Title: Intake of New Zealand blackcurrant powder affects skin-borne volatile organic compounds in middle-aged and older adults

Authors: M.E.T. Willems¹, M. Todaka², M. Banic^{1,3}, M.D. Cook⁴, Y. Sekine²

Affiliations: ¹ Institute of Sport, University of Chichester, United Kingdom, ² Graduate School of Science, Tokai University, Japan, ³ Faculty of Health Sciences and Sport, University of Stirling, United Kingdom, ⁴ Institute of Sport and Exercise Science, University of Worchester, United Kingdom

Corresponding author: Prof Mark Willems, Institute of Sport, University of Chichester, Chichester, United Kingdom, phone: +44 (0)1243 816468, m.willems@chi.ac.uk. Email: m.willems@chi.ac.uk. ORCID: 0000-0003-4385-8739

Abstract

Skin volatile organic compounds (VOCs) can cause body odor or reveal human disease and may result from lipid peroxidation or activity by skin bacteria. We examined the effect of intake of New Zealand blackcurrant (NZBC) powder for 77 skin VOCs in middle-aged and older adults in a cross-over design. Fourteen adults (9 males, age: 55±5 yr) consumed NZBC powder for 7 days (6 g·day⁻¹ with 138.6 mg anthocyanins). Two hours after the last intake, a passive flux sampler with trapping media was applied in the base of the neck for one hour. Gas chromatography-mass spectrometry was used for media analysis. Habitual anthocyanin intake was quantified using a food frequency questionnaire. Compared to control (i.e. no intake of NZBC powder), emission of six skin VOCs, i.e. 2-nonenal, acetic acid, 2-hexanone, 6-methyl-5-hepten-2-one, benzaldehyde, allyl methyl sulphide, were lower by more than 25%. Increases were observed for γ -octanolactone (+184%) and γ -decanolactone (+89%). A trend for decrease was observed for isovaleraldehyde, hexanal and 2-pentanone and an increase for heptanoic acid and γ -nonanolactone. There was a significant correlation with daily habitual dietary anthocyanin intake for control values of hexanal and percentage change of γ -octanolactone. NZBC powder can change emanation of some VOCs in human skin. Analysis of skin VOCs following specific polyphenol intake may address the impact of dietary components to affect internal metabolic processes, body odor and health.

KEYWORDS: volatile organic compounds, metabolism, anthocyanins, lipid peroxidation, body odor, aging

Introduction

In humans, volatile organic compounds (VOCs) are chemicals that are primarily produced by endogenous biochemical processes. VOCs are emitted via urine, faeces, breath and skin (de Lacy Costello et al., 2014). The chemical classification of the VOCs (e.g. carboxylic acids, alcohols, aldehydes, and ketones) is based on functional groups in their chemical structure and it allows the analysis of a VOC profile (Baranska et al., 2013). The VOC profile or even the occurrence of a particular VOC can be altered by disease conditions, e.g. breath VOCs in chronic obstructive pulmonary disease [Van Berkel et al., 2010 (e.g. isoprene)] and Alzheimer (Mazzatenta et al., 2015), and urine and faeces VOCs for colorectal cancer (Bond et al., 2019; Mozdiak et al., 2019), in addition to the risk for development of a pathophysiological condition (Lacey et al., 2018) and body odor, e.g. skin VOC 2-nonenal (Ishino et al., 2010; Jha, 2017). The analysis of breath and skin emissions of VOCs is an emerging research discipline to allow detection of non-invasive biomarkers of internal metabolic processes linked with health and disease.

Skin-borne VOCs [532 named VOCs (de Lacy Costello et al., 2014] can originate from sebaceous and sweat glands and may reveal the outcome of the glands metabolic processes and activity of skin bacteria. However, the endogenous, non-gland, origin of many skin VOCs has not been established. VOC skin emanations can act as an attractant for mosquitos to locate humans (Bernier et al., 2000) but also provide opportunity to detect entrapped humans by building collapse (Agapiou et al., 2013). In addition, some skin-borne VOCs are associated with body odor, e.g. the unsaturated aldehyde 2-nonenal in older adults (Haze et al., 2001).

In healthy humans, the normal physiological and biochemical processes that contribute to an endogenous origin of VOCs can be affected by many lifestyle factors, e.g. smoking, psychological stress, exercise, and diet. For example, a gluten free diet over a period of 4 weeks altered the VOC profile of exhaled breath (Baranska et al., 2013). In addition, the VOC profile of exhaled breath is also affected by certain foods such as garlic, leeks and coffee (Krilaviciute et al., 2019). Garlic intake is probably one of the best-known foods to affect exhaled breath as well as having a positive effect on perceived body odor hedonicity (Fialová et al., 2016) and maybe due to anti-oxidant and anti-microbial properties affecting the skin emission of VOCs (Sato et al., 2020). Coffee also affected the VOC profile of urine (Mack et al., 2019). In contrast, sugar beet pectin supplementation (15g/day for 4 weeks) did not affect exhaled VOC profiles in healthy young adults and healthy elderly (An et al., 2019). However, evidence is emerging how our food choices affects an individual VOC profile. Our knowledge, however, on the effect of specific dietary components on emanation and thus production of VOCs is very much in its infancy. Such information may provide support for existing and future development of dietary guidelines. Current dietary guidelines emphasize a regular intake of fruits and vegetables. The health benefits by intake of many colourful fruits and vegetables have been linked with the polyphenol content, with many studies emphasizing the role of the flavonoid anthocyanin (Lila et al., 2016). Many in vitro studies provided support for anti-oxidant and anti-inflammatory effects of anthocyanins. In addition, reviews have summarized the effects of anthocyanins on the gut microbiome and linked with reduced risk for disease (e.g. Speer et al., 2020; Hair et al., 2021). We have shown that intake of anthocyanin-rich NZBC powder enhanced insulin sensitivity in healthy individual (Willems et al., 2017). Intake of anthocyanin-rich NZBC powder also shifted the exercise-induced lactate curve in endurance-trained participants (Willems et al., 2015). As such, the potential for anthocyanins to affect biochemical and physiological processes may influence the production of VOCs. However, the effect of anthocyanin intake in humans on the skin emission of VOCs is not known.

Therefore, the aim of the present study was to examine the effect of anthocyanin-rich NZBC powder on emission of VOCs from the skin in middle-aged and older adults. For example, in middle-aged and older adults, body odor is associated with emission of 2-nonenal and likely due to a change of the anti-oxidant defence system with aging. It was hypothesised that emission of 2-nonenal would be reduced with intake of blackcurrant anthocyanins by lowering lipid peroxidation. However, this study is also explorative in nature and aims to examine whether a rich-anthocyanin containing supplement can change the emission rate of skin VOCs with unclear origin in middle-aged and older adults.

