

1 **Phylogenomics and biogeography of the world's thrushes (Aves, *Turdus*): New evidence**
2 **for a more parsimonious evolutionary history.**

3 Romina Batista^{1,2,3*}, Urban Olsson^{3,4}, Tobias Andermann^{3,4}, Alexandre Aleixo⁵, Camila
4 Cherem Ribas^{6, †}, Alexandre Antonelli^{3,4,7 †}

5
6 ¹ Programa de Pós-Graduação em Genética, Conservação e Biologia Evolutiva, PPG GCBEv
7 - Instituto Nacional de Pesquisas da Amazônia- INPA Campus II. Av. André Araújo, 2936 –
8 Petrópolis - CEP 69067-375 - Manaus - AM, Brazil

9 ² Coordenação de Zoologia, Laboratório de Biologia Molecular, Museu Paraense Emílio
10 Goeldi, CEP 66077-830, Belém - PA, Brazil

11 ³ Gothenburg Global Biodiversity Centre, Box 461, SE-405 30 Gothenburg, Sweden

12 ⁴ Department of Biological and Environmental Sciences, University of Gothenburg,
13 SE-413 19 Gothenburg, Sweden

14 ⁵ Finnish Museum of Natural History, University of Helsinki, P.O. Box 17, 00014, Helsinki,
15 Finland

16 ⁶ Instituto Nacional de Pesquisas da Amazônia, INPA Campus II. Av. André Araújo 2936,
17 CEP 69060-000, Manaus-AM, Brazil

18 ⁷ Royal Botanic Gardens, Kew, Richmond, Surrey, TW9 3AE, U.K.

19
20 † Joint senior authors

21 *E-mail: rominassbatista@gmail.com

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26 **Abstract**

27 To elucidate the relationships and spatial range evolution across the world of the bird genus
28 *Turdus* (Aves), we produced a large genomic dataset comprising ca. 2 million nucleotides for
29 ca. 100 samples representing 53 species, including over 2,000 loci. We estimated time-
30 calibrated maximum likelihood and multispecies coalescent phylogenies and carried out
31 biogeographic analyses. Our results indicate that there have been considerably fewer trans-
32 oceanic dispersals within the genus *Turdus* than previously suggested, such that the
33 Palaeartic clade did not originate in America and the African clade was not involved in the
34 colonization of the Americas. Instead, our findings suggest that dispersal from the Western
35 Palaeartic via the Antilles to the Neotropics might have occurred in a single event, giving
36 rise to the rich Neotropical diversity of *Turdus* observed today, with no reverse dispersals to
37 the Palaeartic or Africa. Our large multilocus dataset, combined with dense species-level
38 sampling and analysed under probabilistic methods, brings important insights into historical
39 biogeography and systematics, even in a scenario of fast and spatially complex
40 diversification.

41
42 **Keywords:** *Turdus*, dispersal, next-generation sequencing, species radiations.
43

44 Introduction

45

46 The family Turdidae (Aves, Passeriformes) comprises 21 genera distributed worldwide,
47 where *Turdus* is the most species-rich genus (86 spp. [1]), accounting for half of the family's
48 species diversity. Species in the genus are generally similar in shape, but show large variation
49 in plumage colouration and ecology, exhibiting a variety of colour patterns and inhabiting
50 almost all biomes in the world – including, savannas, alpine areas and both tropical and
51 temperate forests [1]. This genus contains many common and familiar species throughout the
52 world, such as the Blackbird (*Turdus merula*), the American Robin (*Turdus migratorius*) and
53 the Clay-colored Thrush or yigüirro (*Turdus grayi*).

54

55 Previous attempts to resolve phylogenetic relationships within *Turdus* have yielded
56 conflicting phylogenetic hypotheses, and, consequently, different biogeographical
57 interpretations [2-4]. One of their main limitations so far has been the reliance on a very
58 limited number of independent genetic markers, making the analyses vulnerable to stochastic
59 effects. Voelker *et al.* [2], using only mitochondrial DNA, recovered four main clades
60 (African, Central America-Caribbean, South American and Eurasian), but their topology
61 lacked nodal support for all but the Eurasian and a South American clade, with the latter also
62 including an African species. Based on their results, the authors suggested including the
63 genera *Platycichla*, *Cichlerminia* and *Nesocichla* within *Turdus*. In another study, Nylander
64 *et al.* [3] analysed a combined data set from mitochondrial and nuclear markers and inferred
65 one Eurasian, three American and three African clades. Their results led those authors to
66 suggest that: i) the apparent non-monophyly of the African taxa might be illusory; ii) several
67 independent dispersal events have taken place from an African source to South America; and
68 iii) that *Platycichla*, *Cichlerminia* and *Nesocichla* should indeed be placed within *Turdus*.
69 Due to the partially conflicting results of these studies, the evolutionary relationships and
70 biogeographical history within *Turdus* remain unresolved.

71

72 The phylogenetic uncertainties concern not only relationships and biogeographic history
73 among the major clades in *Turdus*, but also the taxonomic and phylogeographic implications
74 of recently diverged polytypic species and species complexes. Recent phylogeographic
75 studies of species complexes within *Turdus* revealed additional taxonomic complexity at the
76 level previously regarded as intraspecific, indicating that current taxonomy underestimates
77 the true species diversity within the genus [5-8].

78

79 In summary, the controversies surrounding the early diversification and biogeography of
80 *Turdus*, the circumscription and relationships among major clades and species, and our
81 insufficient knowledge of intraspecific divergences and species limits, make *Turdus* a prime
82 candidate for testing the utility of phylogenomic approaches for tackling evolutionary
83 questions among and within bird species.

84

85 In this study, we address the following questions: 1) Can phylogenomic data confidently
86 delimit major clades within *Turdus* and elucidate the relationships among them? 2) How
87 many trans-Atlantic dispersals happened during the diversification of *Turdus*, leading to its
88 current cosmopolitan distribution? and 3) Has the African continent functioned as a source
89 for intercontinental dispersals? To address these questions, we infer the phylogeny of the
90 genus *Turdus* based on novel and extensive genomic data (ultraconserved elements (UCEs),
91 intragenic and untranslated regions, respectively) and multiple analytical approaches
92 (maximum likelihood, species tree inference, ancestral area reconstructions). To maximize
93 taxonomic sampling, we also compiled and analysed a dataset of sequences of the

94 mitochondrial marker cytochrome *b* (*cytb*), for all currently recognized species within the
95 genus [2, 3, 5, 6, 9].

96

97 **Methods**

98 A full description of the data and methods can be found in the electronic supplementary
99 material (Table S1, S2, S3, S4, Fig. S1). The overall sampling included 115 individuals from
100 53 species (~62%) of 86 currently recognized species within the genus *Turdus* [1]. We used a
101 UCE probe set developed by Faircloth *et al.* [10], and developed additional specific probes
102 for *Turdus* based on 49 of the loci described in Backström *et al.* [11].

103

104 **Phylogenomic and divergence time estimates**

105 We performed unpartitioned concatenated maximum likelihood (ML) analyses in RAxML
106 8.2.9 [12] for two data sets: 1) a matrix including UCEs (two different thresholds of missing
107 data, 75% and 85%); and 2) a matrix including the additional non-UCE loci (intra-genic and
108 untranslated regions). We assessed support for the best ML topology by performing 150
109 nonparametric bootstrap replicates using the autoMRE option in RAxML with the GTR
110 GAMMA site-rate substitution model.

111

112 To avoid loci that could bias species tree analyses, we calculated the number of parsimony-
113 informative sites (PIS) for each UCE locus and created a subset of UCE loci having a number
114 of PIS in the upper quartile of the range (every locus with equal or more than 65 PIS). For the
115 additional non-UCE loci, we included all markers recovered. We performed nonparametric
116 bootstrapping by sites for each locus using RAxML with GTR-GAMMA model of site-rate
117 substitution and sorted the bootstrapped gene trees following Seo [13], an approach for
118 estimating phylogeny and calculating bootstrapping using multilocus sequence data in the
119 context of a distance method. To infer the species tree and calculate nodal support from the
120 frequency of splits in trees constructed, we input the results to ASTRAL-II 4.7.8 [14]. We set
121 ASTRAL to run 150 bootstrap replicates of species trees. We also performed a species tree
122 analysis using SVDquartets [15] for the UCE loci, including all loci recovered. By analysing
123 fewer loci for the species tree, we were able to compare the results using different
124 approaches.

125

126 To estimate divergence times, we used two late Miocene fossils from Hungary, correlated to
127 the Mammal Neogene zone 13 (MN13) (7.2-4.9 mya). These two fossils were compared to *T.*
128 *viscivorus*, *T. pilaris* and *T. torquatus*, and *T. merula* and *T. philomelos*, respectively. We
129 tentatively interpret this as that they represent forms contemporary with the stem node of the
130 main global radiation of thrushes, and use them for setting the minimum age for the clade
131 including *all Turdus species minus T. philomelos and T. viscivorus* (Fig. 3). To fix a
132 maximum age for this same node, we used the minimum age for the split between Regulidae
133 and Certhioidea (17.2 Ma), based on fossil data and assigned in [16] (Fig. 3). As an
134 additional secondary calibration point, we added the split between Muscicapidae and
135 Turdidae, with a confidence interval of 22-13.5 Mya gleaned from [16] (Fig. 3). A UCE ML
136 tree (the topology recovering highest support values overall) was time-calibrated under
137 penalized likelihood using treePL [17]. The rate smoothing parameter was set to 100. The
138 smoothing value was established using the cross-validation parameter available in treePL, we
139 left the cvstart = 1000 (default) and cvstop = 0.1 (default).

140

141 **Biogeographic analyses**

142 To infer the spatial range evolution for *Turdus*, we used Biogeography with Bayesian (and
143 likelihood) Evolutionary Analysis in R Scripts – BioGeoBEARS, version 0.2.1 [18]. We

144 used the time-calibrated phylogeny for *Turdus* calculated in treePL and coded each species as
145 present or absent, according to their ranges following del Hoyo [1]. Eight biogeographical
146 areas were defined based on the zoogeographical realms estimated in [19], with adaptations
147 (Fig. 3): 1. Afrotropical (Afr), 2. Western Palaearctic (WP), 3. Eastern Palaearctic (EP); 4.
148 Oriental (O), 5. Nearctic (N), 6. Panamanian (C); 7. Antilles (Ant); 8. South America (SA).
149

150 **Results**

151 **Phylogenetic relationships**

152 Our results using different thresholds of concatenated UCEs datasets (75% and 85%)
153 recovered *Turdus viscivorus* (historical samples GNM8238 and GNM172, from Sweden) and
154 *T. philomelos* (U4329, from Belarus) as incremental sisters to all other *Turdus* species (Fig.
155 1, Fig. S2 and Fig. S3). In the main clade, all taxa from the Afrotropical Region, including *T.*
156 *pelios*, previously contentiously placed as part of the South American radiation, form a
157 strongly supported monophyletic radiation (Node A, Fig. 1, Fig. S2), that is sister to a clade
158 composed of all other species. The Palaearctic clade contains *T. iliacus* and *T. merula*, with
159 strong support. In previous studies, these two species have been found in varying
160 phylogenetic positions that suggest different biogeographical histories from other Western
161 Palaearctic species [2,3]. We consider the following cut-offs for bootstrap values: 50-74%
162 weak; 75-84% moderate and >85% as strong support (Fig. S2, Fig. 1).
163

