

Note

NMR spectral analysis of some spirostanoids[†]

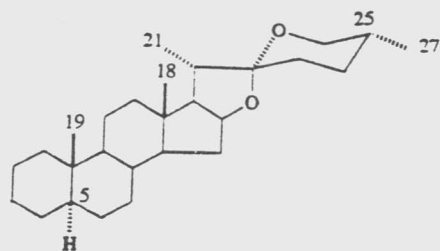
Pawan K Agrawal

Central Institute of Medicinal and Aromatic plants
Lucknow-226 015, India

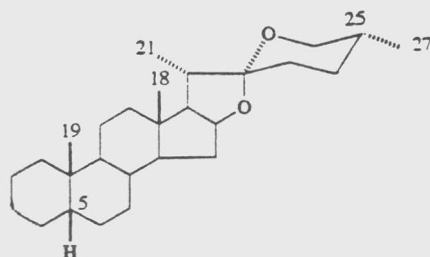
Received 1 December 1997;
accepted 12 February 1998

An approach based upon NOE between H₃-19 and H-5 has been proposed for the establishment of 5β- and 5α- stereochemistry at A/B ring junction in spirostanoids and the recently reported ¹³C NMR shielding data for several steroidal enones has been analyzed.

As a part of our continuing effort to understand the ¹³C NMR spectral behaviour of steroidal saponinins and steroidal saponins¹⁻⁹, we have earlier suggested that the ¹³C NMR chemical shift of the C-22 resonance acts as a reporter for the identification of the parent skeletal type¹⁰⁻¹². The variation in parent skeleton generally arises from the configuration at C-5, i.e. the A/B ring junction and the configuration of the methyl group at C-25 as naturally occurring spirostanoids characterized so far, possess B/C *trans*, C/D *trans* and D/E *cis* ring junctions. Depending upon the *trans* and *cis* relationship between H₃-19 and H-5, the spirostanoids have been grouped into 5α (A) and 5β (B) types respectively (Figure 1). Recently, we proposed a correlation between the ¹H and ¹³C NMR chemical shifts of ring F resonances and orientation of the 27-methyl group for the discrimination of 25*R*/25*S* stereochemistry¹³. Despite the fact that ¹³C NMR chemical shifts of C-5, C-7, C-9 and C-19 are quite distinctive for the identification of stereochemistry at A/B ring junction^{11,12}, we are involved in determining additional NMR approaches applicable for ascertaining 5α- and 5β-stereochemistry. In the present communication, we wish to report the



A



B

significance of ¹H-¹H NOE (NOESY) spectroscopy for the determination of stereochemistry at ring junctions as well as analysis of recently reported ¹³C NMR data for several functionalized spirostanoids¹⁴.

In view of our homo- and heteronuclear NMR studies on 25*R*/25*S* epimeric pair of 5β spirostanoids (smilagenin and sarsasapogenin)¹⁵ together with the literature reports¹⁶⁻²⁰ led us to deduce several conclusions: (i) NOE cross peaks are usually observed for equatorial-axial vicinal and 1,3- *syn*-diaxial proton pair, (ii) NOE is absent between vicinal *trans* diaxial protons, (iii) NOE connectivity between H-8/H₃-18, H₃-19; H-9/H-14, H-17; and H₃-18/H-20 are common for B/C *trans*, C/D *trans*, and D/E *cis* ring junctions respectively, (iv) NOE between H₃-19/H-2a, H-4a; and for H₃-19/H-5 are characteristic for A/B *trans* (5α) and *cis* (5β) respectively. Thus, once the ¹H NMR assignments are evident, presence and absence of NOESY cross peak between H₃-19 and H-5 could be utilized for identifying 5β(B) and 5α(A) subgroups of spirostanoids.

In a recent publication¹⁴, DeNinno and McCarthy have reported ¹³C NMR chemical shift

[†] Part 47 in the series, 'NMR Spectral Investigations', for part 46 see ref. 13.

Table I—¹³C NMR chemical shifts for steroidal sapogenins 1-6

Position	25R 3, 11 (oxo) ₂	25R 3-oxo Δ ¹	25R 3,11 (oxo) ₂ Δ ¹	25R 3β-OH- 3-Me Δ ¹	25R 3β-OH 3-Me 11-oxo Δ ¹	25R 1β, 2 β, 3 β-(OH) ₃ 3-Me 11-oxo
	1	2	3	4	5	6
C-1	37.0	158.3	159.3	136.2	136.6	75.2
C-2	37.9	127.4	127.5	132.4	132.4	73.6
C-3	211.4	200.2	199.4	70.6	70.6	74.6
C-4	44.3	39.8	38.0	44.0	43.5	41.3
C-5	46.9	44.3	44.2	39.9	41.8	36.0
C-6	28.2	27.5	27.0	28.0	27.5	27.4
C-7	32.4	31.7	36.9	32.1	32.6	36.7
C-8	35.2	39.0	40.6	35.2	36.9	39.9
C-9	60.7	50.0	59.8	51.1	60.7	67.0
C-10	36.9	35.3	59.7	29.4	37.3	57.3
C-11	209.7	21.0	209.4	21.0	209.8	211.0
C-12	57.6	40.9	57.3	42.1	57.4	57.6
C-13	44.2	40.6	44.3	40.3	44.4	44.3
C-14	55.5	56.2	55.3	56.2	55.7	55.5
C-15	31.2	31.5	32.1	31.7	31.2	32.2
C-16	80.5	80.7	80.4	80.3	80.6	80.6
C-17	63.9	62.2	60.7	62.2	61.4	60.8
C-18	17.2	16.2	17.3	16.7	17.2	17.2
C-19	11.1	13.1	13.7	15.0	15.4	12.7
C-20	41.8	41.6	41.8	41.6	41.5	41.9
C-21	14.2	14.2	14.2	14.3	14.2	14.1
C-22	109.2	109.4	109.2	109.6	109.2	109.2
C-23	31.3	31.4	31.2	31.4	31.2	31.4
C-24	28.7	28.8	28.7	28.8	28.7	28.7
C-25	30.2	30.3	30.2	30.4	30.2	30.2
C-26	66.9	66.9	66.9	66.8	66.9	66.9
C-27	17.1	17.0	17.1	17.3	17.1	17.1

