The role of Alternative Lengthening of Telomeres in human cancer

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A thesis submitted to the University of Sydney in fulfilment of the requirements for the degree of Doctor of Philosophy

The Children's Medical Research Institute

Faculty of Medicine

The University of Sydney

June, 2006

Statement of Originality

The contents of this thesis have not been presented for the award of a degree or diploma at this or any other university. The data presented are the original work of the author except where specifically indicated in the text.

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Acknowledgements

First and foremost I would like to thank my supervisor, Roger Reddel, for his support, and guidance. Roger is one of the most impressive people I have been associated with and has been an excellent role model. Roger's persistent censoring of my scientific talks has probably been beneficial to both our careers. I would also like to thank my associate supervisor, Tracy Bryan. I have the highest respect for her expert guidance and wisdom.

I am grateful to Peter Rowe and the Children's Medical Research Institute for giving me the opportunity and support to complete my PhD in an environment that fosters science. I am also grateful to the NHMRC for their scholarship support.

I thank all the people who have contributed in many ways during this work, including the following people. Especially, Axel Neumann, Sara Cole, Alessandra Muntoni, Elizabeth Collins, Paul Wang, Sharon Cunningham and all members of the Cancer research Group and the CMRI who have shared their scientific expertise with me and made this an enjoyable place to work. Tony Henwood who provided advice on tumour preparation, Jennifer Peat and Katrina Williams who provided advice on statistical analysis, and John Melki and Mark Hughes who provided advice on Microarray analysis.

Thank you to Daniel Catchpoole, Lyra Pearson, Aileen Baillie, Sue Camara, Vicky Hakin-Smith, Thomas Carroll, David Levy, Suzanne Jackson, the Westmead

Children's Hospital Tumour Bank and members of the Royds laboratory for tumour samples and data without which this work would not be possible.

I am grateful to Nicola Royle, Nadia Zaffaroni, Curt Harris and Wei Qin Jiang for giving me the opportunity to collaborate with their interesting projects.

And finally I am eternally grateful to my parents for their endless support and encouragement.

Publications

- 1. <u>Henson, J.D.</u>, Neumann, A.A., Yeager, T.R., and Reddel, R.R. (2002). Alternative lengthening of telomeres in mammalian cells. Oncogene 21, 598-610.
- 2. <u>Henson, J.D.</u>, Hannay, J.A., McCarthy, S.W., Royds, J.A., Yeager, T.R., Robinson, R.A., Wharton, S.B., Jellinek, D.A., Arbuckle, S.M., Yoo, J., Robinson, B.G., Learoyd, D.L., Stalley, P.D., Bonar, S.F., Yu, D., Pollock, R.E., and Reddel, R.R. (2005). A robust assay for alternative lengthening of telomeres (ALT) in tumors demonstrates the significance of ALT in sarcomas and astrocytomas. Clin. Cancer Res. 11, 217-225.
- 3. Jeyapalan, J.N., Varley, H., Foxon, J.L., Pollock, R.E., Jeffreys, A.J., <u>Henson, J.D.</u>, Reddel, R.R., and Royle, N.J. (2005). Activation of the ALT pathway for telomere maintenance can affect other sequences in the human genome. Hum. Mol. Genet. 14, 1785-1794.
- 4. Jiang, W.Q., Zhong, Z.H., <u>Henson, J.D.</u>, Neumann, A.A., Chang, A.C.M., and Reddel, R.R. (2005). Suppression of alternative lengthening of telomeres by Sp100-mediated sequestration of MRE11/RAD50/NBS1 complex. Mol. Cell. Biol. 25, 2708-2721.
- 5. Costa, A., Daidone, M.G., Daprai, L., Villa, R., Cantù, S. Pilotti, S., Mariani, L., Gronchi, A., <u>Henson, J.D.</u>, Reddel, R.R. and Zaffaroni, N. Presence and Clinical Relevance of Telomere Maintenance Mechanisms in Liposarcoma. Cancer Research (Manuscript submitted, 2006)
- 6. Jiang, W.Q., Zhong, Z.H., <u>Henson, J.D.</u> and Reddel, R.R. (2005). Screen of candidate genes required for ALT by RNA interference and methionine restriction. Oncogene (Manuscript submitted 2006)
- 7. Bowman, E.D., Mechanic, L., <u>Henson, J.D.</u>, Reddel, R.R., Welsh, J., Khan, M., Hewitt, S., Dennis, P. and Harris. C.C. AKT phosphorylation and telomere lengthening in lung cancer prognosis (Manuscript in preparation).