Methods

Participants (Caucasian, male: n=9, female: n=5) were recruited from staff of the University of Chichester in the United Kingdom (age: 55±5 yr, mean±SD, range 49-64 yr). The study was approved in accord with the Research ethics policy of the University of Chichester (ethical approval code: 1718_29). Written informed consent by the participants was obtained after the procedures and aims of the study were explained. The study used a cross-over design. Skin sampling was done in April and May (Zhang et al., 2005) during a working day between 8 am and 2 pm. During the one hour of skin sampling, participants were involved in office desk type activities and collection time was similar for each participant for the control and NZBC conditions. For the NZBC powder condition, participants consumed the NZBC powder for 7 days (Sujon Berries, Nelson, New Zealand, 6 g·day⁻¹ with 138.6 mg anthocyanin, 49 mg Vit C, 5.2 g of carbohydrates and total phenolic content of 271.6 mg per serving) (Willems et al., 2015). Participants were advised to dissolve the powder in water. Two hours after the last intake on day 7, a passive flux sampler (Kimura et al., 2016; Sekine et al., 2018; Umezawa et al., 2018) was applied to the skin in the base of the neck for one hour. Participants were advised not to use hot water or soap for 48 hrs on the skin sampling location to exclude potential exogenous sources of skin VOCs (Gallagher et al., 2008). For the control condition, which preceded the NZBC powder condition, the same participants did not take a placebo as there can be no bias on the outcome measures of the study. The diet of the participants was not controlled. Habitual dietary anthocyanin intake was quantified using a food frequency questionnaire with anthocyanin sources listed in Phenol explorer (Neveu et al., 2010).

Skin samplers were shipped from Tokai University (Japan). The shipment contained two blank samplers during the flight travel from Japan to the United Kingdom and back to correct for potential contamination by VOCs by the travel conditions. The sampler consisted of a polypropylene screw cap, trapping media (MonoTrap®, DCC18, GL Science, Japan) and O-ring as a stopper. Gas chromatography-mass spectrometry was used for media analysis of VOCs according to procedures in Kimura et al. (2016). Samplers for analysis by gas chromatography-mass spectrometry were numbered to keep researchers blind to the treatments. Briefly, VOCs were extracted with 500 µL of dichloromethane after 15-min ultrasonic extraction. All extracts were analyzed on the mass selective detector, JMS-Q1050GC MkII (JEOL, Japan) interfaced to gas chromatograph, Agilent model 7890B. One μ L injections of sample extracts, blank extracts, and quantitation standards were made with a split ratio of 35:1 onto DB-IMS (30 m×0.25 mm I.D.×0.25 µm film thickness, Agilent technologies, USA). Helium was used as the carrier gas with a flow rate of 1.0 mL·min⁻¹. The injector port was maintained at 280 °C. The extracts were analyzed using the following column temperature program: 50 °C hold for 8 min, increase at 6 °C · min⁻¹ to 120 °C and increase at 20 °C · min⁻¹ to 280 °C hold for 2 min. Data on most sample extracts were acquired using real time SIM. The emission flux (i.e. emission rate per area) of volatile compounds, E $(ng \cdot cm^{-2} \cdot h^{-1})$ was obtained by

$$E = \frac{W}{St} \tag{1}$$

where W is a collection amount of analyte (ng) by the PFS, S is an effective cross-section of the trapping media (0.594 cm^2) and t is a sampling duration (h). Travel blanks were subtracted when observed.

Statistical analysis

Normality was checked with a D'Agostino and Pearson omnibus normality test (Graphpad Prism 5 for Windows). Two-tailed Wilcoxon signed rank test (i.e. no Gaussian distribution) or two-tailed paired t-test were used for non-parametric and parametric testing, respectively. Cohen's d effect sizes were calculated and considered trivial (d < 0.2), small (d = 0.2-0.49), moderate (d = 0.5-0.79) and large (d \ge 0.8), respectively (Cohen et al, 1988). For the VOCs for which a change or trend of change was observed with intake of NZBC powder, Pearson correlations were calculated between habitual dietary anthocyanin intake and 1) absolute values of VOCs in the control condition, 2) the observed percentage changes of skin VOCs. In addition, Pearson correlations were calculated between the skin VOCs that changed with intake of NZBC powder. Data are reported as mean±SDs with 95% confidence intervals and box-and-whisker plots for significantly altered VOCs. Significance was accepted at P < 0.05 with $0.05 \ge P \le 0.1$ interpreted according to guidelines by Curran-Everett and Benos (2004).

Results

In total, the emission rate of 77 skin VOCs were analysed with eight compounds showing a significant change (P < 0.05) and five compounds a trend for change ($0.05 \ge P \le 0.1$) after 7-days intake of NZBC powder (Table 1). Table 2 provides the emission rate of the skin VOCs that did not change after 7-day intake of NZBC powder.

Aldehydes

In the NZBC powder condition, the unsaturated aldehyde 2-nonenal was reduced with large effect size (control: 4.66 ± 3.44 , 95% CI [2.69, $6.51 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; NZBC powder: 2.38 ± 1.46 , 95% CI [1.54, $3.22 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; Wilcoxon signed rank; d = -0.87; P = 0.030) (Fig. 1A) with eight participants (~57%) having decreases more than 30% with an average of $69\pm16\%$ (range 32-82%). The two participants with the highest 2-nonenal emission in the control condition, i.e. 9.41 and 13.81 ng $\cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$, showed reductions of 80% and 82%, respectively. Habitual dietary anthocyanin intake was not significantly correlated with control 2-nonenal values (r = -0.28, P = 0.33) and percentage change in 2-nonenal with intake of NZBC powder (r = 0.36, P = 0.21).

Benzaldehyde was reduced with large effect size (control: 2.67 ± 1.54 , 95% CI [1.78, $3.56 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; NZBC powder: 1.45 ± 0.93 , 95% CI [0.91, 1.99 ng $\cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; two-tailed t-test; d = - 0.96; P = 0.04) (Fig. 1B) with nine participants having lower values with NZBC powder intake. The average decrease of those 9 participants was $66\pm29\%$ (range 13-99%). Dietary anthocyanin intake was not significantly correlated with control benzaldehyde values (r = 0.10, P = 0.74) and percentage change in benzaldehyde with intake of NZBC powder (r = 0.06, P = 0.84).

A weak trend for a decrease was observed for isovaleraldehyde but with moderate effect size (control: 7.56 ± 3.37 , 95% CI [5.61, 9.50 ng·cm⁻²·hr⁻¹], NZBC powder: 5.41 ± 2.26 , 95% CI [4.10, 6.72 ng·cm⁻²·hr⁻¹]; two-tailed t-test; d = - 0.75; P = 0.09) with 10 participants (~71%) having lower values with NZBC powder intake. The average decrease of those 10 participants was 47±20% (range 23-83%). Dietary anthocyanin intake was not significantly correlated with control isovaleraldehyde values (r = - 0.44, P = 0.12) and percentage change in isovaleraldehyde with intake of NZBC powder (r = - 0.36, P = 0.20).

A strong trend for a decrease was observed for hexanal with large effect size (control: $8.77\pm4.41, 95\%$ CI [6.22, 11.31 ng·cm⁻²·hr⁻¹]; NZBC powder: $5.65\pm3.25, 95\%$ CI [3.77, 7.52 ng·cm⁻²·hr⁻¹]; two-tailed t-test; d = -0.81; P = 0.06) with 11 participants (~79%) having lower values with NZBC powder intake. The average decrease of those 11 participants was $44\pm29\%$ (range 4-85%) with 6 participants having larger decreases than the average. Dietary anthocyanin intake was correlated with control hexanal values (r = - 0.53, P = 0.047) and with a trend for percentage change in hexanal with intake of NZBC powder (r = 0.47, P = 0.09). The % change in hexanal correlated with the % change in γ -nonanolactone (r = 0.65, P = 0.022) and the % change in γ -decanolactone (r = 0.54, P = 0.047). These percentage change in for VOCs were the only ones that provided significant correlations.

