ARTICLE



Neurobiological roots of psychopathy

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Abstract

Psychopathy is an extreme form of antisocial behavior, with about 1% prevalence in the general population, and 10–30% among incarcerated criminal offenders. Although the heritability of severe antisocial behavior is up to 50%, the genetic background is unclear. The underlying molecular mechanisms have remained unknown but several previous studies suggest that abnormal glucose metabolism and opioidergic neurotransmission contribute to violent offending and psychopathy. Here we show using iPSC-derived cortical neurons and astrocytes from six incarcerated extremely antisocial and violent offenders, three nonpsychopathic individuals with substance abuse, and six healthy controls that there are robust alterations in the expression of several genes and immune response-related molecular pathways which were specific for psychopathy. In neurons, psychopathy was associated with marked upregulation of *RPL10P9* and *ZNF132*, and downregulation of *CDH5* and *OPRD1*. In astrocytes, *RPL10P9* and *MT-RNR2* were upregulated. Expression of aforementioned genes explained 30–92% of the variance of psychopathic symptoms. The gene expression findings were confirmed with qPCR. These genes may be relevant to the lack of empathy and emotional callousness seen in psychopathy, since several studies have linked these genes to autism and social interaction.

Introduction

In developed countries, a relatively small group of antisocial recidivistic offenders commits the majority of all

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violent crimes. The prevalence of antisocial personality disorder (ASPD) is 1–3% in the general population and 40–70% in prison populations, and the corresponding figures for its most severe manifestation, psychopathy, are about 1% in the general population and 10–30% among incarcerated offenders [1–5]. ASPD is characterized by aggression, hostility, callousness, manipulativeness, deceitfulness, and impulsivity, and psychopathy is an

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extreme manifestation of ASPD. Severe antisocial and criminal behavior has a substantial genetic component [6]. This far only one study has reported contributing genes reaching genome-wide significance for ASPD [7], although two studies have found association between singlenucleotide polymorphisms and broad spectrum of antisocial behavior [8, 9]. LINC00951, the gene associated with imprisoned offenders with ASPD, codes for long intergenic noncoding RNA, which is expressed especially in the frontal cortex and cerebellum. Its function is not known [7], but it has been linked to autoimmune disease [10]. The gene linked to adult antisocial behavior, ABCB1, is also highly expressed in the brain, and implicated in substance abuse [9]. No underlying molecular pathways of severe antisocial and criminal behavior are known, but there is preliminary evidence on dysregulation of the endogenous opioid system and brain opioid receptors [11–13] in antisocial individuals. Also abnormal glucose metabolism leading to hypoglycemia has been observed as the strongest predictor for violent crimes [14]. A recent study has also found association between immune-related gene sets and antisocial behavior [9]. We aimed to study the neurobiological background of psychopathy by using induced pluripotent stem cell (iPSC)-derived cortical neurons and astrocytes, and included also nonpsychopathic substance abusers in addition to healthy individuals as control groups in order to distinguish the putative role of the coexisting substance dependence.

Material and methods

We generated and fully characterized iPSC lines from six antisocial violent offenders, three nonviolent substance abusers and six control subjects without antisocial traits or substance abuse disorders. Due to the explanatory nature of the study, no power analysis based on predefined effect size was done. The clinical and sociodemographic characteristics of the study subjects are shown in Table 1. We chose to differentiate the cells into cortical neurons expressing markers of glutamatergic and GABAergic neurons and to astrocytes. Methods for iPSC production and their characterization, derivation of neurons and astrocytes and their analyses are reported in detail in Supplementary Material.

Table 1 Clinical and sociodemographic characteristics of study subjects

	Age	Diagnosis	Number of committed homicides	Number of violent crimes	PCL-R score
Subject 1	30	Antisocial personality disorder, ADHD, alcohol dependence, benzodiazepine abuse, multiple sclerosis, asthma	2	19	37.0
Subject 2	42	Antisocial personality disorder, alcohol dependence	3	4	Not available
Subject 3	49	Antisocial personality disorder, alcohol dependence	2	11	30.0
Subject 4	43	Antisocial personality disorder, alcohol dependence, polysubstance dependence	2	7	33.7
Subject 5	30	Antisocial personality disorder, alcohol dependence, opioid dependence, cannabis dependence, benzodiazepine dependence, amphetamine dependence	3	8	36.0
Subject 6	47	Antisocial personality disorder, borderline personality disorder, paranoid personality disorder, alcohol dependence, polysubstance dependence, amphetamine dependence, hepatitis C	2	9	37.0
Subject 7	38	Alcohol dependence	0	0	2
Subject 8	25	Alcohol dependence	0	0	3
Subject 9	31	Alcohol dependence, cannabis dependence, bulimia	0	0	11
Subject 10	44	None	0	0	3
Subject 11	28	None	0	0	2
Subject 12	28	None	0	0	1
Subject 13	47	None	0	0	3
Subject 14	26	None	0	0	2
Subject 15	51	None	0	0	1

All individuals were males. Subjects 1–6 are violent offenders, 7–9 are individuals with substance abuse but without criminal behavior, and 10–15 are healthy controls. The biological fathers of Subject 1, Subject 3, and Subject 5 had prison convictions due to violent and nonviolent crimes. None of the biological mothers had been convicted into prison

