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1	The release of DOC from seagrass wrack and its implications for trophic connectivity
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13	The export of old leaves and stems (wrack) from seagrass meadows provides a mechanism for
14	trophic connectivity among coastal ecosystems. Since little of this wrack is consumed by
15	mesograzers, leached DOC may determine the importance of wrack as a trophic subsidy.
16	However, few studies have examined the effect of seagrass type or age on the release of DOC
17	or its bioavailability. We examined the amount and composition of DOC released from
18	different wrack (Posidonia sinuosa, Amphibolis antarctica and the alga Laurencia sp.). We
19	then examined the effect of age on DOC leaching from <i>P. sinuosa</i> wrack. The bio-availability
20	of the DOC was also assessed using a bacterial bioassay.
21	
22	The rate of DOC leaching from <i>Posidonia sinuosa</i> leaves decreased exponentially with time.
23	According to that exponential model, about 50% of the total DOC release occurred in the first
24	14 days and it would require a further 2.94 years to release the same amount again. Fresh
25	algae (Laurencia sp.) leached the greatest amount of DOC in the first 16 h (6.7 mg kg ⁻¹ FW
26	wrack), followed by fresh P. sinuosa leaves (1.7 mg kg ⁻¹ FW), A. antarctica leaves (1.1 mg

kg⁻¹) and stems (0.6 mg kg⁻¹), 4 wk old *P. sinuosa* (67 mg kg⁻¹) and fine detritus (74 mg kg⁻¹).
In all cases, the composition of the DOC was similar and dominated by the hydrophilic
component (in *P. sinuosa*, predominantly sugars and amino acids). Leachates from all fresh
wrack supported bacterial growth over 24 h. Leachate from older wrack either failed to
support bacterial growth, or only supported it for a limited time. Given the exponential decay
in DOC release rate, the interacting timescales of transport and leaching will affect the value
of wrack as a vector for trophic subsidies.

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36 INTRODUCTION

37 Seagrass meadows are conspicuous and highly productive components of coastal ecosystems 38 worldwide (Green & Short 2003). A portion of seagrass production is continually shed as old 39 leaves, which can contribute significantly to wrack (detached macrophyte) accumulations in 40 adjacent coastal habitats (Kirkman & Kendrick, 1997; Mateo, 2010). Export rates of detached 41 leaves from meadows varies enormously but can be as high as 100% of leaf production 42 (Cebrian & Duarte, 2001; Mateo, 2010; Mateo et al. 2006). Given that these older leaves also 43 contain nutrients other than carbon, even after re-sorption prior to shedding (e.g., Prado et al., 44 2008) this wrack export represents a significant potential loss of nutrients from the habitat and 45 a potential trophic subsidy to adjacent recipient habitats, particularly in oligotrophic 46 environments where alternative sources of nutrient may be limited.

47

48 Despite this potential, there is little published evidence of seagrass wrack being an important 49 source of nutrient to adjacent habitats. Several studies concluded that seagrass wrack was 50 unlikely to be a significant contributor to meso-grazer production in recipient habitats, 51 including unvegetated marine habitats (Hyndes & Lavery 2005), beach ecosystems (Ince et 52 al., 2007), surf zones (Crawley et al., 2009) and within seagrass meadows (Smit et al., 2005; 53 Smit et al., 2006). Wrack typically has a large proportion of macro-algae, and many 54 mesograzers demonstrate a preference for this over seagrass detritus (Doropoulos et al., 2009), likely due to the lower C:N ratio of algae. This suggests that if the nutrients within 55 56 seagrass wrack are to be recycled within meadows or provide a subsidy to adjacent systems, then mechanisms other than direct consumption of seagrass detritus must be important, 57 58 microbial pathways utilizing dissolved organic carbon (DOC) being among the most likely. 59 60 Fluxes of DOC to overlying water are higher in seagrass meadows than adjacent unvegetated meadows (Barrón & Duarte 2009). Up to 50% of this DOC is consumed by bacteria (Ziegler 61 62 & Benner 1999), which can be rapidly transferred to higher trophic levels through 63 consumption by flagellates and ciliates (Robertson et al., 1982). The seagrass leaves themselves are a major contributor to this DOC flux. Through exudation or autolysis and 64 65 leaching, seagrass leaves typically release 2-10% of net primary production (Moriarty & Pollard 1982, Barrón & Duarte 2009), and this leaf-derived DOC has been shown to support 66 bacterial production (Brylinsky, 1977; Kaldy et al., 2006). 67 68 69 Because seagrass leaves degrade much more slowly (Klumpp & Vandervalk 1984, Moore & 70 Fairweather 2006) than the rate at which they are transported by currents and storm-driven 71 advection, DOC leaching may occur over extended periods of time and encompass a range of 72 different habitats. This may provide a mechanism of cross-habitat trophic subsidy not 73 dependent on the direct consumption of leaves. Thus, the loss of DOC from seagrass leaves is 74 potentially a key contributor to the total DOC flux from seagrass ecosystems and cross-habitat 75 trophic subsidies.

76

Almost all of the studies that have examined the loss of DOC from seagrass leaves have
focused on living or fresh leaves (e.g. Brylinsky, 1977; Robertson et al., 1982; Wetzel &

79 Penhale, 1979). Yet there is evidence that the age of detached leaves has a significant impact 80 on DOC leakage and in the composition (and therefore bioavailability) of that DOC (Maie et 81 al., 2006; Velimirov, 1986), which may be crucial for the transfer of seagrass-derived 82 nutrients to adjacent ecosystems. Furthermore, the amount and rate of DOC leaching, and the 83 ability of microbes to utilise the leachate, varies among different vascular plants (Benner et 84 al., 1986; Maie et al., 2006), suggesting that the export and bioavailability of seagrass-derived 85 DOC may be species-dependent. Among the seagrasses, inter-specific differences could 86 relate to the anatomy of the plants (e.g. membranous, leafy species such as *Posidonia* spp. 87 versus heavily lignified species such as Amphibolis spp.) or the amounts and forms of soluble 88 compounds within the tissues. These differences among species in DOC leaching and its 89 apparent bioavailability led Maie et al. (2006) to call for more studies into the bioavailability 90 of the DOC fractions that are released from macrophyte leaves. Further, the potential 91 significance of seagrass as a source of DOC to the coastal zone, coupled with the rapid 92 decline in seagrass cover in recent decades (Green & Short 2003), prompted Barrón & Duarte 93 (2009) to call for more studies on seagrasses to understand the export of DOC from these 94 systems and its significance.

