Finding new genes causing motor neuron diseases

# Finding new genes causing motor neuron diseases

Sumana Gopinath

Thesis submitted for the degree of

**Doctor of Philosophy** 

August 2006

Under the supervision of

Professor Garth A. Nicholson

Dr.Marina L. Kennerson

**Faculty of Medicine**,

**University of Sydney** 

## **Statement of authenticity**

No part of the work described in this thesis has been submitted in fulfilment of the requirement for any other academic degree or qualification. Except where duly acknowledged, all experimental work was performed by the author.

Sumana Gopinath

#### Abstract

Neurodegenerative disorders are a diverse group of disorders that affect specific subsets of neurons. Motor neuron diseases, neurodegenerative disorders of motor neurons, are seen commonly as sporadic cases and less frequently as familial disease forms. The familial forms show genetic and phenotypic heterogeneity. Clinically motor neuron diseases may be seen as rapidly progressive disorders like amyotrophic lateral sclerosis, ALS or slowly progressive disorders like hereditary motor neuropathies, HMN.

The only proven causes for motor neuron diseases are gene mutations that lead to motor neuron degeneration in familial disease forms. Only some of these genes have been identified and have contributed greatly to our understanding of the neurobiology of familial and sporadic disease forms. Identification of additional disease causing genes would help enhance our knowledge of the pathophysiological mechanisms underlying all forms of motor neuron disorders, which would lead to early diagnoses, effective prophylaxis and efficient therapies for these disorders.

This study aimed to find gene mutations that cause rapid and slowly progressive familial motor neuron disorders in Australian families and to determine their relevance to sporadic forms of motor neuron disease.

The familial forms of ALS show reduced disease penetrance, that is, not all gene mutation carriers manifest the disease. This study examines ALS penetrance in a group of Australian families. The most frequently observed mutations in ALS families are

cytosolic superoxide dismutase/SOD1 gene mutations. In a collection of ALS families in our centre, families without the common SOD1 gene mutations were genotyped for other ALS genes and loci and studied using genetic linkage and haplotype analyses. Studies in a large Australian ALS family further confirmed genetic heterogeneity in non-SOD familial ALS, all known autosomal dominant ALS genes and chromosomal loci were excluded as cause of disease in this family. Such families can be studied further to identify additional disease genes and loci mapped in other ALS families. These families represent powerful resources for identification of additional ALS genes. Identifying the pathogenic genes in families with reduced disease penetrance may be more relevant to sporadic forms of disease.

dHMN is a chronic neurodegenerative disorder predominantly affecting motor neurons. In a large Australian dHMN family, all the known dHMN genes and chromosomal loci were excluded as cause of disease. A genome wide microsatellite screen was performed in this family and genetic linkage was established to a novel 12.98 Mb locus on chromosome 7q34.2-q36. Candidate genes in this large interval will be screened based on their function and expression profile. Identification of a new dHMN locus provides the basis for future identification of a novel gene involved in motor neuron degeneration.

Genes in dHMN have been shown to be pathogenic in ALS and Charcot Marie Tooth syndromes. The new locus for dHMN mapped in this project would lead to identification of a novel dHMN gene, which may elucidate the pathogenesis underlying a wide range of neurodegenerative disorders.

### Acknowledgements

Firstly, I would like to express my gratitude to all members of the families I have studied. Their support, willingness to be examined at all times and trust has inspired my work. This includes the 14 year old who organised a mufti day in her high school to support our research.

I thank my supervisors, Garth Nicholson and Marina Kennerson, for their support and encouragement, and the opportunities they provided me in the field of science. Marina, I do hope that you have, by now, recovered from editing my thesis. My mentors, Indrani Rajan (from high school in Kalpakkam, India, who initiated me into the realms of molecular sciences and continues to support me from Houston now), Suzanne Hodgkinson, Elizabeth McCusker and Prof. John Morris have always encouraged free thinking. I have earned a great friend in Danqing Zhu, who has been a great help throughout my PhD candidature, both inside the lab and outside. I thank all my friends at Concord: from the Molmed lab, Peter Lorentzos, who helped me settle down in a nonclinical world, Annette Berryman the shock absorber of the lab, Stella Christodoulou for FALS data and sample collection, Helen, Jayamala, Marivic, Patricia, Danijela and Anne; from the neurosciences lab, Alex Drew for several discussions on genetics and computers, Ben, Simon and Ian; Peter Lieu for help in statistics; Cindy my predecessor who was always available to guide me with her own recent PhD candidature experience; and other friends at the ANZAC particularly Mark, Mimi, Pat, James, Christine and Annet.