PCL-R psychopathy checklist revised

Description of subjects

Six male offenders were identified by the history of their criminal convictions from the Finnish National Crime Register and recruited through the penal system and classified as extremely violent offenders as described in Tiihonen et al. [5]. Three individuals having substance dependence without violent behavior were recruited from the local substance abuse rehabilitation center, and six healthy controls were recruited from the staff of Niuvanniemi Hospital. The participants were interviewed with Structured Clinical Interview for DSM-IV-Disorders to exclude individuals with a psychosis diagnosis, and to assess whether or not the subject fulfilled criteria for ASPD. Also, any history of substance abuse (alcohol, heroin, buprenorphine, amphetamine, cannabis, other) was obtained through a questionnaire. The history of criminal convictions was obtained from the National Crime Register [5]. Psvchopathy ratings with the Hare Psychopathy Checklist revised (PCL-R) [1] were done by accredited rater OV using official crime register data and forensic mental examination reports (violent offenders), and clinical interview (individuals with substance dependence, healthy controls). Informed consent was obtained from all subjects. This study was approved by the Ethics Committee for Pediatrics, Adolescent Medicine and Psychiatry, Hospital District of Helsinki and Uusimaa, and the Criminal Sanctions Agency of Finland.

Generation of hiPSCs and their characterization

The hiPSC lines were derived from individuals' skin fibroblasts (Supplementary Table 1, Supplementary Figs. 1 and 2). The fibroblasts were isolated and expanded in fibroblast culture media containing Iscove's DMEM media (Thermo Fisher Scientific) with 20% fetal bovine serum, 1% Penicillin-Streptomycin and 1% nonessential amino acids. iPSC reprogramming was performed by the CytoTunE-iPS 2.0 Sendai Reprogramming Kit (Thermo Fisher Scientific) according to the manufacturer's instructions. The iPSCs were grown on Matrigelcoated dishes (BD Biosciences) in E8 medium (Gibco). Medium was changed every other day and hiPSC colonies were enzymatically passaged using 0.5 mM EDTA (Gibco). The pluripotency of hiPSCs was confirmed by expression of pluripotent markers using immunocytochemistry (Oct-4, Sox2, TRA-1-81, and SSEA4) and qPCR (OCT-4, SOX-2, NANOG, and LIN-28). The embryoid body formation assay showed hiPSCs properties to differentiate into all three germ layers. In this assay, hiPSCs were proliferated in low-adherent plates for 2 weeks after which the EBs were plated down on Matrigel-coated plates for an additional two weeks. The expression of smooth muscle actin positive cells (mesoderm), BIIITubulin-positive cells (ectoderm), and alpha-fetoprotein positive cells (endoderm) was confirmed by immunocytochemistry. The clearance of Sendai virus was measured by qPCR, and United Medix Laboratories Ltd in Helsinki (Finland) confirmed normal karyotype of each cell line.

hiPSC differentiation to neural precursor cells (NPCs) and cortical neurons

Neural differentiation was performed according to Hicks et al. [15] with minor modifications. hiPSC colonies growing on Matrigel-coated plates are exposed to dual SMAD inhibitors (10 µM SB431542 and 200 nM LDN-193189) for 10 days in neural differentiation medium containing a 1:1 mix of DMEM/F12 and Neurobasal medium supplemented with 1% B27 supplement, 0.5% N2 supplement, 2 mM Glutamax, 50 IU/ml penicillin, and 50 μg/ml streptomycin (all from Gibco). After the induction, 25 ng/ml bFGF (R&D Systems) was added for additional 2 days to expand the differentiated neuroepithelial cells in rosettes. Rosettes were detached and plated into ultralow attachment dishes (Corning) in neural sphere medium (NSM), consisting of a 1:1 mix of DMEM/F12 and Neurobasal medium supplemented with 1% N₂ supplement, 2 mM Glutamax, 50 IU/ml penicillin, and 50 µg/ml streptomycin (all from Gibco) supplemented with 25 ng/ml bFGF. During the differentiation, half of the medium was renewed every other day and the spheres were manually cut once a week to maintain NPC population. For experimental purposes, NPCs were dissociated with Accutase and plated in NSM media onto PORN/Matrigel-coated plates (with density $2-3 \times 10^6$ cells/6 cm dish; 1×10^6 cells/6-well plate or 100,000 cell/24-well plate). The neurons were matured for 1 week before experiments. Immunocytochemistry results showing the fractions of glutamatergic and GABAergic cells are presented in Supplementary Fig. 3.

hiPSC differentiation to astrocytes

We have adapted a previously published protocol for the differentiation of hiPSC-derived astrocytes [16]. Briefly, due to the same origin of neurons and astrocytes, hiPSCs were differentiated into neuroepithelial cells by using the same procedure as for neuron differentiation for the first 10 days. The neural progenitors were detached to ultralow attachment dishes and expanded in astrocyte sphere medium, i.e., DMEM/F12 medium supplemented with 1% N₂ supplement, 2 mM Glutamax, 50 IU/ml penicillin, and 50 µg/ml streptomycin (all from Gibco), 5000 KY/ml Heparin (LEO), 10 ng/ml bFGF and 10 ng/ml EGF (both from R&D Systems). Half of the medium was renewed every other day

and the spheres were manually cut once a week. According to our experience, this method generates a homogenous population of astrocyte progenitor cells within 4 months of differentiation. The astrocytes were further maturated on Matrigel-coated plates by treatment with 10 ng/ml CNTF and 10 ng/ml BMP4 (both from PeproTech) for 1 week in a density of 600,000 cells/6-well plate or 80,000 cell/24-well plate.

Methods concerning immunocytochemistry, RNA isolation, gene expression profile, qRT-PCR, and quantitative proteomic analysis are described in detail in Supplementary Material.