Ketones

A decrease in skin emission rate was observed for the ketone 6-methyl-5-hepten-2-one (control: 6.55 ± 3.04 , 95% CI [4.79, $8.31 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; NZBC powder: 3.40 ± 1.96 , 95% CI [2.26, $4.53 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; two-tailed t-test; d = -1.23; P = 0.005) (Fig. 1C) with 12 participants (~85%) having lower values with NZBC powder intake. The average decrease of those 12 participants was $58\pm23\%$ (range 28-94%). Habitual dietary anthocyanin intake was not significantly correlated with control 6-methyl-5-hepten-2-one values (r = 0.42, P = 0.14) and percentage change in 6-methyl-5-hepten-2-one with intake of NZBC powder (r = - 0.09, P = 0.76).

2-hexanone was decreased with large effect size (control: 1.84 ± 0.77 , 95% CI [1.40, $2.29 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; NZBC powder: 1.26 ± 0.58 , 95% CI [0.92, $1.59 \text{ ng} \cdot \text{cm}^{-2} \cdot \text{hr}^{-1}$]; two-tailed t-test; d = -0.86; P = 0.03) (Fig. 1D) with 11 participants (~79%) having lower values. The average decrease of those 11 participants was $41\pm27\%$ (range 1-87%). Habitual dietary anthocyanin intake was not significantly correlated with 2-hexanone values in the control

condition (r = 0.05, P = 0.87) and percentage change in 2-hexanone with intake of NZBC powder (r = 0.20, P = 0.49).

There was a weak trend for a decrease of the ketone 2-pentanone with moderate effect size (control: 1.95 ± 0.93 , 95% CI [1.42, 2.49 ng·cm⁻²·hr⁻¹]; NZBC powder: 1.28 ± 0.97 , 95% CI [0.71, 1.84 ng·cm⁻²·hr⁻¹]; two-tailed t-test; d = -0.71; P = 0.09) with 11 participants (~79%) having lower values. The average decrease of those 11 participants was $54\pm30\%$ (range 8-88%). Habitual dietary anthocyanin intake was not significantly correlated with control 2-pentanone values in the control condition (r = - 0.23, P = 0.43) and percentage change in 2-pentanone with intake of NZBC powder (r = - 0.25, P = 0.47).

Carboxylic acids

With NZBC powder, a decrease in skin emission rate was observed for the carboxylic acid acetic acid with large effect size (control: 580 ± 207 , 95% CI [461, 700 ng·cm⁻²·hr⁻¹]; NZBC powder: 369 ± 117 , 95% CI [301, 436 ng·cm⁻²·hr⁻¹]; two-tailed t-test; d = -1.26; P = 0.006) (Fig. 1E) with 11 participants (~79%) having lower values. The average decrease of those 11 participants was $43\pm20\%$ (range 1-68%). Habitual dietary intake of anthocyanin was not significantly correlated with control acetic acid values (r = 0.15, P = 0.61) and percentage change in acetic acid with intake of NZBC powder (r = 0.26, P = 0.37).

There was a strong trend for an increase in skin emission rate for heptanoic acid with moderate effect size (n=13, control: 1.69 ± 1.24 , 95% CI [0.94, 2.43 ng·cm⁻²·hr⁻¹]; NZBC powder: 2.74 ± 1.63 , 95% CI [1.76, $3.72 \text{ ng·cm}^{-2}\cdot\text{hr}^{-1}$]; d = 0.73; P = 0.065) with 7 participants (~54%) having higher values. The average increase of those 7 participants was 389±400% (range 23-1217%), indicating for heptanoic acid substantial individual variability in response. Habitual dietary intake of anthocyanin was not significantly correlated with control heptanoic

acid values (r = 0.29, P = 0.32) and percentage change of heptanoic acid with intake of NZBC powder (r = -0.40, P = 0.17).

Cyclic esters

With NZBC powder, an increase in emission rate for γ -octanolactone was observed (control: 0.54±0.76, 95% CI [0.11, 0.98 ng·cm⁻²·hr⁻¹]; NZBC powder: 1.54±1.23, 95% CI [0.83, 2.25 ng·cm⁻²·hr⁻¹]) with large effect size (d = 0.98, P = 0.023) (Fig. 2A), and 11 participants (~79%) having higher values. The average increase for those 11 participants was 2801±5730% (range 29-18391%), indicating for γ -octanolactone substantial variation among responders. Habitual dietary intake of anthocyanins was not significantly correlated with control γ -octanolactone values (r = - 0.22, P = 0.46). However, the percentage change of γ -octanolactone with intake of NZBC powder was significantly correlated with habitual dietary intake of anthocyanins (r = - 0.71, P = 0.005), with the significance obtained by two *extreme* responders.

An increase for the emission rate of γ -decanolactone was observed (control: 0.46±0.24, 95% CI [0.33, 0.60 ng·cm⁻²·hr⁻¹]; NZBC powder: 0.87±0.47, 95% CI [0.60, 1.15 ng·cm⁻²·hr⁻¹] (Fig. 2B) with large effect size (d = 1.11, P = 0.015), and 11 participants (~79%) having higher values. The average increase for those 11 participants was 243±245% (range 8-686%). Habitual dietary intake of anthocyanins was not significantly correlated with control γ -decanolactone (r = - 0.29, P = 0.31) and percentage change of γ -decanolactone with intake of NZBC powder (r = - 0.38, P = 0.18).

A strong trend for an increase was observed for γ -nonanolactone (n=12 as in 2 participants γ -nonanolactone could not be detected in the control condition, control: 1.09±0.97, 95% CI [0.48, 1.71 ng·cm⁻²·hr⁻¹]; NZBC powder: 1.78±1.27, 95% CI [0.97, 2.57 ng·cm⁻²·hr⁻¹] with moderate effect size (d = 0.62, P = 0.063), and 8 participants (~67%) having higher values. The average increase for those 8 participants was $168\pm100\%$ (range 31-308%). Habitual dietary intake of anthocyanins was not significantly correlated with control γ -nonanolactone (r = 0.17, P = 0.61) and percentage change of γ -nonanolactone with intake of NZBC powder (r = 0.12, P = 0.71).

Sulfur containing compound

A decrease for emission rate of allyl methyl sulfide was observed (control: 0.83 ± 0.19 , 95% [0.72, 0.94 ng·cm⁻²·hr⁻¹]; NZBC powder: 0.61 ± 0.23 , 95% [0.48, 0.75 ng·cm⁻²·hr⁻¹] (Fig. 1F) with large effect size (d = -1.01, P = 0.015), and 11 participants (~79%) having lower values. The average decrease for those 11 participants was $39\pm19\%$ (range 13-78%). Habitual dietary intake of anthocyanins was not significantly correlated with control allyl methyl sulfide (r = -0.12, P = 0.69) and percentage change of allyl methyl sulfide with intake of NZBC powder (r = -0.12, P = 0.69).

