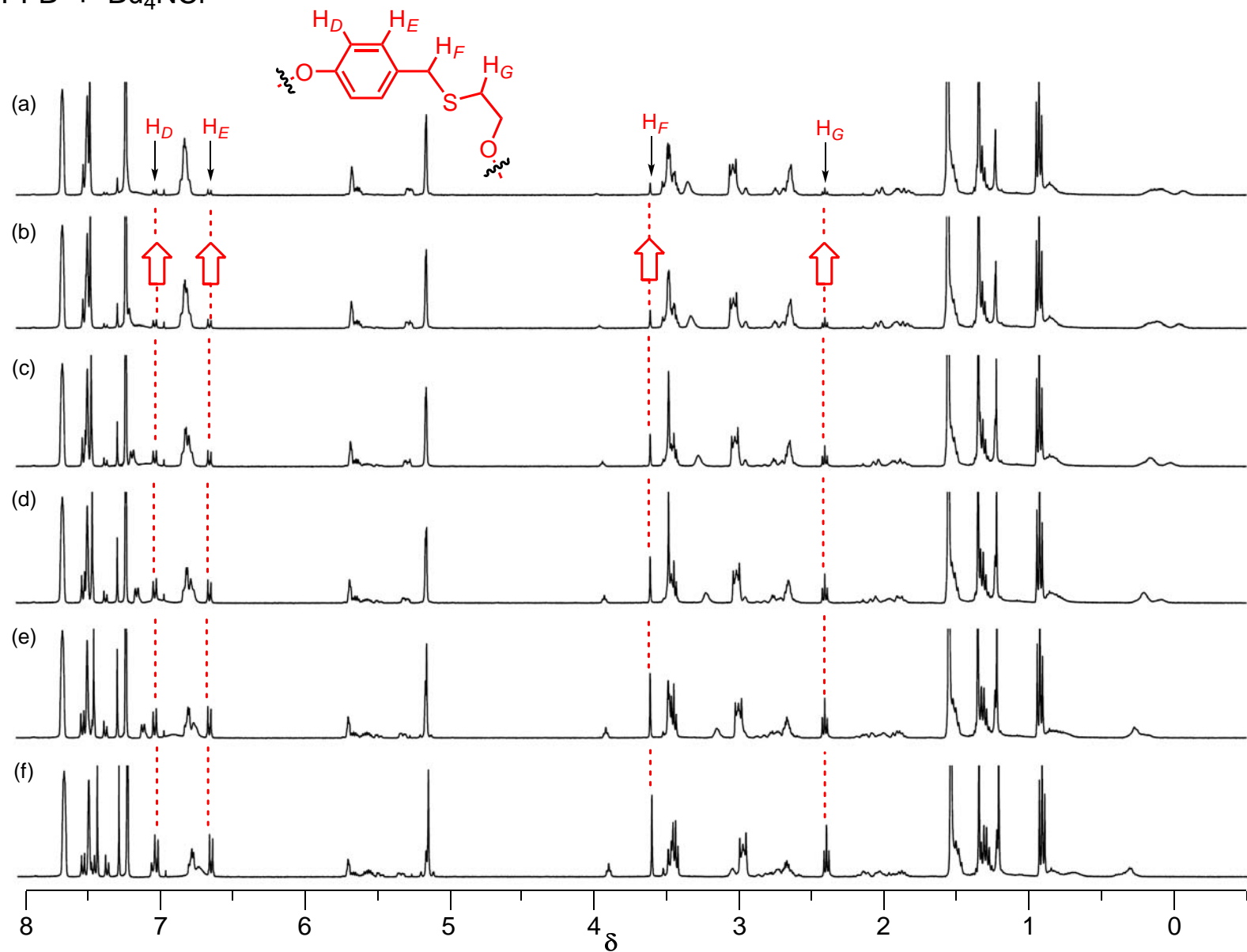


Fig. <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 298 K) revealing the dissociation of the macrocycle **5** from an equimolar mixture of Bu<sub>4</sub>NCl and 7-H• TPFB (4 mM) over time; a) 4, b) 7, c) 13, d) 19, e) 28, f) 43 min.

