

Rapid Communication

Novel synthesis of *D-ribo*-[2*S*,3*S*,4*R*]-2-*N*-palmitoyl-2-aminoheptadecane-1,3,4-triol[†]

Samiksha Katiyar, S Paul & S N Suryawanshi*

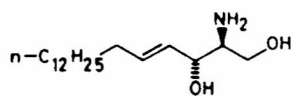
Division of Medicinal Chemistry, Central Drug Research Institute, Lucknow 226 001, India

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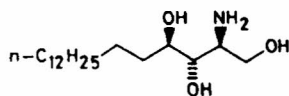
Novel synthesis of *D-ribo*-[2*S*,3*S*,4*R*]-2-*N*-palmitoyl-2-aminoheptadecane from *D-mannose* is described.

D-erythro-Sphingosine [(2*S*,3*R*,4*E*)-2-amino-1,3-diol] **1** and phytosphingosine [(2*S*,3*S*,4*R*)-2-amino-1,3,4-triol] **2** are the major backbone components of glycosphingolipids, which play important roles in biological processes on cell surfaces¹. Phytosphingosine was first detected in fungi², plants³, yeasts⁴ and other organisms⁵. Later, phytosphingosine was also detected in vertebrate species⁶ and sea organisms^{7,8}. It is also present in mammalian tissues, for example in kidney⁹, liver¹⁰, uterus¹¹, intestine¹², skin¹³, blood plasma¹⁴ and thyrocytes¹⁵. Sphingosines have attracted considerable interest as potent inhibitors of protein kinase C, an essential enzyme in cell regulation and signal transduction¹⁶. Various synthetic approaches to optically active phytosphingosine have been reported¹⁷. In this communication we report the synthesis of a phytosphingosine analog from *D-mannose*.

Diacetone mannose **3**, easily available from *D-mannose* on Wittig olefination with dodecyltriphenylphosphorane (dodecyltriphenylphosphonium

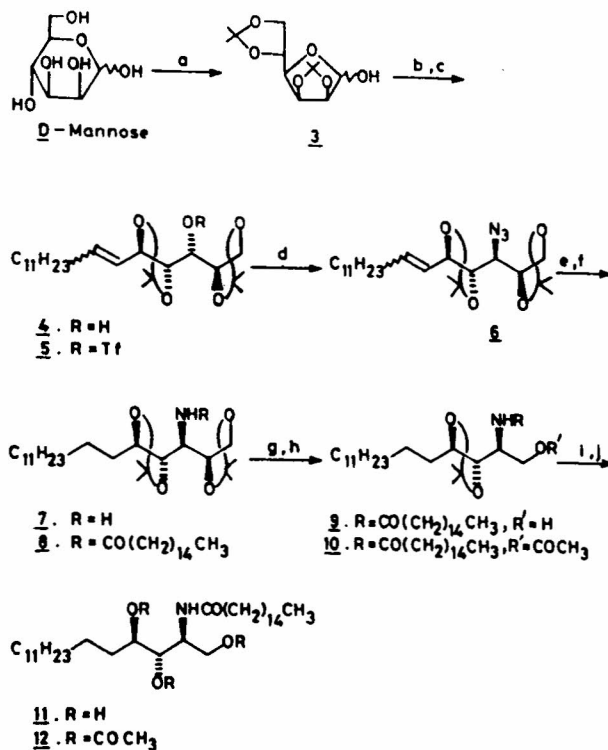


D-erythro-sphingosine **1**



Phytosphingosine **2**

bromide, KOBut, toluene, 16 hr) furnished **4** (cf. Scheme I) as an *E/Z* (40:60) mixture in 80% yield. Reaction of olefin **4** with 1.2 equivalents of Tf₂O and pyridine at 0 °C furnished the triflate **5**, which on further treatment with NaN₃ in DMF furnished azide **6** in 80% yield. IR absorption of **6** at 2100 cm⁻¹ supported its assigned structure. Azide **6** on hydrogenation (H₂, 10% Pd-C, EtOAc at 50 psi) furnished the amine **7** in 65% yield. Palmitoylation of amine **7** (*p*-nitrophenyl palmitate, Py, 80 °C) furnished the palmitate **8** in quantitative yield. The deprotection/oxidation of terminal isopropylidene group in **8** with periodic acid in ethyl acetate followed by hydride reduction (NaBH₄, ethanol) furnished the ceramide **9** in 60% yield. Deprotection of **9** with 70% aqueous acetic acid furnished the



Reagents : a) $\text{C}_{12}\text{H}_{25}\text{CH}=\text{CHPh}_3\text{P}^+\text{Br}^-$, t - KOBut, Toluene; b) Tf₂O, Pyridine; c) NaN₃, DMF; d) H₂, 10% Pd-C, EtOAc; e) CH₃(CH₂)₁₄COO-C₆H₄-NO₂, pyridine; f) H₂, 10% Pd-C, EtOAc; g) NaBH₄, EtOH; h) 70% Aq, AcOH; i) Ac₂O, Py.