Discussion

The present study provides first evidence that consumption of anthocyanin-rich NZBC powder affects the emission rate of volatile organic compounds from human skin. Out of the 77 skin VOCs that were analysed, 13 VOCs (i.e. 18%) showed a significant change or a trend for change with 9 out of 13 (i.e. 69%) with large effect size. In the present study, the daily intake of 6 gram of NZBC powder could have been obtained with about 80-100 gram of fresh NZBCs, suggesting ecological validity with application for consideration of daily blackcurrant intake or potentially other anthocyanin-sources to be part of a healthy diet. The polyphenolic profile of blackcurrant is made up primarily of four anthocyanins, i..e. cyanidin-*3-O*-glucoside, cyanidin-*3-O*-rutinoside, delphinidin-*3-O*-glucoside and delphinidin-*3-O*-rutinoside (Kapasakalidis et al., 2006). It needs to be noted, however, that all berries have a

very distinct polyphenol composition, e.g. blackberry and blueberry contain 2 and 13 anthocyanins, respectively (Lee et al., 2015). It is therefore possible that our observations on changes in emission rate of skin VOCs are unique with respect to the anthocyanin profile of blackcurrant, and maybe due to conversion of the absorbed anthocyanins to phenolic acids by colonic bacteria (Aura et al., 2005). Future work may address the emission rate of skin VOCs of berries with different polyphenol compositions.

Emission of VOCs that are associated with body odor in older adults were lowered, i.e. 2-nonenal (Haze et al., 2001) and 6-methyl-5-hepten-2-one (Ozeki et al., 2016). 2nonenal has a greasy and grassy odor that is perceived as unpleasant (Haze et al., 2001). Haze et al. (2001) provided evidence that 2-nonenal is produced by the oxidative degradation of unsaturated fatty acids, i.e. by lipid peroxidation of fatty acids produced in the sebaceous glands (Ishino et al., 2010). Observations of 2-nonenal skin emissions have been observed in Japanese over 39 years of age (Haze et al., 2001) and non-Japanese populations between 41-70 years of age (Gallagher et al., 2008), so likely differences in dietary intake (i.e. higher intake of marine-based foods in Japanese) between those groups was not a likely explanation for the production of 2-nonenal in older adults. Age-induced lipid peroxidation is upregulated in older adults independent of dietary intake and therefore expected to be present in the participants of the present study. Intake of NZBC in humans has been shown to reduce lipid peroxidation by lowering the amounts of exercise-induced plasma carbonyls (Lyall et al., 2009). The present study provides therefore indirect observations of the ability of blackcurrant intake to reduce biochemical consequences of age-related oxidative stress in the sebaceous glands. The ketone 6-methyl-5-hepten-2-one results from oxidative degradation of the sebaceous gland product squalene (Ozeki et al., 2016). Lower values for the emission rate of the skin VOC 6-methyl-5-hepten-2-one maybe due to reduced synthesis of squalene and subsequently a smaller amount of squalene being available for oxidative degradation. 6methyl-5-hepten-2-one is suppressed in older adults (mean age 84.5 years) by inhibition of squalene peroxidation by 6 weeks use of soap containing the anti-fungal miconazole nitrate (Ozeki et al., 2016). Overall, the reduction of the body odor components 2-nonenal and 6-methyl-5-hepten-2-one by intake of NZBC powder may have application for the well-being of older adults with concern for their body odor.

Emission of acetic acid by the skin has been associated with foot malodour (Caroprese et al., 2009) and body odor in young adults (Lam et al., 2018). The acetic acid is thought the result of bacterial metabolic activity that results in transformation of secretions by the eccrine glands. Cutaneous Propionibacterium and staphylococcus epidermidis synthesize acetic acid (Lam et al., 2018; Piwowarek et al., 2018). Therefore, if NZBC inhibits activity of propionibacterium and staphylococcus epidermidis may affect skin health due to their role in acne (Wang et al., 2016). Interestingly, tea polyphenols are suggested to reduce sebum production in the skin and have potential for acne treatment (Saric et al., 2016). Other phenolic compounds, such as ferulic acid which is a metabolite of cyanidin-3-*O*-glucoside (De Ferrars et al., 2014) have been proposed as well with potential for skin disorders (Działo et al., 2016). However, we cannot exclude that reduced sebum production occurred by intake of NZBC powder.

Benzaldehyde is the aromatic aldehyde linked with almond flavour (Oliveira et al., 2019) and was reduced with intake of NZBC powder. Benzaldehyde is also a VOC released by muscle cells (Mochalski et al., 2014), but its function within the muscle cells is not defined. Benzaldehyde is also produced in skin flora by Pseudomonas aeruginosa (Timm et al., 2018) and associated with skin infections. It is possible that the intake of NZBC powder reduces pseudomomonas aeruginosa on human skin and contributing to a maintenance of a healthy skin. In support, protocatechuic acid, an anthocyanin-derived metabolite, leads to death of Pseudomonas aeruginosa in in-vitro conditions (Ajiboye et al., 2017). However, as

far as we know, the implication of the reduction in the VOC human skin emission of benzaldehyde by intake of NZBC powder is not clear. Future work could address the role of anthocyanins on skin VOCs in people with skin disorders.

Our understanding on the endogenous origin of 2-hexanone detected by skin emission is absent. Inhalation of atmospheric 2-hexanone should be avoided. In plastic fabric workers, for example, occupational exposure to 2-hexanone (methyl butyl ketone) caused acute mixed motor and sensory neuropathy (Landrigan et al., 1980). When present in urine, it was suggested by Mochalski and Unterkofler (2016) that 2-hexanone could be the product of 2hexanol oxidation. In urine, 2-hexanone was considered a potential biomarker for lung cancer (Santos et al., 2017). In breath, 2-hexanone and 2-pentanone were changed in response to hypoxic conditions (7620 m, 8% O2) (Harsman et al., 2015). It was suggested that 2hexanone (and 2-heptanone) may be due to beta-oxidation of unsaturated fatty acids in the lungs (Harsman et al., 2015). However, different processes can be responsible for volatile compounds emanated from skin and breath.

Allyl methyl sulphide was reduced by the intake of NZBC powder. Allyl methyl sulphide is a bioactive compound derived from garlic (Castro et al., 2010), can be measured as a skin VOC following garlic intake (Sato et al., 2020) and suggested to have antioxidant effects in combination with other organosulfur compounds (Fanelli et al., 1998). However, the implication of the decrease in allyl methyl sulphide by intake of NZBC powder is not clear, but may affect body odor.

Five skin VOCs (4 with moderate and 1 with large effect size) showed a trend for change with intake of NZBC powder. Isovaleraldehyde, also known as 3-methylbutanal, has been shown in vitro that its formation is by interaction between human leucocyte antigen and skin microflora, and it suggested to contribute body odor (Savelev et al., 2008). Isovaleraldehyde was decreased by intake of NZBC powder. A trend for a decrease was also observed for hexanal. In general, hexanal is a product of lipid oxidation (Frankel, 1980) and the skin VOC hexanal is the consequence of oxidation of human skin lipids. Hexanal is also taken as an oxidative stress marker in exhaled breath VOC in workers exposed to silica (Jalali et al., 2016). Therefore, it is possible that the intake of NZBC powder reduced lipid peroxidation of fatty acids in the sebaceous glands. 2-pentanone was reduced with intake of NZBC powder and just as benzaldehyde also a VOC released by muscle cells (Mochalski et al., 2014). The implications of a decrease in isovaleraldehyde, hexanal, and 2-pentanone by intake of NZBC powder are not clear. Two skin VOCs, i.e. heptanoic acid and γ nonanolactone, showed a trend for an increase by intake of NZBC powder, with the implications also not clear.