95

96 In this paper, we compare the amount, composition and bioavailability of DOC released from 97 different types of seagrass wrack. We also examined the effect of wrack age on the amount of 98 DOC released and its bioavailability. The main objectives were to: 1) determine the amount 99 and functional composition of DOC released from different wrack materials. We tested the 100 hypotheses that the amount of DOC released will vary among different types of wrack and 101 that functional composition will vary among types of wrack; 2) examine whether the amount 102 and composition of DOC released from *Posidonia sinuosa* wrack depended on wrack age. We 103 tested the hypothesis that the amount of DOC released would diminish with age of wrack and 104 that the composition would differ among wrack of different ages; and 3) assess the bio105 availability of DOC released from wrack and whether this is affected by wrack age. We

106 hypothesised that bacterial biomass would increase more rapidly when grown in leachate than

107 blank solution, and more rapidly in leachate from fresh wrack than aged wrack.

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- 109

110 MATERIALS & METHODS

111 Study region

112 The study was conducted on wrack accumulations in Geographe Bay (Fig 1), a 100 km wide 113 north-facing embayment, on the south-western coast of Australia. It is a relatively protected 114 bay with extensive beds of seagrass Posidonia sinuosa Cambridge & Kuo, and Amphibolis 115 antarctica (Labillardiere) Sonder et Ascherson ex Ascherson. Posidonia and Amphibolis are 116 the dominant meadow-forming genera of seagrasses in south-west Australia. *Posidonia* spp. 117 produce above-ground shoots with 1-3 strap-like leaves that are periodically shed. 118 Amphibolis spp. have heavily lignified and persistent stems on which clusters of short leaves 119 are borne (Ducker et al., 1977). The stems are often heavily covered by epiphytes, 120 particularly red (rhodophyte) macro-algae such as Laurencia sp (Lavery & Vanderklift 2002). 121 Extensive accumulations of detached seagrass leaves and stems are typical of the region, 122 especially in winter (McMahon et al., 1997). The bay is exposed to the NW-winds that 123 characterise the early phase of storms in the region. This exposure results in the transport of 124 wrack throughout the Bay, providing a high degree of connectivity among seagrass and other 125 habitats in the region. The wrack is typically dominated by *P. sinuosa*, with significant 126 amounts (up to 30% by weight) of A. antarctica at times and generally a low amount (but 127 occasionally up to 15%) of red algae (McMahon et al., in review). Geographe Bay is an 128 oligotrophic waterbody and, as such, the decomposition of wrack in sub-tidal and beach 129 habitats may be a vital source of recycled nutrients (Robertson & Lenanton 1984).

131 Release of DOC from different wrack material

132 The effect of wrack type on the amount and composition of DOC leachate was examined by 133 comparing DOC leached from wrack commonly found in the region: the seagrasses Posidonia 134 sinuosa (leaves) and Amphibolis antarctica (leaves and stems); the red algae Laurencia sp., 135 which is common as both a free-living algae and epiphytic on seagrasses; and the fine 136 particulate organic fraction (>0.1 - <1 mm) of beach-cast wrack that had been on the beach 137 for at least 2 months. Fresh samples of *P. sinuosa*, *A. antarctica* and *Laurencia sp.* were 138 collected from meadows in 0.5 m depth. Beach-cast wrack was collected from Busselton 139 Beach, Geographe Bay (S 33° 39.317', E 115 ° 16.812') and then sieved to separate the fine 140 fraction. Samples were stored at 4°C, for no more than 12 hours, until leachate was collected. 141 142 The effect of wrack age on DOC leaching was examined using one wrack type, P. sinuosa 143 leaves. We focused on *P. sinuosa* since this was the dominant component of the beach cast 144 wrack, typically accounting for more than 60% by biomass. Fresh samples of *P. sinuosa* were 145 collected from a meadow in 0.5 m depth. The fresh material was transported immediately to 146 the laboratory and stored in a cool room at 4°C for a maximum of 2 days until leachate was 147 collected. For 'aged' material, approximately 500 g of fresh P. sinuosa was placed in nylon 148 mesh litterbags (mesh size < 5 mm) which were then placed on the surface of wrack 149 accumulations on the beach, held in place by pickets driven into the wrack accumulations, and 150 exposed to the ambient weather conditions. Replicate bags (n=3) were removed after 1, 2, and 151 4 weeks and returned to the laboratory for leachate experiments. 152 153 **DOC** leachate extraction

Each wrack type and age was incubated to extract DOC. All leaves of both seagrass species
and the *Laurencia* thallus were lightly scraped with a razor blade to remove algal epiphytes.
For each wrack type, four replicate samples of 100-150 g wet weight plant material were

placed in 0.5 L of sterile, artificial seawater (ASW: Red Sea Salt[™] at 35 ppt) in acid washed 2 157 158 L glass beakers. The beakers were sealed with scientific-grade rubber stoppers and incubated 159 for 16 h at 18°C on shaker trays and with periodic, gentle agitation. Blanks were prepared as 160 described above but with no wrack. After 16 h, the leachate was filtered through a series of Whatman GFF filter papers and then a 0.45 µm, hydrophilic polypropylene membrane filter 161 162 (Pall Life Sciences) and analysed for total DOC with a Shimadzu Total Organic Carbon 163 Analyser 9000. Scraped leaves were used in the experiment to avoid DOC release from 164 epiphytic algae. However, to test for any effect of scraping on DOC release, simultaneous incubations were performed on unscraped leaves with epiphytes gently removed by hand. 165 166 Even with the removal of epiphytic algae and animals, a microbial biofilm is likely to remain 167 on the leaf surface, which can reduce the flux of DOC to the surrounding waters through 168 direct uptake (Maie et al. 2006). Consequently, the changes in DOC concentration of the 169 incubating water are referred to as Net DOC release.

170

171 The initial leaching experiments indicated high release rates of DOC from fresh seagrass 172 wrack in the first 1-2 weeks of aging. To obtain increased temporal resolution of early 173 leaching rates, the experiment was repeated using *P. sinuosa* leaves to determine the change 174 in release rate over this initial period of high DOC leaching. Three replicate leaf samples (180 175 g wet weight) were incubated in 1.4 L of ASW in acid washed 2 L glass beakers. A blank of 176 ASW was incubated at the same time. Samples of the leachate were collected after 1, 3, 5, 7, 177 10, 14 days of incubation, and the ASW replaced each time. The leachate was filtered and 178 analysed as described above.

179

180 Characterization of DOC

181 DOC composition was characterized by fractionation using open column chromatography
182 following modified methods of Chow *et al.* (2004) and Cleveland *et al.* (2004). The leachate

183 was fractionated into three components: 1) hydrophobic DOC (fulvic and humic acids;

184 Hughes, 2007) by retention on DAX-8 resin; 2) transphilic DOC, by retention on the XAD-4

resin; and 3) hydrophilic DOC, the eluent passing through the DAX-8 and XAD-4 column

186 (Thurman & Malcolm 1981). The hydrophilic fraction is composed predominantly of low

187 molecular weight compounds, including carbohydrates and amino acids (Cleveland et al.