Scheme I

[†] CDRI Communication No. 5769

ceramide 11¹⁸ in quantitative yield. The ceramide 11 on acetylation (Ac₂O, Py) furnished the triacetate 12 (Scheme I) in quantitative yield.

In conclusion, the C-17 analog of phytoceramide has been efficiently synthesised from D-mannose.

Acknowledgement

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References

- 1 Kaufer J M & Hakomori S, *Handbook of lipid research*, Vol. 3. *Sphingolipid biochemistry*, (Plenum Press, New York) 1983.
- 2 Oda T, *J Pharm Soc Jpn*, 72, 1952, 142.
- 3 Carter H E, Clemer W D, Lands W D M, Mueller K L & Tomizava H H, *J Biol Chem*, 206 1954, 613.
- 4 Thorpe S R & Swely C C, *Biochemistry*, 6, 1967, 887.
- 5 Dearborn D G, Smith S & Korn E D, *J Biol Chem*, 251, 1976, 2996.
- 6 Nishimura K, *Comp Biochem Physiol A: Comp Biochem*, 86, 1987, 149.
- 7 Yu-Teh L, Hirabayashi Y, DeGasperi R, Yu R K, Ariga T Koerner T A W & Li S C, *Biol Chem*, 259, 1984, 8980.
- 8 Smirnova G P, Kochetkov N K & Sadovskaya V L, *Biochem Biophys Acta*, 920, 1987, 47; Kubo H, Jiang G J, Jrie A, Morita M, Matsubara T & Hoshi M, *J Biochem*, 7, 1968, 2296
- 9 Karison K A, Samuelsson B E & Steen G O, *Acta Chem Scand*, 22, 1968, 1361; Carter H E & Hirschberg, *Biochemistry*, 7, 1968, 2296.
- 10 Barenholz Y & Gatt S, *Biochem Biophys Res Commun*, 27, 1967, 319.
- 11 Takamatsu K, Mikami M, Kiguchi K, Nozava S & Iwamori M, *Biochim Biophys Acta*, 1165, 1992, 177.
- 12 Okabe K, Keenan R W & Schmidt G, *Biochim Biophys Res Commun*, 31, 1968, 137; Svennerholm L, Fredman P, Mansson J E, Nilsson O & Holmgren J, *J Adv Exp Med Biol*, 152, 1982, 333.
- 13 Wertz P W, Miethke M C, Long S A, Stauss J S & Downing D T, *J Invest Dermatol*, 84, 1985, 410.
- 14 Vance D E & Sweely C G, *Lipid Res*, 8, 1967, 621.
- 15 Bouchon B, Portoukalian J, Origazzi J & Bornet H, *Biochem Biophys Res Commun*, 143, 1987, 827.
- 16 (a) Gigg R & Gigg J, *J Chem Soc (C)*, 1966, 1876.
(b) Gigg J, Gigg R & Warren C D, *J Chem Soc (C)*, 1966, 1872.
(c) Birk R, Jung K H & Schmidt R R, *Liebig's Ann Chem*, 1994, 83.
(d) Murakami T, Minamikawa H & Hato M, *Tet Lett*, 35, 1994, 745.
(e) Wild R & Schmidt R R, *Liebig's Ann Chem* 1995, 755.
(f) Schmidt R R & Macer T, *Carbohydrate Res*, 74, 1988, 169.
- 17 (a) Hannun Y A & Bell R M, *Science*, 243, 1989, 500.
(b) Merrill A H, Nimkar S, Menaldino D, Hannun Y A, Loomis C, Bell R M, Tyagi S R, Lambeth J D, Stevens V L, Hunter R & Liotta D C, *Biochemistry*, 28, 1989, 3138.
- 18 The elemental analysis and spectral data for the new compounds were in accordance with the structures assigned, and only selected data are listed.

12: IR (KBr, cm⁻¹): 3431, 3020, 2927, 1739, 1676; [α]_D²⁰ +12.5 (c 1, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 0.89 (t, J = 6.00 Hz, 6H), 1.25 (s, 50H), 2.04 (s, 6H), 2.08(s, 3H), 2.20(m, 2H), 4.00(dd, J = 3.00, 12.00 Hz, 1H), 4.30 (dd, J = 5.00, 10.00 Hz, 1H), 4.50(m, 1H), 4.92(dt, J = 3.00, 7.00 Hz, 1H), 5.12(dd, J = 4.00, 7.00 Hz, 1H), 5.95 (d, J = 10.00 Hz, 1H); ¹³C NMR (400 MHz, CDCl₃): δ 13.99 (q), 28.61(q) 28.61(q), 28.89(q), 22.61(t), 25.49(t), 28.33(t), 29.29-21.63(t), 31.88(t), 36.73(t), 47.58(d), 62.69(t), 72.36(d), 72.99(d), 169.92(s), 170.68(s), 170.91(s), 172.74(s); MS: m/z 668 (M⁺+1), 608.