The present study did not employ dietary control or a compulsory wash-out period with restricted intake of habitual polyphenols. Participants in the present study were allowed to adhere to their normal hygienic procedures except for 48 h in which they were instructed not to use hot water and personal care products on the skin sampling site. This is the first study to show that even without dietary control or a wash-out period for habitual polyphenol intake that VOC emission by skin can be affected by intake of an anthocyanin-rich berry powder, indicating an effect of the NZBC powder supplement. However, it needs to be noted that the blackcurrant powder also contained vitamin C which may have affected the emission rate of the skin VOCs. It is also possible that the increase in some VOCs were due to volatile compounds present in the NZBC powder (Liu et al., 2018; Marsol-Vall et al., 2018). In addition, the participants in the present study were supplemented for 7 days with the same amount of NZBC powder which could potentially have affected the observed variation in change of some skin VOCs. In addition, we can also not exclude an effect of the final intake of NZBC powder 2 hours before VOC emission measurements. Dietary control and use of specific anthocyanin-rich extracts may be required for standardization of future studies to isolate the anthocyanin-effects on the emission rate of skin VOCs It may be of interest as well that future studies would address potential sex differences for the response by intake of NZBC powder on VOCs. Our study used a small convenience sample with only 9 males and 5 females. Finally, this study examined a large number of VOCs without an explicit hypothesis for change. However, the presence of the large effect sizes seems to suggest an absence of type I error, and therefore does not necessarily require a multiple comparison analysis with alpha-adjustment (Bender and Lange, 1999; Ranganathan et al., 2016). Finally, it would be of interest to examine dose-response effects of anthocyanin intake on the emission rate of skin VOCs whether more VOCs would respond with large effect size, in addition to more VOCs providing significant changes.

It is concluded that the volatile profile of skin emissions was affected by the intake of anthocyanin-rich New Zealand blackcurrant powder. Future work may address the link between anthocyanin intake by berry consumption, anthocyanin-derived metabolites and VOCs as biomarkers for health, disease and body odor.

Funding sources: Sujon New Zealand blackcurrant powder was provided by Gibb Holdings (Nelson) Limited (New Zealand).

Acknowledgement: We thank William Floyd, advisor to the Japan Blackcurrant Association for bringing attention to the issue of body odor in older adults.

Declaration of interest: none

Financial support: This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

Author contributions: Mark Willems: Conceptualization, Investigation, Writing – Original draft, Project Administration; Michihito Todaka: Formal analysis; Milena Banic: Writing – Original draft, Matthew Cook: Formal analysis; Yoshika Sekine: Conceptualization, Formal analysis, Resources, Writing – Review & Editing, Project Administration.

References

Agapiou A, Mikedi K, Karma S, Giotaki ZK, Kolostoumbis D, Papageorgiou C et al. Physiology and biochemistry of human subjects during entrapment. J Breath Res 2013;7(1):016004. https://doi.org/10.1088/1752-7155/7/1/016004.

Ajiboye TO, Habibu RS, Saidu K, Haliru FZ, Ajiboye HO, Aliyu NO et al. Involvement of oxidative stress in protocatechuic acid-mediated bacterial lethality. Microbiology open 2017;6(4). https://doi.org/10.1002/mbo3.472.

An R, Wilms E, Smolinska A, Hermes GDA, Masclee AAM, de Vos P et al. Sugar Beet Pectin Supplementation Did Not Alter Profiles of Fecal Microbiota and Exhaled Breath in Healthy Young Adults and Healthy Elderly. Nutrients 2019;11(9),pii: E2193.

https://doi.org/10.3390/nu11092193.

Aura AM, Martin-Lopez P, O'Leary KA, Williamson G, Oksman-Caldentey KM, Poutanen K, Santos-Buelga C. In vitro metabolism of anthocyanins by gut microflora. Eur J Nutr 2005;44(3):133-42. https://doi.org/10.1007/s00394-004-0502-2.

Baranska A, Tigchelaar E, Smolinska A, Dallinga JW, Moonen EJ, Dekens JA et al. Profile of volatile organic compounds in exhaled breath changes as a result of gluten-free diet. J Breath Res 2013;7(3):03b7104. https://doi.org/10.1088/1752-7155/7/3/037104.

Bender R, Lange S. Multiple test procedures other than Bonferroni's deserve wider use. BMJ 1999;318(7183):600-1. https://doi.org/10.1136/bmj.318.7183.600a.

Bernier UR, Kline DL, Barnard DR, Schreck CE, Yost RA. Analysis of human skin emanations by gas chromatography/mass spectrometry. 2. Identification of volatile compounds that are candidate attractants for the yellow fever mosquito (Aedes aegypti). Anal Chem 2000;72(4):747-56.

Bond A, Greenwood R, Lewis S, Corfe B, Sarkar S, O'Toole P et al. Volatile organic compounds emitted from faeces as a biomarker for colorectal cancer. Aliment Pharmacol Ther 2019;49(8):1005-12. https://doi.org/10.1111/apt.15140.

Caroprese A, Gabbanini S, Beltramini C, Lucchi E, Valgimigli L. HS-SPME-GC-MS analysis of body odor to test the efficacy of foot deodorant formulations. Skin Res Technol 2009;15(4):503-10. https://doi.org/10.1111/j.1600-0846.2009.00399.x.

Castro C, Lorenzo AG, González A, Cruzado M. Garlic components inhibit angiotensin IIinduced cell-cycle progression and migration: Involvement of cell-cycle inhibitor p27(Kip1) and mitogen-activated protein kinase. Mol Nutr Food Res 2010;54(6):781-7.

https://doi.org/10.1002/mnfr.200900108.

Cohen J (1988). Statistical Power Analysis for the Behavioral Sciences. Routledge.

Curran-Everett D, Benos DJ. Guidelines for reporting statistics in journals published by the

American Physiological Society. Am J Physiol Regul Integr Comp Physiol

2004;287(2):R247-9. https://doi.org/10.1152/ajpregu.00346.2004.

De Ferrars RM, Czank C, Zhang Q, Botting NP, Kroon PA, Cassidy A et al. The pharmacokinetics of anthocyanins and their metabolites in humans. Br J Pharmacol 2014;171(13):3268-82. https://doi.org/10.1111/bph.12676.

De Lacy Costello B, Amann A, Al-Kateb H, Flynn C, Filipiak W, Khalid T et al. A review of the volatiles from the healthy human body. J Breath Res 2014;8(1):014001. https://doi.org/10.1088/1752-7155/8/1/014001.

Działo M, Mierziak J, Korzun U, Preisner M, Szopa J, Kulma A. The Potential of Plant Phenolics in Prevention and Therapy of Skin Disorders. Int J Mol Sci 2016;17(2):160. https://doi.org/10.3390/ijms17020160.

Fanelli SL, Castro GD, de Toranzo EG, Castro JA. Mechanisms of the preventive properties of some garlic components in the carbon tetrachloride-promoted oxidative stress. Diallyl sulfide; diallyl disulfide; allyl mercaptan and allyl methyl sulfide. Res Commun Mol Pathol Pharmacol 1998;102(2):163-74.

Fialová J, Roberts SC, Havlíček J. Consumption of garlic positively affects hedonic perception of axillary body odour. Appetite 2016;97:8-15.

perception of annualy couly outful reported 2010,5710

https://doi.org/10.1016/j.appet.2015.11.001.

