



A checklist of chromosome numbers and karyotypes of Amazonian freshwater fishes

Jorge Ivan Rebelo PORTO (1), Eliana FELDBERG (1),
Céleste Mutuko NAKAYAMA (1),
José das Neves FALCÃO (2)

RÉSUMÉ

CATALOGUE DES NOMBRES DE CHROMOSOMES ET CARYOTYPES DES POISSONS DU BASSIN AMAZONIEN

Une liste des nombres de chromosomes de 211 espèces de poissons du bassin amazonien est présentée. Ces espèces, appartenant à 5 ordres, sont caractérisées par une grande variabilité du nombre de chromosomes ($2n = 22$ à $2n = 134$), par la présence de chromosomes sexuels et de polymorphisme chromosomique chez certaines espèces, et enfin par une grande spécificité des bandes C et des sites des régions de l'organisateur du nucléole.

MOTS CLÉS : Chromosomes — Poissons — Eaux douces — Bassin amazonien.

ABSTRACT

A CHECKLIST OF CHROMOSOME NUMBERS AND KARYOTYPES OF AMAZONIAN FRESHWATER FISHES

A checklist of chromosome numbers of Amazonian freshwater fishes is presented. 211 nominal species belonging to 5 orders have had their haploid/diploid number listed. These species are characterized by a high karyotypic diversity including a wide chromosome number range ($2n=22$ to $2n=134$), different sex chromosomal mechanisms, chromosomal polymorphisms and nearly always a species-specific pattern of C-banding and nucleolar organizer regions.

KEY WORDS : Chromosomes — Fishes — Freshwaters — Amazon Basin.

RESUMEN

LISTA DE LOS NUMEROS CROMOSÓMICOS Y CARIOTIPOS DE PECES AMAZONICOS DE AGUA DULCE

Es presentada una lista con los datos cromosómicos de peces de la cuenca amazonica. 211 especies pertenecientes a 5 ordenes ya fueron estudiados citogeneticamente, determinandose por lo menos su numero cromosómico haploide/diploide. La variación del numero diploide va desde $2n=22$ hasta $2n=134$. Tambien fueron observado mecanisimos cromosómicos sexuales y polimorfismos cromosómicos en algunas especies, seguido de padrones casi siempre especie-specificos de la banda-C y de las regiones organizadoras del nucleolo.

PALABRAS CLAVES : Cromosomos — Peces — Agua dulce — Cuenca amazonica.

(1) Instituto Nacional da Pesquisas da Amazônia, Coordenação de Pesquisas em Biologia Aquática, Cx Postal 478, Manaus-AM, Brasil, CEP 69083.

(2) Universidade do Amazonas, Instituto de Biociências, Departamento de Biologia, Estrada do Contorno S/N, Manaus-AM, Brasil, CEP 69000.

INTRODUCTION

In the neotropics, chromosomal data have contributed to the studies of the biology, genetics and systematics of fishes. This is particularly true with respect to species characterization and diagnosis (BERTOLLO, 1978; ALMEIDA-TOLEDO, 1978), detection of inter and intraspecific polymorphism (GIULIANO-CAETANO and BERTOLLO, 1988; OLIVEIRA *et al.*, 1990a), data on sex chromosome systems (MOREIRA FILHO *et al.*, 1980; GALETTI Jr. *et al.*, 1981; BERTOLLO *et al.*, 1983; GALETTI Jr. and FORESTI, 1986; FELDBERG *et al.*, 1987; FALCÃO, 1988), supernumerary chromosomes (FALCÃO *et al.*, 1984; PAULS, 1985; VENERE and GALETTI Jr., 1985; OLIVEIRA *et al.*, 1988b; FORESTI *et al.*, 1989; ERDTMAN *et al.*, 1990) and natural triploidy (ALMEIDA-TOLEDO *et al.*, 1985; VENERE and GALETTI Jr., 1985; GIULIANO-CAETANO and BERTOLLO, 1990). All of the above mentioned findings are important for the understanding of fish or even animal chromosomal evolution.

Checklists containing chromosome data of fish have been published since 1971 (GYLDENHOLM and SCHEEL, 1971; CHIARELLI and CAPANNA, 1973; DENTON, 1973; PARK, 1974; OJIMA *et al.*, 1976; GOLD *et al.*, 1980; SOLA *et al.*, 1981; OLIVEIRA *et al.*, 1988a). SOLA *et al.* (1981) stated that the elaboration of fish chromosome lists is important because they allow for easy access to condensed information and identification of the cytogenetic characters important for the elaboration of evolutionary and phylogenetic models.

The Amazon basin is an excellent field for ichthyogenetic study (ALMEIDA-VAL *et al.*, 1991) and we believe that a checklist of Amazonian fish chromosome data will contribute to a better understanding of the diversity of fish species.

CYTOGENETIC FEATURES OF AMAZONIAN FISHES

In 1988, OLIVEIRA *et al.* listed the chromosome formulae of 433 neotropical fish species. In this study the origin of 55 % of the fish is unknown, some of them undoubtedly obtained from aquarium dealers.

In elaborating our list, we attempted to identify, among the species of unknown origin, the Amazonian freshwater species, and we updated the taxonomic status of many species. Thus, we have listed species belonging to the Amazon basin based on the compilation of OLIVEIRA *et al.* (*op. cit.*), and new data obtained by us (Table I). For the taxonomy and origin of the species with diploid/haploid numbers

already determined, several papers were consulted: GOLDSTEIN (1973), GÉRY (1977), NIJSSEN and ISBRUCKER (1980), KULLANDER (1983, 1986), ORTEGA and VARI (1986), GOULDING *et al.* (1988), BURGESS (1989). The classification of the higher taxa of the listed species is arranged according to GREENWOOD *et al.* (1966) and LAUDER and LIEM (1983) with the exception of the Serrasalminae which is considered at a family level, according to GÉRY (1977).

211 nominal species belonging to the Amazonian ichthyofauna have had their diploid/haploid number determined. However, few of these species have had their karyotypes described. These listed species correspond to 49 % of the neotropical fish species listed by OLIVEIRA *et al.* (*op. cit.*). We also detected that a vast amount of cytogenetic data is available in abstracts of Brazilian scientific meetings and others not readily available sources.

Diploid numbers

Diploid numbers ($2n$) of Amazonian fishes range widely: $2n = 22$ (*Nannostomus unifasciatus*) to $2n = 134$ (*Corydoras aeneus*). Also, species differ in having single or multiple nucleolar organizer regions (NORs) as well as different sex chromosomal mechanisms, including multiple systems. This karyotype diversity is apparently correlated with the rich specific diversity of Amazonian ichthyofauna.

Karyologically, the characiforms are the most studied fish group of the Amazon, followed by siluriforms (including the gymnotoids - electric fishes), perciforms (cichlids) and then on a minor scale, osteoglossiforms (bonytongue fishes) and lepidosireniforms (lungfishes).

In the characiforms, the diploid numbers vary from $2n = 22$ to $2n = 102$. However, in the families Anostomidae, Curimatidae, Prochilodidae, Hemiodidae and Chilodidae the chromosome numbers and karyotype morphology are very similar, usually with $2n = 54$, and meta-submetacentric (M-SM) chromosomes in their karyotypes and single NORs. On the other hand, there are karyotypically divergent families such as the Erythrinidae, Lebiasinidae, Characidae and Serrasalminae each of which shows diversification in their diploid numbers, karyotype morphologies and multiple NORs. Erythrinidae, Lebiasinidae, Characidae plus Ctenolucidae are characterized by diploid numbers smaller than $2n = 54$ (considered primitive in characiforms). Serrasalminae, on the other hand, contains species with diploid numbers equal to or greater than $2n = 54$. This cytogenetic feature of serrasalmids ($2n = 54$) has lead AREFJEV (1989) and PORTO *et al.* (1989, 1991) to consider them a distinct group at a family level, and not a

subfamily of Characidae, as postulated by some authors.

In the siluriforms, different from characiforms, there are few cytogenetic studies, although this group also has a large number of species in the Amazon basin. Diploid numbers of siluriforms vary from $2n=24$ to $2n=134$. The available data indicate that they are characterized by high karyotypic diversity, especially the families Callichthyidae (Siluroidei) and Sternopygidae (Gymnotoidei). It should be pointed out that the karyotypic diversity detected in Callichthyidae is related to gene duplication and/or polyploid events (OLIVEIRA *et al.*, *op. cit.*).

In the perciforms, especially the cichlids, the diploid numbers vary from $2n=38$ ($n=19$) to $2n=60$. Almost all species present $2n=48$, dominated by subtelo-acrocentric (ST-A) chromosomes, although there are some different diploid numbers and karyotype morphologies in this group, for example $2n=60$ predominating M-SM chromosomes.

