

# BMJ Open Multiorgan impairment in low-risk individuals with post-COVID-19 syndrome: a prospective, community-based study

Andrea Dennis,<sup>1</sup> Malgorzata Wamil,<sup>2,3</sup> Johann Alberts,<sup>4</sup> Jude Oben,<sup>5,6</sup> Daniel J Cuthbertson,<sup>7</sup> Dan Wootton,<sup>8,9</sup> Michael Crooks,<sup>10,11</sup> Mark Gabbay,<sup>12</sup> Michael Brady,<sup>1,13</sup> Lyth Hishmeh,<sup>14</sup> Emily Attree,<sup>15</sup> Melissa Heightman,<sup>16</sup> Rajarshi Banerjee,<sup>1</sup> Amitava Banerjee ,<sup>16,17,18</sup> On behalf of COVERSCAN study investigators

**To cite:** Dennis A, Wamil M, Alberts J, *et al*. Multiorgan impairment in low-risk individuals with post-COVID-19 syndrome: a prospective, community-based study. *BMJ Open* 2021;**11**:e048391. doi:10.1136/bmjopen-2020-048391

► Prepublication history and additional material for this paper are available online. To view these files, please visit the journal online (<http://dx.doi.org/10.1136/bmjopen-2020-048391>).

RB and AB are joint senior authors.

Received 26 December 2020  
Revised 25 February 2021  
Accepted 11 March 2021



© Author(s) (or their employer(s)) 2021. Re-use permitted under CC BY. Published by BMJ.

For numbered affiliations see end of article.

## Correspondence to

Dr Amitava Banerjee;  
[ami.banerjee@ucl.ac.uk](mailto:ami.banerjee@ucl.ac.uk)

## ABSTRACT

**Objective** To assess medium-term organ impairment in symptomatic individuals following recovery from acute SARS-CoV-2 infection.

**Design** Baseline findings from a prospective, observational cohort study.

**Setting** Community-based individuals from two UK centres between 1 April and 14 September 2020.

**Participants** Individuals  $\geq 18$  years with persistent symptoms following recovery from acute SARS-CoV-2 infection and age-matched healthy controls.

**Intervention** Assessment of symptoms by standardised questionnaires (EQ-5D-5L, Dyspnoea-12) and organ-specific metrics by biochemical assessment and quantitative MRI.

**Main outcome measures** Severe post-COVID-19 syndrome defined as ongoing respiratory symptoms and/or moderate functional impairment in activities of daily living; single-organ and multiorgan impairment (heart, lungs, kidneys, liver, pancreas, spleen) by consensus definitions at baseline investigation.

**Results** 201 individuals (mean age 45, range 21–71 years, 71% female, 88% white, 32% healthcare workers) completed the baseline assessment (median of 141 days following SARS-CoV-2 infection, IQR 110–162). The study population was at low risk of COVID-19 mortality (obesity 20%, hypertension 7%, type 2 diabetes 2%, heart disease 5%), with only 19% hospitalised with COVID-19. 42% of individuals had 10 or more symptoms and 60% had severe post-COVID-19 syndrome. Fatigue (98%), muscle aches (87%), breathlessness (88%) and headaches (83%) were most frequently reported. Mild organ impairment was present in the heart (26%), lungs (11%), kidneys (4%), liver (28%), pancreas (40%) and spleen (4%), with single-organ and multiorgan impairment in 70% and 29%, respectively. Hospitalisation was associated with older age ( $p=0.001$ ), non-white ethnicity ( $p=0.016$ ), increased liver volume ( $p<0.0001$ ), pancreatic inflammation ( $p<0.01$ ), and fat accumulation in the liver ( $p<0.05$ ) and pancreas ( $p<0.01$ ). Severe post-COVID-19 syndrome was associated with radiological evidence of cardiac damage (myocarditis) ( $p<0.05$ ).

## Strengths and limitations of this study

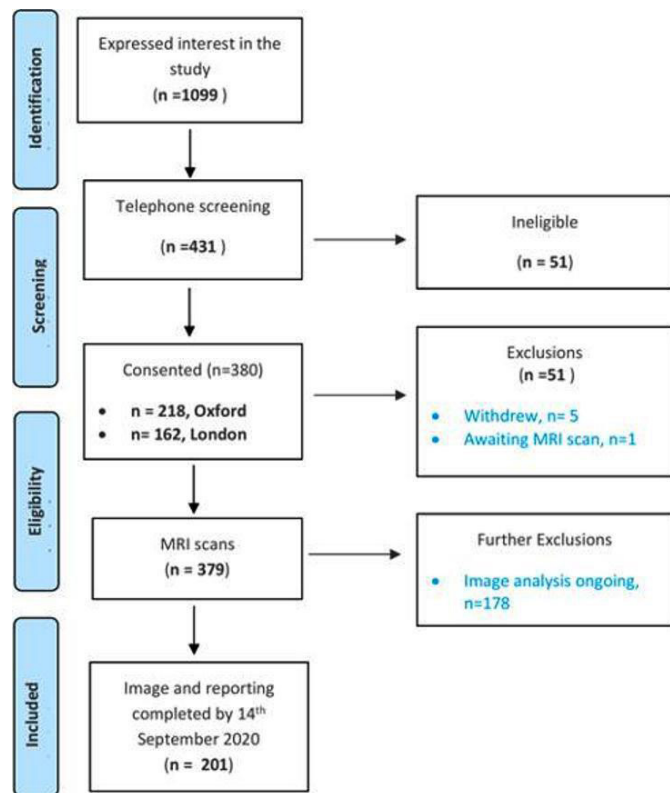
- This is an ongoing, prospective, longitudinal COVID-19 recovery study with biochemical and imaging characterisation of organ function, starting in April 2020 before recognition of ‘long-COVID’, proper testing availability and prospective COVID-19-related research.
- By recruiting ambulatory patients with broad inclusion criteria, we focused on a real-world population at lower risk of COVID-19 severity and mortality.
- Healthy controls were included for comparison, not individuals with postinfluenza symptoms, COVID-19 without symptoms or from general clinics, which further studies may explore.
- The study population was not ethnically diverse despite disproportionate COVID-19 impact in non-white individuals.
- To limit interaction and exposure between the trial team and the patients, pulse oximetry, spirometry, MRI assessment of the brain and muscle function were not included from the outset.

**Conclusions** In individuals at low risk of COVID-19 mortality with ongoing symptoms, 70% have impairment in one or more organs 4 months after initial COVID-19 symptoms, with implications for healthcare and public health, which have assumed low risk in young people with no comorbidities.

**Trial registration number** NCT04369807; Pre-results.

## INTRODUCTION

Early in the COVID-19 pandemic, research and clinical practice focused on pulmonary manifestations.<sup>1</sup> There is increasing evidence for direct multiorgan effects,<sup>2–7</sup> as well as indirect effects on other organ systems and disease processes, such as cardiovascular diseases and cancers, through changes in healthcare delivery and patient behaviours.<sup>8–10</sup> The



**Figure 1** Flow from recruitment to enrolment of 201 patients with post-COVID-19 syndrome.

clear long-term impact on individuals and health systems underlines the urgent need for a whole body approach with assessment of all major organ systems following SARS-CoV-2 infection. Quantitative MRI has recently been used to show multiorgan impairment in individuals post-COVID-19 hospitalisation,<sup>11</sup> but has not been used in non-hospitalised individuals.

COVID-19 is the convergence of an infectious disease, undertreated non-communicable diseases and social determinants of health, described as a ‘syndemic’.<sup>12</sup> Pre-existing non-communicable diseases and risk factors predict poor COVID-19 outcomes, whether intensive care admission or mortality.<sup>10</sup> Research has emphasised acute SARS-CoV-2 infection, hospitalised individuals and COVID-19 mortality,<sup>13–15</sup> which is likely to underestimate the true burden of COVID-19-related disease. Among those surviving acute infection, 10% report persistent symptoms for 12 weeks or longer after initial infection (‘long-COVID’, or ‘post COVID-19 syndrome’, PCS).<sup>16</sup> However, PCS is yet to be fully defined.<sup>17–20</sup> Neither severity of symptoms, nor medium-term and long-term pathophysiology across organ systems, nor the appropriate control populations are understood.

UK government policies have emphasised excess mortality risk in moderate-risk and high-risk conditions, including ‘shielding’<sup>10</sup> and commissioning of a risk calculator to identify those at highest risk of COVID-19 severity and mortality.<sup>21</sup> These policies assume that younger individuals without apparent underlying conditions are at low risk. However, unlike symptoms following critical illness<sup>22</sup>

or acute phase of other coronavirus infections,<sup>23</sup> symptoms in PCS are commonly reported in individuals with low COVID-19 mortality risk, for example, female, young and no chronic comorbidities.<sup>14</sup> The potential scale of PCS in ‘lower-risk’ individuals, representing up to 80% of the population,<sup>3</sup> necessitates urgent policies across countries to monitor,<sup>24</sup> treat<sup>19</sup> and pay<sup>25</sup> for long-term implications of COVID-19 and to mitigate impact on healthcare utilisation and economies.

Therefore, in a pragmatic, prospective cohort study of individuals with persistent symptoms at least 4 weeks following recovery from acute SARS-CoV-2 infection and at low risk of COVID-19 mortality, we investigated (1) the prevalence of multiorgan impairment, compared with healthy, age-matched controls; (2) the associations between typical COVID-19 symptoms and multiorgan impairment; and (3) the associations between hospitalisation, severity of symptoms and multiorgan impairment.

## METHODS

### Patient population and study design

In an ongoing, prospective study, participants were recruited to the study following expression of interest on the study registration website. Participants learnt about the study through advertisement on social media or via recommendations from clinicians from four participant identification centres, the latter usually applied to patients who had been hospitalised. Assessment took place at two UK research imaging sites (Perspectum, Oxford; and Mayo Clinic Healthcare, London) between 1 April 2020 and 14 September 2020, completing baseline assessment by 14 September 2020 (figure 1). Participants with laboratory-confirmed SARS-CoV-2 infection (tested SARS-CoV-2-positive by oropharyngeal/nasopharyngeal swab by reverse-transcriptase PCR (n=62), a positive antibody test (n=63), or with strong clinical suspicion of infection with typical symptoms/signs and assessed as highly likely to have COVID-19 by two independent clinicians (n=73)) were eligible for enrolment. Exclusion criteria were symptoms of active respiratory viral infection (temperature >37.8°C or three or more episodes of coughing in 24 hours), hospital discharge in the last 7 days, and contraindications to MRI, including implanted pacemakers, defibrillators, other metallic implanted devices and claustrophobia. All participants gave written informed consent.

### Assessment of PCS

Assessment included patient-reported validated questionnaires (quality of life, EQ-5D-5L,<sup>26</sup> and Dyspnoea-12<sup>27</sup>) and fasting biochemical investigations (listed in online supplemental methods). PCS was classified as ‘severe’ (defined as persistent breathlessness, score of ≥10 on Dyspnoea-12, or reported moderate or greater problems with usual activities on EQ-5D-5L) or ‘moderate’. These thresholds were selected as the Dyspnoea-12 has been correlated with the Medical Research Council (MRC)

**Table 1** Baseline demographics and symptoms of 201 low-risk individuals with post-COVID-19 syndrome

	All patients (N=201)	Healthy controls (n=36)	P value	Not hospitalised (n=163)	Hospitalised (n=37)	P value	Moderate PCS (n=77)	Severe PCS (n=116)	P value
Age (years), mean (SD)	44 (11.0)	39 (12.4)	0.013	43 (10.9)	50 (10.0)	0.001	45 (12.2)	44 (10.0)	0.419
Female, n (%)	142 (70.6)	14 (38.9)	0.032	118 (72.4)	23 (62.2)	0.302	51 (66.2)	85 (73.3)	0.374
BMI (kg/m <sup>2</sup> ), median (IQR)	25.7 (22.7–28.1)	23.2 (21.4–23.1)	<0.001	25.3 (22.7–27.7)	27.2 (23.1–31.0)	0.063	25.8 (22.7–27.9)	25.4 (22.5–28.2)	0.639
Ethnicity									
White	176 (87.6)	33 (91.7)		148 (90.8)	28 (75.7)		67 (87.0)	106 (91.4)	0.178
Mixed	3 (1.5)	0 (0)	0.904	3 (1.8)	0 (0)	0.016	1 (1.3)	2 (1.7)	
South Asian	7 (3.5)	3 (8.3)		4 (2.5)	3 (8.1)		5 (6.5)	0 (0)	
Black	4 (2.0)	0 (0)		1 (0.6)	2 (5.4)		2 (2.6)	2 (1.7)	
Comorbidities and risks									
Smoking									0.244
Never	133 (66.2)	20 (60.6)		108 (66.3)	24 (64.9)		55 (71.4)	72 (61.7)	
Current	6 (3.0)	8 (24.2)	<0.001	6 (3.7)	0 (0)	0.641	3 (3.9)	3 (2.6)	
Ex-smoker	62 (30.8)	5 (15.2)		49 (30.1)	13 (35.1)		19 (24.7)	41 (35.3)	
Healthcare worker	64 (31.8)	4 (12.1)	0.009	50 (30.7)	13 (35.1)	0.695	33 (42.9)	28 (24.1)	0.007
Asthma	37 (18.4)	0 (0)	0.002	34 (20.9)	3 (8.1)	0.099	13 (16.9)	22 (19.0)	0.849
BMI									
≥25 kg/m <sup>2</sup>	113 (56.5)	7 (20)		87 (53.7)	25 (67.6)	0.144	46 (60.5)	62 (53.4)	0.374
≥30 kg/m <sup>2</sup>	40 (20.0)	0 (0)		28 (17.3)	12 (32.4)	0.066	16 (21.1)	24 (20.7)	1.000
Hypertension	13 (6.5)	0 (0)	0.001	11 (6.7)	2 (5.4)	1.000	6 (7.8)	7 (6.0)	0.771
Diabetes	4 (2.0)	0 (0)	0.104	4 (2.5)	0 (0.0)	1.000	4 (5.2)	0 (0.0)	0.024
Previous heart disease	9 (4.5)	0 (0)	0.001	8 (4.9)	1 (2.7)	1.000	3 (3.9)	5 (4.3)	1.000
Symptoms									
Fatigue	196 (98.0)			159 (97.5)	37 (100.0)	1.000	73 (96.1)	115 (99.1)	0.302
Shortness of breath	176 (88.0)			141 (86.5)	35 (94.6)	0.262	58 (76.3)	112 (96.6)	<0.0001
Muscle ache	173 (86.5)			142 (87.1)	31 (83.8)	0.597	66 (86.8)	101 (87.1)	1.000
Headache	165 (82.5)			138 (84.7)	27 (73.0)	0.098	56 (73.7)	102 (87.9)	0.019
Joint pain	156 (78.0)			127 (77.9)	29 (78.4)	1.000	56 (73.7)	94 (81.0)	0.284
Chest pain	152 (76.0)			128 (78.5)	24 (64.9)	0.090	47 (61.8)	98 (84.5)	0.001
Cough	146 (73.0)			117 (71.8)	29 (78.4)	0.539	55 (72.4)	84 (72.4)	1.000
Fever	144 (72.0)			113 (69.3)	31 (83.8)	0.104	51 (67.1)	86 (74.1)	0.329
Sore throat	143 (71.5)			120 (73.6)	23 (62.2)	0.165	50 (65.8)	86 (74.1)	0.256

Continued



Table 1 Continued

	All patients (N=201)	Healthy controls (n=36)	P value	Not hospitalised (n=163)	Hospitalised (n=37)	P value	Moderate PCS (n=77)	Severe PCS (n=116)	P value
Diarrhoea	118 (59.0)			91 (55.8)	27 (73.0)	0.065	40 (52.6)	76 (65.5)	0.097
Abnormal pain	108 (54.0)			91 (55.8)	17 (45.9)	0.361	30 (39.5)	75 (64.7)	0.001
Wheezing	98 (49.0)			75 (46.0)	23 (62.2)	0.101	30 (39.5)	64 (55.2)	0.039
Inability to walk	80 (40.0)			58 (35.6)	22 (59.5)	0.009	24 (31.6)	50 (43.1)	0.130
Runny nose	68 (34.0)			55 (33.7)	13 (35.1)	0.85	24 (31.6)	41 (35.3)	0.642
Time interval									
Initial symptoms to assessment (days), median (IQR)	141 (110–162)			141 (112–163)	138 (97–150)	0.106	121 (89–158)	145 (121–163)	0.001
COVID-19-positive to assessment (days), median (IQR)	71 (41–114)			68 (35–112)	105 (59–126)	0.012	60 (43–98)	78 (34–119)	0.305

Data are presented as count (%). Comparisons between managed at home versus hospitalised and between moderate versus PCS were conducted using Fisher's exact test BMI, body mass index; PCS, post-COVID-19 syndrome.

dyspnoea grade, where level 3 warrants referral to rehabilitation services,<sup>27</sup> and with EQ-5D-5L, less than 8% of the general population report moderate or greater problems with usual activities.<sup>28</sup>

### Multiorgan impairment in PCS compared with healthy controls

We selected MRI as the imaging modality (as in UK Biobank) due to (1) safety (no radiation exposure, no need for intravenous contrast and minimal contact with the radiographer); (2) quantitative reproducibility (>95% acquisition and image processing success rate); (3) capacity for information sharing (digital data repository for independent analysis and research); and (4) rapid scalability (35 min scan to phenotype lung, heart, kidney, liver, pancreas and spleen). Multiorgan MRI data were collected at both study sites (Oxford: MAGNETOM Aera 1.5T; Mayo Healthcare London: MAGNETOM Vida 3T; both from Siemens Healthcare, Erlangen, Germany). The COVERSCAN multiparametric MRI assessment typically required 35 min per patient, including the lungs, heart, liver, pancreas, kidneys and spleen, by standardised methodology (online supplemental file 1). In brief, we assessed inflammation of the heart, kidneys, liver and pancreas with quantitative T1 relaxation mapping; lung function was characterised with a dynamic structural T2-weighted lung scan estimating lung capacity; ectopic fat accumulation in the liver and pancreas from proton density fat fraction; and volume of the liver and spleen measured from T1-weighted structural scan.

To determine impairment in each organ, we compared MRI-derived measurements from the heart, lungs, kidneys, liver, pancreas and spleen with reference ranges (online supplemental table 1), which were established as mean±2 SD from the healthy, age-matched control subjects (n=36) and validated by scoping literature review.<sup>11</sup> We defined organ impairment if quantitative T1 mapping was outside the reference ranges for the heart, kidney, liver and pancreas, reduced estimated lung capacity from dynamic measurements in the lungs, or there was evidence of hepatomegaly, splenomegaly or ectopic fat accumulation.

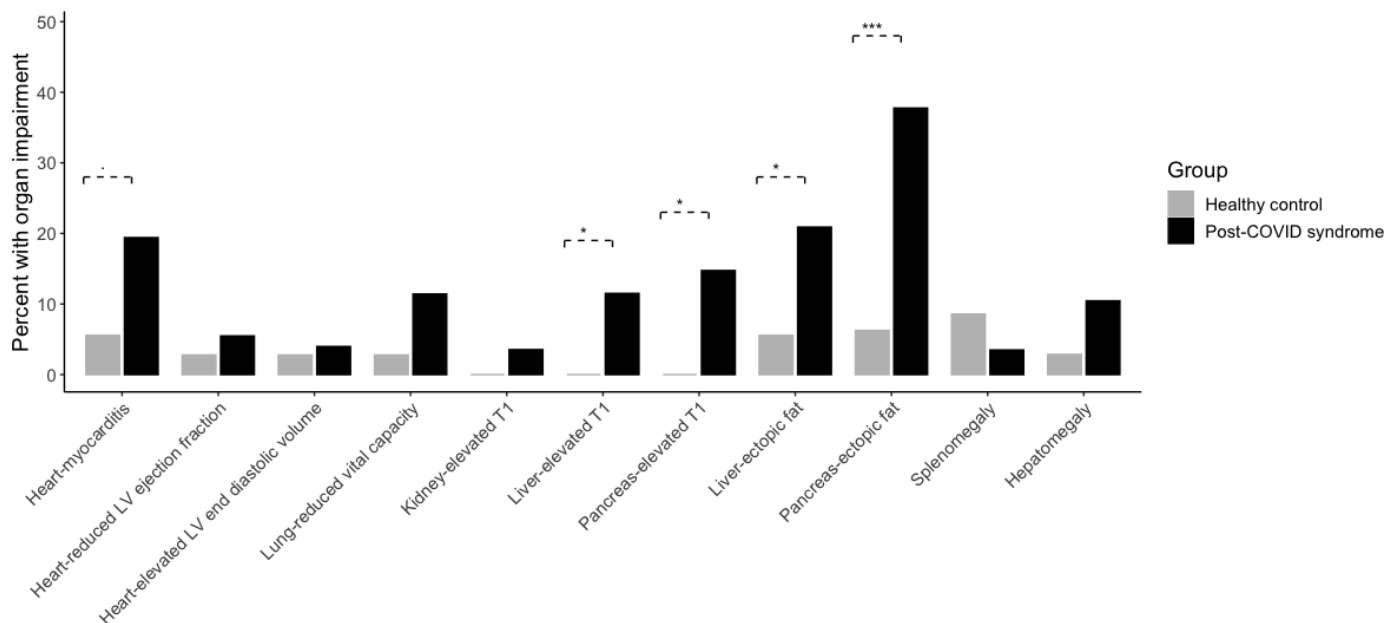
### Symptoms and multiorgan impairment

Associations between organ impairment and symptoms were visually assessed using a heat map, dividing those with impairments to an organ into columns and colouring the rows by percentage of reported symptoms.