- 188 2004).
- 189

DAX-8 Superlite (Sigma-Aldrich) and XAD-4 Amberlite (Sigma Aldrich) resin columns were prepared in a similar manner, following the manufacturer's instructions. The resin was mixed with water (milli-Q) to form a slurry and poured into a glass column (30 x 1.5 cm) fitted with a tap. The bed volume for the resin was approximately 20 mL. The column was conditioned by passing 200 mL Milli-Q water though the column drop-wise, followed by six alternating washes of 0.1 M NaOH (40 mL) and 0.1 M HCl (40 mL), again eluted drop-wise. The final wash was with 0.1 M HCl, to leave the column acidified.

197

198 The DOC leachate (200 mL) was acidified with 35% HCl to pH 2 then passed through the DAX-8 resin (dropwise or at $< 3 \text{ mL min}^{-1}$). The first 10 mL of eluent was discarded (as it 199 200 was simply displaced acid). When most of the leachate had been applied to the column, 0.1 M 201 HCl (2 bed volumes or 40 mL) was applied to the top of the column. The acid was passed 202 through the column dropwise to elute all but the hydrophobic fraction. The eluent (leachate 203 and acid) was then passed through the XAD-4 column in a similar manner, in this instance the 204 acid eluting the hydrophilic fraction. A sample of de-ionised (Milli-Q) water was passed 205 through the columns and treated in an identical way as the leachates to act as an analytical 206 blank.

208	The hydrophobic and transphilic fractions retained on the DAX-8 and XAD-4 resins,
209	respectively, were elute with 0.1 M NaOH. The volume of base added was typically 5 bed
210	volumes (100 mL) or until the absorbance of the eluent at 254 nm was similar to the blank,
211	indicating an absence of DOC.
212	
213	The total volume of each fraction was recorded. All pre-filtered leachate samples and
214	fractionated samples were acidified with 35% HCl to pH 2 and analyzed for total DOC with a
215	Shimadzu Total Organic Carbon Analyzer 9000. The UV absorbance for each of the acidified
216	samples was also recorded at 254 nm on a Shimadzu UV-1601 spectrophotometer.
217	
218	
219	Wrack composition on beaches
220	The DOC leaching studies were conducted on wrack collected in sub-tidal habitats and
221	incubated in submerged conditions, typical of sub-tidal seagrass wrack. Initial results showed
222	differences in the net release of DOC from wrack of different ages. Since large amounts of
223	wrack accumulate on beaches, we examined the age of beach-cast wrack to determine
224	whether beach wrack was likely to have arrived while it was 'fresh' (and with higher net DOC
225	release rates), and, therefore, whether the bulk of DOC leaching (and potential trophic
226	subsidy) occurs in sub-tidal habitats or on beaches. The composition of beach wrack was
227	determined at three sites on five occasions over the period of maximum wrack accumulation
228	on beaches (May-Oct; McMahon et al., in review). Samples were collected at Forrest Beach,
229	Volunteer Marine Rescue and Geographe Sailing Club (Fig. 1) on 19 th - 22 nd May, 9 th - 11 th
230	June, 12 th - 15 th August, 22 nd - 25 th September and 20 th - 22 nd October, 2008. At each site and
231	time, four replicate wrack accumulations were sampled. About 0.001 m ³ of wrack was
232	collected from the surface of the accumulation with a quadrat and from the sediment
233	immediately below the accumulation with a corer (90 mm I.D. x 10 cm deep). The wrack was

rinsed to remove sand and sorted into categories based on the estimated age of wrack. Age

235 was defined as either old (no green leaves) or new (green leaves or stem) on the basis of their

colour: pilot work showed that moist leaves above the surface of the sediment turned brown

237 within 2 weeks (*P. sinuosa*) or 2 - 4 weeks (*A. antarctica*) (Oldham et al., in review).

238

Bioavailability of DOC

We used a bacterial bioassay to test the bioavailability of the filtered DOC leachate produced by *P. sinuosa* leaves, *A. antarctica* leaves and stem, *Laurencia* sp. and fine particulate wrack during the 16 h. incubations, using the methods of Cleveland et al. (2007). The response of a bacterial inoculum to the different DOC leachates was observed as growth rate over a 24 h. period.

245

246 Filtered DOC leachate (200 mL) was combined with a bacterial inoculum (2 mL) in acid-247 washed glass flasks, wrapped and capped in aluminum foil. A bacterial inoculum was created 248 by combining 100 g of moist beach sediment, 100 g of moist wrack and 800 mL sterile 249 artificial seawater. This was left in the dark for 24 h at 18°C and then filtered through a 250 Whatman 3 filter paper with the filtrate used as the bacterial inoculum. For each DOC 251 leachate, four replicates and four blanks (200 ml ASW + 2 ml bacterial inoculum) were 252 incubated at 25°C. Triplicate 1 mL sub-samples were taken after 0, 3, 18 and 24 h of 253 incubation and fixed with 0.5% glutaraldehyde for 15 min in the dark (Marie et al. 1997) then 254 stored in liquid nitrogen until further processing. Heterotrophic bacterial cell counts were 255 determined on a FACS Canto II flow cytometer. Samples were diluted with TE buffer (1:50 dilution) and stained with SYBR Green I for 15 min. at 80°C. Acquisition was run for 2 min 256 at a speed of 1 μ l s⁻¹. Data were stored as FCS 2.0 files and cell counts (cells mL⁻¹) were 257 258 calculated using the CYTOWIN 4.3 software.

261 Statistical Analysis

262 A one-way ANOVA was used to test for differences in the total amount of DOC released 263 among different types of wrack, with wrack type as fixed factor. A two-way ANOVA was 264 then used to test for differences among wrack types and DOC fraction on the total amount of 265 DOC released, with wrack type and DOC fraction treated as fixed factors. A one-way 266 ANOVA was used to test for significant effects of wrack age on the total DOC released, with 267 age as fixed factor. A two-way ANOVA was then used to test for effects of wrack age and 268 form of DOC on the amount of DOC released, with age and DOC fraction as fixed factors. A 269 repeated measures ANOVA was used to test for significant effects of scraping on the release 270 of DOC from leaves over time, with scraping a fixed factor.

271

The assumption of homogeneity of variances was tested using Cochran's test. When variances were heterogeneous, data were Ln- transformed, or arcsin-transformed for proportions and percentage values. Where significant main effects were detected, post-hoc comparisons (Tukey's) were conducted to determine the sources of significant variation.

276

277

278 **RESULTS**

279 Amount and composition of DOC

280 The net DOC leaching from wrack over 16 hours differed significantly among wrack types,

with fresh algae (*Laurencia sp.*) leaching about four times the DOC released by fresh *P*.

sinuosa leaves, six times that released by A. antarctica leaves, more than 11 times that

released by A. antarctica stems and more than 90 times that released by the fine fraction of

- 284 natural wrack accumulations (Table 1). The recovery of DOC after fractionation into
- 285 hydrophobic, transphilic and hydrophilic fractions was high, ranging from 78-94% (Table 1).