My project was funded by grants from the MNDRI, Australia and an Australian postgraduate award from the University of Sydney. The MNDA, Victoria awarded me the Nina Buscombe award 2005 for travel assistance to attend the ALS symposium in Dublin, December 2005.

None of this work would have been possible without encouragement and support from my family, Amma and appa for believing that I could embark on this project and coercing it, Keerthy who emphasised that "I must do what enjoy doing" and for being "composed" through the 3.5 years, my sisters and brother for being with me. Appa, your statistical support and graphs have encouraged me to take up similar projects again. Ashoreans-85, your countless mails and endless news helped me stay awake through several nights and let me peep at the bright side of life, especially at times when I needed reminding. Most of all, Sukrut Mysore, who has shared every excitement and anxiety of mine; even edited my work and greatly supported me merely by his presence. This work is dedicated to my son, Sukrut.

#### **Publications**

#### **Papers**

Identification of a new locus for distal hereditary motor neuropathy on chromosome 7q34.2-q36. Submitted for publication to Neurology.

**Genetic causes of ALS: review article**. Submitted to the International journal of Neuroscience.

Haplotype studies in small ALS families support genetic heterogeneity in familial ALS. (in preparation)

**Disease penetrance in familial ALS varies from high tolow merging with sporadic disease.** (in preparation).

#### Abstracts

**Finding new genes causing diseases of motor neurones**. Presented at the Genome Conference, Lorne, Victoria, Australia, February 2004.

**Localising new genes causing Amyotrophic Lateral Sclerosis**. Presented at the Genome Conference, Lorne, Victoria, Australia, February 2004.

Searching for a chromosomal locus for distal hereditary motor neuropathy in an Australian family. Presented at the University of Sydney, Fourth research conference 2004: "From Cell to Society 4" at Leura, NSW, November 2004.

**Finding new genes causing amyotrophic lateral sclerosis**. Presented at the 15<sup>th</sup> International ALS/MND symposium, Philadelphia, December 2004.

**Searching for a new locus associated with hereditary motor neuropathy**. Presented at the University of Sydney, Neuroscience Showcase, August 2005.

Haplotype studies in small ALS families: usefulness in the search for new genes causing ALS. Presented at the 18<sup>th</sup> World Congress of Neurology, Sydney, November 2005 and the 16<sup>th</sup> international ALS/MND symposium Dublin, December 2005.

**Search for a novel chromosomal locus causing distal hereditary motor neuropathy in an Australian family**. Presented at the 18<sup>th</sup> World Congress of Neurology, Sydney, November 2005.

Haplotype studies are useful in small ALS families to search for new gene mutations. Presented at the 26<sup>th</sup> annual meeting of the Australian Neuroscience Society, Sydney, January 2006.

**Novel chromosomal locus for a form of distal hereditary motor neuropathy.** Presented at the annual meeting of Australian Neurologists, Canberra, May 2006. Awarded the James Lance Young Investigator award for best oral presentation.

# Table of contents

Cont	tents										Page
State	ement of a	uthenti	city								ii
Abst	ract										iii
Ackr	nowledgen	nents									v
Publ	ications fr	om thi	s work								vii
Tabl	e of conter	nts									ix
List	of figures	•	•	•	•		•	•	•	•	xiv
List	of tables		•					•	•		xvi
List	of abbrevi	ations	used								xvii
1.	General	Intro	duction	•	•	•	•		•	•	1
1.1	Overviev	V	•	•	•	•	•		•	•	2
1.2	Hypothes	sis and	l aims	•	•	•	•	•	•	•	3
	1.2.1	Over	view of	thesis	•	•	•	•	•	•	5
1.3	Amyotro	phic la	ateral scl	erosis,	ALS	•	•	•	•	•	5
	1.3.1	Intro	duction	•	•	•	•	•	•		5
	1.3.2	Aetic	ological	factors	underly	ing AL	S	•	•		7
	1.3.3	Gene	es causin	g ALS	•		•	•	•	•	9
	1.3	.3.1	Supero	xide di	smutase	21	•	•	•		9
	1.3	.3.2	Alsin	•	•		•	•	•	•	12
	1.3	.3.3	Senata	xin	•	•	•	•	•		13
	1.3	.3.4	VAPB	•	•		•	•	•	•	14
	1.3	.3.5	Dynact	in	•	•	•	•	•		15
	1.3	.3.6	Angiog	genin	•		•	•	•		16
	1.3	.3.7	Other r	isk fact	tor gene	S	•	•	•	•	16
	1.3	.3.8	FALS	chromo	somal l	oci witł	n unider	ntified g	genes	•	19
	1.3.4	Mecl	hanisms	of cell	death in	ALS	•	•	•	•	22
	1.3.5	The g	genetic s	pectrur	n of AL	S: bene	efits and	l limitat	ions		
			of fam	ily stud	ies		•			•	23