7-H• TPFB + Bu<sub>4</sub>NBr

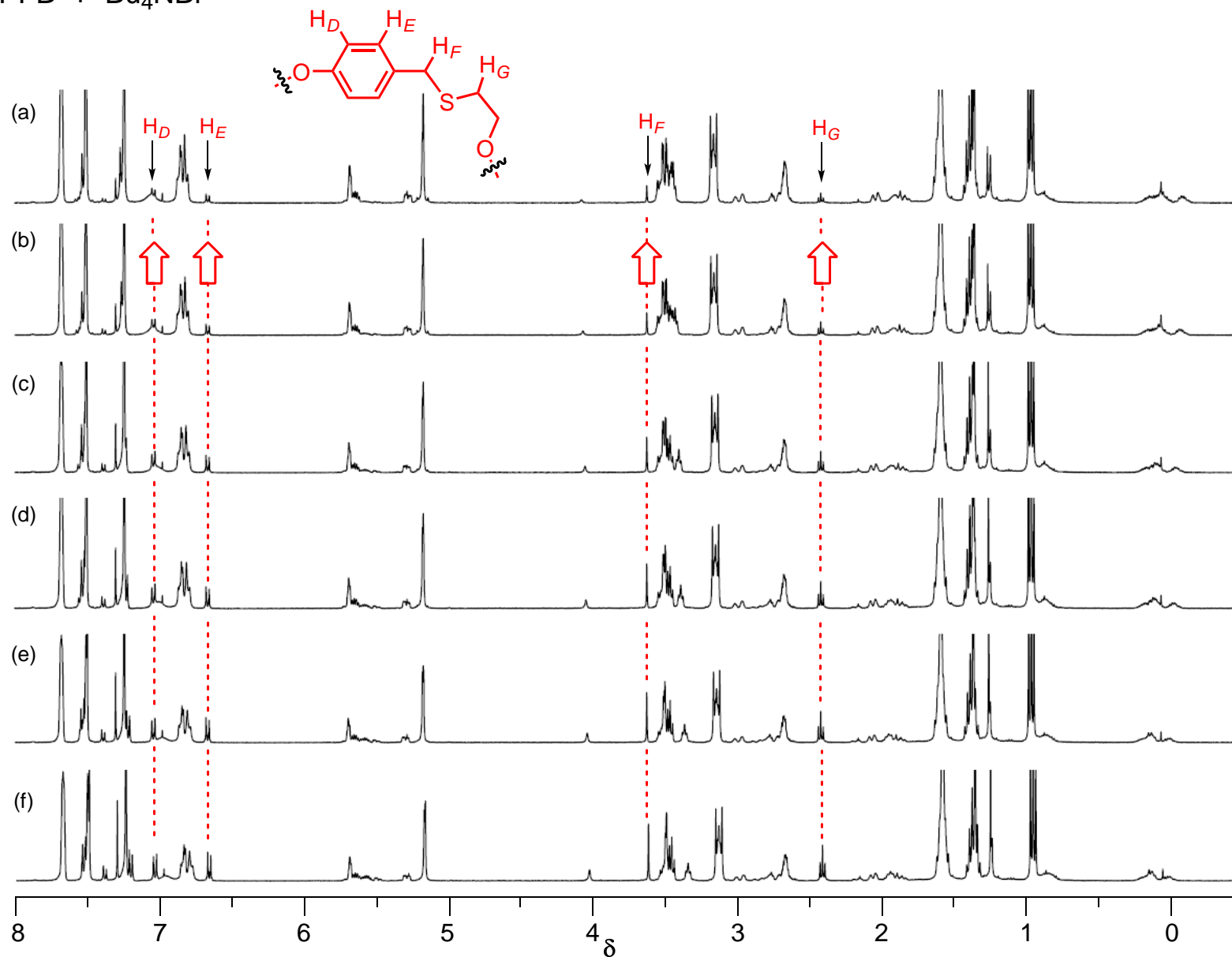


Fig. <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 298 K) revealing the dissociation of the macrocycle **5** from an equimolar mixture of Bu<sub>4</sub>NBr and 7-H• TPFB (4 mM) over time; a) 14, b) 20, c) 32, d) 38, e) 50, and f) 56 min.



