Development of a non-invasive technique to determine reproductive hormones in cetaceans

Carolyn Hogg

PhD Thesis University of Sydney August 2005

This thesis is submitted to the University of Sydney in fulfilment of the requirement for the Degree of Doctor of Philosophy.

The work presented in this thesis is, to the best of my knowledge and belief, original except as acknowledged in the text. I hereby declare that I have not submitted this material, either in full or in part, for a degree at this or any other institution.

O----h--- 11-----

Carolyn Hogg 31 August 2005



Acknowledgements

Before I take the time to thank all of those that helped me succeed in this project I would like to thank all of those who said it would never work. This was probably the single most important factor that kept me going through the long hours and late nights of problem solving in the lab. My inherent belief that it would work and the knowledge that there was only a select few who believed me made the tedious hours pass by and it all worth it in the end.

I am completely indebted to my fabulous supervisors, Dr Tracey Rogers and Dr Russell Vickers, without whom this small and crazy idea would never have blossomed. Thank you to Tracey for believing in me when everyone else did not for your guidance, support and friendship. I will always be grateful. To Russell, for knowing that you are able to find the smallest and strangest things in saliva if only you have a secure grasp of chemistry and the tenacity to pursue it. Thank you for your support. You helped me discover that I could achieve anything if I was willing to chase it. Thank you to my Associate Supervisor Professor Chis Maxwell for valuable comments on the chapters. Without the great assistance of Sonia Gu, many of the finer points and inner workings of the LC-MS may never have been understood. Sonia was a great teacher, colleague and friend, who sat there and watched as this entire tale unfolded and became a reality. To Tanya Grassi, who kept me sane during those long months of getting the testosterone method to work. Tanya was there through the ups and downs of stability. Thank you for your reassurance that I was not mad and the results were correct and that the rest of them were in fact all wrong. Thank you for the hours of support and the copious amounts of red wine.

This program was made possible by a number of other amazing people. Thank you to Professor Laurie Mather for allowing whale snot to take over the corner of the lab for the past 3.5 years. Thank you also to the staff and management of Sea World Gold Coast for the use of their animals. For always being willing to provide saliva and blow at a moment's notice and ensuring that it all ran smoothly. Thanks in particular to Trevor Long and Guy Bedford and the dolphin trainers of Dolphin Cove and Dolphin Bay, Rosco, Clare, Tasha, Shari, Spike, Rachel, Macca, Todd, Kylie and everyone else.

The aim of this study was to design a method to collect blow from free-swimming whales. Thanks must go to all those who made humpback whale blow collection possible. Thank you to the members of HARC, namely Doug Cato (DTSO), Mike Noad (University of Queensland) and Dale Stokes (Scripps Oceanographic Institute) who made the field seasons possible. Thanks also to Tony White (DSTO) and Dave Clement for their comments on the original pole and bracket. Perhaps the greatest thank you goes to Alex Shorter and Kira Barton (Woods Hole Oceanographic Institute) who believed that with their pole and my snot collector anything was possible. Without their enthusiasm of the first season I believe that there would not have been a second. Thank you to Patrick Miller, Nicolletta Biasonetti and Jim Partan (WHOI) who were the extraordinary crew of the second season and the tagging/snot collecting experts. Thanks also to Michael Moore and his brother in-law who discovered the finer points of carbon fibre poles. However, perhaps the biggest thank you must go to Dave Paton, the most brilliant boat handler around whales I have ever seen. Thank you for your support, humour and long discussions at sea. The whale seasons would not have been the same if it weren't for a few others, Josh Smith (PhD partner-in-crime) who was always willing to share a beer and commiserate, Ceri Morris, Becca Dunlop, Alice Pope and Michaela Ciaglia.

Thanks must also go to my fellow PhD students in the lab, Angelo Jayammane, Hamish Ross, and David Lee, who travelled with me along the journey of character building, late nights and the insanity that constitutes a PhD. To the post docs who kept reminding me that failed experiments eventually become successful ones if you persevere for long enough. Dr Kristina Vikman and Dr Michelle Gerke, thank you for the words of wisdom and chats over coffee.

Lastly but not least to my family and friends who were there every step of the way. To Mum, Dad, Dunc and Ant for never really understanding what I was on about but still being so very proud of me. To Brad for believing that I was the most interesting dinner conversation in years; Susan, for making me meet my milestones; and Sue for always telling me this is what I was destined to achieve. Most importantly, thank you to Jeff, for his lack of interest in the great leviathans and who showed me that there was more to life than whales, who put up with the late night lab runs, waiting in the car when I needed to "just switch something off", for listening to the problems and trying to offer solutions, for giving me the biggest incentive to finish in a timely fashion and most of all for just being there. Thank you from the bottom of my heart.

