

## Supplementary Information for

### CD11c<sup>+</sup>CD88<sup>+</sup>CD317<sup>+</sup> myeloid cells are critical mediators of persistent CNS autoimmunity

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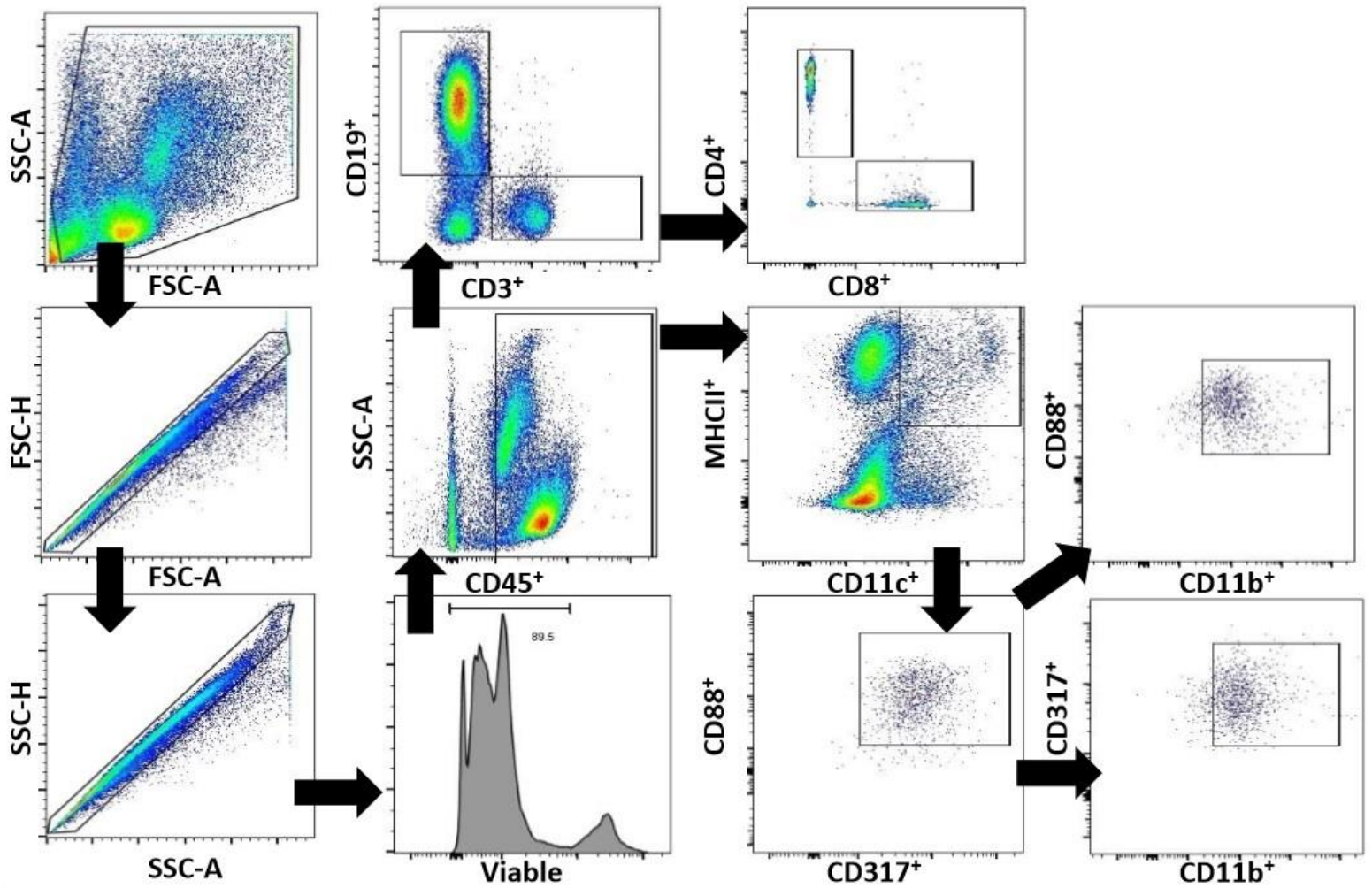
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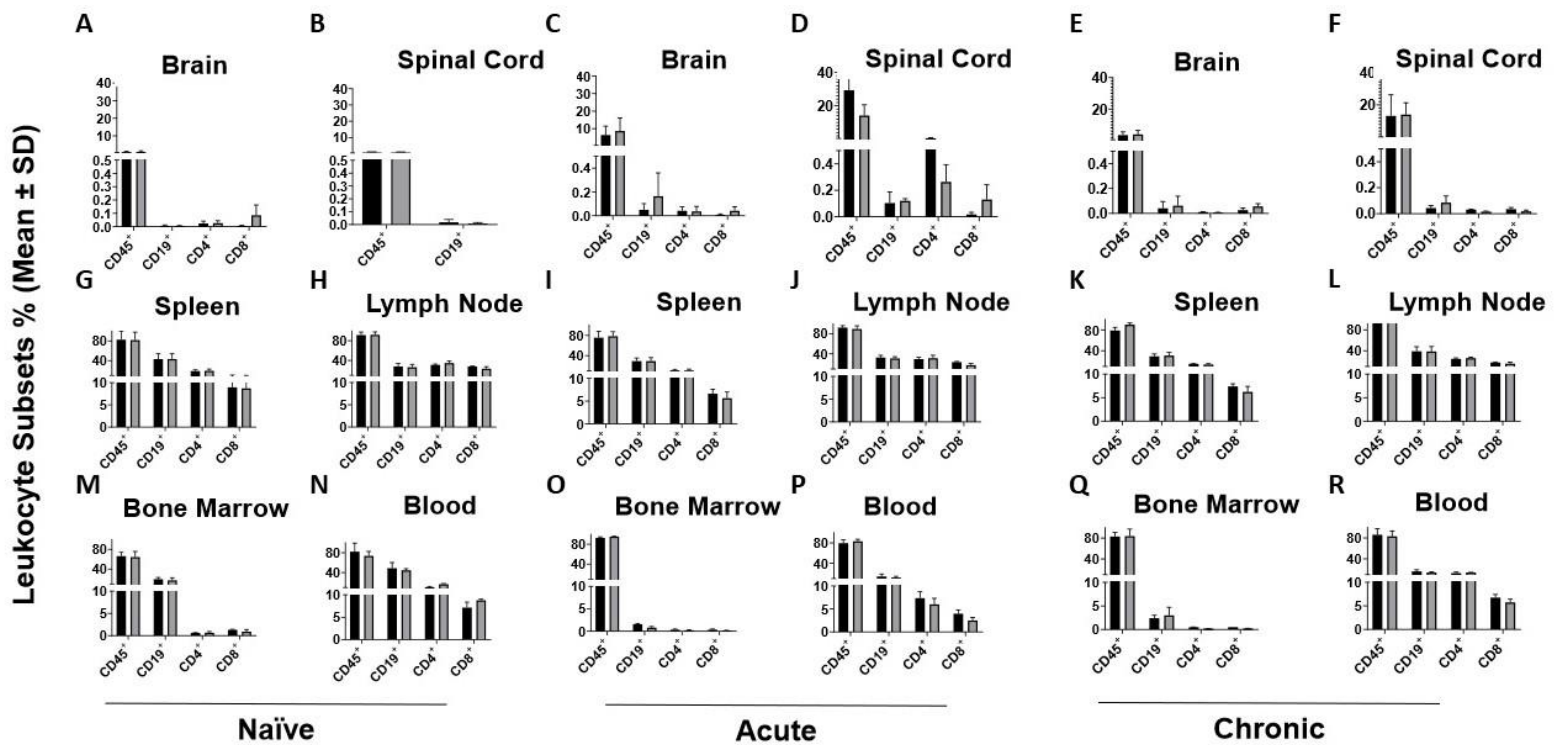
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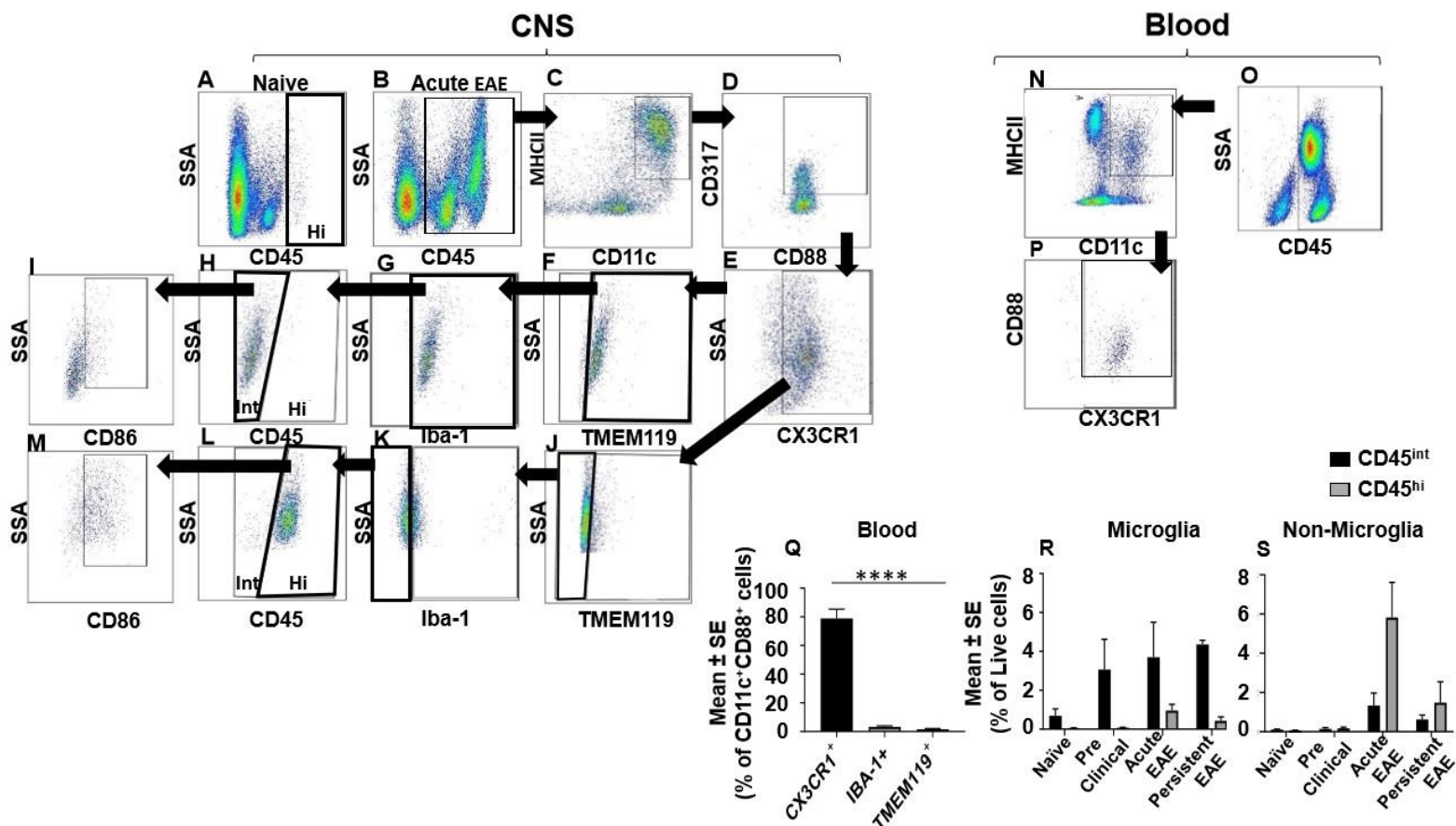
Figures S1 to S3



**Supplementary Figure 1.** Flow cytometry study gating strategy. Cells were gated according to morphology side scatter (SSC-A) vs forward scatter (FSC-A). Doublets were excluded (FSC-A vs FSC-H and SSC-A vs SSC-H). Live cells were selected using the viability dye. Flow cytometry density plots showing gating strategy used to identify CD11c<sup>+</sup>CD88<sup>+</sup>CD317<sup>+</sup> cells (CD45<sup>+</sup>MHCII<sup>+</sup>D11c<sup>+</sup>CD88<sup>+</sup>CD317<sup>+</sup>), B cells (CD45<sup>+</sup>CD3<sup>+</sup>CD19<sup>+</sup>), CD4<sup>+</sup> T cells (CD45<sup>+</sup>CD3<sup>+</sup>CD4<sup>+</sup>) and CD8<sup>+</sup> T cells (CD45<sup>+</sup>CD3<sup>+</sup>CD8<sup>+</sup>). Each sample contains a minimum of 50 x 10<sup>3</sup> live events.



**Supplementary Figure 2.** The composition of leukocytes outside the CD11c<sup>+</sup> lineage is not altered in CD11c.Cre<sup>+/-</sup>/ITGA4<sup>fl/fl</sup> mice. The mean ± standard deviation (SD) of leukocyte subsets (%) from total 50 x 10<sup>3</sup> recorded viable cells in different compartments, including (A-F) brain and spinal cord, (G-L) spleen and lymph node and (M-R) bone marrow and blood, is presented during naïve, acute and persistent clinical actively-induced experimental autoimmune encephalomyelitis (EAE) in CD11c.Cre<sup>+/-</sup>/ITGA4<sup>fl/fl</sup> mice or C57BL/6 wild type (WT) controls (N =6 experimental animals per group; data show pooled analysis of all study cohorts). There was no difference in frequency of cell types including CD45<sup>+</sup>, CD19<sup>+</sup>, CD4<sup>+</sup> and CD8<sup>+</sup> cells between the two groups during different stages of the disease (P value > 0.05).



**Supplementary Figure 3:** Characterization of CD11c<sup>+</sup>CD88<sup>+</sup>CD317<sup>+</sup> cells and microglia using *ex vivo* flow cytometry. (A) CD45<sup>hi</sup> cells were not abundantly present in naïve CNS tissue. (B) From CD45<sup>+</sup> cells, (C) CD11c<sup>+</sup>MHCII<sup>+</sup> and (D) CD88<sup>+</sup>CD317<sup>+</sup> cells were selected. Next, (E) CX3CR1<sup>+</sup> cells were gated for, and selected for (F) expression of TMEM119<sup>+</sup> or (J) the absent expression of TMEM119<sup>-</sup>. (G) Iba-1<sup>+</sup> cells were selected from TMEM119<sup>+</sup> cells. (H, R) The majority of CX3CR1<sup>+</sup>TMEM119<sup>+</sup>Iba-1<sup>+</sup> cells defined as parenchymal microglia localized in the CD45<sup>int</sup> gate. (K) Iba-1<sup>-</sup> cells were selected from TMEM119<sup>-</sup> cells. (L, S) The majority of CX3CR1<sup>+</sup>TMEM119<sup>-</sup>Iba-1<sup>-</sup> cells defined as non-microglia localized in the CD45<sup>hi</sup> gate. (I) Fewer CD45<sup>int</sup> microglia expressed CD86 compared to (M) CD45<sup>hi</sup> non-microglia. (O, N) CD45<sup>+</sup>CD11c<sup>+</sup>CD88<sup>+</sup> cells in the blood were mostly (P) CX3CR1<sup>+</sup>, but (Q) TMEM-119<sup>-</sup> and Iba-1<sup>-</sup>.