286 For all wrack types, the hydrophilic fraction dominated the total DOC (37-68%), followed by 287 the hydrophobic fraction (17-31%) and the transphilic fraction (4-11%). Nonetheless, there 288 were subtle, but significant, differences in the percentage contribution that hydrophobic and 289 hydrophilic components made to the total DOC, but not the transphilic component (Table 1), 290 reflected in a significant interaction between wrack type and DOC fraction (2-way ANOVA 291 Wrack Type x DOC fraction d.f.=8,59; p < 0.001). The proportion of DOC present as 292 hydrophilic (and presumably the most bio-available) DOC was highest in fresh P. sinuosa 293 leaves (68%) followed by Laurencia and Amphibolis tissues (53-61%) and least in the fine 294 fraction of beach-cast wrack (37%).

295

296 Influence of aging of P. sinuosa wrack on DOC leaching

297 The net DOC leaching from *Posidonia sinuosa* leaves declined with increasing age of the 298 wrack (Table 2). Fresh and one week old leaves released similar amounts of DOC (>1400 mg kg⁻¹) over 16 h, at least 10 times the amount released after 2 weeks of aging and 20 times that 299 300 released after 4 weeks. The composition of the leachate released by leaves of different ages 301 varied subtly and not systematically (Table 2), with a significant interactive effect of wrack 302 age and DOC fraction (2-way ANOVA, d.f. = 6,35, p<0.001; and Table 2 for post-hoc 303 pairwise comparisons). However, in all cases the leachate was dominated by the hydrophilic 304 fraction (48-67%), followed by the hydrophobic (16-27%) and the transphilic (11-17%) 305 fractions.

306

307 DOC release during the first 14 days

308 The rate of net DOC released from fresh *P. sinuosa* leaves during the first 14 days was

309 affected by scraping (Fig 2), with a significant Time x Scraping interaction (p<0.05). For

310 scraped leaves, the leaching rate (A_t) was described by a single-stage exponential decay with

311 a half-life of 1.8 days: $A_t = 752 e^{(-0.317t)}$. For unscraped leaves, the net release rate of DOC

was describe by a two-stage model, with an increasing rate of DOC release for the first 5 days, after which the leaching rate was describe by a exponential decay with a half-life of 1.65 days: $A_t = 1610 e^{(-0385t)}$, which approached the decay curve for the scraped leaves.

315

316 Despite the differences in initial net DOC release rates, the total mass of DOC released 317 (M_{DOC}) from scraped and unscraped leaves was similar over the initial high release period 318 (first 5 days: 1650 ± 122 vs. 1600 ± 77.0 mg kg⁻¹) and then the full 14 days (1920 ± 131 vs. 319 1740 ± 80.0 mg kg⁻¹) of the experiment, indicating that the effect of scraping the leaves was 320 minimal in terms of quantity of DOC leached.

321

The accumulated mass released over the 14 days of incubation approached 2000 mg (Fig 3), with the rate of release dramatically slowing by day 14. Assuming that the mechanism of DOC release remained constant over time, the curve fit to the full dataset (scraped and unscraped leaves, $M_{DOC} = 716 + 470$ Ln(t)) predicts that a further 1100 days (3 years) would be required to release the next 2000 mg.

327

328 Composition of wrack on beaches

329 The wrack accumulating on Geographe Bay beaches was typically dominated by old material 330 (Fig 4). In May, the period just prior to the first autumn - winter storms, the wrack on the sand 331 surface and that within the underlying beach sediments was dominated by old material 332 (generally > 90%). During the winter storm period (June - September) the proportion of fresh 333 wrack increased in both zones, reaching 25-30% at the surface of accumulations but was 334 always less than 10% in the sediment layer below accumulations. By October (spring), the 335 proportion of fresh wrack had declined in all accumulations, approaching 5% in the surface 336 layer and negligible in the sediment layer.

338 **Bioavailability of DOC leachates**

339 For leachates from all types of fresh wrack material, there were significant exponential 340 increases in bacterial abundance over time following inoculation (Fig 5; Table 3; in all cases 341 p<0.001. The Fine Fraction from beach cast wrack also showed a significant increase in 342 bacterial abundance over time (p < 0.05), though the rate of increase was much smaller. In 343 leachate from one month old *Posidonia* leaves, there was no significant increase in bacterial 344 abundance over time. In all cases, when the number of bacteria in the blank incubations was 345 plotted against time the slope was not significantly different to zero, indicating little or no 346 bacterial growth, except for the Fresh Posidonia and Laurencia leachate incubations, where 347 there was a significant exponential decay in bacterial abundance. 348 349 The age of *Posidonia* wrack affected the ability of the leachate to support bacterial growth. 350 For leachate from fresh *Posidonia* leaf material, the linear increase in bacterial abundance over a 24 h period had an average slope of 2.04×10^5 cells h⁻¹. For the leachate from 4 weeks 351 352 old Posidonia leaves, bacterial abundance increased for the first 18 hours, though the slope over this period was less than half that in the leachate from fresh leaves (9.20 x 10^4 cells h⁻¹), 353 354 and declined thereafter.

355

356

357

358 **DISCUSSION**

359 Effect of wrack type on DOC release

360 Total net DOC released varied among types of wrack (algae> Posidonia leaves > Amphibolis

361 leaves > *Amphibolis* stems > fine fraction). This is consistent with studies that found release

362 rates were higher from algae than seagrasses (e.g. Brylinsky, 1977). Algae have less structural

363 carbon, and therefore more storage carbon, per unit biomass, which would account for this

364 difference. Within the different types of seagrass wrack there were also differences in release 365 rates of DOC. Fresh *Posidonia* leaves had the largest release rate of DOC, followed by A. antarctica leaves and then stems. The stems of Amphibolis are vertical rhizomes and serve as 366 367 a major storage organ for carbohydrates. In the closely related species Amphibolis griffithii, soluble sugars account for 15-20% DW, and starches account for 2-3% DW of the rhizome 368 369 (Lavery et al. 2009). On this basis, we might expect higher fluxes of soluble carbohydrate 370 compounds from the stems than the leaves. However, the lower net release rate for stems may 371 reflect higher levels of other soluble compounds in the leaves, especially proteins associated 372 with photosynthesis, and stronger barriers to diffusion, since stems are highly lignified and 373 contain large amounts of vascular tissue. This may also explain the differences among leaves, 374 since Posidonia sinuosa leaves and those of A. griffithii typically have similar levels of 375 soluble sugars and starches (*P. sinuosa* = 2-4% DW soluble sugars and 5-10% starches; 376 Collier, et al. 2009); A. griffithii = 5-15% soluble sugars and 2-3% starch; Lavery et al. 2009). 377

The relatively low release rates from aged *Posidonia* leaves (4 weeks old) and the fine
particulate fraction of wrack (at least 2 months old) reflects the effect of aging on DOC
release and the significant loss of DOC which occurs in the first few days of leaching.
However, despite the low rate of bacterial growth on leachate from the 4 weeks old *Posidonia*leaves, the initial growth over 18 hours confirms that the DOC leachate was bioavailable.
The decline in bacterial biomass after 18 hours indicates that it was more likely a function of
the mass of DOC in the leachate than the composition that affected bacterial growth.