Executive Summary

Reproductive physiology plays a vital role in population growth and vitality. Baseline data on reproductive physiology and a comprehensive knowledge of breeding biology are essential to conservation management. Great whales have been hunted from the 16th century to the present day. Although many populations are increasing there are populations with low or declining reproductive rates. In 2001 it was recommended to the International Whaling Commission that new techniques be developed to assess the internal physiology of great whales. This study, based on this recommendation, aims to develop analytical methods to assess reproductive hormones in cetacean blow samples and determine the feasibility of its use with free-swimming great whales.

A method for the assessment of steroid hormone concentrations using liquid chromatography-mass spectrometry (LC-MS) was developed and validated. These methods were then used to determine testosterone and progesterone concentrations in saliva and blow of bottlenose dolphins. The stability of testosterone and progesterone was found to be a major issue. Without inhibitors, hormone concentrations increased by up to 65% over three hours at 21°C. Storing samples at low temperatures (-20°C or -80°C) slowed but did not cease the rate of change. The addition of inhibitors, manganese chloride and amoxycillin potassium/clavulanate, improved the stability of testosterone and progesterone. It is proposed that when using dolphin saliva and blow samples to measure reproductive hormones the samples are extracted as soon as possible after collection to prevent degradation. This study highlighted the need to address steroid hormone stability prior to any long-term biological program, to ensure that changes seen in hormone concentration are due to biological activity rather than storage.

A technique to collect blow samples from free-swimming great whales was developed. This technique, in conjunction with the specially developed LC-MS methods allowed for the determination of testosterone and progesterone concentrations in humpback whale blow. The techniques developed in this study to determine reproductive hormones in cetacean saliva and blow have applications for both captive and wild population studies. In captive institutions, saliva and/or blow can be used to monitor reproductive cycling in both females and males. As it is non-invasive it can be used on a daily basis with minimal stress to the animals. The use of blow sampling has the capacity to improve our understanding of reproductive cycling in great whales as it can be used to sample animals in both the breeding and feeding areas. This technique may allow us to now examine whether reproductive dysfunction is playing a role in the slow recovery of critically endangered species such as the North Atlantic right whale.

Table of Contents

Acknowle	edgements	i
Executive	e Summary	iii
Table of	Contents	v
List of Ta	bles	X
List of Fig	gures	xii
Abbrevia	tions Used	xv
Scientific	Names	xvii
•	1 - Introduction	1
1.2 Whale	es and whaling	5
1.3 Horm	ones	9
1.3.1	Steroid hormones	10
1	1.3.1.1 Cholesterol	10
1	1.3.1.2 Steroid synthesis	13
1	1.3.1.3 Enzymes in steroid synthesis	16
1	1.3.1.4 Transport	16
1.3.2	Androgens	22
1.3.3	Progestins and oestrogens	25
1.4 Cetac	ean reproduction	27
1.4.1	Females	28
1.4.2	Males	30
1.5 Curre	ent hormonal sampling techniques	31
1.5.1	Blood	32
1.5.2	Faecal	33
1.5.3	Urine	33
1.5.4	Saliva	34
1.5.5	Other infrequently used hormone sampling methods	35
1.6 Aims	of this study	36

	•	2 – General Methodology noassays	38
		spectrometry	
	2.2.1	History	
2	2.2.2	Chromatography-Mass spectrometry	
2	2.2.3	Mass spectrometric analysis	
2.3	lmmur	noassays vs. LC-MS	
		al method development	
2	2.4.1	Stability	50
2	2.4.2	Internal standard	50
2	2.4.3	Other validation criteria	54
	-	3 – Testosterone uction	55
3.2	Materi	als and Methods	57
3	3.2.1	Chemicals and solutions	57
3	3.2.2	LC analysis	58
3	3.2.3	LC-MS methods	58
3	3.2.4	Sample collection and preparation	61
3	3.2.5	Validation	62
3	3.2.6	Statistical analysis	63
3.3	Result	s	64
3	3.3.1	Validation	64
3	3.3.2	Dolphin samples (endogenous testosterone concentrations)	68
3.4	Discus	ssion	71
3	3.4.1	Validation	71
3	3.4.2	Dolphin samples (endogenous testosterone concentrations)	72
3.5	Concl	usion	73
Cha	apter	4 – Testosterone Stability	
	•	uction	74
4.2 l	Materi	als and Methods	76
4	4.2.1	Sample preparation	76
4	4.2.2	LC-MS assay	77
,	122	Inhibitors	77

