

文章编号: 1001-6880(2010)04-0544-04

分离自木霉 *Trichoderma* sp. PT2 的一个新倍半萜糖苷

杜希萍^{1,2}, 李瑶瑶¹, 鲁春华^{1*}, 郑忠辉¹, 沈月毛¹¹厦门大学生命科学学院 细胞生物学与肿瘤细胞工程教育部重点实验室 福建省药物工程实验室, 厦门 361005²集美大学生物工程学院, 集美 361021

摘要: 从海藻真菌木霉 *Trichoderma* sp. PT2 菌体提取物中分离到一个新倍半萜糖苷类化合物 Trichodem oside (1)。并通过 1D-, 2D-NMR 波谱数据鉴定了化合物的结构, 并应用 MTT 法对化合物 1 的细胞毒活性进行了研究。

关键词: 倍半萜糖苷; Trichodem oside; 木霉; 结构解析; 生物活性

中图分类号: R93

文献标识码: A

A Novel Sesquiterpene Glucoside from *Trichoderma* sp. PT2

DU Xiping^{1,2}, LI Yaoyao¹, LU Chunhua^{1,*}, ZHENG Zhonghui¹, SHEN Yue-mao¹¹Key Laboratory of the Ministry of Education for Cell Biology and Tumor Cell Engineering; Fujian Engineering

Laboratory for Pharmaceuticals, School of Life Science, Xiamen University, Xiamen, 361005, China

²Bioengineering College of Jimei University, Jimei 361021, China

Abstract A novel sesquiterpene glucoside, trichodem oside (1), was isolated from the mycelia extract of a filamentous fungus *Trichoderma* sp. PT2. The structure was elucidated by spectroscopic analyses including 1D-, 2D-NMR data and mass spectrometric analyses. Compound 1 displayed weak growth inhibition against human HeLa cells.

Key words sesquiterpene glucoside; Trichodem oside; *Trichoderma* sp.; structure determination; cytotoxicity

Introduction

The fungal genus *Trichoderma* is a widespread saprophyte that occurs almost ubiquitously. Because of the useful secondary metabolites produced by this genus, many strains have received considerable attention as biocontrol agents^[1,2]. Some secondary metabolites including N-acylated peptides, peptibols^[3,4], koniginins A-G^[5-8], octaketide-derived compounds, two butenolide compounds^[9,10], and demethylsobicillin and oxosobicillinol^[11] were reported as antibiotics with various activities. During the course of examining fungi for biologically active natural products, the strain PT2 from *Blidingia minima* Kylin, collected at Wuyu Island, Zhangzhou of Fujian Province, was isolated. The ethyl

acetate extract of the fermentation showed strong antimicrobial activities against KB and Raji cell lines^[12]. Cyclopeptides were the active constituent^[13]. Our further research on the chemical components of *Trichoderma* sp. PT2 yielded one new compound. Here the isolation, structure elucidation and bioactivity assay of this compound was described.

Results and Discussion

Fermentation (10 L) was carried out at 28 °C for 20 d without agitation. After filtration, the mycelia were extracted exhaustively with acetone. Acetone was evaporated *in vacuo* and the crude extract was partitioned between methanol and petroleum ether. The methanol solution was collected and evaporated to dryness *in vacuo* to afford 17 g extract. The extract was purified over RP-18, Sephadex LH-20 and silica gel columns to afford a novel sesquiterpene glucoside.

Trichodem oside (1) was obtained as a white powder with $[\alpha]_D^{20} + 5.76$ (c 0.26, MeOH). The molecular formula C₂₃H₃₉NO₈ was determined according to the

Received December 28, 2009; Accepted April 21, 2010

Foundation Item: This work was financially supported by China Ocean Resource R&D Association (Grant DYXM-115-02-2-13) and the National High Technology Research and Development Program of China (863 No. 2006AA09Z410).

