December 2014

Diversity of Anopheles species and trophic behavior of putative malaria vectors in two malaria endemic areas of northwestern Thailand

Krajana Tainchum¹, Wanapa Ritthison¹, Thipwara Chuaycharoensuk¹, Michael J. Bangs^{1, 2}, Sylvie Manguin³, and Theeraphap Chareonviriyaphap¹⊠

¹Department of Entomology, Faculty of Agriculture, Kasetsart University, Bangkok 10900, Thailand, faasthc@ku.ac.th ²Public Health & Malaria Control Department, International SOS, Jl. Kertajasa, Kuala Kencana, Papua 99920, Indonesia ³Institut de Recherche pour le Développement, UMR-MD3, Montpellier 34093, France

Received 28 June 2014; Accepted 2 September 2014

ABSTRACT: We determined the species diversity, blood-feeding behavior, and host preference of *Anopheles* mosquitoes in two malaria endemic areas of Tak (Mae Sot District) and Mae Hong Son (Sop Moei District) Provinces, located along the Thai border with Myanmar, during a consecutive two-year period. Anopheline mosquitoes were collected using indoor and outdoor human-landing captures and outdoor cow-baited collections. Mosquitoes were initially identified using morphological characters, followed by the appropriate multiplex AS-PCR assay for the identification of sibling species within Anopheles (Cellia) complexes and groups present. Real-time PCR was performed for parasite-specific detection in mosquitoes (*Plasmodium* spp. and Wuchereria bancrofti). A total of 7,129 Anopheles females were captured, 3,939 from Mae Sot and 3,190 from Sop Moei, with 58.6% and 37% of all anophelines identified as An. minimus, respectively. All three malaria vector complexes were detected in both areas. One species within the Minimus Complex (An. minimus) was present along with two related species in the Funestus Group, (An. aconitus, An. varuna), two species within the Dirus Complex (An. dirus, An. baimaii), and four species within the Maculatus Group (An. maculatus, An. sawadwongporni, An. pseudowillmori, and An. dravidicus). The trophic behavior of An. minimus, An. dirus, An. baimaii, An. maculatus, and An. sawadwongporni are described herein. The highest An. minimus densities were detected from February through April of both years. One specimen of An. minimus from Mae Sot was found positive for Plasmodium vivax. Journal of Vector Ecology 39 (2): 424-436. 2014.

Keyword Index: Anopheles dirus, Anopheles minimus, Anopheles maculatus, species complexes, malaria, Thailand.

INTRODUCTION

After years of intensive, well-organized malaria control activities in Thailand, malaria still remains prevalent in vulnerable areas, especially along the less developed international borders with Myanmar. Approximately 85% of all reported malaria cases in the country occur in this poorly monitored border region, primarily related to transient employment opportunities or occupational activities including agriculture, hunting, and gem mining. Several primary malaria vector species in Thailand are abundant in forested areas along the international border (Chareonviriyaphap et al. 2003, Corbel et al. 2013). Bancroftian filariasis is also endemic along the Thai-Myanmar border but of low prevalence. However, the detection of this parasite is not uncommon among Burmese working in Thailand (Khamboonruang et al. 1987, Jitpakdi et al. 1998, Bhumiratana et al. 2002). Nocturnally subperiodic (NSP) Wuchereria bancrofti is transmitted by a wide variety of mosquito species including anophelines also capable of malaria transmission (Pothikasikorn et al. 2008).

Of the 73 known Anopheles species in Thailand, various sibling species in the Minimus and Dirus Complexes and the Maculatus Group have been recognized as the main malaria vectors in the country (Rattanarithikul et al. 2006, Sinka et al. 2011). Molecular techniques based on polymerase chain reaction (PCR) have allowed for precise and reliable differentiation and identification of the sibling species of medical importance (Manguin et al. 2008, Sinka et al. 2011). Among these 73 species, five to six species, depending on the literature, are incriminated as primary malaria vectors in Thailand (Rattanarithikul et al. 2006), including Anopheles baimaii (previously An. dirus D) (Green et al. 1991) and Anopheles dirus (Rosenberg et al. 1990, Green et al. 1991) of the Dirus Complex, Anopheles minimus (previously An. minimus A) (Rattanarithikul et al. 1996a) of the Minimus Complex, and Anopheles pseudowillmori (Green et al. 1991), An. maculatus and An. sawadwongporni of the Maculatus Group (Saeung 2012). Additionally, An. campestris and An. epiroticus (Sundaicus Complex) have also been incriminated as potential malaria vectors in Thailand (Apiwathnasor et al. 2002). Wuchereria bancrofti develops experimentally in An. minimus and An. maculatus (Pothikasikorn et al. 2008). Natural infections of this parasite have also been found in An. minimus, An. maculatus and An. vagus in Thailand (Pothikasikorn et al. 2008).