385

The composition of the DOC leached from different species of wrack was similar. This may partly reflect the level of resolution in our chemical characterisation of the leachates. The high % recovery of DOC following fractionation gives confidence that we have not underestimated a significant portion of the DOC. Maie et al. (2006) found differences in concentrations of sugars and phenols in leachate from a range of aquatic plants they studied.
However, the plants they studied covered a wide phylogenetic range, from algal periphyton to
freshwater macrophytes, mangroves and seagrasses. In comparison, our wrack was all
derived from seagrasses, with the exception of the alga *Laurencia*.

394

395 The similarity in leachate quality from all wrack types, including that of aged wrack, indicates 396 that the quality of DOC that seagrass wrack contributes to recipient habitats is likely to be 397 similar, irrespective of the type or age of the wrack, though the mass contributed will decline 398 rapidly with age. It was surprising that the hydrophilic portion (which contains sugars, amino 399 acids, small molecular weight fatty acids and other compounds likely to be more labile) 400 continued to form a significant proportion (more than 50%) of the DOC leached from aged P. 401 sinuosa and the fine fraction. It is not clear whether, in the older wrack, the low molecular 402 weight component is derived directly from the wrack, or is contained in exudates from 403 bacteria growing on the wrack or in suspension. In any case, this makes little difference in 404 terms of the potential benefit of the input to recipient ecosystems. If it is derived from 405 exudates of bacteria growing on the wrack, then it is possible that this input of readily 406 bioavailable DOC could persist for months, though at a very slow rate.

407

408 Mass & Timescale of DOC Release

The release rate of DOC declined rapidly with age of *Posidonia* wrack. About 50% (2000 mg kg⁻¹ FW wrack) was released in the first 14 days. Assuming that the mechanism of DOC release remains constant over the decay period of the wrack, it would take in the order of thousands of days to release the next 2000 mg kg⁻¹ FW wrack. This assumption may not be the case but, nonetheless, it is clear that the rate of DOC release will fall dramatically after the first days. The *Posidonia* leaves used in our studies contained about 33% carbon DW or 11% FW (using a DW;FW ratio of 0.34; unpublished data) so the total mass of carbon released

over 14 days was about 1.8% of the leaf carbon, and if we project out to 2000 days, 3.6% of 416 417 the leaf carbon. These values are similar to the total fixed carbon lost through DOC excretion 418 reported for Posidonia oceanica (Velimirov 1986) but much less than the 48% estimated by 419 Kirkman & Reid (1979) for *Posidonia australis*, which has very similar leaf structure to P. 420 sinuosa (Cambridge & Kuo 1982). Kirkman & Reids' (1979) estimate was likely to have 421 severely over-estimated the leaching of DOC from leaves. They used leaves with necrotic 422 tissue and with a full complement of epiphytes which would have contributed to DOC 423 leakage, and they measured the rates over two hours, which are likely to produce much higher 424 estimates of loss that would occur for aged wrack tissue, as shown by our results.

425

426 Temporal variation in DOC release rates have been reported for other seagrasses. Velimirov 427 (1986) found that the young, green portions of *Posidonia oceanica* leaves released negligible 428 amounts of DOC but high loss was observed for older, brown leaves, while the rate of DOC 429 leaching from Thalassia testudinum leaves declined exponentially with age, with 84% of the 430 DOC leached in the first two weeks (Maie et al. 2006). In our case, the initial rate of DOC 431 release was enhanced by scraping leaves to remove epiphytes. This could be due to the 432 removal of epiphytes or damage to the leaf surface. Epiphytic organisms reduce the release of 433 DOC from seagrass leaves to surrounding water (Velimirov 1986, Wetzel & Penhale 1979), 434 presumably through assimilation of the DOC. Scraping is also likely to disrupt the tough 435 cuticle and thick epidermal leaves of *P. sinuosa* (Cambridge & Kuo, 1982), enhancing 436 diffusive losses of cellular DOC. However, this effect was limited mainly to day 1, with the 437 total mass of DOC released over the first 5 days (the period of highest initial release rates) 438 similar in both types of leaf, providing confidence that scraping had little effect on the total 439 amount of DOC released beyond the first day. Despite the potential for scraping to introduce 440 an experimental artefact, it may be representative of the condition of naturally shed leaves 441 which will have damage to the leaf surface, particularly the necrotic upper part of the leaves,

through the action of grazers and abrasion by sediments as they are transported in bedload andsuspended transport.

444

The bacterial growth in the assays demonstrates the bioavailability of the leachate DOC released from fresh seagrass and algae, with exponential increases in abundance indicating bacterial growth. Bacteria grown on seagrass DOC leachate can rapidly be converted into bacterial aggregates that are consumed by ciliates and flagellates at a much faster rate than the residual particulate organic carbon (Robertson et al. 1982). We did not enumerate the bacteria on the surface of the wrack, but these typically are much more abundant that bacteria in suspension.

452

453 **Relative contribution of wrack to DOC in Geographe Bay**

454 While it was beyond the scope of this study to produce a full DOC budget for Geographe Bay, 455 sufficient information is available to compare the potential contribution of wrack with some 456 other sources of DOC (Table 1) to the study area. Oldham et al. (2010) estimated a total 457 annual wrack production of 16,900 t DW of Posidonia leaf wrack and 15, 700 t of Amphibolis wrack from this region (i.e. an area of 60 km^2 with an average depth of 5 m - the area of 458 459 seagrass coverage to the 10 m depth contour). At the initial DOC release rates recorded in our 460 study, this mass of wrack would contribute 191 kg of DOC to the study region in one day. Actively growing phytoplanktton can leak between 0.0005 - 0.055 pmol DOC cell⁻¹ d⁻¹ 461 462 (Biddanda & Benner 1997). Coastal waters to the north of Geographe Bay typically have about 91,000 cells L^{-1} of phytoplankton (Hanson et al. 2006). Assuming a similar cell count 463 464 in Geographe Bay, phytoplankton in the study area would, at most, contribute 0.19 - 21 kg 465 DOC per day, between 1 and 3 orders of magnitude less than seagrass wrack. 466