### Hospitalisation, severity and multiorgan impairment

We compared mean differences in quantitative organ metrics for hospitalised versus not hospitalised and moderate versus severe PCS using Kruskal-Wallis test (Fisher's exact test for differences in binary outcomes). We defined multiorgan impairment as ≥2 organs with metrics outside the reference range. We investigated the associations between multiorgan impairment and (1) being hospitalised and (2) severe PCS with multivariate





**Figure 2** Percentage of patients (black) and controls (grey) with individual organ measures outside of the predefined normal range. Lines represent significant difference in the proportions between the two groups, with \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . LV, left ventricular.

logistic regression models, adjusting for age, sex and body mass index (BMI).

### Patient and public involvement and engagement

Patients and the public have directly and indirectly informed our research, from design to dissemination, with regular updates and webinars, including question and answer sessions with patients. Several clinician coauthors were indirectly informed by their patients in the COVERSCAN study (RB, AB) or PCS clinics (DW, MH, MC), who are members of organisations such as Long Covid SOS (eg, LH) and UKDoctors#Longcovid (eg, EA). LH and EA have been involved in the research, interpretation of results, understanding implications of our results and providing critical feedback to the manuscript.

### Statistical analysis

We performed all analyses using R V.3.6.1, using descriptive statistics to summarise baseline characteristics and considering a  $p$  value less than 0.05 as statistically significant. Mean and SD were used for normally distributed continuous variables, median with IQR for non-normally distributed variables, and frequency and percentage for categorical variables. For group-wise comparison for absolute values between cases and healthy controls, we used Kruskal-Wallis test.

## RESULTS

### Overall study population

#### Baseline characteristics

The study included 201 individuals (full details regarding hospitalisation:  $n=199$ ; full questionnaire data to assign PCS severity:  $n=193$ ). The mean age was 44.0 (range 21–71) years and the median BMI was 25.7 (IQR 23–28).

Of the individuals, 71% were female, 88% were white, 32% were healthcare workers and 19% had been hospitalised with COVID-19. Assessments (symptoms, blood and MRI) had a median of 141 (IQR 110–162) days after initial symptoms. Medical history included smoking (3%), asthma (19%), obesity (20%), hypertension (7%), diabetes (2%) and prior heart disease (5%). The healthy control group had a mean age of 39 years (range 20–70), 40% were female, with a median BMI of 23 (IQR: 21–25) (table 1).

Regardless of hospitalisation, the most frequently reported symptoms were fatigue (98%), shortness of breath (88%), muscle ache (87%) and headache (83%) (table 1). Of the individuals, 99% had four or more and 42% had ten or more symptoms. Of individuals 70% reported  $\geq 13$  weeks off paid employment. Of the incidental structural findings observed on MRI ( $n=56$ ), three were cardiac (atrial septal defect, bicuspid aortic valve and right atrial mass), one renal (hydronephrosis) and the rest were benign cysts.

Haematological investigations, including mean corpuscular haemoglobin concentration (24%), and renal, liver and lipid biochemistry, including potassium (38%), alanine transferase (14%), lactate dehydrogenase (17%), triglycerides (11%) and cholesterol (42%), were abnormally high in  $\geq 10\%$  of individuals. Bicarbonate (10%), phosphate (11%), uric acid (11%) and transferrin saturation (19%) were abnormally low in  $\geq 10\%$  of individuals (online supplemental table 1).

### Single-organ and multiorgan impairment in PCS compared with healthy controls

Organ impairment was more common in PCS than healthy controls (figure 2 and online supplemental figure

**Table 2** Evidence of organ impairment in 201 low-risk individuals with post-COVID-19 syndrome

Measurement	All patients (N=201)	Healthy controls (n=36)	P value	Not hospitalised (n=163)	Hospitalised (n=37)	P value	Moderate PCS (n=77)	Severe PCS (n=116)	P value
<b>Heart</b>									
Left ventricular ejection fraction (%)									
Normal (>51%)	190 (95.0)	35 (97.2)	0.699	155 (95.7)	33 (89.1)	0.124	72 (93.5)	111 (95.7)	0.353
Impaired (≤51%)	11 (5.0)	1 (2.8)		7 (4.3)	4 (10.1)		5 (6.4)	5 (4.3)	
Left ventricular end diastolic volume (mL)									
>264 mL in Men; >206 mL in Women	8 (4.0)	1 (2.8)	1.00	4 (2.5)	4 (10.8)	0.040	4 (5.2)	4 (3.4)	0.715
<b>Evidence of myocarditis</b>									
≥3 segments with high T1 (≥1229 ms at 3T; ≥1015 ms at 1.5T)	39 (19.4)	2 (5.6)	0.053	30 (18.4)	8 (21.6)	0.647	9 (11.7)	29 (25.0)	0.027
<b>Lungs</b>									
Deep breathing fractional area change	(n=17 missing)			(n=13 missing)	(n=3 missing)		(n=8 missing)	(n=7 missing)	
<31%	21 (11.4)	1 (2.8)	0.138	17 (11.3)	4 (11.8)	1	7 (10.1)	13 (11.9)	0.811
<b>Kidneys</b>									
Kidney cortex T1	(n=3 missing)			(n=3 missing)			(n=2 missing)		
Normal (<1652 ms at 3T; <1227 ms at 1.5T)	191 (96.5)	36 (100.0)	0.599	155 (96.9)	35 (94.6)	0.618	74 (98.7)	112 (96.6)	0.65
Impaired (≥1652 ms at 3T; ≥1227 ms at 1.5T)	7 (3.5)	0 (0.0)		5 (3.1)	2 (5.4)		1 (1.3)	4 (3.4)	
<b>Pancreas</b>									
Pancreatic inflammation (T1 in ms)	(n=11 missing)	(n=13 missing)		(n=7 missing)	(n=4 missing)		(n=4 missing)	(n=6 missing)	
Normal <803 ms	162 (85.3)	23 (100.0)	0.049	139 (89.1)	22 (66.7)	0.002	60 (82.2)	95 (86.4)	0.530
Impaired ≥803 ms	28 (14.7)	0 (0)		17 (10.9)	11 (33.3)		13 (17.8)	15 (13.6)	
<b>Pancreatic fat</b>									
Normal <4.6%	122 (62.2)	30 (93.8)	<0.001	107 (66.9)	14 (40.0)	0.004	44 (57.9)	72 (63.7)	0.449
Impaired ≥4.6%	74 (37.8)	2 (6.2)		53 (33.1)	21 (60.0)		32 (42.1)	41 (36.3)	
<b>Liver</b>									
Liver inflammation (cT1 in ms)	(n=1 missing)			(n=1 missing)			(n=1 missing)		
Normal <784 ms	177 (88.5)	36 (100)	0.030	148 (91.4)	28 (75.7)	0.018	69 (90.8)	101 (87.1)	0.494
Impaired ≥784 ms	23 (11.5)	0 (0)		14 (8.6)	9 (24.3)		7 (9.2)	15 (12.9)	
<b>Liver fat</b>									
Normal <4.8%	159 (79.1)	34 (94.4)	0.034	134 (82.2)	24 (64.9)	0.026	61 (79.2)	91 (78.4)	1
Impaired ≥4.8%	42 (20.9)	2 (5.4)		29 (17.8)	13 (35.1)		16 (20.8)	25 (21.6)	
Liver volume	(n=1 missing)								

Continued

Table 2 Continued

Measurement	All patients (N=201)	Healthy controls (n=36)	P value	Not hospitalised (n=163)	Hospitalised (n=37)	P value	Moderate PCS (n=77)	Severe PCS (n=116)	P value
Normal <1935 mL	180 (89.6)	34 (97.1)	0.214	154 (94.5)	25 (67.6)	<0.0001	68 (88.3)	104 (89.7)	0.816
Impaired ≥1935 mL	21 (10.4)	1 (2.9)		9 (5.5)	12 (32.4)		9 (11.7)	12 (10.3)	
Spleen									
Spleenic volume (mL)		(n=1 missing)							
Normal <350 mL	194 (96.5)	32 (91.4)	0.172	160 (98.2)	33 (89.2)	0.023	74 (96.1)	112 (96.6)	1
Impaired ≥350 mL	7 (3.5)	3 (8.6)		3 (1.8)	4 (10.8)		3 (3.9)	4 (3.4)	

Data are presented as count (%). Comparisons between managed at home versus hospitalised and between moderate versus PCS were conducted using Fisher's exact test. PCS, post-COVID-19 syndrome.

1). Impairment was present in the heart in 26% (myocarditis 19%, systolic dysfunction 9%), lung in 11% (reduced vital capacity), kidney in 4% (inflammation), liver in 28% (12% inflammation, 21% ectopic fat, 10% hepatomegaly), pancreas in 40% (15% inflammation, 38% ectopic fat) and spleen in 4% (splenomegaly) (figure 2 and table 2). Of the individuals, 70% had impairment in at least one organ and 29% had multiorgan impairment, with overlap across multiple organs (figure 3). Impairment in the liver, heart or lungs was associated with further organ impairment in 63%, 62% and 48% of individuals, respectively (figure 3).

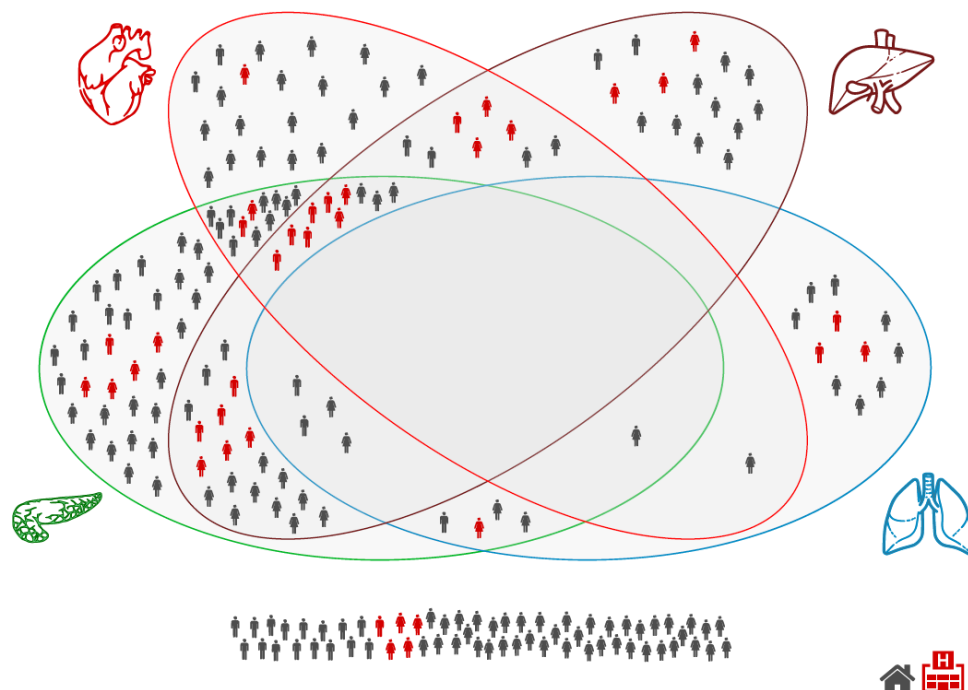
### Symptoms and multiorgan impairment

Hepatic and pulmonary impairment frequently clustered together, with fatigue, muscle aches, fever and cough commonly reported. Impairment in particular organs was associated with particular symptoms—pancreas: diarrhoea, fever, headache and dyspnoea; heart: headache, dyspnoea and fatigue; and kidney: wheezing, runny nose, diarrhoea, cough, fever, headache, dyspnoea and fatigue (figure 4).

### Hospitalisation, severity and multiorgan impairment

The hospitalised group were older ( $p=0.001$ ), had higher BMI ( $p=0.063$ ), and were more likely to be non-white ( $p=0.016$ ) and to report 'inability to walk' ( $p=0.009$ ) than non-hospitalised individuals. There were no other statistically significant differences between risk factors or symptoms between the groups. Impairment of the liver, pancreas (eg, ectopic fat in the pancreas and liver, hepatomegaly) and  $\geq 2$  organs was higher in hospitalised individuals (all  $p<0.05$ ) (figure 3 and table 2). In multivariate analyses, adjusting for age, sex and BMI, liver volume remained significantly associated with hospitalisation ( $p=0.001$ ). Hospitalised individuals had high triglycerides (30% vs 7.2%,  $p=0.002$ ), cholesterol (60% vs 38%,  $p=0.04$ ) and low-density lipoprotein-cholesterol (57% vs 31%,  $p=0.01$ ), and low transferrin saturation (38% vs 15%,  $p=0.01$ ), compared with non-hospitalised individuals. erythrocyte sedimentation rate (ESR) (13%), bicarbonate (12%), uric acid (16%), platelet count (13%) and high-sensitivity C-reactive protein (CRP) (15%) were high in  $\geq 10\%$  of hospitalised individuals.

Of the individuals, 60% ( $n=120$ ) had severe PCS, with 52% reporting persistent, moderate problems undertaking usual activities (level 3 or greater in the relevant EQ-5D-5L question; 34% reported Dyspnoea-12 score  $\geq 10$ ). Of those with severe PCS, 84% were not hospitalised and 73% were female. There were no differences in age, BMI or ethnicity between the groups. Individuals with severe PCS were more likely to report shortness of breath ( $p<0.001$ ), headache ( $p=0.019$ ), chest pain ( $p=0.001$ ), abdominal pain ( $p=0.001$ ) and wheezing ( $p=0.039$ ). Of those with 'severe' PCS, 25% had myocarditis compared with 12% with moderate PCS (unadjusted: 0.023; adjustment for age, sex and BMI:  $p=0.04$ ; online supplemental figure 2). Severe PCS was associated with higher mean



**Figure 3** Multiorgan impairment in low-risk individuals with post-COVID-19 syndrome by gender and hospitalisation.

cell haemoglobin concentration (28% vs 17%), cholesterol (46.2% vs 32.8%), CRP (10% vs 3.8%) and ESR (10% vs 6%) than moderate PCS, but these differences were not statistically significant (online supplemental table 3). Muscle aches, fever and coughing were common in severe PCS, and headache was common in individuals with inflammation of the pancreas (figure 4).

## DISCUSSION

We report three findings in the first COVID-19 recovery study to evaluate medium-term, multiorgan impairment. First, in low-risk individuals, there were chronic symptoms and mild impairment in the heart, lung, liver, kidney and pancreas 4 months post-COVID-19, compared with healthy controls. Second, cardiac impairment was more common in severe PCS. Third, we demonstrate feasibility and potential utility of community-based multiorgan assessment for PCS.

### Comparison with other studies

Common symptoms were fatigue, dyspnoea, myalgia, headache and arthralgia, despite low risk of COVID-19 mortality or hospitalisation. COVID-19 impact models have included age, underlying conditions and mortality, but not morbidity, multiorgan impairment and chronic diseases.<sup>29 30</sup> Even in non-hospitalised individuals, up to 10% of those infected have PCS,<sup>15 31</sup> but studies of extrapulmonary manifestations emphasise acute illness.<sup>32</sup> We describe mild rather than severe organ impairment, but the pandemic's scale and high infection rates in lower risk individuals signal medium-term and longer-term COVID-19 impact, which cannot be ignored in healthcare or policy spheres.

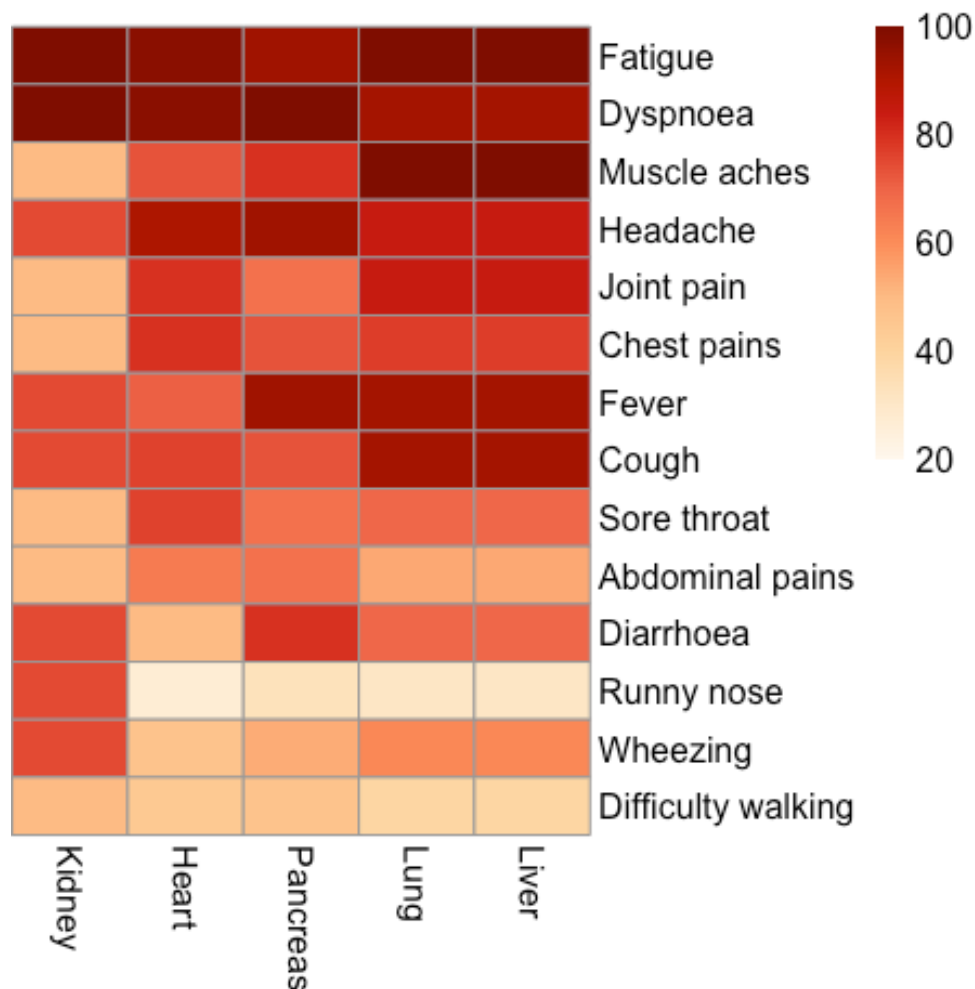
Acute myocarditis and cardiogenic shock<sup>33</sup> are documented in hospitalised patients with COVID-19.<sup>6</sup> In American athletes, recent COVID-19 was associated with myocarditis.<sup>34</sup> Although causality cannot be attributed and postviral syndromes have included similar findings,<sup>21</sup> we show that a quarter of low-risk individuals with PCS have mild systolic dysfunction or myocarditis. The significance of these findings and the associations with contemporaneous abnormal echocardiography findings and long-term myocardial fibrosis and impairment are unknown. Cardiac impairment, a risk factor for severe COVID-19, may have a role in PCS. Two further findings that deserve investigation are pancreatic abnormalities, given the excess diabetes risk reported in PCS,<sup>15</sup> and the preponderance of healthcare workers at increased PCS risk (as observed for COVID-19 mortality), possibly due to higher viral burden.

PCS is likely to be a syndrome rather than a single condition. Despite an immunological basis for individual variations in COVID-19 progression and severity,<sup>35</sup> prediction models have high rates of bias, perform poorly,<sup>36</sup> and focus on respiratory dysfunction and decisions for ventilation in acutely unwell patients, rather than multiorgan function. Ongoing long-term studies<sup>37</sup> exclude non-hospitalised, low-risk individuals. During a pandemic, we studied subclinical organ impairment in PCS, showing low rates of incidental findings. As specialist PCS services are rolled out,<sup>38 39</sup> multiorgan assessment, monitoring and community pathways have potential roles during and beyond COVID-19, but need to be evaluated.

