Deep ocean nutrients during the Last Glacial Maximum deduced from sponge silicon isotopic compositions

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8 Abstract

9 The relative importance of biological and physical processes within the Southern Ocean for the storage of carbon and atmospheric pCO_2 on glacial-interglacial 10 timescales remains uncertain. Understanding the impact of surface biological 11 production on carbon export in the past relies on the reconstruction of the nutrient 12 supply from upwelling deep-waters. In particular, the upwelling of silicic acid 13 14 $(Si(OH)_4)$ is tightly coupled to carbon export in the Southern Ocean via diatom productivity. Here, we address how changes in deep-water Si(OH)₄ concentrations 15 can be reconstructed using the silicon isotopic composition of deep-sea sponges. We 16 report δ^{30} Si of modern deep-sea sponge spicules and show that they reflect seawater 17 Si(OH)₄ concentration. The fractionation factor of sponge δ^{30} Si compared to seawater 18 δ^{30} Si shows a positive relationship with Si(OH)₄, which may be a growth rate effect. 19 20 Application of this proxy in two down-core records from the Scotia Sea reveals that 21 $Si(OH)_4$ concentrations in the deep Southern Ocean during the Last Glacial 22 Maximum (LGM) were no different than today. Our result does not support a coupling of carbon and nutrient build up in an isolated deep-ocean reservoir during 23

24	the LGM. Our data, combined with records of stable isotopes from diatoms, are only
25	consistent with enhanced LGM Southern Ocean nutrient utilization if there was also
26	a concurrent reduction in diatom silicification or a shift from siliceous to organic-
27	walled phytoplankton.
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29 Keywords: Porifera, spicule, silicic acid, deep-water, silicon cycle, glacial

32 Debate still surrounds the relative importance of physical and biological mechanisms behind glacial-interglacial variations in atmospheric carbon dioxide (pCO₂; reviewed by 33 34 Sigman and Boyle, 2000). The Southern Ocean has been implicated in the regulation of 35 greenhouse gases through both types of mechanism. Firstly, a physical reduction in the 36 ventilation of deep waters due to greater glacial sea-ice cover and ocean stratification, 37 enhanced in a cooler ocean, would result in less outgassing of CO₂ to the atmosphere (e.g. 38 de Boer et al., 2007). Secondly, an increase in biological export, accompanied by 39 enhanced burial of carbon, would also reduce atmospheric pCO₂ during glacials (e.g. 40 Kohfeld et al., 2005). A "biogeochemical divide" has been proposed, whereby biological export in the Antarctic Zone of the Southern Ocean regulates CO₂ directly, whereas export 41 42 in the Subantarctic Zone controls global preformed nutrient supply, such that different 43 regions around Antarctica may drive or respond to climate change by different mechanisms 44 (Marinov et al., 2006). An understanding of Southern Ocean nutrients is clearly required 45 to distinguish the physical and biological mechanisms that impact pCO₂ over glacial-46 interglacial timescales.

The concentration of silicic acid, $[Si(OH)_4]$, in deep-waters is governed by tectonics and silicate weathering on long timescales (>10⁴ years) and by ocean productivity and ocean circulation on glacial-interglacial timescales (10³-10⁴ years; Ragueneau et al., 2000; Falkowski et al., 2004). As such the Si cycle is a synergistic driver of, and respondent to, the carbon (C) cycle and global climatic change. In the modern surface ocean, biological precipitation of amorphous silica (opal) by diatoms is the dominant process that removes 53 Si(OH)₄ from seawater, efficiently transporting silica and organic C to the seafloor. The 54 partitioning of C and Si between the surface and deep-ocean is controlled by the export and 55 remineralization of this biological material relative to vertical mixing rates (Toggweiler et 56 al., 1999; Ragueneau et al., 2000).

57 Diatom blooms rely on upwelling sources of Si(OH)₄ because efficient utilization 58 strips almost all of the Si from surface waters (Ragueneau et al., 2000). The nutrient 59 composition of upwelling waters, in particular the ratio of Si to other major nutrients, plays 60 a strong role in the population structure of phytoplankton growing in surface waters and 61 the biological pumping of C to deep-water (Yool and Tyrell, 2003; Falkowski et al., 2004). 62 Furthermore, reduced vertical mixing, or enhanced stratification, results in an increase in 63 deep-water C, and a corresponding reduction in atmospheric pCO₂. (Toggweiler, 1999). 64 Quantifying changes in the Si(OH)₄ content of deep-water is an important step towards 65 understanding the link between the sequestration of C, Si and other nutrients over glacial 66 cycles (Brzezinski et al., 2002; Matsumoto et al., 2002). The Southern Ocean is a key 67 location for studying paleo-Si(OH)₄ because of the regional and global importance of Si-68 based productivity and its potential sensitivity to well documented proximal climatic 69 changes (Anderson et al., 2002).

The silicon isotopic composition (δ^{30} Si) of biogenic opal provides a direct method for quantifying seawater Si(OH)₄ budgets (de la Rocha et al., 1997; de la Rocha, 2003; Beucher et al., 2007). Siliceous sponges (Phylum Porifera, Classes Demospongea and Hexactinella) produce needle-like skeletal elements, spicules, composed of hydrated amorphous silica. Uptake of ambient Si(OH)₄ occurs via a sodium transporter, which 75 resembles active transporters isolated from other metazoans (Schroeder et al., 2004). 76 Biosilicification in sponges is controlled by two enzymes silicatein, which promotes 77 condensation reactions, and silicase, which dissolves silica (Müller et al., 2007). A 78 previously published study shows that the uptake of Si(OH)₄ results in fractionation of Si isotopes. such that spicules have some of the lightest δ^{30} Si signatures known in natural 79 systems (de la Rocha, 2003). The δ^{30} Si of sponge spicules is a potential proxy to quantify 80 81 whole ocean changes in Si cycling over timescales longer than the residence time of Si in 82 the oceans (~15 ka), as well as changes in intermediate and deep-water Si(OH)₄ 83 composition on shorter timescales (de la Rocha, 2003; de la Rocha and Bickle, 2005).

