## 24-Ethyl-cholest-22-en-3α-ol and other constituents from the roots of *Holoptelea integrifolia*

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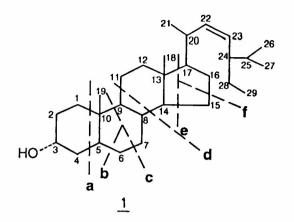
and

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From the combined pet ether-benzene extract of roots of *Holoptelea integrifolia*, a rare cholesterol derivative identified as 24-ethyl-cholest-22-en-3 $\alpha$ -ol has been isolated alongwith *n*-tetracosane,  $\alpha$ -amyrin, epifriedelinol,  $\beta$ -amyrin,  $\beta$ -sitosterol and friedelin. Characterization have been made on the basis of chemical and spectral evidences.

Holoptelea integrifolia (Roxb.) Planch. (Fam. Ulmaceae) is a large glabrous deciduous tree distributed throughout the greater parts of India up to an altitude of 800 m. A survey of literature revealed some work on the phytochemistry of leaves<sup>1</sup>, stem bark<sup>1-4</sup>, heart wood<sup>5,6</sup> and seeds<sup>7</sup> and antiviral activity of the crude leaf sap<sup>8</sup>. This communication describes the isolation and identification of phytochemicals from the roots.

The sterol 1, white granules, m.p. 193-95°, showed  $M^+$  at m/z 414 corresponding to the molecular formula  $C_{29}H_{50}O$ . It showed IR absorption bands for hydroxyl (3440 cm<sup>-1</sup>), olefinic bond (1650 cm<sup>-1</sup>) and *gem*-dimethyl group (1380 and 1370 cm<sup>-1</sup>). Its <sup>1</sup>H NMR spectrum exhibited singlets at  $\delta$  0.67 and 0.78 due to angular methyl groups at C-18 and C-19, a doublet at  $\delta$  0.98 due to C-21 methyl group and multiplets at  $\delta$  5.12 (2H) corresponding to two olefinic protons, 4.04 for H-3 proton, 2.08 (2H) for the allylic protons at C-20 and C-24, 0.81 (9H) for the methyl group at C-29 and isopropyl group and in the region 1.24-1.88 (26H) for the remaining methine and methylene protons. A broad singlet at  $\delta$  2.55



(1H) which disappeared on deuteration confirmed the presence of an -OH group. In the mass spectrum, in addition to the molecular ion peak at m/z 414, peaks due to characteristic steroid fragmentations a, b, c, d, e and f at m/z 342, 303, 290, 165, 233 and 248 respectively indicated the presence of a saturated steroid nucleus<sup>9</sup>. Peaks at m/z 275 [M<sup>+</sup>-C<sub>10</sub>H<sub>19</sub>, side chain, 81%] and 273  $[M^+$  - side chain -2H, 43%] with the high intensity of the former which was formed by the allylic cleavage indicated the presence of a  $\Delta^{22(23)}$  double bond<sup>10</sup>. The position of the olefinic bond was further confirmed by the peak at m/z 302 which was produced by cleavage of the C<sub>20</sub>-C<sub>22</sub> bond together with a one hydrogen transfer from the charge retaining fragment. The peaks at m/z 399  $[M^{+}-CH_{3}]$ , 396  $[M^{+}-H_{2}O]$ , 381  $[M^{+}-(H_{2}O + CH_{3})]$ and 371  $[M^+-CH(CH_3)_2]$  were also observed. Based on these evidences, compound 1 appeared to be 24-ethyl-cholest-22-en-3-ol.

The orientation of hydroxyl group at C-3 wasfound to be  $\alpha$ . In the <sup>1</sup>H NMR spectrum the H-3 signal appeared at  $\delta$  4.04 which fulfilled the requirement for 3 $\beta$ -H accounting for the fact that equatorial protons attached to hydroxyl substituted carbon atoms appear at lower fields than the axial protons in epimeric alcohols<sup>11</sup>. The same was also confirmed by the mass spectrum where an intense peak appeared at m/z 343, as only those structures with  $\alpha$ -oriented hydroxyl group readily undergo cleavage of the A ring<sup>12</sup>. A survey of literature revealed that this steroid 24-ethyl-cholest-22-en- $3\alpha$ -ol has been isolated earlier from the sponge  $3\alpha$ -hydroxy group are very rare in the higher plants.