# 7-H• TPFB + Bu<sub>4</sub>NI

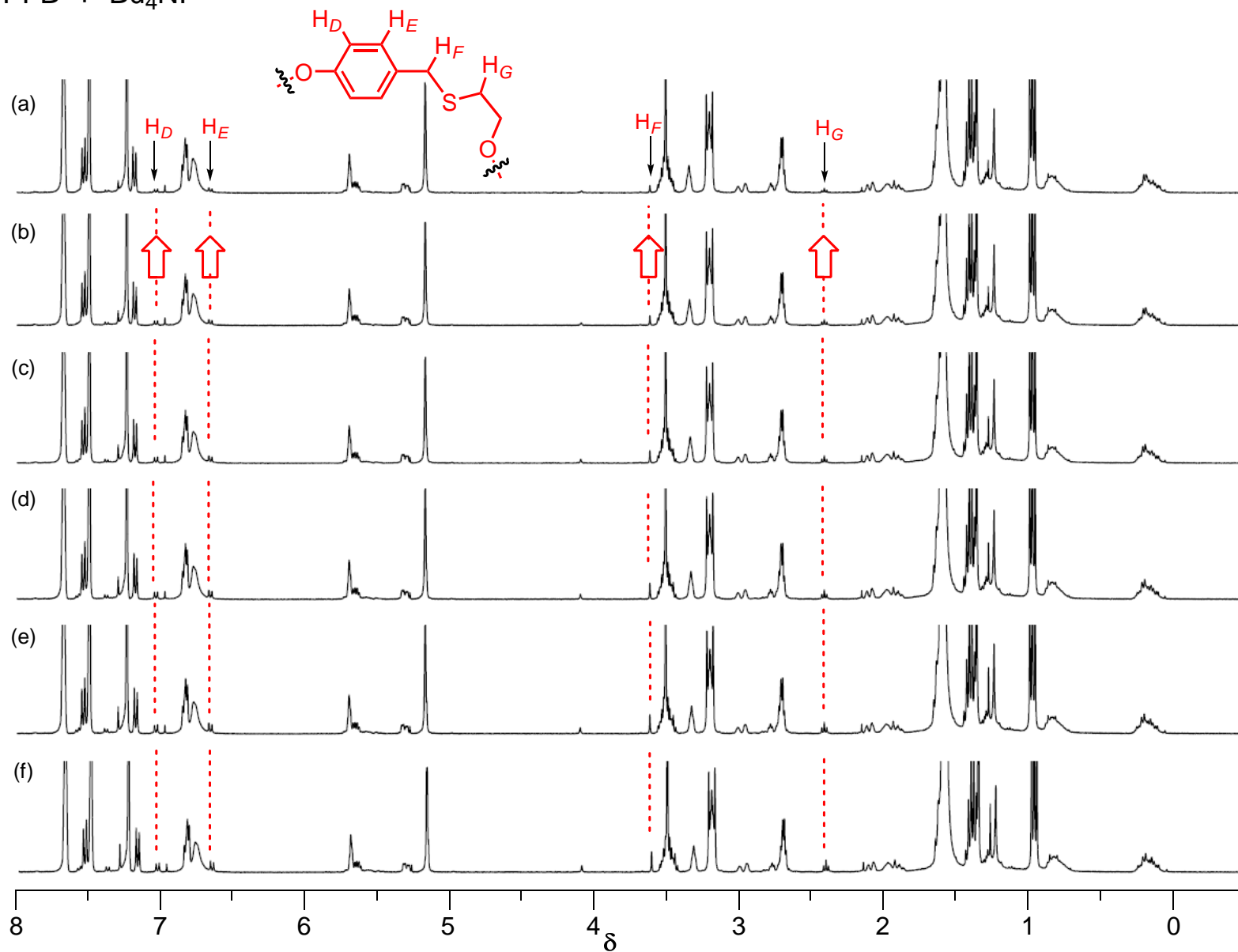


Fig. <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 298 K) revealing the dissociation of the macrocycle **5** from an equimolar mixture of Bu<sub>4</sub>NI and 7-H• TPFB (4 mM) over time; a) 21, b) 30, c) 39, d) 48, e) 57, and f) 63 min.