Results

Supplementary Tables 2 and 3 show all differentially expressed genes in neurons and astrocytes, respectively, up to nominal significance (p < 0.05) between violent offenders and control subjects. Since cultivation of neurons failed from cells of one healthy control, there were 14 individuals in the analyses concerning neurons, and 15 individuals in the analyses concerning astrocytes. Differentially expressed genes surviving correction for multiple comparisons in cortical neurons are shown in Table 2. Of these genes, in neurons, ribosomal RPL10P9 pseudogene showed over tenfold upregulation in violent offenders as compared with healthy controls and nonviolent individuals with substance abuse. Also zinc finger protein 132 (ZNF132) gene was markedly upregulated, and cadherin 5 (CDH5) gene markedly downregulated among the neurons derived from cells of violent offenders. Figure 1 displays the qPCR replications of these results (except for RPL10P9 due to no suitable primers being available), and correlations between gene expression levels and psychopathy score (PCL-R). Pearson's correlations between gene expression and PCL-R score were 0.67 (p = 0.013, N = 13; see Fig. 2) for RPL10P9, 0.96 (p = 0.000, N = 13) for ZNF132, -0.65(p = 0.015, N = 13) for CDH5, and -0.55 (p = 0.05, N =13) for opioid receptor delta 1(OPRD1). CDH13 gene encoding a cadherin that regulates axon growth during neural differentiation, has been previously linked to extremely violent behavior [5], but it did not achieve statistically significance (p = 0.24) in this study, possibly because it is most prominently expressed by oligodendrocytes in the brain. Although the result for RPL10P9 could not be verified with qPCR due to missing suitable primers, the same result for significant correlation with PCL-R score and upregulation of the gene in the gene array was also discovered in the astrocytes differentiated from the hiPSCs lines [Pearson's correlation 0.66 (p = 0.007, N = 14)] (Fig. 2). Altogether, the data indicate the robustness of this finding and underline the importance of RPL10P9 in the pathophysiology of psychopathy. In the astrocytes, also mitochondria encoded 16S RNA (*MT-RNR*) 2 showed a four- to sixfold upregulation of RNA expression in the violent offenders (Fig. 2).

Interestingly, opioid receptor delta 1 (OPRD1) gene was upregulated in nonviolent offenders as compared with normal controls, but the expression for *OPRD1* was even lower in the violent offenders as compared with the two other groups. although these results reached only borderline statistical significance (Fig. 1c, Suppl. Table 2). As the OPRD1 gene codes for an opioid receptor delta protein, a protein involved in mediating the effects of opioids often used for substance abuse purposes, and as many violent antisocial offenders often suffer from substance abuse disorders, it was striking that the expression level of this protein was low in the violent offenders. Thus, we did further qPCR analysis for OPRD1 in order to see whether differences between the groups arise in this more accurate analysis. The qPCR analysis pointed toward the same trend of lower expression of *OPRD1* in the violent offender group, although this result again reached only borderline statistical significance. Taking all together, ZNF132, RPL10P9, CDH5, and OPRD1 genes explained 30–92% of variance of the psychopathy symptomatology, as measured by the PCL-score. Moreover, in fibroblasts, no differences between the studied groups were detected (Suppl. Fig. 4). These findings point out that ZNF132 is mainly overexpressed in neurons and most likely affect the transcriptional regulation of other genes. Results from pathway analyses from neurons are displayed in Supplementary Table 4, and show enrichment in several immune responserelated pathways. No statistically significant enrichments were observed in astrocytes.

Data from proteomic analysis are shown in Fig. 3. Here the largest effect sizes were observed for opioid-binding protein/cell-adhesion molecule (OPCML) in the proteomic analysis and for PSMD3, PEG10, and PCDH19 in phosphoproteome analysis. Of the proteins with significantly higher levels in the proteome analysis, OPCML was the most elevated protein in the violent offenders as compared with controls (6.6-fold change, $p = 9.5 \times 10^{-3}$). However, also nonviolent substance abusers showed higher OPCML values than controls, indicating that this finding may not be necessarily specific for psychopathy but could be associated with substance dependence.

In the phosphoproteome analysis, in the violent criminals, paternally expressed 10 (PEG10) levels were 51-fold, protocadherin 19 (PCDH19) 37-fold, spectrin beta, nonerythrocytic 5 (SPTBN5) 17-fold, and acyl-CoA synthetase long chain family member 4 (ACSL4) 7-fold higher than controls. Concerning phosphoproteins with lower levels in the violent offenders, levels of a proteasome 26S subunit non-ATPase 3 (PSMD3) were 203-fold, and Myosin 1E (MYO1e) 19-fold lower than in controls.

Table 2 Transcriptome analysis of differentially expressed genes in hiPSCs-derived cortical neurons