5.15.25.3	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 6 Result 5.3.1 5.3.2 5.3.3 5.3.4	5 – Female Reproductive Hormones uction Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation Statistical analysis Progesterone validation Oestradiol validation Oestrone validation Dolphin samples (endogenous progesterone concentrations)	96101102102103104105105109
5.1 5.2	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 5.2.6 Result 5.3.1 5.3.2 5.3.3	Oestrous cycle	96101102102103104105105
5.1 5.2	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 5.2.6 Result 5.3.1 5.3.2	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation Statistical analysis Progesterone validation Oestradiol validation	96101102102102103104105105
5.1 5.2	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 5.2.6 Result 5.3.1	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation Statistical analysis Progesterone validation	96101102102103104105
5.1 5.2	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 5.2.6 Result	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation Statistical analysis	96101102102103104
5.1 5.2	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5 5.2.6	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation Statistical analysis	96101102102102103
5.1	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4 5.2.5	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation Validation	96101102102102
5.1	5.1.1 Materi 5.2.1 5.2.2 5.2.3 5.2.4	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods Sample collection and preparation	96101102102
5.1	5.1.1 Materi 5.2.1 5.2.2 5.2.3	Oestrous cycle als and Methods Chemicals and solutions LC analysis LC-MS methods	96101102
5.1	5.1.1 Materi 5.2.1 5.2.2	Oestrous cycle als and Methods Chemicals and solutions LC analysis	96101102
5.1	Introd 5.1.1 Materi 5.2.1	Oestrous cycle	96 101
5.1	Introd 5.1.1 Materi	Oestrous cycleals and Methods	96 101
5.1	Introd 5.1.1	Uction Oestrous cycle	96
	Introd	uction	
	•	-	96
+.ე	COLICI	u3iVII	93
		ssionusion	
A A		.3.4.4 Blow at -80°C	
		.3.4.3 Saliva at -80°C	
		.3.4.2 Blow at -20°C	
		.3.4.1 Saliva at -20°C	
		Long-term storage	
		.3.3.2 Blow at 21°C	
		.3.3.1 Saliva at 21°C	
	4.3.3	Short-term storage	
	4.3.2	Freeze-thaw experiments	
	4.3.1	Inhibitors	
4.3		'S	
	4.2.7	Statistical analysis	
	4.2.6	Long-term storage	
		-	
	4.2.5	Short-term storage	70

	•	6 – Progesterone Stability	440
		uction	
		als and Methods	
	.2.1	Sample preparation	
_	.2.2	LC-MS assay	
6	.2.3	Inhibitors	
6	.2.4	Freeze-thaw experiments	
6	.2.5	Short-term storage	. 122
6	.2.6	Long-term storage	. 122
6	.2.7	Statistical analysis	. 122
6.3 R	Resul	ts	. 123
6	.3.1	Freeze-thaw experiments	. 123
6	.3.2	Short-term storage	. 123
	6	.3.2.1 Saliva at 21°C	. 123
	6	.3.2.2 Blow at 21°C	. 126
6	.3.3	Long-term storage	. 127
	6	.3.3.1 Saliva at -20°C	. 127
	6	.3.3.2 Blow at -20°C	. 127
	6	.3.3.3 Saliva at -80°C	. 129
	6	.3.3.4 Blow at -80°C	. 129
6.4 D)iscu:	ssion	. 131
6.5 C	oncl	usion	. 133
	-	7 – Whale Blow Collection: is it feasible?	135
7	.1.1	Humpback whale reproduction	. 136
7.2 N	<i>l</i> lateri	als and Methods	. 139
7	.2.1	Sample collection materials	. 139
7	.2.2	Blow sample collection	. 140
7	.2.3	Blow sample analysis	. 146
7	.2.4	0-100% gradient analysis	. 148
7.3 R	Resul	ts	. 151
7	.3.1	Sample collection materials	. 151
7	.3.2	Blow sample analysis	. 151
7	.3.3	0-100% gradient analysis	. 152

7.4 Discu	ussion	159
7.4.1	Sample collection materials	159
7.4.2	Blow sample analysis	159
7.4.3	0-100% gradient analysis	161
7.5 Cond	clusion	162
-	er 8 – General Discussion	
8.1 Analy	ytical chemistry	163
8.2 Stabi	ility	164
8.3 Whal	le blow collection	167
8.4 Cond	clusion	169
Referer	nces	171
Append		
Appendi	ix 1	190
Appendi	ix 2	199