Presentations

- 1. <u>Henson, J.D.</u>, Yeager, T.R., Robinson, R.A., Yoo, J., Robinson, B.G., Learoyd, D.L., Jellinek, D.A., Wharton, S.B., Royds, J.A., Eeles, R. and Reddel, R.R. Investigating Alternative Lengthening of Telomeres (ALT) in human tumors by detection of ALT-Associated PML Bodies. Lorne Cancer Conference, 2002. (Winner of Poster Prize)
- 2. <u>Henson, J.D.</u>, Yeager, T.R., Robinson, R.A., Yoo, J., Robinson, B.G., Learoyd, D.L., Jellinek, D.A., Wharton, S.B., Royds, J.A. and Reddel, R.R. Investigating Alternative Lengthening of Telomeres (ALT) in human tumors by detection of ALT-Associated PML Bodies. Australian Society for Medical Research, XIth NSW Scientific Meeting, 2002.
- 3. <u>Henson, J.D.</u>, Yeager, T.R., Arbuckle, S.M., Robinson, R.A., Yoo, J., Jellinek, D.A., Wharton, S.B., Royds, J.A., McCarthy, S.W., Eeles, R. and Reddel, R.R. Investigating Alternative Lengthening of Telomeres (ALT) in human tumors by detection of ALT-Associated PML Bodies. Australian Telomere Workshop I, 2002.
- 4. <u>Henson, J.D.</u>, Yeager, T.R., McCarthy, S.W., Pollock, R.E., Arbuckle, S.M., Robinson, R.A., Yoo, J., Robinson, B.G., Learoyd, D.L., Jellinek, D.A., Wharton, S.B., Royds, J.A., Eeles, R. and Reddel, R.R. Investigating Alternative Lengthening of Telomeres (ALT) in human tumors by detection of ALT-Associated PML Bodies (APBs). Lorne Cancer Conference, 2003.
- 5. <u>Henson, J.D.</u>, Hannay, J.A., McCarthy, S.W., Royds, J.A., Yeager, T.R., Robinson, R.A., Wharton, S.B., Jellinek, D.A., Arbuckle, S.M., Yoo, J., Robinson, B.G., Learoyd, D.L., Stalley, P.D., Bonar, S.F., Yu, D., Pollock, R.E., and Reddel, R.R. A robust assay for alternative lengthening of telomeres (ALT) in tumors demonstrates the significance of ALT in sarcomas and astrocytomas. Australian Telomere Workshop II, 2004.
- 6. <u>Henson, J.D.</u>, Hannay, J.A., Royds, J.A., McCarthy, S.W., Yeager, T.R., Nuemann, A.A., Robinson, R.A., Wharton, S.B., Jellinek, D.A., Arbuckle, S.M., Yoo, J., Harris, C.C., Robinson, B.G., Learoyd, D.L., Stalley, P.D., Bonar, S.F., Yu, D., Pollock, R.E., and Reddel, R.R. Development of an assay for Alternative Lengthening of Telomeres (ALT). Children's Hospital Westmead, Grand Rounds, 2004.
- 7. <u>Henson, J.D.</u>, Yeager, T.R., McCarthy, S.W., Pollock, R.E., Arbuckle, S.M., Bonar, S.F., Robinson, R.A., Yoo, J., Jellinek, D.A., Wharton, S.B., Royds, J.A., Robinson, B.G., Learoyd, D.L., Eeles, R. and Reddel, R.R. A robust assay for alternative lengthening of telomeres (ALT) in tumors demonstrates the significance of ALT in sarcomas and astrocytomas. Australian Health and Medical Research Congress, 2004.