### Implications for research, clinical practice and public health

Our findings have three research implications. First, as countries face second waves, COVID-19 impact models





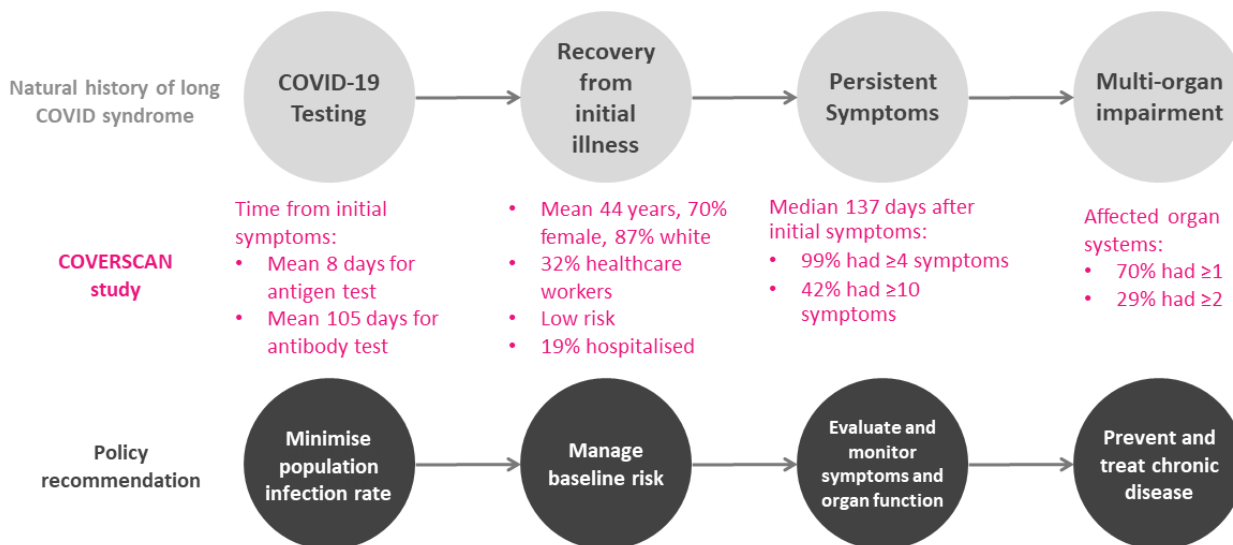
**Figure 4** Percentage of reported symptoms during the acute phases of the illness within those with evidence of organ impairment for each organ separately. Darker red indicates higher percentage of reported symptoms per impaired organ. There are no distinct patterns of symptoms relating to each impaired organ, but a high burden of symptoms in individuals is highlighted.

should include PCS, whether quality of life, healthcare utilisation or economic effects. Second, there is urgent need for multiorgan assessment, including blood and imaging, as well as primary and secondary care data linkage, to define PCS. Third, longitudinal studies of clustering of symptoms and organ impairment will inform health services research to plan multidisciplinary care pathways. There are three management implications. First, we signal the need for multiorgan monitoring in at least the medium term, especially extrapulmonary sequelae. Care pathways involving MRI (with limited access in many clinical settings) need evaluation versus other modalities to detect organ impairment (eg, spirometry, N-terminal pro B-type natriuretic peptide (NT-pro-BNP), ECG, echocardiography, ultrasound and blood investigations). Second, until effective vaccines and treatments are widely available, ‘infection suppression’ (eg, social distancing, masks, physical isolation) is the prevention strategy. Third, whether understanding baseline risk or multiorgan complications, PCS requires management across specialties (eg, cardiology, gastroenterology) and

disciplines (eg, epidemiology, diagnostics, laboratory science) (figure 5).

### Limitations

There are some limitations. First, our cardiac MRI protocol excluded gadolinium contrast due to concerns regarding COVID-19-related renal complications, relying on native T1 mapping to characterise myocardial inflammation non-invasively (previously validated for acute myocarditis).<sup>40</sup> Second, for organ impairment, we show association, not causation, and incidental findings are possible in asymptomatic individuals<sup>41</sup>; however, our findings are strengthened by comparison with healthy, age-matched controls, although not matched for sex or baseline comorbidities. Third, for pragmatic reasons, our controls were scanned using 1.5T, but we used 3T ranges as described in an analogous study with similar acquisition protocols. Therefore, we may be under-representing the true proportion of impairment in those individuals with PCS scanned at 3T. Fourth, further studies may explore different controls, for example, individuals with



**Figure 5** Natural history of post-COVID-19 syndrome, the COVERSCAN study in low-risk individuals (N=201) and policy recommendations.

postinfluenza symptoms, COVID-19 without symptoms or from general clinics. We will investigate duration, trajectory, complications and recovery for specific symptoms and organ impairment in the follow-up phase. Fifth, our study population was not ethnically diverse, despite disproportionate COVID-19 impact in non-white individuals. Sixth, to limit interaction and exposure between the trial team and the patients, pulse oximetry, spirometry, MRI assessment of the brain and muscle function were not included from the outset.

## CONCLUSIONS

Our study suggests PCS has a physiological basis, with measurable patient-reported outcomes and organ impairment. Future research should address longer-term follow-up of organ function beyond symptoms and blood investigations, even in lower risk individuals; prioritisation for imaging, investigation and referral; and optimal care pathways. Health system responses should emphasise infection suppression and management of pre-COVID-19 and post-COVID-19 risk factors and chronic diseases.

### Author affiliations

<sup>1</sup>Perspectum, Oxford, UK

<sup>2</sup>Department of Cardiology, Great Western Hospital Foundation NHS Trust, Swindon, UK

<sup>3</sup>Department of Cardiology, Oxford University Hospitals NHS Foundation Trust, Oxford, UK

<sup>4</sup>Alliance Medical, Warwick, UK

<sup>5</sup>Department of Gastroenterology, Guy's and St Thomas' NHS Foundation Trust, London, UK

<sup>6</sup>Institute for Liver and Digestive Health, University College London, London, UK

<sup>7</sup>Institute of Cardiovascular and Metabolic Medicine, University of Liverpool, Liverpool, UK

<sup>8</sup>Institute of Infection and Global Health, University of Liverpool, Liverpool, UK

<sup>9</sup>Department of Respiratory Research, Liverpool University Hospitals NHS Foundation Trust, Liverpool, UK

<sup>10</sup>Department of Respiratory Medicine, Hull and East Yorkshire Hospitals NHS Trust, Hull, UK

<sup>11</sup>Institute of Clinical and Applied Health Research, University of Hull, Hull, UK

<sup>12</sup>Institute of Population Health Sciences, University of Liverpool, Liverpool, UK

<sup>13</sup>Department of Oncology, University of Oxford, Oxford, UK

<sup>14</sup>Long COVID SOS, Oxford, UK

<sup>15</sup>UKDoctors#Longcovid, London, UK

<sup>16</sup>Department of Medicine, University College London Hospitals NHS Foundation Trust, London, UK

<sup>17</sup>Institute of Health Informatics, University College London, London, UK

<sup>18</sup>Department of Cardiology, Barts Health NHS Trust, London, UK

**Twitter** Amitava Banerjee @amibanerjee1

**Contributors** Study design: AD, RB, JA, COVERSCAN team. Patient recruitment: RB, COVERSCAN team. Data collection: MW, COVERSCAN team. Data analysis: AD, AB, COVERSCAN team. Data interpretation: AB, AD, MW, RB. Initial manuscript drafting: AB, AD, RB. Critical review of early and final versions of the manuscript: all authors including JO and DJC. Specialist input: MW, AB (cardiology); RB, MH, DW, MC, DJC (general medicine); MH, MC, DW (long COVID-19); MB, RB (imaging); AD (statistics); AB (epidemiology/public health); MG (primary care); JA (healthcare management); LH, EA (patient and public involvement).

**Funding** This work was supported by the UK's National Consortium of Intelligent Medical Imaging (Industry Strategy Challenge Fund), Innovate UK (Grant 104688) and the European Union's Horizon 2020 research and innovation programme (agreement no 719445). The research was designed, conducted, analysed and interpreted by the authors independently of the funding sources.

**Competing interests** AD, RB and MB are employees of Perspectum.

**Patient consent for publication** Not required.

**Ethics approval** The study has received ethical approval (20/SC/0185).

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Data availability statement** Data are available upon reasonable request from the corresponding author.

**Supplemental material** This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those

of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

**Open access** This is an open access article distributed in accordance with the Creative Commons Attribution 4.0 Unported (CC BY 4.0) license, which permits others to copy, redistribute, remix, transform and build upon this work for any purpose, provided the original work is properly cited, a link to the licence is given, and indication of whether changes were made. See: <https://creativecommons.org/licenses/by/4.0/>.

#### ORCID iD

Amitava Banerjee <http://orcid.org/0000-0001-8741-3411>

#### REFERENCES

- World Health Organization. Clinical management of severe acute respiratory infection (SARI) when COVID-19 disease is suspected. interim guidance 13 March 2020, 2020. Available: <https://apps.who.int/iris/handle/10665/331446>
- Pavon AG, Meier D, Samim D, *et al*. First documentation of persistent SARS-CoV-2 infection presenting with late acute severe myocarditis. *Can J Cardiol* 2020;36:1326.e5–1326.e7.
- Puntmann VO, Carerj ML, Wieters I. Outcomes of cardiovascular magnetic resonance imaging in patients recently recovered from coronavirus disease 2019 (COVID-19). *JAMA Cardiol* 2019;2020:1265–73.
- Tabary M, Khanmohammadi S, Araghi F, *et al*. Pathologic features of COVID-19: a Concise review. *Pathol Res Pract* 2020;216:153097.
- Alqahtani SA, Schattenberg JM. Liver injury in COVID-19: the current evidence. *United European Gastroenterol J* 2020;8:509–19.
- Somasundaram NP, Ranathunga I, Ratnasamy V, *et al*. The impact of SARS-CoV-2 virus infection on the endocrine system. *J Endocr Soc* 2020;4:1–22.
- Farouk SS, Fiaccadori E, Cravedi P, *et al*. COVID-19 and the kidney: what we think we know so far and what we don't. *J Nephrol* 2020;33:1213–8.
- Lai A, Pasea L, Banerjee A. Estimating excess mortality in people with cancer and multimorbidity in the COVID-19 emergency. *BMJ Open* 2020;10:e043828.
- Banerjee A, Pasea L, Harris S, *et al*. Estimating excess 1-year mortality associated with the COVID-19 pandemic according to underlying conditions and age: a population-based cohort study. *The Lancet* 2020;395:1715–25.
- Banerjee A, Chen S, Pasea L. Excess deaths in people with cardiovascular diseases during the COVID-19 pandemic. *Eur J Prev Cardiol* 2020.
- Raman B, Cassar MP, Tunnicliffe EM, *et al*. Medium-Term effects of SARS-CoV-2 infection on multiple vital organs, exercise capacity, cognition, quality of life and mental health, post-hospital discharge. *EClinicalMedicine* 2021;31:100683.
- Horton R. Offline: COVID-19 is not a pandemic. *The Lancet* 2020;396:874.
- Shovlin CL, Vizcaychipi MP. Implications for COVID-19 triage from the ICNARC report of 2204 COVID-19 cases managed in UK adult intensive care units. *Emerg Med J* 2020;37:332–3.
- Docherty AB, Harrison EM, Green CA, *et al*. Features of 20 133 UK patients in hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: prospective observational cohort study. *BMJ* 2020;369:m1985–12.
- Williamson EJ, Walker AJ, Bhaskaran K, *et al*. Factors associated with COVID-19-related death using OpenSAFELY. *Nature* 2020;584:430–6.
- Office for National Statistics. The prevalence of long COVID symptoms and COVID-19 complications, 2020. Available: <https://www.ons.gov.uk/news/statementsandletters/theprevalenceoflongcovidssymptomsandcovid19complications>
- del Rio C, Collins LF, Malani P. Long-Term health consequences of COVID-19. *JAMA* 2020;324:1723.
- Carfi A, Bernabei R, Landi F, *et al*. Persistent symptoms in patients after acute COVID-19. *JAMA* 2020;324:603–5.
- Nabavi N. Long covid: how to define it and how to manage it. *BMJ* 2020;370:m3489.
- Greenhalgh T, Knight M, A'Court C, *et al*. Management of post-acute covid-19 in primary care. *BMJ* 2020;13:m3026.
- National Institute for Health Research. New risk prediction model could help improve guidance for people shielding from COVID-19, 2020. Available: <https://www.nihr.ac.uk/news/new-risk-prediction-model-could-help-improve-guidance-for-people-shielding-from-covid-19/25096>
- Hill AD, Fowler RA, Pinto R, *et al*. Long-term outcomes and healthcare utilization following critical illness – a population-based study. *Crit Care* 2016;20:1–10.
- Perrin R, Riste L, Hann M, *et al*. Into the looking glass: post-viral syndrome post COVID-19. *Med Hypotheses* 2020;144:110055.
- George PM, Barratt SL, Condliffe R, *et al*. Respiratory follow-up of patients with COVID-19 pneumonia. *Thorax* 2020;75:1009–16.
- Jiang DH, McCoy RG. Planning for the Post-COVID syndrome: how payers can mitigate long-term complications of the pandemic. *J Gen Intern Med* 2020;35:3036–9.
- Janssen MF, Pickard AS, Golicki D, *et al*. Measurement properties of the EQ-5D-5L compared to the EQ-5D-3L across eight patient groups: a multi-country study. *Qual Life Res* 2013;22:1717–27.
- Yorke J, Moosavi SH, Shuldham C, *et al*. Quantification of dyspnoea using descriptors: development and initial testing of the Dyspnoea-12. *Thorax* 2010;65:21–6.
- Hobbins A, Barry L, Kelleher D, *et al*. The health of the residents of Ireland: population norms for Ireland based on the EQ-5D-5L descriptive system – a cross sectional study. *HRB Open Res* 2018;1:22.
- Gupta A, Madhavan MV, Sehgal K, *et al*. Extrapulmonary manifestations of COVID-19. *Nat Med* 2020;26:1017–32.
- Palmer K, Monaco A, Kivipelto M, *et al*. The potential long-term impact of the COVID-19 outbreak on patients with non-communicable diseases in Europe: consequences for healthy ageing. *Aging Clin Exp Res* 2020;32:1189–94.
- Menni C, Valdes AM, Freidin MB, *et al*. Real-Time tracking of self-reported symptoms to predict potential COVID-19. *Nat Med* 2020;26:1037–40.
- Mandal S, Barnett J, Brill S, *et al*. "Long-COVID": a cross-sectional study of persisting symptoms, biomarker and imaging abnormalities following hospitalisation for COVID-19. *Thorax* 2020;thoraxjnl-2020-215818.
- Chau VQ, Giustino G, Mahmood K, *et al*. Cardiogenic shock and hyperinflammatory syndrome in young males with COVID-19. *Circ Hear Fail* 2020:556–9.
- Rajpal S, Tong MS, Borchers J, *et al*. Cardiovascular magnetic resonance findings in competitive athletes recovering from COVID-19 infection. *JAMA Cardiol* 2020:5–7.
- Mathew D, Giles JR, Baxter AE, *et al*. Deep immune profiling of COVID-19 patients reveals distinct immunotypes with therapeutic implications. *Science* 2020;369:eabc8511.
- Wynants L, Van Calster B, Collins GS, *et al*. Prediction models for diagnosis and prognosis of covid-19: systematic review and critical appraisal. *BMJ* 2020;369:m1328.
- PHOSP-COVID. Post-HOSPitalisation COVID-19 study, 2020. Available: <https://www.phosp.org/>
- NHS. NHS to offer 'long covid' sufferers help at specialist centres, 2020. Available: <https://www.england.nhs.uk/2020/10/nhs-to-offer-long-covid-help/>
- National Institute for Health and Care Excellence. COVID-19 rapid guideline: managing the long-term effects of COVID-19, 2020. Available: <https://www.nice.org.uk/guidance/ng188>
- Ferreira VM, Piechnik SK, Dall'Armellina E, *et al*. Native T1-mapping detects the location, extent and patterns of acute myocarditis without the need for gadolinium contrast agents. *J Cardiovasc Magn Reson* 2014;16:1–11.
- Gibson LM, Paul L, Chappell FM, *et al*. Potentially serious incidental findings on brain and body magnetic resonance imaging of apparently asymptomatic adults: systematic review and meta-analysis. *BMJ* 2018;14:k4577.

**Web Supplementary Materials**

Supplementary methods	2
Supplementary references	4
Supplementary results	6
Figure S1: Comparison of patients to control quantitative image derived measures in a subset of those scanned at 1.5T	6
Figure S2: Organ impairment in severe versus moderate post COVID syndrome (n=201)	7
Table S1: Reference ranges to define organ impairment	7
Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by hospitalisation or managed at home	8
Table S3: Blood investigations in 201 low-risk individuals, sub-divided by those with severe of moderate post-COVID syndrome	13



## **Supplementary methods**

### **Blood investigations**

Blood investigations included: full blood count, serum biochemistry (sodium, chloride, bicarbonate, urea, creatinine, bilirubin, alkaline phosphatase, aspartate transferase, alanine transferase, lactate dehydrogenase, creatinine kinase, gamma-glutamyl transpeptidase, total protein, albumin, globulin, calcium, magnesium, phosphate, uric acid, fasting triglycerides, cholesterol (total, HDL, LDL), iron, iron-binding capacity (unsaturated and total) and inflammatory markers (erythrocyte sedimentation rate, ESR; high sensitivity-C-Reactive Protein, CRP) (TDL laboratories, London).

### **Imaging**

All the imaging methods can be deployed on standard clinical MRI scanners and are generally expedited approaches of methods previously demonstrated in the scientific literature that unless stated each utilise a short (<14seconds) breath-hold.

Cardiac imaging involved complete coverage of the heart with a short-axis stack (to the valve plane) of cine images acquired using cardiac gating, this acquisition mirrors that in UK Biobank and is a standardized approach(S1). Three short-axis cardiac T1 maps are acquired using the MOLLI-T1 approach at the basal, mid and apical levels of the left ventricle.

Liver and pancreas imaging used the LiverMultiScan acquisition protocol (Perspectum, Oxford, UK), which involves 3 single 2D axial slice breath-held acquisitions that separately are sensitive to the fat content (proton density fat fraction, or PDFF), to T2\* (which is representative of liver iron content) and a MOLLI-T1 measurement (providing a measurement of tissue water), additionally a volumetric scan was used that covers the entire liver(S2).

Two dynamic cine MR acquisitions of the lung were acquired in the coronal plane with a 306.91 ms temporal resolution: one 40 s acquisition with the patient instructed to breathe normally and a second 30 s acquisition with the patient instructed to breathe deeply.

Kidney imaging used a single coronal view that was able to image both kidneys, imaging contrasts were MOLLI-T1, T2\* (for blood oxygen level assessment), and diffusion imaging that was acquired during free-breathing in 2minutes.

### **Image Analysis**

Cardiac MRI Analysis: Experienced cardiac MRI analysts used CVI42 (Cardiovascular Imaging Inc, Canada) to manually trace the end-diastolic and end-systolic phases in each of the short-axis views, following the standard UK BioBank evaluation approach as previously described(S3). This analysis yielded: For both the left and the right ventricle; End diastolic volume, End systolic volume, Stroke volume and Ejection Fraction. Additionally left ventricular muscle mass and wall thickness are determined from the function data. Cardiac T1 was determined for each of the 16 cardiac segments (of the AHA 17 segment model)(S4).

Liver Images were analysed by data analysts experienced at using the LiverMultiScan (Perspectum, Oxford, UK) software. This yielded global metrics in each liver of PDFF (proton density fat fraction), T2\*, and cT1 (cT1 is a measurement of T1 that has been corrected for the confounding effects of iron and standardised to 3 Tesla; it is elevated with disease).

Pancreas images were analysed in a similar manner to the above except the software used was not FDA-cleared and iron correction was not performed. The output T1 was standardized to 3 Tesla.

Lung cine imaging allowed the measurement of the area of the left and right lungs through the breathing cycle in the coronal plane, which used automated methods that were reviewed by image analysts. The periodicity of the area fluctuations was used to determine the respiratory rate. All analysis was performed in-house using MATLAB based tools. The method was validated by measuring the correlation between the change in area and the forced vital capacity, the latter being measured using spirometry.

Patient respiration was assessed by imaging a single 2D coronal slice of the lungs over 30 seconds using a dynamic cine MRI acquisition, during which the patient instructed to breathe deeply.

Kidney images were assessed using in-house tools to fit the parametric maps and allow trained analysts to make measurements. The T2\* maps were analysed by the Twelve Layer Concentric Object (TLCO) approach that generates a gradient of relaxation values, in the other evaluations the cortex and medulla were manually segmented using the MOLLI-T1 map or the b=0 (in the case of diffusion) to guide the boundary.

In all cases the volumetric assessments utilised an initial in-house developed machine-learning driven segmentation, and then a manual step that may be used to fine tune boundaries. This approach was also used in the body composition analysis, which for reasons of speed was performed only in a single slice (an axial view that passes through L3 of the spine) in this work.