467 In contrast, living seagrass represents a very large DOC source relative to wrack.

468 Velimirov (1986) compared the DOC release from healthy, living and scenescent seagrass 469 leaves; healthy leaves released 0.2% of that released by scenescent leaves. However, the 470 biomass of living meadow is much higher than that of wrack. Assuming a release rate comparable to that given by Velimirov for *Posidonia oceanica* (0.006 mg DOC $g^{-1} h^{-1}$) and 471 472 using McMahon et al.'s (1997) reported biomass for Posidonia sinuosa in Geographe Bay (115-470 g dw m^{-2}), live seagrass in the study area would provide 990 – 4000 kg DOC per 473 474 day to the study area, 10-40 times that of wrack. Of course, this is a fixed source of DOC, 475 compared with the more mobile nature of wrack, which permits inter-habitat connectivity. 476

477 Surface beach sands can be a significant source of DOC to the water column in high-energy environments, with a net flux of 4-22 mmol DOC m⁻² d⁻¹ (Heymans & McLachlan 1996, 478 D'Andrea et al. 2002, Avery et al. 2012). Geographe Bay has an approximately 0.5 m diurnal 479 tide range and a mean beach slopes of about 0.06 m m^{-1} (Oldham et al. 2010). Using the 480 lower slope estimate, over the 30 km stretch of beach an area of 2.5 x 10^5 m⁻² of beach face 481 482 would be inundated each day, providing an estimated flux of DOC in the order of 12-66 kg DOC d⁻¹. While significant, this is a smaller source than, and is likely to be most significant 483 484 to the surf zone adjacent to the beach unlike wrack, which can be transported and gradually 485 release DOC over a wider area.

486

487 Implications for Trophic connectivity

The exponential-decay model of DOC release from *P. sinuosa* leaves indicates that there will be significant temporal variation in the release of DOC from leaves shed by plants, with significant implications for trophic connectivity. Wrack is constantly being produced in seagrass meadows, but in the case of *Posidonia* and *Amphibolis* leaves significant water velocities, in excess of 0.15 m s⁻¹, are required to suspend wrack, allowing it to be transported away from the meadow (Oldham et al. in review). Typically, this results in wrack 494 accumulating in offshore meadows during quiescent periods (spring through to early autumn 495 in our study site) and leaves being transported to beaches and other habitats during storm 496 events (McMahon et al., in review), typically in autumn and winter. Leaves shed in spring 497 and summer may, therefore, slowly degrade within the meadow for several weeks or months, 498 with the majority of DOC released within the meadow itself. Adjacent habitats will only 499 receive wrack in a high DOC-leaching phase under two scenarios: 1) during unusual storm 500 events which are sufficiently energetic to dislodge and transport living material and when 501 fresh wrack may constitute a significant portion of the total; and 2) during normal autumn-502 winter storms, when it will only constitute a small proportion of the total wrack exported (i.e. 503 that shed in the previous two weeks).

504

505 The above suggests that in our system and outside of storm events, when the timescale of 506 leaching is typically much faster than the timescale of transport, wrack may be of limited 507 value in supporting trophic subsidies. However, we have noted relatively high DOC 508 concentrations in porewaters beneath wrack accumulations. While fresh wrack was never the 509 dominant component of wrack accumulations, it frequently accounted for 25-30% of the mass 510 during winter (when storm conditions dominate). During this time, beach accumulations can persist for several weeks reaching biomasses of 4 kg m⁻² under natural conditions but as much 511 as 19 kg m⁻² in areas affected by coastal structures (McMahon in prep). Assuming 4 kg m⁻² of 512 wrack with 30% fresh material approximately 4.1 g of DOC m^{-2} would be released over two 513 514 weeks, which our data shows is capable of supporting bacterial growth. Thus, while seagrass 515 wrack may have relatively little value as a source of trophic connectivity during periods of 516 quiescent hydrodynamics, it may still be important during periods of higher energy and faster 517 transport, leading to the formation of biogeochemical hot moments (sensu McClain et al. 518 2003). Furthermore, under quiescent hydrodynamics, seagrass detritus may contribute, even if 519 at slow DOC release rates, to the sedimentary organic carbon pool of offshore habitats

including oligotrophic unvegetated habitats, as suggested by Ziegler & Benner (1999). This
could also apply to beaches and other recipient habitats if the wrack is buried and therefore
can persist in these habitats for sufficiently long periods to allow an accumulation of DOC.
This demonstrates a complex interaction of timescales of transport (or residence times) and
timescales of leaching which must be undertaken into account when considering the potential
for trophic subsidies.

526

527 Conclusions

528 We conclude that *Posidonia sinuosa* and *Amphibolis antarctica* seagrass wrack leaches 529 bioavailable DOC. We also conclude that, for P. sinuosa, there is an initial rapid release of 530 DOC within the first days-weeks followed by an extended period of low release rates. As 531 similar theorem of DOC release have been shown for other seagrasses such as *P. oceanica* 532 (Velimirov 1986) and Thalassia testudinum (Maie et al. 2006), it is likely that wrack from 533 many species of seagrass will demonstrate similar patterns of DOC release. Despite 534 differences in the rate of DOC release from different types and ages of wrack, the 535 composition was similar and it was bioavailable even when released from old wrack, though 536 the amount released would limit bacterial growth. Given the known consumption of bacterial 537 aggregates by higher levels of the foodweb, the leaching of DOC is one means of recycling 538 the nutrients in seagrass detritus. The interaction of the timescales of transport and the 539 timescale of leaching will be critical in determining the value of wrack as a vector for trophic subsidies. When fresh wrack is released during periods of rapid hydrodynamic transport, it 540 541 has the potential to release most of its DOC into recipient habitats. However, during 542 quiescent periods, the rapid leaching will result in most of the DOC being recycled within the 543 seagrass meadow. Further work is required to determine the importance of bacterial growth 544 on the surface of wrack and in suspension as a sink for seagrass DOC, and the efficiency of its 545 subsequent incorporation into the food web of recipient ecosystems.