164 While support for both the African and the Eurasian clades is strong, support for a
165 monophyletic group [Node D, Fig. 1] in the Americas is weak. The important first splits
166 within this clade are weakly supported, but the topology suggests a first divergence between
167 Antillean taxa (a strongly supported clade including *T. aurantius* from Jamaica, *T. plumbeus*
168 *schistaceus* from Cuba and *T. plumbeus ardosiacus* from Puerto Rico) and remaining taxa
169 (Fig. 1). The following basal divergences are all between taxa now inhabiting the Nearctic,
170 Panamanian or Antillean regions [19]. Although the divergence pattern is mostly weakly
171 supported, a biogeographical structure may be discernible. A strongly supported clade
172 comprises mainly species found in Central America (*T. rufitorques*, *T. nigrescens*, *T.*
173 *infuscatus*), but also *T. migratorius*, a species widely distributed in North America. An
174 exclusively Panamanian clade (strongly supported) consists of two subspecies of *T. plebejus*
175 (*rafaelenses* from Nicaragua and *plebejus* from Panama). A large and strongly supported
176 monophyletic Neotropical radiation (node E; Fig. 1) includes the early (but poorly supported)
177 sequential divergence of two species from the Antilles (*T. swalesi* from Hispaniola and *T.*
178 *lherminieri* from Dominica), and is further subdivided into two main subclades
179 corresponding to highland and lowland species, respectively.
180

181 The topology based on 49 additional loci (Fig. S4) includes a concatenated matrix of 65,574
182 bp, using a taxon set with 105 individuals. Bootstrap (BS) support for a monophyletic *Turdus*
183 is 100% (Fig. S4). *Turdus viscivorus* is sister to the remainder of the clade with strong
184 support. The next lineage to diverge is *T. philomelos*, which in turn is sister to the remainder
185 of the clade. A basal split separates two clades with low BS support values – one large group
186 includes species from the Palaearctic, Africa, Nearctic, Panamanian, as well as the Antillean
187 regions, and is sister to a group which includes species from the Antilles (*T. swalesi* and *T.*
188 *lherminieri*) and all samples from the continental South America Region (Fig. S4, weakly
189 supported). Support for a monophyletic African radiation is strong, and *T. swalesi* and *T.*
190 *lherminieri* are recovered (although with weak support) as part of the Neotropical radiation,
191 as for the UCE data set (Node E, Fig. 1, Fig. S2 and Fig. S3).
192

193 The analysis of the *cytb* data set (Fig. S5) recovers a monophyletic *Turdus* clade (excluding
194 *Otocichla mupinensis* and *Psophocichla litsitsirupa*, which were recovered as sisters to the
195 *Turdus* clade and which have both previously been included in *Turdus*), where *T. viscivorus*
196 and *T. philomelos* are again the first lineages to diverge, being sisters to the remainder of the
197 clade with high support. Within the main clade, most early diverging nodes are unsupported,
198 but otherwise there are many similarities to previous studies [2, 3]. African species are
199 divided into six clades, with *T. pelios* sister to Antillean species and a Eurasian clade with the
200 exception of *T. merula* and *T. iliacus*. Unlike previous studies [2, 3], there is for the first time
201 high support for a monophyletic South American clade based on a single mitochondrial
202 locus. A number of unexpectedly deep or shallow divergences of potential taxonomic interest
203 are highlighted in Fig. S5 and discussed in the supplementary material.

204

205 The UCE species tree (Fig. S6, with gene trees of 483 UCE loci as input in ASTRAL)
206 recovers a similar topology to the concatenated UCEs and *cytb* regarding *T. viscivorus* and *T.*
207 *philomelos*. The placement of these species was also recovered, although in reverse order
208 compared to concatenated UCEs and *cytb*. Here, *T. viscivorus* and *T. philomelos* are sisters to
209 a strongly supported clade, which consists of two major groups: one including Panamanian,
210 Western Palaearctic, Antillean, African, Eastern Palaearctic; and one South American group
211 (Node D, Fig. 1), with a few Panamanian and Antillean species, plus two Eastern Palaearctic
212 species, *Turdus eunomus* and *T. naumanni*. Support for the diverging order is weak, but
213 several of the same groups as in Fig. 1 are recovered with strong support, e.g. *T. plebejus*, the
214 Central American clade (*T. rufitorques*, *T. nigrescens*, *T. infuscatus*, *T. migratorius*), the
215 African clade and the Eurasian clade. Also, the Neotropical clade is similar to the topology in
216 Fig. 1, including the division between the highland and lowland clades.

217

218 The UCE species tree (Fig. S7, with 1,931 UCE loci as input in SVDquartets) shows *T.*
219 *lherminieri* from the Antilles as sister species to all other species of the crown group. Again,
220 this analysis recovers most of the same clades as the other analyses performed in this study,
221 while there is no resolution regarding the phylogenetic relationships among these large
222 groups. The similarities between analyses are the recovery of a monophyletic African group,
223 a monophyletic Eurasian group, a monophyletic Panamanian group and a generally similar
224 Neotropical clade.

225

226 The species tree built from the 49 additional loci (Fig. S8) (with gene trees of each additional
227 locus as input) recovers a tree topology where the species *Turdus viscivorus* and *T.*
228 *philomelos* are again the first to diverge, followed by two main groups (Fig. S8): a
229 moderately supported Afrotropical clade sister to the remainder of the thrushes from all non-
230 African regions. In this tree, the main topology is insufficiently supported, but major clades
231 such as the Eurasian clade including *T. iliacus* and *T. merula* are recovered, although support
232 is weak. The American radiation is recovered as monophyletic, but again with weak support.
233 Although there are differences in detail, major patterns in the Panamanian and Neotropical
234 clades proposed by previous analyses shown here are corroborated.

235

236 **Divergence time estimates**

237 The crown age of *Turdus* is estimated to c. 9.3 million years (Ma) (Fig. 3, Fig. S9), in
238 late Miocene. The first event of the major global radiation starts c. 7.2 million years
239 ago (Mya), corresponding to the divergence of the African clade, rapidly followed by the
240 divergence at c. 5.7 Mya between the Palaearctic and Oriental clades from the clade
241 comprising species with distributions in the New World. Around 5.3 Mya, the New World

242 radiation begins. Panamanian and South American clades first begin to diverge at c. 4.4 Mya
243 and 4.9 Mya, respectively.

244

245 **Geographical ancestral range reconstruction**

246 Including the “jump dispersal” (j) parameter in the DEC model (Fig. 3), implemented in
247 BioGeoBEARS, suggests a similar scenario as the DEC reconstruction (not shown), but
248 causes the reconstructions at the nodes to become less noisy. Independent dispersals
249 originating in the Western Palaeartic and moving towards the Eastern Palaeartic continuing
250 to the Oriental region are suggested. The reconstruction of the New World radiation
251 (including N, C, GA and SA, see Fig. 3 for abbreviations) suggests that it originated from a
252 single dispersal event from the Western Palaeartic to the Antilles, although weak clade
253 support makes this conclusion tentative (Fig. 1). From the Antilles, one or two dispersals to
254 the Central America region are inferred, but the interpretation is complicated. From the
255 Panamanian region, one lineage (*T. migratorius*) was recovered to have dispersed to North
256 America. The radiation in South America is inferred to have originated directly from an
257 ancestor dispersing from the Great Antilles. Separate dispersals back to the Central America
258 region from South America are implied for the ancestors of *T. assimilis* and *T. grayi* (Fig. 3).

259

260 **Discussion**

261

262 **Data considerations**

263 The different evolutionary scenarios that have been suggested by studies of *Turdus* [2, 3]
264 propose conflicting scenarios of how they spread across the world. Most likely, the main
265 reasons for the discrepancies are the differences in the amount and source of genetic data,
266 combined with the stochasticity introduced by the short internodes caused by the high rate of
267 speciation during the radiation. The UCE topology (ML) with 1,931 concatenated loci (75%
268 completeness of taxa) is the best currently available overview of the *Turdus* radiation, due to
269 the highest number of loci sampled and the better resolution recovered to date (Fig. 1 and
270 Fig. S2). The concatenated UCE dataset shows reduced incongruence and improved
271 resolution of the topology, as already documented for this kind of approach [20]. The more
272 limited genetic sampling used by Voelker *et al.* [2] and Nylander *et al.* [3] makes these
273 analyses more vulnerable to disagreement between the genealogy of the loci analysed and the
274 species tree, as well as to the effects of various evolutionary processes such as incomplete
275 lineage sorting, gene flow and hybridization. By exploring the different kinds of datasets
276 obtained by genomic sampling, we were able to estimate a novel phylogenetic hypothesis
277 based on 1,931 UCEs (Fig. 1, Fig. S2 and Fig. S3). These data suggest a much simpler and
278 parsimonious dispersal scenario compared to previous studies.

279

280 **Divergence time estimates**

281 The age of the split between *Oenanthe* and *Turdidae* was estimated to 22 Mya, corresponding
282 to the upper extent of the confidence limit given for this split in Fig. 2 in Oliveros *et al.* [16].
283 This places the radiation of *Turdidae* within a time frame compatible with several recent
284 studies [16, 21-24]. Furthermore, the age of divergences between species are also compatible
285 with the standard 2.1% per million years divergence rate in the cytochrome b gene [25].

286

287 **Phylogenetic implications and the historical biogeography of *Turdus***

288 Our time-calibrated phylogeny based on 1,931 UCEs (Fig. 3, Fig. S9) sheds new light on the
289 relationships among the majority of species in the genus *Turdus*. Previous studies suggested a
290 series of trans-Atlantic dispersal events, both from Africa to the New World and from the
291 Caribbean Islands to the Old World. Our findings challenge these hypotheses and that the

292 African thrushes acted as a source for dispersal to the New World [3, 26], and suggest a
293 different scenario for the evolution of the genus *Turdus*. Although we cannot completely rule
294 out the possibility of more than one trans-Atlantic dispersal (see *Phylogenetic incongruences*
295 *leading to alternative phylogenetic histories* in the electronic supplementary material), the
296 topologies allowing for more than one transatlantic dispersal are unsupported in that respect,
297 and in some details most likely spurious. Our statistically most well supported results based
298 on 1,931 UCEs differ most importantly from previous hypotheses in that i) the African
299 radiation is monophyletic; ii) Eurasian taxa did not originate from a west to east transatlantic
300 dispersal; and iii) that the American taxa originate from a single dispersal from the Old
301 World (Western Palaearctic) to the Americas, a result recovered also with low support in the
302 analysis of 49 independent loci.

303
304 Ancestral lineages are here inferred to having occurred in the Western Palaearctic, and from
305 there, dispersals to the Afrotropical, Eastern Palaearctic and South American regions took
306 place. The Afrotropical radiation is inferred to have been entirely *in situ*, with no indication
307 that it had any role in the colonization of South America (Fig. 3, Fig. S9). The radiation in the
308 Old World seems to have originated in the Western Palaearctic, with a subsequent spread
309 towards the Eastern Palaearctic and Oriental regions on at least two separate occasions (Fig.
310 3, Fig. S9). We note, however, that genomic sampling of Palaearctic species is incomplete,
311 making these conclusions tentative. Contrary to previous hypotheses, *Turdus pelios* is
312 recovered in the African clade and is no longer suggested to be more closely related to
313 species from the Caribbean (Fig.2, Fig. 3), thus contradicting both a close relationship with
314 other taxa from South America [2] as well as a reversed transatlantic dispersal event, as
315 inferred by both Nylander *et al.* [3] and Voelker *et al.* [2]. *T. philomelos* and *T. viscivorus*
316 represent ancient lineages that seem to have been unaffected by the processes driving the
317 global radiation from the Palaearctic region (Fig. 1 and Fig. 3, Fig. S2-S9). *Turdus iliacus* is
318 inferred to be sister to *T. merula* and both are part of the Palaearctic clade, with strong
319 support. In contrast, both Voelker *et al.* [2] and Nylander *et al.* [3] recovered *T. iliacus* within
320 the New World clade, as sister to *T. plebejus*, a Panamanian species.