Here, we investigate the Si isotope composition of modern sponges and ambient waters from the Southern Ocean, and find a clear relationship between $[Si(OH)_4]$ and sponge spicule $\delta^{30}Si$ and Si isotope fractionation. We have then applied this calibration to sponge spicules picked from two cores in the Scotia Sea (Figure 1) to determine if $[Si(OH)_4]$ changed in response to the major climatic shifts since the Last Glacial Maximum (LGM).

90 2. Methods

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2.1. Field setting and sample materials

The modern day Southern Ocean has large $[Si(OH)_4]$ gradients (Pollard et al., 2002) and an abundance of living sponges. Therefore, we have used a transect across the Drake Passage and Scotia Sea to undertake a calibration of sponge $\delta^{30}Si$ fractionation as a function of ambient $[Si(OH)_4]$. We focus our reconstruction of past Si(OH)_4 on the Scotia

96 Sea, which plays a disproportionately important role in global oceanography. It contains 97 both Antarctic and Subantarctic Zone waters, and acts as a bathymetric channel for the 98 Antarctic Circumpolar Current (ACC) and major oceanographic fronts. This flow results 99 in intense mixing and modification between the well ventilated and Si-rich Weddell Sea 100 derived deep-waters and the rest of the global oceans (Naveira Garabato et al., 2002). For 101 the calibration, we collected and analysed living specimens from a north-south transect 102 across the Scotia Sea and Drake Passage, encompassing a range of [Si(OH)₄] (12 to 120 103 µM) and depths (300 to 2500 m; Figure 1A, 1B, Table 1). The [Si(OH)₄] increases 104 polewards and with depth through a combination of water-mass mixing, remineralization 105 and an isopycnal gradient (Figure 1B; Pollard et al., 2002). Sponges were collected aboard the R/V Nathaniel B. Palmer by either benthic trawl or dredge, and dried or frozen for 106 107 transit (April-May 2008). Water samples for Si(OH)₄ analysis were collected in niskin 108 bottles attached to deep-water CTD casts and a towed camera system (WHOI TowCam), 109 filtered immediately through 0.4 µm polycarbonate membranes (Whatman) and stored in 110 pre-cleaned HDPE bottles.

Additional samples were collected from coastal West Antarctic Peninsula (Figure 1) and the North Atlantic. Core material was obtained from two cores in the Scotia Sea, from south of the ACC (Piston core PC034, 1652m) and within the ACC (Kaston core KC081, 3662m; Figure 1; Table 2). All core material was sampled from the British Antarctic Survey.

116 2.2. Laboratory methods

117 The modern spicules were initially separated from organic matter by repeatedly heating 118 and sonicating in concentrated HNO₃ and H₂O₂ (Analar). Sediment grains were removed 119 by picking until visual inspection showed the spicules to be clear of detritus. The spicules 120 were then cleaned of any remaining organic matter and surface contaminants in a class 100 121 laboratory by heating in 50% quartz-distilled HNO₃/10% quartz-distilled HCl. Sediment 122 samples were deflocculated in sodium hexametaphosphate (5% w/v), sieved at 200 µm and 123 rinsed thoroughly in 18 MΩ.cm Milli-Q water. Approximately 20-30 spicules were hand picked, rinsed with reagent grade methanol to remove clays, and heated with H₂O₂ to 124 125 remove organic matter.

The sponge spicules were then dissolved by heating in 0.2 M NaOH (Analar) at 100 °C for 3 days (Cardinal et al., 2007). The solution was then acidified to pH~2 using 0.2 N thermally distilled HCl. Quantitative separation of Si from major ions was achieved using a cation exchange resin (BioRad AG50W-X12; Georg et al., 2006), and diluted to 300-600 ppb Si depending on machine sensitivity.

The Si isotope analyses were carried out for Si isotopes ²⁸Si, ²⁹Si and ³⁰Si using the *NuPlasma HR* MC-ICP-MS (University of Oxford) in medium resolution mode. The samples were bracketed with a concentration-matched NBS28 standard (Georg et al., 2006, 2009; Reynolds et al., 2007), and isotope ratios calculated according to Equation 1.

135
$$\delta^{\mathsf{x}} Si = \left(\left[\frac{R_{sam}}{R_{nbs28}} \right] - 1 \right) \times 1000 \tag{1}$$

136 Each sample was measured 8 times on the mass spectrometer, and mean ratios calculated. 137 Standards were checked before every batch run to ensure accuracy (either "diatomite" with δ^{30} Si = +1.26‰ (0.08), δ^{29} Si = +0.65‰ (0.03) or "Big Batch" with δ^{30} Si = -10.67‰ (0.08), 138 δ^{29} Si=-5.48‰ (0.05), parentheses indicating $2\sigma_{SE}=2\sigma_{SD}/\sqrt{n}$; Reynolds et al., 2007). Repeat 139 140 dissolutions, and repeat aliquots of the same dissolution, indicate an adequate level of 141 reproducibility comparable to previous studies (repeat measurements agree within 142 $\sim \pm 0.15\%$). Subsamples from the same modern sponge specimens were also analysed using 143 in-line gas fluorination followed by Isotope Ratio Mass Spectroscopy (IRMS; Finnigan 144 MAT 253) by M. Leng at the NERC Isotope Geosciences Laboratory (NIGL, Nottingham, UK), yielding δ^{30} Si values within error of the measurements made at the University of 145 146 Oxford by MC-ICP-MS.

147 The $[Si(OH)_4]$ of water samples from the site of sponge collection were analysed at the 148 WHOI nutrient facility, and were consistent with Southern Ocean data from existing 149 databases (Schlitzer, 2000; Garcia et al., 2006). If there were no water samples collected in 150 the vicinity of the sponges, then [Si(OH)₄] was estimated from existing data (Schlitzer, 151 2000; Garcia et al., 2006). For isotopic analysis, Si was quantitatively separated from 152 seawater using a modified Mg co-precipitation technique (Cardinal et al., 2005; Reynolds 153 et al., 2006). 2% by volume of 1M NaOH (Aristar) was added to 10-15 ml of seawater to 154 precipitate Mg(OH)₂, shaken, left for 1 hour, centrifuged and the supernatant transferred to 155 a new, clean tube. To ensure quantitative yields, the process was repeated twice adding 156 1% by volume NaOH to the supernatant each time. The precipitate was washed twice with 157 ~0.001M NaOH to remove anions, which can suppress intensity on the MC-ICP-MS. Ion 158 chromatography was used to show the precipitate wash effectively removed excess Cl⁻ and 159 F⁻ ions. The precipitate was then dissolved in 5% thermally distilled HNO₃ or HCl. The Si 160 was then purified using cation exchange as above; as before, matrix tests show the resin 161 effectively removes Na and other cations. The δ^{30} Si was measured using the same 162 protocol as for the spicules.