- 8. <u>Henson, J.D.</u>, Hannay, J.A., McCarthy, S.W., Royds, J.A., Yeager, T.R., Robinson, R.A., Wharton, S.B., Jellinek, D.A., Arbuckle, S.M., Yoo, J., Robinson, B.G., Learoyd, D.L., Stalley, P.D., Bonar, S.F., Yu, D., Pollock, R.E., and Reddel, R.R. A robust assay for alternative lengthening of telomeres (ALT) in tumors demonstrates the significance of ALT in sarcomas and astrocytomas. Children's Cancer Institute Australia, Seminar, 2005.
- 9. <u>Henson, J.D.</u>, Hannay, J.A., McCarthy, S.W., Royds, J.A., Yeager, T.R., Robinson, R.A., Wharton, S.B., Jellinek, D.A., Arbuckle, S.M., Yoo, J., Robinson, B.G., Learoyd, D.L., Stalley, P.D., Bonar, S.F., Yu, D., Pollock, R.E., and Reddel, R.R. A robust assay for alternative lengthening of telomeres (ALT) in tumors demonstrates the significance of ALT in sarcomas and astrocytomas. Australian Institute for Medical Scientists, National Scientific Meeting, 2005.

Summary

Activation of a telomere maintenance mechanism is a vital step in the development of most cancers and provides a target for the selective killing of cancer cells. Cancers can use either telomerase or Alternative Lengthening of Telomeres (ALT) to maintain their telomeres and inhibition of either telomere maintenance mechanism can cause cancer cells to undergo senescence or apoptosis. Although telomerase inhibitors are undergoing clinical trials, on commencing this study very little was known about the role of ALT in cancer, what proteins were involved in its mechanism and regulation and how it could be targeted clinically. The primary aim of this thesis was to develop an assay for ALT suitable for examining archived tumour specimens and to begin using it to examine the prevalence and clinical significance of ALT in cancer. This assay and gene expression analysis was also used to identify genes that are involved in or associated with the activation of the ALT mechanism, to contribute towards the overall goal of an ALT cancer therapy.

The ALT mechanism involves recombination mediated replication and ALT cells have a marked increase in a range of recombinational events specifically at their telomeres. Presumably, as a consequence of this the telomere lengths of ALT cells are very heterogeneous and on average long. This can be detected by terminal restriction fragment (TRF) Southern analysis, which has been used previously as the definitive test for ALT activity. However, TRF analysis requires intact genomic DNA and is unsuitable for tumour specimens which are commonly archived by paraffin embedding. Another hallmark of ALT is ALT-associated PML bodies (APBs) which are the subset of PML bodies that contain telomeric DNA. Work done in this study to

consolidate APBs as a hallmark of ALT, combined with published data, showed 29/31 ALT[+], 3/31 telomerase[+] and 0/10 mortal cell lines/strains are APB[+]. The three APB[+]/telomerase[+] cell lines identified here had an order of magnitude lower frequency of APB[+] nuclei than the ALT[+] cell lines. APBs may be functionally linked to the ALT mechanism and contain the recombination proteins that are thought to be involved in the ALT mechanism. This study, in collaboration with Dr W-Q Jiang, strengthened this functional link by demonstrating that loss of ALT activity (as determined by TRF analysis) coincided with the disruption of APBs.

The detection of APBs was developed into a robust assay for ALT in archived tumour specimens using a technique of combined immunofluorescence and telomere fluorescence *in situ* hybridisation. It was demonstrated that the APB assay concurred exactly with the standard assay for ALT (TRF analysis) in 60 tumours for which TRF analysis gave unequivocal results. The APB assay may be a more appropriate technique in the case of tumour specimen heterogeneity, which may explain why the APB assay was able to give definitive results when TRF analysis was equivocal. We demonstrated that intratumoral heterogeneity for ALT does exist and this could explain why about 3% of tumours in this study were APB[+] but with more than a ten-fold reduction in the frequency of APB[+] nuclei. This study also made the novel discovery of single stranded C-rich telomeric DNA inside APBs which potentially could be used to make the APB assay more suitable for routine pathology laboratory use.