Finally, in the osteoglossiforms, three species of osteoglossids were karyotyped and their diploid numbers vary from $2n=54$ to $2n=56$. In lepidosireniformes, a single species was karyotyped and a small diploid number ($2n=38$) was detected, with enormous chromosomes and an extremely high DNA value.

Nucleolar organizer region (NOR)

The NOR chromosome data from Amazonian freshwater fish families presented in Table I makes it possible to detect three groups in terms of patterns of specific localization of NORs sites: 1) single NORs, 2) multiple NORs, and 3) both single and multiple NORs. In the first group, the following families can be listed: Osteoglossidae, Anostomidae, Curimatidae, Prochilodidae, Hemiodidae, Pimelodidae, Aptereronotidae, Sternopygidae and Cichlidae; in the second group: Serrasalminidae and Lebiasinidae; and in the third group: Erhythrinidae, Characidae and Callichthyidae.

C-banding

Most of the species analyzed thus far present C-bands at, or around, the centromeres (centro/pericentromeric band). Less frequently they occur at the chromosome tips (telomeric band). Positive C-bands can also be found associated with NOR regions (in almost all of the serrasalminid species) or along chromosome arms (interstitial band) or as entirely heterochromatic short or long chromosome arms (particularly the sex chromosomes). All types of C-

bands can occur in the same species. Thus, when different species are compared, almost always a species-specific pattern of constitutive heterochromatin distribution is apparent.

Sex chromosomes

Based on cytological heteromorphy, heterologous sex chromosomes (ZZ/ZW and XX/XY₁Y₂ types) have been described for 6 Amazonian species, corresponding to 2.8% of the total number of species listed. The occurrence of the same kind of sex chromosome mechanisms (ZZ/ZW) in different genera (*Triporthus*, *Semaprochilodus* and *Eigenmannia*) shows that this kind of heterogamety has evolved several times among the Amazonian Ostariophysi fishes. FALCÃO (1988) suggests that in *Triporthus*, the W chromosome differentiated through a heterochromatinization process, followed by deletions.

Hibridization and Triploidy

Artificial crossing has shown that serrasalminid species, *Mylossoma duriventris* X *Colossoma macropomum* and *Piaractus brachypomus* X *Colossoma macropomum*, have a capacity to hybridize even when the taxa belongs to distinct genera (KOSSOWSKI *et al.*, 1983 and NAKAYAMA *et al.*, *pers. comm.*, respectively).

In the Amazon basin two cases of natural triploidy have been reported in fish: *Eigenmannia* sp. and *Hoplerythinus unitaeniatus*. In both cases diploid and triploid forms were found. The fertilization of a non-reduced ovule by a haploid spermatozoon was considered to be the most probable origin of the triploids observed (ALMEIDA-TOLEDO *et al.*, 1985; GIULIANO-CAETANO and BERTOLLO, 1990).

CONCLUSION

There still are too few cytogenetic data on Amazonian fishes. The lack of published karyograms or metaphase photographs as well as the problem of taxonomic determination and origin of analyzed material constitute the main problems in the cytogenetic study of Amazonian fishes, especially in the characiforms whose cytogenetic data forms a substantial part of the checklist. Thus, these data must be considered with care.

Considering the existence of approximately 3000 species in the Amazon Basin and the high karyotypic diversity detected so far, cytogenetic studies of

Amazonian fishes should be carried out preferably on monophyletic groups of fishes, taking into account their biogeography and the utilization of high resolution chromosome banding. In this way, it will be possible to determine the karyotypic characters that better elucidate the taxonomic and evolutionary problems.

ACKNOWLEDGEMENTS

We thank Dr. James PATTON for his comments on the manuscript and Dr. Michel JÉGU for his French translation. This research was supported by CNPq-Pig, Inpa/SCT, Eletro-norte and Orstom.

Manuscrit accepté par le Comité de rédaction le 15 février 1993

REFERENCES

- ALMEIDA-TOLEDO (L. F.), 1978. — *Contribuição à citogenética dos Gymnolidae (Pisces, Ostariophysi)*. Tese de Doutorado. Instituto de Biociências, Universidade de São Paulo, 128 p.
- ALMEIDA-TOLEDO (L. F.), FORESTI (F.) and TOLEDO-FILHO (S.A.), 1981. — Constitutive heterochromatin and nucleolus organizer region in the knifefish, *Apteronotus albifrons* (Pisces, Apterontidae). *Experientia*, 37 : 953-954.
- ALMEIDA-TOLEDO (L. F.), FORESTI (F.) and TOLEDO-FILHO (S.A.), 1985. — Spontaneous triploidy and NOR activity in *Eigenmannia* sp (Pisces, Sternopygidae) from the Amazon basin. *Genetica*, 66 : 85-88.
- ALMEIDA-TOLEDO (L. F.), FORESTI (F.), TOLEDO-FILHO (S.A.), BERNARDINO (G.), FERRARI (W.) and ALCANTARA (R. C. G.), 1987. — Cytogenetic studies on *Colossoma milrei*, *Colossoma macropomum* and their interspecific hybrid. *Proc. World Symp. on Selection, Hybridization, and Genetic Engineering in Aquaculture* : 189-195.
- ALMEIDA-VAL (V. M. F.), VAL (A. L.), FELDBERG (E.), CARACIOLO (M. C. M.) and PORTO (J. I. R.), 1991. — Evolução de peixes da Amazônia : aspectos genéticos e adaptativos. In : *Bases científicas para estratégias de preservação e desenvolvimento da Amazônia : fatos e perspectivas*. A. L. Val, R. Figliuolo et E. Feldberg (eds.). Inpa. Manaus : 281-292.
- AREFJEV (V. A.), 1989. — Chromosome sets of four characid fish species (Teleostei, Characidae). *Zool. Zh.*, 68 (5) : 82-91.
- AREFJEV (V. A.), 1990 a — Karyotypic diversity of characid families (Pisces, Characidae). *Caryologia*, 43 (3-4) : 291-304.
- AREFJEV (V. A.), 1990 b — Problems of karyotypic variability in the family Characidae (Pisces, Characiformes) with the description of somatic karyotypes for six species of tetras. *Caryologia*, 43 (3-4) : 305-319.
- BERTOLLO (L. A. C.), 1978. — *Estudos citogenéticos no gênero Hoplias Gill, 1903 (Pisces, Erythrinidae)*. Tese de Doutorado, Departamento de Genética e Matemática Aplicada à Biologia, Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo. 163 p.
- BERTOLLO (L. A. C.), 1988. — As regiões organizadoras de nucléolo na família Erythrinidae. *Res. II Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 20.
- BERTOLLO (L. A. C.) and MOREIRA FILHO (O.), 1983. — Possível ocorrência de uma nova espécie no gênero *Hoplias* (Pisces, Erythrinidae) : evidências morfológicas e citogenéticas. *Ciência e Cultura (Supl.)*, 36 : 786.
- BERTOLLO (L. A. C.), TAKAHASHI (C. S.), ALMEIDA-TOLEDO (L. F.), GALETTI Jr. (P. M.), FERRARI (I.), MOREIRA FILHO (O.) and FORESTI (F.), 1980. — Estudos citogenéticos em peixes da região amazônica. I. Ordem Cypriniformes. *Ciência e Cultura (Supl.)*, 32 : 735.
- BERTOLLO (L. A. C.), TAKAHASHI (C. S.) and MOREIRA FILHO (O.), 1983. — Multiple sex chromosome in the genus *Hoplias* (Pisces, Erythrinidae). *Cytologia*, 48 : 1-12.
- BURGESS (W. E.), 1989. — *An atlas of freshwater and marine catfishes. A preliminary survey of the Siluriformes*. T.F.H. Publications, Inc. : 326-367.
- CESTARI (M. M.), FERREIRA (R.) and GALETTI Jr. (P.M.), 1990. — Complemento cariotípico de duas espécies de peixes ornamentais : *Chilodus punctatus* (Chilodontidae) e *Anostomus anostomus* (Anostomidae) (Characiformes). *Res. III Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 3.
- CHIARELLI (A. B.) and CAPANNA (E.), 1973. — Checklist of fish chromosomes. In : *Cytotaxonomy and vertebrate evolution*. A. B. Chiarelli et E. Capanna (eds.). Academic Press, New York : 206-232.
- DELLA-ROSA (V. A.), BERTOLLO (L. A. C.), FERRARI (I.), TAKAHASHI (C. S.), MOREIRA FILHO (O.) and FORESTI (F.), 1980. — Estudos citogenéticos em peixes da Amazônia. II. Ordem Siluriformes. *Ciência e Cultura (Supl.)*, 32 : 735.
- DENTON (T. E.), 1973. — *Fish chromosome methodology*. Charles C. Thomas Publishers. Illinois. 166 p.
- ERDTMAN (B.), CALCAGNOTO (D.), RABOLINI (L.) and MÁLABARBA (L. R.), 1990. — Variabilidade cromossômica em *Callichthys callichthys* (Callichthyidae, Siluriformes, Pisces). *Ciência e Cultura (Supl.)*, 42 : 452-453.