- 163 3. Results and discussion
- 164 3. 1. Modern calibration

Modern Southern Ocean sponge δ^{30} Si ranges from -0.70% to -4.13% (Figure 2A; 165 166 Table 3), in agreement with the limited published data from archived sponges (de la Rocha, 2003). A three isotope plot of all the silicon isotope data collected (δ^{29} Si vs. δ^{30} Si) has a 167 slope of 0.51 (± 0.01) calculated by model II regression (parentheses denotes the 95%) 168 confidence interval), which is consistent with that anticipated for kinetic equilibrium 169 (0.509 or 0.511 depending on whether Si or SiO₂ undergoes fractionation; Reynolds et al., 170 2007; Figure 2C). We find an inverse linear functional relationship between sponge δ^{30} Si 171 and $[Si(OH)_4]$ (r²= 0.75) according to Equation 2 (parentheses denotes the 95% confidence 172 interval, such that a single measurement of δ^{30} Si_{sponge} can give a Si(OH)₄ concentration to 173 approximately $\pm 20 \mu$ M). 174

175
$$[Si(OH)_4] = -30.3(\pm 8.2)\delta^{30}Si_{sponge} - 13.79$$
 (2)

176 The measured seawater δ^{30} Si_{Si(OH)4} in the collected waters agree well with modeled (Table 177 4; Wischmeyer et al., 2003; Reynolds, 2009) and published data from other sectors of the 178 Southern Ocean (de la Rocha et al., 2000). Our $\delta^{30}Si_{Si(OH)4}$ values were used to calculate a 179 fractionation factor, ϵ (Equation 3; de la Rocha, 2003), which ranges from -2.5 to -5.3‰ 180 and increases with ambient Si(OH)₄ (r²=0.56; Figure 3A).

181
$$\varepsilon \approx \delta^{30} Si_{sponge} - \delta^{30} Si_{Si(OH)_4}$$
(3)

Subsamples from the same specimen, and of two co-existing specimens of the same species, show δ^{30} Si is homogeneous within and between individuals bathed in the same water mass (Figure 2B). We cannot rule out a ~0.5‰ species-specific offset in fractionation, but it is small compared to the effect of environmental controls (~4‰; Figure 2A).

187 Other factors that are known to influence biomineralization co-vary with [Si(OH)₄] 188 in the Southern Ocean (e.g. temperature and pH; Foo et al., 2004). However, the samples 189 from coastal West Antarctic Peninsula and the deep North Atlantic show similar isotope 190 fractionations for a given $[Si(OH)_4]$ compared to sponges from the deep Southern Ocean 191 despite being collected under very different temperature, salinity and pH conditions 192 (Figure 3B-D). Consistent with growth under low Si(OH)₄ conditions, and in line with our 193 data, shallow-water sponges from a low nutrient tropical shelf show relatively heavy isotopic compositions (δ^{30} Si_{sponge} from -2 to +0.5‰; Vroon et al., 2004). Our interpretation 194 that $[Si(OH)_4]$ is the dominant control over $\delta^{30}Si_{sponge}$ appears to hold in a variety of 195 196 oceanographic settings, providing a robust proxy for paleo-[Si(OH)₄].

197 The environmental control over δ^{30} Si in sponges is likely to be a physiological 198 growth rate response to varying ambient Si(OH)₄ concentrations because ε is variable, and 199 correlates with Si(OH)₄ (Figure 3A). This control is not unexpected because silicification 200 is known to depend on Si(OH)₄ availability. Sponges produce different types of spicules 201 under Si limiting and replete conditions (Maldonado et al., 1999). Further, sponge culture 202 studies show there is a positive correlation between Si(OH)₄ availability and uptake rates, 203 and no apparent relationship with temperature (Frølich & Barthel, 1997). If the 204 fractionation process occurs at the site of Si uptake, greater uptake rates may lead to a 205 Alternatively, Si(OH)₄ availability may control the internal greater fractionation. 206 biochemical pathways involved during silicification, which may in turn determine isotopic 207 fractionation. For example, ambient Si(OH)₄ is a known modulating factor that regulates 208 the expression of silicate and silicase, and induces expression of genes for other enzymes 209 involved in biosilicification (Perovic-Ottstadt et al., 2005; Müller et al., 2007). Further 210 work is required to understand the biosilicification process and, in particular, the reactions 211 that result in isotopic fractionation.

212 The Si isotope fractionation observed during the formation of sponge spicules is 213 greater than the fractionation observed during silicification of diatom opal ($\varepsilon = -0.8$ to -214 2.1%; Cardinal et al., 2005, 2007). Although diatoms use uptake transporters and 215 condensation/dissolution enzymes distinct to sponges, the mechanisms are somewhat 216 similar and may be homologous (Foo et al., 2004). However, we suggest the two groups 217 evolved Si acquisition mechanisms, which likely impact Si isotopes differentially, at distinct points in Earth history. Siliceous sponges originated in the Precambrian (Love et 218 219 al, 2009), with a relatively low affinity for Si (Frølich & Barthel, 1997), at a time when 220 weathering of silicate rocks resulted in high oceanic Si(OH)₄ (Siever, 1992). In contrast, diatoms evolved in the Jurassic (Sims et al., 2006) with a higher affinity for Si, probably
because biological utilization by other siliceous organisms resulted in lower oceanic
Si(OH)₄ (Maldonado et al., 1999).