### Supplementary references

- S1. Petersen SE, Matthews PM, Francis JM, Robson MD, Zemrak F, Boubertakh R, Young AA, Hudson S, Weale P, Garratt S, Collins R, Piechnik S, Neubauer S. UK Biobank's cardiovascular magnetic resonance protocol. *J Cardiovasc Magn Reson*. 2016 Feb 1;18:8.
- S2. Banerjee R, Pavlides M, Tunnicliffe EM, Piechnik SK, Sarania N, Philips R, Collier JD, Booth JC, Schneider JE, Wang LM, Delaney DW, Fleming KA, Robson MD, Barnes E, Neubauer S. Multiparametric magnetic resonance for the non-invasive diagnosis of liver disease. *J Hepatol*. 2014 Jan;60(1):69-77. doi: 10.1016/j.jhep.2013.09.002.
- S3. Petersen SE, Aung N, Sanghvi MM, Zemrak F, Fung K, Paiva JM, Francis JM, Khanji MY, Lukaschuk E, Lee AM, Carapella V, Kim YJ, Leeson P, Piechnik SK, Neubauer S. Reference ranges for cardiac structure and function using cardiovascular magnetic resonance (CMR) in Caucasians from the UK Biobank population cohort. *J Cardiovasc Magn Reson*. 2017 Feb 3;19(1):18.
- S4. Cerqueira MD, Weissman NJ, Dilsizian V, Jacobs AK, Kaul S, Laskey WK, Pennell DJ, Rumberger JA, Ryan T, Verani MS; American Heart Association Writing Group on Myocardial Segmentation and Registration for Cardiac Imaging. Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart. A statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation*. 2002 Jan 29;105(4):539-42.
- S5. Kawel-Boehm N, Maceira A, Valsangiacomo-Buechel ER, Vogel-Claussen J, Turkbey EB, Williams R, Plein S, Tee M, Eng J, Bluemke DA. Normal values for cardiovascular magnetic resonance in adults and children. *J Cardiovasc Magn Reson*. 2015 Apr 18;17(1):29.
- S6. Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P; ESC Scientific Document Group. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur Heart J*. 2016 Jul 14;37(27):2129-2200.
- S7. Tsao CW, Lyass A, Larson MG, Cheng S, Lam CS, Aragam JR, Benjamin EJ, Vasan RS. Prognosis of adults with borderline left ventricular ejection fraction. *JACC Heart Fail*. 2016 Jun;4(6):502-10.
- S8. Chalasani, Naga, et al. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 67.1 (2018): 328-357
- S9. Mojtahed A, Kelly C, Herlihy A, et al. Reference range of liver corrected T1 values in a population at low risk for fatty liver disease-a UK Biobank sub-study, with an appendix of interesting cases. *Abdominal Radiol* 2019; 44: 72–84.
- S10. Jayaswal AN, Levick C, Selvaraj EA, et al. Prognostic value of multiparametric MRI, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int* 2020; in press. DOI:doi:10.1111/liv.14625.
- S11. Jayaswal ANA, Levick C, Selvaraj EA, Dennis A, Booth JC, Collier J, Cobbold J, Tunnicliffe EM, Kelly M, Barnes E, Neubauer S, Banerjee R, Pavlides M. Prognostic value of multiparametric magnetic

resonance imaging, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int.* 2020 Jul 30. doi: 10.1111/liv.14625

S12. Chouhan MD, Firmin L, Read S, Amin Z, Taylor SA. Quantitative pancreatic MRI: a pathology-based review. *Br J Radiol.* 2019 Jul;92(1099):20180941.

S13. Harrington KA, Shukla-Dave A, Paudyal R, Do RKG. MRI of the Pancreas. *J Magn Reson Imaging.* 2020 Apr 17. doi: 10.1002/jmri.27148.

S14. Gillis KA, McComb C, Patel RK, et al. Non-contrast renal magnetic resonance imaging to assess perfusion and corticomedullary differentiation in health and chronic kidney disease. *Nephron* 2016; 133: 183–92.

S15. Peperhove M, Vo Chieu VD, Jang M-S, et al. Assessment of acute kidney injury with T1 mapping MRI following solid organ transplantation. *Eur Radiol* 2018; 28: 44–50.

S16. Chow KU, Luxembourg B, Seifried E, Bonig H. Spleen size is significantly influenced by body height and sex: establishment of normal values for spleen size at us with a cohort of 1200 healthy individuals. *Radiology* 2015; 279: 306–13.

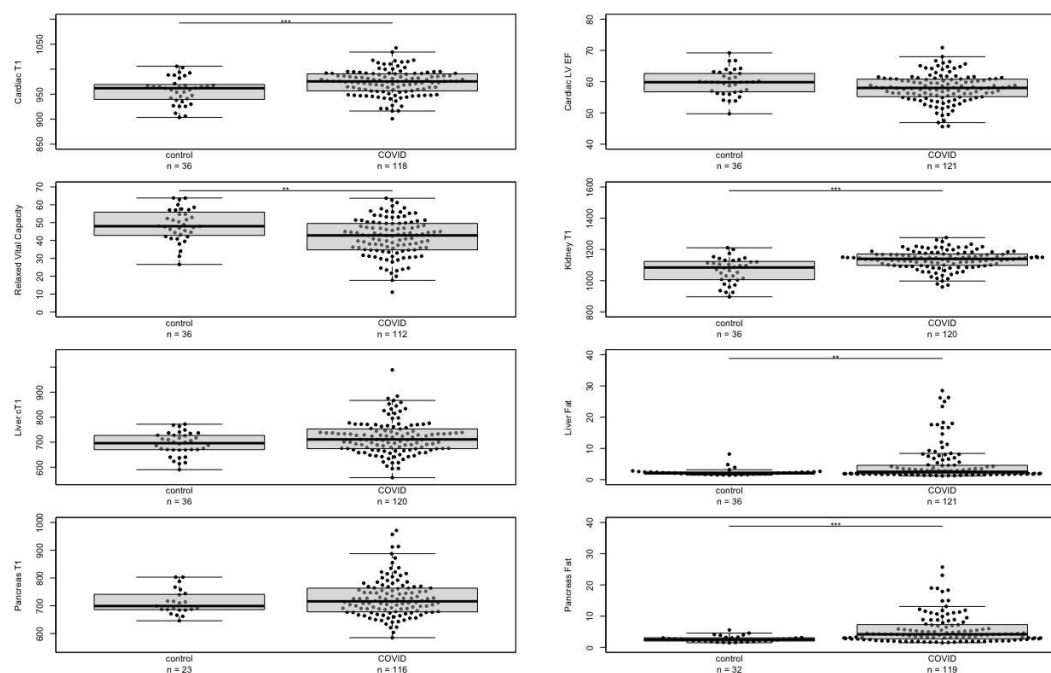


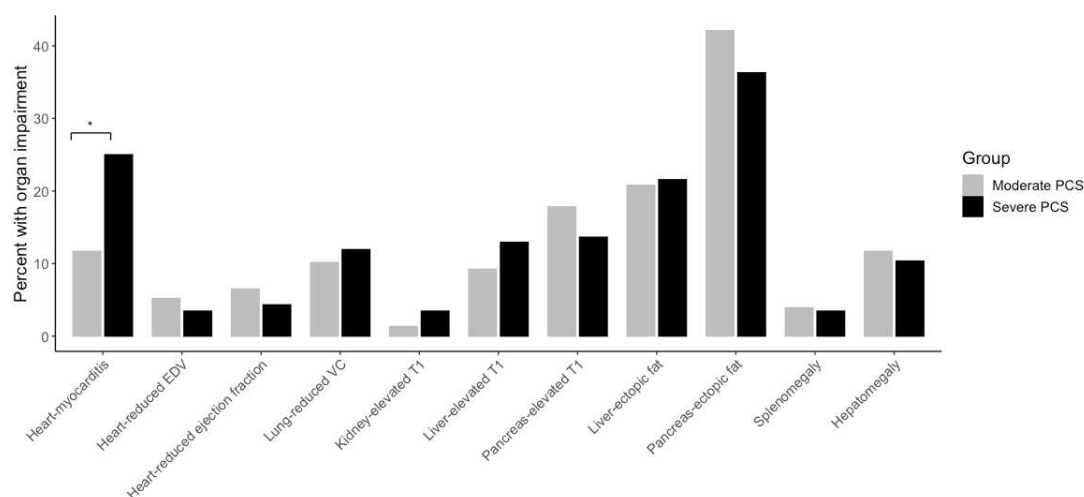
## Supplementary results

### Sub-group analysis

Data from healthy participants (n=36) scanned on the 1.5T Siemens MRI scanner were compared to the sub-group of patients (N=121) scanned on the same MRI machine. Median global cardiac T1 was elevated in the patient group (979 ms versus 962ms,  $P=0.001$ ). Lung fractional area difference, a measure of relaxed vital capacity, was significantly lower in the patient group (41% versus 48%,  $P<.001$ ). Kidney inflammation (1148 vs 1084 ms,  $p <0.001$ ) was significantly elevated in the patients as were markers of organ fat (liver 2.6% versus 2.1%,  $p=0.008$ ; pancreas: 4.3% versus 2.5%,  $p<0.001$ ) (**Figure S1**).

**Figure S1:** Box plots showing median and interquartile ranges for the healthy control group and the patient group for those scanned at 1.5T. Comparisons between groups were performed using two-sided Kolmogorov-Smirnov (KS) tests. Significance stars are \*  $P<.05$ ; \*\*  $P<.01$ , \*\*\* $P<.001$ .



**Figure S2:** Organ impairment in severe versus moderate post COVID syndrome (n=201)**Table S1:** Reference ranges for organ impairment, defined as a value that was greater than the mean plus 2 standard deviations of that from the control group for most; mean minus 2 standard deviations for left ventricular ejection fraction and lung fractional area difference for the 1.5T scans. For the 3T scans, this was the value as reported by Raman et al (2020).

	1.5T Reference range	3T reference range
Left ventricular ejection fraction (LVEF) (S4-S7)	≤ 51.5%	----
Increased end-diastolic volume (S4-S7)	≥ 264ml in men ≥ 206ml in women	----
Myocarditis (S4-S7)	≥ 1015 ms	≥ 1238ms
Deep breathing fractional area change*	≤ 31%	----
Liver volume (S8-S11)	≤ 1.93L	----
Liver fat (S8-S11)	≥ 4.8%	----
Liver inflammation (S8-S11)	≥ 784 ms	----
Pancreatic fat (S12-S13)	≥ 4.6%	----
Pancreatic inflammation (S12-13)	≥ 803ms	----
Renal Cortical T1(S14-S15)	≥ 1227ms	≥ 1652ms
Spleen volume(S16)	≤ 0.35L	----

\* Our lung imaging protocol captured 2D dynamic imaging of the lungs as the patient breathes. We delineated the lungs at maximum inspiration and again at maximum expiration and take the difference to give a proxy of 'vital capacity', which correlates well with forced vital capacity ( $r = 0.61$ ,  $P < .001$ ) from spirometry. Given the measure was associated with body size, we divided the difference in maximum inspiration and expiration by maximum inspiration to give a normalised 'lung ejection fraction'. In order to assess whether an individual's 'lung ejection fraction' was abnormal, it was measured in 39 controls, characterising a healthy normal range of the mean  $\pm$  2 standard deviations, with a lower score representing poorer lung health. 31% (0.31) was the lower limit for normal from our controls and therefore selected as the threshold for respiratory impairment.

**Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by those who were hospitalised versus those who were managed at home**

Measurement	All	Managed at home	Hospitalised	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.575
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.287
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	174 (97.8%)	142 (97.3%)	32 (100%)	1
• Abnormal low ( < 80 fL )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	174 (97.8%)	143 (97.9%)	31 (96.9%)	0.249
• Abnormal low ( < 26 pg )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	0 (0%)	1 (3.1%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	135 (75.8%)	109 (74.7%)	26 (81.2%)	0.501
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	43 (24.2%)	37 (25.3%)	6 (18.8%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	161 (91%)	129 (89%)	32 (100%)	0.218
• Abnormal low ( < 11.5 )	10 (5.6%)	10 (6.9%)	0 (0%)	
• Abnormal high ( > 15 )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	166 (93.3%)	138 (94.5%)	28 (87.5%)	0.152
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	177 (99.4%)	145 (99.3%)	32 (100%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 13 fL )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>White cell count</b>				

• Normal ( 3 - 10 x10 <sup>9</sup> /L )	172 (96.6%)	140 (95.9%)	32 (100%)	0.593
• Abnormal low ( < 3 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 10 x10 <sup>9</sup> /L )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Neutrophils</b>				
• Normal ( 2 - 7.5 x10 <sup>9</sup> /L )	163 (91.6%)	133 (91.1%)	30 (93.8%)	1
• Abnormal low ( < 2 x10 <sup>9</sup> /L )	12 (6.7%)	10 (6.8%)	2 (6.2%)	
• Abnormal high ( > 7.5 x10 <sup>9</sup> /L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Lymphocytes</b>				
• Normal ( 1.2 - 3.65 x10 <sup>9</sup> /L )	161 (90.4%)	130 (89%)	31 (96.9%)	0.316
• Abnormal low ( < 1.2 x10 <sup>9</sup> /L )	17 (9.6%)	16 (11%)	1 (3.1%)	
• Abnormal high ( > 3.65 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal ( 0.2 - 1 x10 <sup>9</sup> /L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.2 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Eosinophils</b>				
• Normal ( 0 - 0.4 x10 <sup>9</sup> /L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	1
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.4 x10 <sup>9</sup> /L )	6 (3.4%)	5 (3.4%)	1 (3.1%)	
<b>Basophils</b>				
• Normal ( 0 - 0.1 x10 <sup>9</sup> /L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.1 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal ( 1 - 20 mm/hr )	164 (91.1%)	136 (91.9%)	28 (87.5%)	0.491
• Abnormal low ( < 1 mm/hr )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 mm/hr )	16 (8.9%)	12 (8.1%)	4 (12.5%)	
<b>Sodium</b>				
• Normal ( 135 - 145 mmol/L )	173 (97.2%)	141 (96.6%)	32 (100%)	1
• Abnormal low ( < 135 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 145 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Potassium</b>				
• Normal ( 3.5 - 5.1 mmol/L )	108 (62.1%)	87 (61.3%)	21 (65.6%)	0.692
• Abnormal low ( < 3.5 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5.1 mmol/L )	66 (37.9%)	55 (38.7%)	11 (34.4%)	
<b>Chloride</b>				
• Normal ( 98 - 107 mmol/L )	171 (96.1%)	139 (95.2%)	32 (100%)	1
• Abnormal low ( < 98 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 107 mmol/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	150 (84.3%)	125 (85.6%)	25 (78.1%)	0.169
• Abnormal low ( < 22 mmol/L )	18 (10.1%)	15 (10.3%)	3 (9.4%)	
• Abnormal high ( > 29 mmol/L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Urea</b>				



• Normal ( 1.7 - 8.3 mmol/L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	161 (90.4%)	134 (91.8%)	27 (84.4%)	0.219
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.7%)	9 (6.2%)	3 (9.4%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	175 (98.3%)	144 (98.6%)	31 (96.9%)	0.45
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	168 (94.4%)	137 (93.8%)	31 (96.9%)	0.161
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	8 (4.5%)	8 (5.5%)	0 (0%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	162 (93.1%)	133 (93.7%)	29 (90.6%)	0.464
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (6.9%)	9 (6.3%)	3 (9.4%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	151 (84.8%)	125 (85.6%)	26 (81.2%)	0.603
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14%)	19 (13%)	6 (18.8%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	142 (80.7%)	118 (81.9%)	24 (75%)	0.236
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.8%)	5 (3.5%)	0 (0%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (16.5%)	21 (14.6%)	8 (25%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	163 (91.6%)	132 (90.4%)	31 (96.9%)	0.642
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	13 (7.3%)	12 (8.2%)	1 (3.1%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	165 (92.7%)	136 (93.2%)	29 (90.6%)	0.461
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.1%)	6 (4.1%)	3 (9.4%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	173 (97.2%)	143 (97.9%)	30 (93.8%)	0.22
• Abnormal low ( < 63 g/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
• Abnormal high ( > 83 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Albumin</b>				
• Normal ( 34 - 50 g/L )	167 (93.8%)	136 (93.2%)	31 (96.9%)	0.692
• Abnormal low ( < 34 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 50 g/L )	11 (6.2%)	10 (6.8%)	1 (3.1%)	
<b>Globulin</b>				

• Normal ( 19 - 35 g/L )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 19 g/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 35 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Calcium</b>				
• Normal ( 2.2 - 2.6 mmol/L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	0.43
• Abnormal low ( < 2.2 mmol/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 2.6 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
<b>Magnesium</b>				
• Normal ( 0.6 - 1 mmol/L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.6 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Phosphate</b>				
• Normal ( 0.87 - 1.45 mmol/L )	150 (84.3%)	121 (82.9%)	29 (90.6%)	0.518
• Abnormal low ( < 0.87 mmol/L )	23 (12.9%)	21 (14.4%)	2 (6.2%)	
• Abnormal high ( > 1.45 mmol/L )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
<b>Uric acid</b>				
• Normal ( 266 - 474 umol/L in men; 175 - 363 umol/L in women )	148 (83.1%)	124 (84.9%)	24 (75%)	0.067
• Abnormal low ( < 266 umol/L in men; < 175 umol/L in women )	19 (10.7%)	16 (11%)	3 (9.4%)	
• Abnormal high ( > 474 umol/L in men; > 363 umol/L in women )	11 (6.2%)	6 (4.1%)	5 (15.6%)	
<b>Triglycerides</b>				
• Normal ( < 2.3 mmol/L )	10 (100%)	8 (100%)	2 (100%)	N/A
• Abnormal high ( > 2.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal ( < 2.3 mmol/L )	149 (88.7%)	128 (92.8%)	21 (70%)	0.002
• Abnormal high ( > 2.3 mmol/L )	19 (11.3%)	10 (7.2%)	9 (30%)	
<b>Cholesterol</b>				
• Normal ( < 5 mmol/L )	4 (40%)	3 (37.5%)	1 (50%)	1
• Abnormal high ( > 5 mmol/L )	6 (60%)	5 (62.5%)	1 (50%)	
<b>Fasting cholesterol</b>				
• Normal ( < 5 mmol/L )	98 (58.3%)	86 (62.3%)	12 (40%)	0.04
• Abnormal high ( > 5 mmol/L )	70 (41.7%)	52 (37.7%)	18 (60%)	
<b>HDL cholesterol</b>				
• Normal ( 0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women )	106 (59.6%)	87 (59.6%)	19 (59.4%)	0.075
• Abnormal low ( < 0.9 mmol/L in men; < 1.2 mmol/L in women )	16 (9%)	10 (6.8%)	6 (18.8%)	
• Abnormal high ( > 1.5 mmol/L in men; > 1.7 mmol/L in women )	56 (31.5%)	49 (33.6%)	7 (21.9%)	
<b>LDL cholesterol</b>				
• Normal ( < 3 mmol/L )	113 (64.9%)	100 (69.4%)	13 (43.3%)	0.011
• Abnormal high ( > 3 mmol/L )	61 (35.1%)	44 (30.6%)	17 (56.7%)	
<b>Iron</b>				
• Normal ( 10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women )	164 (92.1%)	135 (92.5%)	29 (90.6%)	0.22
• Abnormal low ( < 10.6 umol/L in men; < 6.6 umol/L in women )	4 (2.2%)	2 (1.4%)	2 (6.2%)	

• Abnormal high ( > 28.3 umol/L in men; > 26 umol/L in women )	10 (5.6%)	9 (6.2%)	1 (3.1%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal ( 41 - 77 umol/L )	172 (97.2%)	141 (97.2%)	31 (96.9%)	1
• Abnormal low ( < 41 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 77 umol/L )	5 (2.8%)	4 (2.8%)	1 (3.1%)	
<b>Transferrin saturation</b>				
• Normal ( 20 - 55 % )	139 (78.5%)	120 (82.8%)	19 (59.4%)	0.011
• Abnormal low ( < 20 % )	34 (19.2%)	22 (15.2%)	12 (37.5%)	
• Abnormal high ( > 55 % )	4 (2.3%)	3 (2.1%)	1 (3.1%)	
<b>High sensitivity CRP</b>				
• Normal ( 0 - 5 mg/L )	146 (92.4%)	124 (93.9%)	22 (84.6%)	0.112
• Abnormal low ( < 0 mg/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5 mg/L )	12 (7.6%)	8 (6.1%)	4 (15.4%)	

**Table S3: Blood investigations in 201 low-risk individuals sub-divided by those with severe or moderate post-COVID syndrome (PCS)**

Measurement	All	Moderate PCS	Severe PCS	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	168 (97.1%)	64 (100%)	104 (95.4%)	0.274
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.825
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	4 (2.3%)	2 (3.1%)	2 (1.8%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	170 (98.3%)	62 (96.9%)	108 (99.1%)	0.556
• Abnormal low ( < 80 fL )	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	170 (98.3%)	61 (95.3%)	109 (100%)	0.049
• Abnormal low ( < 26 pg )	2 (1.2%)	2 (3.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	131 (75.7%)	53 (82.8%)	78 (71.6%)	0.103
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	42 (24.3%)	11 (17.2%)	31 (28.4%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	157 (91.3%)	59 (92.2%)	98 (90.7%)	0.339
• Abnormal low ( < 11.5 )	10 (5.8%)	2 (3.1%)	8 (7.4%)	
• Abnormal high ( > 15 )	5 (2.9%)	3 (4.7%)	2 (1.9%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	161 (93.1%)	59 (92.2%)	102 (93.6%)	0.417
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	172 (99.4%)	64 (100%)	108 (99.1%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	



• Abnormal high (> 13 fl)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>White cell count</b>				
• Normal (3 - 10 x10 <sup>9</sup> /L)	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.671
• Abnormal low (< 3 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 10 x10 <sup>9</sup> /L)	6 (3.5%)	3 (4.7%)	3 (2.8%)	
<b>Neutrophils</b>				
• Normal (2 - 7.5 x10 <sup>9</sup> /L)	159 (91.9%)	57 (89.1%)	102 (93.6%)	0.468
• Abnormal low (< 2 x10 <sup>9</sup> /L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
• Abnormal high (> 7.5 x10 <sup>9</sup> /L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
<b>Lymphocytes</b>				
• Normal (1.2 - 3.65 x10 <sup>9</sup> /L)	156 (90.2%)	56 (87.5%)	100 (91.7%)	0.43
• Abnormal low (< 1.2 x10 <sup>9</sup> /L)	17 (9.8%)	8 (12.5%)	9 (8.3%)	
• Abnormal high (> 3.65 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal (0.2 - 1 x10 <sup>9</sup> /L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.2 x10 <sup>9</sup> /L)	1 (0.6%)	0 (0%)	1 (0.9%)	
• Abnormal high (> 1 x10 <sup>9</sup> /L)	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Eosinophils</b>				
• Normal (0 - 0.4 x10 <sup>9</sup> /L)	167 (96.5%)	63 (98.4%)	104 (95.4%)	0.415
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.4 x10 <sup>9</sup> /L)	6 (3.5%)	1 (1.6%)	5 (4.6%)	
<b>Basophils</b>				
• Normal (0 - 0.1 x10 <sup>9</sup> /L)	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.1 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal (1 - 20 mm/hr)	160 (91.4%)	62 (93.9%)	98 (89.9%)	0.416
• Abnormal low (< 1 mm/hr)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 20 mm/hr)	15 (8.6%)	4 (6.1%)	11 (10.1%)	
<b>Sodium</b>				
• Normal (135 - 145 mmol/L)	168 (97.1%)	63 (98.4%)	105 (96.3%)	1
• Abnormal low (< 135 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 145 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Potassium</b>				
• Normal (3.5 - 5.1 mmol/L)	105 (62.1%)	35 (56.5%)	70 (65.4%)	0.255
• Abnormal low (< 3.5 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5.1 mmol/L)	64 (37.9%)	27 (43.5%)	37 (34.6%)	
<b>Chloride</b>				
• Normal (98 - 107 mmol/L)	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low (< 98 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 107 mmol/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	