321
322 We find evidence for the Antilles as the location of the first colonization event (Fig. 3). In the
323 ancestral range reconstruction shown in Fig. 3, an Antillean distribution receives the highest
324 probability, but Central America is also a possibility. *Turdus* thus seems to be an example of
325 a long-distance dispersal from the Palaearctic where the birds first settled in the Caribbean
326 region and then proceeded to radiate and colonize the mainland. These new data receive
327 higher phylogenetic support than previous studies, and generate a both more parsimonious
328 explanation and a more logical scenario for the colonization of the Americas by *Turdus*.

329
330 The initial events in the colonisation of the Americas seem to be centred around the
331 Caribbean region. The most likely scenario according to the ancestral range reconstruction
332 (Fig. 3) is a dispersal from a founder population in the Greater Antilles to Central America,
333 resulting in one limited radiation including the ancestors of the *T. rufitorques* clade and in the
334 *T. plebejus* lineage, followed by a reverse dispersal back to the Antilles from the Panamanian
335 region by the ancestors of *T. swalesi* and *T. lherminieri*. In an alternative scenario, including
336 two dispersals from the Greater Antilles to the Panamanian region, the ancestor of the clade
337 including *T. rufitorques* would have had to disperse first, followed by the ancestor of *T.*
338 *plebejus*, while the ancestors of *T. swalesi* and *T. lherminieri* would emanate from dispersal
339 events within the Antilles.

340

341 Thrushes probably entered the Panamanian region by the time a land bridge was already fully
342 or partly established [27-29]. There is an ongoing debate on the emergence of the
343 Panamanian land bridge and the timing and processes underlying the Great American Biotic
344 Interchange [29], which is partly due to difficulties in the interpretation of the exposure of
345 land and shallow waters [28]. The closure of the Panamanian land bridge has had an
346 important impact on the diversification of a variety of New World birds, as shown by Smith
347 and Klicka [30]. Weir *et al.* [31], Töpel *et al.* [32], Antonelli *et al.* [33], Oliveros *et al.* [34],
348 among others, have shown a marked increase in the rates of interchange of South and Central
349 American birds, after or in connection with a land bridge completion. These studies suggest
350 that the route of most dispersal events was primarily south to north.

351
352 Whilst a common pattern for birds, thrushes are likely to only have been marginally affected
353 by the emergence of the Panamanian uplift. The colonization of South America is instead
354 inferred to be a dispersal event directly from the Antilles, with no evidence of any connection
355 to the Panamanian land bridge. This is not surprising, since these birds evidently dispersed to
356 and between the Antillean islands, and could thus have dispersed between the Panamanian or
357 Antillean regions and continental South America independently of a land bridge. This result
358 shows similarities to findings from studies of other taxa. For instance, Sturge *et al.* [35]
359 showed, based on mitochondrial DNA (cytochrome b and ND2), that the colonization of
360 South America by New World orioles (*Icterus*) is likely to have involved ancestors from the
361 Caribbean, and a recent biogeographical reconstruction recovered reverse colonization by
362 short-faced bats from the islands (e.g. Antilles) to South America [36]. Other examples
363 include the genus *Megascops* (Aves, Strigidae) endemic to the New World with a likely
364 origin in the Caribbean Islands [37].

365
366 By improving the taxonomic and geographic sampling within South America, we recovered
367 for the first time a single, monophyletic South American clade within *Turdus*. In contrast,
368 Nylander *et al.* [3] inferred three South American clades, and thus suggested independent
369 colonization events. Our results regarding the South American radiation are similar to those
370 of Voelker *et al.* [2] in this respect, apart from the African species (*T. pelios*) that was part of
371 the American clade in their analysis but is part of the African clade in this study (Fig. 1, Fig.
372 2, Fig. S2).

373
374 According to our phylogenetic hypothesis, the South American clade is divided into two
375 main groups. One comprises mainly highland species (mostly Andean and Tepuian, but one
376 Atlantic Forest species) and corresponds to an Andean clade already identified in Voelker,
377 Rohwer [2]. The second main clade includes species from the lowlands in Amazonia, Chaco,
378 Cerrado and Pampas, as well as Central America (*T. grayi* and *T. assimilis*). Paraphyly of *T.*
379 *olivater* was detected, with *T. olivater kemptoni* (U4322, Venezuela) being recovered as sister
380 to *T. fuscater fuscater* (U4285, Bolivia) and the Tepuian *T. olivater roraimae* sister to these
381 two. This pattern has been documented in a previous study using a multilocus dataset [37].
382 Similarly, *T. fuscater* appears as paraphyletic, with *T. fuscater gigantoides* (U4287, Peru)
383 being sister to *T. chiguanco conradi* (U4279, Peru) [38]. From continental South America, a
384 number of independent reversals – dispersal events to both the Antilles and the Panamanian
385 region – are inferred, which in the latter case could have been aided by the land bridge.

386
387 Future studies could focus on clarifying the geographical range evolution of *Turdus* within
388 South America and along altitudinal zones, as well as investigating the influence of landscape
389 and climatic changes on their past and current distribution and genetic diversity.

390

391 **Conclusions**

392 By employing broad genomic data with dense taxonomic sampling (see Fig. S10 for an
393 overview of all approaches used in this study), we were able to not only provide an improved
394 phylogenomic framework for continued taxonomy and classification revisions, but also a
395 starting point for more detailed studies of the morphological, behavioural and ecological
396 evolution in thrushes. In addition, our results provide strong evidence to reconsider ongoing
397 controversies on the early evolution and range evolution of *Turdus*, including the logical but
398 novel inference of a monophyletic *in situ* radiation in Africa, no American origin of the major
399 Palearctic radiation and a single rather than multiple trans-Atlantic dispersals giving rise to
400 the American radiation. In summary, phylogenomic analyses are not only useful at higher
401 taxonomic levels [39], but also hold the potential to uncover new insights into the
402 diversification and biogeographic history of rapid species radiations.

403

404 **References**

- 405 [1] del Hoyo, J., Elliott, A., Sargatal, J., Christie, D.A. & Kirwan, G. 2019 *Handbook of the*
406 *Birds of the World Alive*, Lynx Edicions.
- 407 [2] Voelker, G., Rohwer, S., Bowie, R.C.K. & Outlaw, D.C. 2007 Molecular systematics of a
408 speciose, cosmopolitan songbird genus: Defining the limits of, and relationships among, the
409 *Turdus* thrushes. *Molecular Phylogenetics and Evolution* **42**, 422-434.
410 (doi:10.1016/j.ympev.2006.07.016).
- 411 [3] Nylander, J.A., Olsson, U., Alstrom, P. & Sanmartin, I. 2008 Accounting for phylogenetic
412 uncertainty in biogeography: a Bayesian approach to dispersal-vicariance analysis of the
413 thrushes (Aves: *Turdus*). *Syst Biol* **57**, 257-268. (doi:10.1080/10635150802044003).
- 414 [4] Nagy, J., Végvári, Z. & Varga, Z. 2019 Phylogeny, migration and life history: filling the
415 gaps in the origin and biogeography of the *Turdus* thrushes. *Journal of Ornithology* **160**, 529-
416 543. (doi:10.1007/s10336-019-01632-3).
- 417 [5] Melo, M., Bowie, R.C.K., Voelker, G., Dallimer, M., Collar, N.J. & Jones, P.J. 2010
418 Multiple lines of evidence support the recognition of a very rare bird species: the Príncipe
419 thrush. *Journal of Zoology*, no-no. (doi:10.1111/j.1469-7998.2010.00720.x).
- 420 [6] Núñez-Zapata, J. & Peterson, A.T. 2016 Pleistocene diversification and speciation of
421 White-throated Thrush (*Turdus assimilis*; Aves: Turdidae). *Journal of Ornithology*.
- 422 [7] Cerqueira, P.V., Santos, M.P.D. & Aleixo, A. 2016 Phylogeography, inter-specific limits
423 and diversification of *Turdus ignobilis* (Aves: Turdidae). *Mol Phylogenet Evol* **97**, 177-186.
424 (doi:10.1016/j.ympev.2016.01.005).
- 425 [8] Avendano, J.E., Arbelaez-Cortes, E. & Cadena, C.D. 2017 On the importance of
426 geographic and taxonomic sampling in phylogeography: A reevaluation of diversification and
427 species limits in a Neotropical thrush (Aves, Turdidae). *Mol Phylogenet Evol* **111**, 87-97.
428 (doi:10.1016/j.ympev.2017.03.020).
- 429 [9] Bowie, R.C.K., Bloomer, P., Clancey, P.A. & Crowe, T.M. 2003 The Karoo Thrush
430 (*Turdus smithi* Bonaparte 1850), a southern African endemic. *Ostrich* **74**, 1-7.
431 (doi:10.2989/00306520309485364).
- 432 [10] Faircloth, B.C., McCormack, J.E., Crawford, N.G., Harvey, M.G., Brumfield, R.T. &
433 Glenn, T.C. 2012 Ultraconserved elements anchor thousands of genetic markers spanning
434 multiple evolutionary timescales. *Syst Biol* **61**, 717-726. (doi:10.1093/sysbio/sys004).
- 435 [11] Backström, N., Fagerberg, S. & Ellegren, H. 2008 Genomics of natural bird populations:
436 a gene-based set of reference markers evenly spread across the avian genome. *Mol Ecol* **17**,
437 964-980. (doi:10.1111/j.1365-294X.2007.03551.x).
- 438 [12] Stamatakis, A. 2014 RAxML version 8: a tool for phylogenetic analysis and post-
439 analysis of large phylogenies. *Bioinformatics* **30**, 1312-1313.
440 (doi:10.1093/bioinformatics/btu033).