3.2. Downcore data

We use the modern relationship between $[Si(OH)_4]$ and $\delta^{30}Si_{sponge}$, and analyses of spicules hand-picked from two Scotia Sea sediment cores, to investigate whether deepwater $[Si(OH)_4]$ changed within the ACC on glacial-interglacial timescales, and to address the potential consequences for atmospheric pCO₂ levels in the past.

Our modern core-top sample from piston core PC034 (Figure 4A; $\delta^{30}Si_{sponge} = -$ 229 230 $3.86\pm0.17\%$, $\pm2\sigma_{SD}$) agrees well with the living sponges collected from the southern 231 Scotia Sea, growing in $[Si(OH)_4]$ of approximately 100 μ M (Figure 2A, 4B). The 232 voungest section of the northern site (KC081) has been dated at ~6 ka (Figure 4A), so we 233 are unable to measure a core-top sample at the site. In both cores, the Last Glacial 234 Maximum (LGM) value is similar to the modern, suggesting deep-water [Si(OH)₄] was not 235 significantly different at the LGM compared to today at either site (within $\sim 20 \mu$ M). The 236 similarity between LGM and modern [Si(OH)₄] in Southern Ocean deep-waters provides 237 insight into processes occurring in different regions of the water column on glacial-238 interglacial timescales.

The fractionation factor, ε , would be the most appropriate parameter to record because it would account for changes in seawater $\delta^{30}Si_{Si(OH)4}$. However, $\delta^{30}Si_{Si(OH)4}$ of seawater cannot be constrained directly downcore, and so here we present $\delta^{30}Si_{sponge}$ and assume that δ^{30} Si_{Si(OH)4} of deep waters is constant over glacial-interglacial cycles in accord with recent box modeling (Georg et al., 2009).

244 There are two important caveats when interpreting our downcore data. Firstly, age 245 models for cores in the Southern Ocean are notoriously challenging due to poor 246 preservation of benthic foraminifera and other dateable carbonates. Here, we apply the 247 best dating constraints available, based on radiocarbon measurements and stratigraphic 248 markers, locating the LGM using the abundance of diatoms and the radiolarian 249 Cycladophora davisiana (Figure 4A). The limited number of tie-points results in an 250 inherent degree of uncertainty on the age model. Secondly, as is a common problem with 251 any sedimentary proxy, there is a possibility that core components, such as spicules, 252 undergo differential transport and are of different ages to surrounding grains. However, 253 sponges are benthic and, in life, comprise a large proportion of sticky organic matter, 254 making significant post-mortem transportation of spicules less likely than, for example, 255 planktonic diatoms or foraminifera. Opal-specific dating methods may resolve any issues 256 arising from particulate transportation (Ingalls et al., 2004).

3.2.1. Deep processes: the glacial carbon reservoir

A leading hypothesis for the reduced pCO_2 during the glacial is the presence of an isolated reservoir of C-rich water in the Pacific and Southern Oceans (Marchitto and Broecker 2006; Marchitto et al., 2007). Radiocarbon records from Baja California and the Equatorial Pacific point towards a deglacial degassing of old C from a reservoir isolated during the LGM (Marchitto et al., 2007; Stott et al., 2009). Deglacial age deep-sea corals 263 from the Drake Passage, dated to Heinrich Event 1 (~16.7 ka), also show radiocarbon 264 depletion, but not to such a great extent as the Baja California record (Robinson & van de 265 Flierdt, 2009). Benthic foraminiferal records from the Southern Ocean show very light δ^{13} C signatures, indicative of a high nutrient content or significant changes in air-sea 266 267 exchange of inorganic C (Curry et al., 1998; Marchitto and Broecker, 2006; Marchitto et Specifically, there is evidence for a strong vertical δ^{13} C gradient at 268 al., 2007). 269 approximately 2.5km to the north of the polar front in the South Atlantic during the glacial 270 (Hodell et al., 2003). However, records of benthic foraminiferal Cd/Ca, a proxy for 271 dissolved phosphate (P), do not support a significant change between the nutrient content 272 of the modern and LGM deep Southern Ocean (Boyle, 1992). This discrepancy has led to 273 considerable debate in the paleoceanographic community surrounding the continued 274 presence of nutrient-poor North Atlantic derived waters in the ACC. One possibility, 275 consistent with a continued presence of NADW in the Glacial Southern Ocean, is that the light δ^{13} C was associated either with changes in air-sea exchange of C (Broecker, 1993; 276 277 Lynch-Stieglitz & Fairbanks, 1994; Marchitto & Broecker, 2006) or organic matter with 278 high C:P content (Arrigo, 1999; Elderfield & Rickaby, 2000). Alternatively, several studies have highlighted caveats and artifacts associated with both δ^{13} C in regions of high 279 280 organic matter accumulation rate (Mackensen et al., 1993) and Cd/Ca where significant 281 dissolution has occurred (McCorkle et al., 1995). When outliers in Southern Ocean 282 datasets of individual foraminiferal shells of a single benthic species, potentially arising 283 from changes in productivity in high organic accumulation rate regions, are removed the resulting "representative" δ^{13} C brings δ^{13} C more inline with Cd/Ca. These data, together 284

with ²³¹Pa/²³⁰Th records of water mass export (Yu et al, 1996), are consistent Glacial
NADW entering the ACC followed by transportation to the North Pacific in a similar
fashion to the modern ocean (Matsumoto & Lynch-Stieglitz, 1999).