<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	147 (85%)	55 (85.9%)	92 (84.4%)	0.946
• Abnormal low ( < 22 mmol/L )	16 (9.2%)	6 (9.4%)	10 (9.2%)	
• Abnormal high ( > 29 mmol/L )	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Urea</b>				
• Normal ( 1.7 - 8.3 mmol/L )	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	156 (90.2%)	59 (92.2%)	97 (89%)	0.705
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.9%)	3 (4.7%)	9 (8.3%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.9%)	2 (3.1%)	3 (2.8%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	170 (98.3%)	63 (98.4%)	107 (98.2%)	1
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	164 (94.8%)	59 (92.2%)	105 (96.3%)	0.185
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	7 (4%)	3 (4.7%)	4 (3.7%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.2%)	2 (3.1%)	0 (0%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	157 (92.9%)	59 (93.7%)	98 (92.5%)	1
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (7.1%)	4 (6.3%)	8 (7.5%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	146 (84.4%)	56 (87.5%)	90 (82.6%)	0.512
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14.5%)	7 (10.9%)	18 (16.5%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	137 (80.1%)	51 (81%)	86 (79.6%)	0.24
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.9%)	0 (0%)	5 (4.6%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (17%)	12 (19%)	17 (15.7%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	159 (91.9%)	56 (87.5%)	103 (94.5%)	0.28
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	12 (6.9%)	7 (10.9%)	5 (4.6%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	161 (93.1%)	60 (93.8%)	101 (92.7%)	0.426
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.2%)	4 (6.2%)	5 (4.6%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	168 (97.1%)	63 (98.4%)	105 (96.3%)	0.792

• Abnormal low (< 63 g/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	
• Abnormal high (> 83 g/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
<b>Albumin</b>				
• Normal (34 - 50 g/L)	162 (93.6%)	59 (92.2%)	103 (94.5%)	0.538
• Abnormal low (< 34 g/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 50 g/L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
<b>Globulin</b>				
• Normal (19 - 35 g/L)	168 (97.1%)	61 (95.3%)	107 (98.2%)	0.616
• Abnormal low (< 19 g/L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 35 g/L)	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Calcium</b>				
• Normal (2.2 - 2.6 mmol/L)	167 (96.5%)	62 (96.9%)	105 (96.3%)	0.525
• Abnormal low (< 2.2 mmol/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high (> 2.6 mmol/L)	4 (2.3%)	2 (3.1%)	2 (1.8%)	
<b>Magnesium</b>				
• Normal (0.6 - 1 mmol/L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.6 mmol/L)	1 (0.6%)	1 (1.6%)	0 (0%)	
• Abnormal high (> 1 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Phosphate</b>				
• Normal (0.87 - 1.45 mmol/L)	145 (83.8%)	55 (85.9%)	90 (82.6%)	0.824
• Abnormal low (< 0.87 mmol/L)	23 (13.3%)	8 (12.5%)	15 (13.8%)	
• Abnormal high (> 1.45 mmol/L)	5 (2.9%)	1 (1.6%)	4 (3.7%)	
<b>Uric acid</b>				
• Normal (266 - 474 umol/L in men; 175 - 363 umol/L in women)	145 (83.8%)	53 (82.8%)	92 (84.4%)	0.804
• Abnormal low (< 266 umol/L in men; < 175 umol/L in women)	18 (10.4%)	8 (12.5%)	10 (9.2%)	
• Abnormal high (> 474 umol/L in men; > 363 umol/L in women)	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Triglycerides</b>				
• Normal (< 2.3 mmol/L)	10 (100%)	6 (100%)	4 (100%)	N/A
• Abnormal high (> 2.3 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal (< 2.3 mmol/L)	144 (88.3%)	52 (89.7%)	92 (87.6%)	0.802
• Abnormal high (> 2.3 mmol/L)	19 (11.7%)	6 (10.3%)	13 (12.4%)	
<b>Cholesterol</b>				
• Normal (< 5 mmol/L)	4 (40%)	3 (50%)	1 (25%)	0.571
• Abnormal high (> 5 mmol/L)	6 (60%)	3 (50%)	3 (75%)	
<b>Fasting cholesterol</b>				
• Normal (< 5 mmol/L)	96 (58.9%)	39 (67.2%)	57 (54.3%)	0.135
• Abnormal high (> 5 mmol/L)	67 (41.1%)	19 (32.8%)	48 (45.7%)	
<b>HDL cholesterol</b>				
• Normal (0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women)	103 (59.5%)	38 (59.4%)	65 (59.6%)	0.539
• Abnormal low (< 0.9 mmol/L in men; < 1.2 mmol/L in women)	16 (9.2%)	4 (6.2%)	12 (11%)	

• Abnormal high (> 1.5 mmol/L in men; > 1.7 mmol/L in women)	54 (31.2%)	22 (34.4%)	32 (29.4%)	
<b>LDL cholesterol</b>				
• Normal (< 3 mmol/L)	111 (65.7%)	45 (72.6%)	66 (61.7%)	0.18
• Abnormal high (> 3 mmol/L)	58 (34.3%)	17 (27.4%)	41 (38.3%)	
<b>Iron</b>				
• Normal (10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women)	160 (92.5%)	57 (89.1%)	103 (94.5%)	0.337
• Abnormal low (< 10.6 umol/L in men; < 6.6 umol/L in women)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 28.3 umol/L in men; > 26 umol/L in women)	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal (41 - 77 umol/L)	167 (97.1%)	60 (93.8%)	107 (99.1%)	0.064
• Abnormal low (< 41 umol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 77 umol/L)	5 (2.9%)	4 (6.2%)	1 (0.9%)	
<b>Transferrin saturation</b>				
• Normal (20 - 55 %)	135 (78.5%)	50 (78.1%)	85 (78.7%)	0.283
• Abnormal low (< 20 %)	33 (19.2%)	11 (17.2%)	22 (20.4%)	
• Abnormal high (> 55 %)	4 (2.3%)	3 (4.7%)	1 (0.9%)	
<b>High sensitivity CRP</b>				
• Normal (0 - 5 mg/L)	141 (92.2%)	50 (96.2%)	91 (90.1%)	0.223
• Abnormal low (< 0 mg/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5 mg/L)	12 (7.8%)	2 (3.8%)	10 (9.9%)	

**Web Supplementary Materials**

Supplementary methods	2
Supplementary references	4
Supplementary results	6
Figure S1: Comparison of patients to control quantitative image derived measures in a subset of those scanned at 1.5T	6
Figure S2: Organ impairment in severe versus moderate post COVID syndrome (n=201)	7
Table S1: Reference ranges to define organ impairment	7
Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by hospitalisation or managed at home	8
Table S3: Blood investigations in 201 low-risk individuals, sub-divided by those with severe of moderate post-COVID syndrome	13



## **Supplementary methods**

### **Blood investigations**

Blood investigations included: full blood count, serum biochemistry (sodium, chloride, bicarbonate, urea, creatinine, bilirubin, alkaline phosphatase, aspartate transferase, alanine transferase, lactate dehydrogenase, creatinine kinase, gamma-glutamyl transpeptidase, total protein, albumin, globulin, calcium, magnesium, phosphate, uric acid, fasting triglycerides, cholesterol (total, HDL, LDL), iron, iron-binding capacity (unsaturated and total) and inflammatory markers (erythrocyte sedimentation rate, ESR; high sensitivity-C-Reactive Protein, CRP) (TDL laboratories, London).

### **Imaging**

All the imaging methods can be deployed on standard clinical MRI scanners and are generally expedited approaches of methods previously demonstrated in the scientific literature that unless stated each utilise a short (<14seconds) breath-hold.

Cardiac imaging involved complete coverage of the heart with a short-axis stack (to the valve plane) of cine images acquired using cardiac gating, this acquisition mirrors that in UK Biobank and is a standardized approach(S1). Three short-axis cardiac T1 maps are acquired using the MOLLI-T1 approach at the basal, mid and apical levels of the left ventricle.

Liver and pancreas imaging used the LiverMultiScan acquisition protocol (Perspectum, Oxford, UK), which involves 3 single 2D axial slice breath-held acquisitions that separately are sensitive to the fat content (proton density fat fraction, or PDFF), to T2\* (which is representative of liver iron content) and a MOLLI-T1 measurement (providing a measurement of tissue water), additionally a volumetric scan was used that covers the entire liver(S2).

Two dynamic cine MR acquisitions of the lung were acquired in the coronal plane with a 306.91 ms temporal resolution: one 40 s acquisition with the patient instructed to breathe normally and a second 30 s acquisition with the patient instructed to breathe deeply.

Kidney imaging used a single coronal view that was able to image both kidneys, imaging contrasts were MOLLI-T1, T2\* (for blood oxygen level assessment), and diffusion imaging that was acquired during free-breathing in 2minutes.

### **Image Analysis**

Cardiac MRI Analysis: Experienced cardiac MRI analysts used CVI42 (Cardiovascular Imaging Inc, Canada) to manually trace the end-diastolic and end-systolic phases in each of the short-axis views, following the standard UK BioBank evaluation approach as previously described(S3). This analysis yielded: For both the left and the right ventricle; End diastolic volume, End systolic volume, Stroke volume and Ejection Fraction. Additionally left ventricular muscle mass and wall thickness are determined from the function data. Cardiac T1 was determined for each of the 16 cardiac segments (of the AHA 17 segment model)(S4).

Liver Images were analysed by data analysts experienced at using the LiverMultiScan (Perspectum, Oxford, UK) software. This yielded global metrics in each liver of PDFF (proton density fat fraction), T2\*, and cT1 (cT1 is a measurement of T1 that has been corrected for the confounding effects of iron and standardised to 3 Tesla; it is elevated with disease).

Pancreas images were analysed in a similar manner to the above except the software used was not FDA-cleared and iron correction was not performed. The output T1 was standardized to 3 Tesla.

Lung cine imaging allowed the measurement of the area of the left and right lungs through the breathing cycle in the coronal plane, which used automated methods that were reviewed by image analysts. The periodicity of the area fluctuations was used to determine the respiratory rate. All analysis was performed in-house using MATLAB based tools. The method was validated by measuring the correlation between the change in area and the forced vital capacity, the latter being measured using spirometry.

Patient respiration was assessed by imaging a single 2D coronal slice of the lungs over 30 seconds using a dynamic cine MRI acquisition, during which the patient instructed to breathe deeply.

Kidney images were assessed using in-house tools to fit the parametric maps and allow trained analysts to make measurements. The T2\* maps were analysed by the Twelve Layer Concentric Object (TLCO) approach that generates a gradient of relaxation values, in the other evaluations the cortex and medulla were manually segmented using the MOLLI-T1 map or the b=0 (in the case of diffusion) to guide the boundary.

In all cases the volumetric assessments utilised an initial in-house developed machine-learning driven segmentation, and then a manual step that may be used to fine tune boundaries. This approach was also used in the body composition analysis, which for reasons of speed was performed only in a single slice (an axial view that passes through L3 of the spine) in this work.

### Supplementary references

- S1. Petersen SE, Matthews PM, Francis JM, Robson MD, Zemrak F, Boubertakh R, Young AA, Hudson S, Weale P, Garratt S, Collins R, Piechnik S, Neubauer S. UK Biobank's cardiovascular magnetic resonance protocol. *J Cardiovasc Magn Reson*. 2016 Feb 1;18:8.
- S2. Banerjee R, Pavlides M, Tunnicliffe EM, Piechnik SK, Sarania N, Philips R, Collier JD, Booth JC, Schneider JE, Wang LM, Delaney DW, Fleming KA, Robson MD, Barnes E, Neubauer S. Multiparametric magnetic resonance for the non-invasive diagnosis of liver disease. *J Hepatol*. 2014 Jan;60(1):69-77. doi: 10.1016/j.jhep.2013.09.002.
- S3. Petersen SE, Aung N, Sanghvi MM, Zemrak F, Fung K, Paiva JM, Francis JM, Khanji MY, Lukaschuk E, Lee AM, Carapella V, Kim YJ, Leeson P, Piechnik SK, Neubauer S. Reference ranges for cardiac structure and function using cardiovascular magnetic resonance (CMR) in Caucasians from the UK Biobank population cohort. *J Cardiovasc Magn Reson*. 2017 Feb 3;19(1):18.
- S4. Cerqueira MD, Weissman NJ, Dilsizian V, Jacobs AK, Kaul S, Laskey WK, Pennell DJ, Rumberger JA, Ryan T, Verani MS; American Heart Association Writing Group on Myocardial Segmentation and Registration for Cardiac Imaging. Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart. A statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation*. 2002 Jan 29;105(4):539-42.
- S5. Kawel-Boehm N, Maceira A, Valsangiacomo-Buechel ER, Vogel-Claussen J, Turkbey EB, Williams R, Plein S, Tee M, Eng J, Bluemke DA. Normal values for cardiovascular magnetic resonance in adults and children. *J Cardiovasc Magn Reson*. 2015 Apr 18;17(1):29.
- S6. Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P; ESC Scientific Document Group. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur Heart J*. 2016 Jul 14;37(27):2129-2200.
- S7. Tsao CW, Lyass A, Larson MG, Cheng S, Lam CS, Aragam JR, Benjamin EJ, Vasan RS. Prognosis of adults with borderline left ventricular ejection fraction. *JACC Heart Fail*. 2016 Jun;4(6):502-10.
- S8. Chalasani, Naga, et al. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 67.1 (2018): 328-357
- S9. Mojtahed A, Kelly C, Herlihy A, et al. Reference range of liver corrected T1 values in a population at low risk for fatty liver disease-a UK Biobank sub-study, with an appendix of interesting cases. *Abdominal Radiol* 2019; 44: 72–84.
- S10. Jayaswal AN, Levick C, Selvaraj EA, et al. Prognostic value of multiparametric MRI, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int* 2020; in press. DOI:doi:10.1111/liv.14625.
- S11. Jayaswal ANA, Levick C, Selvaraj EA, Dennis A, Booth JC, Collier J, Cobbold J, Tunnicliffe EM, Kelly M, Barnes E, Neubauer S, Banerjee R, Pavlides M. Prognostic value of multiparametric magnetic

resonance imaging, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int.* 2020 Jul 30. doi: 10.1111/liv.14625

S12. Chouhan MD, Firmin L, Read S, Amin Z, Taylor SA. Quantitative pancreatic MRI: a pathology-based review. *Br J Radiol.* 2019 Jul;92(1099):20180941.

S13. Harrington KA, Shukla-Dave A, Paudyal R, Do RKG. MRI of the Pancreas. *J Magn Reson Imaging.* 2020 Apr 17. doi: 10.1002/jmri.27148.

S14. Gillis KA, McComb C, Patel RK, et al. Non-contrast renal magnetic resonance imaging to assess perfusion and corticomedullary differentiation in health and chronic kidney disease. *Nephron* 2016; 133: 183–92.

S15. Peperhove M, Vo Chieu VD, Jang M-S, et al. Assessment of acute kidney injury with T1 mapping MRI following solid organ transplantation. *Eur Radiol* 2018; 28: 44–50.

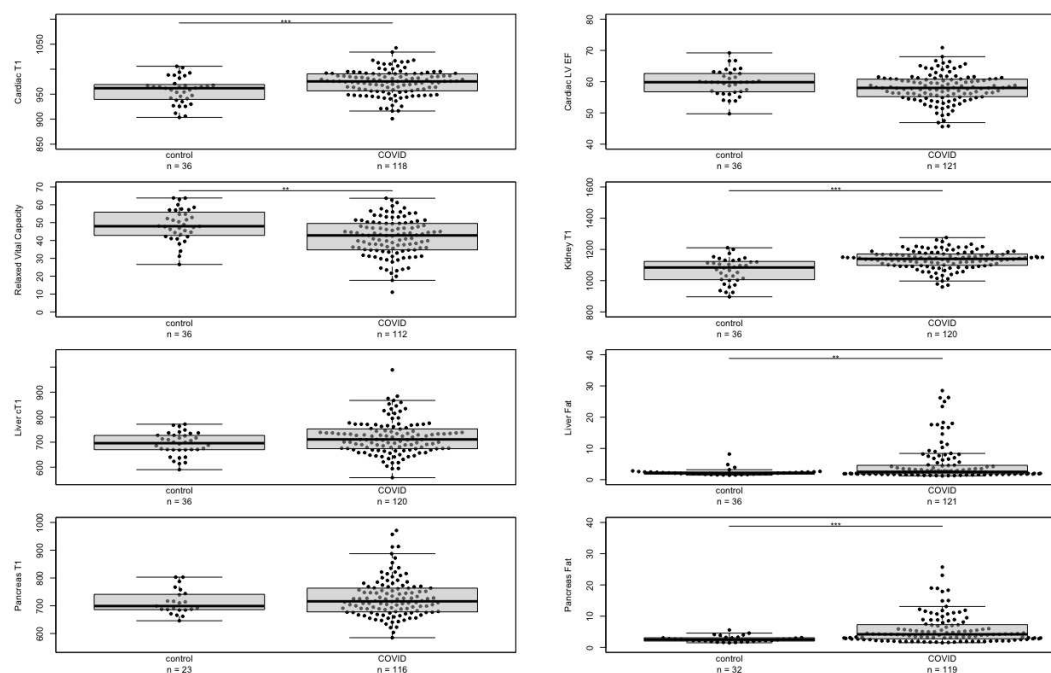
S16. Chow KU, Luxembourg B, Seifried E, Bonig H. Spleen size is significantly influenced by body height and sex: establishment of normal values for spleen size at us with a cohort of 1200 healthy individuals. *Radiology* 2015; 279: 306–13.

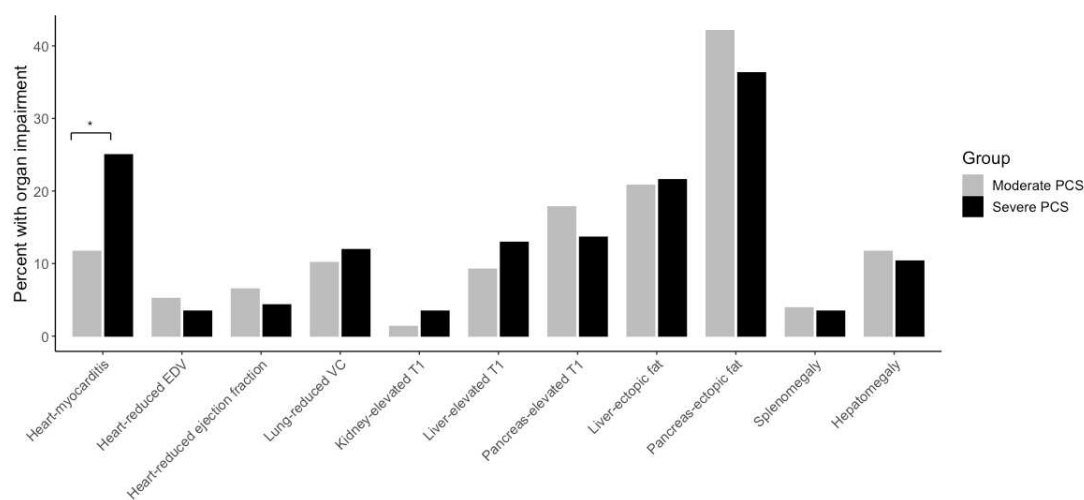
## Supplementary results

### Sub-group analysis

Data from healthy participants (n=36) scanned on the 1.5T Siemens MRI scanner were compared to the sub-group of patients (N=121) scanned on the same MRI machine. Median global cardiac T1 was elevated in the patient group (979 ms versus 962ms,  $P=0.001$ ). Lung fractional area difference, a measure of relaxed vital capacity, was significantly lower in the patient group (41% versus 48%,  $P<.001$ ). Kidney inflammation (1148 vs 1084 ms,  $p <0.001$ ) was significantly elevated in the patients as were markers of organ fat (liver 2.6% versus 2.1%,  $p=0.008$ ; pancreas: 4.3% versus 2.5%,  $p<0.001$ ) (**Figure S1**).

**Figure S1:** Box plots showing median and interquartile ranges for the healthy control group and the patient group for those scanned at 1.5T. Comparisons between groups were performed using two-sided Kolmogorov-Smirnov (KS) tests. Significance stars are \*  $P<.05$ ; \*\*  $P<.01$ , \*\*\* $P<.001$ .