Frankel EN. Lipid oxidation. Prog Lipid Res 1980;19(1-2):1-22

Gallagher M, Wysocki CJ, Leyden JJ, Spielman AI, Sun X, Preti G. Analyses of volatile organic compounds from human skin. Br J Dermatol 2008;159(4):780-91.

https://doi.org/10.1111/j.1365-2133.2008.08748.x.

Hair R, Sakaki JR, Chun OK. Anthocyanins, Microbiome and Health Benefits in Aging. Molecules 2021;26(3):535. https://doi.org/10.3390/molecules26030537.

Harshman SW, Geier BA, Fan M, Rinehardt S, Watts BS, Drummond LA et al. The

identification of hypoxia biomarkers from exhaled breath under normobaric conditions. J

Breath Res 2015;9(4):047103. https://doi.org/10.1088/1752-7155/9/4/047103.

Haze S, Gozu Y, Nakamura S, Kohno Y, Sawano K, Ohta H et al. 2-Nonenal newly found in human body odor tends to increase with aging. J Invest Dermatol 2001;116(4):520-4.

https://doi.org/10.1046/j.0022-202x.2001.01287.x.

Ishino K, Wakita C, Shibata T, Toyokuni S, Machida S, Matsuda S et al. Lipid peroxidation generates body odor component trans-2-nonenal covalently bound to protein in vivo. J Biol Chem 2010:285(20):15302-13. https://doi.org/10.1074/jbc.M109.068023.

Jalali M, Zare Sakhvidi MJ, Bahrami A, Berijani N, Mahjub H. Oxidative Stress Biomarkers in Exhaled Breath of Workers Exposed to Crystalline Silica Dust by SPME-GC-MS. J Res Health Sci 2016;16(3):153-61.

Jha SK. Characterization of human body odor and identification of aldehydes using chemical sensor Rev Anal Chem 2017;36(2), 20160028. https://doi.org/10.1515/revac-2016-0028. Kapasakalidis PG, Rastall RA, Gordon MH. Extraction of polyphenols from processed black currant (Ribes nigrum L.) residues. J Agric Food Chem 2006;31;54(11):4016-21. https://doi.org/10.1021/jf0529991.

Kimura K, Sekine Y, Furukawa S, Takahashi M, Oikawa D. Measurement of 2-nonenal and diacetyl emanating from human skin surface employing passive flux sampler-GCMS system. J Chromatogr B Analyt Technol Biomed Life Sci 2016;1028:181-5.

https://doi.org/10.1016/j.jchromb.2016.06.021.

Krilaviciute A, Leja M, Kopp-Schneider A, Barash O, Khatib S, Amal H et al. Associations of diet and lifestyle factors with common volatile organic compounds in exhaled breath of average-risk individuals. J Breath Res 2019;13(2):026006. https://doi.org/10.1088/1752-7163/aaf3dc.

Lacey JRN, Kidel C, van der Kaaij JM, Brinkman P, Gilbert-Kawai ET, Grocott MPW et al. The Smell of Hypoxia: using an electronic nose at altitude and proof of concept of its role in the prediction and diagnosis of acute mountain sickness. Physiol Rep 2018:6(17):e13854. https://doi.org/10.14814/phy2.13854. Lam TH, Verzotto D, Brahma P, Ng AHQ, Hu P, Schnell D et al. Understanding the microbial basis of body odor in pre-pubescent children and teenagers. Microbiome 2018;6(1):213. https://doi.org/10.1186/s40168-018-0588-z.

Landrigan PJ, Kreiss K, Xintaras C, Feldman RG, Heath CW Jr. Clinical epidemiology of occupational neurotoxic disease. Neurobehav Toxicol 1980;2(1):43-8.

Lee SG, Vance TM, Nam TG, Kim DO, Koo SI, Chun OK. Contribution of Anthocyanin Composition to Total Antioxidant Capacity of Berries. Plant Foods Hum Nutr 2015;70(4):427-32. https://doi.org/10.1007/s11130-015-0514-5.

Lila MA, Burton-Freeman B, Grace M, Kalt W. Unraveling Anthocyanin Bioavailability for Human Health. Annu Rev Food Sci Technol 2016;7:375-93. https://doi.org/10.1146/annurevfood-041715-033346.

Liu Y, Wang S, Ren J, Yuan G, Li Y, Zhang B, Zhu B. Data on free and bound volatile compounds in six *Ribes nigrum* L. blackcurrant cultivars. Data Brief 2018;17:926-37. https://doi.org/10.1016/j.dib.2018.01.090.

Lyall KA, Hurst SM, Cooney J, Jensen D, Lo K, Hurst RD et al. Short-term blackcurrant extract consumption modulates exercise-induced oxidative stress and lipopolysaccharidestimulated inflammatory responses. Am J Physiol Regul Integr Comp Physiol 2009;297(1):R70-81. https://doi.org/10.1152/ajpregu.90740.2008.

Mack CI, Egert B, Liberto E, Weinert CH, Bub A, Hoffmann I et al. Robust Markers of Coffee Consumption Identified Among the Volatile Organic Compounds in Human Urine. Mol Nutr Food Res 2019;63(10):e1801060. https://doi.org/10.1002/mnfr.201801060. Marsol-Vall A, Kortesniemi M, Karhu ST, Kallio H, Yang B. Profiles of Volatile Compoundsin Blackcurrant (Ribes nigrum) Cultivars with a Special Focus on the Influences of Growth Latitude and Weather Conditions. J Agric Food Chem 2018;66(28):7485-7495. https://doi.org/10.1021/acs.jafc.8b02070. Mazzatenta A, Pokorski M, Sartucci F, Domenici L, Di Giulio C. Volatile organic compounds (VOCs) fingerprint of Alzheimer's disease. Respir Physiol Neurobiol 2015;209:81-4. https://doi.org/10.1016/j.resp.2014.10.001.

Mochalski P, Al-Zoairy R, Niederwanger A, Unterkofler K, Amann A. Quantitative analysis of volatile organic compounds released and consumed by rat L6 skeletal muscle cells in vitro. J Breath Res 2014;8(4):046003. https://doi.org/10.1088/1752-7155/8/4/046003.

Mochalski P, Unterkofler K. Quantification of selected volatile organic compounds in human urine by gas chromatography selective reagent ionization time of flight mass spectrometry (GC-SRI-TOF-MS) coupled with head-space solid-phase microextraction (HS-SPME). Analyst 2016;141(15):4796-803. https://doi.org/10.1039/c6an00825a.

Mozdiak E, Wicaksono AN, Covington JA, Arasaradnam RP. Colorectal cancer and adenoma screening using urinary volatile organic compound (VOC) detection: early results from a single-centre bowel screening population (UK BCSP). Tech Coloproctol 2019:23(4):343-51.

https://doi.org/10.1007/s10151-019-01963-6.

Neveu V, Perez-Jiménez J, Vos F, Crespy V, du Chaffaut L, Mennen L et al. Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. Database (Oxford) 2010;2010:bap024. https://doi.org/10.1093/database/bap024.

Oliveira I, Malheiro R, Meyer AS, Pereira JA, Gonçalves B. Application of chemometric tools for the comparison of volatile profile from raw and roasted regional and foreign almond cultivars (*Prunus dulcis*). J Food Sci Technol 2019;56(8):3764-76.

https://doi.org/10.1007/s13197-019-03847-x.