Our δ^{30} Si results from KC081, situated greater than 3km, below the glacial δ^{13} C gradient, 288 (Hodell et al., 2003), do not show a change between the modern and the LGM, so we infer 289 290 that, any isotopically light C in the deep Southern Ocean was not associated with higher 291 levels of Si(OH)₄. Furthermore, our results are consistent with the benthic foraminiferal 292 Cd/Ca records, suggesting there were insignificant changes in both refractory and labile 293 nutrient concentrations in Southern Ocean deep-waters during the LGM. In this case, the light δ^{13} C signature of the LGM Southern Ocean could have been caused by a reduction in 294 air-sea exchange of inorganic C (CO₂) due to poor ventilation, or changes in the surface 295 296 temperature or residence time of subducting deep waters (e.g. Broecker, 1993; Lynch-Stieglitz & Fairbanks, 1994; Mackensen, 2001). For example, the isolation of a deep-297 water mass, due to stratification or sea-ice cover, could lead to a depletion in $\delta^{13}C$ and a 298 decrease in atmospheric CO₂ without a concurrent change in nutrients (Toggweiler, 1999). 299

Alternatively, the light δ^{13} C could originate from the remineralization of organic C that is not associated with significant quantities of Si(OH)₄ or P. This decoupling could be a result of physiological changes within the diatom populations, or shifts in the phytoplankton population structure, during the LGM. Firstly, extensive field and laboratory experiments have shown that diatoms have both lower cellular Si:N and P:N when grown under Fe replete conditions than when Fe stressed or limited (Timmermans et al., 2004; Brzezinksi et al., 2005; Price, 2005). The higher Fe conditions existing in the 307 Southern Ocean during glacials, due to enhanced dust supply (e.g. Kohfeld et al., 2005), 308 are thought to promote lower Si:N uptake ratios in diatoms compared to Fe-stressed 309 diatoms growing during interglacials (Brzezinksi et al., 2002). Secondly, the non-siliceous 310 dinoflagellate *Phaeocystis* shows a high N:P uptake ratio during modern Southern Ocean 311 blooms. If *Phaeocystis* blooms became more dominant in stratified waters during the LGM 312 (Arrigo, 1999; Elderfield and Rickaby 2000), then export of the associated organic C, and subsequent remineralization in deep-waters, could result in a depletion in $\delta^{13}C$ and an 313 314 increase in NO₃ without a concurrent increase in either $Si(OH)_4$ or P. These two scenarios 315 are not mutually exclusive, and both may have contributed to the nutrient conditions 316 prevailing in the glacial Southern Ocean.

317 3.2.2. Surface processes: glacial nutrient utilization

318 In addition to the deep-water signal, our results provide further information about 319 biological utilization in surface waters at the LGM. The Silicic Acid Leakage Hypothesis 320 (SALH) proposes that reduced productivity, due the physiological response of phytoplankton to Fe fertilization, caused a reduction in the Si to N uptake ratio in the 321 322 Southern Ocean during the LGM (Brzezinski et al., 2002; Matsumoto et al., 2002). 323 According to the SALH, a pool of excess Si(OH)₄ in Antarctic surface waters was exported 324 to lower latitudes, via intermediate water, promoting diatom production at the expense of 325 carbonate-producing coccolithophores. In this scenario, there would be an increase in 326 export of organic C with respect to inorganic C in the low latitudes associated with a rise in 327 ocean alkalinity and lowered atmospheric pCO₂ (Brzezinski et al., 2002; Matsumoto et al., 2002). Diatom δ^{30} Si records show a 0.5% change across the deglaciation, which have 328

329 been interpreted as a reduction in the fraction of Si(OH)₄ utilized in the Antarctic (de la 330 Rocha et al., 1997), subantarctic and subtropics during the LGM (Beucher et al., 2007). Since whole ocean δ^{30} Si changes are unlikely over this timescale (Georg et al., 2009), our 331 332 new record demonstrates that the decrease in Si(OH)₄ surface depletion in the subantarctic 333 region (Beucher et al., 2007) was not caused by changes in the [Si(OH)₄] concentration of 334 upwelling waters (Figure 4B). Instead it must have been a consequence of either an 335 increase in upwelling intensity or reduced surface utilization. We argue above that the 336 geochemical evidence points towards an increase in stratification in the ACC south of the polar front during the LGM, so the diatom δ^{30} Si data is best explained by changes in the 337 efficiency of surface utilization (Beucher et al., 2007). Records of nitrogen isotopes ($\delta^{15}N$) 338 of diatom-bound organic matter indicate an increase in utilization of nitrate in surface 339 waters (Sigman et al., 1999; Robinson et al., 2004). This can be reconciled with the 340 diatom δ^{30} Si data if there was a large-scale physiological change in diatoms reducing Si:N 341 342 and P:N uptake ratios (e.g. Brzezinksi et al., 2005), or an increase in productivity by non-343 siliceous phytoplankton, such as *Phaeocystis* (Arrigo, 1999; Elderfield and Rickaby, 2000).

344 3.2.3. Implications for glacial pCO₂

Our new sponge δ^{30} Si data, when combined with other geochemical proxies, bring new insight into the mechanisms behind the lower glacial atmospheric pCO₂. Deep-water C and macronutrients are decoupled on glacial-interglacial timescales as a result of physical processes, such as ocean stratification or sea-ice cover, which lock-up C in the deep ocean and lower atmospheric pCO₂ (Toggweiler, 1999; Stephens and Keeling, 2000). 350 An increase in the biological export of C from the surface to the deep ocean is consistent 351 with all of the currently available data if there was either a concurrent decrease in diatom 352 silicification, or a shift away from siliceous to organic-walled phytoplankton production, in 353 the Southern Ocean. Whilst this could increase the C:Si and C:P content of the deep 354 ocean, without significant mineral ballast a lightly-silicified diatom or *Phaeocystis* 355 dominated biological pump would not export as C efficiently as one dominated by heavily-356 silicified diatoms (Jin et al., 2006), limiting the impact on atmospheric pCO₂. However, 357 the excess Si(OH)_s resulting from a decrease in surface utilization in the Southern Ocean 358 could have been exported away from the subantarctic via mode waters without impacting 359 the deep-water Si inventory. This would have the potential to impact atmospheric pCO_2 by 360 increasing the relative productivity of diatoms in the lower latitudes (Matsumoto et al., 361 2002; Bradtmiller et al., 2007).