- FALCÃO (J. N.), 1988. — *Caracterização cariológica em peixes do gênero Triportheus (Teleostei, Characiformes, Characidae)*. Tese de Doutorado. Departamento de Genética e Matemática Aplicada à Biologia, Faculdade de Medicina de Ribeirão Preto, USP. São Paulo. 137 p.
- FALCÃO (J. N.), MOREIRA FILHO (O.) and BERTOLLO (L. A. C.), 1984. — An additional chromosome in two fish species. *Rev. Brasil. Genet.*, VII (1) : 109-118.
- FELDBERG (E.) and BERTOLLO (L. A. C.), 1985 a. — Karyotypes of 10 species of Neotropical cichlids (Pisces, Perciformes). *Caryologia*, 38 (3-4) : 257-268.
- FELDBERG (E.) and BERTOLLO (L. A. C.), 1985 b. — Nucleolar organizing regions in some species of Neotropical cichlid fish (Pisces, Perciformes). *Caryologia*, 38, (3-4) : 319-324.
- FELDBERG (E.), BERTOLLO (L. A. C.), ALMEIDA-TOLEDO (L. F.), FORESTI (F.), MOREIRA FILHO (O.) and SANTOS (A. F.), 1987. — Biological aspects of Amazonian fishes. IX. Cytogenetic studies in two species of the genus *Semaprochilodus* (Pisces, Prochilodontidae). *Genome*, 29 (1) : 1-4.
- FELDBERG (E.), PORTO (J. I. R.) and BERTOLLO (L. A. C.), 1992. — Karyotype evolution in Curimatidae (Teleostei, Characiformes) of the Amazon region. I. Studies on the genera *Curimata*, *Psectrogaster*, *Steindachnerina* and *Curimatella*. *Rev. Brasil. Genet.* 15 (2) : 369-383.
- FELDBERG (E.), PORTO (J.I.R.), BERTOLLO (L. A. C.) and NAKAYAMA (C. M.), in press. — Karyotype evolution in Curimatidae (Teleostei, Characiformes) from the Amazon region. II. Centric fissions in the genus *Potamorhina*. *Genome*, 36.
- FELDBERG (E.), PORTO (J. I. R.) and NAKAYAMA (C. M.), 1990. — Caracterização cariotípica em *Guianacara* sp (Perciformes, Labroidei, Cichlidae) do rio Trombetas-PA. *Res. III Simp. de Citog. Evol. e Aplic. Peixes Neotrop.* : 39.
- FENOCCHIO (A. S.) and BERTOLLO (L. A. C.), 1987. — Estudos citogenéticos preliminares de Siluriformes da bacia Amazônica. *Ciência e Cultura (Supl.)*, 39 : 732.
- FENOCCHIO (A. S.) and BERTOLLO (L. A. C.), 1992 — Karyotype similarities among Pimelodidae (Pisces, Siluriformes) from the Brazilian Amazon region. *Cytobios*, 69 : 41-46.
- FORESTI (F.), 1987. — *Estudos cromossômicos em Gymnotiformes (Pisces, Ostariophysi)*. Tese de Livre Docência. Instituto Básico de Biologia Médica e Agrícola, Universidade Estadual Júlio de Mesquita Filho, Botucatu, São Paulo. 171 p.
- FORESTI (F.), ALMEIDA-TOLEDO (L. F.) and TOLEDO-FILHO (S. A.), 1984. — Chromosome studies in *Gymnotus carapo* and *Gymnotus* sp (Pisces, Gymnotidae). *Caryologia*, 37 : 141-146.
- FORESTI (F.), ALMEIDA-TOLEDO (L. F.) and TOLEDO-FILHO (S. A.), 1989. — Supernumerary chromosome system, C-banding pattern characterization and multiple nucleolar organizer regions in *Moenkhausia sanctaflomenae* (Pisces, Characidae). *Genetica*, 79 : 107-114.
- GALETTI JR. (P. M.) and FORESTI (F.), 1986. — Evolution of ZZ/ZW system in *Leporinus* (Pisces, Anostomidae). Role of constitutive heterochromatin. *Cytogenet. Cell Genet.*, 43 : 43-46.
- GALETTI JR. (P. M.), FORESTI (F.), BERTOLLO (L.A.C.) and MOREIRA FILHO (O.), 1981. — Heteromorphic sex chromosomes in three species of the genus *Leporinus* (Pisces, Anostomidae). *Cytogen. Cell Genet.*, 28 : 138-142.
- GALETTI JR. (P. M.), MESTRINER (C. A.), VENERE (P. C.) and FORESTI (F.), 1991. — Heterochromatin and karyotype reorganization in fish of the family Anostomidae (Characiformes). *Cytogen. Cell Genet.*, 56 : 116-121.
- GÉRY (J.), 1977. — *Characoids of the world*. T.F.H. Pub. Inc., New Jersey, USA. 672 p.
- GIULIANO-CAETANO (L.), 1986. — *Estudo citogenético em Hoplerythrinus unitaeniatus (Pisces, Erythrinidae) de diferentes bacias hidrográficas brasileiras*. Dissertação de Mestrado. Departamento de Ciências Biológicas. Universidade Federal de São Carlos — SP. 84 p.
- GIULIANO-CAETANO (L.) and BERTOLLO (L. A. C.), 1988. — Karyotype variability in *Hoplerythrinus unitaeniatus* (Characiformes, Erythrinidae). I. Chromosome polymorphism in the rio Negro population (Manaus, state of Amazonas). — *Rev. Brasil. Genet.*, 11 (2) : 299-306.
- GIULIANO-CAETANO (L.) and BERTOLLO (L. A. C.), 1990. — Karyotypic variability in *Hoplerythrinus unitaeniatus* (Pisces, Characiformes, Erythrinidae). II. Occurrence of natural triploidy — *Rev. Brasil. Genet.*, 13 (2) : 231-237.
- GOLD (J. R.), KAREL (W. J.) and STRAND (M. R.), 1980. — Chromosome formulae of North American fishes. *The Progress. Fish-Cult.*, 42 (1) : 10-23.
- GOLDSTEIN (R. J.), 1973. — *Cichlids of the world*. T.F.H. Publications Co. USA, 382 p.
- GOULDING (M.), CARVALHO (M. L.) and FERREIRA (E. G.), 1988. — *Rio Negro : rich life in poor water*. The Hague : SPB Academic Publishing. 200 p.
- GREENWOOD (P. H.), ROSEN (D. E.), WEITZMAN (S. H.) and MYERS (G. S.), 1966. — Phyletic studies of teleostean fishes, with a provisional classification of living forms. *Bull. Amer. Mus. Nat. Hist.*, 131 : 339-455.
- GYLDENHOLM (A. O.) and SCHEEL (J. J.), 1971. — Chromosome numbers of fishes. I. *J. Fish Biol.*, 3 : 479-486.
- HINEGARDNER (R.) and ROSEN (D. E.), 1972. — Cellular DNA content and the evolution of Teleostean fishes. *Am. Nat.*, 106 : 621-644.
- HOWELL (W. M.), 1972. — Somatic chromosomes of the black ghost knifefish, *Apteronotus albifrons* (Pisces, Apterontidae). *Copeia*, 1972 : 191-193.
- HUDSON (R. G.), 1976. — A comparison of karyotypes and erythrocyte DNA quantities of several species of catfish (Siluriformes) with phylogenetic implications. In Le-grande (W. H.), 1981. — Chromosomal evolution in