1.4	Heredita	ary motor neuropathies, HMN	•	•	26
	1.4.1	Introduction			26
	1.4.2	Genes causing distal HMN		•	27
	1.	4.2.1 Heat shock proteins, <i>HSP 22</i> and <i>HSP 27</i>		•	28
	1.	4.2.2 Glycyl tRNA synthetase, GARS .			29
	1.	4.2.3 Seipin			30
	1.4.3	Study of HMN: Relevance to ALS and other neurodegenerative disorders .			33
1.5	Identific	cation of disease genes by genetic linkage studies			34
	1.5.1	Genetic linkage			35
	1.5.2	Genetic Linkage analysis			36
	1.5.3	Two-point linkage analysis			38
	1.5.4	Multipoint linkage analysis			39
	1.5.5	Simulation studies			40
	1.5.6	Haplotype analyses			40
2.	Genera	l materials and methods			42
<b>2.</b> 2.1		I materials and methods			42 43
	Materia				
	Materia 2.1.1.	ls and equipment			43
	Materia 2.1.1. 2.1.2.	ls and equipment		•	43 43
	Materia 2.1.1. 2.1.2. 2.1.3.	ls and equipment		•	43 43 44
	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresis		• • • •	43 43 44 44
	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA ladders		• • • •	43 43 44 44 45
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA laddersMaterials for tissue culture	· · · ·	• • • • •	43 43 44 44 45 45
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6. DNA ex	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA laddersMaterials for tissue cultureGeneral equipment	· · · · ·	· · · ·	43 43 44 44 45 45 45
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6. DNA ex 2.2.1.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA laddersMaterials for tissue cultureGeneral equipment	· · · · ·	· · · ·	43 43 44 44 45 45 46 47
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6. DNA ex 2.2.1. 2.2.2.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA laddersMaterials for tissue cultureGeneral equipmentNetractionNaterials	· · · ·	· · · · ·	43 43 44 44 45 45 46 47 47
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6. DNA ex 2.2.1. 2.2.2. 2.2.3.	Is and equipmentMaterials for DNA extractionMaterials for Polymerase Chain Reaction (PCR)Materials for gel electrophoresisDNA laddersMaterials for tissue cultureGeneral equipmentAttractionPhenol chloroform extractionPuregene extraction kit	· · · ·	· · · · ·	43 43 44 44 45 45 46 47 47 48
2.1	Materia 2.1.1. 2.1.2. 2.1.3. 2.1.4. 2.1.5. 2.1.6. DNA ex 2.2.1. 2.2.2. 2.2.3. Polymer	Is and equipment       .       .       .         Materials for DNA extraction       .       .         Materials for Polymerase Chain Reaction (PCR)         Materials for gel electrophoresis       .         DNA ladders       .       .         Materials for tissue culture       .       .         Materials for tissue culture       .       .         General equipment       .       .         Ktraction       .       .         Phenol chloroform extraction       .       .         Puregene extraction kit       .       .         Checking the quality and quantity of DNA extractor       .         rase Chain Reaction (PCR)       .       .		· · · · ·	43 43 44 44 45 45 46 47 47 48 49