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553 **Reference**

- Avery GB, Kieber RJ, Taylor KJ, Dixon JL (2012) Dissolved organic carbon release from
 surface sand of a high energy beach along the Southeastern Coast of North Carolina,
 USA. Marine Chemistry 132-133:23–27
- Barrón C, Duarte C (2009) Dissolved organic matter release in a Posidonia oceanica meadow.
 Marine Ecology Progress Series 374:75–84
- Benner R, Peele ER, Hodson RE (1986) Microbial utilization of dissolved organic matter
 from leaves of the red mangrove, Rhizophora mangle, in the fresh creek estuary,
 Bahamas. Estuarine Coastal and Shelf Science 23:607–619
- Biddanda B, Benner R (1997) Carbon, nitrogen, and carbohydrate fluxes during the
 production of particulate and dissolved organic matter by marine phytoplankton.
 Limnology and Oceanography 42:506–518
- Brylinsky M (1977) Release of dissolved organic matter by some marine macrophytes.
 Marine Biology 39:213–220
- 567 Cambridge M, Kuo J (1982) Morphology, anatomy and histochemistry of the Australian
 568 seagrasses of the genus Posidonia Konig (Posidoniaceae) III . Posidonia sinuosa
 569 Cambridge & Kuo. Aquatic Botany 14:1–14
- 570 Cebrian J, Duarte CM (2001) Detrital stocks and dynamics of the seagrass Posidonia oceanica
 571 (L.) Delile in the Spanish Mediterranean. Aquatic Botany 70:295–309
- 572 Chow CWK, Fabris R, Drikas M (2004) A rapid fractionation technique to characterise
 573 natural organic matter for the optimisation of water treatment processes. The Journal of
 574 Water Supply 53:85–92
- 575 Cleveland CC, Neff JC, Townsend AR, Hood E (2004) Composition, dynamics and fate of
 576 leached dissolved organic matter in terrestrial ecosystems: Results from a decomposition
 577 experiment. Ecosystems 7:275–285
- 578 Cleveland CC, Nemergut DR, Schmidt SK, Townsend AR (2007) Increases in soil respiration
 579 following labile carbon additions linked to rapid shifts in soil microbial community
 580 composition. Biogeochemistry 82:229–240
- Collier CJ, Lavery PS, Ralph PJ, Masini RJ (2009) Shade-induced response and recovery of
 the seagrass Posidonia sinuosa. Journal of Experimental Marine Biology and Ecology
 370:89–103
- 584 Crawley KR, Hyndes GA, Vanderklift MA, Revill AT, Nichols PD (2009) Allochthonous
 585 brown algae are the primary food source for consumers in a temperate, coastal
 586 environment. Marine Ecology Progress Series 376:33–44
- 587 D'Andrea AF, Aller RC, Lopez GR (2002) Organic matter flux and reactivity on a South
 588 Carolina sandflat: The impacts of porewater advection and macrobiological structures.
 589 Limnology & Oceanography 47:1056–1070

- Doropoulos C, Hyndes GA, Lavery PS, Tuya F (2009) Dietary preferences of two seagrass
 inhabiting gastropods : Allochthonous vs autochthonous resources. Estuarine, Coastal
 and Shelf Science 83:13–18
- 593 Ducker SC, Foord NJ, Knox RB (1977) Biology of Australian Seagrasses: the Genus
 594 Amphibolis C. Agardh (Cymodoceaceae. Australian Journal of Botany:67–95
- 595 Green EP, Short FT (2003) World Atlas of Seagrasses. UNEP-WCMC, Cambridge
- Hanson C, Clementson L, Thompson P (2006) Phytoplankton community structure. In:
 Keesing JK, Heine JN, Babcock RC, Craig PD, Koslow JA (eds) Strategic research Fund
 for the Marine Environment Final Report. Volume 2: The SRFME core projects.
 Strategic research Fund for the Marine Environment, CSIRO, p 71–80
- Heymans JJ, McLachlan A (1996) Carbon Budget and Network Analysis of a High-energy
 Beach/Surf-zone Ecosystem. Estuarine Coastal and Shelf Science 43:485–505
- Hyndes GA, Lavery PS (2005) Does transported seagrass provide an important trophic link in
 unvegetated, nearshore areas? Estuarine Coastal and Shelf Science 63:633–643
- Ince R, Hyndes GA, Lavery PS, Vanderklift MA (2007) Marine macrophytes directly
 enhance abundances of sandy beach fauna through provision of food and habitat.
 Estuarine Coastal and Shelf Science 74:77–86
- Kaldy JE, Eldridge PM, Cifuentes LA, Jones WB (2006) Utilization of DOC from seagrass
 rhizomes by sediment bacteria : 13C-tracer experiments and modeling. Marine Ecology
 Progress Series 317:41–55
- Kirkman H, Kendrick GA (1997) Ecological significance and commercial harvesting of
 drifting and beach-cast macro-algae and seagrasses in Australia : a revie. Journal of
 Applied Phycology 9:311–326
- Kirkman H, Reid D (1979) A study of the role of the seagrass Posidonia australis in the
 carbon budget of an estuary. Aquatic Botany 7:173–183
- Klumpp DW, Vandervalk A (1984) Nutritional quality of seagrasses (Posidonia australis and Heterozostera tasmanica) - comparison between species and stages of decomposition.
 Marine Biology Letters:67–83
- Lavery PS, Mcmahon K, Mulligan M, Tennyson A (2009) Interactive effects of timing ,
 intensity and duration of experimental shading on Amphibolis griffithii. Marine Ecology
 394:21–33
- Lavery PS, Vanderklift MA (2002) A comparison of spatial and temporal patterns in
 epiphytic macroalgal assemblages of the seagrasses Amphibolis griffithii and Posidonia
 coriacea. Marine 236:99–112
- Maie N, Jaffé R, Miyoshi T, Childers DL (2006) Quantitative and qualitative aspects of
 dissolved organic carbon leached from senescent plants in an oligotrophic wetland.
 Biogeochemistry 78:285–314

- Marie D, Partensky F, Jacquet S (1997) Enumeration and cell cycle analysis of natural populations of marine picoplankton by flow cytometry using the nucleic acid stain SYBR Green. Applied and Environmental Microbiology 63:186–193
 Mateo MA (2010) Beach-Cast Cymodocea nodosa Along the Shore of a Semienclosed Bay: Sampling and Elements to Assess Its Ecological Implications. Journal of Coastal Research 262:283–291
- Mateo MA, Cebrian J, Dunton K, Mutchler T (2006) Carbon Flux in Seagrass Ecosystems. In:
 Larkum A, Orth R, Duarte C (eds) Seagrasses: Biology, Ecology and Conservation.
 Springer-Verlag, Netherlands, p 159–192
- McClain ME, Boyer EW, Dent CL, Gergel SE, Grimm NB, Groffman PM, Hart SC, Harvey
 JW, Johnston C a., Mayorga E, McDowell WH, Pinay G (2003) Biogeochemical hot
 spots and hot moments at the interface of terrestrial and aquatic ecosystems. Ecosystems
 639 6:301–312
- McMahon K, Young E, Montgomery S, Cosgrove J, Wilshaw J, Walker DI (1997) Status of a
 shallow seagrass system, Geographe Bay, south-western Australia. Journal of the Royal
 Society of Western Australia 80:255–262
- Moore TN, Fairweather PG (2006) Decay of multiple species of seagrass detritus is
 dominated by species identity, with an important influence of mixing litters. Oikos
 114:329–337
- Moriarty DJW, Pollard PC (1982) Diel variation of bacterial productivity in seagrass (Zostera capricorni) beds measured by rate of thymidine incorporation into DNA. Marine Biology 173:165–173
- Oldham CE, Lavery PS, McMahon K, Pattiratchi C, Chiffings TW (2010) Seagrass wrack
 dynamics in Geographe Bay, Western Australia. Nedlands, Australia
- Prado P, Collier C, Lavery PS (2008) 13C and 15N translocation within and among shoots in
 two Posidonia species from Western Australia. Marine Ecology Progress Series 361:69–
 82
- Robertson AI, Lenanton RCJ (1984) Fish community structure and food chain dynamics in
 the surfzone of sandy beaches: the role of detached macrophyte detritus. Journal of
 Experimental Marine Biology and Ecology 84:265–283
- Robertson M, Mills A, Zieman J (1982) Microbial synthesis of detritus-like particulates from
 dissolved organic carbon released by tropical seagrasses. Marine Ecology Progress
 Series 7:279–285
- Smit AJ, Brearley A, Hyndes GA, Lavery PS, Walker DI (2005) Carbon and nitrogen stable
 isotope analysis of an Amphibolis griffithii seagrass bed. Estuarine Coastal and Shelf
 Science 65:545–556
- 663 Smit AJ, Brearley A, Hyndes G a., Lavery PS, Walker DI (2006) δ15N and δ13C analysis of a
 664 Posidonia sinuosa seagrass bed. Aquatic Botany 84:277–282