- North American catfishes (Siluriformes, Ictaluridae) with particular emphasis on the madtoms, *Noturus*. *Copeia*, 1981 : 33-52.
- KOSSOWSKI (G.), BRACAMONTE (N. O.) and VELASCO (J. Q.), 1983. — Karyotype of the hybrid of *Colossoma macropomum* (female) (Cuvier, 1818) x *Mylossoma duriventris* (male) (Cuvier, 1818) and their progenies (Pisces, Cypriniformes, Characidae). *Acta Cient. Venez.*, 34 (2) : 173-175.
- KULLANDER (S. O.), 1983. — A revision of the South American cichlid genus *Cichlasoma* (Teleostei : Cichlidae). Stockholm, 296 p.
- KULLANDER (S. O.), 1986. — Cichlid fishes of the Amazon river drainage of Peru. Stockholm, 431 p.
- LAUDER (G. V.) and LIEM (K. F.), 1983. — The evolution and interrelationships of the Actinopterygian fishes. *Bull. Mus. Comp. Zool.*, 150 (3) : 95-197.
- LUEKEN (M.) and FOERSTER (W.), 1969. — Chromosomenuntersuchungen bei fischen mit einer vereinfachten. In Gyldenholm, (A.O.) and Scheel, (J.J.), 1971. — Chromosome numbers of fishes. I. *J. Fish Biol.*, 3 : 479-486.
- MARCON (J. L.), FELDBERG (E.), PORTO (J. I. R.) and AFFONSO (E. G.), 1992. — Estudos fisiológicos e citogenéticos em peixes do gênero *Dianema* (Siluriformes, Callichthyidae) da Amazônia central. *Res. IV Simp. Citon. Evol. e Aplic. Peixes Neotrop.* : 34.
- MESTRINER (C. A.) and GALETTI Jr. (P. M.), 1987. — Estudos cromossômicos em *Schizodon fasciatus* (Anostomidae, Characiformes). *Ciência e Cultura (Supl.)*, 39 : 787.
- MOREIRA FILHO (O.), BERTOLLO (L. A. C.), FERRARI (I.), TAKAHASHI (C. S.) and FORESTI (F.), 1980. — Estudos citogenéticos em peixes da região amazônica. III. Ordem Perciformes. *Ciência e Cultura (Supl.)*, 32 : 734.
- MURAMOTO (J.), OHNO (S.) and ATKIN (M. B.), 1968. — On the diploid state of the fish order Ostariophysi. *Chromosoma*, 24 : 59-66.
- NAKAYAMA (C. M.), JÉGU (M.) and FELDBERG (E.), 1986. — Caracterização citotaxonomica de algumas espécies do gênero *Serrasalmus* (Serrasalmidae) da Amazônia. *Res. XIII Congresso Brasileiro de Zoologia* : 335.
- NAKAYAMA (C. M.), JÉGU (M.), FELDBERG (E.) and PORTO (J. I. R.), 1992. — Diferenciação cromossômica em duas espécies crípticas de *Serrasalmus* (Characiformes, Serrasalmidae) da Amazônia central. *Res. IV Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 12.
- NAKAYAMA (C.M.), FELDBERG (E.) and PORTO (J. I. R.), 1988 a. — Variabilidade de NORs e número cromossômico no gênero *Serrasalmus* (Serrasalmidae, Characiformes). *Res. XV Congresso Brasileiro de Zoologia* : 370.
- NAKAYAMA (C. M.), FELDBERG (E.) and PORTO (J. I. R.), 1988 b. — Padrão de banda C em 4 espécies do gênero *Serrasalmus* (Characiformes, Serrasalmidae) de bacia Amazônica. *Res. II Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 25.
- NAKAYAMA (C. M.), FELDBERG (E.) and PORTO (J. I. R.), 1990. — Caracterização cromossômica em peixes da família Serrasalmidae : *Colossoma macropomum* e *Piaractus brachypomus* (Myleinae). *Res. III Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 23.
- NIJSSSEN (H.) and ISBRUCKER (I. J. H.), 1980. — A review of the genus *Corydoras* (Lacepède, 1803) (Pisces, Siluriformes, Callichthyidae), *Bijdragen Tol de Dierkunde*, 50 (1) : 190-220.
- OHNO (S.) and ATKIN (N. B.), 1966. — Comparative DNA values and chromosome complements in eight species of fishes. *Chromosoma*, 18 : 455-456.
- OJIMA (Y.), UENO (L.) and HAYASHI (M.), 1976. — A review of the chromosome numbers in fishes. *La Kromosomo II* (1) : 19-47.
- OLIVEIRA (C.), 1987. — *Estudos citogenéticos no gênero Corydoras* (Pisces, Siluriformes, Callichthyidae). Dissertação de Mestrado. Departamento de Biologia, Instituto de Biociências, Universidade de São Paulo. 154 p.
- OLIVEIRA (C.), ALMEIDA-TOLEDO (L. F.), FORESTI (F.), BRITSKI (H.) and TOLEDO-FILHO (S. A.), 1988 a. — Chromosome formulae of Neotropical freshwater fishes. *Rev. Brasil. Genet.*, 11 (3) : 577-624.
- OLIVEIRA (C.), ALMEIDA-TOLEDO (L. F.), FORESTI (F.) and TOLEDO-FILHO (S. A.), 1988 b. — Supernumerary chromosomes, Robertsonian rearrangement and multiple NORs in *Corydoras aeneus* (Pisces, Siluriformes, Callichthyidae). *Caryologia*, 41 (3-4) : 227-236.
- OLIVEIRA (C.), ALMEIDA-TOLEDO (L. F.) and TOLEDO-FILHO (S. A.), 1990 a. — Comparative cytogenetic analysis of three cytotypes of *Corydoras nattereri* (Pisces, Siluriformes, Callichthyidae). *Cytologia*, 55 : 21-26.
- OLIVEIRA (C.), ALMEIDA-TOLEDO (L. F.) and TOLEDO-FILHO (S.A.), 1990 b. — Estudos citogenéticos na família Callichthyidae (Pisces, Siluriformes) : os gêneros *Dianema* e *Brochis*. *Res. III Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 27.
- OLIVEIRA (M. I. B.) and FALCÃO (J. N.), 1992. — Caracterização cariotípica em algumas espécies de peixes dos igarapés do município de Manaus. *Res. I Congresso de Iniciação Científica do Amazonas* : 99.
- ORTEGA (H.) and VARI (R.), 1986. — Annotated checklist of the freshwater fishes of Peru. *Smithsonian Contributions to Zoology*, 437 : 25 p.
- PARK (E. H.), 1974. — A list of the chromosome numbers of fishes. *Coll. Rev. Coll. Lib. Arts & Sciences. Seoul Nat. Univ.*, 20 (1-2) : 346-372.
- PAULS (E.), 1985. — Considerações sobre a evolução cromossômica e sistema de cromossomos supranumerários em espécies do gênero *Prochilodus* (Pisces, Prochilodontidae). Tese de Doutorado. Departamento de Ciências Biológicas, Universidade Federal de São Carlos, São Paulo, Brasil. 156 p.