The APB assay was used to show that ALT is a significant concern for oncology. ALT was utilised in approximately one quarter of glioblastoma multiforme (GBM),

one third of soft tissue sarcomas (STS) including three quarters of malignant fibrous histiocytomas (MFH), half of osteosarcomas and one tenth of non-small cell lung carcinomas (NSCLC). Furthermore, the patients with these ALT[+] tumours had poor survival; median survivals were 2 years for ALT[+] GBM, 4 years for ALT[+] STS including 3.5 years for ALT[+] MFH and 5 years for ALT[+] osteosarcoma. ALT[+] STS and osteosarcomas were also just as aggressive as their ALT[-] counterparts in terms of grade and patient outcome. ALT status was not found to be associated with response to chemotherapy in osteosarcomas or survival in STS. ALT was however, less prevalent in metastatic STS.

The APB assay was a prognostic indicator for GBM and was correlated with three fold increased median survival in GBM (although this survival was still poor). ALT was more common in lower grade astrocytomas (88% ALT[+]) than GBM (24% ALT[+]) and ALT[+] GBM had an identical median age at diagnosis to that reported for secondary GBM. It is discussed that these data indicate that ALT was indirectly associated with secondary GBM and is possibly an early event in its progression from lower grade astrocytoma. This is relevant because secondary GBM have distinct genetic alterations that may facilitate activation of the ALT mechanism.

Putative repressors of ALT could explain why this study found that ALT varied among the different STS subtypes. ALT was common in MFH (77%), leiomyosarcoma (62%) and liposarcoma (33%) but rare in rhabdomyosarcoma (6%) and synovial sarcoma (9%). ALT was not found in colorectal carcinoma (0/31) or thyroid papillary carcinoma (0/17) which have a high prevalence of telomerase

activity and a reduced need for a telomere maintenance mechanism (low cell turnover), respectively.

A yeast model of ALT predicts that one of the five human RecQ helicases may be required for ALT. Using the APB assay to test for the presence of ALT in tumours from patients with known mutations in either WRN or RECQL4 it was demonstrated that neither of these RecQ helicases is essential for ALT. Although p53 and mismatch repair (MMR) proteins have been suggested to be possible repressors of ALT, there was no apparent increase in the frequency of ALT in tumours from patients with a germline mutation in p53 codon 273 or in colorectal carcinomas that had microsatellite instability and thus MMR deficiency. Also contrary to being a repressor of ALT but consistent with its ability to interact with a protein involved in the ALT mechanism, the MMR protein MLH1, was demonstrated to be present in the APBs of an ALT[+] cell line.

To further test for genes that may be involved in the ALT mechanism or associated with its activation, RNA microarray was used to compare the gene expression of 12 ALT[+] with 12 matched telomerase[+] cell lines; 240 genes were identified that were significantly differentially expressed (p<0.005) between the ALT[+] and telomerase[+] cell lines. Only DRG2 and SFNX4 were significantly differentially expressed after adjusting for the estimated false positive rate. Overall, DRG2, MGMT and SATB1 were identified as most likely to be relevant to the ALT[+] tumours and Western analysis indicated that DRG2 and MGMT levels were down-regulated after activation of ALT and up-regulated after activation of telomerase, whereas SATB1 protein levels appeared to be up-regulated after immortalisation but to a higher degree

with activation of ALT compared to telomerase. Since lack of MGMT is known to be a determinant of temozolomide sensitivity in GBM, the possibility that ALT and the APB assay could be used to predict temozolomide sensitivity is discussed. The microarray data was consistent with MGMT expression being suppressed by EGF (p < 0.05), indicating that caution may be needed with combining EGFR inhibitors with temozolomide in ALT cancers. One ALT[+] cell line which did not express MGMT had TTAA sequence in its telomeres. This could possibly have resulted from mutations due to lack of MGMT expression and a possible role for MGMT in the ALT mechanism is discussed.

Further analysis of the microarray data identified two groups of co-regulated genes (p $< 5 \times 10^{-5}$): CEBPA, TACC2, SFXN4, HNRPK and MGMT, and SIGIRR, LEF1, NSBP1 and SATB1. Two thirds of differentially expressed genes were down-regulated in ALT. Chromosomes 10 and 15 had a bias towards genes with lower expression in ALT while chromosomes 1, 4, 14 and X had a bias towards genes with higher expression levels in ALT.