**Figure S2:** Organ impairment in severe versus moderate post COVID syndrome (n=201)**Table S1:** Reference ranges for organ impairment, defined as a value that was greater than the mean plus 2 standard deviations of that from the control group for most; mean minus 2 standard deviations for left ventricular ejection fraction and lung fractional area difference for the 1.5T scans. For the 3T scans, this was the value as reported by Raman et al (2020).

	1.5T Reference range	3T reference range
Left ventricular ejection fraction (LVEF) (S4-S7)	≤ 51.5%	----
Increased end-diastolic volume (S4-S7)	≥ 264ml in men ≥ 206ml in women	----
Myocarditis (S4-S7)	≥ 1015 ms	≥ 1238ms
Deep breathing fractional area change*	≤ 31%	----
Liver volume (S8-S11)	≤ 1.93L	----
Liver fat (S8-S11)	≥ 4.8%	----
Liver inflammation (S8-S11)	≥ 784 ms	----
Pancreatic fat (S12-S13)	≥ 4.6%	----
Pancreatic inflammation (S12-13)	≥ 803ms	----
Renal Cortical T1(S14-S15)	≥ 1227ms	≥ 1652ms
Spleen volume(S16)	≤ 0.35L	----

\* Our lung imaging protocol captured 2D dynamic imaging of the lungs as the patient breathes. We delineated the lungs at maximum inspiration and again at maximum expiration and take the difference to give a proxy of 'vital capacity', which correlates well with forced vital capacity ( $r = 0.61$ ,  $P < .001$ ) from spirometry. Given the measure was associated with body size, we divided the difference in maximum inspiration and expiration by maximum inspiration to give a normalised 'lung ejection fraction'. In order to assess whether an individual's 'lung ejection fraction' was abnormal, it was measured in 39 controls, characterising a healthy normal range of the mean  $\pm$  2 standard deviations, with a lower score representing poorer lung health. 31% (0.31) was the lower limit for normal from our controls and therefore selected as the threshold for respiratory impairment.



**Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by those who were hospitalised versus those who were managed at home**

Measurement	All	Managed at home	Hospitalised	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.575
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.287
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	174 (97.8%)	142 (97.3%)	32 (100%)	1
• Abnormal low ( < 80 fL )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	174 (97.8%)	143 (97.9%)	31 (96.9%)	0.249
• Abnormal low ( < 26 pg )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	0 (0%)	1 (3.1%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	135 (75.8%)	109 (74.7%)	26 (81.2%)	0.501
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	43 (24.2%)	37 (25.3%)	6 (18.8%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	161 (91%)	129 (89%)	32 (100%)	0.218
• Abnormal low ( < 11.5 )	10 (5.6%)	10 (6.9%)	0 (0%)	
• Abnormal high ( > 15 )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	166 (93.3%)	138 (94.5%)	28 (87.5%)	0.152
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	177 (99.4%)	145 (99.3%)	32 (100%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 13 fL )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>White cell count</b>				

• Normal ( 3 - 10 x10 <sup>9</sup> /L )	172 (96.6%)	140 (95.9%)	32 (100%)	0.593
• Abnormal low ( < 3 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 10 x10 <sup>9</sup> /L )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Neutrophils</b>				
• Normal ( 2 - 7.5 x10 <sup>9</sup> /L )	163 (91.6%)	133 (91.1%)	30 (93.8%)	1
• Abnormal low ( < 2 x10 <sup>9</sup> /L )	12 (6.7%)	10 (6.8%)	2 (6.2%)	
• Abnormal high ( > 7.5 x10 <sup>9</sup> /L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Lymphocytes</b>				
• Normal ( 1.2 - 3.65 x10 <sup>9</sup> /L )	161 (90.4%)	130 (89%)	31 (96.9%)	0.316
• Abnormal low ( < 1.2 x10 <sup>9</sup> /L )	17 (9.6%)	16 (11%)	1 (3.1%)	
• Abnormal high ( > 3.65 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal ( 0.2 - 1 x10 <sup>9</sup> /L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.2 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Eosinophils</b>				
• Normal ( 0 - 0.4 x10 <sup>9</sup> /L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	1
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.4 x10 <sup>9</sup> /L )	6 (3.4%)	5 (3.4%)	1 (3.1%)	
<b>Basophils</b>				
• Normal ( 0 - 0.1 x10 <sup>9</sup> /L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.1 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal ( 1 - 20 mm/hr )	164 (91.1%)	136 (91.9%)	28 (87.5%)	0.491
• Abnormal low ( < 1 mm/hr )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 mm/hr )	16 (8.9%)	12 (8.1%)	4 (12.5%)	
<b>Sodium</b>				
• Normal ( 135 - 145 mmol/L )	173 (97.2%)	141 (96.6%)	32 (100%)	1
• Abnormal low ( < 135 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 145 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Potassium</b>				
• Normal ( 3.5 - 5.1 mmol/L )	108 (62.1%)	87 (61.3%)	21 (65.6%)	0.692
• Abnormal low ( < 3.5 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5.1 mmol/L )	66 (37.9%)	55 (38.7%)	11 (34.4%)	
<b>Chloride</b>				
• Normal ( 98 - 107 mmol/L )	171 (96.1%)	139 (95.2%)	32 (100%)	1
• Abnormal low ( < 98 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 107 mmol/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	150 (84.3%)	125 (85.6%)	25 (78.1%)	0.169
• Abnormal low ( < 22 mmol/L )	18 (10.1%)	15 (10.3%)	3 (9.4%)	
• Abnormal high ( > 29 mmol/L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Urea</b>				

• Normal ( 1.7 - 8.3 mmol/L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	161 (90.4%)	134 (91.8%)	27 (84.4%)	0.219
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.7%)	9 (6.2%)	3 (9.4%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	175 (98.3%)	144 (98.6%)	31 (96.9%)	0.45
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	168 (94.4%)	137 (93.8%)	31 (96.9%)	0.161
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	8 (4.5%)	8 (5.5%)	0 (0%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	162 (93.1%)	133 (93.7%)	29 (90.6%)	0.464
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (6.9%)	9 (6.3%)	3 (9.4%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	151 (84.8%)	125 (85.6%)	26 (81.2%)	0.603
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14%)	19 (13%)	6 (18.8%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	142 (80.7%)	118 (81.9%)	24 (75%)	0.236
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.8%)	5 (3.5%)	0 (0%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (16.5%)	21 (14.6%)	8 (25%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	163 (91.6%)	132 (90.4%)	31 (96.9%)	0.642
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	13 (7.3%)	12 (8.2%)	1 (3.1%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	165 (92.7%)	136 (93.2%)	29 (90.6%)	0.461
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.1%)	6 (4.1%)	3 (9.4%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	173 (97.2%)	143 (97.9%)	30 (93.8%)	0.22
• Abnormal low ( < 63 g/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
• Abnormal high ( > 83 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Albumin</b>				
• Normal ( 34 - 50 g/L )	167 (93.8%)	136 (93.2%)	31 (96.9%)	0.692
• Abnormal low ( < 34 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 50 g/L )	11 (6.2%)	10 (6.8%)	1 (3.1%)	
<b>Globulin</b>				

• Normal ( 19 - 35 g/L )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 19 g/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 35 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Calcium</b>				
• Normal ( 2.2 - 2.6 mmol/L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	0.43
• Abnormal low ( < 2.2 mmol/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 2.6 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
<b>Magnesium</b>				
• Normal ( 0.6 - 1 mmol/L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.6 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Phosphate</b>				
• Normal ( 0.87 - 1.45 mmol/L )	150 (84.3%)	121 (82.9%)	29 (90.6%)	0.518
• Abnormal low ( < 0.87 mmol/L )	23 (12.9%)	21 (14.4%)	2 (6.2%)	
• Abnormal high ( > 1.45 mmol/L )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
<b>Uric acid</b>				
• Normal ( 266 - 474 umol/L in men; 175 - 363 umol/L in women )	148 (83.1%)	124 (84.9%)	24 (75%)	0.067
• Abnormal low ( < 266 umol/L in men; < 175 umol/L in women )	19 (10.7%)	16 (11%)	3 (9.4%)	
• Abnormal high ( > 474 umol/L in men; > 363 umol/L in women )	11 (6.2%)	6 (4.1%)	5 (15.6%)	
<b>Triglycerides</b>				
• Normal ( < 2.3 mmol/L )	10 (100%)	8 (100%)	2 (100%)	N/A
• Abnormal high ( > 2.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal ( < 2.3 mmol/L )	149 (88.7%)	128 (92.8%)	21 (70%)	0.002
• Abnormal high ( > 2.3 mmol/L )	19 (11.3%)	10 (7.2%)	9 (30%)	
<b>Cholesterol</b>				
• Normal ( < 5 mmol/L )	4 (40%)	3 (37.5%)	1 (50%)	1
• Abnormal high ( > 5 mmol/L )	6 (60%)	5 (62.5%)	1 (50%)	
<b>Fasting cholesterol</b>				
• Normal ( < 5 mmol/L )	98 (58.3%)	86 (62.3%)	12 (40%)	0.04
• Abnormal high ( > 5 mmol/L )	70 (41.7%)	52 (37.7%)	18 (60%)	
<b>HDL cholesterol</b>				
• Normal ( 0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women )	106 (59.6%)	87 (59.6%)	19 (59.4%)	0.075
• Abnormal low ( < 0.9 mmol/L in men; < 1.2 mmol/L in women )	16 (9%)	10 (6.8%)	6 (18.8%)	
• Abnormal high ( > 1.5 mmol/L in men; > 1.7 mmol/L in women )	56 (31.5%)	49 (33.6%)	7 (21.9%)	
<b>LDL cholesterol</b>				
• Normal ( < 3 mmol/L )	113 (64.9%)	100 (69.4%)	13 (43.3%)	0.011
• Abnormal high ( > 3 mmol/L )	61 (35.1%)	44 (30.6%)	17 (56.7%)	
<b>Iron</b>				
• Normal ( 10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women )	164 (92.1%)	135 (92.5%)	29 (90.6%)	0.22
• Abnormal low ( < 10.6 umol/L in men; < 6.6 umol/L in women )	4 (2.2%)	2 (1.4%)	2 (6.2%)	

• Abnormal high ( > 28.3 umol/L in men; > 26 umol/L in women )	10 (5.6%)	9 (6.2%)	1 (3.1%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal ( 41 - 77 umol/L )	172 (97.2%)	141 (97.2%)	31 (96.9%)	1
• Abnormal low ( < 41 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 77 umol/L )	5 (2.8%)	4 (2.8%)	1 (3.1%)	
<b>Transferrin saturation</b>				
• Normal ( 20 - 55 % )	139 (78.5%)	120 (82.8%)	19 (59.4%)	0.011
• Abnormal low ( < 20 % )	34 (19.2%)	22 (15.2%)	12 (37.5%)	
• Abnormal high ( > 55 % )	4 (2.3%)	3 (2.1%)	1 (3.1%)	
<b>High sensitivity CRP</b>				
• Normal ( 0 - 5 mg/L )	146 (92.4%)	124 (93.9%)	22 (84.6%)	0.112
• Abnormal low ( < 0 mg/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5 mg/L )	12 (7.6%)	8 (6.1%)	4 (15.4%)	

**Table S3: Blood investigations in 201 low-risk individuals sub-divided by those with severe or moderate post-COVID syndrome (PCS)**

Measurement	All	Moderate PCS	Severe PCS	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	168 (97.1%)	64 (100%)	104 (95.4%)	0.274
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.825
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	4 (2.3%)	2 (3.1%)	2 (1.8%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	170 (98.3%)	62 (96.9%)	108 (99.1%)	0.556
• Abnormal low ( < 80 fL )	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	170 (98.3%)	61 (95.3%)	109 (100%)	0.049
• Abnormal low ( < 26 pg )	2 (1.2%)	2 (3.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	131 (75.7%)	53 (82.8%)	78 (71.6%)	0.103
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	42 (24.3%)	11 (17.2%)	31 (28.4%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	157 (91.3%)	59 (92.2%)	98 (90.7%)	0.339
• Abnormal low ( < 11.5 )	10 (5.8%)	2 (3.1%)	8 (7.4%)	
• Abnormal high ( > 15 )	5 (2.9%)	3 (4.7%)	2 (1.9%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	161 (93.1%)	59 (92.2%)	102 (93.6%)	0.417
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	172 (99.4%)	64 (100%)	108 (99.1%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	



• Abnormal high (> 13 fl)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>White cell count</b>				
• Normal (3 - 10 x10 <sup>9</sup> /L)	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.671
• Abnormal low (< 3 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 10 x10 <sup>9</sup> /L)	6 (3.5%)	3 (4.7%)	3 (2.8%)	
<b>Neutrophils</b>				
• Normal (2 - 7.5 x10 <sup>9</sup> /L)	159 (91.9%)	57 (89.1%)	102 (93.6%)	0.468
• Abnormal low (< 2 x10 <sup>9</sup> /L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
• Abnormal high (> 7.5 x10 <sup>9</sup> /L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
<b>Lymphocytes</b>				
• Normal (1.2 - 3.65 x10 <sup>9</sup> /L)	156 (90.2%)	56 (87.5%)	100 (91.7%)	0.43
• Abnormal low (< 1.2 x10 <sup>9</sup> /L)	17 (9.8%)	8 (12.5%)	9 (8.3%)	
• Abnormal high (> 3.65 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal (0.2 - 1 x10 <sup>9</sup> /L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.2 x10 <sup>9</sup> /L)	1 (0.6%)	0 (0%)	1 (0.9%)	
• Abnormal high (> 1 x10 <sup>9</sup> /L)	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Eosinophils</b>				
• Normal (0 - 0.4 x10 <sup>9</sup> /L)	167 (96.5%)	63 (98.4%)	104 (95.4%)	0.415
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.4 x10 <sup>9</sup> /L)	6 (3.5%)	1 (1.6%)	5 (4.6%)	
<b>Basophils</b>				
• Normal (0 - 0.1 x10 <sup>9</sup> /L)	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.1 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal (1 - 20 mm/hr)	160 (91.4%)	62 (93.9%)	98 (89.9%)	0.416
• Abnormal low (< 1 mm/hr)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 20 mm/hr)	15 (8.6%)	4 (6.1%)	11 (10.1%)	
<b>Sodium</b>				
• Normal (135 - 145 mmol/L)	168 (97.1%)	63 (98.4%)	105 (96.3%)	1
• Abnormal low (< 135 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 145 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Potassium</b>				
• Normal (3.5 - 5.1 mmol/L)	105 (62.1%)	35 (56.5%)	70 (65.4%)	0.255
• Abnormal low (< 3.5 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5.1 mmol/L)	64 (37.9%)	27 (43.5%)	37 (34.6%)	
<b>Chloride</b>				
• Normal (98 - 107 mmol/L)	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low (< 98 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 107 mmol/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	

<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	147 (85%)	55 (85.9%)	92 (84.4%)	0.946
• Abnormal low ( < 22 mmol/L )	16 (9.2%)	6 (9.4%)	10 (9.2%)	
• Abnormal high ( > 29 mmol/L )	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Urea</b>				
• Normal ( 1.7 - 8.3 mmol/L )	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	156 (90.2%)	59 (92.2%)	97 (89%)	0.705
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.9%)	3 (4.7%)	9 (8.3%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.9%)	2 (3.1%)	3 (2.8%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	170 (98.3%)	63 (98.4%)	107 (98.2%)	1
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	164 (94.8%)	59 (92.2%)	105 (96.3%)	0.185
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	7 (4%)	3 (4.7%)	4 (3.7%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.2%)	2 (3.1%)	0 (0%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	157 (92.9%)	59 (93.7%)	98 (92.5%)	1
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (7.1%)	4 (6.3%)	8 (7.5%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	146 (84.4%)	56 (87.5%)	90 (82.6%)	0.512
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14.5%)	7 (10.9%)	18 (16.5%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	137 (80.1%)	51 (81%)	86 (79.6%)	0.24
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.9%)	0 (0%)	5 (4.6%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (17%)	12 (19%)	17 (15.7%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	159 (91.9%)	56 (87.5%)	103 (94.5%)	0.28
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	12 (6.9%)	7 (10.9%)	5 (4.6%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	161 (93.1%)	60 (93.8%)	101 (92.7%)	0.426
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.2%)	4 (6.2%)	5 (4.6%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	168 (97.1%)	63 (98.4%)	105 (96.3%)	0.792

• Abnormal low (< 63 g/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	
• Abnormal high (> 83 g/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
<b>Albumin</b>				
• Normal (34 - 50 g/L)	162 (93.6%)	59 (92.2%)	103 (94.5%)	0.538
• Abnormal low (< 34 g/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 50 g/L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
<b>Globulin</b>				
• Normal (19 - 35 g/L)	168 (97.1%)	61 (95.3%)	107 (98.2%)	0.616
• Abnormal low (< 19 g/L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 35 g/L)	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Calcium</b>				
• Normal (2.2 - 2.6 mmol/L)	167 (96.5%)	62 (96.9%)	105 (96.3%)	0.525
• Abnormal low (< 2.2 mmol/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high (> 2.6 mmol/L)	4 (2.3%)	2 (3.1%)	2 (1.8%)	
<b>Magnesium</b>				
• Normal (0.6 - 1 mmol/L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.6 mmol/L)	1 (0.6%)	1 (1.6%)	0 (0%)	
• Abnormal high (> 1 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Phosphate</b>				
• Normal (0.87 - 1.45 mmol/L)	145 (83.8%)	55 (85.9%)	90 (82.6%)	0.824
• Abnormal low (< 0.87 mmol/L)	23 (13.3%)	8 (12.5%)	15 (13.8%)	
• Abnormal high (> 1.45 mmol/L)	5 (2.9%)	1 (1.6%)	4 (3.7%)	
<b>Uric acid</b>				
• Normal (266 - 474 umol/L in men; 175 - 363 umol/L in women)	145 (83.8%)	53 (82.8%)	92 (84.4%)	0.804
• Abnormal low (< 266 umol/L in men; < 175 umol/L in women)	18 (10.4%)	8 (12.5%)	10 (9.2%)	
• Abnormal high (> 474 umol/L in men; > 363 umol/L in women)	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Triglycerides</b>				
• Normal (< 2.3 mmol/L)	10 (100%)	6 (100%)	4 (100%)	N/A
• Abnormal high (> 2.3 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal (< 2.3 mmol/L)	144 (88.3%)	52 (89.7%)	92 (87.6%)	0.802
• Abnormal high (> 2.3 mmol/L)	19 (11.7%)	6 (10.3%)	13 (12.4%)	
<b>Cholesterol</b>				
• Normal (< 5 mmol/L)	4 (40%)	3 (50%)	1 (25%)	0.571
• Abnormal high (> 5 mmol/L)	6 (60%)	3 (50%)	3 (75%)	
<b>Fasting cholesterol</b>				
• Normal (< 5 mmol/L)	96 (58.9%)	39 (67.2%)	57 (54.3%)	0.135
• Abnormal high (> 5 mmol/L)	67 (41.1%)	19 (32.8%)	48 (45.7%)	
<b>HDL cholesterol</b>				
• Normal (0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women)	103 (59.5%)	38 (59.4%)	65 (59.6%)	0.539
• Abnormal low (< 0.9 mmol/L in men; < 1.2 mmol/L in women)	16 (9.2%)	4 (6.2%)	12 (11%)	

• Abnormal high (> 1.5 mmol/L in men; > 1.7 mmol/L in women)	54 (31.2%)	22 (34.4%)	32 (29.4%)	
<b>LDL cholesterol</b>				
• Normal (< 3 mmol/L)	111 (65.7%)	45 (72.6%)	66 (61.7%)	0.18
• Abnormal high (> 3 mmol/L)	58 (34.3%)	17 (27.4%)	41 (38.3%)	
<b>Iron</b>				
• Normal (10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women)	160 (92.5%)	57 (89.1%)	103 (94.5%)	0.337
• Abnormal low (< 10.6 umol/L in men; < 6.6 umol/L in women)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 28.3 umol/L in men; > 26 umol/L in women)	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal (41 - 77 umol/L)	167 (97.1%)	60 (93.8%)	107 (99.1%)	0.064
• Abnormal low (< 41 umol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 77 umol/L)	5 (2.9%)	4 (6.2%)	1 (0.9%)	
<b>Transferrin saturation</b>				
• Normal (20 - 55 %)	135 (78.5%)	50 (78.1%)	85 (78.7%)	0.283
• Abnormal low (< 20 %)	33 (19.2%)	11 (17.2%)	22 (20.4%)	
• Abnormal high (> 55 %)	4 (2.3%)	3 (4.7%)	1 (0.9%)	
<b>High sensitivity CRP</b>				
• Normal (0 - 5 mg/L)	141 (92.2%)	50 (96.2%)	91 (90.1%)	0.223
• Abnormal low (< 0 mg/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5 mg/L)	12 (7.8%)	2 (3.8%)	10 (9.9%)	

**Web Supplementary Materials**

Supplementary methods	2
Supplementary references	4
Supplementary results	6
Figure S1: Comparison of patients to control quantitative image derived measures in a subset of those scanned at 1.5T	6
Figure S2: Organ impairment in severe versus moderate post COVID syndrome (n=201)	7
Table S1: Reference ranges to define organ impairment	7
Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by hospitalisation or managed at home	8
Table S3: Blood investigations in 201 low-risk individuals, sub-divided by those with severe of moderate post-COVID syndrome	13

## **Supplementary methods**

### **Blood investigations**

Blood investigations included: full blood count, serum biochemistry (sodium, chloride, bicarbonate, urea, creatinine, bilirubin, alkaline phosphatase, aspartate transferase, alanine transferase, lactate dehydrogenase, creatinine kinase, gamma-glutamyl transpeptidase, total protein, albumin, globulin, calcium, magnesium, phosphate, uric acid, fasting triglycerides, cholesterol (total, HDL, LDL), iron, iron-binding capacity (unsaturated and total) and inflammatory markers (erythrocyte sedimentation rate, ESR; high sensitivity-C-Reactive Protein, CRP) (TDL laboratories, London).