While the number of cryptic species has increased, Thailand has been identifying sibling species within the complexes for several decades, yet information on the distribution, ecology and behavior of many of these species remains poor or lacking. This information is quite critical in defining the vector capacity of each species (Takken and Verhulst 2013). Knowledge on mosquito behavior is crucial to understanding the epidemiology of vector transmission throughout the range of each species and for assisting national and international efforts in vector-borne disease control. Numerous observations on trophic behavioral and seasonal abundance have been conducted on Anopheles complexes (sensu lato) or related groups (Ismail et al. 1974, Harbach et al. 1987, Ratanatham et al. 1988, Rattanarithikul et al. 1996a, Chareonviriyaphap et al. 2003). Unfortunately, most of these observations relied on species that were identified by morphological characters alone, as molecular assays capable of reliably identifying each sibling species were not readily available or still non-existent. During the past few decades, studies on malaria vectors in Thailand using molecular identification assays have increased and provided precise identification to the species level, as well as the recognition of additional Anopheles species and species complexes in Thailand (Baimai et al. 1984, 1988, Baimai 1989, Green et al. 1990, Green et al. 1992, Poopittayasataporn and Baimai 1995, Walton et al. 1999, Garros et al. 2004, Rattanarithikul et al. 2006, Dusfour et al. 2007, Walton et al. 2007, Poolprasert et al. 2008, Saeung et al. 2008, Thongsahuan et al. 2009, Eamkum et al. 2014).

Previous studies have described the biting activity of *An*. minimus and An. harrisoni in malaria-endemic Tak (Tisgratog et al. 2012) and Kanchanaburi Provinces (Sungvornyothin et al. 2006), and both biting activity and host preference of An. dirus and An. baimaii in Kanchanaburi Province (Tananchai et al. 2012). However, additional investigations on the bionomics and blood feeding activities of sympatric malaria vectors belonging to species complexes are needed. In the present study, molecular identification assays were combined with the systematic descriptions of the trophic behavior, biting activity, seasonal abundance, and parasite infections of individual sibling species in the two most malaria endemic areas of Mae Sot (Tak Province) and Sop Moei (Mae Hong Son Province), northwestern Thailand. Additionally, overall anopheline species diversity from these two areas along the Thai-Myanmar border is described.

MATERIALS AND METHODS

Collection sites

Adult mosquito collections were conducted every two months during three consecutive nights, for a period of two years in one village each in Mae Sot District (Tak Province) and Sop Moei District (Mae Hong Son Province) located in northwestern Thailand (Figure 1).

The Mae Sot field site has a western boundary with Kayin State, Myanmar and located at approximately 471 m above sea level. There were five isolated hill tribes living in the area, the most common ethnic group being Karen (> 85%). Rubber plantations, fruit orchards and other agricultural crops surrounded most of the study site. A narrow, slow running stream, approximately 0.5 m in average depth and 2 m wide, ran across the village.

The Sop Moei study site neighbors Myanmar, Shan State to the north and Kayin and Kayah States to the west. The field site is located approximately 50 m from the Ngao River and

one kilometer from Mae Ngao National Park. The study site is surrounded by secondary forest at approximately 126 m above sea level. There are several tribal groups in this area in which the most common is Thai.