- PAULS (E.) and BERTOLLO (L.A.C.), 1990. — Distribution of a super numerary chromosome system and aspects of karyotypic evolution in the genus *Prochilodus* (Pisces, Prochilodontidae). *Genetica*, 81 : 117-123.
- Porto (J. I. R.), 1992. — *Estudos citotaxonômicos em peixes da família Hemiodidae (Ostariophysi, Characiformes), da Amazônia central*. Dissertação de Mestrado. Programa de Pós-Graduação INPA-FUA. Manaus-AM. 103 p.
- PORTO (J. I. R.) and FELDBERG (E.), 1988. — Considerações citotaxonômicas em *Callichthys callichthys* (Siluriformes, Callichthyidae) da Amazônia central. *Res. II Simp. Citog. Evol. Aplic. Peixes Neotrop.* : 8.
- PORTO (J. I. R.) and FELDBERG (E.), 1992 a. — Ocorrência de novos citótipos nos gêneros *Hoplosternum* e *Callichthys* (Siluriformes, Callichthyidae) da bacia amazônica. *Rev. Brasil. Genet.*, 15 (1) Supplement 2 : 81.
- PORTO (J. I. R.) and FELDBERG (E.), 1992 b. — Comparative cytogenetic study of the armored catfishes of the genus *Hoplosternum* (Siluriformes, Callichthyidae). *Rev. Brasil. Genet.*, 15 (2) : 359-367.
- PORTO (J. I. R.), FELDBERG (E.), NAKAYAMA (C. M.) and JÉGU (M.), 1989. — Análise cariotípica na família Serrasalmididae (Ostariophysi, Characiformes) : Aspectos evolutivos. *Ciência e Cultura (Supl.)* 41 : 714.
- PORTO (J. I. R.), FELDBERG (E.), NAKAYAMA (C. M.), MAIA (R. O.) and JÉGU (M.), 1991. — Cytotaxonomic analysis in the Serrasalmididae (Ostariophysi, Characiformes). *Bull. Zool. Mus. Univ. Amsterdam (abstracts)* : 66.
- POST (A.), 1965. — Vergleichende untersuchungen der chromosomenzahlen bei süsswasser teleosteein. *Z. Zool. Syst. Evol. Forsch.* 3 : 47-93.
- RAMIREZ-GIL (H.) and FELDBERG (E.), 1992. — Características cromossômicas de *Callophysus macropterus* (Siluriformes, Pimelodidae) da Amazônia Central (Ilha da Marchantaria e Anavilhanas). *Res. IV Simp. Citog. Evol. e Aplic. Peixes Neotrop.* : 31.
- SANGUINO (E. C. B.) and FALCÃO (J. N.), 1992. — Caracterização das regiões organizadoras de nucléolos de algumas espécies de peixes dos igarapés do município de Manaus. *Res. I Congresso de Iniciação Científica do Amazonas* : 100.
- SANTOS (A. F.), FELDBERG (E.) and BERTOLLO (L. A. C.), 1985. — Estudos citogenéticos em peixes da região amazônica : *Brycon* sp (Bryconinae, Characidae). *Res. XII Congresso Brasileiro de Zoologia* : 173.
- SCHEEL (J. J.), 1972. — The chromosomes of the third neon tetras. *Trop. Fish Hobbyist*, 20 (4) : 60-65.
- SCHEEL (J. J.), 1973. — *Fish chromosome and their evolution*. Internal reports of Danmarks Akvarium, Charlottenslund, Danmark, 22 p.
- SCHEEL (J. J.) and CHRISTENSEN (B.), 1970. — The chromosomes of the two common neon tetras. *Trop. Fish Hobbyist*, 19 (7) : 24-31.
- SCHEEL (J. J.), SIMONSEN (V.) and GYLDENHOLM (A. O.), 1972. — The karyotypes and some electrophoretic patterns of fourteen species of the genus *Corydoras*. *Z. Zool. Syst. Evol. Forsh.*, 10 : 144-152.
- SOLA (L.), CATAUDELLA (S.) and CAPANNA (E.), 1981. — New developments in vertebrate cytotoxicology. III. Karyology of bony fishes : a review. *Genetica*, 54 : 285-328.
- SUZUKI (A.), TAKI (Y.) and URUSHIDO (T.), 1982. — Karyotypes of two species of Arowana, *Osteoglossum bicirrhosum* and *O. ferreirai*. *Japan. J. Ichthyol.*, 29 (2) : 220-222.
- THOMPSON (K. W.), 1979. — Cytotoxicology of 41 species of Neotropical Cichlidae. *Copeia*, 4 : 679-691.
- TURNER (B. J.), DIFFOOT (N.), RASCH (E.M.), 1992. — The callichthyid catfish *Corydoras aeneus* is an unresolved diploid-tetraploid sibling species complex. *Ichthyol. Explor. Freshwaters*, 3 (1) : 17-23.
- URUSHIDO (T.), TAKAHASHI (E.) and TAKI (Y.), 1975. — Karyotypes of three species of fishes in the order Osteoglossiformes. *Chromosome Information Service*, 18 : 20-22.
- UYENO (T.), 1973. — A comparative study of chromosomes in the teleostean fish order Osteoglossiformes. In Suzuki A., Taki Y. and Urushido T., 1982. — Karyotypes of two species of Arowana, *Osteoglossum bicirrhosum* and *O. ferreirai*. *Japan. J. Ichthyol.*, 29 (2) : 220-222.
- VENERE (P. C.), 1988. — Nota sobre os cromossomos de algumas espécies do baixo rio Tocantins (Reservatório de Tucuruí, PA). *Res. II Simp. Citog. Evol. Aplic. Peixes Neotrop.* : 36.
- VENERE (P. C.) and GALETTI Jr. (P. M.), 1985. — Natural triploidy and chromosome B in the fish *Curimata modesta* (Curimatidae, Characiformes). *Rev. Brasil. Genet.*, 7 (4) : 681-687.
- VENERE (P. C.) and GALETTI Jr. (P. M.), 1986 a. — Aspectos citogenéticos em Anostomídeos da região Amazônica. *Ciência e Cultura (Supl.)*, 38 : 934.
- VENERE (P. C.) and GALETTI Jr. (P. M.), 1986 b. — Estudos cromossômicos em *Leporinus* (Anostomidae) de diferentes bacias hidrográficas. *Res. I Simp. Citog. Evol. Aplic. Peixes Neotrop.* : 62.
- VENERE (P. C.) and GALETTI Jr. (P. M.), 1989. — Chromosome relationships of some Neotropical Characiformes of the family Curimatidae. *Rev. Brasil. Genet.*, 12 (1) : 17-25.

TABLE I (1)

| ORDER FAMILY Species | NOR | | | | | | | | CBA | | | | reference |
|---|-----|----|-----------------|------|------|------|----|----------|-----|---|---|------|----------------------------------|
| | n | 2n | KF | numb | pair | type | am | location | I | P | T | HSex | |
| OSTEOGLOSSIFORMES | | | | | | | | | | | | | |
| ARAPAIMIDAE | | | | | | | | | | | | | |
| <i>Arapaima gigas</i> | - | 56 | 4M+12SM-ST+40T | - | - | - | - | - | - | - | - | - | Urushido <i>et al.</i> (1975) |
| OSTEOGLOSSIDAE | | | | | | | | | | | | | |
| <i>Osteoglossum bicirrhosum</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>O. bicirrhosum</i> | - | 56 | 1SM+1ST+54A | - | - | - | - | - | - | - | - | - | Uyeno (1973) |
| <i>O. bicirrhosum</i> | - | 56 | 3ST+53A | 2 | - | ST,A | p | TERM | - | - | - | - | Suzuki <i>et al.</i> (1982) |
| <i>O. ferreirai</i> | - | 54 | 2M+4SM+14ST+34A | - | - | - | - | - | - | - | - | - | Suzuki <i>et al.</i> (1982) |
| CHARACIFORMES | | | | | | | | | | | | | |
| ANOSTOMIDAE | | | | | | | | | | | | | |
| <i>Abramites hypselonotus</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Anostomus anostomus</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>A. anostomus</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>A. anostomus</i> | 27 | 54 | 54M-SM | - | - | - | - | - | - | - | - | - | Ojima <i>et al.</i> (1976) |
| <i>A. anostomus</i> | - | 54 | 54M-SM | - | - | - | - | - | - | - | - | - | Cestari <i>et al.</i> (1990) |
| <i>A. ternetzi</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Leporinus affinis</i> | - | 54 | 54M-SM | - | - | - | - | - | - | - | - | - | Venere (1988) |
| <i>L. brunneus</i> | - | 54 | 54M-SM | 2 | - | - | - | - | - | - | - | - | Venere & Galetti Jr. (1986b) |
| <i>L. cylindriciformes</i> | - | 54 | 54M-SM | 2 | - | - | - | - | - | - | - | - | Venere & Galetti Jr. (1986b) |
| <i>L. friderici</i> | - | 54 | 32M+22SM | 2 | 2 | M | p | TERM. | X | X | - | - | Galetti Jr. <i>et al.</i> (1991) |
| <i>L. ortomaculatus</i> | - | 54 | - | 2 | - | - | - | - | - | - | - | - | Venere & Galetti Jr. (1986b) |
| <i>L. tigrinus</i> | - | 54 | - | - | - | - | - | - | - | - | - | - | Venere (1988) |
| <i>Rhytidodus microlepis</i> | - | 54 | - | - | - | - | - | - | - | - | - | - | Bertollo <i>et al.</i> (1980) |
| <i>Schizodon fasciatus</i> | - | 54 | 54M-SM | - | - | - | - | - | - | - | - | - | Mestriner & Galetti Jr. (1987) |
| <i>S. fasciatus</i> | - | 54 | 28M+26SM | 2 | 12 | M | q | TERM. | - | X | X | - | Galetti Jr. <i>et al.</i> (1991) |
| <i>Schizodon sp.</i> | - | 54 | 54M-SM | 2 | - | - | - | - | - | - | - | - | Venere & Galetti Jr. (1986a) |
| CHARACIDAE | | | | | | | | | | | | | |
| BRYCONINAE | | | | | | | | | | | | | |
| <i>Brycon cf. cephalus</i> | - | 50 | 20M+22SM+8ST | 2 | - | ST | q | TERM. | X | X | X | - | Santos <i>et al.</i> (1985) |
| <i>B. cf. erythropterum</i> | - | 50 | 20M+22SM+8ST | 2 | - | ST | q | TERM. | X | X | X | - | Santos <i>et al.</i> (1985) |
| <i>B. cf. pesu</i> | - | 50 | 20M+22SM+8ST | 2 | - | ST | q | TERM. | X | X | X | - | Feldberg <i>et al.</i> , unpubl. |
| CHALCIDIINAE | | | | | | | | | | | | | |
| <i>Chalceus macrolepidotus</i> | - | 54 | 32M+22ST | - | - | - | - | - | - | - | - | - | Muramoto <i>et al.</i> (1968) |
| <i>C. macrolepidotus</i> | - | 52 | 44M-SM+8ST-T | - | - | - | - | - | - | - | - | - | Ojima <i>et al.</i> (1976) |
| CHEIRODONTINAE | | | | | | | | | | | | | |
| <i>C. troemneri</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Megalomphodus megalopterus</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>M. megalopterus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>M. sweglesi</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Microschemobrycon casiquiare</i> | 21 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Paracheirodon axelrodi</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>P. axelrodi</i> ¹ | - | 52 | - | - | - | - | - | - | - | - | - | - | Scheel & Christensen (1970) |
| <i>P. axelrodi</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>P. innesi</i> | - | 36 | - | - | - | - | - | - | - | - | - | - | Lueken & Foerster (1969) |
| <i>P. innesi</i> | - | 32 | - | - | - | - | - | - | - | - | - | - | Scheel & Christensen (1970) |
| <i>P. innesi</i> | 16 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>P. simulans</i> ² | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>P. simulans</i> | - | 50 | - | - | - | - | - | - | - | - | - | - | Scheel (1972) |
| <i>Pristella riddlei</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>P. riddlei</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| IGUANODECTINAE | | | | | | | | | | | | | |
| <i>Iguanodectes spilurus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| STETHAPPRIONINAE | | | | | | | | | | | | | |
| <i>Poptella compressa</i> ³ | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| TETRAGONOPTERINAE | | | | | | | | | | | | | |
| <i>Bryconella pallidifrons</i> ⁴ | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Ctenobrycon aff. hauxwellianus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Exodon paradoxus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Hemigrammus analis</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. erythrozonus</i> | - | 48 | - | - | - | - | - | - | - | - | - | - | Lueken & Foerster (1969) |