#### Finding new genes causing motor neuron diseases

	2.3.3.	Method	•	•	•	•	•	•	•	52
	2.3.4.	Troubleshoo	oting							54
2.4.	Primer of	design .								54
2.5.	Gel elec	ctrophoresis								56
	2.5.1.	Agarose gel	electrop	horesis						56
	2.5.2.	Denaturing	polyacry	lamide	gel ele	ctropho	oresis, P	AGE		57
2.6.	Autorad	liography			•					58
2.7.	Genoty	pe data analys	sis .							59
	2.7.1.	Cyrillic								59
	2.7.2.	Selection of	families	for get	notypin	g using	g SIMLI	NK		59
	2.7.3.	Microsatelli	te marke	r analy	sis					60
	2.7.4.	Linkage ana	lysis							60
	2.7.5.	Haplotype c	onstructi	on and	analysi	is .				61
2.8	Tissue c	culture .								61
	2.8.1	Isolation of	lymphoc	ytes fro	om who	le bloo	od.	•		61
	2.8.2	Transformat	ion of ly	mphoc	ytes					63
	2.8.3	Maintenance	e of Lym	phobla	sts			•		63
	2.8.4	Freezing dov	wn cells	•				•		64
	2.8.5	Reculturing	cells from	m froze	en stock	ζ.				64
3.	Recruit	ment of FAI	S famili	es and	searea	ation o	studies			65
3.1	Overvie								•	66
3.2	Method		•					•	•	67
5.2	3.2.1	Ascertainme					•		•	67
	3.2.1	SOD1 muta		•			•	•	•	68
	3.2.2	Estimation of		-				sis	•	68
3.3	Results				0 0	U	•		•	70
5.5	3.3.1	Classificatio						•	•	70
	3.3.2	Mutations in							•	70
	3.3.2	Penetrance i					ALS Iai	·	•	72
	3.3.4	Segregation					-	·	•	78
	J.J. <del>T</del>	Segregation	rano m I		annies	•	•	•	•	70

3.4	Discu	ssion	•	•	•	•	•	•	81
4.	Gene	tic linkage studies in 1	FALS	familie	es.				85
4.1	Overv	view							86
4.2	Mater	ials and methods	•						86
	4.2.1	Family selection	•						86
	4.2.2	Simulation analysis	•						87
	4.2.3	Genetic analysis	•						87
	4.2.4	Statistical analysis	•						87
	4.2.5	Haplotype analysis	•						88
4.3	Result	ts	•						89
	4.3.1	Family selection	•						89
	4.3.2	Simulation analysis	•						91
	4.3.3	Clinical results	•						91
	4.3.4	Linkage analysis	•						94
	4.3.5	Haplotype analyses	•						97
	4.3.6	Exclusion of other lo	ci in f	amily 1	0.				99
	4.3.7	Genetic linkage and	haplot	ype ana	lyses ii	n other ]	FALS fa	amilies	101
4.4	Discu	ssion							103
5.	Clinic	cal assessment and ge	netic ]	linkage	analys	sis in a (	dHMN	family	107
5.1	Overv	view							108
5.2	Subje	cts and methods .							108
	5.2.1	Family selection							108
	5.2.2	Simulation analysis							109
	5.2.3	Genetic analysis	•						109
	5.2.4	Statistical analysis							110
5.3	Resul	ts				•			110
	5.3.1	Family selection							110
	5.3.2	Simulation analysis							112
	5.3.3	Clinical features	•		•				113

#### Finding new genes causing motor neuron diseases

	5.3.4	Genetic linkag	ge anal	ysis	•	•	•	•	•	120
5.4	Discus	ssion .								122
6. Ide	entificat	ion of a novel l	locus fo	or dHN	IN on o	chromo	some 7	q 34.2-	q36	123
6.1 O	verview		•	•	•	•	•	•	•	124
6.2 M	ethods		•	•	•		•		•	125
	6.2.1	Genome wide	screen	ing	•	•	•	•	•	125
	6.2.2	Statistical ana	lysis	•	•	•	•		•	125
	6.2.3	Transcript ma	ap and	selectio	n of ge	nes	•		•	126
6.3 R	esults		•	•	•				•	127
	6.3.1	Linkage anal	ysis of	genome	e wide s	screen d	ata		•	127
	6.3.2	Confirming di	isease 1	ocus on	chrom	osome	7q34.2-	36	•	129
	6.3.3	Defining the c by haplotype				disease				131
	6.3.4	Screening add	litional	dHMN	famili	es for li	nkage t	o novel	locus	134
	6.3.5	Bioinformatic of candidate	•	is of the			us and	selectio	on	138
6.4 D	iscussio	n								139
7. Ge	neral di	scussion and f	uture d	lirectio	ns					143
7.1 In	nportanc	e of studying n	eurode	generat	ive disc	orders				144
7.2 Fa	umilial A	LS studies								145
7.3 St	udies w	ith non-ALS m	otor ne	uron dis	sorders		•		•	148
7.4 Fi	ıture dir	ections .	•	•	•	•	•	•	•	149
Biblic	ography	· .								152
Appe	ndix		•	•	•	•	•	•	•	166
		rosatellite ampl		-				•	•	167
	2 Two	-point LOD sco	ores on	genom	e wide	microsa	tellite s	screen		
		in family 54	perform	ned at A	GRF			•		168