- 441 [13] Seo, T.K. 2008 Calculating bootstrap probabilities of phylogeny using multilocus
442 sequence data. *Mol Biol Evol* **25**, 960-971. (doi:10.1093/molbev/msn043).
- 443 [14] Mirarab, S., Reaz, R., Bayzid, M.S., Zimmermann, T., Swenson, M.S. & Warnow, T.
444 2014 ASTRAL: genome-scale coalescent-based species tree estimation. *Bioinformatics* **30**,
445 i541-548. (doi:10.1093/bioinformatics/btu462).
- 446 [15] Chifman, J. & Kubatko, L. 2014 Quartet inference from SNP data under the coalescent
447 model. *Bioinformatics* **30**, 3317-3324. (doi:10.1093/bioinformatics/btu530).
- 448 [16] Oliveros, C.H., Field, D.J., Ksepka, D.T., Barker, F.K., Aleixo, A., Andersen, M.J.,
449 Alström, P., Benz, B.W., Braun, E.L., Braun, M.J., et al. 2019 Earth history and the passerine
450 superradiation. *Proceedings of the National Academy of Sciences*, 201813206.
451 (doi:10.1073/pnas.1813206116).
- 452 [17] Smith, S.A. & O'Meara, B.C. 2012 treePL: divergence time estimation using penalized
453 likelihood for large phylogenies. *Bioinformatics* **28**, 2689-2690.
454 (doi:10.1093/bioinformatics/bts492).
- 455 [18] Matzke, N.J. 2014 Model selection in historical biogeography reveals that founder-event
456 speciation is a crucial process in Island Clades. *Syst Biol* **63**, 951-970.
457 (doi:10.1093/sysbio/syu056).
- 458 [19] Holt, B.G., Lessard, J.-P., Borregaard, M.K., Fritz, S.A., Araújo, M.B., Dimitrov, D.,
459 Fabre, P.-H., Graham, C.H., Graves, G.R., Jönsson, K.A., et al. 2013 An Update of Wallace's
460 Zoogeographic Regions of the World. *Science* **339**, 74-78. (doi:10.1126/science.1228282).
- 461 [20] Salichos, L. & Rokas, A. 2013 Inferring ancient divergences requires genes with strong
462 phylogenetic signals. *Nature* **497**, 327-331. (doi:10.1038/nature12130).
- 463 [21] Claramunt, S. & Cracraft, J. 2015 A new time tree reveals Earth history's imprint on the
464 evolution of modern birds. *Sci Adv* **1**, e1501005. (doi:10.1126/sciadv.1501005).
- 465 [22] Moyle, R.G., Oliveros, C.H., Andersen, M.J., Hosner, P.A., Benz, B.W., Manthey, J.D.,
466 Travers, S.L., Brown, R.M. & Faircloth, B.C. 2016 Tectonic collision and uplift of Wallacea
467 triggered the global songbird radiation. *Nat Commun* **7**, 12709. (doi:10.1038/ncomms12709).
- 468 [23] Prum, R.O., Berv, J.S., Dornburg, A., Field, D.J., Townsend, J.P., Lemmon, E.M. &
469 Lemmon, A.R. 2015 A comprehensive phylogeny of birds (Aves) using targeted next-
470 generation DNA sequencing. *Nature* **526**, 569-573. (doi:10.1038/nature15697).
- 471 [24] Selvatti, A.P., Gonzaga, L.P. & Russo, C.A. 2015 A Paleogene origin for crown
472 passerines and the diversification of the Oscines in the New World. *Mol Phylogenet Evol* **88**,
473 1-15. (doi:10.1016/j.ympev.2015.03.018).
- 474 [25] Weir, J.T. & Schluter, D. 2008 Calibrating the avian molecular clock. *Mol Ecol* **17**,
475 2321-2328. (doi:10.1111/j.1365-294X.2008.03742.x).
- 476 [26] Voelker, G., Rohwer, S., Outlaw, D.C. & Bowie, R.C.K. 2009 Repeated trans-Atlantic
477 dispersal catalysed a global songbird radiation. *Global Ecology and Biogeography* **18**, 41-49.
478 (doi:10.1111/j.1466-8238.2008.00423.x).
- 479 [27] Montes, C., Cardona, A., McFadden, R., Moron, S.E., Silva, C.A., Restrepo-Moreno, S.,
480 Ramirez, D.A., Hoyos, N., Wilson, J., Farris, D., et al. 2012 Evidence for middle Eocene and
481 younger land emergence in central Panama: Implications for Isthmus closure. *Geological*
482 *Society of America Bulletin* **124**, 780-799. (doi:10.1130/b30528.1).
- 483 [28] Bacon, C.D., Silvestro, D., Jaramillo, C., Smith, B.T., Chakrabarty, P. & Antonelli, A.
484 2015 Biological evidence supports an early and complex emergence of the Isthmus of
485 Panama. *Proceedings of the National Academy of Sciences* **112**, 6110-6115.
486 (doi:10.1073/pnas.1423853112).
- 487 [29] Jaramillo, C., Montes, C., Cardona, A., Silvestro, D., Antonelli, A. & Bacon, C.D. 2017
488 Comment (1) on "Formation of the Isthmus of Panama" by O'Dea et al. *Sci Adv* **3**, e1602321.
489 (doi:10.1126/sciadv.1602321).

- 490 [30] Smith, B.T. & Klicka, J. 2010 The profound influence of the Late Pliocene Panamanian
491 uplift on the exchange, diversification, and distribution of New World birds. *Ecography*.
492 (doi:10.1111/j.1600-0587.2009.06335.x).
- 493 [31] Weir, J.T., Bermingham, E. & Schluter, D. 2009 The Great American Biotic Interchange
494 in birds. *Proceedings of the National Academy of Sciences* **106**, 21737-21742.
495 (doi:10.1073/pnas.0903811106).
- 496 [32] Töpel, M., Zizka, A., Calio, M.F., Scharn, R., Silvestro, D. & Antonelli, A. 2017
497 SpeciesGeoCoder: Fast Categorization of Species Occurrences for Analyses of Biodiversity,
498 Biogeography, Ecology, and Evolution. *Syst Biol* **66**, 145-151. (doi:10.1093/sysbio/syw064).
- 499 [33] Antonelli, A., Zizka, A., Carvalho, F.A., Scharn, R., Bacon, C.D., Silvestro, D. &
500 Condamine, F.L. 2018 Amazonia is the primary source of Neotropical biodiversity. *Proc Natl*
501 *Acad Sci U S A*. (doi:10.1073/pnas.1713819115).
- 502 [34] Oliveros, C.H., Andersen, M.J., Hosner, P.A., Mauck, W.M., Sheldon, F.H., Cracraft, J.
503 & Moyle, R.G. 2019 Rapid Laurasian diversification of a pantropical bird family during the
504 Oligocene-Miocene transition. *Ibis*. (doi:10.1111/ibi.12707).
- 505 [35] Sturge, R.J., Jacobsen, F., Rosensteel, B.B., Neale, R.J. & Omland, K.E. 2009
506 Colonization of South America from Caribbean Islands Confirmed by Molecular Phylogeny
507 with Increased Taxon Sampling. *The Condor* **111**, 575-579. (doi:10.1525/cond.2009.080048).
- 508 [36] Tavares, V.d.C., Warsi, O.M., Balseiro, F., Mancina, C.A. & Dávalos, L.M. 2018 Out of
509 the Antilles: Fossil phylogenies support reverse colonization of bats to South America.
510 *Journal of Biogeography* **45**, 859-873. (doi:10.1111/jbi.13175).
- 511 [37] Dantas, S.M., Weckstein, J.D., Bates, J.M., Krabbe, N.K., Cadena, C.D., Robbins, M.B.,
512 Valderrama, E. & Aleixo, A. 2016 Molecular systematics of the new world screech-owls
513 (Megascops: Aves, Strigidae): biogeographic and taxonomic implications. *Mol Phylogenet*
514 *Evol* **94**, 626-634. (doi:10.1016/j.ympev.2015.09.025).
- 515 [38] Valderrama, E., Pérez-Emán, J.L., Brumfield, R.T., Cuervo, A.M., Cadena, C.D. &
516 Patten, M. 2014 The influence of the complex topography and dynamic history of the
517 montane Neotropics on the evolutionary differentiation of a cloud forest bird (*Premnoplex*
518 *brunnescens*, Furnariidae). *Journal of Biogeography* **41**, 1533-1546. (doi:10.1111/jbi.12317).
- 519 [39] Zhang, G. & Li, C. & Li, Q. & Li, B. & Larkin, D.M. & Lee, C. & Storz, J.F. &
520 Antunes, A. & Greenwold, M.J. & Meredith, R.W., et al. 2014 Comparative genomics
521 reveals insights into avian genome evolution and adaptation. *Science* **346**, 1311-1320.
522 (doi:10.1126/science.1251385).

523

524 **Data accessibility**

525 All DNA sequence data produced in this project are made freely available at NCBI Sequence
526 Read Archive (SRA) under the BioProject accession PRJNA574741 (Table S1). The
527 project's documentation, comprising origin of all data, download information, and basic
528 information, is freely available on GitHub: [https://github.com/RominaSSBatista/Turdus-](https://github.com/RominaSSBatista/Turdus-Project)
529 [Project](https://github.com/RominaSSBatista/Turdus-Project).

530

531 **Authors' contributions**

532 R.B., C.R., A. Aleixo, A. Antonelli and U.O. conceived, designed and coordinated the study
533 and helped draft the manuscript.; R.B and U.O carried out the molecular lab work; R.B., T.A
534 and U.O participated in data analysis; R.B. led the writing with contributions from all
535 authors. All authors gave final approval for publication.

536

537 **Competing interests**

538 The authors have no competing interests to declare.

539

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573

574 **Figure legends**

575

576 **Figure 1. Phylogenies of *Turdus*.** Topology with branch lengths, based on concatenated
577 maximum likelihood analysis of 1,931 UCE loci. Numbers on nodes indicate maximum
578 likelihood bootstrap support. Nodes without numbers indicate maximum likelihood bootstrap
579 support > 75%. Letters A-E depict the main nodes mentioned in the text. Branch in red
580 represent a historical sample (toepads) included in this study. Zoogeographical regions,
581 according to the current distribution of *Turdus* species, are provided to the right of the tree.
582 Shaded boxes in grey show the subdivision of Old/New World areas and two main groups
583 within South America (highland and lowland).

584

585 **Figure 2. Biogeographic hypotheses for *Turdus*.** Summarized from (a) Voelker *et al.*, 2007
586 (2 mitochondrial genes), asterisks above nodes indicate Bayesian posterior probability ≥ 0.95 ,
587 numbers below nodes indicate maximum likelihood bootstrap support (>50%); (b) Nylander
588 *et al.*, 2008 (2 mitochondrial and 2 nuclear genes), numbers above nodes indicate posterior

589 probability (>0.95); (c) This study (genomic sampling of 1,931 UCEs), numbers below nodes
590 indicate maximum likelihood bootstrap values. Only major clades are shown.

591

592 **Figure 3. Ancestral range reconstruction.** *Turdus* time-calibrated phylogeny, based on
593 1,931 UCEs, calibrated with one fossil calibration (black arrow), and with range evolution
594 estimated under the DEC+j model. Pie charts show the probability of each range. All nodes
595 including secondary colours represent the uncertainty to recover a unique ancestral range for
596 certain nodes within the phylogeny. The map shows the biogeographical regions used for the
597 analyses (adapted from Holt et al., 2013).

Table S1. List of taxa, locality and voucher information.