362 3.2.4. The deglaciation

363 During the deglacial (after ~ 18 kyr), our records indicate a depletion in deep-water 364 $[Si(OH)_4]$ both within and south of the ACC (Figure 4B). In the late Holocene, deep-water 365 $[Si(OH)_4]$ recovers to modern values. Given the uncertainty on the age models, we cannot 366 pin point the exact timing of these changes but the decline is a robust observation in both 367 cores. Our best estimate for the maximum rate of change of deep-water $[Si(OH)_4]$ is 368 approximately 10 μ M ky⁻¹

We hypothesize two possible mechanisms behind this decline in deep-water
Si(OH)4: large scale changes in nutrients arising from decomposition of biogenic particles

371 in the deep Southern Ocean ("remineralized nutrients") or localized changes in unutilized nutrients subducted during deep-water formation ("preformed nutrients"; Marinov et al., 372 373 2006). Firstly, our data are consistent with storage of Si(OH)₄ in the form of a large-scale 374 opal deposition in the Southern Ocean during the deglacial (10-16 kyr), caused by winddriven upwelling of water enriched in Si(OH)₄ relative to the surface. Enhanced opal 375 fluxes during the deglaciation, coupled with 231 Pa/ 230 Th activity ratios, have been observed 376 377 in Pacific and Atlantic Sectors of the Southern Ocean and equatorial regions (Anderson et 378 al., 2009). Such a significant and widespread burial of opal could have resulted in a drop in 379 deep ACC remineralized Si(OH)₄, whilst the enhanced upwelling drove a concurrent rise 380 in pCO₂ (Anderson et al., 2009). The decline in deep-water $[Si(OH)_4]$ would have led to a 381 negative feedback, by limiting the amount of productivity that could be supported by 382 upwelling water and an eventual restoration of export production rates, opal fluxes and 383 [Si(OH)₄] gradients over time. Such changes in remineralized nutrients cannot change 384 global deep-water [Si(OH)₄] over timescales of less than the residence time of Si in the 385 whole ocean (~15 ky; Georg et al., 2009). However, we constructed a simple one-box model for the deep Southern Ocean alone (south of 50° S, > 400 m depth, area ~ 25 x 10^{12} 386 m^2), using a mean modern [Si(OH)₄] of 100 µM and mean depth of 4000 m (Garcia et al., 387 2006), and modern opal accumulation fluxes of ~ 0.2 mol Si m⁻² yr⁻¹ (Pondaven et al., 388 2000). Using this model, we estimate changes in the opal burial rate relative to Si inputs of 389 390 $\sim 20\%$ (less than observed in core TN057-13-4PC; Figure 4b, Anderson et al., 2009) could 391 cause changes of the order of 10 µM in 1000 years, which are sufficient to explain our 392 observations.

393 Alternatively, the rapid rates in change indicate the decline in deep-water $[Si(OH)_4]$ 394 during the deglaciation was at least in part a result of localized inputs of preformed 395 nutrients. For example, the retreat of the Weddell Sea ice shelf during the deglacial and 396 early Holocene "Hypsithermal" may have changed deep-water formation processes and led 397 to a decline in the input of highly Si-enriched Weddell Sea Deep Water (WSDW) into the 398 ACC (Yoon et al., 2007). This could explain both the rapid decline in deep-water $Si(OH)_4$ 399 and the apparent divergence in the records between ~ 6-10 ky (higher [Si(OH)₄] from the 400 southern core compared to the northern core; Figure 4B). Resumption of WSDW input to 401 the ACC could be in part responsible for the recovery in deep-water Si(OH)₄ in the later 402 Holocene. Future research should focus on constraining this climatically important time, 403 in order to understand the response of Southern Ocean deep-water circulation during 404 periods of global warming.

405 4. Summary and conclusions

In summary, we show the potential for combined δ^{30} Si records from diatoms and 406 sponges to constrain the surface and deep components of the Si biological pump. We 407 investigate fractionation by modern deep-sea sponges, and find the $\delta^{30}Si$ of spicules and 408 409 fractionation with respect to ambient seawater correlate with the ambient $[Si(OH)_4]$. We 410 then apply this calibration to two downcore records from the Scotia Sea. Our results show 411 that the deep-waters of the LGM Southern Ocean were not enriched in Si(OH)₄, suggesting 412 that the isolated reservoir of old, isotopically depleted C was not associated with an 413 increase in either Si(OH)₄ or P. Instead, the Southern Ocean resulted in lower atmospheric 414 pCO₂ during the glacial largely due to reduction in air-sea exchange of C via a physical

415 process. If there was an enhanced glacial biological pump in the Southern Ocean, it was 416 likely to have been dominated by lightly-silicified diatoms or organic-walled 417 phytoplankton (e.g. *Phaeocystis*) resulting in a concurrent increase in surface nitrate 418 utilization, and a change in deep-water nutrient ratios that may not have been associated 419 with a considerable increase in pump efficiency. Utilization of Si-rich upwelling waters by 420 diatoms was lower during the LGM, providing a surplus of Si(OH)₄ that could then be 421 exported to lower latitudes via intermediate waters, where enhanced export production 422 could lower atmospheric CO₂. During the deglaciation, there is a robust and significant 423 decrease in deep-water Si(OH)₄, which could be associated with a pulse of opal burial 424 linked with wind-driven upwelling and concurrent rise in pCO₂, or a change in water mass 425 inputs.

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 thermohaline circulation inferred from radiochemical data. Nature. 379, 689-694.
- 619
- 620
- 621
- 622 The authors would like to thank the captain and crew of the R/V Nathaniel B. Palmer (cruise NBP0805;
- 623 Program Manager Thomas Wagner). Thanks also to Rhian Waller (University of Hawaii), Laura Schejter

624 (INIDEP, Argentina), Jade Berman (University of Victoria, Wellington), Nicky White (University of
625 Cambridge) and Andy Clarke (British Antarctic Survey) for sponge samples and help with identification.

- 626 Thanks to Chris Siebert, Helen Williams, Sune Nielsen, Ros Armytage and Paul Savage (University of
- 627 Oxford) for assistance in the laboratory, Melanie Leng (NIGL) for additional silicon isotope measurements,
- 628 Maureen Auro and Paul Henderon (WHOI) for assistance with nutrient analyses and Kathy Scanlon (USGS)
- 629 for map production. Core material and age model from Claire Allen and Claus-Dieter Hillenbrand (British
- 630 Antarctic Survey). Thanks to Louisa Bradtmiller (WHOI), editor Peter deMenocal and three anonymous
- 631 reviewers for constructive reviews and discussion. Cruise NBP0805 was funded by NSF Office of Polar
- 632 Programs (OPP) Antarctic Sciences (grant number ANT-0636787). Data from the Palmer LTER data archive
- 633 were supported by Office of Polar Programs, NSF grants OPP-9011927, OPP-9632763 and OPP-0217282.
- 634 The work was funded by the Natural Environment Research Council (NERC) grant NE/F005296/1 and an
- 635 Antarctic Science Bursary. The authors declare no competing financial interests.
- 636 Correspondence and requests for materials should be addressed to K.R.H. (e-mail: kathh@earth.ox.ac.uk).