# List of figures

Figure		Р	age
1.1 Classification of degenerative motor neuron disorders .		. 2	
1.2 Disease spectrum in ALS		. 2:	5
1.3 Overlap of genetic causes between ALS, HMN and CMT		. 34	4
1.4 Schematic representation of two-point LOD scores .		. 39	9
1.5 Multipoint linkage analysis		. 39	9
2.1 Schematic representation of PCR technique		. 50	0
2.2 Amplification of target DNA sequence in PCR		. 5	1
2.3 Steps in gel electrophoresis		. 5'	7
2.4 Differential migration across a continuous gradient of Ficoll-Pad	que .	. 62	2
3.1 Expansion of FALS dataset in the Department of Molecular Me	dicine,		
Concord Hospital		. 7	1
3.2 DNA samples available from affected individuals in non-SOD f	amilies.	. 7	1
3.3 SOD1 mutations in the Australian FALS cohort		. 72	2
3.4 Histogram for disease penetrance in Australian FALS families		. 7:	5
3.5 Range of penetrance in Australian FALS families .		. 70	6
3.6 Histogram for disease penetrance in various SOD1 mutations		. 7′	7
3.7 Histogram of segregation ratio in Australian FALS families		. 73	8
3.8 Normal distribution of frequency of segregation ratio in			
SOD1 positive and non-SOD FALS		. 7	9
4.1 Age dependent penetrance of FALS in family 10 .		. 8	8
4.2 FALS pedigrees chosen for further analyses		. 9	0
4.3 Cumulative frequency of disease occurrence in family 10		. 92	2
4.4 Multipoint linkage at FALS loci in family 10		. 9	6
4.5 Pedigree of family 10 with affected individuals genotyped at AI	LS6 loci	ıs 9'	7
4.6 Schematic representation of haplotypes in family 10 at ALS6 lo	cus .	. 98	8
4.7 Family 10 haplotypes in autosomal dominant FALS loci		. 10	00
4.8 Pedigree of family 86		. 10	01

4.9 Family 86 haplotypes in autosomal dominant FALS loci		•	•	102
5.1 dHMN families chosen for SIMLINK analysis .		•		111
5.2 Detailed pedigree of dHMN family 54		•		113
5.3 Pes cavus in affected individuals				114
5.4 Toe deformities in family 54				115
5.5 Muscle wasting in family 54				115
5.6 Normal feet and legs in unaffected members of family 5	4			115
5.7 Sural nerve biopsy in subject II: 10		•		119
6.1 Summary of peak LOD scores by chromosome on genor	ne wide	e		
screen in family 54		•		128
6.2 Schematic representation of partial linkage map on chro	mosom	e 7q34.	2-q36	129
<ul><li>6.2 Schematic representation of partial linkage map on chro</li><li>6.3 Multipoint localisation for novel dHMN locus .</li></ul>		-	-	129 132
	•	•	-	
<ul><li>6.3 Multipoint localisation for novel dHMN locus</li><li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li></ul>	locus	•		132
<ul><li>6.3 Multipoint localisation for novel dHMN locus</li><li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li><li>6.5 Linkage analysis in family 655 at the novel locus</li></ul>	locus	•	•	132 133
<ul> <li>6.3 Multipoint localisation for novel dHMN locus .</li> <li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li> <li>6.5 Linkage analysis in family 655 at the novel locus</li> <li>6.6 Linkage analysis in family 44 at the novel locus .</li> </ul>	locus	•	•	132 133 135
<ul> <li>6.3 Multipoint localisation for novel dHMN locus .</li> <li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li> <li>6.5 Linkage analysis in family 655 at the novel locus</li> <li>6.6 Linkage analysis in family 44 at the novel locus .</li> <li>6.7 Linkage analysis in family 353 at the novel locus</li> </ul>	locus	•	•	132 133 135 136
<ul> <li>6.3 Multipoint localisation for novel dHMN locus</li> <li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li> <li>6.5 Linkage analysis in family 655 at the novel locus</li> <li>6.6 Linkage analysis in family 44 at the novel locus</li> <li>6.7 Linkage analysis in family 353 at the novel locus</li> <li>6.8 Linkage analysis in family 702 at the novel locus</li> </ul>	locus	· · ·		132 133 135 136 136
<ul> <li>6.3 Multipoint localisation for novel dHMN locus .</li> <li>6.4 Family 54 pedigree with haplotypes at the novel dHMN</li> <li>6.5 Linkage analysis in family 655 at the novel locus</li> <li>6.6 Linkage analysis in family 44 at the novel locus .</li> <li>6.7 Linkage analysis in family 353 at the novel locus</li> <li>6.8 Linkage analysis in family 702 at the novel locus</li> </ul>	locus	· · ·		<ol> <li>132</li> <li>133</li> <li>135</li> <li>136</li> <li>136</li> <li>137</li> </ol>