ID	Taxon	Locality	Voucher	Tissue Number	Country	Locality	Lat	Long	Level	Type	NCBI
U1172	<i>Tardus inflexus</i> Lafresque, 1844	Bufo Museum	UWBM 104175	DAB 4706	Guatemala	Departamento de Totonicapán, montañas noroeste Totonicapán	14.9108	-91.3656	INGROUP	Tissue	SAMN12944381
U1180	<i>Tardus inflexus</i> Lafresque, 1844	Bufo Museum	UWBM 104127	DK 0761	Panama	Provincia de Chiriquí, Parque Nacional Volcan Barú	8.8449	-82.3918	INGROUP	Tissue	SAMN12944382
U1419	<i>Tardus albicollis assimilis</i> Cabanis, 1850	Bufo Museum	UWBM 104125	DHB 4586	Guatemala	Departamento de Quezaltenango Santa María de Jesús 7 km SSW Finca de Santa María	14.3119	-91.5169	INGROUP	Tissue	SAMN12944383
U1422	<i>Tardus albicollis assimilis</i> Cabanis, 1850	Bufo Museum	UWBM 5433	DAB 837	Argentina	Provincia de Tucumán, San Miguel de Tucumán, El Infiernillo	23.475	-65.524	INGROUP	Tissue	SAMN12944384
U1464	<i>Tardus eraii</i> Boulenger, 1838	Bufo Museum	UWBM 18010	DHB 4341	Guatemala	Departamento de Retalhoulén, San Felipe Retalhoulén, 5 km S, Finca El Nido	14.2026	-91.5961	INGROUP	Tissue	SAMN12944385
U1468	<i>Tardus eraii</i> Boulenger, 1838	Bufo Museum	UWBM 10667	BFS 0710	Mexico	Campeche, Municipio de la Candelaria	19.1204	-99.1461	INGROUP	Tissue	SAMN12944386
U1477	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 5292	SAR 674	South Africa	KwaZulu-Natal Province, Melmoth	-28.59	31.44	INGROUP	Tissue	SAMN12944387
U2273	<i>Tardus amarucochilensis</i> Cabanis, 1850	Loisiana State University Museum of Natural Science	LSMZJ 7372	BOLV 163	Bolivia	Santa Cruz Department, Da Marayo de Chiquitos 126 km ENE San José	-17.7895	-63.1828	INGROUP	Tissue	SAMN12944388
U4334	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	LSMZJ 2992	SAK 044	Bolivia	Granit mine Province, U.S. Guantanamo Naval Base Castillo Hill	-7.52129	-82.0000	INGROUP	Tissue	SAMN12944389
U4442	<i>Tardus rufiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 31619	BOLV 163	Bolivia	Santa Cruz Department, Prov. El Tambo 14 km SE Comana 18 doores 00'23" 54 doores 20"	-18.0604	-64.43	INGROUP	Tissue	SAMN12944390
U2338	<i>Tardus aruanensis</i> Anderson, Hartlett, 1923	Department of Biology and Environmental Science, University of Gothenburg	DZUZ 2338		Antarctica	Aranzaz 2007-06-04 PFL	40.232	-64.9861	INGROUP	Tissue	SAMN12944391
U4176	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	LSMZJ 2992		South Africa	Amantsoo Ferryboat, CERRO DEL LA NEBLINA CAMP VIL 1800M	32.3796	-31.37	INGROUP	Tissue	SAMN12944392
U4345	<i>Tardus serranus</i> fuscicornatus (Chapman, 1912)	Loisiana State University Museum of Natural Science	LSMZJ 12152		Ecuador	Pachicha Province, MINDO	-0.0486	-78.7525	INGROUP	Tissue	SAMN12944393
U4225	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 4993		Denmark	Silkeborg, Tårnvej 10 km Karolinska Hospital	55.2258	11.9939	INGROUP	Tissue	SAMN12944394
U3899	<i>Tardus rufiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 31691		Japan	Francisco Morazan Department, Cerro YUYUCA Zamorano Bio logical reeducation station	-14.067	-87.0831	INGROUP	Tissue	SAMN12944395
U4220	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 41621		Panama	Rio del Tono Province, Chiriquí to Chiriquí Grande road	8.7919	-82.2094	INGROUP	Tissue	SAMN12944396
U4143	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 92132		Uruguay	Florida, Guacabiville UF Campus	-34.1843	-65.3828	INGROUP	Tissue	SAMN12944397
U4319	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 17111		Ecuador	Esmeraldas Province, EL PLACER CA 700M	-2.81047	-79.9899	INGROUP	Tissue	SAMN12944398
U4231	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 17130		Ecuador	Volcanal Country, San Aro 7 km SE Kooner	55.2258	11.9939	INGROUP	Tissue	SAMN12944399
U4300	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 224		Sweden	Province of Öland Östergötland, Öland	56.2777	16.9948	INGROUP	Tissue	SAMN12944400
U4337	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 42823		Vietnam	Everest Station 5 km SE West	-15.7378	107.6167	INGROUP	Tissue	SAMN12944401
U4322	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 7379		Bolivia	Carapaz Department, CERRO DEL LA NEBLINA CAMP VIL 1800M	32.3796	-65.778	INGROUP	Tissue	SAMN12944402
U4155	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 54448		Argentina	Provincia de Corrientes, Corrientes, Manuel Drouot	-27.8419	-58.1194	INGROUP	Tissue	SAMN12944403
U4236	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 5599		Malaysia	Wilayah Perak, Kuala Kangsar	6.0444	-103.7508	INGROUP	Tissue	SAMN12944404
U4339	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 61301		Peru	Piura Department, Ouedesha Cahallón 3 km NE El Tocto	-5.7839	-89.7671	INGROUP	Tissue	SAMN12944405
U4279	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 63070		Peru	Puno Department, 14 km NW Camino 19 Doores 16.555 70 Doores 28.27W	-15.275	-70.4717	INGROUP	Tissue	SAMN12944406
U4305	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 70072		United States of America	Florida, Gainesville UF Campus	29.6415	-82.3858	INGROUP	Tissue	SAMN12944407
U4292	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 70280		Bolivia	Bent Department, 38K W. FRINIDAD 175M	-14.628	-64.9000	INGROUP	Tissue	SAMN12944408
U4297	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 4719		Bolivia	Santa Cruz Department, Soriano de Huanchabamba 21km SE, Cantata Aro Hvi	-14.5	-60.62	INGROUP	Tissue	SAMN12944409
U4285	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 2284		Bolivia	La Paz Department, P. Saverda 83 km by road E, Charazani Cerro Asunta Pira	-14.407	-68.3961	INGROUP	Tissue	SAMN12944410
U4259	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 23164		Russia	Belarus: Dzerzhinsk Raion Forest Sta	53.45	27.2	INGROUP	Tissue	SAMN12944411
U4281	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 4008		Bolivia	Maudslayi v. Amaruta Chilea, Benavick Peninsula near mouth of Rio Santa Maria de 2 km S	-13.8775	-67.4167	INGROUP	Tissue	SAMN12944412
U4287	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 7073		Peru	Huancayo Department, UNCHOG PASS NNW ACCOMAY 3450M	-13.924	-71.6932	INGROUP	Tissue	SAMN12944413
U4115	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 2823		Peru	Provincia de Tarma, Tarma	-10.5333	-76.3833	INGROUP	Tissue	SAMN12944414
U4279	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 92399	JAG 1919	Argentina	Provincia de Tucumán, Tal del Valle, 20 km S, 6 km E	-26.8517	-65.7092	INGROUP	Tissue	SAMN12944415
U4114	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 47149		Peru	San Martín Department, Ca 22 km ENE Florida	-5.72706	-77.5028	INGROUP	Tissue	SAMN12944416
U4182	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 23111		Peru	Santa Cruz Department, Prov. Florida 21.2 km E Samitara	-18.1775	-64.9474	INGROUP	Tissue	SAMN12944417
U4185	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 9564	GAV 1777	South Africa	Haitian Department, Kimbelen, 95 km S, 45 km W	-29.633	-24.1	INGROUP	Tissue	SAMN12944418
U4318	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 1851		Argentina	Provincia de Tucumán, La Florida 800 Pta. Amancio Hwy 11 road km from Olmos	-23.828	-65.11	INGROUP	Tissue	SAMN12944419
U4317	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 6432		Peru	Tumbes Department, Campo verde 13 doores	-3.84567	-80.1764	INGROUP	Tissue	SAMN12944420
U4299	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 4476		Peru	Cajamarca Department, La Junta junction no. Tabacona and Chaudine	-5.8033	-78.7716	INGROUP	Tissue	SAMN12944421
U4309	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 27265		Costa Rica	Montes de Orosi, 1 km N Via MRG, Cerro Alegre	9.8031	-82.0417	INGROUP	Tissue	SAMN12944422
U4321	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 4849		Guatemala	Silvone Koonam Mountain 6 km W Koonam	4.9333	-89.9	INGROUP	Tissue	SAMN12944423
U4192	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	UWBM 70177	DAB 858	Argentina	Provincia de Tucumán, Corrientes, Manuel Drouot	-19.5149	-65.7092	INGROUP	Tissue	SAMN12944424
U4170	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 69222	DAB 1466	Nicaragua	Departamento de Managua, Managua, Chocoye en	11.99	-86.26	INGROUP	Tissue	SAMN12944425
U4188	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 101022	BFS 0809	Guatemala	Provincia de Chiriquí, Parque Nacional Volcan Barú	8.8449	-82.3918	INGROUP	Tissue	SAMN12944426
U4187	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 56139	DAB 1292	Nicaragua	Rosario Department, Prov. Florida 21.2 km E Samitara	-18.1775	-64.9474	INGROUP	Tissue	SAMN12944427
U4166	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 94174	DHB 2387	Honduras	Silvone Koonam Mountain 6 km W Koonam	4.9333	-89.9	INGROUP	Tissue	SAMN12944428
U4194	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 10177	SAK 044	Bolivia	Provincia de Corrientes, Corrientes, Manuel Drouot	-0.0486	-78.7525	INGROUP	Tissue	SAMN12944429
U4352	<i>Tardus albiventris</i> Lafresque, 1844	Martin Mdo collection	Not available		Gulf of Guinea	Departamento de Santa Cruz, Provincia Villafraude, Valdeverde, San Lorenzo	-10.077	-61.9157	INGROUP	Tissue	SAMN12944430
U4175	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 10460	DHB 479	Malawi	Príncipe	1.6498	7.391472	INGROUP	Tissue	SAMN12944431
U4315	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 1333	DHB 479	Malawi	North-Western Region, Mwanza, 20 km SW, Thambani Forest Reserve	-15.4508	34.64481	INGROUP	Tissue	SAMN12944432
U4275	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 21425		Peru	Cabo Rico Llanos Costa 63 km NNW mouth Arroyo Cruz	-10.899	-81.2287	INGROUP	Tissue	SAMN12944433
U4176	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 1858	JAG 1820	Guatemala	Panama Province, Cerro Campana Abo de Campana Parque Nacional W Cam	8.71372	-79.9528	INGROUP	Tissue	SAMN12944434
U4190	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 104079	DHB 4410	Guatemala	Departamento de Quezaltenango, Quezaltenango, Xela, El Baril	14.283	-91.89	INGROUP	Tissue	SAMN12944435
U4292	<i>Tardus albiventris</i> Lafresque, 1844	Bufo Museum	UWBM 104175		Guatemala	Cajamarca Department, Ca Cha NNW San Jose de Lourdes 5 doores 04.7 S 78 doores 57.9 W	-5.07167	-78.966	INGROUP	Tissue	SAMN12944436
TAM5	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 70901 (113720)	BR 34012	Brazil	Rondonia, Japurá	-5.8064	-60.8317	INGROUP	Tissue	SAMN12944437
TAM2	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	LEGEIA 428	P230	Brazil	Park. Parque Zoológico de Curitiba	-6.06139	-50.06	INGROUP	Tissue	SAMN12944438
TAL2	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 77372 (23059)	SEC 310	Brazil	Amarantosa, São Gabriel de Cachoeira PFB93	-5.465	-63.072	INGROUP	Tissue	SAMN12944439
U098	<i>Tardus albiventris</i> Lafresque, 1844	Loisiana State University Museum of Natural Science	LSMZJ 2710		Cameroon	Oued Province, Airport at Ta andi, 20 km W Foumban	-5.86038	-10.7758	INGROUP	Tissue	SAMN12944440
TAL15	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 65655 (F2260)	AMA NA 126	Brazil	Park. Itaipua FNP, Amami Mambembe	-2.53023	-57.5749	INGROUP	Tissue	SAMN12944441
TN2	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 69756 (F10951)	M 48 099	Brazil	Rondonia, Manaus	-8.01177	-57.8497	INGROUP	Tissue	SAMN12944442
TN1	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 61073 (F4289)	PEM 065	Brazil	Park. Monte Alegre Parque Estadual Monte Alegre Arouche	-2.02447	-54.20999	INGROUP	Tissue	SAMN12944443
TRK15	<i>Tardus albiventris</i> Lafresque, 1844	Divisão de Avés do Museu de Zoologia da Universidade Estadual de Feira de Santana	DAMUFZ 1524	00962	Brazil	Bahia, Morro do Chané	-11.8225	-41.12	INGROUP	Tissue	SAMN12944444
TRF10	<i>Tardus albiventris</i> Lafresque, 1844	Divisão de Avés do Museu de Zoologia da Universidade Estadual de Feira de Santana	DAMUFZ 1524	00962	Brazil	Bahia, Andaraí Marinho	-12.7699	-41.1567	INGROUP	Tissue	SAMN12944445
TRF11	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 71015 (LZLF 091)	CAS108	Brazil	Park. Castelo do Papa Ferreira Bonito	-5.80208	-41.5358	INGROUP	Tissue	SAMN12944446
TRK2	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 7587 (F16152)	UFAC 1286	Brazil	Paraná, Marabá	-8.01177	-57.8497	INGROUP	Tissue	SAMN12944447
TRK3	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 6354 (F16043)	AMZ 537	Brazil	Açor, Rio Branco Estrada do Duvaldi, Fazenda São Raimundo	-9.95987	-67.7326	INGROUP	Tissue	SAMN12944448
TRF12	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	UWBM 101193	BTS 244	Mexico	Mato Grosso, Nova Brasilândia mineira distrito Rio Jurema Fazenda Getório	-10.8282	-58.2367	INGROUP	Tissue	SAMN12944449
TRF13	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 61363 (F2981)	UFAC 803	Brazil	Chiriquí, Arriac, small foss of ovide on forest	16.2083	-91.9352	INGROUP	Tissue	SAMN12944450
TRF14	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 7587 (F16152)	UFAC 1286	Brazil	Açor, Rio Branco Transacção AC990 km 70 Ramal Jurema km 11	-9.96088	-68.4258	INGROUP	Tissue	SAMN12944451
TRF15	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 66844 (F19449)	ORX 034	Brazil	Lozoya Department, Iba Paro, Rio Amancio opposite Av. avsa on 80 km BE Iquitos	-5.7061	-71.2807	INGROUP	Tissue	SAMN12944452
TRF16	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 61065 (F4292)	PEM 069	Brazil	Park. Ontemina Lago Saracua Comandante Caniba	1.76389	-56.2268	INGROUP	Tissue	SAMN12944453
TRF17	<i>Tardus albiventris</i> Lafresque, 1844	Museo Parnaseo Emilio Goeldi	MPEJ 61065 (F42								

Table S2 List of additional loci used, including code or locus name, description, location in the chromosome and source when information is available.