7-H• TPFB + Bu<sub>4</sub>NPF<sub>6</sub>

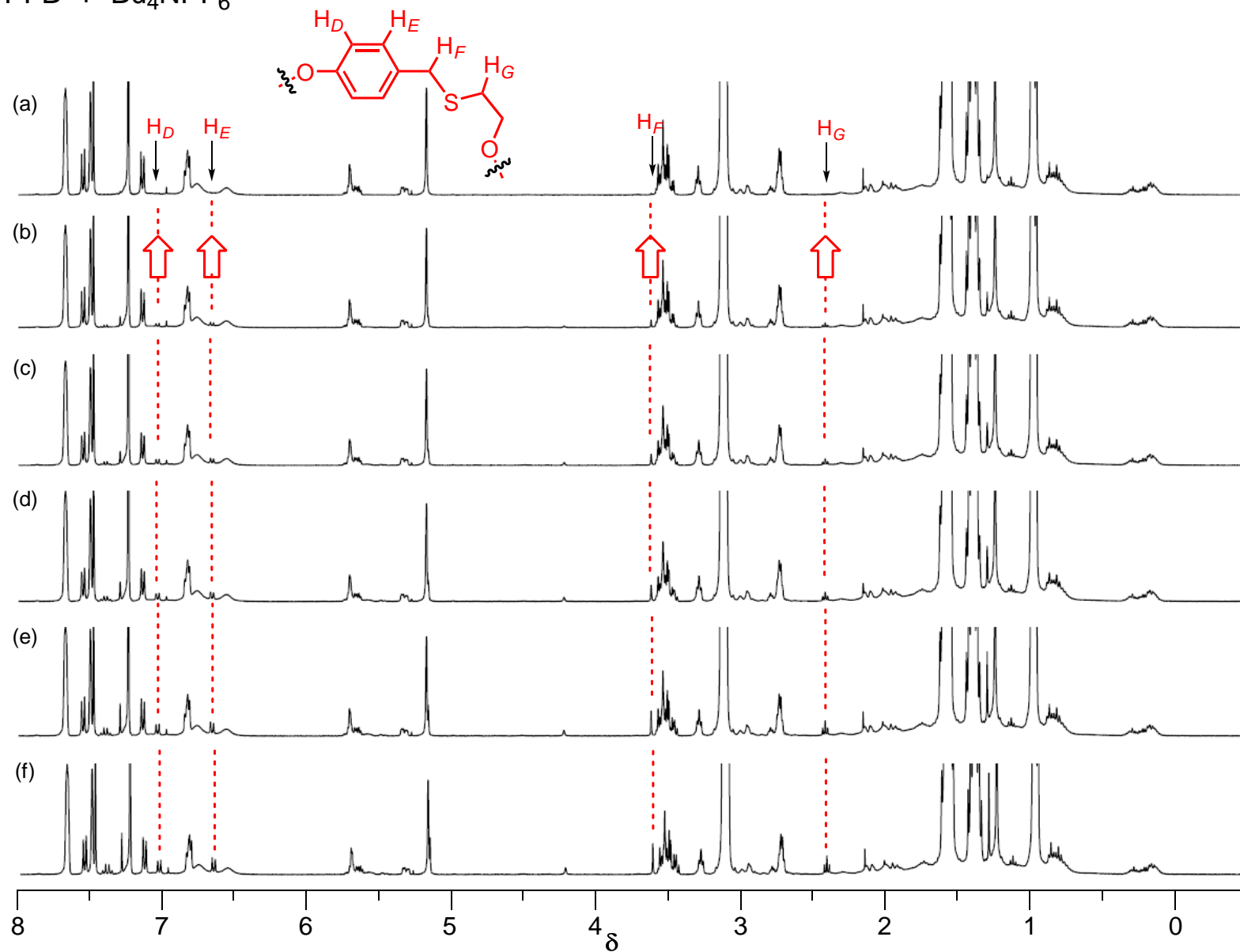


Fig. <sup>1</sup>H NMR spectra (400 MHz, CDCl<sub>3</sub>, 298 K) revealing the dissociation of the macrocycle **5** from a mixture of Bu<sub>4</sub>NPF<sub>6</sub> (24 mM) and 7-H• TPFB (4 mM) over time; a) 0.4, b) 2.5, c) 4.6, d) 6.7, e) 8.8, and f) 10.9 h.