# List of tables

## Table

## Page

1.1 FALS genes and chromosomal loci	•	10
1.2 Harding classification of dHMN		27
1.3 Chromosomal loci and genes in dHMN		32
2.1 Reagents used for PCR optimisation reaction		52
2.2 Standard temperature cycles used for PCR		53
2.3 Reagents used in "hot" radiolabelled PCR		53
2.4 Estimated electrophoresis time at 65 W for different sized PCR produc	ets	58
3.1 Segregation analyses for penetrance in SOD1 positive FALS families		73
3.2 Segregation analyses for penetrance in non SOD FALS families		74
3.3 Comparison of Gaussian parameters for segregation ratio in		
SOD1 positive and non-SOD FALS	•	80
4.1 Results of simulation analysis for linkage in FALS families .	•	91
4.2 Clinical features of ALS in family 10	•	93
4.3 Two-point and multipoint linkage at autosomal dominant		
ALS loci in family 10	•	95
4.4 Two-point linkage results at ALS loci in family 86	•	101
5.1 Simulation data on power analysis for selected dHMN families .	•	112
5.2 Clinical features of neuropathy in affected members of family 54	•	116
5.3 Nerve conduction studies in affected members of family 54 .	•	117
5.4 Two-point LOD scores between disease and dHMN loci in family 54	•	120
5.5 Exclusion of autosomal dominant dHMN loci in family 54 .		121
6.1 Markers with two-point LOD score $> 1.5$ on genome wide screen		127
6.2 Two-point and multipoint LOD scores between novel dHMN locus		
and microsatellite markers on chromosome 7q34.2-q36 .		130
6.3 Candidate genes for dHMN in chromosome 7q34.2-q36.		138

## Abbreviations

AD	Autosomal dominant
AGRF	Australian Genome Research facility
ALS	amyotrophic lateral sclerosis
AOA2	Ataxia ocular apraxia -2
APEX nuclease	Apurinic/apyrimidic endonuclease
AR	Autosomal recessive
CMAP	Compound Muscle action potential
CMT	Charcot Marie Tooth syndrome
CNTF	Ciliary neurotrophic growth factor
CRGH	Concord Repatriation General Hospital
CSAHS	Central Sydney area health service
CSF	Cerebrospinal fluid
DCTN1	Dynactin 1
dHMN	Distal hereditary motor neuropathy
DNA	Deoxyribonucleic acid
dNTPs	deoxynucleotide triphosphates
EAAT2	Excitatory amino acid transporter 2
EBV	Ebstein Barr virus
EDTA	Ethylene diamino tetra acetic acid
EMG	Electromyography
FALS	Familial ALS
FDA	Federal drug authority
FTD	Fronto-temporal dementia
GC content	Guanine and cytosine content
HLA	Human leucocyte antigens
HMN	Hereditary motor neuropathy (ies)
HSP	Heat shock protein
LL	Lower limbs
LOD	Linkage of odds
MEP	Magentic evoked potentials
MND	Motor neuron disease
NAIP	Neuronal apoptosis inhibitory protein
NF	Neurofilament
NIPPV	Non-invasive intermittent positive pressure ventilation
OMIM	Online Mendelian inheritance in man
PAGE	Polyacrilamide gel electrophoresis
PCR	Polymerase chain reaction
PD	Parkinson's disease
PON	Paroxonase
RFLP	Restriction fragment length polymorphisms
RNA	Ribonucleic acid
SALS	Sporadic ALS
SMA	Spinal muscular atrophy
SNAP	Sensory Nerve action potential
SOD1	Superoxide dismutase 1
SPG	Spastic paraplegia

TE	Tris EDTA
tRNA	Transfer RNA
UBI	Ubiquinated inclusions
UCSC	University of California at Santacruz (Human Genome browser)
UL	Upper limbs
VAPB	Vesicle associated membrane protein B and C
VEGF	Vascular endothelial growth factor