Number	code/locus	description	Chromosome	Source
1	24813			1 Backström et al., 2008
2	26698	Histone-binding protein RBBP7 (retinoblastoma-binding protein 7) (RBBP-7) (retinoblastoma-binding protein p46). (Source:UniProt/SwissProt)		1 Backström et al., 2008
3	27331			1 Backström et al., 2008
4	23989	tumour differentially expressed 2 (Source:RefSeq_peptide; Acc:NP_001026245)		3 Backström et al., 2008
5	17483	High mobility group protein B2 (high mobility group protein 2) (HMG- 2) (Source:UniProt/SwissProt; Acc:P26584)		4 Backström et al., 2008
6	25149	Carboxypeptidase Z precursor (EC 3.4.17.-) (CPZ) (cCPZ) (Source:UniProt/SwissProt; Acc:Q8QGP3)		4 Backström et al., 2008
7	15724	acyl-coenzyme A dehydrogenase, short/branched chain (Source:RefSeq_peptide; Acc:NP_001026411)		6 Backström et al., 2008
8	18142	proteasome (prosome, macropain) 26S subunit, non-ATPase, 14 (Source:RefSeq_peptide; Acc:NP_001026427)		7 Backström et al., 2008
9	6419	adenosine deaminase-like (Source:RefSeq_peptide; Acc:NP_001025718)		10 Backström et al., 2008
10	3862	WD repeat protein 24 (Source:UniProt/SwissProt; Acc:Q5ZMV9)		14 Backström et al., 2008
11	1658	Shwachman–Bodian–Diamond syndrome (Source:RefSeq_peptide; Acc:NP_001006211)		19 Backström et al., 2008
12	8235	Hypothetical protein (Source:UniProt/SPTREMBL; Acc:Q5ZKN8)	unplaced Scaffold	Backström et al., 2008
13	4550	not in Ensembl database	-	Backström et al., 2008
14	VLDLR7	very low density lipoprotein receptor	Z	Backström et al., 2008
15	BRM12	SMARCA2 SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily a, member 2	Z	Backström et al., 2008
16	13403	proteasome (prosome, macropain) 26S subunit, ATPase, 2 (Source:RefSeq_peptide; Acc:NP_001006225)		1 Backström et al., 2008
17	21652	L-lactate dehydrogenase B chain (EC 1.1.1.27) (LDH-B). (Source:UniProt/SwissProt; Acc:P00337)		1 Backström et al., 2008
18	10005			2 Backström et al., 2008
19	25969			2 Backström et al., 2008
20	13093			3 Backström et al., 2008
21	9949			4 Backström et al., 2008
22	20207	Hypothetical protein (Source:UniProt/SPTREMBL; Acc:Q5ZJ03)		5 Backström et al., 2008
23	3978	RasGEF domain family, member 1 A (Source:RefSeq_peptide; Acc:NP_001026390)		6 Backström et al., 2008
24	4557	acyl-coenzyme A dehydrogenase, long chain (Source:RefSeq_peptide; Acc:NP_001006511)		7 Backström et al., 2008
25	3399	Pleckstrin homology domain-containing family B member 2 (Evectin-2) (Source: UniProt/SwissProt; Acc:Q5F352)		9 Backström et al., 2008
26	13907			9 Backström et al., 2008
27	16934	Selenoprotein T precursor (Source:UniProt/SwissProt; Acc:Q5ZJN8)		9 Backström et al., 2008
28	13336	eukaryotic translation initiation factor 3, subunit 1 alpha, 35 kDa (Source:RefSeq_peptide; Acc:NP_001012789)		10 Backström et al., 2008
29	1304	Casein kinase II subunit alpha' (EC 2.7.11.1) (CK II) (Source:UniProt/SwissProt; Acc:P21869)		11 Backström et al., 2008
30	12260	dihydropyrimidinase-like 3 (Source:RefSeq_peptide; Acc:NP_989824)		13 Backström et al., 2008
31	6938			14 Backström et al., 2008
32	2108			15 Backström et al., 2008
33	12619	Pescadillo homologue 1 (Source:UniProt/SwissProt; Acc:O00541)		15 Backström et al., 2008
34	9663			19 Backström et al., 2008
35	7793	Phosphopantothenate–cysteine ligase (EC 6.3.2.5) (phosphopantothenoylcysteine synthetase) (PPC synthetase) (Source:UniProt/SwissProt)		21 Backström et al., 2008
36	504	hypothetical protein LOC418842 (Source:RefSeq_peptide; Acc:NP_001006270)		23 Backström et al., 2008
37	2496			27 Backström et al., 2008
38	5915			28 Backström et al., 2008
39	14718	Superkiller viralicidic activity 2-like 2 (EC 3.6.1.-) (ATP-dependent helicase SKIV2L2) (Source:UniProt/SwissProt; Acc:P4228)		Z Backström et al., 2008
40	12884	dihydroipoamide dehydrogenase (Source:RefSeq_peptide; Acc:NP_001025898)		1 Backström et al., 2008
41	15506	Contactin-1 precursor (neural cell recognition molecule F11). (Source:UniProt/SwissProt; Acc:P14781)		1 Backström et al., 2008
42	10623	vasoactive intestinal peptide receptor 2 (Source:RefSeq_peptide; Acc:NP_001014970)		2 Backström et al., 2008
43	22187	Myelin basic protein (MBP) (Source:UniProt/SwissProt; Acc:P15720)		2 Backström et al., 2008
44	26472	Collagen alpha-1(XIV) chain precursor (undulin) (Source:UniProt/SwissProt; Acc:P32018)		2 Backström et al., 2008
45	25442	ubiquitin-conjugating enzyme E2, J1 (UBC6 homologue, yeast) (Source:RefSeq_peptide; Acc:NP_990094)		3 Backström et al., 2008
46	17367	26S protease regulatory subunit 4 (P26s4) (proteasome 26S subunit ATPase 1) (Source:UniProt/SwissProt; Acc:Q90732)		5 Backström et al., 2008
47	7248			8 Backström et al., 2008
48	17388	mago-nashi homologue, proliferation-associated (Source:RefSeq_peptide; Acc:NP_989656)		8 Backström et al., 2008
49	MC1R	melanocortin 1 receptor		11 Backström et al., 2008
50	ACA			Harvey, 2015
51	ACO1	aconitase 1, soluble	Z	Harvey, 2015
52	ALD	aldolase	no info	Harvey, 2015
53	ARNTL	aryl hydrocarbon receptor nuclear translocator like	no info	Harvey, 2015
54	BDNF	brain derived neurotrophic factor	5	Harvey, 2015
55	CALB1	calbindin 1	2	Harvey, 2015
56	CHC (CLTC)	clathrin heavy chain	19	Harvey, 2015
57	CLK	clock	no info	Harvey, 2015
58	CLOCK	clock circadian regulator.	4	Harvey, 2015
58	CLTCL1	clathrin heavy chain like 1.	22	Harvey, 2015
59	CMYC	v-myc avian myelocytomatosis viral oncogene homolog	2	Harvey, 2015
60	CSDE1	cold shock domain containing E1	26	Harvey, 2015
61	DCOH (PCBD1)	pterin-4 alpha-carbinolamine dehydratase 1	6	Harvey, 2015
62	CSNK1E	casein kinase 1 epsilon	1	Harvey, 2015
63	DDX5	DEAD-box helicase 5	Unplaced Scaffold	Harvey, 2015
64	EEF	eukaryotic translation elongation factor		Harvey, 2015
65	EGR1	early growth response 1	13	Harvey, 2015
66	EIF5	eukaryotic translation initiation factor 5	5	Harvey, 2015
67	ENO1	enolase 1, (alpha)	21	Harvey, 2015
68	GRIA2	glutamate ionotropic receptor AMPA type subunit 2	4	Harvey, 2015
69	HMG (HMGN4)	high mobility group nucleosomal binding domain 4	23	Harvey, 2015
70	HNRNPA2B1	heterogeneous nuclear ribonucleoprotein A2/B1	2	Harvey, 2015
71	HOXA	homeobox A cluster		Harvey, 2015
72	IRF1	interferon regulatory factor 1	13	Harvey, 2015
73	KCNQ5	potassium voltage-gated channel subfamily Q member 5	3	Harvey, 2015
74	MUSK	muscle, skeletal, receptor tyrosine kinase	Z	Harvey, 2015
75	NAA60 (NAT15)	N(alpha)-acetyltransferase 60, NatF catalytic subunit	14	Harvey, 2015
76	NGF	nerve growth factor	26	Harvey, 2015
77	NTF3	neurotrophin 3	1	Harvey, 2015
78	ODC1	ornithine decarboxylase 1	3	Harvey, 2015
79	PARK7	Parkinson disease (autosomal recessive, early onset) 7	21	Harvey, 2015
80	PDE6B	phosphodiesterase 6B	Z	Harvey, 2015
81	PER2	period circadian clock 2	9	Harvey, 2015
82	PPP2CB	protein phosphatase 2 catalytic subunit beta	4	Harvey, 2015
83	PSMA2	proteasome subunit alpha 2	2	Harvey, 2015
83	PSMA	proteasome		Harvey, 2015
84	RAG1	recombination activating 1	5	Harvey, 2015
85	RAG2	recombination activating 2	5	Harvey, 2015
86	RAPGEF1	Rap guanine nucleotide exchange factor 1	17	Harvey, 2015
87	RHOD	ras homolog family member D	Unplaced Scaffold	Harvey, 2015

88	SBDS	SBDS ribosome assembly guanine nucleotide exchange factor	19 Harvey, 2015
89	SEPT2	septin 2	9 Harvey, 2015
90	SOMA		Harvey, 2015
91	SUCLG1	succinate-CoA ligase alpha subunit	4 Harvey, 2015
92	TXNDC12	thioredoxin domain containing 12	8 Harvey, 2015
93	VDAC2	voltage dependent anion channel 2	6 Harvey, 2015
94	VIM	vimentin	2 Harvey, 2015
95	ALDOB	aldolase, fructose-bisphosphate B	Z Harvey, 2015

Table S3. Quality control after trimming using the PHYLUCE pipeline.