This work has developed a robust assay for ALT in tumour specimens which was then used to show the significance of ALT in sarcomas, astrocytomas and NSCLC. It has also identified genes that could possibly be molecular targets for the treatment of ALT[+] cancers.

Abbreviations

ADP adenosine diphosphate

AEBSF 4-(2-aminoethyl)-benzenesulphonyl fluoride hydrochloride

ALT Alternative Lengthening of Telomeres

APB ALT-associated PML body

AT ataxia telangiectasia

ATCC American Type Culture Collection

BCNU bis-(2-chloroethyl)-nitrosourea

base pair

BS Bloom syndrome

BSA bovine serum albumin

BUR base unpairing region

CF cystic fibrosis

CHAPS 3-[(3-cholamidopropyl)-dimethylammonio]-1-

propanesulphonate

CI confidence interval

CO-FISH chromosome orientation fluorescence *in situ* hybridization

cpm counts per minute

CSL Commonwealth Serum Laboratories

DEPC diethyl pyrocarbonate

DABCO 1,4 Diazabicyclo(2.2.2)octane

DAPI 4',6-Diamidino-2-phenylindole dihydrochloride

DMSO dimethylsulphoxide

DMEM Dulbecco's Modified Eagle Medium

DNA deoxyribonucleic acid

DNAse deoxyribonuclease

dNTP deoxynucleoside 5'-triphosphate

DSB double strand break

DTT dithiothreitol

ECTR extra-chromosomal telomeric repeats

EDTA Ethylenediaminetetraacetate

EGTA Ethylene glycol-bis(2-aminoethylether)-N,N,N',N'- tetraacetate

ExoI Exonuclease I

ExoIII Exonuclease III

FBS foetal bovine serum

FISH fluorescence *in situ* hybridisation

FITC fluorescein isothiocyanate

GBM glioblastoma multiforme

HBS HEPES-Buffered Saline

HJ Holliday junction

HMT histone methyltransferases

HPV Human Papilloma Virus

HR homologous recombination

HRP horseradish peroxidase

hTERT human Telomerase Reverse Transcriptase

hTR human Telomerase RNA

LB lysis buffer

LFS Li Fraumeni syndrome

LHC Laboratory of Human Carcinogenesis

LHC-BM Laboratory of Human Carcinogenesis basal medium

LNS Lesch-Nyhan syndrome

MAR matrix attachment region

MBN Mung bean nuclease

MFH malignant fibrous histiocytoma

MMR mismatch repair

MOPS 3-(N-morpholino)propane-sulphonic acid

MRN MRE11/ RAD50/ NBS1 complex

MSI microsatellite instability

MSS microsatellite stable

NCI-Frederick National Cancer Institute at Frederick Division of Cancer

Treatment Tumor Repository

NER nucleotide excision repair

NHEJ non-homologous end joining

NIGMS National Institute of General Medical Sciences Human Genetic

Cell Repository

NP40 Nonidet P-40

NSCLC non small cell lung carcinoma

O⁶-AG O⁶-alkylguanine

O⁶-MG O⁶-methylguanine

PAGE Polyacrylamide gel electrophoresis

PBS phosphate buffered saline

PD population doubling

TWEEN-20 polyoxyethylene-sorbitan monolaurate

PVP-40 polyvinylpyrrolidone 40

RNA ribonucleic acid

RNAse ribonuclease

RPMI Roswell Park Memorial Institute

RT room temperature

RTS Rothmund-Thomson syndrome

SBTI soy bean trypsin inhibitor

SCE sister chromatid exchange

SDS sodium dodecyl sulphate

SIR semi interquartile range

SMC structural maintenance of chromosome (protein)

SSC sodium chloride-trisodium citrate buffer

STS soft tissue sarcoma

SV40 Simian virus 40

TBE tris-borate-EDTA buffer

TE tris-EDTA buffer

TEMED N,N,N',N'-tetramethylethylenediamine

TMM telomere maintenance mechanism

TNE tris-sodium chloride-EDTA buffer

TRAP Telomere Repeat Amplification Protocol

TRD telomeric rapid deletion

TRF terminal restriction fragment

TRIS tris(hydroxymethyl)aminomethane

v/v volume for volume

WHO World Health Organisation

WS Werner syndrome

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