Ozeki C, Moro O. A study of the suppression of body odour in elderly subjects by anti-fungal agents. Int J Cosmet Sci 2016;38(3):312-8. https://doi.org/10.1111/ics.12295.

Piwowarek K, Lipińska E, Hać-Szymańczuk E, Kieliszek M, Ścibisz I. Propionibacterium spp.-source of propionic acid, vitamin B12, and other metabolites important for the industry.
Appl Microbiol Biotechnol 2018;102(2):515-38. https://doi.org/10.1007/s00253-017-8616-7.
Ranganathan P, Pramesh CS, Buyse M. Common pitfalls in statistical analysis: The perils of multiple testing. Perspect Clin Res 2016;7(2):106-7. https://doi.org/10.4103/2229-3485.179436.

Santos PM, Del Nogal Sánchez M, Pozas ÁPC, Pavón JLP, Cordero BM. Determination of ketones and ethyl acetate-a preliminary study for the discrimination of patients with lung cancer. Anal Bioanal Chem 2017;409(24):5689-96. https://doi.org/10.1007/s00216-017-0508-2.

Saric S, Notay M, Sivamani RK. Green Tea and Other Tea Polyphenols: Effects on Sebum Production and Acne Vulgaris. Antioxidants (Basel) 2016;6(1),pii:E2.

https://doi.org/10.3390/antiox6010002.

Sato S, Sekine Y, Kakumu Y, Hiramoto T. Measurement of diallyl disulfide and allyl methyl sulfide emanating from human skin surface and influence of ingestion of grilled garlic. Sci Rep 2020; 10(1):465. https://doi.org/10.1038/s41598-019-57258-1.

Savelev SU, Antony-Babu S, Roberts SC, Wang H, Clare AS, Gosling LM et al.. Individual variation in 3-methylbutanal: a putative link between human leukocyte antigen and skin microflora. J Chem Ecol 2008;34(9):1253-7. https://doi.org/10.1007/s10886-008-9524-1. Sekine Y, Sato, S, Kimura K, Sato H, Nakai S, Yanagisawa Y. Detection of tobacco smoke emanating from human skin surface of smokers employing passive flux sampler - GCMS system. J. Chromatogr. B 2018;1092:394-401.

https://doi.org/10.1016/j.jchromb.2018.06.038.

Speer H, D'Cunha NM, Alexopoulos NI, McKune AJ, Naumovski N. Anthocyanins and Human Health – A Focus on Oxidative Stress, Inflammation and Disease. Antioxidants (Basel) 2020; 9(5):366. https://doi.org/10.3390/antiox9050366.

Timm CM, Lloyd EP, Egan A, Mariner R, Karig D. Direct Growth of Bacteria in Headspace Vials Allows for Screening of Volatiles by Gas Chromotography Mass Spectrometry Front Microbiol 2018;9:491. https://doi.org/10.3389/fmicb.2018.00491.

Umezawa K, Sekine Y, Kimura K, Asai S, Saito T, Inokuchia S et al. Emanation of fenitrothion from skin surface of patient who attempted to suicide by acute poisoning. Official J Japan Soc Lab Med 2018;66(9):949-56.

Van Berkel JJ, Dallinga JW, Möller GM, Godschalk RW, Moonen EJ, Wouters EF et al. A profile of volatile organic compounds in breath discriminates COPD patients from controls. Respir Med 2010;104(4):557-63. https://doi.org/10.1016/j.rmed.2009.10.018.

Wang Y, Kao MS, Yu J, Huang S, Marito S, Gallo RL et al. A Precision Microbiome Approach Using Sucrose for Selective Augmentation of Staphylococcus epidermidis Fermentation against Propionibacterium acnes. Int J Mol Sci 2016;17(11). pii: E1870. https://doi.org/10.3390/ijms17111870.

Willems MET, Myers SD, Gault ML, Cook MD. Beneficial physiological effects with intake of blackcurrant intake in endurance athletes. Int J Sport Nutr Exerc Metab 2015;25(4):367-374. https://doi.org/10.1123/ijsnem.2014-0233.

Willems MET, Dos Santos Silva J, Cook MD, Blacker SD. Beneficial effects on fasting insulin and postprandial responses through 7-day intake of New Zealand blackcurrant powder. Functional Foods in Health and Disease 2017;7(7):483-493.

https://doi.org/10.31989/ffhd.v7i7.335.

Zhang ZM, Cai JJ, Ruan GH, Li GK. The study of fingerprint characteristics of the emanations from human arm skin using the original sampling system by SPME-GC/MS. J

Accepted version, 21/03/2021, Journal of Dietary Supplements

Chromatogr B Analyt Technol Biomed Life Sci 2005;822(1-2):244-52.

https://doi.org/10.1016/j.jchromb.2005.06.026.

Table legends

Table 1. Volatile organic compounds emitted by the skin that were changed (P<0.05) or showed a trend for change $(0.05 \ge P \le 0.1)$ by intake of New Zealand blackcurrant powder for 7 days in middle-aged and older adults (male: n=9, female: n=5, age: 55±5 yr). + and - indicates an increase and decrease of the emission rate, respectively.

Volatile organic compounds	CAS number	Direction of change	Effect size	P-value
		by blackcurrant		
Aldehydes				
2-Nonenal	2463-53-8	(-)	-0.87	0.030
Isovaleraldehyde	590-86-3	(-)	-0.75	0.093
Benzaldehyde	100-52-7	(-)	-0.96	0.040
Hexanal	66-25-1	(-)	-0.81	0.062
Ketones				
2-Hexanone	591-78-6	(-)	-0.86	0.031
6-Methyl-5-hepten-2-one	110-93-0	(-)	-1.23	0.005
2-Pentanone	107-87-9	(-)	-0.71	0.088
Carboxylic acid				
Acetic acid	64-19-7	(-)	-1.26	0.006
Heptanoic acid	111-14-8	(+)	0.73	0.065
Cyclic esters				
γ-octanolactone	104-50-7	(+)	0.98	0.023
γ-decanolactone	706-14-9	(+)	1.11	0.015
γ-nonanolactone	104-61-0	(+)	0.62	0.063
Sulphur containing				
compounds				
Allyl methyl sulfide	10152-76-8	(-)	-1.01	0.015

Table 2. Volatile organic compounds emitted by the skin (dermal emission flux in $ng \cdot cm^{-2} \cdot h^{-1}$) that were not changed (P>0.05) by 7-day intake of New Zealand blackcurrant powder in middle-aged and older adults ((male: n=9, female: n=5, age: 55±5 yr). Cohen's d effect sizes were trivial (d < 0.2), small (d = 0.2-0.49) or moderate (d = 0.5-0.79). Data are mean± SD.