ENSGO0000233913					
RPL10P9 ZNF132 RPL10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 TTGAX MT-TS1 TTGAX MT-TS1 TTGAX MT-TS1 TGAX MT					
ZNF132 ZNF132 RPL10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 ITGAX MT-TS1 ITGAX MT-TP6 LILRB2 OR2T11 ASIP	Some protein III neelidogene y	606 86	3 433	8 26E-10	4 29F-05
ZNF132 RPL10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 ITGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM21B SNORD115-5 MTND2P28 MTND2P2		79.387	2.447	3.48E-09	9.06E-05
RPL 10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 TTGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM21B SNORD115-5 MTND2P28 MTND	nc finger protein 132	138.380	2.407	4.37E-08	7.57E-04
RPL 10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 TIGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND2P28 MTND2P28 MTND2P28 MTND2P28 MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP		21.891	1.526	3.39E-06	4.40E-02
RPL10P9 DNM1P46 MT-RNR1 MYPN ABCC2 APOL3 ITGAX MT-TS1 TWEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND					
DNMIP46 MT-RNR I MYPN ABCC2 APOL3 ITGAX MT-TSI TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MT	Ribosomal protein L10 pseudogene 9	606.86	3.686	6.13E - 10	1.84E - 05
MT-RNR I MYPN ABCC2 APOL3 ITGAX MT-TSI TWEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 M	namin 1 pseudogene 46	16.689	-1.595	1.70E - 07	2.13E-03
MYPN ABCC2 APOL3 ITGAX MT-TS1 TWEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 M	Mitochondrially encoded 12 S RNA	3236.254	-2.687	2.12E-07	2.13E-03
ABCC2 APOL3 ITGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND3P2 MTND3P2 MTND3P3 TREM1	vopalladin	7.066	-2.990	4.20E - 07	3.16E - 03
APOL3 ITGAX MT-TS1 TWEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MT	ATP binding cassette subfamily C member 2	19.849	-1.972	1.76E - 06	1.01E-02
ITGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28		13.244	-2.741	2.02E - 06	1.01E-02
ITGAX MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28		4.804	-2.446	5.86E - 06	1.82E-02
MT-TS1 TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND2P	egrin subunit alpha X	13.142	-2.747	6.24E - 06	1.82E-02
TMEM217 CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND	Mitochondrially encoded tRNA serine 1 (UCN)	17.046	-2.165	7.14E-06	1.82E-02
CDH5 NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND2P28 CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	ansmembrane protein 217	40.619	-1.633	7.58E-06	1.82E-02
NSFP1 CBWD1 FAM221B SNORD115-5 MTND2P28 MTND2P28 CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	dherin 5	7.101	-2.516	8.16E - 06	1.82E-02
CBWD1 FAM221B SNORD115-5 MTND2P28 KIAA1324L CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP		7.076	2.313	8.31E - 06	1.82E-02
FAM221B SNORD115-5 MTND2P28 KIAA1324L CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	OBW domain containing 1	320.593	1.208	8.47E - 06	1.82E-02
SNORD115-5 MTND2P28 KIAA1324L CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	Family with sequence similarity 221 member B	7.214	-2.535	1.06E - 05	2.09E-02
MTND2P28 KIAA1324L CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	Small nucleolar RNA, C/D box 115-5	28.237	2.062	1.11E-05	2.09E-02
KIAA1324L CLEC20A GPRCSA MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	Mitochondrially encoded NADH:ubiq.	86.559	-2.017	1.36E - 05	2.24E-02
KIAA1324L CLEC20A GPRCSA MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP		9.175	-2.653	1.46E - 05	2.24E-02
KIAA1324L CLEC20A GPRC5A MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP		5.636	-2.501	1.49E - 05	2.24E-02
CLEC20A GPRCSA MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	AA1324 like	1178.161	1.248	1.56E - 05	2.24E-02
CLEC20A GPRCSA MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP		6.711	2.063	1.56E - 05	2.24E-02
GPRCSA MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	C-type lectin domain containing 20A	3.726	-2.730	1.70E - 05	2.33E-02
MGAM MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	G protein-coupled receptor class C group 5 member A	19.264	-2.356	2.02E - 05	2.56E-02
MT-ATP6 LILRB2 OR2T11 SCARNA3 TREM1 ASIP	ıltasE–glucoamylase	42.608	-1.951	2.05E - 05	2.56E-02
LILRB2 OR2T11 SCARNA3 TREM1 ASIP	Mitochondrially encoded ATP synthase 6	629.862	-1.705	2.53E - 05	3.04E-02
OR2T11 SCARNA3 TREM1 ASIP	Leukocyte immunoglobulin like receptor B2	2.933	-2.720	2.95E - 05	3.41E - 02
SCARNA3 TREMI ASIP	Olfactory receptor family 2 subfamily T member 11	4.182	-2.634	3.13E-05	3.46E - 02
SCARNA3 TREM1 ASIP		5.112	-2.644	3.22E - 05	3.46E - 02
SCARNA3 TREM1 ASIP		79.387	1.852	3.47E - 05	3.59E-02
TREMI ASIP	nall Cajal body-specific RNA 3	35.106	1.208	3.58E - 05	3.59E-02
ASIP	Triggering receptor expressed on myeloid cells 1	5.602	-2.372	4.35E-05	4.10E-02
ENSG00000187812	outi signaling protein	3.816	-2.555	4.36E - 05	4.10E-02
		7.086	-2.614	4.61E - 05	4.20E-02
	nc finger protein 132	138.380	1.920	5.01E - 05	4.43E-02
ENSG0000060709 RIMBP2 RIMS binding protein 2	MS binding protein 2	152.792	1.619	5.68E - 05	4.88E-02

"Violent" indicates violent offenders, and "nonviolent" indicates individuals with substance abuse but without criminal behavior Only genes with adjusted p-value < 0.05 and at least twofold up- or downregulation are presented