Collection methods

Standard landing collection techniques were used such as indoor and outdoor human-baited landing captures and cattle-baited outdoor trap collections. Indoor and outdoor human landing collections (HLC) were conducted by two teams of four people each. The first team worked from 18:00 to 24:00, followed by a second team from midnight to 06:00. Two people sat inside the house, while the other two took up a position outside approximately 100 m from the same hut. Human-landing collections occurred for 45 min each hour followed by a 15 min resting period. The collectors rotated hourly between indoor and outdoor collections. Cattle bait collections were conducted by one separate collector for 15 min each hour. Additional details on human landing collection methods were described previously (Tisgratog et al. 2012). Collected mosquitoes were kept in a clean plastic cup covered with netting and provided 10% sugar solution soaked on cotton. The following morning, mosquitoes were taken back to the field laboratory for morphological identification. Hourly ambient outdoor air temperatures and relative humidity were recorded. Rainfall data was collected from the Mae Sot and Sop Moei District meteorological stations, located approximately 10 km from each village. The human and animal use protocols for this study were approved by the Ethical Research Committee, Chulalongkorn University, Bangkok, Thailand (No. 0961/56).

Mosquito species identification

Each female mosquito was intially sorted using morphological keys (Rattanarithikul et al. 2006), individually stored in a clean, labelled 1.5 ml non-silicone tube, and immediately frozen and stored in a liquid nitrogen tank. Mosquito samples were sent to the Department of Entomology, Kasetsart University, in Bangkok. Individual An. minimus s.l., An. dirus s.l. and An. maculatus s.l. were placed in a DNA extraction tube and homogenized in 50 ml of extraction buffer (0.2 M sucrose, 0.1M Tris-HCl at pH 8.0, 50mM EDTA and 0.5% SDS). A volume of 7 μ l of 8 mM KOAc (pH 9.0) was added and the tube placed on ice for 30 min. The sample was centrifuged at 12,000 rpm for 20 min and the supernatant was removed to a clean tube. Then, 100 µl of 100% ethanol was added and the samples placed at 4° C for 30 min. Samples were spun at 12,000 rpm for 20 min at 4° C. The supernatant was again cleaned using 150 µl of 70% ethanol and centrifuged at 12,000 rpm for 5 min at 4° C, and again with 100% ethanol and centrifuged at 12,000 rpm for 5 min at 4° C. The resultant pellet was dried at room temperature for 20 min before being re-suspended in 100 µl of TE buffer and stored at -20°C, based on Linton et al. (2001) and Manguin et al. (2002).

Species complex or group specific multiplex allelespecific polymerase chain reaction (AS-PCR) assays were used for molecular species identification within the 1) Minimus

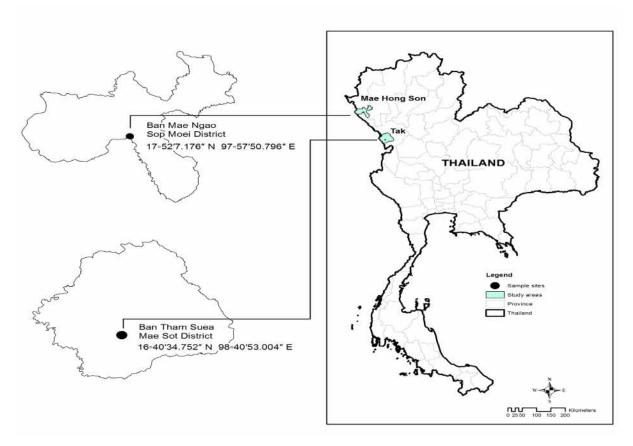


Figure 1. Study site locations in Mae Sot District, Tak Province and Sop Moei District, Mae Hong Son Province.

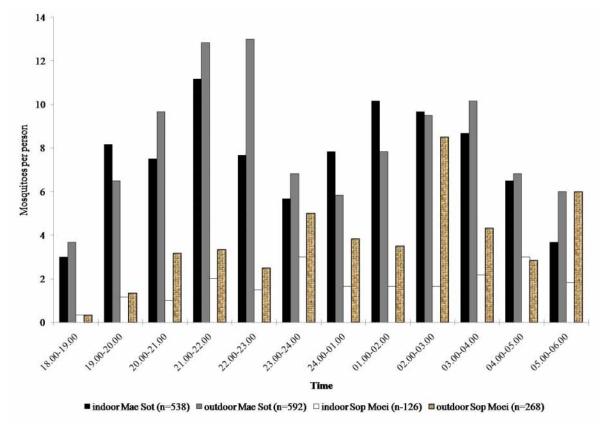


Figure 2. Mean indoor and outdoor human-landing rates by hour for *Anopheles minimus* in Mae Sot and Sop Moei.