- Thurman EM, Malcolm RM (1981) Preparative isolation of aquatic humic substances.
 Environmental Science and Technology 15:463–466
- Velimirov B (1986) DOC dynamics in a Mediterranean seagrass system. Marine Ecology
 Progress Series 28:21–41
- Wetzel RL, Penhale PA (1979) Transport of carbon and excretion of dissolved organic carbon
 by leaves and roots/rhizomes in seagrasses and their epiphytes. Aquatic Botany 6:149–
 158
- 672 Ziegler S, Benner R (1999) Dissolved organic carbon cycling in a subtropical seagrass 673 dominated lagoon. Marine Ecology Progress Series 180:149–160
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- 675

676	Table 1 Dissolved organic carbon composition in leachates derived from different wrack
677	material (means \pm sd). Within each class of DOC (totals, hydrophobic, transphilic and
678	hydrophilic), shared subscript letters indicate no significant differences among wrack types
679	($\alpha = 0.05$). <i>Posidonia</i> = <i>P. sinuosa</i> , <i>Amphibolis</i> = <i>A. antarctica</i> ; Fine fraction = $0.1 - 1.0$
680	mm size class of natural wrack accumulations.

Wr

Wrack Type	DOC released in 16 h (mg kg ⁻¹ FW wrack; n=4 in all cases)				DOC Recovery
	TOTAL	Hydrophobic	Transphilic	Hydrophilic	(/*)
Fresh Laurencia	6 749 ±278 _a	$1\ 998 \pm 64\ 30\%_a$	675 ± 95 10% _a	3 554 ± 213 53% _a	93%
Fresh Posidonia	$1~724~{\pm}76_{\rm b}$	298 ± 14	138 ± 6	1173 ± 58	93%
leaves		17% _b	8% _b	68% _b	
Fresh Amphibolis	$1\ 102\ \pm 24\ _{\rm c}$	284 ± 26	48 ± 4	676 ± 21	91%
leaves		26% _b	4% _{bc}	61% _c	
Fresh Amphibolis	588 ±31 _d	180 ± 13	58 ± 4	312 ± 29	94%
stems		31% _c	10% _{bc}	53% _d	
Fine fraction	74 ±1 e	$\begin{array}{c} 22\pm3\\ 30~\%_{d} \end{array}$	8 ± 5 11 % c	27 ± 1 37% _e	78%

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687	
688	Table 2 Dissolved organic carbon composition of leachate from P. sinuosa leaves of different ages.
689	ANOVA revealed a significant interaction of Age and DOC fraction (p <0.001). Shared subscript
690	letters indicate no significant difference in mass of DOC released (Tukey's test; p>0.05) among
691	treatments within each class of DOC.
692	

Age		DOC release over 16 h (mg kg ⁻¹ FW wrack)			
	TOTAL	Hydrophobic	Transphilic	Hydrophilic	
Fresh1	$1\ 419\pm93\ _a$	223 ± 18 16% _a	$\frac{181 \pm 17}{13\%_a}$	$\begin{array}{c} 855\pm45\\ 60\%_a \end{array}$	89%
1 week old	$1\ 627\pm192\ _a$	417 ± 13 26% _{bc}	277 ± 32 17% _a	783 ± 120 48% _b	91%
2 weeks old	$133\pm17~_{b}$	36 ± 4 27% _b	$\frac{19 \pm 4}{14\%}_{a}$	67 ± 7 50% _b	91%
4 weeks old	67 ± 2 c	12 ± 1 18% _c	7.2 ± 0.2 $11\%_a$	45 ± 1 67% _a	96%

- **Table 3** Exponential curve fits describing the change in abundance of bacterial cell abundance over69924 hr in leachates from different types of wrack following bacterial inoculation. * = significant at700 $p \ge 0.05$; ** $p \le 0.01$. In all cases, x = time in hours.

Wrack Type	Correlation	r	р
Posidonia leaf (Fresh)	$4.54 \text{ x } 10^5 \text{ e}^{0.109 \text{ x}}$	0.95	***
Laurencia (Fresh)	$1.65 \ge 10^6 e^{0.101 x}$	0.90	***
Amphibolis leaf (Fresh)	$1.26 \text{ x } 10^6 \text{ e}^{0.076 \text{ x}}$	0.89	***
Amphibolis stem (Fresh)	$1.23 \text{ x } 10^6 \text{ e}^{0.086 \text{ x}}$	0.97	***
Posidonia (Old)	$3.92 \text{ x } 10^4 \text{ e}^{0.129 \text{ x}}$	0.34	nsd
Fine Fraction	$7.64 \text{ x } 10^4 \text{ e}^{0.084 \text{ x}}$	0.51	*
Blank (Fresh Posidonia	$9.58 \ge 10^4 e^{-0.011x}$	-	***
& Laurencia)		0.83	



Figure 1 Map of Geographe Bay, Western Australia, showing the location of the three sites used to

- sample beach wrack composition.



- 715 Figure 2 Net DOC release rates from scraped and unscraped *P. sinuosa* leaves during 14-
- 716 day incubations. The regression for unscraped leaves is for days 5-14.



Figure 3 Cumulative net mass of DOC released over time







Figure 4 The composition (new versus old) of wrack on beaches of Geographe Bay fromMay-October 2008.



Figure 5 Abundance of heterotrophic bacterial cells following addition of an inoculum to DOC leachates from different wrack types.