Volatile organic	CAS number	Control	New Zealand	Effect size	P-value
compounds		Control	blackcurrant	(Cohen's d	i vulue
Alcohols				(
1-Propanol	71-23-8	2.56 ± 1.81	2.74 ± 1.63	0.11	0.37
1-Butanol	71-36-3	5.27 ± 2.69	4.84 ± 2.72	-0.16	0.68
1-Pentanol	71-41-0	7.63 + 4.62	6.52 + 4.70	-0.24	0.56
1-Hexanol	111-27-3	3.77 ± 1.70	3.38 ± 1.65	-0.23	0.58
1-Heptanol	111-70-6	1.97 ± 1.21	2.97 ± 2.07	0.59	0.16
1-Octanol	111-87-5	4.98 ± 3.63	6.68 ± 6.03	0.34	0.39
1-Nonenal	143-08-8	3.56 ± 1.82	4.64 ± 3.06	0.43	0.31
1-Decanol	112-30-1	6.99 + 4.73	7.16 + 3.23	0.04	0.91
2-Ethyl-1-hexanol	104-76-7	0.07 ± 0.12	0.12 ± 0.20	0.30	0.19
Acetoin	513-86-0	2.40 + 2.26	1.73 ± 1.15	-0.37	0.37
Aldehvdes					
Acetaldehyde	75-07-0	1.78 ± 0.13	1.84 ± 0.12	0.48	0.28
Propanal	123-38-6	2.35 ± 0.91	2.54 ± 0.67	0.24	0.59
Butanal	123-72-8	1.42 ± 1.08	1.21 ± 0.47	-0.25	0.45
Valeraldehvde	110-62-3	1.05 ± 0.64	0.93 ± 0.42	-0.22	0.61
Heptanal	111-71-7	5.52 ± 2.85	5.36 ± 3.16	-0.05	0.90
Octanal	124-13-0	2.25 ± 1.52	2.59 ± 1.64	0.22	0.60
Nonanal	124-19-6	6.24 ± 3.28	6.83 ± 5.11	0.14	0.74
Decanal	112-31-2	4.11 ± 3.45	5.88 ± 4.57	0.44	0.29
2-Hexenal	505-57-7	2.38 ± 1.18	1.96 ± 2.07	-0.25	0.53
Vanillin	121-33-5	1.20 ± 0.90	0.90 ± 0.85	-0.34	0.36
Carboxylic acids					
Propanoic acid	79-09-4	4.55 ± 3.08	4.44 ± 2.46	-0.04	0.91
Butanoic acid	107-92-6	3.60 ± 2.28	4.35 ± 1.72	0.37	0.39
Isovaleric acid	503-74-2	1.62 ± 0.94	1.40 ± 0.90	0.24	0.59
Valeric acid	109-52-4	1.32 ± 0.95	0.81 ± 0.61	-0.64	0.23
Hexanoic acid	142-62-1	1.96 ± 1.10	1.53 ± 0.89	-0.43	0.26
Octanoic acid	124-07-2	1.27 ± 1.12	1.31 ± 0.89	0.04	0.91
Nonanoic acid	112-05-0	3.65 ± 2.09	3.60 ± 2.19	-0.02	0.85
Decanoic acid	334-48-5	1.61 ± 0.95	1.96 ± 1.89	0.23	0.58
Ketones					
Acetone	67-64-1	1.42 ± 0.54	1.47 ± 0.43	0.10	0.59
2-Butanone	78-93-3	1.81 ± 0.75	1.74 ± 0.60	-0.10	0.83
2-Heptanone	110-43-0	1.43 ± 1.05	1.24 ± 1.16	-0.17	0.69
2-Octanone	111-13-7	1.42 ± 1.04	1.54 ± 1.03	0.12	0.72
2-Nonanone	821-55-6	1.59 ± 1.31	2.23 ± 2.18	0.36	0.35
2-Decanone	693-54-9	2.39 ± 1.35	2.50 ± 2.21	0.06	0.88
2-Undecanone	112-12-9	3.27 ± 1.77	2.59 ± 1.94	-0.37	0.23
2-Dodecanone	6175-49-1	0.97 ± 0.57	0.74 ± 0.62	-0.39	0.32
2-Tridecanone	593-08-8	1.76 ± 0.99	1.16 ± 1.21	-0.54	0.72
2-Tetradecanone	2345-27-9	1.76 ± 1.31	1.43 ± 0.93	-0.29	0.36
2-Pentadecanone	2345-28-0	2.67 ± 1.64	2.05 ± 1.23	-0.43	0.35
Acid esters					
Ethyl acetate	141-78-6	3.47 ± 2.27	5.12 ± 5.23	0.41	0.33
Benzyl acetate	140-11-4	0.11 ± 0.08	0.09 ± 0.08	-0.25	0.50
cis-3-Hexenyl acetate	3681-71-8	1.89 ± 1.89	1.52 ± 0.66	-0.26	0.38
Benzyl hydrocarbons					

Accepted version, 21/03/2021, Journal of Dietary Supplements

Ethyl benzene	100-41-4	0.33 ± 0.72	0.15 ± 0.12	-0.35	0.32
m,p-Xylene	106-42-3 108-	0.28 ± 0.77	0.07 ± 0.06	-0.39	0.34
	38-3				
o-Xylene	95-47-6	0.20 ± 0.16	0.24 ± 0.16	0.25	0.55
Styrene	100-42-5	0.29 ± 0.44	0.14 ± 0.20	-0.44	0.28
Halogen containing					
volatile					
p-Dichlorobenzene	106-46-7	0.04 ± 0.03	0.03 ± 0.04	-0.28	0.79
Indoles					
Indole	120-72-9	0.28 ± 0.29	0.33 ± 0.20	0.20	0.59
Skatole	83-34-1	0.09 ± 0.06	0.07 ± 0.04	0.20	0.34
Terpenes					
α-Pinene	2437-95-8	0.45 ± 0.46	0.35 ± 0.32	-0.25	0.56
β-Pinene	127-91-3	0.26 ± 0.17	0.23 ± 0.15	-0.19	0.46
d,l-Limonene	5989-27-5	0.51 ± 0.64	0.27 ± 0.22	-0.50	0.20
	5989-54-8				
<u> </u>					
Sulfur compounds					
Methyl mercaptan	74-93-1	2.40 ± 1.70	3.34 ± 3.07	0.38	0.25
Diallyl disulfide	2179-57-9	1.67 ± 1.23	1.49 ± 0.95	-0.16	0.68
Cyclic esters					
γ-Hexanolactone	695-06-7	0.47 ± 0.41	0.46 ± 0.54	-0.02	0.91
γ-Heptanolactone	105-21-5	0.26 ± 0.20	0.54 ± 0.86	0.45	0.35
γ-Undecanolactone	104-67-6	0.25 ± 0.19	0.21 ± 0.18	-0.22	0.61
Other					
Geosmin	19700-21-1	0.19 ± 0.11	0.20 ± 0.17	0.07	0.80
1,3-Butanediol	107-88-0	2.43 ± 1.11	2.84 ± 1.21	0.35	0.37
Phenol	108-95-2	0.64 ± 0.46	0.57 ± 0.33	0.18	0.63
Butylated hydroxytoluene	128-37-0	0.67 ± 0.27	$\overline{0.85\pm0.58}$	0.40	0.34

Figure legends

Figure 1. Box-and-whisker plot for emission flux of the skin volatile organic compound 2nonenal (A), benzaldehyde (B), 6-methyl-5-hepten-2-one (C), 2-hexanone (D), acetic acid (E) and allyl methyl sulfide (F). New Zealand blackcurrant (NZBC) powder. Intake of New NZBC powder was for 7 days. *, emission flux was lower with NZBC powder (P < 0.05). Data of 14 participants.



Figure 2. Box-and-whisker plot for emission flux of the skin volatile organic compound γ -octanolactone (A) and γ -decanolactone (B). Intake of New Zealand blackcurrant powder

(NZBC) was for 7 days. *, emission flux was higher with NZBC powder (P < 0.05). Data of 14 participants.