* Corresponding author. Tel: 86-532-88963253; E-mail: ahua096@xmu.edu.cn

quasi molecular ion peaks at m/z 480.5 $[M + Na]^+$ and 937.3 $[2M + Na]^+$ in ESI-MS, and NMR data. The presence of a α -glycosyl N-acyl moiety was revealed by the 1H NMR signals at δ 4.97 (d, 3.5 H, H-1'), 3.88 (dd, 10.2, 3.1, H-2'), 3.76 (dd, 13.4, 4.0, H-3'), 3.64 (t, 9.4, H-4'), 3.45 (t, 8.9, H-5'), 3.78 (d, 5.0, H-6'), 1.98 (s, 3H, H-8'), and the ^{13}C NMR signals at δ 98.8 (C-1'), 54.6 (C-2'), 72.0 (C-3'), 72.8 (C-4'), 71.0 (C-5'), 61.7 (C-6'), 172.9 (C-7'), 22.5 (C-8')^[14]. The HMBC correlations of H-1'/C-3', H-2'/C-1', C-3' and C-7', H-3'/C-4' and C-5', H-6'/C-5', and H-8'/C-7' confirmed this conclusion (Table 1).

Besides the α -glycosyl N-acyl moiety, the ^{13}C NMR spectra of **1** (Table 1) displayed three quaternary carbons, three methylenes (one oxygenated), five methines (two olefinic and two oxygenated), and four methyls indicating the presence of a sesquiterpene moiety. The following HMBC correlations were observed: H-13 (δ 0.97) / C-3, C-4, C-5 and C-14; H-14 (δ 0.70) / C-3, C-4, C-5 and C-13; H-12 (δ 1.68) / C-5, C-6 and C-1. In combination with the 1H , 1H -COSY correlations between H-1/H-2 and H-2/H-3, the fragment b was established (Fig. 1). The fragment c could be also postulated from HMBC experiment. The HMBC correlations from H-8/C-5, C-7, C-9, C-10 and C-15

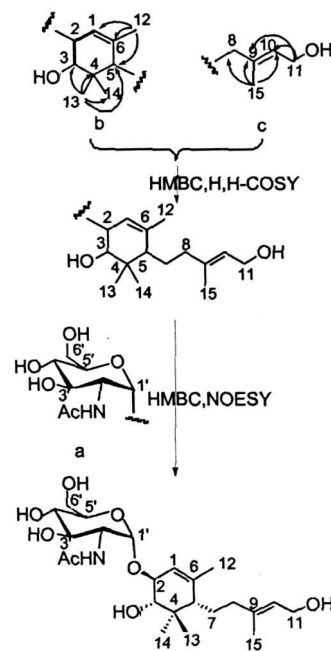


Fig. 1 The fragments a, b and c of compound **1**, and the selected HMBC correlations (H \rightarrow C) and 1H , 1H -COSY correlations

revealed that fragments b and c were joined together. Therefore, the structure of the sesquiterpene moiety was determined. The glycosyl was located at C-2 in view of the HMBC correlation H-1' (δ 4.97) / C-2 (δ 81.1). Thus, the structure of **1** was determined as shown in figure 1. The relative configuration of **1** was determined by NOESY cross signals: H-3/H-5, H-3/H₃-13, H-2/H-14.

Table 1 The NMR data of compound **1** (CDCl₃, ppm, J in Hz)

No	δ_H	δ_C	HMBC	1H , 1H -COSY
1	5.28 (s, 1H)	122.0 (d)	C-3, C-5, C-12	H-2
2	3.88 (dd, 10.2, 3.1, 1H)	81.1 (d)	C-1'	H-3, H-1
3	3.34 (d, 8.6, 1H)	78.7 (d)	C-2, C-4, C-5, C-13, C-14	H-2
4	-	40.0 (s)	-	-
5	1.73 (br s, 1H)	49.7 (d)	-	H-7
6	-	139.4 (s)	-	-
7	1.37 (m, 1H), 1.20 (m, 1H)	25.8 (t)	C-6, C-8	H-8
8	2.18 (m, 1H), 1.95 (m, 1H)	41.1 (t)	C-5, C-7, C-9, C-10, C-15	H-7
9	-	138.9 (s)	-	-
10	5.35 (t, 6.5, 1H)	123.9 (d)	C-8, C-11, C-15	H-11
11	4.09 (d, 6.7, 2H)	58.9 (t)	C-9, C-10	H-12
12	1.68 (s, 3H)	21.9 (q)	C-1, C-5, C-6	-