TABLE I (2)

| | | | | | | | | | | | | | | | |
|---|----|-----|------------------|-----|------|------|-----|----------|---|---|---|----|---------------|---|-----------------------------------|
| <i>H. erythrozonus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. gracilis</i> ⁵ | 24 | 48 | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. hyanuary</i> | - | 52 | 22M-SM+30ST-A | - | - | - | - | - | - | - | - | - | - | - | Arefjev(1990b) |
| <i>H. marginatus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. micropterus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. ocellifer</i> | 24 | 48 | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. ocellifer</i> | 19 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. pulcher</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. pulcher</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. rhodostomus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. rhodostomus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. rodwayi</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. schmardae</i> | - | 52 | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. schmardae</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. stictus</i> ⁶ | - | 52 | - | - | - | - | - | - | - | - | - | - | - | - | Scheel & Christensen (1970) |
| <i>H. stictus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. unilineatus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. vorderwinkleri</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Hyphessobrycon agulha</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. agulha</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. belloti</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. bentosi bentosi</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. aff. copelandi</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. erythro stigma</i> ⁷ | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. haraldschultzi</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. heterorhabdus</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. loretoensis</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. peruvianus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. pulchripinnis</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. pulchripinnis</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. scholzei</i> | - | 50 | 8M+20SM+8ST+14A | - | - | - | - | - | - | - | - | - | - | - | Arefjev (1990b) |
| <i>H. serpae</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| <i>H. serpae</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. stictus</i> | - | 52 | - | - | - | - | - | - | - | - | - | - | - | - | Gyldenholm & Scheel (1971) |
| <i>H. stictus</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>H. tropis</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Inpaichthys kerri</i> | - | 52 | 12M+26SM+14ST-A | - | - | - | - | - | - | - | - | - | - | - | Arefjev (1989) |
| <i>Moenkhausia colletti</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>M. oligolepis</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Thayeria boehkei</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>T. obliqua</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Post (1965) |
| TRIPORTHEINAE | | | | | | | | | | | | | | | |
| <i>Triportheus albus</i> | - | 52 | 14M+20SM+14ST+4A | 1-4 | 6,11 | ST | p | SUB,TER | - | X | X | ZW | Falcão (1988) | | |
| <i>T. culter</i> | - | 52 | 14M+16SM+16ST+6A | - | - | - | - | - | - | - | - | - | - | - | Falcão (1988) |
| <i>T. elongatus</i> | - | 52 | 22M+12SM+16ST+2A | 1-4 | 3,11 | ST | p | TERM. | - | X | X | ZW | Falcão (1988) | | |
| <i>T. flavus</i> | - | 52 | 22M+14SM+12ST+4A | 1-4 | 6,7 | M,ST | p,q | SUB,INT | - | X | X | ZW | Falcão (1988) | | |
| <i>T. pictus</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| CHILODIDAE | | | | | | | | | | | | | | | |
| <i>Chilodus punctatus</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. punctatus</i> | - | 54 | 54M,SM | - | - | - | - | - | - | - | - | - | - | - | Cestari <i>et al.</i> (1990) |
| CTENOLUCIIDAE | | | | | | | | | | | | | | | |
| <i>Boulengerella sp</i> | - | 36 | - | - | - | - | - | - | - | - | - | - | - | - | Porto <i>et al.</i> , unpubl. |
| CURIMATIDAE | | | | | | | | | | | | | | | |
| <i>Curimata cyprinoides</i> | - | 54 | 44M+10SM | 2 | 3 | M | q | TERM. | - | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>C. kneri</i> | - | 54 | 40M+12SM+2ST | 2 | 15 | ST | p | TERM. | - | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>C. inornata</i> | - | 54 | 40M+14SM | 2 | 4 | SM | p | INTERST. | - | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>C. ocellata</i> | - | 56 | 40M+16SM | 2 | 22 | SM | p | INTERST. | - | - | - | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>C. vittata</i> | - | 54 | 42M+12SM | 1 | 9 | SM | q | TERM. | - | - | - | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>Curimatella alburna</i> | - | 54 | 46M+8SM | 2 | 14 | M | q | TERM. | - | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>C. meyeri</i> | - | 54 | 46M+8SM | 2 | 9 | M | q | TERM. | - | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>Curimatopsis aff. macrolepis (CYT A)</i> | 26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. aff. macrolepis (CYT B)</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Cyphocharax cf. spilura</i> | - | 54 | - | 2 | 10 | M | q | TERM. | - | - | - | - | - | - | Venere & Galetti Jr. (1989) |
| <i>Potamorhina altamazonica</i> | 51 | 102 | 2M+2SM+98A | 2 | 5 | A | q | TERM. | X | X | X | - | - | - | Feldberg <i>et al.</i> (in press) |
| <i>P. latior</i> | - | 56 | 58M+2SM+2ST | 2 | 27 | M | q | TERM. | X | X | X | - | - | - | Feldberg <i>et al.</i> (in press) |
| <i>P. pristigaster</i> | - | 54 | 42M+12SM | 2 | 16 | SM | q | TERM. | - | X | - | - | - | - | Feldberg <i>et al.</i> (in press) |
| <i>Psectrogaster rutiloides</i> | - | 54 | 42M+12SM | 2 | 11 | M | q | TERM. | X | X | X | - | - | - | Feldberg <i>et al.</i> (1992) |
| <i>Steindachnerina leuciscus</i> | - | 54 | 48M+6SM | 2 | 16 | M | q | TERM. | - | - | - | - | - | - | Feldberg <i>et al.</i> (1992) |
| ERYTHRINIDAE | | | | | | | | | | | | | | | |
| <i>Erythrinus erythrinus</i> | - | 52 | - | 1-8 | - | M | q,p | TERM. | - | - | - | - | - | - | Bertollo (1988) |
| <i>Hoplerethrinus unitaeniatus</i> | - | 48 | 48M-SM | - | - | - | - | - | - | - | - | - | - | - | Giuliano-Caetano (1986) |

TABLE I (3)