## **Experimental Section**

General. Melting points are uncorrected. IR spectra were recorded (as KBr pellets) on a Perkin-Elmer model 557 spectrophotometer, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra on model Jeol FX 90Q at 89.55 MHz and 22.49 MHz respectively, using CDCl<sub>3</sub> and DMSO- $d_6$  as solvents and TMS as an internal standard and EIMS spectra on a Hitachi Model RMU 6E Mass Spectrometer.

Column chromatography was carried out using silica gel (60-120 mesh). TLC was carried out on silica gel G chromatoplates and 2% ceric ammonium sulphate as spraying agent.

**Plant material.** The plant material was collected from the Campus, University of Rajasthan, Jaipur during July 1994 and identified at the Botany Department of our University.

**Extraction.** Air-dried, powdered *H. integrifolia* (1.5 kg, roots) was exhaustively extracted with ethanol (95%) on a steam bath for  $8 \times 3$  hr. The concentrated ethanolic extract (60 g) was fractionated with pet. ether followed by benzene which on concentration afforded 2.5 g and 10.5 g fractions respectively. Both the fractions gave similar spots on TLC (benzene:EtOAc, 1:1), hence they were mixed together.

**Isolation of compounds.** The combined pet. etherbenzene fraction on CC over silica gel (260 g) gave following compounds.

*n*-Tetracosane.  $M^+$  338, m.p. 52-54° (EtOAc) (Eluent:pet.ether, 0.44 g).

α-**Amyrin.**  $M^+$  426, m.p. 183-84° (MeOH) (Eluent: pet. ether: C<sub>6</sub>H<sub>6</sub>, 4:1, 0.76 g).

**Epifriedelinol.**  $M^+$  428, m.p. 277-78° (CHCl<sub>3</sub>-MeOH), (Eluent: pet. ether: C<sub>6</sub>H<sub>6</sub>, 2:3, 0.56 g).

**Lupeol.**  $M^+$  426, m.p. 210-11° (CHCl<sub>3</sub>-MeOH), (Eluent: pet. ether: C<sub>6</sub>H<sub>6</sub>, 2:3, 0.53 g).

β-**Amyrin.**  $M^+$  426, m.p. 196-97° (CHCl<sub>3</sub>-MeOH), (Eluent:pet. ether: C<sub>6</sub>H<sub>6</sub>, 1:4, 0.52 g).

β-Sitosterol. M<sup>+</sup> 414, m.p. 135-36° (CHCl<sub>3</sub>-MeOH), (Eluent: pet. ether: C<sub>6</sub>H<sub>6</sub>, 1:4, 0.42 g).

**Friedelin.**  $M^+$  426, m.p. 258-60° (CHCl<sub>3</sub>-MeOH), (Eluent: C<sub>6</sub>H<sub>6</sub>, 0.98 g).

24-Ethyl-cholest-22-en- $3\alpha$ -ol 1. White granules, M<sup>+</sup> 414, m.p. 193-95° (CHCl<sub>3</sub>-MeOH), acetate (Ac<sub>2</sub>O/Py), m.p. 125-27° (MeOH), (Eluent: EtOAc, 0.72 g).

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## References

- Misra G, Bhatnagar S C & Nigam S K, Planta Med, 26, 1974, 394.
- 2 Gupta N C, Singh B & Bhakuni D S, Phytochemistry, 8, 1969, 79.
- 3 Biwas K M & Malik H, J Indian Chem Soc, 63, 1986, 448.
- 4 Mondal D N, Barik B R, Dey A K, Patra A & Kundu A B, Indian Drugs, 31, 1994, 69.
- 5 Misra G, Bhatnagar S C & Nigam S K, Planta Med, 27, 1975, 299.
- 6 Misra G, Bhatnagar S C & Nigam S K, Planta Med 31, 1977, 232.
- 7 Singh P & Sharma S, Current Research on Medicinal and Aromatic Plants, 17, 1995, 171.
- 8 Tripathi R N, Tripathi R K R & Pandey D K, *Environment India*, 4, **1981**, 86.
- 9 Budzikiewicz H & Djerassi C, *J Am Chem Soc*, 84, 1962, 1430.
- 10 Wyllie S G & Djerassi C, J Org Chem, 33, 1968, 305.
- 11 Seldes A M, Gros E G, Suarez A, Ravirosa J & San-Martin A, Tetrahedron, 44, 1988, 1359.
- 12 Friedland S S, Lane G H, Longman R T, Train K E & O'Neal M, J Anal Chem, 31, 1959, 169.