13	0.97 (s 3H)	24.4 (q)	C-3, C-4 C-5, C-14	-
14	0.70 (s 3H)	14.1 (q)	C-3, C-4 C-5, C-13	-
15	1.63 (s 3H)	16.1 (q)	C-8, C-9 C-10	-
1'	4.97 (d, 3.5 1H)	98.8 (d)	C-3', C-1	H-2'
2'	3.88 (dd 3.1, 10.2 1H)	54.6 (d)	C-1', C-3', C-7'	H-1', H-3'
3'	3.76 (dd 13.4 4.0 1H)	72.0 (d)	C-4', C-5'	H-2'
4'	3.64 (t 9.4 1H)	72.8 (d)	C-2', C-5'	H-5'
5'	3.45 (t 8.9 1H)	71.0 (d)	C-3', C-6'	H-4'
6'	3.78 (d, 5.0 2H)	61.7 (t)	C-5'	H-5'
7'	-	172.9 (s)	-	-
8'	1.98 (s 3H)	22.5 (q)	C-7'	-

Bioassays

The cytotoxic activity against the Hela cell line of compound 1 was measured 72 h post treatment by the MTT method^[15]. Compound 1 (10 µg/mL) displayed weak growth inhibition (12%).

Experimental

General

Precoated TLC plates were from Qingdao Haiyang Chemical Factory, Qingdao P. R. China. For column chromatography (CC), silica gel (200-300 and 80-100 mesh Qingdao), silica gel 60 (Merck), RP-18 (Merck), and Sephadex LH-20 gel (Amersham Biosciences) were used. Optical rotation was measured on a Perkin-Elmer 341 polarimeter with CHCl₃ as solvent. NMR Spectra were recorded on a Bruker DRX-500 spectrometer (δ in ppm rel to Me₄Si in Hz), and MS spectra on a Finnigan LCQ-Advantage and VG Auto-Spec-3000 mass spectrometers.

Fermentation of the Strain

The strain PT2 was identified as *Trichoderma* sp according to its ITS sequence of rDNA (ITS1-5.8S-ITS2). The strain was incubated on slope of 50% seawater potato dextrose agar (PDA) media in a test tube at 28 °C for 7 d to afford seed cultures. Then the seed cultures were washed with sterile water, and the spores were transferred to 3000 mL Erlenmeyer flasks each containing 1000 mL of 50% seawater potato dextrose broth (PDB) for static incubation, room temperature, 20 d.

Extraction and isolation

The flask cultures were filtered and the mycelia were extracted with acetone exhaustively. The acetone solution was collected and evaporated in vacuo to afford a crude extract. Then dissolved in methanol and partitioned between petroleum ether and methanol. The brown oil (17 g) was obtained from the methanol phase.

The extract (17 g) was subjected to MPLC (170 g RP-18) eluted with H₂O, 30%, 50%, 70%, and 100% methanol respectively. Then five fractions (Fr S1-S5) were obtained. Fr S3 (98 mg) was subjected to CC (45 g Sephadex LH-20, MeOH). All fractions were analyzed by TLC (EtOAc/MeOH 1:2), and pooled into 3 portions (Fr S31-S33). Fr S33 (74 mg) was further separated over CC (silica gel CHCl₃/acetone 10:1, 5:1, 1:1, 1:2) to yield 1 (5 mg).