### **Imaging**

All the imaging methods can be deployed on standard clinical MRI scanners and are generally expedited approaches of methods previously demonstrated in the scientific literature that unless stated each utilise a short (<14seconds) breath-hold.

Cardiac imaging involved complete coverage of the heart with a short-axis stack (to the valve plane) of cine images acquired using cardiac gating, this acquisition mirrors that in UK Biobank and is a standardized approach(S1). Three short-axis cardiac T1 maps are acquired using the MOLLI-T1 approach at the basal, mid and apical levels of the left ventricle.

Liver and pancreas imaging used the LiverMultiScan acquisition protocol (Perspectum, Oxford, UK), which involves 3 single 2D axial slice breath-held acquisitions that separately are sensitive to the fat content (proton density fat fraction, or PDFF), to T2\* (which is representative of liver iron content) and a MOLLI-T1 measurement (providing a measurement of tissue water), additionally a volumetric scan was used that covers the entire liver(S2).

Two dynamic cine MR acquisitions of the lung were acquired in the coronal plane with a 306.91 ms temporal resolution: one 40 s acquisition with the patient instructed to breathe normally and a second 30 s acquisition with the patient instructed to breathe deeply.

Kidney imaging used a single coronal view that was able to image both kidneys, imaging contrasts were MOLLI-T1, T2\* (for blood oxygen level assessment), and diffusion imaging that was acquired during free-breathing in 2minutes.

### **Image Analysis**

Cardiac MRI Analysis: Experienced cardiac MRI analysts used CVI42 (Cardiovascular Imaging Inc, Canada) to manually trace the end-diastolic and end-systolic phases in each of the short-axis views, following the standard UK BioBank evaluation approach as previously described(S3). This analysis yielded: For both the left and the right ventricle; End diastolic volume, End systolic volume, Stroke volume and Ejection Fraction. Additionally left ventricular muscle mass and wall thickness are determined from the function data. Cardiac T1 was determined for each of the 16 cardiac segments (of the AHA 17 segment model)(S4).

Liver Images were analysed by data analysts experienced at using the LiverMultiScan (Perspectum, Oxford, UK) software. This yielded global metrics in each liver of PDFF (proton density fat fraction), T2\*, and cT1 (cT1 is a measurement of T1 that has been corrected for the confounding effects of iron and standardised to 3 Tesla; it is elevated with disease).

Pancreas images were analysed in a similar manner to the above except the software used was not FDA-cleared and iron correction was not performed. The output T1 was standardized to 3 Tesla.



Lung cine imaging allowed the measurement of the area of the left and right lungs through the breathing cycle in the coronal plane, which used automated methods that were reviewed by image analysts. The periodicity of the area fluctuations was used to determine the respiratory rate. All analysis was performed in-house using MATLAB based tools. The method was validated by measuring the correlation between the change in area and the forced vital capacity, the latter being measured using spirometry.

Patient respiration was assessed by imaging a single 2D coronal slice of the lungs over 30 seconds using a dynamic cine MRI acquisition, during which the patient instructed to breathe deeply.

Kidney images were assessed using in-house tools to fit the parametric maps and allow trained analysts to make measurements. The T2\* maps were analysed by the Twelve Layer Concentric Object (TLCO) approach that generates a gradient of relaxation values, in the other evaluations the cortex and medulla were manually segmented using the MOLLI-T1 map or the b=0 (in the case of diffusion) to guide the boundary.

In all cases the volumetric assessments utilised an initial in-house developed machine-learning driven segmentation, and then a manual step that may be used to fine tune boundaries. This approach was also used in the body composition analysis, which for reasons of speed was performed only in a single slice (an axial view that passes through L3 of the spine) in this work.

### Supplementary references

- S1. Petersen SE, Matthews PM, Francis JM, Robson MD, Zemrak F, Boubertakh R, Young AA, Hudson S, Weale P, Garratt S, Collins R, Piechnik S, Neubauer S. UK Biobank's cardiovascular magnetic resonance protocol. *J Cardiovasc Magn Reson*. 2016 Feb 1;18:8.
- S2. Banerjee R, Pavlides M, Tunnicliffe EM, Piechnik SK, Sarania N, Philips R, Collier JD, Booth JC, Schneider JE, Wang LM, Delaney DW, Fleming KA, Robson MD, Barnes E, Neubauer S. Multiparametric magnetic resonance for the non-invasive diagnosis of liver disease. *J Hepatol*. 2014 Jan;60(1):69-77. doi: 10.1016/j.jhep.2013.09.002.
- S3. Petersen SE, Aung N, Sanghvi MM, Zemrak F, Fung K, Paiva JM, Francis JM, Khanji MY, Lukaschuk E, Lee AM, Carapella V, Kim YJ, Leeson P, Piechnik SK, Neubauer S. Reference ranges for cardiac structure and function using cardiovascular magnetic resonance (CMR) in Caucasians from the UK Biobank population cohort. *J Cardiovasc Magn Reson*. 2017 Feb 3;19(1):18.
- S4. Cerqueira MD, Weissman NJ, Dilsizian V, Jacobs AK, Kaul S, Laskey WK, Pennell DJ, Rumberger JA, Ryan T, Verani MS; American Heart Association Writing Group on Myocardial Segmentation and Registration for Cardiac Imaging. Standardized myocardial segmentation and nomenclature for tomographic imaging of the heart. A statement for healthcare professionals from the Cardiac Imaging Committee of the Council on Clinical Cardiology of the American Heart Association. *Circulation*. 2002 Jan 29;105(4):539-42.
- S5. Kawel-Boehm N, Maceira A, Valsangiacomo-Buechel ER, Vogel-Claussen J, Turkbey EB, Williams R, Plein S, Tee M, Eng J, Bluemke DA. Normal values for cardiovascular magnetic resonance in adults and children. *J Cardiovasc Magn Reson*. 2015 Apr 18;17(1):29.
- S6. Ponikowski P, Voors AA, Anker SD, Bueno H, Cleland JGF, Coats AJS, Falk V, González-Juanatey JR, Harjola VP, Jankowska EA, Jessup M, Linde C, Nihoyannopoulos P, Parissis JT, Pieske B, Riley JP, Rosano GMC, Ruilope LM, Ruschitzka F, Rutten FH, van der Meer P; ESC Scientific Document Group. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur Heart J*. 2016 Jul 14;37(27):2129-2200.
- S7. Tsao CW, Lyass A, Larson MG, Cheng S, Lam CS, Aragam JR, Benjamin EJ, Vasan RS. Prognosis of adults with borderline left ventricular ejection fraction. *JACC Heart Fail*. 2016 Jun;4(6):502-10.
- S8. Chalasani, Naga, et al. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 67.1 (2018): 328-357
- S9. Mojtahed A, Kelly C, Herlihy A, et al. Reference range of liver corrected T1 values in a population at low risk for fatty liver disease-a UK Biobank sub-study, with an appendix of interesting cases. *Abdominal Radiol* 2019; 44: 72–84.
- S10. Jayaswal AN, Levick C, Selvaraj EA, et al. Prognostic value of multiparametric MRI, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int* 2020; in press. DOI:doi:10.1111/liv.14625.
- S11. Jayaswal ANA, Levick C, Selvaraj EA, Dennis A, Booth JC, Collier J, Cobbold J, Tunnicliffe EM, Kelly M, Barnes E, Neubauer S, Banerjee R, Pavlides M. Prognostic value of multiparametric magnetic

resonance imaging, transient elastography and blood-based fibrosis markers in patients with chronic liver disease. *Liver Int.* 2020 Jul 30. doi: 10.1111/liv.14625

S12. Chouhan MD, Firmin L, Read S, Amin Z, Taylor SA. Quantitative pancreatic MRI: a pathology-based review. *Br J Radiol.* 2019 Jul;92(1099):20180941.

S13. Harrington KA, Shukla-Dave A, Paudyal R, Do RKG. MRI of the Pancreas. *J Magn Reson Imaging.* 2020 Apr 17. doi: 10.1002/jmri.27148.

S14. Gillis KA, McComb C, Patel RK, et al. Non-contrast renal magnetic resonance imaging to assess perfusion and corticomedullary differentiation in health and chronic kidney disease. *Nephron* 2016; 133: 183–92.

S15. Peperhove M, Vo Chieu VD, Jang M-S, et al. Assessment of acute kidney injury with T1 mapping MRI following solid organ transplantation. *Eur Radiol* 2018; 28: 44–50.

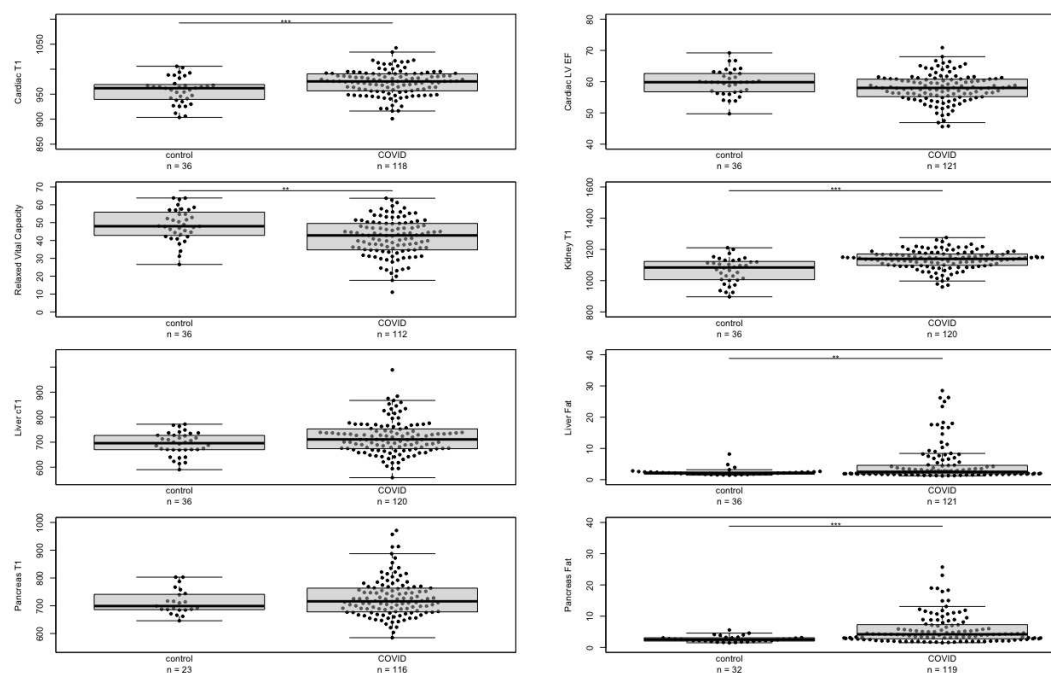
S16. Chow KU, Luxembourg B, Seifried E, Bonig H. Spleen size is significantly influenced by body height and sex: establishment of normal values for spleen size at us with a cohort of 1200 healthy individuals. *Radiology* 2015; 279: 306–13.

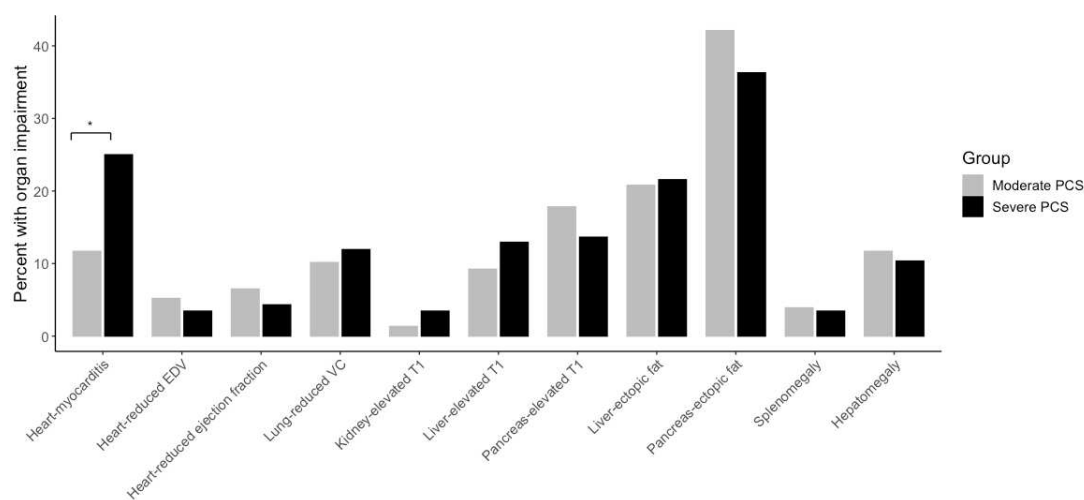
## Supplementary results

### Sub-group analysis

Data from healthy participants (n=36) scanned on the 1.5T Siemens MRI scanner were compared to the sub-group of patients (N=121) scanned on the same MRI machine. Median global cardiac T1 was elevated in the patient group (979 ms versus 962ms,  $P=0.001$ ). Lung fractional area difference, a measure of relaxed vital capacity, was significantly lower in the patient group (41% versus 48%,  $P<.001$ ). Kidney inflammation (1148 vs 1084 ms,  $p <0.001$ ) was significantly elevated in the patients as were markers of organ fat (liver 2.6% versus 2.1%,  $p=0.008$ ; pancreas: 4.3% versus 2.5%,  $p<0.001$ ) (**Figure S1**).

**Figure S1:** Box plots showing median and interquartile ranges for the healthy control group and the patient group for those scanned at 1.5T. Comparisons between groups were performed using two-sided Kolmogorov-Smirnov (KS) tests. Significance stars are \*  $P<.05$ ; \*\*  $P<.01$ , \*\*\* $P<.001$ .



**Figure S2:** Organ impairment in severe versus moderate post COVID syndrome (n=201)**Table S1:** Reference ranges for organ impairment, defined as a value that was greater than the mean plus 2 standard deviations of that from the control group for most; mean minus 2 standard deviations for left ventricular ejection fraction and lung fractional area difference for the 1.5T scans. For the 3T scans, this was the value as reported by Raman et al (2020).

	1.5T Reference range	3T reference range
Left ventricular ejection fraction (LVEF) (S4-S7)	≤ 51.5%	----
Increased end-diastolic volume (S4-S7)	≥ 264ml in men ≥ 206ml in women	----
Myocarditis (S4-S7)	≥ 1015 ms	≥ 1238ms
Deep breathing fractional area change*	≤ 31%	----
Liver volume (S8-S11)	≤ 1.93L	----
Liver fat (S8-S11)	≥ 4.8%	----
Liver inflammation (S8-S11)	≥ 784 ms	----
Pancreatic fat (S12-S13)	≥ 4.6%	----
Pancreatic inflammation (S12-13)	≥ 803ms	----
Renal Cortical T1(S14-S15)	≥ 1227ms	≥ 1652ms
Spleen volume(S16)	≤ 0.35L	----

\* Our lung imaging protocol captured 2D dynamic imaging of the lungs as the patient breathes. We delineated the lungs at maximum inspiration and again at maximum expiration and take the difference to give a proxy of 'vital capacity', which correlates well with forced vital capacity ( $r = 0.61$ ,  $P < .001$ ) from spirometry. Given the measure was associated with body size, we divided the difference in maximum inspiration and expiration by maximum inspiration to give a normalised 'lung ejection fraction'. In order to assess whether an individual's 'lung ejection fraction' was abnormal, it was measured in 39 controls, characterising a healthy normal range of the mean  $\pm$  2 standard deviations, with a lower score representing poorer lung health. 31% (0.31) was the lower limit for normal from our controls and therefore selected as the threshold for respiratory impairment.

**Table S2: Blood investigations in 201 low-risk individuals with post-COVID syndrome, sub-divided by those who were hospitalised versus those who were managed at home**

Measurement	All	Managed at home	Hospitalised	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.575
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	170 (95.5%)	140 (95.9%)	30 (93.8%)	0.287
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	174 (97.8%)	142 (97.3%)	32 (100%)	1
• Abnormal low ( < 80 fL )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	174 (97.8%)	143 (97.9%)	31 (96.9%)	0.249
• Abnormal low ( < 26 pg )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	0 (0%)	1 (3.1%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	135 (75.8%)	109 (74.7%)	26 (81.2%)	0.501
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	43 (24.2%)	37 (25.3%)	6 (18.8%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	161 (91%)	129 (89%)	32 (100%)	0.218
• Abnormal low ( < 11.5 )	10 (5.6%)	10 (6.9%)	0 (0%)	
• Abnormal high ( > 15 )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	166 (93.3%)	138 (94.5%)	28 (87.5%)	0.152
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	177 (99.4%)	145 (99.3%)	32 (100%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 13 fL )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>White cell count</b>				

• Normal ( 3 - 10 x10 <sup>9</sup> /L )	172 (96.6%)	140 (95.9%)	32 (100%)	0.593
• Abnormal low ( < 3 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 10 x10 <sup>9</sup> /L )	6 (3.4%)	6 (4.1%)	0 (0%)	
<b>Neutrophils</b>				
• Normal ( 2 - 7.5 x10 <sup>9</sup> /L )	163 (91.6%)	133 (91.1%)	30 (93.8%)	1
• Abnormal low ( < 2 x10 <sup>9</sup> /L )	12 (6.7%)	10 (6.8%)	2 (6.2%)	
• Abnormal high ( > 7.5 x10 <sup>9</sup> /L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Lymphocytes</b>				
• Normal ( 1.2 - 3.65 x10 <sup>9</sup> /L )	161 (90.4%)	130 (89%)	31 (96.9%)	0.316
• Abnormal low ( < 1.2 x10 <sup>9</sup> /L )	17 (9.6%)	16 (11%)	1 (3.1%)	
• Abnormal high ( > 3.65 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal ( 0.2 - 1 x10 <sup>9</sup> /L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.2 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 x10 <sup>9</sup> /L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Eosinophils</b>				
• Normal ( 0 - 0.4 x10 <sup>9</sup> /L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	1
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.4 x10 <sup>9</sup> /L )	6 (3.4%)	5 (3.4%)	1 (3.1%)	
<b>Basophils</b>				
• Normal ( 0 - 0.1 x10 <sup>9</sup> /L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 0 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 0.1 x10 <sup>9</sup> /L )	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal ( 1 - 20 mm/hr )	164 (91.1%)	136 (91.9%)	28 (87.5%)	0.491
• Abnormal low ( < 1 mm/hr )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 mm/hr )	16 (8.9%)	12 (8.1%)	4 (12.5%)	
<b>Sodium</b>				
• Normal ( 135 - 145 mmol/L )	173 (97.2%)	141 (96.6%)	32 (100%)	1
• Abnormal low ( < 135 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 145 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Potassium</b>				
• Normal ( 3.5 - 5.1 mmol/L )	108 (62.1%)	87 (61.3%)	21 (65.6%)	0.692
• Abnormal low ( < 3.5 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5.1 mmol/L )	66 (37.9%)	55 (38.7%)	11 (34.4%)	
<b>Chloride</b>				
• Normal ( 98 - 107 mmol/L )	171 (96.1%)	139 (95.2%)	32 (100%)	1
• Abnormal low ( < 98 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 107 mmol/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	150 (84.3%)	125 (85.6%)	25 (78.1%)	0.169
• Abnormal low ( < 22 mmol/L )	18 (10.1%)	15 (10.3%)	3 (9.4%)	
• Abnormal high ( > 29 mmol/L )	10 (5.6%)	6 (4.1%)	4 (12.5%)	
<b>Urea</b>				



• Normal ( 1.7 - 8.3 mmol/L )	178 (100%)	146 (100%)	32 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	161 (90.4%)	134 (91.8%)	27 (84.4%)	0.219
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.7%)	9 (6.2%)	3 (9.4%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.8%)	3 (2.1%)	2 (6.2%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	175 (98.3%)	144 (98.6%)	31 (96.9%)	0.45
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	168 (94.4%)	137 (93.8%)	31 (96.9%)	0.161
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	8 (4.5%)	8 (5.5%)	0 (0%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	162 (93.1%)	133 (93.7%)	29 (90.6%)	0.464
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (6.9%)	9 (6.3%)	3 (9.4%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	151 (84.8%)	125 (85.6%)	26 (81.2%)	0.603
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14%)	19 (13%)	6 (18.8%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	142 (80.7%)	118 (81.9%)	24 (75%)	0.236
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.8%)	5 (3.5%)	0 (0%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (16.5%)	21 (14.6%)	8 (25%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	163 (91.6%)	132 (90.4%)	31 (96.9%)	0.642
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.1%)	2 (1.4%)	0 (0%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	13 (7.3%)	12 (8.2%)	1 (3.1%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	165 (92.7%)	136 (93.2%)	29 (90.6%)	0.461
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	4 (2.2%)	4 (2.7%)	0 (0%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.1%)	6 (4.1%)	3 (9.4%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	173 (97.2%)	143 (97.9%)	30 (93.8%)	0.22
• Abnormal low ( < 63 g/L )	3 (1.7%)	2 (1.4%)	1 (3.1%)	
• Abnormal high ( > 83 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Albumin</b>				
• Normal ( 34 - 50 g/L )	167 (93.8%)	136 (93.2%)	31 (96.9%)	0.692
• Abnormal low ( < 34 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 50 g/L )	11 (6.2%)	10 (6.8%)	1 (3.1%)	
<b>Globulin</b>				