samples	reads	total base pairs	mean length	95 CI length	minimum	maximum	median
U2338	1012978	249219091	246,0261635	0,065984449	40	301	272
U3899	723404	176746449	244,3260599	0,079944499	40	301	271
U4155	1287700	311751988	242,0998587	0,059042297	40	301	266
U4159	899776	224353255	249,3434533	0,069347335	40	301	277
U4162	949773	236893800	249,4214933	0,066665403	40	301	275
U4163	1324780	319777258	241,3814052	0,057819107	40	301	263
U4164	757123	183335173	242,1471452	0,076157202	40	301	264
U4166	1179423	292508760	248,0100524	0,06174044	40	301	276
U4168	1366917	343577070	251,3518158	0,055260561	40	301	279
U4170	1573272	385549507	245,062206	0,052688229	40	301	271
U4172	1123582	271886712	241,9820823	0,062141555	40	301	264
U4176	1225960	297311565	242,5132672	0,060936417	40	301	267
U4177	1410551	356772030	252,9309681	0,053420887	40	301	280
U4179	2036667	512876677	251,8215678	0,045834905	40	301	281
U4185	1004514	249679575	248,5575861	0,067771511	40	301	278
U4187	129766	31861427	245,5298537	0,189743909	40	301	273
U4188	966152	237380702	245,697056	0,067969219	40	301	272
U4189	627462	157289243	250,6753285	0,082290656	40	301	278
U4192	789627	193558595	245,1266167	0,078486925	40	301	275
U4273	1767776	447079437	252,905027	0,047753256	40	301	280
U4279	876435	214418802	244,6488353	0,070362741	40	301	268
U4281	1080731	270373140	250,1761678	0,061467404	40	301	276
U4285	1202545	299028546	248,663082	0,058614511	40	301	275
U4287	1954004	491557894	251,5644257	0,04734589	40	301	282
U4292	955476	234938942	245,8868062	0,067068208	40	301	270
U4297	1068930	266340548	249,1655656	0,064316475	40	301	277
U4299	1738881	437056955	251,3437981	0,04609414	40	301	274
U4305	656016	155954409	237,7295813	0,075634587	40	301	250
U4309	995238	249848514	251,0439855	0,066259004	40	301	281
U4312	740502	185157722	250,0435137	0,077636681	40	301	280
U4314	1067878	269002783	251,9040405	0,062940745	40	301	281
U4315	852825	207899878	243,7778888	0,075507931	40	301	272
U4317	1519039	384864087	253,3602409	0,051862643	40	301	282
U4319	1253453	312132253	249,0179153	0,057395171	40	301	275
U4320	658636	156238357	237,2150277	0,083116438	40	301	257
U4321	1068882	266691344	249,5049444	0,064832551	40	301	279
U4322	1555790	381173729	245,0033289	0,055951634	40	301	274
U4325	1014928	243430082	239,8496071	0,0665418	40	301	261
U4329	1002038	246944943	246,4426928	0,066288071	40	301	272
U4331	1056110	257277409	243,6085341	0,064296491	40	301	267
U4332	749929	177747541	237,0191591	0,079481137	40	301	258
U4334	738961	183623528	248,4887944	0,076282947	40	301	275
U4336	1132500	279425814	246,7336106	0,061010828	40	301	271
U4337	661535	151643152	229,2292199	0,07972637	40	301	241
U4338	1128316	284932377	252,5288811	0,061412377	40	301	283
U4339	1116113	276157009	247,4274639	0,06213276	40	301	273
U4340	1108044	277731444	250,6501944	0,062876366	40	301	280
U4345	1089228	261793754	240,3479841	0,063607622	40	301	262
Bali	972331	240064047	246,8953957	0,075113069	40	301	288
BTC580	1015780	251742031	247,8312538	0,072659513	40	301	289

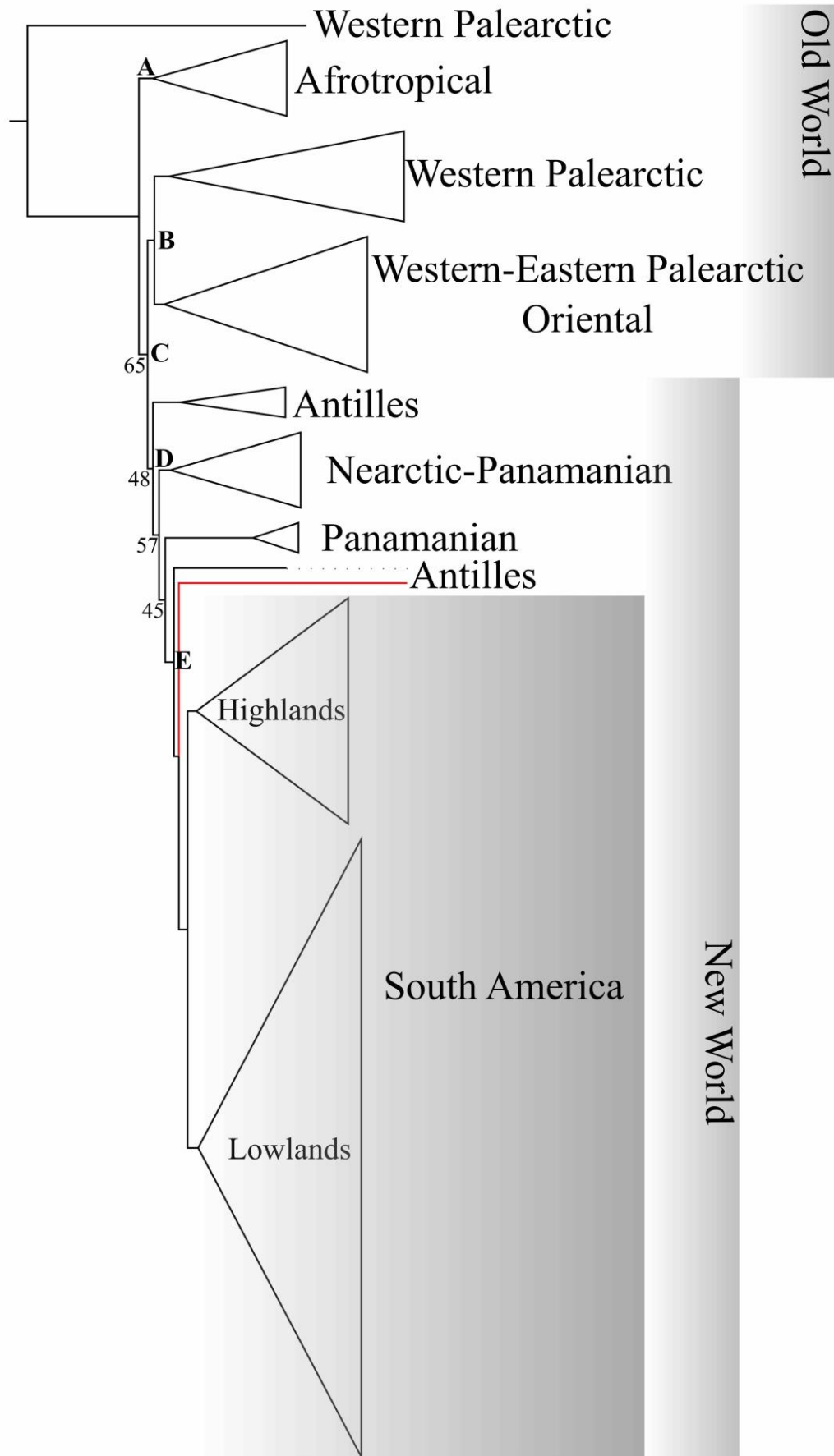
U3572 (DOT)	844655	205482219	243,2735484	0,079833087	40	301	278
GFB998	783593	194040288	247,6289196	0,084171305	40	301	291
TAL13	660694	160566677	243,0272971	0,091345092	40	301	279
TAL24	662921	159339921	240,3603461	0,094269565	40	301	278
TAM2	1333741	327995771	245,9216377	0,062408152	40	301	282
TAM6	554234	132696165	239,4226356	0,103640966	40	301	276
TFL2	999161	245756812	245,9631751	0,073415753	40	301	283
TFL5	1291805	317845459	246,0475528	0,064492984	40	301	285
TFU1	1127747	271945895	241,140872	0,071772996	40	301	279
TFU13	837988	206955122	246,9666893	0,080893935	40	301	288
THX1	771404	186454885	241,7084757	0,085309501	40	301	278
THX5	1039185	253021311	243,4805266	0,073142114	40	301	281
TIG13	1429444	353707763	247,4442951	0,061253086	40	301	289
TIG3	1187264	293311726	247,0484458	0,066813717	40	301	286
TLAW3	1216394	300832662	247,3151479	0,066392139	40	301	288
TLAW4	1176451	292581023	248,6980104	0,06701537	40	301	290
TLEU5	953831	232058063	243,2905441	0,073135996	40	301	275
TNU1	1171769	286225526	244,267877	0,068899959	40	301	282
TNU2	1300434	319696378	245,8382186	0,064571004	40	301	284
TRUF1	1277649	314908687	246,4751172	0,065082313	40	301	286
TRUF10	898326	220267968	245,1982554	0,078477401	40	301	285
TSCZ1	972815	239772819	246,4731927	0,075236152	40	301	287
TSCZ3	1290572	318869578	247,0761631	0,064437429	40	301	288
U1097	1015709	248193955	244,3553764	0,074707565	40	301	284
U1098	1369418	340283669	248,4878021	0,061801585	40	301	289
U3456	899380	222090348	246,9371656	0,078348176	40	301	289
U4053	904782	225849404	249,6174813	0,074640473	40	301	289
U4054	1011573	250525131	247,6589737	0,072591434	40	301	287
U4148	1031444	255451986	247,6644258	0,072275341	40	301	289
U4156	993954	245212052	246,7036221	0,07212532	40	301	284
U4169	914091	223217448	244,1960899	0,07755667	40	301	283
U4175	977089	239697091	245,3175617	0,072925125	40	301	280
U4178	1455722	358442686	246,2301772	0,061471731	40	301	287
U4190	988804	244917158	247,6902986	0,073643052	40	301	289
U4194	1200273	295761379	246,4117572	0,066907662	40	301	286
U4275	1097999	268805266	244,8137621	0,070054603	40	301	282
U4280	663484	158895714	239,4868814	0,089020859	40	301	269
U4282	1214791	291814350	240,2177412	0,069624553	40	301	278
U4294	1208730	294713244	243,8205753	0,064532772	40	301	274
U4301	773100	179627165	232,346611	0,090192909	40	301	266
U4303	1159540	284384082	245,255948	0,068752336	40	301	284
U4308	684353	166663298	243,5341089	0,090811918	40	301	283
U4316	797714	192346148	241,1216902	0,085506322	40	301	279
U4335	1165730	281942185	241,8589082	0,068763556	40	301	277
U4352	864871	214873353	248,445552	0,078630804	40	301	290
GNM10084	1322336	135556747	102,5130882	0,035593745	40	251	95
GNM11291	1202387	125566144	104,4307232	0,036002436	40	251	98
GNM11676	14686	2416075	164,515525	0,564243961	40	251	161
GNM172	937721	107982291	115,1539648	0,041604034	40	251	111
GNM5292	720579	86542147	120,1008453	0,049512035	40	251	116
GNM5706	440600	56814946	128,9490377	0,067567384	40	251	125
GNM5775	871031	106943615	122,7781962	0,045441672	40	251	119
GNM5778	603851	74314547	123,0676889	0,056159831	40	251	119

GNM6136	1195342	144383360	120,7883267	0,038238317	40	251	117
GNM6245	1232489	158432632	128,5468933	0,039926145	40	251	126
GNM6270	1257352	142359579	113,2217382	0,035974002	40	251	110
GNM7449	1035648	113034325	109,1435748	0,040692702	40	251	103
GNM7455	1053628	127802809	121,297848	0,047917574	40	251	114
GNM780	913165	115588688	126,5802872	0,046651912	40	251	123
GNM8238	1878746	304161769	161,8961632	0,035690294	40	251	162
GNM8584	1312628	169481538	129,116199	0,038678763	40	251	126
GNM8762	1205711	257416626	213,4977835	0,041426022	40	251	228
U1774	1780967	374256176	210,1421172	0,035183842	40	251	224
U3599	1440180	177049843	122,9359129	0,03817315	40	251	118
U3649	2062348	267366087	129,6415964	0,032526451	40	251	126

Table S4. Quality control after assembly using the PHYLUCE pipeline.