| | | | | | | | | | | | | | | | |
|---|-------|----|-----------------|------|----|------|-----|----------|---|---|---|-----------------|----------------|----|--|
| <i>H. unitaeniatus</i> (CYT A) | 24 | 48 | 48M-SM | - | - | - | - | - | - | - | - | - | - | - | Giuliano-Caetano (1986) |
| <i>H. unitaeniatus</i> (CYT B) | 24 | 48 | 46N-SM+2ST-A | 3-4 | - | M,A | q | INTERST. | - | - | - | - | - | - | Giuliano-Caetano (1986) |
| <i>H. unitaeniatus</i> (CYT C) | 24 | 48 | 46N-SM+2ST-A | 2 | - | M | q | TER,INT. | X | X | X | - | - | - | Giuliano-Caetano (1986) |
| <i>H. unitaeniatus</i> (CYT D) | 24 | 48 | 47M-SM+1ST-A | 4 | - | M,A | q | INTERST. | X | X | X | - | - | - | Giuliano-Caetano (1986) |
| <i>H. unitaeniatus</i> (Tryploid) | - | 72 | 69M-SM+3ST-A | 3 | - | M-SM | q | TERM. | - | - | - | - | - | - | Giuliano-Caetano (1986) |
| <i>Hoplias malabaricus</i> | - | 40 | - | - | - | - | - | - | - | - | - | - | - | - | Bertollo <i>et al.</i> (1980) |
| <i>Hoplias</i> sp. | 20/21 | 41 | 29M+11SM+1A (M) | - | - | - | - | - | - | - | - | XY ₁ | Y ₂ | - | Bertollo <i>et al.</i> (1983) |
| | - | 40 | 30M+10SM (F) | - | - | - | - | - | - | - | - | - | - | XX | Bertollo <i>et al.</i> (1983) |
| <i>Hoplias</i> sp. | - | 50 | - | - | - | - | - | - | - | - | - | - | - | - | Bertollo & Moreira F ^o (1983) |
| GASTEROPELECIDAE | | | | | | | | | | | | | | | |
| <i>Carnegiella strigata</i> | 25/26 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. strigata</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Gasteropelecus sternicla</i> | 27 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| HEMIODIDAE | | | | | | | | | | | | | | | |
| ANODINAE | | | | | | | | | | | | | | | |
| <i>Anodus elongatus</i> | - | 54 | 24M+26SM+4ST | 2 | 26 | ST | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| <i>Anodus melanopogon</i> | - | 54 | 20M+28SM+6ST | 2 | 25 | ST | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| <i>Anodus</i> sp. | - | 54 | 24M+24SM+6ST | 2 | 25 | ST | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| BIVIBRANCHIINAE | | | | | | | | | | | | | | | |
| <i>Argonectes scapularis</i> | - | 54 | 50M-SM+4ST | 2 | - | ST | q | TERM. | - | - | - | - | - | - | Porto, unpubl. |
| HEMIODINAE | | | | | | | | | | | | | | | |
| <i>Hemiodus immaculatus</i> | 27 | 54 | 22M+26SM+6ST | 2 | 25 | ST | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| <i>H. cf. microlepis</i> | - | 54 | 20M+30SM+4ST | 2 | - | SM | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| <i>H. ocellatus</i> | 27 | 54 | 26M+24SM+4ST | 2 | 15 | SM | q | SUBT. | - | - | - | - | - | - | Porto (1992) |
| <i>H. unimaculatus</i> | 27 | 54 | 26M+24SM+4ST | 2 | 17 | SM | q | TERM. | - | - | - | - | - | - | Porto (1992) |
| LEBIASINIDAE | | | | | | | | | | | | | | | |
| <i>Copeina guttata</i> | 21 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Copella amoldi</i> (CYT A) | 22 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Copella amoldi</i> (CYT B) | 22 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Copella nattereri</i> | 18 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Nannostomus beckfordi</i> (CYT A) | 22 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. beckfordi</i> (CYT B) | 18 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. beckfordi</i> (CYT C) | - | 42 | 2M+40A | - | - | - | - | - | - | - | - | - | - | - | Arefjev (1990a) |
| <i>N. erythrurus</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. marginatus</i> | 21 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. trifasciatus</i> (CYT A) | 19 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. trifasciatus</i> (CYT B) | 15 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. trifasciatus</i> (CYT C) | 12 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. eques</i> ⁸ | - | 34 | 34A | - | - | - | - | - | - | - | - | - | - | - | Arefjev (1990a) |
| <i>N. harrisoni</i> ⁹ | 20 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>N. unifasciatus</i> ¹⁰ | 11 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Pyrrhulina</i> sp. | - | 42 | 2M+2SM+38ST-A | - | - | - | - | - | - | - | - | - | - | - | Oliveira & Falcão (1992) |
| <i>Pyrrhulina</i> sp. | - | 42 | - | 2-4 | - | - | - | - | - | - | - | - | - | - | Sanguino & Falcão (1992) |
| PROCHILODIDAE | | | | | | | | | | | | | | | |
| <i>P. nigricans</i> | - | 54 | 40M+14SM | 2 | 2 | M | q | INTERST. | - | X | - | - | - | - | Pauls & bertollo (1990) |
| <i>Semaprochilodus insignis</i> | 27 | 54 | 40M+14SM | 2 | 3 | M | p | INTERST. | - | X | - | - | - | - | Feldberg <i>et al.</i> (1987) |
| <i>S. taeniurus</i> | 27 | 54 | 40M+14SM | 2 | 3 | M | p | INTERST. | X | X | X | ZW | - | - | Feldberg <i>et al.</i> (1987) |
| SERRASALMIDAE | | | | | | | | | | | | | | | |
| <i>Catoprion mento</i> | - | 60 | - | - | - | - | - | - | - | - | - | - | - | - | Porto <i>et al.</i> , unpubl. |
| <i>Colossoma macropomum</i> ¹¹ | 27 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. macropomum</i> | - | 54 | - | - | - | - | - | - | - | - | - | - | - | - | Bertollo <i>et al.</i> (1980) |
| <i>C. macropomum</i> | - | 54 | 18M+36SM | - | - | - | - | - | - | - | - | - | - | - | Kossowski <i>et al.</i> (1983) |
| <i>C. macropomum</i> | - | 54 | 20M+34SM | 1-4 | - | M | q | TERM. | - | X | X | - | - | - | Almeida-Toledo <i>et al.</i> (1987) |
| <i>C. macropomum</i> | - | 54 | 26M+28SM | 1-4 | - | M | q | TER,INT | - | X | X | - | - | - | Nakayama <i>et al.</i> (1990) |
| <i>Metynnis argenteus</i> | 31 | - | - | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>M. lippincottianus</i> | - | 62 | 30M+30SM+2A | - | - | - | - | - | - | - | - | - | - | - | Arefjev (1989) |
| <i>M. schreitmueelleri</i> ¹² | - | 62 | 60M-SM+2ST | - | - | - | - | - | - | - | - | - | - | - | Ojima <i>et al.</i> (1976) |
| <i>M. sp.</i> | - | 62 | 54M-SM+8ST-A | 3-4 | - | ST-A | q | INTERST. | - | - | - | - | - | - | Porto <i>et al.</i> (1989;1991) |
| <i>Mylesinus paraschomburgkii</i> | - | 58 | - | 6-12 | - | ST-A | p | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>Myleus pacu</i> | - | 58 | 40M-SM+18ST-A | 5-9 | - | ST-A | p | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>M. rubripinnis</i> | - | 58 | - | 5-8 | - | ST-A | p | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>M. shomburgkii</i> | - | 58 | 42M-SM+16ST-A | 5-8 | - | ST-A | p | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>Mylossoma aureum</i> | - | 54 | 54M-SM | 6-14 | - | M,SM | p,q | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>M. duriventris</i> | - | 54 | 50M-SM+4T | - | - | - | - | - | - | - | - | - | - | - | Ojima <i>et al.</i> (1976) |
| <i>M. duriventris</i> | - | 54 | 18M+34SM+2A | - | - | - | - | - | - | - | - | - | - | - | Kossowski <i>et al.</i> (1983) |
| <i>M. duriventris</i> | - | 54 | 54M-SM | 6-14 | - | M,SM | p,q | TERM. | - | - | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>n. gen.</i> ¹³ | - | 58 | 42M-SM+16ST-A | 6-12 | - | ST-A | p | TERM. | - | X | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |

TABLE I (4)