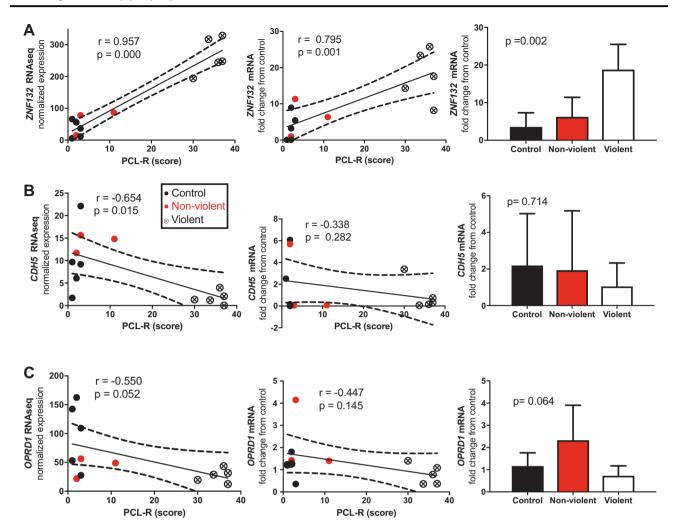


Fig. 1 RNA expression analyses of hiPSCs-derived cortical neurons for a *ZNF132*, **b** *CDH5*, and **c** *OPRD1* genes. The first graph represents correlation with normalized expression levels and the second with gene expression levels validated by quantitative RT-PCR (qRT-

PCR). The column graph presents mRNA expression levels of gene of interest measured by qRT-PCR. *r* indicates Pearson correlation coefficient. "Violent" indicates violent offenders, and "nonviolent" indicates individuals with substance abuse but without criminal behavior

Discussion

To our knowledge, this is the first study to reveal significant alterations in gene expression related to psychopathy. Our results showed that expression levels of *RPL109*, *ZNF132*, *CDH5*, and *OPRD1* genes in neurons explained 30–92% of the severity of psychopathy, and *RPL109* expression was significantly associated with degree of psychopathy also in astrocytes. It is remarkable that all the aforementioned genes except *OPRD1* have been previously linked to autism [17–22], and might thus contribute to the emotional callousness and lack of empathy observed in psychopathic violent offenders. The strongest association was observed for *ZNF132*, a member of zinc finger Kröppel family associated with several developmental and malignant disorders [23]. It has been also reported that autism gene *CHD8* modifies the expression of *ZNF132* [18]. The exact

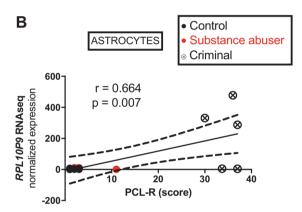
function of *ZNF132* is unknown but it may be involved in transcriptional regulation. Interestingly, the highest expression levels of *ZNF132* mRNA were seen in cortical neurons of violent subjects, while in hiPSCs, no difference between violent and nonviolent subjects was observed. *ZNF132* is expressed highly in the cerebellum [24], and a recent study has found that cerebellum can regulate social behavior by controlling dopamine release [25], suggesting that this may contribute to mental disorders, such as autism and schizophrenia. Our results imply that cerebellum may also have a role in severe antisocial behavior.

We observed enrichment in several immune responserelated pathways. This is an interesting finding since a recent study on adult antisocial behavior found enrichment in 7 gene sets, most of which being immune related [9]. This suggests that altered immune response contributes to the pathophysiology of antisocial behavior.

A ASTROCYTES violent vs. control

	HGNC		Average	Log2		Adjusted
Ensembl ID	symbol	Gene description	expression	foldchange	P-value	p-value
ENSG00000233913	RPL10P9	ribosomal protein L10 pseudogene 9	101,870	2,373	8,210E-09	0,000
ENSG00000210082	MT-RNR2	mitochondrially encoded 16S RNA	26374,371	2,039	4,859E-07	0,005

violent vs. control + non-violent							
	HGNC		Average	Log2		Adjusted	
Ensembl ID	symbol	Gene description	expression	foldchange	P-value	p-value	
ENSG00000123080	CDKN2C	cyclin dependent kinase inhibitor 2C	585,550	1,374	4,088E-10	0,000	
ENSG00000233913	RPL10P9	ribosomal protein L10 pseudogene 9	101,870	2,608	1,715E-09	0,000	
ENSG00000210082	MT-RNR2	mitochondrially encoded 16S RNA	26374,371	2,469	1,053E-08	0,000	
ENSG00000280660			20,006	-2,149	2,360E-07	0,001	
ENSG00000266066	POLRMTP1		9,748	1,744	1,916E-06	0,006	
ENSG00000069696	DRD4	dopamine receptor D4	97,963	-1,874	2,069E-06	0,006	
ENSG00000100033	PRODH	proline dehydrogenase 1	67,209	-1,835	2,439E-06	0,006	
ENSG00000108231	LGI1	leucine rich glioma inactivated 1	138,973	-1,864	4,405E-06	0,009	
ENSG00000023892	DEF6	DEF6, guanine nucleotide exchange factor	5,139	-1,841	9,024E-06	0,016	
ENSG00000241213	LINC02024	long intergenic non-protein coding RNA 2024	34,935	-1,221	1,160E-05	0,019	
ENSG00000171759	PAH	phenylalanine hydroxylase	12,072	-1,348	2,884E-05	0,039	
ENSG00000281406	BLACAT1	bladder cancer associated transcript 1 (non-protein coding)	101,401	-1,205	3,648E-05	0,046	
ENSG00000234323	LINC01505	long intergenic non-protein coding RNA 1505	26,206	1,769	4,320E-05	0,046	
ENSG00000172785	CBWD1	COBW domain containing 1	441,282	1,154	4,677E-05	0,046	
ENSG00000164241	C5orf63	chromosome 5 open reading frame 63	11,721	-1,768	4,990E-05	0,046	
ENSG00000246214			23,684	-1,390	5,040E-05	0,046	
ENSG00000156689	GLYATL2	glycine-N-acyltransferase like 2	36,035	1,229	5,717E-05	0,050	