Tables

Chapter 2
Table 2.1: Substances tested as internal standards 52
Chapter 3
Table 3.1: Intra-batch and inter-batch precision and accuracy for
testosterone in dolphin saliva and blow
Table 3.2: Endogenous testosterone concentrations of male
bottlenose dolphins
Chapter 4
Table 4.1: Changes in spiked testosterone concentration in bottlenose dolphin
saliva and blow at 21°C with no inhibitors or inhibitors added
Chapter 5
Table 5.1: Intra- and inter-batch precision and accuracy for progesterone
in dolphin saliva and blow
Chapter 6
Table 6.1: Changes in spiked progesterone concentration in bottlenose
dolphin saliva and blow at 21°C with no inhibitors or inhibitors added 125
Table 6.2: Changes in spiked progesterone concentration in bottlenose
dolphin saliva and blow at -20°C with no inhibitors or inhibitors added
Table 6.3: Changes in spiked progesterone concentration in bottlenose
dolphin saliva and blow at -80°C with no inhibitors or inhibitors added

Table 7.1: a) Humpback whale blow samples collected from	
Peregian Beach in 2003	149
b) Humpback whale blow samples collected from	
Peregian Beach in 2004	150
Table 7.2: Positive $([M+]^+)$ and negative $([M+H]^-)$ mass-to-charge ratios (r)	<i>n/z</i>) fo
some of the precursors and metabolites of testosterone and progesterone	158

List of Figures

Figure 1.1: Six areas of the Antarctic as designated by the International	
Whaling Commission (IWC)	6
Figure 1.2: Five ring cholesterol structure and its derivatives	11
Figure 1.3: Relationship between peptide hormones and steroid hormones	14
Figure 1.4: The principal pathways of human steroid hormone synthesis	17
Figure 1.5a: 21-Carbon pathway for steroid metabolism.	20
Figure 1.5b: Androgen and oestrogen metabolism	21
Figure 1.6: Negative feedback system of the hypothalamic-pituitary-gonadal	24
Figure 1.7: a) The progestins. b) The oestrogens	26
Chapter 2	
Figure 2.1: Chromatogram of dolphin blow in scan mode	42
Figure 2.2: Chromatogram and mass spectrum of dolphin blow in SIM mode	44
Figure 2.3: Formic acid has the same m/z as testosterone	48
Figure 2.4: Acetic acid has the same m/z as oestradiol	49
Figure 2.5: a) MS chromatogram of lidocaine standard. b) MS chromatogram	
of lidocaine, testosterone standard and mixed dolphin saliva	53
Chapter 3	
Figure 3.1: Determination of testosterone m/z by direct infusion	
into the mass spectrometer	59
Figure 3.2: MS chromatogram of saliva sample from dolphin Sirius	
showing the native testosterone peak	60
Figure 3.3: Retention times of testosterone and internal standard	65
Figure 3.4: Direct infusion of cotton gauze into the mass spectrometer	66
Figure 3.5: a) MS chromatogram of endogenous testosterone in dolphin saliva.	
(b) MS chromatogram of endogenous testosterone in dolphin blow	69

Figure 4.1: a) Changes in 50 ng/ml spiked testosterone in bottlenose	
dolphin saliva and blow at 21°C. b) Changes in 0.5 ng/ml spiked	
testosterone in bottlenose dolphin saliva and blow at 21°C	80
Figure 4.2: Differences in testosterone concentration over six hours	
using different MnCl ₂ concentrations	82
Figure 4.3: (a) Stability of saliva spiked with testosterone (5 ng/ml) at	
21°C with Augmentin® and MnCl ₂ over 18 hours. (b) Stability of blow	
spiked with testosterone (50 ng/ml) at 21°C with Augmentin® and	
MnCl₂ over 18 hours	83
Figure 4.4: a) Three cycles of freezing-thawing saliva and blow	
with 50 ng/ml testosterone. b) Three cycles of freezing-thawing saliva and	
blow with 5 ng/ml testosterone	86
Chapter 5	
Figure 5.1: Outline of the oestrous cycle	98
Figure 5.2: Determination of progesterone m/z by direct infusion	106
Figure 5.3: Retention times of progesterone and internal standard	107
Figure 5.4: a) Determination of oestrone m/z by direct infusion.	
b) Determination of oestradiol m/z by direct infusion	110
Figure 5.5: Retention times of a) oestradiol and b) oestrone	111
Figure 5.6: a) MS chromatogram of progesterone in	
pregnant dolphin saliva; and b) progesterone in pregnant dolphin blow	113
Figure 5.7: a) MS chromatogram of progesterone in dolphin saliva.	
b) MS chromatogram of progesterone in dolphin blow	114
Chapter 6	
Figure 6.1: a) Three cycles of freezing-thawing saliva and blow	
with 50 ng/ml progesterone. b) Three cycles of freezing-thawing saliva and	
blow with 0.5 ng/ml progesterone	124