data for several spirostane derivatives, (5 α , 25R)-spirostan-3,11-dione **1**, (5 α , 25R)-spirostan-1-en-3-one **2**, (5 α , 25R)-spirostan-1-en-3,11-dione **3**, (3R, 5 α , 25R)-3-hydroxy-3-methyl-spirost-1-ene **4**, (3R, 5 α , 25R)-3-hydroxy-3-methyl-spirost-1-en-11-one **5**, and (3R, 5 α , 25R) 1,2,3-trihydroxy-3-methyl-spirostan-11-one **6**. The analysis of the ¹³C NMR data of these compounds, was considered worthwhile in view of the reported cholesterol absorption inhibiting activity of diglycoside derived from 11-ketotigogenin²¹. Our earlier studies on steroidal sapogenins¹⁰⁻¹², facilitated the analysis and ¹³C NMR assignments are presented in Table I. The analysis of the ¹³C NMR data led to the several inferences: (i) substituents such as oxo, hydroxyl and olefinic bond, consistent with our studies on steroidal alkaloids²¹ and steroidal sapogenins¹⁰⁻¹², suggest that these modifies the

chemical shifts of carbons occupying α , β , and γ position respectively, (ii) Δ^1 olefinic bond causes, (a) an upfield shift (12 ppm) of C-3 (**1** vs. **3**), (b) downfield shift of C-19 by about 2 ppm, (c) marginal effect (0.3 ppm) on the chemical shift of C-11; (iii) appearance of ring F resonances at almost identical position ± 0.1 ppm in all the compounds (**1-6**) reflects absence of any kind of long-range interactions with 11-oxo group as well as existence of chair conformation of ring-F.

Acknowledgement

The author is thankful to Dr Sushil Kumar, Director, CIMAP for constant encouragement.

References

- 1 Agrawal P K, Singh S B & Thakur R S, *Indian J Pharm Sci*, 46, 1984, 158.

- 2 Schneider H J, Buchheit, U & Agrawal P K, *Tetrahedron*, 40, 1984, 1017.
- 3 Mahmood U, Agrawal P K & Thakur R S, *Phytochemistry*, 24, 1985, 2446.
- 4 Agrawal P K & Thakur R S, *Indian J Chem*, 25B 1986 469.
- 5 Agrawal P K, Mahmood U & Thakur R S, *Heterocycles*, 29, 1989, 1895.
- 6 Uniyal G C, Agrawal P K, Thakur R S & Sati O P, *Phytochemistry*, 29 1990, 937.
- 7 Uniyal G C, Agrawal P K, Sati O P & Thakur R S, *Phytochemistry*, 30, 1990, 1336.
- 8 Uniyal G C, Agrawal P K, Sati O P & Thakur R S, *Phytochemistry*, 30, 1990, 4187.
- 9 Agrawal P K, *Indian J Chem*, 25B, 1996, 278.
- 10 Agrawal P K, Jain D C, Gupta R K & Thakur R S, *Phytochemistry* 24, 1985, 2479.
- 11 Agrawal P K, Jain D C & Pathak A K, *Magn Reson Chem*, 33, 1995, 923.
- 12 Agrawal P K, in *Saponins Used in Food and Agriculture*, edited by W Waller & K Yamasaki (Plenum Press, New York), 1996, p 299.
- 13 Agrawal P K, Morris G A & Bunsawansong D, *Phytochemistry*, 1997, in press.
- 14 DeNinno M P & McCarthy K E, *Tetrahedron* 53, 1997, 11007.
- 15 Agrawal P K, Morris G A & Bunsawansong D, *Magn Reson Chem*, 1997, 441.
- 16 Mimaki Y, Kuroda M, Nakamura O, Sashida Y, Satou T, Koike K & Nikaido T, *Chem Pharm Bull*, 45, 1997, 558.
- 17 Mimaki Y, Kuroda M, Nakamura O & Sashida Y, *J Nat Prod*, 60, 1997, 592.
- 18 Yoshikawa M, Murakami T, Komatsu H, Murakami N, Yamahara, J & Matsuda H, *Chem Pharm Bull*, 45, 1997, 81.
- 19 Mimaki Y, Kameyama A, Kuroda M, Sashida Y, Hirano T, Oka K, Koike K & Nikaido T, *Phytochemistry*, 44, 1997, 305.
- 20 McCarthy P A, DeNinno M P, Morehouse L A, Chandler C E, Bangerter F W, Wilson T C, Urban F J, Walinsky S W, Cosgrove P G, Duplantier K C, Etienne J B, Fowler M A, Lambert J F, O'Donnell J P, Pezzullo S L, Watson Jr H A, Wilkins R W, Zaccaro L M & Zawistoski M P, *J Med Chem*, 39, 1996, 1935.
- 21 Agrawal P K, Srivastava S K & Gaffield W, *Alkaloids: Chemical and Biological Perspectives*, Vol 7, edited by S W Pelletier (Springer, New York), 1991, p 49.