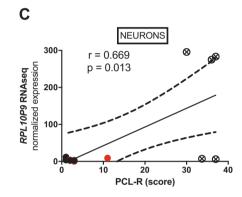


Fig. 2 Transcriptome analyses of differentially expressed genes in hiPSCs-derived astrocytes. **a** The genes with adjusted *p*-value < 0.05 and at least twofold up- or downregulation are presented in the table. The correlation of PCL-R score with normalized expression levels for

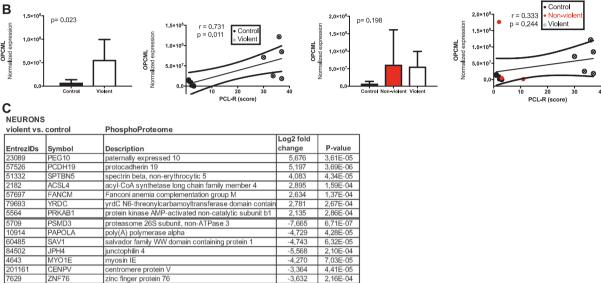
RPL10P9 in **b** astrocytes and **c** neurons. *r* indicates the Pearson correlation coefficient. "Violent" indicates violent offenders, and "nonviolent" indicates individuals with substance abuse but without criminal behavior

In proteomic analysis, the most robust finding was upregulation of OPCML. It has been shown to have an accessory role in opioid receptor function, and the gene encoding the protein is highly conserved in mammals. In rats, the accessory role to activate opioid receptors has been shown to be specific for the mu receptor ligands. Differences in *OPCML* gene expression have also been detected in patients with schizophrenia, although protein level measurements from post mortem brains have not differed between patients and healthy controls [26].

In phosphoprotein analysis, several proteins were upregulated. Of these, PEG10 is a paternally imprinted gene that uses a rare mechanism for encoding for two different protein products by using the -1 ribosomal frameshift translation, which is well known from retroviruses and retrotransposons, but is extremely rare in humans [27]. In adult mice, the protein is expressed only in the brain and

testes, and blocks TGF-B signaling. A paternally imprinted gene such as this one could explain why psychopathy is inherited from father to son. In this study, three of six offenders had a biological father convicted into prison, while none of the mothers had been imprisoned. PCDH19 is a protocadherin, which has been linked to epilepsy [28], autism [29] and behavioral problems, aggression, and photosensitivity. PCDH19 is thought to be a calciumdependent cell-adhesion protein that is primarily expressed in the brain, and has been shown to cause a decrease in the amount of neurosteroids, including adrenocorticotropic hormone, in females. ACSL4 has been associated with X-chromosome linked mental retardation [30] as well as insulin secretion [31]. On the other hand, PSMD3 and MYO1e were substantially downregulated compared with controls. Of these, PSMD3 is an enzyme, an aberration of which contributes to pathogenesis of neurodevelopmental

NEURONS violent vs. control Proteome Log2 fold P-value EntrezIDs Symbol Description change 4978 9,50E-03 opioid binding protein/cell adhesion molecule like XPNPEP2 1,960 3,91E-03 X-prolyl aminopeptidase 2 3034 histidine ammonia-lyase 10457 GPNMB -2,040 -1,770 3.60E-03 glycoprotein nmb DECR2 26063 2.4-dienovl-CoA reductase 2 4.95E-04 HIST2H3C//HIST histone cluster 2 H3 family member c//histone cluster 2H3A//HIST2H3 2 H3 family member a//histone cluster 2 H3 family 126961 member d -1,453 3,75E-03 minichromosome maintenance complex binding protein -1,249 6,44E-03 В 1.5×10 2.0×10 n= 0.023 r = 0.731• Contro p = 0.0111.5×10 1.0×10 OPCML alized expre OPCML 1 0×10 5.0×10



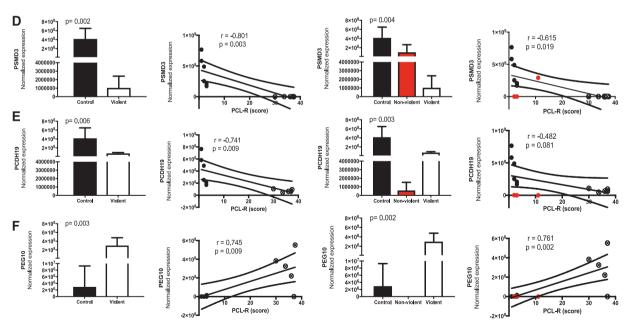


Fig. 3 Proteomic analyses of differentially expressed proteins and phosphoproteins in hiPSCs-derived cortical neurons. **a** Proteins with adjusted *p*-value < 0.05 and at least twofold up- or downregulation are presented in the table. **b** The normalized expression of opioid-binding protein/cell-adhesion molecule (OPCML) and its correlation with PCL-R score. **c** Top ten list of phosphoproteins and normalized

expression and its correlation with PCL-R score for **d** 26S proteasome non-ATPase regulatory subunit 3 (PSMD3), **e** Protocadherin 19 (PCDH19), **f** Retrotransposon-derived protein (PEG10). p-values shown in **a** and **c** are nominal values, and remained statistically significant (p < 0.05) after correction for multiple comparisons in **c**. r indicates the Pearson correlation coefficient

and neurodegenerative disorders [32, 33] and insulin resistance [34]. This finding suggests that downregulation of PSMD3 contributes to abnormal glucose metabolism which results into impulsive violent behavior among severely antisocial individuals as has been reported in several studies [14, 35]. MYO1e has been associated with autism in a single study [36].