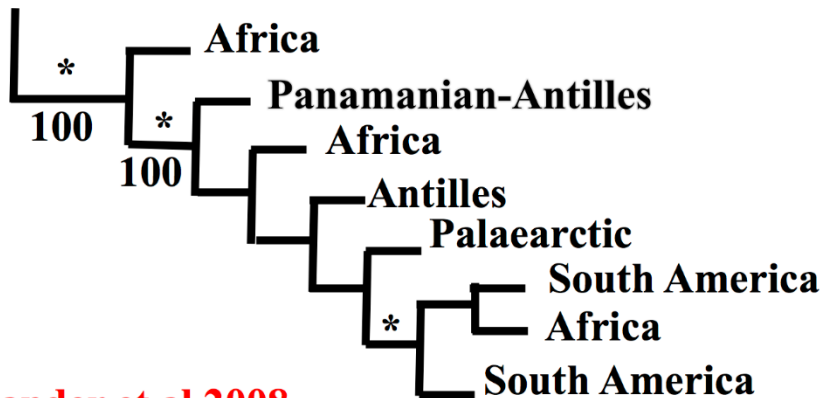
Sample	Reads	Base pairs	Average length	standard error length	Minimum le	Maximum le	Median leng	Contigs > 1kb	Quality Control	Source	Sequencer	Core Facility
GNN10084	3456	993221	287,3903356	3,627150127	224	10795	247	10	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN11291	4232	1218455	287,9146975	2,855316648	224	10795	250	8	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN11676	708	189219	267,2584746	15,08393687	224	10795	240	2	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN1172	5976	1863198	311,7801205	2,705279955	224	10771	265	50	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN5292	4633	1438899	310,5760846	3,167926192	224	10795	260	44	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN5706	5024	1459656	290,5366242	2,544985302	224	10795	250	16	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN5775	7899	2362345	299,0688695	1,914137655	224	10795	257	40	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN5778	4097	1238535	302,3029046	3,309825804	224	10795	254	23	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN6136	11113	3438196	309,3850445	1,969173433	224	10795	263	72	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN6245	16561	4938301	298,1885756	1,137128474	224	10795	260	58	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN6270	7435	2280375	306,7081372	2,075498566	224	10795	266	40	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN7449	4755	1378343	289,8723449	2,638871818	224	10795	252	9	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN7455	12066	3495999	289,7396818	1,416704926	224	10795	248	59	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN780	6558	2083571	317,7143946	3,041325311	224	10795	253	77	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN8238	72422	21270952	293,7084311	1,035346498	224	13561	249	401	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN8584	17006	4996536	293,8101846	0,967417666	224	10795	259	25	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
GNN8762	111014	31409732	282,9348731	0,452416207	224	10795	244	580	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
U1774	181582	52046772	286,6295778	0,34003765	224	10795	247	791	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
U3599	10319	3030288	293,6610137	1,609132539	224	10795	251	53	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
U3649	34803	10034161	288,3131052	0,635076505	224	10771	253	94	After Assembly_Trinity	ToePad	Miseq	SciLifeLab_Stockolm
U2338	24384	10037572	411,6458333	1,812531763	224	10795	300	1846	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U3899	19460	8556950	439,7199383	2,480551729	224	22411	306	1866	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4155	31974	12822211	401,0199224	1,52827837	224	10795	301	2177	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4159	20969	9413181	448,9093901	2,138687991	224	10795	315	2064	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4162	21186	9043093	426,8428679	1,832809718	224	10795	314	1412	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4163	38558	14985148	388,639141	1,292798834	224	10795	298	2143	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4164	19037	8250755	433,4062615	2,154075977	224	10795	308	1625	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4166	28268	12091318	427,7387152	1,677682695	224	10795	316	2116	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4168	30754	12790746	415,905118	1,520896727	224	10795	313	2024	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4170	35884	14195140	395,5841043	1,390655812	224	14874	303	2061	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4172	32346	12685670	392,1866691	1,375693841	224	10795	302	1738	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4176	30929	12645210	408,8463901	1,56323169	224	10795	305	2202	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4177	31012	12748900	411,0957049	1,450653098	224	10795	310	1954	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4179	37563	15384032	408,5738719	1,464648477	224	16715	309	2431	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4185	21076	9527634	452,0608275	2,264952649	224	10795	314	2214	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4187	5086	2684581	527,8373968	4,274614402	224	7448	304	321	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4188	25529	10572284	414,128403	1,741289087	224	10795	307	1850	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4189	15875	7287770	459,072126	2,430166276	224	10795	320	1572	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4192	16711	8095529	484,4431213	2,708794709	224	10795	320	2140	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4273	38783	15582737	401,7929763	1,247858497	224	10795	311	2180	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4279	23161	9691762	418,451794	1,903348859	224	10795	302	1832	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4281	31902	12596746	394,8575638	1,408450923	224	10795	307	1721	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4285	26770	10781203	402,7345163	1,565420908	224	10795	304	1536	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4287	34251	14545114	424,6624624	1,533151453	224	10795	312	2538	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4292	23385	9613101	411,0797947	1,724225214	224	10795	305	1555	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4297	26792	11336077	423,1142505	1,717226983	224	10795	313	2088	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4299	42021	14659312	348,8568097	0,941060265	224	10795	285	840	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4305	20347	7596095	373,3275176	1,558586568	224	10795	286	451	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4309	21303	9327226	437,8362672	2,165971512	224	10795	310	1987	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4312	18675	8440804	451,9841499	2,349710715	224	10795	315	1904	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4314	26471	11029044	416,6462922	1,673396199	224	10795	308	1910	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4315	19708	9114937	462,4993404	2,466906233	224	10795	313	2257	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4317	35523	13944510	392,5487712	1,322364258	224	10795	301	1989	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4319	30498	12319726	403,9519313	1,479933999	224	10795	305	1894	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4320	22411	8927665	398,3608496	1,753780622	224	10795	298	1293	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4321	23496	10197182	433,99651	1,921612904	224	10795	312	2076	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4322	27529	12474860	453,1534019	2,097418237	224	10795	312	2637	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4325	32769	12625182	385,2782203	1,410652051	224	10795	294	1789	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4329	29662	11729526	395,4394849	1,416402337	224	10795	306	1118	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4331	27751	11416246	411,3814277	1,712540609	224	10795	307	1834	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4332	21893	9002860	411,2209382	1,966595132	224	10795	297	1733	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4334	24285	9866532	406,2809141	1,801687626	224	15101	303	1513	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4336	30358	12064233	397,3988076	1,461011644	224	10795	302	1636	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4337	24210	8644012	357,0430401	1,166682931	224	10795	290	155	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4338	23074	10144471	439,6494323	2,11551047	224	18523	311	2159	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4339	29936	12282468	410,2908872	1,694325623	224	17622	308	1976	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4340	22547	10135478	449,5266776	2,175137154	224	10795	312	2295	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U4345	32872	12463774	379,1608055	1,336019021	224	10795	296	1573	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
Bali	19992	9282736	464,322529	2,260311208	224	10795	323	2076	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
BTC580	19110	9078567	475,0689168	2,374200929	224	10795	327	2129	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
U3572 (DOT6948)	27214	11411008	409,388844	1,502948995	224	10795	313	1410	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
GF8998	17380	8351015	480,4956847	2,483969514	224	10795	333	2033	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TAL13	19827	8901209	448,943814	2,151416413	224	10795	321	1818	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TAL24	20083	9207645	458,4795598	2,379901208	224	12359	320	2009	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TAM2	32825	13384564	407,7551866	1,394081637	224	10795	317	1694	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TAM6	22039	9563624	433,9409229	1,935354047	224	10795	318	1763	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TFL2	33305	13422179	403,0079267	1,352955472	224	10795	312	1818	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TFL5	31819	13411644	421,4979729	1,562881799	224	10795	314	2306	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TFLU13	23017	10037061	436,0716427	1,905331934	224	10795	317	1952	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TFU1	25193	11173237	443,5056166	1,927438462	224	10795	320	2216	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
THX1	21640	9456639	436,9981054	2,014737454	224	10795	315	1903	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
THX5	26699	11596004	434,3235327	1,84955501	224	10795	316	2136	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TIG13	35298	14495927	410,6727577	1,437326697	224	13947	313	2268	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TIG3	29587	12352666	417,5031602	1,562072715	224	10795	314	2076	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TLAW3	30561	12884291	421,5925853	1,582360234	224	10795	314	2193	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TLAW4	30823	12696877	411,9286572	1,427957044	224	10795	313	1831	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TLEU5	30646	11967755	390,5160543	1,230950101	224	10795	313	834	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TNU1	35064	14541909	414,7247604	1,412963629	224	13304	319	2166	After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Uppsala
TNU2												

U4178	27323	12450022	455,6608718	2,18621926	224	19226	321	2457 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4190	24774	10863554	438,5062566	1,773225392	224	10795	323	1939 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4194	31133	13059120	419,4623069	1,683706287	224	17392	315	2187 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4275	27442	11628045	423,7316887	1,624657615	224	10795	316	1932 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4280	26386	10356986	392,5182294	1,34246928	224	10795	312	656 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4282	23322	11076678	474,9454592	2,315201092	224	10795	322	2486 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4294	36879	14387855	390,1367987	1,193038453	224	10795	311	1549 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4301	20966	9620042	458,8401221	2,389133012	224	16741	319	2121 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4303	31083	13168664	423,661294	1,685870941	224	18529	315	2274 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4308	21270	9375819	440,800141	2,027721393	224	10795	319	1800 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4316	23503	10262549	436,6484704	1,959544481	224	10795	315	2025 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4335	26100	11071576	424,1983142	1,755386819	224	13862	315	1735 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala
U4352	17474	8319601	476,1131395	2,492831721	224	10795	326	2045 After Assembly_Trinity	FreshTissue	Miseq	SciLifeLab_Upssala



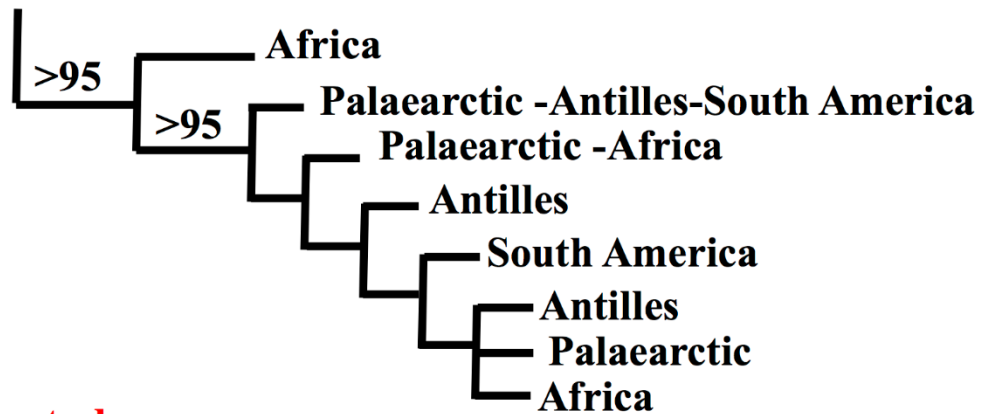
a) Voelker et al 2007

Turdus philomelos (Palearctic)



b) Nylander et al 2008

Turdus philomelos (Palearctic)



c) This study

Turdus philomelos (Palearctic)