Figure 1: A) Location of sample sites in the Southern Ocean and coastal
Antarctica. Grey stars indicate modern sponge sample sites; grey squares indicate
deep-water samples; grey dotted circles show cores PC034 and KC081. Map
production by K. Scanlon (USGS).

- B) Dissolved silicic acid concentrations from across the Drake Passage. Drawnusing Ocean Data View (Schlitzer, 2000).
- Figure 2: A) The relationship between modern sponge silicon isotope composition and ambient seawater Si(OH)₄ concentration (r^2 =0.75; Equation 2). Filled symbols show values for Hexactinellid sponges; open symbols show values for Demosponges. There is no consistent difference between the two classes of sponges, although there may be species-specific fractionation (~0.5‰). Error bars show 2 σ_{SD} .
- B) Isotopic homogeneity between individuals of the same species from one locality, and within an individual. The black symbols show duplicate measurements of spicule silicon isotopes taken from three different specimens; the white triangles show the silicon isotopic composition of two specimens of *Rosella* collected from the same site; the white circles show the silicon isotopic composition of two different types of spicules from the same specimen (dermal and perenchymal). Error bars show $2\sigma_{SD}$.

656 C) A three-isotope plot for all sponge samples. The equation was calculated using 657 model II regression, and numbers in parentheses indicate 95% confidence 658 intervals of the slope and intercept respectively. The relationship between 659 $\delta^{29}Si_{sponge}$ and $\delta^{30}Si_{sponge}$ is consistent with mass dependent fractionation under 660 kinetic equilibrium. Error bars show $2\sigma_{SD}$.

661 Figure 3: Fractionation of silicon isotopes by sponges. The black circles show the 662 fractionation by sponges from the deep Southern Ocean; the grey squares show 663 the fractionation by sponges from the West Antarctic Peninsula; the white triangles 664 show the fractionation by sponges from the deep North Atlantic (assuming Atlantic deep water δ^{30} Si_{Si(OH)4} = 1.6‰; Reynolds, 2009). Fractionation is plotted against 665 ambient A) Si(OH)4 concentrations; B) salinity; C) temperature and D) pH (data 666 from the British Antarctic Survey and Garcia et al., 2006). 667 Error bars are propagated from $\pm 2\sigma_{SD}$ of $\delta^{30}Si_{Si(OH)4}$ and $\delta^{30}Si_{Sponge}$. 668

Figure 4: A) Age models for the two cores based on magnetic susceptibility 669 670 (dashed lines) and diatom abundances (squares). Note for PC034, the shallower 671 peak in magnetic susceptibility has been interpreted as showing the Antarctic 672 For KC081, age constraints are also available from the Climate Reversal. 673 occurrence of the radiolarian Cycladophora davisiana (triangles), which are abundant in glacial sediments, and corrected radiocarbon ages (indicated by black 674 675 arrows) showing core top ~ 6.3 ka, 0.23 mbsf ~ 11.3 ka and 1.59 mbsf ~ 23.5 ka 676 (NERC Publication Code AA-28113-5). Data and interpretation from C. Allen 677 (BAS). Also shown are locations of the cores with respect to major oceanographic fronts, although these can vary by several degrees in the modern setting (SAF = 678 679 Sub-Antarctic Front; PF = Polar Front; SACCF = Southern Antarctic Circumpolar 680 Front; SB = Southern Boundary of the ACC, adapted from Naveira Garabato et al., White symbols correspond to the northern core (KC081) and black 681 2002). 682 symbols to the southern core (PC034).

683 B) Composition of sponges (error bars show $2\sigma_{SD}$) and reconstructed deep-water Si(OH)₄ concentrations (95% confidence intervals ~ 20 μ M; Equation 2). White 684 685 symbols correspond to the northern core (KC081) and black symbols to the southern core (PC034). The white box shows δ^{30} Si of modern sponges found near 686 KC081: the black box shows δ^{30} Si of modern sponges found near PC034. The 687 age model is based on the information in A). Also shown: opal flux and 688 ²³¹Pa/²³⁰Th from core TN057-13-4PC from south of the Polar Front in the Atlantic 689 690 Sector (Anderson et al., 2009); EPICA ice core records of pCO₂ and δ D, a proxy for local temperature (Indermüle et al., 1999; Fluckiger et al., 1999; Monnin et al., 691 692 2001, 2004).

Sample code	Classification	Location	Depth (m)	Method	Lat (°S)	Lon (°W)
NBP0805-TB1-3	Unidentified Demosponge	Burdwood Bank	306- 333	Blake trawl	54.48	62.18
NBP0805-TB1-6	Mycalidae	Burdwood Bank	306- 333	Blake trawl	54.48	62.18
NBP0805-TB1-11	Unidentified Hexactinellid	Burdwood Bank	306- 333	Blake trawl	54.48	62.18
NBP0805-TB3-21	Mycalidae	Burdwood Bank	2215- 2343	Blake trawl	54.91	62.03
NBP0805-TB4-3	Unidentified Demosponge	Burdwood Bank	804- 828	Blake trawl	54.74	62.21
NBP0805-TB4-24	Unidentified Demosponge	Burdwood Bank	804- 828	Blake trawl	54.74	62.21
NBP0805-TB4-27	Rosella	Burdwood Bank	804- 828	Blake trawl	54.74	62.21
NBP0805-T01-27	Rosella	Elephant Island	407- 428	Otter trawl	61.25	56.42
NBP0805-T01-57	Rosella	Elephant Island	407- 428	Otter trawl	61.25	56.42
NBP0805-TO3-100	Unidentified Demosponge	Drake Passage	689- 914	Otter trawl	60.56	65.97
NBP0805-TO3-111	Unidentified Demosponge	Drake Passage	689- 914	Otter trawl	60.56	65.97
NBP0805-DR7-47	Acoelocalyx	Scotia Sea	2109- 2422	Dredge	54.88	40.69
NBP0805-DR13-47	Acoelocalyx	Scotia Sea	2097- 2477	Dredge	59.50	42.50
NBP0805-DR16-47	Acoelocalyx	Scotia Sea	930-	Dredge	61.10	56.45