• Normal ( 19 - 35 g/L )	173 (97.2%)	142 (97.3%)	31 (96.9%)	0.386
• Abnormal low ( < 19 g/L )	3 (1.7%)	3 (2.1%)	0 (0%)	
• Abnormal high ( > 35 g/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
<b>Calcium</b>				
• Normal ( 2.2 - 2.6 mmol/L )	172 (96.6%)	141 (96.6%)	31 (96.9%)	0.43
• Abnormal low ( < 2.2 mmol/L )	2 (1.1%)	1 (0.7%)	1 (3.1%)	
• Abnormal high ( > 2.6 mmol/L )	4 (2.2%)	4 (2.7%)	0 (0%)	
<b>Magnesium</b>				
• Normal ( 0.6 - 1 mmol/L )	176 (98.9%)	144 (98.6%)	32 (100%)	1
• Abnormal low ( < 0.6 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
• Abnormal high ( > 1 mmol/L )	1 (0.6%)	1 (0.7%)	0 (0%)	
<b>Phosphate</b>				
• Normal ( 0.87 - 1.45 mmol/L )	150 (84.3%)	121 (82.9%)	29 (90.6%)	0.518
• Abnormal low ( < 0.87 mmol/L )	23 (12.9%)	21 (14.4%)	2 (6.2%)	
• Abnormal high ( > 1.45 mmol/L )	5 (2.8%)	4 (2.7%)	1 (3.1%)	
<b>Uric acid</b>				
• Normal ( 266 - 474 umol/L in men; 175 - 363 umol/L in women )	148 (83.1%)	124 (84.9%)	24 (75%)	0.067
• Abnormal low ( < 266 umol/L in men; < 175 umol/L in women )	19 (10.7%)	16 (11%)	3 (9.4%)	
• Abnormal high ( > 474 umol/L in men; > 363 umol/L in women )	11 (6.2%)	6 (4.1%)	5 (15.6%)	
<b>Triglycerides</b>				
• Normal ( < 2.3 mmol/L )	10 (100%)	8 (100%)	2 (100%)	N/A
• Abnormal high ( > 2.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal ( < 2.3 mmol/L )	149 (88.7%)	128 (92.8%)	21 (70%)	0.002
• Abnormal high ( > 2.3 mmol/L )	19 (11.3%)	10 (7.2%)	9 (30%)	
<b>Cholesterol</b>				
• Normal ( < 5 mmol/L )	4 (40%)	3 (37.5%)	1 (50%)	1
• Abnormal high ( > 5 mmol/L )	6 (60%)	5 (62.5%)	1 (50%)	
<b>Fasting cholesterol</b>				
• Normal ( < 5 mmol/L )	98 (58.3%)	86 (62.3%)	12 (40%)	0.04
• Abnormal high ( > 5 mmol/L )	70 (41.7%)	52 (37.7%)	18 (60%)	
<b>HDL cholesterol</b>				
• Normal ( 0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women )	106 (59.6%)	87 (59.6%)	19 (59.4%)	0.075
• Abnormal low ( < 0.9 mmol/L in men; < 1.2 mmol/L in women )	16 (9%)	10 (6.8%)	6 (18.8%)	
• Abnormal high ( > 1.5 mmol/L in men; > 1.7 mmol/L in women )	56 (31.5%)	49 (33.6%)	7 (21.9%)	
<b>LDL cholesterol</b>				
• Normal ( < 3 mmol/L )	113 (64.9%)	100 (69.4%)	13 (43.3%)	0.011
• Abnormal high ( > 3 mmol/L )	61 (35.1%)	44 (30.6%)	17 (56.7%)	
<b>Iron</b>				
• Normal ( 10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women )	164 (92.1%)	135 (92.5%)	29 (90.6%)	0.22
• Abnormal low ( < 10.6 umol/L in men; < 6.6 umol/L in women )	4 (2.2%)	2 (1.4%)	2 (6.2%)	

• Abnormal high ( > 28.3 umol/L in men; > 26 umol/L in women )	10 (5.6%)	9 (6.2%)	1 (3.1%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal ( 41 - 77 umol/L )	172 (97.2%)	141 (97.2%)	31 (96.9%)	1
• Abnormal low ( < 41 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 77 umol/L )	5 (2.8%)	4 (2.8%)	1 (3.1%)	
<b>Transferrin saturation</b>				
• Normal ( 20 - 55 % )	139 (78.5%)	120 (82.8%)	19 (59.4%)	0.011
• Abnormal low ( < 20 % )	34 (19.2%)	22 (15.2%)	12 (37.5%)	
• Abnormal high ( > 55 % )	4 (2.3%)	3 (2.1%)	1 (3.1%)	
<b>High sensitivity CRP</b>				
• Normal ( 0 - 5 mg/L )	146 (92.4%)	124 (93.9%)	22 (84.6%)	0.112
• Abnormal low ( < 0 mg/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 5 mg/L )	12 (7.6%)	8 (6.1%)	4 (15.4%)	

**Table S3: Blood investigations in 201 low-risk individuals sub-divided by those with severe or moderate post-COVID syndrome (PCS)**

Measurement	All	Moderate PCS	Severe PCS	p-value
<b>Haemoglobin</b>				
• Normal ( 130 - 170 g/L in men; 115 - 155 g/L in women )	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low ( < 130 g/L in men; < 115 g/L in women )	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high ( > 170 g/L in men; > 155 g/L in women )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Haematocrit (HCT)</b>				
• Normal ( 0.37 - 0.5 in men; 0.33 - 0.45 in women )	168 (97.1%)	64 (100%)	104 (95.4%)	0.274
• Abnormal low ( < 0.37 in men; < 0.33 in women )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 0.5 in men; > 0.45 in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
<b>Red cell count</b>				
• Normal ( 4.4 - 5.8 x10 <sup>12</sup> /L in men; 3.95 - 5.15 x10 <sup>12</sup> /L in women )	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.825
• Abnormal low ( < 4.4 x10 <sup>12</sup> /L in men; < 3.95 x10 <sup>12</sup> /L in women )	4 (2.3%)	2 (3.1%)	2 (1.8%)	
• Abnormal high ( > 5.8 x10 <sup>12</sup> /L in men; > 5.15 x10 <sup>12</sup> /L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Mean cell volume (MCV)</b>				
• Normal ( 80 - 99 fL )	170 (98.3%)	62 (96.9%)	108 (99.1%)	0.556
• Abnormal low ( < 80 fL )	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high ( > 99 fL )	0 (0%)	0 (0%)	0 (0%)	
<b>Mean corpuscular haemoglobin (MCH)</b>				
• Normal ( 26 - 33.5 pg )	170 (98.3%)	61 (95.3%)	109 (100%)	0.049
• Abnormal low ( < 26 pg )	2 (1.2%)	2 (3.1%)	0 (0%)	
• Abnormal high ( > 33.5 pg )	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Mean corpuscular haemoglobin concentration (MCHC)</b>				
• Normal ( 300 - 350 g/L )	131 (75.7%)	53 (82.8%)	78 (71.6%)	0.103
• Abnormal low ( < 300 g/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 350 g/L )	42 (24.3%)	11 (17.2%)	31 (28.4%)	
<b>Red cell distribution width (RDW)</b>				
• Normal ( 11.5 - 15 )	157 (91.3%)	59 (92.2%)	98 (90.7%)	0.339
• Abnormal low ( < 11.5 )	10 (5.8%)	2 (3.1%)	8 (7.4%)	
• Abnormal high ( > 15 )	5 (2.9%)	3 (4.7%)	2 (1.9%)	
<b>Platelet count</b>				
• Normal ( 150 - 400 x10 <sup>9</sup> /L )	161 (93.1%)	59 (92.2%)	102 (93.6%)	0.417
• Abnormal low ( < 150 x10 <sup>9</sup> /L )	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high ( > 400 x10 <sup>9</sup> /L )	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Mean platelet volume (MPV)</b>				
• Normal ( 7 - 13 fL )	172 (99.4%)	64 (100%)	108 (99.1%)	1
• Abnormal low ( < 7 fL )	0 (0%)	0 (0%)	0 (0%)	

• Abnormal high (> 13 fl)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>White cell count</b>				
• Normal (3 - 10 x10 <sup>9</sup> /L)	167 (96.5%)	61 (95.3%)	106 (97.2%)	0.671
• Abnormal low (< 3 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 10 x10 <sup>9</sup> /L)	6 (3.5%)	3 (4.7%)	3 (2.8%)	
<b>Neutrophils</b>				
• Normal (2 - 7.5 x10 <sup>9</sup> /L)	159 (91.9%)	57 (89.1%)	102 (93.6%)	0.468
• Abnormal low (< 2 x10 <sup>9</sup> /L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
• Abnormal high (> 7.5 x10 <sup>9</sup> /L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
<b>Lymphocytes</b>				
• Normal (1.2 - 3.65 x10 <sup>9</sup> /L)	156 (90.2%)	56 (87.5%)	100 (91.7%)	0.43
• Abnormal low (< 1.2 x10 <sup>9</sup> /L)	17 (9.8%)	8 (12.5%)	9 (8.3%)	
• Abnormal high (> 3.65 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Monocytes</b>				
• Normal (0.2 - 1 x10 <sup>9</sup> /L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.2 x10 <sup>9</sup> /L)	1 (0.6%)	0 (0%)	1 (0.9%)	
• Abnormal high (> 1 x10 <sup>9</sup> /L)	1 (0.6%)	1 (1.6%)	0 (0%)	
<b>Eosinophils</b>				
• Normal (0 - 0.4 x10 <sup>9</sup> /L)	167 (96.5%)	63 (98.4%)	104 (95.4%)	0.415
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.4 x10 <sup>9</sup> /L)	6 (3.5%)	1 (1.6%)	5 (4.6%)	
<b>Basophils</b>				
• Normal (0 - 0.1 x10 <sup>9</sup> /L)	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low (< 0 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 0.1 x10 <sup>9</sup> /L)	0 (0%)	0 (0%)	0 (0%)	
<b>Erythrocyte sedimentation rate (ESR)</b>				
• Normal (1 - 20 mm/hr)	160 (91.4%)	62 (93.9%)	98 (89.9%)	0.416
• Abnormal low (< 1 mm/hr)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 20 mm/hr)	15 (8.6%)	4 (6.1%)	11 (10.1%)	
<b>Sodium</b>				
• Normal (135 - 145 mmol/L)	168 (97.1%)	63 (98.4%)	105 (96.3%)	1
• Abnormal low (< 135 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 145 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Potassium</b>				
• Normal (3.5 - 5.1 mmol/L)	105 (62.1%)	35 (56.5%)	70 (65.4%)	0.255
• Abnormal low (< 3.5 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5.1 mmol/L)	64 (37.9%)	27 (43.5%)	37 (34.6%)	
<b>Chloride</b>				
• Normal (98 - 107 mmol/L)	166 (96%)	62 (96.9%)	104 (95.4%)	1
• Abnormal low (< 98 mmol/L)	4 (2.3%)	1 (1.6%)	3 (2.8%)	
• Abnormal high (> 107 mmol/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	

<b>Bicarbonate</b>				
• Normal ( 22 - 29 mmol/L )	147 (85%)	55 (85.9%)	92 (84.4%)	0.946
• Abnormal low ( < 22 mmol/L )	16 (9.2%)	6 (9.4%)	10 (9.2%)	
• Abnormal high ( > 29 mmol/L )	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Urea</b>				
• Normal ( 1.7 - 8.3 mmol/L )	173 (100%)	64 (100%)	109 (100%)	N/A
• Abnormal low ( < 1.7 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 8.3 mmol/L )	0 (0%)	0 (0%)	0 (0%)	
<b>Creatinine</b>				
• Normal ( 66 - 112 umol/L in men; 49 - 92 umol/L in women )	156 (90.2%)	59 (92.2%)	97 (89%)	0.705
• Abnormal low ( < 66 umol/L in men; < 49 umol/L in women )	12 (6.9%)	3 (4.7%)	9 (8.3%)	
• Abnormal high ( > 112 umol/L in men; > 92 umol/L in women )	5 (2.9%)	2 (3.1%)	3 (2.8%)	
<b>Bilirubin</b>				
• Normal ( 0 - 20 umol/L )	170 (98.3%)	63 (98.4%)	107 (98.2%)	1
• Abnormal low ( < 0 umol/L )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 20 umol/L )	3 (1.7%)	1 (1.6%)	2 (1.8%)	
<b>Alkaline phosphatase</b>				
• Normal ( 40 - 129 IU/L in men; 35 - 104 IU/L in women )	164 (94.8%)	59 (92.2%)	105 (96.3%)	0.185
• Abnormal low ( < 40 IU/L in men; < 35 IU/L in women )	7 (4%)	3 (4.7%)	4 (3.7%)	
• Abnormal high ( > 129 IU/L in men; > 104 IU/L in women )	2 (1.2%)	2 (3.1%)	0 (0%)	
<b>Aspartate transferase</b>				
• Normal ( 0 - 37 IU/L in men; 0 - 31 IU/L in women )	157 (92.9%)	59 (93.7%)	98 (92.5%)	1
• Abnormal low ( < 0 IU/L in men; < 0 IU/L in women )	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high ( > 37 IU/L in men; > 31 IU/L in women )	12 (7.1%)	4 (6.3%)	8 (7.5%)	
<b>Alanine transferase</b>				
• Normal ( 10 - 50 IU/L in men; 10 - 35 IU/L in women )	146 (84.4%)	56 (87.5%)	90 (82.6%)	0.512
• Abnormal low ( < 10 IU/L in men; < 10 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 50 IU/L in men; > 35 IU/L in women )	25 (14.5%)	7 (10.9%)	18 (16.5%)	
<b>Lactate dehydrogenase (LDH)</b>				
• Normal ( 135 - 225 IU/L in men; 135 - 214 IU/L in women )	137 (80.1%)	51 (81%)	86 (79.6%)	0.24
• Abnormal low ( < 135 IU/L in men; < 135 IU/L in women )	5 (2.9%)	0 (0%)	5 (4.6%)	
• Abnormal high ( > 225 IU/L in men; > 214 IU/L in women )	29 (17%)	12 (19%)	17 (15.7%)	
<b>Creatinine kinase (CK)</b>				
• Normal ( 38 - 204 IU/L in men; 26 - 140 IU/L in women )	159 (91.9%)	56 (87.5%)	103 (94.5%)	0.28
• Abnormal low ( < 38 IU/L in men; < 26 IU/L in women )	2 (1.2%)	1 (1.6%)	1 (0.9%)	
• Abnormal high ( > 204 IU/L in men; > 140 IU/L in women )	12 (6.9%)	7 (10.9%)	5 (4.6%)	
<b>Gamma glutamyl transferase</b>				
• Normal ( 10 - 71 IU/L in men; 6 - 42 IU/L in women )	161 (93.1%)	60 (93.8%)	101 (92.7%)	0.426
• Abnormal low ( < 10 IU/L in men; < 6 IU/L in women )	3 (1.7%)	0 (0%)	3 (2.8%)	
• Abnormal high ( > 71 IU/L in men; > 42 IU/L in women )	9 (5.2%)	4 (6.2%)	5 (4.6%)	
<b>Total protein</b>				
• Normal ( 63 - 83 g/L )	168 (97.1%)	63 (98.4%)	105 (96.3%)	0.792

• Abnormal low (< 63 g/L)	3 (1.7%)	1 (1.6%)	2 (1.8%)	
• Abnormal high (> 83 g/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
<b>Albumin</b>				
• Normal (34 - 50 g/L)	162 (93.6%)	59 (92.2%)	103 (94.5%)	0.538
• Abnormal low (< 34 g/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 50 g/L)	11 (6.4%)	5 (7.8%)	6 (5.5%)	
<b>Globulin</b>				
• Normal (19 - 35 g/L)	168 (97.1%)	61 (95.3%)	107 (98.2%)	0.616
• Abnormal low (< 19 g/L)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 35 g/L)	2 (1.2%)	1 (1.6%)	1 (0.9%)	
<b>Calcium</b>				
• Normal (2.2 - 2.6 mmol/L)	167 (96.5%)	62 (96.9%)	105 (96.3%)	0.525
• Abnormal low (< 2.2 mmol/L)	2 (1.2%)	0 (0%)	2 (1.8%)	
• Abnormal high (> 2.6 mmol/L)	4 (2.3%)	2 (3.1%)	2 (1.8%)	
<b>Magnesium</b>				
• Normal (0.6 - 1 mmol/L)	171 (98.8%)	63 (98.4%)	108 (99.1%)	0.604
• Abnormal low (< 0.6 mmol/L)	1 (0.6%)	1 (1.6%)	0 (0%)	
• Abnormal high (> 1 mmol/L)	1 (0.6%)	0 (0%)	1 (0.9%)	
<b>Phosphate</b>				
• Normal (0.87 - 1.45 mmol/L)	145 (83.8%)	55 (85.9%)	90 (82.6%)	0.824
• Abnormal low (< 0.87 mmol/L)	23 (13.3%)	8 (12.5%)	15 (13.8%)	
• Abnormal high (> 1.45 mmol/L)	5 (2.9%)	1 (1.6%)	4 (3.7%)	
<b>Uric acid</b>				
• Normal (266 - 474 umol/L in men; 175 - 363 umol/L in women)	145 (83.8%)	53 (82.8%)	92 (84.4%)	0.804
• Abnormal low (< 266 umol/L in men; < 175 umol/L in women)	18 (10.4%)	8 (12.5%)	10 (9.2%)	
• Abnormal high (> 474 umol/L in men; > 363 umol/L in women)	10 (5.8%)	3 (4.7%)	7 (6.4%)	
<b>Triglycerides</b>				
• Normal (< 2.3 mmol/L)	10 (100%)	6 (100%)	4 (100%)	N/A
• Abnormal high (> 2.3 mmol/L)	0 (0%)	0 (0%)	0 (0%)	
<b>Fasting triglycerides</b>				
• Normal (< 2.3 mmol/L)	144 (88.3%)	52 (89.7%)	92 (87.6%)	0.802
• Abnormal high (> 2.3 mmol/L)	19 (11.7%)	6 (10.3%)	13 (12.4%)	
<b>Cholesterol</b>				
• Normal (< 5 mmol/L)	4 (40%)	3 (50%)	1 (25%)	0.571
• Abnormal high (> 5 mmol/L)	6 (60%)	3 (50%)	3 (75%)	
<b>Fasting cholesterol</b>				
• Normal (< 5 mmol/L)	96 (58.9%)	39 (67.2%)	57 (54.3%)	0.135
• Abnormal high (> 5 mmol/L)	67 (41.1%)	19 (32.8%)	48 (45.7%)	
<b>HDL cholesterol</b>				
• Normal (0.9 - 1.5 mmol/L in men; 1.2 - 1.7 mmol/L in women)	103 (59.5%)	38 (59.4%)	65 (59.6%)	0.539
• Abnormal low (< 0.9 mmol/L in men; < 1.2 mmol/L in women)	16 (9.2%)	4 (6.2%)	12 (11%)	



• Abnormal high (> 1.5 mmol/L in men; > 1.7 mmol/L in women)	54 (31.2%)	22 (34.4%)	32 (29.4%)	
<b>LDL cholesterol</b>				
• Normal (< 3 mmol/L)	111 (65.7%)	45 (72.6%)	66 (61.7%)	0.18
• Abnormal high (> 3 mmol/L)	58 (34.3%)	17 (27.4%)	41 (38.3%)	
<b>Iron</b>				
• Normal (10.6 - 28.3 umol/L in men; 6.6 - 26 umol/L in women)	160 (92.5%)	57 (89.1%)	103 (94.5%)	0.337
• Abnormal low (< 10.6 umol/L in men; < 6.6 umol/L in women)	3 (1.7%)	2 (3.1%)	1 (0.9%)	
• Abnormal high (> 28.3 umol/L in men; > 26 umol/L in women)	10 (5.8%)	5 (7.8%)	5 (4.6%)	
<b>Total iron binding capacity (TIBC)</b>				
• Normal (41 - 77 umol/L)	167 (97.1%)	60 (93.8%)	107 (99.1%)	0.064
• Abnormal low (< 41 umol/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 77 umol/L)	5 (2.9%)	4 (6.2%)	1 (0.9%)	
<b>Transferrin saturation</b>				
• Normal (20 - 55 %)	135 (78.5%)	50 (78.1%)	85 (78.7%)	0.283
• Abnormal low (< 20 %)	33 (19.2%)	11 (17.2%)	22 (20.4%)	
• Abnormal high (> 55 %)	4 (2.3%)	3 (4.7%)	1 (0.9%)	
<b>High sensitivity CRP</b>				
• Normal (0 - 5 mg/L)	141 (92.2%)	50 (96.2%)	91 (90.1%)	0.223
• Abnormal low (< 0 mg/L)	0 (0%)	0 (0%)	0 (0%)	
• Abnormal high (> 5 mg/L)	12 (7.8%)	2 (3.8%)	10 (9.9%)	