Trichodemoside [N-((2S, 3S, 4R, 5S)-2-((1S, 4S, 6S)-6-hydroxy-4-((E)-5-hydroxy-3-methylpent-3-enyl)-3,5,5-trimethylcyclohex-2-enyloxy)-tetrahydro-4,5-dihydroxy-6-(hydroxyethyl)-2H-pyran-3-yl)acetamide] [α]_D²⁰ + 5.76 (c 0.26 MeOH), ESI-MS *m/z* 480.5 [M+Na]⁺ and 937.3 [2M+Na]⁺, NMR data see Table 1.

Acknowledgements This work was financially supported by China Ocean Resource R&D Association (Grant DYXM-115-02-2-13) and the National High Technology Research and Development Program of China (863 No 2006AA09Z410).

References

1 Andrade R, Ayer WA, Mebe PP. The metabolites of *Trichoderma*

- chodema longibrachiatum*. Part I. Isolation of the metabolites and the structure of trichodemol *Can J Chem*, 1992, 70: 2526-2535
- 2 Andrade R, Ayer WA, Trifonov LS. The metabolites of *Trichodema longibrachiatum*. Part II. The structures of trichodemolide and sobiquinol *Can J Chem*, 1996, 74: 371-379
 - 3 Rebuffat S, Hliniš, Prigent Y, et al. Isolation and structural elucidation of the 11-residue peptaibol antibiotic harzianin HK VI *J Chem Soc Perkin Trans 1*, 1996, 16: 2021-2027
 - 4 Augeveir Bour J, Rebuffat S, Auvinc, et al. Harzianin HB I: an 11-residue peptaibol from *Trichodema harzianum*: isolation, sequence, solution synthesis and membrane activity. *J Chem Soc Perkin Trans 1*, 1997, 10: 1587-1594
 - 5 Dunlop RW, Simon A, Sivasithanparam K, et al. Chemical studies on the cecropiaceae: a novel A-ring seco triterpene from *Musanga cecropioides* *J Nat Prod*, 1989, 52: 67-74
 - 6 Cutler HG, Himmelsbach DS, Yagen B, et al. Koniginin B: a biologically active congener of koniginin A from *Trichoderma koningii* *J Agric Food Chem*, 1991, 39: 977-980
 - 7 Parker SR, Cutler HG, Schreiner PR. Koniginin C: a biologically active natural product from *Trichoderma koningii* *Biosci Biotech Biochem*, 1995, 59: 1126-1127
 - 8 Cutler HG, Cutler SJ, Ross SA, et al. Koniginin G, a new metabolite from *Trichoderma aureoviride* *J Nat Prod*, 1999, 62: 137-139
 - 9 Amassi F, Ghisalberti EL, Narvey MJ. New antibiotics from strains of *Trichoderma harzianum*. *J Nat Prod*, 1991, 54: 396-402
 - 10 Ghisalberti EL, Rowland CY. Antifungal metabolites from *Trichoderma harzianum*. *J Nat Prod*, 1993, 56: 1799-1804
 - 11 Abe N, Yamamoto K, Hirota K. Novel fungal metabolites demethylsobicillin and oxosobicillinol isolated from *Trichoderma* sp. USF-2690 *Biosci Biotech Biochem*, 2000, 64: 620-622
 - 12 Zhang J, Zheng ZH, Huang YJ, et al. The antimicrobial activity of a lignicolous fungus *J Xiamen Univ Nat Sci*, 2004, 43: 551-556
 - 13 Zhang J. The screening for bioactive strains from lignicolous fungi and the preliminary study on the secondary metabolites from three marine fungi Master Degree Thesis, Xiamen University, China, 2004
 - 14 Vocadlo DJ, Withers SG. Detailed comparative analysis of the catalytic mechanisms of β -N-Acetylglucosaminidases from families 3 and 20 of glycoside hydrolases *Biochemistry*, 2005, 44: 12809-12818
 - 15 Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assay *J Immunol Methods*, 1983, 65: 55-63