			1070			
NBP0805-DR29-47	Acoelocalyx	Drake Passage	758- 759	Dredge	60.56	65.91
NBP0805-DR34-47	Acoelocalyx	Drake Passage	899- 838	Dredge	59.73	68.74
NBP0805-DR35- 111	Unidentified Demosponge	Drake Passage	672- 718	Dredge	59.72	68.88
NBP0805-DR40-47	Acoelocalyx	Drake Passage	1294- 1351	Dredge	59.73	68.93
CRS-956	Unidentified Demosponge	Anvers Island	600	Dredge	64.78	65.30
RB-Mycale	Mycale	Adelaide Island	10	Hand collection	67.57	68.23

Table 1: Samples used in the modern calibration collected from the Southern
Ocean. CRS-956 was collected by R. Waller (University of Hawaii) from near
Anvers Island; RB-Mycale was collected by J. Berman (BAS) from near Adelaide
Island.

Core	Depth (m)	Latitude (°S)	Longitude (°W)
PC034	1652	59.79	39.60
KC081	3662	56.74	42.97

Table 2: Locations of cores used in this study.

Sample code	Т (°С)	S (psu)	рН	Si(OH)4 (μM)	δ ³⁰ Si (‰)	δ ²⁹ Si (‰)	*δ ³⁰ Si (‰)
NBP0805-TB1-3	4.3	34.17	7.96	12	-0.70(0.04)	-0.38(0.02)	-0.36
NBP0805-TB1-6	4.3	34.17	7.96	12	-1.01(0.08)	-0.51(0.05)	-0.52

						-	
Repeat dissolution					-1.15 (0.04)	-0.61 (0.03)	-0.59
NBP0805-TB1-11	4.3	34.17	7.96	12	-1.69(0.03)	-0.84(0.03)	-0.86
NBP0805-TB3-21	1.8	34.69	7.70	97	-2.79(0.02)	-1.45(0.02)	-1.42
NBP0805-TB4-3	3.1	34.35	7.80	56	-1.55(0.03)	-0.83(0.02)	-0.79
NBP0805-TB4-24 Dermal Perenchymal	3.1	34.35	7.80	56	-2.87(0.04) -2.96(0.04)	-1.41(0.02) -1.51(0.01)	-1.46 -1.51
NBP0805-TB4-27	3.1	34.35	7.80	56	-2.54(0.04)	-1.28(0.02)	-1.30
NBP0805-T01-27	1.9	34.65	7.71	90	-3.83(0.05)	-1.94(0.02)	-1.95
NBP0805-T01-57	1.9	34.65	7.71	90	-3.51(0.04)	-1.83(0.02)	-1.79
NBP0805-TO3-100	2.4	34.63	7.80	75	-3.06(0.04)	-1.59(0.02)	-1.56
NBP0805-TO3-111	2.4	34.63	7.80	75	-2.88(0.09)	-1.46(0.06)	-1.47
Repeat dissolution					-3.18 (0.05)	-1.64 (0.03)	-1.62
NBP0805-DR7-47	1.5	35	7.7	120	-3.76(0.05) -3.58(0.02)	-1.92(0.05) -1.77(0.03)	-1.92 -1.82
NBP0805-DR13-47	1.5	35	7.7	110	-3.20(0.05)	-1.59(0.02)	-1.63
NBP0805-DR16-47	1.48	34.72	7.76	96	-4.13(0.05)	-2.10(0.04)	-2.10
NBP0805-DR29-47	2.4	34.63	7.80	100	-3.91(0.04) -3.91(0.05)	-2.02(0.01) -2.02(0.02)	-1.99 -1.99
NBP0805-DR34-47	2.3	34.62	7.80	75	-2.54(0.03)	-1.31(0.02)	-1.30
NBP0805-DR35-111	2.4	34.64	7.80	70	-2.86(0.04)	-1.44(0.03)	-1.46
NBP0805-DR40-47	1.9	34.70	7.72	94	-2.92(0.03)	-1.47(0.01)	-1.49
CRS-956	1.5	34.72		107	-3.36(0.04)	-1.72(0.02)	-1.71

RB-Mycale	-0.4	33	8.19	50	-2.39(0.05)	-1.21(0.03)	-1.22
Repeat dissolution					-2.34(0.03) -2.40(0.03)	-1.21(0.02) -1.24(0.03)	-1.19 -1.22

Table 3: Silicon isotopic composition of modern sponge spicules. Temperature, salinity, pH and nutrient values courtesy of WHOI, BAS, Palmer LTER, WOCE (Schlitzer, 2000) and World Ocean Atlas 05 (WOA05; Garcia et al., 2006). Numbers in parentheses are $2(\sigma/\sqrt{n})$. * δ^{30} Si is the expected δ^{29} Si value calculated assuming mass dependent fractionation:

709 $*\delta^{30}Si = 0.51 \times \delta^{30}Si_{sponge}$

Sample	Lat (°S)	Long (°W)	Depth (m)	Si(OH)4	δ ³⁰ Si
CTD2-1-1	54.88	62.14	2190	96.5	1.06 (0.04)
CTD2-13-7	54.88	62.14	805	56.0	1.28 (0.03)
CTD2-17-9	54.88	62.14	300	11.8	1.76 (0.05)
CAM3-bottom	61.28	56.42	400	95.8	1.03 (0.05)
CAM7-1500	60.55	65.94	1500	100.6	1.10 (0.06)
repeat					1.04 (0.06)
CAM8-600	59.89	68.86	600	40.4	1.40 (0.06)

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Table 4: Seawater Si(OH)₄ and δ^{30} Si values. Si(OH)₄ measured by WHOI nutrient

facility; δ^{30} Si measured using a co-precipitation method. Numbers in parentheses

713 are $2(\sigma/\sqrt{n})$.

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