Figure 7.1: Study site at Peregian Beach, Queensland, Australia	141
Figure 7.2: 13-metre carbon fibre pole used to collect blow samples	
from whales, the pole is bracket mounted to the vessel (a) and steered	
by a crew member (b).	142
Figure 7.3: Blow collection device was a 5-inch bamboo double	
ring with nylon stocking that had been sonicated with 100% acetonitrile	
stretched across it	144
Figure 7.4: a) The blow collection device was mounted to the top of the pole	
b) Collection of a blow that was graded as a medium quality sample	145
Figure 7.5: MS chromatograms of scans of different collection materials	153
Figure 7.6: MS chromatogram showing presence of testosterone in	
humpback whale blow	156
Figure 7.7: Chromatogram showing presence of progesterone in	
humpback whale blow	157

Abbreviations Used

ANOVA Analysis of variance

ART Assisted reproductive technology

CL corpus luteum

Cr Creatinine

Da Daltons

DHT 5α-dihydrotestosterone

EDCs Endocrine disrupting chemicals

EIA Enzyme immunoassay

ESI Electrospray ionisation

FSH Follicle stimulating hormone

GC Gas chromatography

GC-MS Gas chromatography-mass spectrometry

GnRH Gonadotropin releasing hormone

H Proton

hCG Human chorionic gonadotropin

HPLC High performance liquid chromatography

hrs Hours

IWC International Whaling Commission

IS Internal standard

LC Liquid chromatography

LC-MS Liquid chromatography-mass spectrometry

LC-MS-MS Liquid chromatography-mass spectrometry-mass

spectrometry

LH Luteinising hormone

LOD Limit of detection

LOQ Limit of quantification

M Molecular mass

MeOH Methanol

min Minute

mins Minutes

MnCl₂ Manganese chloride

MS Mass spectrometry

mw Molecular weight

m/z Mass-to-charge ratio

NI No inhibitors

PdG Pregnanediol-3α-glucuronide

PTP Plasma transport protein

RE Relative error

RIA Radioimmunoassay

RSD Relative standard deviation

RT Retention time

scc Side chain cleavage

secs Seconds

SIM Selected-ion monitoring mode

SPE Solid phase extraction

SRM Selected-reaction monitoring mode

TOF Time-of-flight mass spectrometry

Scientific Names

Asian elephant Elephas maximus

Bandicoot Isoodon macrourus

Black Rhinoceros Diceros bicornis

Bottlenose dolphin Tursiops truncatus

Bowhead whale Balaena mysticetus

California sea lion Zalophus californianus

Capuchin monkeys Cebus apella

Cattle Bos sp.

Chimpanzee Pan troglodytes schweinfurthii

Common dolphin Delphinus delphis

Dairy cow Bos taurus

False killer whale Pseudorca crassidens

Feral horses Equus caballus

Fin whale Balaenoptera physalus

Goat Capra hircus

Gorillas Gorilla gorilla

Harbour seal Phoca vitulina

Horse Equus caballus

Human Homo sapiens

Humpback whale Megaptera novaeangliae

Killer whale Orcinus orca

Minke whale Balaenoptera acutostrata

Monk seal Monachus schauinslandi

Muriquis Brachteles arachnoides

Nile hippopotamus Hippopotamus amphibius

North Atlatncic right whale Eubalaena glacilias

One-humped camel Camelus dromedarius

Pig Sus scrofa

Polar bear Ursus maritimus

Rat Rattus norvegicus

River buffalo Bubalus bubalis

Sei whale Balaenoptera borealis

Sheep Ovis aries

Short-finned pilot whale Globicephala macrorhynchus

Siberian tigers Panthera tigris altaica

Southern right whale Eubalaena australis

Sperm whale Physeter macrocephalus

Timber Wolf Canis lupis

Western lowland gorilla Gorilla gorilla gorilla

White-tailed deer Odocoileus virginianus

"Instead of predicting new events with great precision, or explaining what has already been found, a great deal of scientific progress is linked to what is deemed possible and likely. Expectations, however vague and intuitive, lay the groundwork for discovery....."

Frans de Waal, 2001.