| | | | | | | | | | | | | | |
|---------------------------------------|-------|-----|-------------------|------|---|-------|-----|----------|---|---|---|---|-------------------------------------|
| <i>Piaractus brachypomus</i> | - | 54 | 34M+20SM | 3-6 | - | M,SM | p,q | TERM. | X | X | X | - | Nakayama <i>et al.</i> (1990) |
| <i>Pristobrycon eigenmanni</i> | - | 60 | 44M-SM+16ST-A | - | - | - | - | - | X | X | X | - | Porto <i>et al.</i> (1989; 1991) |
| <i>P. serrulatus</i> | - | 60 | 44M-SM+16ST-A | 6-10 | - | ST-A | p | TERM. | - | X | X | - | Porto <i>et al.</i> (1989; 1991) |
| <i>P. striolatus</i> | - | 62 | 46M-SM+16ST-A | 6-7 | - | ST-A | p | TERM. | - | - | - | - | Porto <i>et al.</i> (1989; 1991) |
| <i>Pristobrycon sp.</i> | - | 60 | 48M-SM+12ST-A | 6-10 | - | ST-A | p | TERM. | - | - | - | - | Nakayama <i>et al.</i> (1988a) |
| <i>P. sp.</i> | - | 60 | 50M-SM+10ST-A | 6-8 | - | ST-A | p | TERM. | - | - | - | - | Nakayama <i>et al.</i> (1988a) |
| <i>Pygocentrus nattereri</i> | - | 60 | 50M-SM+10ST-A | 6-8 | - | ST-A | p | TERM. | X | X | X | - | Nakayama <i>et al.</i> (1988a,b) |
| <i>Serrasalmus altuvei</i> | - | 60 | 46M-SM+14ST-A | - | - | - | - | - | - | - | - | - | Nakayama <i>et al.</i> , unpubl. |
| <i>S. elongatus</i> | - | 60 | 40M-SM+20ST-A | 6-12 | - | ST-A | p | TERM. | - | X | X | - | Nakayama <i>et al.</i> (1986,1988b) |
| <i>S. hollandi</i> | - | 64 | - | - | - | - | - | - | - | - | - | - | Muramoto <i>et al.</i> (1968) |
| <i>S. manuelli</i> | - | 60 | 44M-SM+16ST-A | 6-10 | - | ST-A | p | TERM. | - | X | X | - | Porto <i>et al.</i> (1991) |
| <i>S. rhombeus</i> | - | 60 | 44M-SM+16ST-A | - | - | - | - | - | - | - | - | - | Nakayama <i>et al.</i> (1992) |
| <i>Serrasalmus sp.1</i> | - | 58 | 46M-SM+12ST-A | 4-8 | - | ST,A | p | TERM. | X | X | X | - | Nakayama <i>et al.</i> (1992) |
| <i>Serrasalmus sp.2</i> | - | 60 | 46M-SM+14ST-A | - | - | - | - | - | - | X | X | - | Nakayama <i>et al.</i> (1988b) |
| <i>S. spilopleura</i> | - | 60 | 44M-SM+16ST-A | 6-10 | - | ST-A | p | TERM. | - | X | X | - | Porto <i>et al.</i> (1991) |
| SILURIFORMES | | | | | | | | | | | | | |
| SILUROIDEI | | | | | | | | | | | | | |
| AGENEIOSIDAE | | | | | | | | | | | | | |
| <i>Ageneiosus brevifilis</i> | - | 56 | - | - | - | - | - | - | - | - | - | - | Fennocchio & Bertollo (1987) |
| <i>Ageneiosus sp.</i> | - | 54 | - | - | - | - | - | - | - | - | - | - | Fennocchio & Bertollo (1987) |
| AUCHENIPTERIDAE | | | | | | | | | | | | | |
| <i>Parauchenipterus cf. galeatus</i> | - | 58 | - | - | - | - | - | - | - | - | - | - | Fennocchio & Bertollo (1987) |
| CALLICHTHYIDAE | | | | | | | | | | | | | |
| <i>Brochis splendens</i> 14 | 49 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>B. splendens</i> | - | 100 | 18M+18SM+20ST+44A | 4 | - | SM,ST | q | TERM. | - | X | - | - | Oliveira <i>et al.</i> (1990b) |
| <i>Callichthys callichthys</i> | - | 52 | - | 2 | - | SM,ST | p | TERM. | - | - | - | - | Porto & Feldberg (1992b) |
| <i>C. callichthys</i> | - | 54 | 46M-SM+8ST-A | 3 | - | SM,ST | p | TERM. | - | - | - | - | Porto & Feldberg (1988) |
| <i>C. callichthys</i> | - | 58 | - | - | - | - | - | - | - | - | - | - | Porto & Feldberg (1992b) |
| <i>Corydoras aeneus</i> 15 | - | 132 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. aeneus</i> | 66 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>Corydoras aeneus</i> | 60 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>C. aeneus</i> 16 | - | 58 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. aeneus</i> | 29 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. aeneus</i> | - | 134 | - | - | - | - | - | - | - | - | - | - | Turner <i>et al.</i> (1992) |
| <i>C. aeneus</i> | - | 56 | - | - | - | - | - | - | - | - | - | - | Turner <i>et al.</i> (1992) |
| <i>C. agassizii</i> | - | 98 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. agassizii</i> | 49 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. arcuatus</i> | - | 46 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. arcuatus</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. elegans</i> | - | 50 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. elegans</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. elegans</i> | 25 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>C. melanistus</i> | - | 46 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. melanistus</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. melanistus</i> | 24 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>C. punctatus</i> | 22-23 | - | - | - | - | - | - | - | - | - | - | - | Hinegardner & Rosen (1972) |
| <i>C. aff. punctatus</i> | - | 102 | 8M+14SM+20ST+60A | 2 | - | SM | p | TERM. | - | X | X | - | Oliveira (1987) |
| <i>C. rabauti</i> | - | 58 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. rabauti</i> | 29 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. reticulatus</i> | - | 74 | 16M+20SM+12ST+26A | 2-3 | - | A | q | TERM. | - | X | X | - | Oliveira (1987) |
| <i>C. schwartzi</i> | - | 46 | - | - | - | - | - | - | - | - | - | - | Scheel <i>et al.</i> (1972) |
| <i>C. schwartzi</i> | 23 | - | - | - | - | - | - | - | - | - | - | - | Scheel (1973) |
| <i>C. schwartzi</i> | - | 46 | 32M+14SM | 2 | - | M | p | TERM. | - | - | - | - | Oliveira (1987) |
| <i>Dianema longibarbis</i> | - | 60 | - | - | - | - | - | - | - | - | - | - | Hudson (1976) |
| <i>D. longibarbis</i> | - | 60 | 8M-SM+52ST-A | 2 | - | SM | p | INTERST. | - | - | - | - | Marcon <i>et al.</i> (1992) |
| <i>D. urostriatum</i> 17 | - | 62 | 8M+4SM+4ST+46A | 2 | - | A | p | TERM. | - | X | X | - | Oliveira <i>et al.</i> (1990b) |
| <i>D. urostriatum</i> | - | 62 | 6M-SM+56ST-A | 2 | - | ST | p | TERM. | - | - | - | - | Marcon <i>et al.</i> (1992) |
| <i>H. littorale</i> | - | 60 | 8M-SM+52ST-A | 2 | - | A | q | TERM. | - | - | - | - | Porto & Feldberg (1992b) |
| <i>H. thoracatum</i> | - | 64 | 30M-SM+34ST-A | 2 | - | A | q | TERM. | - | - | - | - | Porto & Feldberg (1992b) |
| <i>H. aff. thoracatum</i> | - | 66 | - | - | - | - | - | - | - | - | - | - | Porto & Feldberg (1992a) |
| <i>H. sp.</i> | - | 62 | 8M-SM+54ST | 4 | - | M-SM | p | TERM. | - | - | - | - | Porto & Feldberg (1992a) |
| DORADIDAE | | | | | | | | | | | | | |
| <i>Opsodoras humeralis</i> | - | 58 | - | - | - | - | - | - | - | - | - | - | Della-Rosa <i>et al.</i> (1980) |
| <i>Pseudodoras niger</i> | - | 58 | 58M-SM-ST-A | - | - | - | - | - | - | - | - | - | Venere (1988) |
| LORICARIIDAE | | | | | | | | | | | | | |
| <i>Pterygoplichthys multiradiatus</i> | - | 52 | - | - | - | - | - | - | - | - | - | - | Della-Rosa <i>et al.</i> (1980) |
| <i>Loricaria sp.</i> | - | 62 | - | - | - | - | - | - | - | - | - | - | Della-Rosa <i>et al.</i> (1980) |

TABLE I (6)

| | | | | | | | | | | | | | | | | |
|------------------------------|---|----|--------------|---|---|---|---|---|---|---|---|---|---|---|---|--------------------------------|
| <i>S. aequifasciatus</i> | - | 60 | 58M-SM+2ST-T | - | - | - | - | - | - | - | - | - | - | - | - | Thompson (1979) |
| <i>Uaru amphiacanthoides</i> | - | 46 | 8M-SM+38ST-T | - | - | - | - | - | - | - | - | - | - | - | - | Thompson (1979) |
| LEPIDOSIRENIFORMES | | | | | | | | | | | | | | | | |
| LEPIDOSIRENIDAE | | | | | | | | | | | | | | | | |
| <i>Lepidosiren paradoxa</i> | - | 38 | 38M | - | - | - | - | - | - | - | - | - | - | - | - | Ohno & Atkin (1966) |
| <i>L. paradoxa</i> | - | 38 | 38M-SM-ST | - | - | - | - | - | - | - | - | - | - | - | - | Oliveira <i>et al.</i> (1988a) |

1 = *Cheirodon axelrodi*
 5 = *Hemigrammus gracilis*
 9 = *P. harrisoni*
 13 = *Utiaritchthys* sp
 17 = *Dianema urostriata*
 21 = *Geophagus surinamensis*
 25 = *Geophagus jurupari*

2 = *Hyphessobrycon simulans*
 6 = *Hemigrammus stictus*
 10 = *P. unifasciatus*
 14 = *Brochis coeruleus*
 18 = *Apistogramma pertense*
 22 = *Cichlasoma severum*
 26 = *Symphysodon aequifasciata*

3 = *Poptella orbicularis*
 7 = *Hyphessobrycon erythrostigma*
 11 = *Piaractus nigripinnis*
 15 = *Corydoras aneus*
 19 = *Crenicara filamentosa*
 23 = *Cichlasoma coryphaenoides*

4 = *Bryconella palifrons*
 8 = *Poecilobrycon oques*
 12 = *Metynnis hypsauchen*
 16 = *Corydoras schultzei*
 20 = *Crenicara maculatus*
 24 = *Aequidens curviceps*