In conclusion, expression of ZNF132 in neurons and RPL10P9 in both neurons and astrocytes is markedly abnormal among habitually violent offenders and these findings are strongly associated with the degree of psychopathic symptoms. The changes in protein levels observed here point to alteration in insulin sensitivity and glucose metabolism, and previous literature has shown that abnormal glucose metabolism is the only predictor for violent crimes which can surpass the accuracy of PCL-R [35]. We also observed changes in the opioid system, which has been shown to support prosocial functions, such as empathy, among humans and nonhuman primates [12, 13, 37, 38]. Our results showing a decrease in the expression of opioid delta receptor gene are in line with these previous findings. A recent theory suggests that a deficient endogenous opioid system contributes to antisocial personality, proposing that antisocial individuals attempt to stimulate their dysfunctional opioid system by the rewarding effect of substance abuse, and impulsive, sensationseeking, aggressive, and promiscuous behavior [11]. Our data suggest that dysfunction of the opioid system contributes to the phenotype of psychopathy, supporting the recently presented idea that partial opioid receptor agonists, such as (+)-naloxone might be the first effective treatment for psychopathy [11].

Data availability

All data needed to evaluate the conclusions in the manuscript are provided in the manuscript or the supplementary material.

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Author contributions JT and JK conceived the study. ŠL planned and supervised iPSC lines characterizations, differentiation the neurons and sample preparation for RNA and protein sequencing. IO and OV performed skin biopsies and rating of symptoms, KAP derived iPSC lines with YG and characterized them. IH grew and differentiated neurons and PLJV characterized them. MK differentiated the neurons, and prepared RNA and protein samples for sequencing, run qRT-PCR.

JT wrote the first draft of the manuscript with help of JK, ŠL prepared the figures and tables, and all authors contributed to the final version.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Hare RD. The Hare Psychopathy Checklist Revised (PCL-R).
 2nd ed. Toronto, ON, Canada: Multi-Health Systems; 2003.
- 2. Fazel S, Danesh J. Serious mental disorder in 23000 prisoners: a systematic review of 62 surveys. Lancet. 2002;359:545–50.
- Gibbon S, Duggan C, Stoffers J, Huband N, Völlm BA, Ferriter M, et al. Psychological interventions for antisocial personality disorder. Cochrane Database Syst Rev. 2010;16:CD007668.
- Vitale JE, Smith SS, Brinkley CA, Newman JP. The reliability and validity of the Psychopathy Checklist – Revised in a sample of female offenders. Crim Justice Behav. 2002;29:202–31.
- Tiihonen J, Rautiainen MR, Ollila HM, Repo-Tiihonen E, Virkkunen M, Palotie A, et al. Genetic background of extreme violent behavior. Mol Psychiatry. 2015;20:786–92.
- Ferguson CJ. Genetic contributions to antisocial personality and behavior: a meta-analytic review from an evolutionary perspective. J Soc Psychol. 2010;150:160–80.
- Rautiainen MR, Paunio T, Repo-Tiihonen E, Virkkunen M, Ollila HM, Sulkava S, et al. Genome-wide association study of antisocial personality disorder. Transl Psychiatry. 2016;6:e883.
- Tielbeek JJ, Johansson A, Polderman TJC, Rautiainen MR, Jansen P, Taylor M, et al. Genome-wide association studies of a broad spectrum of antisocial behavior. JAMA Psychiatry. 2017;74:1242–50.
- Salvatore JE, Edwards AC, McClintick JN, Bigdeli TB, Adkins A, Aliev F, et al. Genome-wide association data suggest ABCB1 and immune-related gene sets may be involved in adult antisocial behavior. Transl Psychiatry. 2015;5:e558.
- McIntosh LA, Marion MC, Sudman M, Comeau ME, Becker ML, Bohnsack JF, et al. Genome-wide association meta-analysis reveals novel juvenile idiopathic arthritis susceptibility loci. Arthritis Rheuma. 2017;69:2222–32.
- Bandelow B, Wedekind D. Possible role of a dysregulation of the endogenous opioid system in antisocial personality disorder. Hum Psychopharmacol. 2015;30:393–415.
- Nummenmaa L, Tuominen L. Opioid system and human emotions. Br J Pharm. 2018;175:2737–49.

