# Stem rust resistance in South African wheat cultivars

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# ABSTRACT

The appearance and anticipated spread of race TTKS (syn. Ug99) of Puccinia graminis f. sp. tritici have renewed interest in breeding for durable resistance to stem rust of wheat. In an attempt to determine the current status of stem rust resistance in South African (SA) bread wheat, 67 cultivars and lines were tested with US and East African races of P. graminis f. sp. tritici. Entries were also screened with DNA markers associated with Sr24 (Sr24#50) and Sr31 (iag95). Sr2 DNA marker (stm559n) data were compared with seedling chlorosis scores to validate the use of this marker in SA genotypes. Most cultivars interacted differentially with the races tested. DNA marker analysis confirmed the presence of Sr31 in seven and Sr24 in 12 entries. Stm559n for Sr2 reliably amplified the correct allele in most local and control lines. However, in several instances the Sr2-associated allele was amplified in presumably non-Sr2 carrying cultivars. This study emphasized that diversification of resistance sources is needed as few SA wheat entries appear to have a broadbased resistance to stem rust.

## **INTRODUCTION**

Stem rust, caused by Puccinia graminis f. sp. tritici, has been an important disease of bread wheat in South Africa (SA) for many years (Pretorius et al., 2007). Over the past three decades regional epidemics have occurred as a result of breakdown of genes such as Sr9e and Sr24 in wheat and Sr27 and SrSatu in triticale. At present stem rust occurs mostly on spring wheat and triticale grown in the winter rainfall areas of the Western Cape. Despite its regular occurrence, information on the genetic base of stem rust resistance in leading SA wheat cultivars and breeding lines is limited. The objective of this study was to determine the status of stem rust resistance in advanced germplasm. In view of recent pathogenic adaptation for virulence in East Africa (Jin et al., 2008), emphasis was placed on the occurrence of Sr24 and Sr31 in SA wheats.

#### MATERIALS AND METHODS

A collection of 54 wheat cultivars and 13 breeding lines was tested for seedling resistance to stem rust races BCCB, MCCF, QFCS, QTHJ, RCRS, RKQQ, TPMK, TTTT, TTKSK and TTKST at the USDA Cereal Disease Laboratory in St Paul, USA. Infection types (ITs) were scored according to a 0 to 4 scale (Stakman et al., 1962)

14 days after inoculation. ITs of 2 and lower were considered indicative of resistance and 3 to 4 as a susceptible host response. The collection was also screened at the University of the Free State, SA for the expression of seedling chlorosis, a phenotype reported to be linked to Sr2 (Brown, 1997). Plants were inoculated with stem rust isolate UVPgt55 12 days after sowing and, following a dew period, kept at 25 to 28°C in a greenhouse. Seedling chlorosis was rated 16 days after inoculation. Suneca (Sr2) and Morocco (susceptible) were included as controls. Entries were tested for the Sr2marker stm559n, which replaced stm559tgag (Hayden et al., 2004; M.J. Hayden, University of Adelaide, personal communication), as well as the Sr31 marker iag95 (Mago et al., 2002; 2005) and the Sr24 marker Sr24#50 (Mago et al., 2005).

## **RESULTS AND DISCUSSION**

Fifty three entries were susceptible to at least one of the 10 stem rust races tested, implying non-durable, hypersensitive resistance in the majority of cultivars. Vulnerability to the African races in particular was emphasized by 42 entries being susceptible to either one of races TTKSK and TTKST. The cultivars Duzi, Caledon, Elands, Pan 3404, Pan 3492, Pan 3364, SST047, SST57, SST94, SST347, SST399 and Steenbras, and experimental lines Mon Exp 2 and Mon Exp 3, were resistant to all races.

Typical seedling chlorosis was observed on the Sr2 control Suneca. However, the relationship between seedling chlorosis and the stm559n marker was not conclusive. Either seedling chlorosis was not equally expressed in the different wheats or the marker detected a similar allele in non-Sr2 genotypes. Nonetheless, several genotypes apparently lacking Sr2 were identified for marker-assisted introgression of this gene. Further work is required to confidently detect Sr2 through marker and seedling assays. In addition, these tests have to be confirmed by stem rust response and pseudo-black chaff expression in the field. Marker analysis confirmed Sr24 in 12 entries. This frequency is lower than expected as 20 entries were postulated to contain the gene based on the IT range (1 to 2) normally associated with Sr24. Although Sr31 is not commonly used in South African wheat breeding, seven entries contained the iag95 marker. All entries carrying this marker had low ITs typical of the Sr31 phenotype to the US races. Except for line Mon Exp 3 (IT 2-), all Sr31 wheats were fully susceptible as seedlings to TTKSK and TTKST.

Sixteen entries, five of which were resistant to all races, were postulated to carry the *SrTmp* gene. Despite being non-durable elsewhere, this gene is still effective in SA. Caution should thus be exercised in cultivars classified as resistant under SA conditions as some of them may rely on single gene resistance. Genetic studies, incorporating both seedling and adult-plant resistance, need to be conducted on resistant cultivars. Breeders could then discern between complex and monogenic resistance and use sources with the best potential of durability in their programs. It is widely accepted that resistance based on hypersensitive seedling resistance is not durable. If effective seedling genes are used, they have to be protected in backgrounds displaying adequate adult plant resistance.

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#### REFERENCES

- Brown, G.N. 1997. The inheritance and expression of leaf chlorosis associated with gene *Sr2* for adult plant resistance to wheat stem rust. Euphytica 95:67-71.
- Hayden, M.J., Kuchel, H and Chalmers, K.J. 2004. Sequence tagged microsatellites for the *Xgwm533* locus provide new diagnostic markers to select for the presence of stem rust resistance gene *Sr2* in bread wheat (*Triticum aestivum* L.). Theor. Appl. Genet. 1009:1641-1647.
- Jin, Y., Szabo, L., Pretorius, Z.A., Singh, R.P. & Fetch, T. 2008. Detection of virulence to Sr24 within race TTKS of *Puccinia graminis* f. sp. *tritici*. Plant Dis. 92:923-926.
- Mago, R., Spielmeyer, W., Lawrence, G.J., Lagudah, E.S., Ellis, J.G. and Pryor, A. 2002. Identification and mapping of molecular markers linked to rust resistance genes located on chromosome 1RS of rye using wheat-rye translocation lines. Theor. Appl. Genet. 104:1317-1324.
- Mago, R., Miah, H., Lawrence, G.J., Wellings, C.R., Spielmeyer, W., Bariana, H.S., McIntosh, R.A., Pryor, A.J. and Ellis, J.G. 2005. High-resolution mapping and mutation analysis separate the rust resistance genes Sr31, Lr26 and Yr9 on the short arm of rye chromosome 1. Theor. Appl. Genet. 112:41-50.
- Pretorius, Z.A., Pakendorf, K.W., Marais, G.F., Prins, R. and Komen, J.S. 2007. Challenges for sustainable control of cereal rust diseases in South Africa. Aust. J. Agric. Res. 58:593-601.
- Stakman, E.C., Stewart, D.M. and Loegering, W.Q. 1962. Identification of physiologic races of

*Puccinia graminis* var. *tritici*. U.S. Dept. Agric., ARS E-617. 53pp.