- Nummenmaa L, Tuominen L, Dunbar R, Hirvonen J, Manninen S, Arponen E, et al. Social touch modulates endogenous μ-opioid system activity in humans. Neuroimage. 2016;138:242–7.
- Virkkunen M, Rissanen A, Franssila-Kallunki A, Tiihonen J. Low non-oxidative glucose metabolism and violent offending: an 8-year prospective follow-up study. Psychiatry Res. 2009; 168:26–31.
- Hicks AU, Lappalainen RS, Narkilahti S, Suuronen R, Corbett D, Sivenius J, et al. Transplantation of human embryonic stem cellderived neural precursor cells and enriched environment after cortical stroke in rats: cell survival and functional recovery. Eur J Neurosci. 2009;29:562–74.
- Oksanen M, Petersen AJ, Naumenko N, Puttonen K, Lehtonen Š, Gubert Olivé M, et al. PSEN1 mutant iPSC-derived model reveals severe astrocyte pathology in Alzheimer's disease. Stem Cell Rep. 2017;9:1885–97.
- O'Roak BJ, Vives L, Girirajan S, Karakoc E, Krumm N, Coe BP, et al. Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations. Nature. 2012;485:246–50.
- Wang P, Mokhtari R, Pedrosa E, Kirschenbaum M, Bayrak C, Zheng D, et al. CRISPR/Cas9-mediated heterozygous knockout of the autism gene CHD8 and characterization of its transcriptional networks in cerebral organoids derived from iPS cells. Mol Autism. 2017;8:11.
- Klauck SM, Felder B, Kolb-Kokocinski A, Schuster C, Chiocchetti A, Schupp I, et al. Mutations in the ribosomal protein gene RPL10 suggest a novel modulating disease mechanism for autism. Mol Psychiatry. 2006;11:1073–84.
- Chiocchetti A, Pakalapati G, Duketis E, Wiemann S, Poustka A, Poustka F, et al. Mutation and expression analyses of the ribosomal protein gene RPL10 in an extended German sample of patients with autism spectrum disorder. Am J Med Genet A. 2011;155A:1472-5.
- Chiocchetti AG, Haslinger D, Boesch M, Karl T, Wiemann S, Freitag CM, et al. Protein signatures of oxidative stress response in a patient specific cell line model for autism. Mol Autism. 2014;5:10.
- Redies C, Hertel N, Hübner CA. Cadherins and neuropsychiatric disorders. Brain Res. 2012;1470:130–44.
- Tommerup N, Vissing H. Isolation and fine mapping of 16 novel human zinc finger-encoding cDNAs identify putative candidate genes for developmental and malignant disorders. Genomics. 1995;27:259–64.
- 24. Gtex portal. https://gtexportal.org/home/gene/ZNF132.
- Carta I, Chen CH, Schott AL, Dorizan S, Khodakhah K. Cerebellar modulation of the reward circuitry and social behavior. Science. 2019;363:eaav0581.
- Umeda-Yano S, Hashimoto R, Yamamori H, Weickert CS, Yasuda Y, Ohi K, et al. Expression analysis of the genes identified

- in GWAS of the postmortem brain tissues from patients with schizophrenia. Neurosci Lett. 2014;568:12–16.
- Lux H, Flammann H, Hafner M, Lux A. Genetic and molecular analyses of PEG10 reveal new aspects of genomic organization, transcription and translation, PLoS ONE, 2010;5:e8686.
- Kolc KL, Sadleir LG, Scheffer IE, Ivancevic A, Roberts R, Pham D, et al. A systematic review and meta-analysis of 271 PCDH19variant individuals identifies psychiatric comorbidities, and association of seizure onset and disease severity. Mol Psychiatry. 2019;24:241–51.
- Breuillard D, Leunen D, Chemaly N, Auclair L, Pinard JM, Kaminska A, et al. Autism spectrum disorder phenotype and intellectual disability in females with epilepsy and PCDH-19 mutations. Epilepsy Behav. 2016;60:75–80.
- Meloni I, Muscettola M, Raynaud M, Longo I, Bruttini M, Moizard MP, et al. FACL4, encoding fatty acid-CoA ligase 4, is mutated in nonspecific X-linked mental retardation. Nat Genet. 2002;30:436–40.
- Ansari IH, Longacre MJ, Stoker SW, Kendrick MA, O'Neill LM, Zitur LJ, et al. Characterization of Acyl-CoA synthetase isoforms in pancreatic beta cells: Gene silencing shows participation of ACSL3 and ACSL4 in insulin secretion. Arch Biochem Biophys. 2017;618:32–43.
- 32. Kim JH, Shinde DN, Reijnders MRF, Hauser NS, Belmonte RL, Wilson GR, et al. De novo mutations in SON disrupt RNA splicing of genes essential for brain development and metabolism, causing an intellectual-disability syndrome. Am J Hum Genet. 2016;99:711–9.
- Kim JE, Hong YH, Kim JY, Jeon GS, Jung JH, Yoon BN, et al. Altered nucleocytoplasmic proteome and transcriptome distributions in an in vitro model of amyotrophic lateral sclerosis. PLoS ONE. 2017;12:e0176462.
- Zheng JS, Arnett DK, Parnell LD, Lee YC, Ma Y, Smith CE, et al. Genetic variants at PSMD3 interact with dietary fat and carbohydrate to modulate insulin resistance. J Nutr. 2013;143:354–61.
- 35. Virkkunen M, Rawlings R, Tokola R, Poland RE, Guidotti A, Nemeroff C, et al. CSF biochemistries, glucose metabolism, and diurnal activity rhythms in alcoholic, violent offenders, fire setters, and healthy volunteers. Arch Gen Psychiatry. 1994;51:20–27.
- Hu H, Coon H, Li M, Yandell M, Huff CD. VARPRISM: incorporating variant prioritization in tests of de novo mutation association. Genome Med. 2016;8:91.
- Nummenmaa L, Manninen S, Tuominen L, Hirvonen J, Kalliokoski KK, Nuutila P, et al. Adult attachment style is associated with cerebral μ-opioid receptor availability in humans. Hum Brain Mapp. 2015;36:3621–8.
- Rütgen M, Seidel EM, Silani G, Riečanský I, Hummer A, Windischberger C, et al. Placebo analgesia and its opioidergic regulation suggest that empathy for pain is grounded in self pain. Proc Natl Acad Sci USA. 2015;112:E5638–E5646.