Complex and related species, including Anopheles minimus, An. harrisoni, An. aconitus, An. varuna and An. pampanai (Sungvornyothin et al. 2006), 2) Dirus Complex, including Anopheles dirus, An. cracens, An. scanloni, An. baimaii and An. nemophilous (Walton et al. 1999), and 3) Maculatus Group, including Anopheles maculatus, An. sawadwongporni, An. pseudowillmori, An. dravidicus, An. rampae (former Form K) (Walton et al. 2007, Somboon et al. 2011).

Real Time PCR detection for *Plasmodium* species and *Wuchereria bancrofti* in mosquitoes

Real-time PCR was performed with a Roche LightCycler 480 (Software Version LCS480 1.5.0.39) using TaqMan reagents and hydrolysis probes for the detection of Plasmodium falciparum, P. vivax, and P. knowlesi, following a slightly modified methodology of Divis et al. (2010), and Wuchereria bancrofti immature stages using methods of Rao et al. (2006). Two reactions were separately performed including a first round of screening reaction for the detection of all Plasmodium species (Primers; Plasmo 1, Plasmo 2 and the Plasmodium screening probe) from Rougemont et al. (2004), Plasmodium vivax (Primers; VIV-F, VIV-R and probe VIV-PB) from Perandin et al. (2004) and Wuchereria bancrofti (Primers; LDR1-F, LDR2-R and probe WB-PB) from Rao et al. (2006). For the second round, a specific reaction was processed for the detection of P. falciparum (Primers; FAL-F, FAL-R and FAL probe) from Perandin et al. (2004) and P. knowlesi Primers; Plasmo 1 and 2 from Rougemont et al. (2004) and PK probe from Divis et al. (2010).

Data analysis

Numbers of primary malaria vectors collected by different collection methods were analyzed using non-parametric tests. A Wilcoxon test and Pearson's correlation analyses were used to investigate the interaction between the number of mosquitoes and environmental parameters. All data were analyzed with a level of significance set at 0.05% (p< 0.05), using a SPSS statistical package (ver. 17.0, SPSS, Chicago, IL).

RESULTS

Species composition and abundance

A total of 7,129 Anopheles females was collected during the two-year sampling period of February, 2011 to January, 2013. This sample included 22 species in the subgenus Cellia (6,156 specimens), and a single species and three groups in the subgenus Anopheles (n=973) (Table 1). Of 5,148 specimens within the two complexes and the Maculatus Group identified by molecular assays, 3,278 were derived from Mae Sot and 1,870 from Sop Moei field sites. The Minimus Complex and related species included An. minimus, An. aconitus and An. varuna; the Dirus Complex was composed of An. dirus and An. baimaii; and the Maculatus Group was represented by An. maculatus, An. sawadwongporni, An. pseudowillmori and An. dravidicus. Within the subgenus Cellia, the most abundant species in Mae Sot and Sop Moei was An. minimus accounting for 58.6% and 37.0% of all recorded anophelines, respectively

and 49.6% of the total sample from both sites combined (Table 1). Anopheles maculatus represented 25.4% and 20.4% of the total anopheline sample by site in Mae Sot and Sop Moei, respectively, and 23.3% of total for both sites. In both sites, An. dirus and An. baimaii were rarely collected, 0.5% and 0.6%, respectively in Mae Sot and 0.4% for both species in Sop Moei. The other Anopheles taxa in the subgenus Cellia included An. annularis s.l., An. culicifacies s.l., An. jamesii, An. jeyporiensis, An. karwari, An. kochi s.l., An. nivipes, An. philippinensis, An. splendidus, An. stephensi, An. subpictus, An. tessellatus and An. vagus, all of which were identified by morphological criteria only. In the subgenus Anopheles, a species belonging to the Anopheles hyrcanus group was the most prevalent with 46.7% and 45.0% collected in Mae Sot and Sop Moei, respectively and representing 45.6% of the total sample in the subgenus, followed by the Anopheles barbirostris Group (36.0% in Mae Sot, 21.9% in Sop Moei), for a total of 27% of collection for both sites. In addition, specimens of the Anopheles umbrosus group and Anopheles peditaeniatus were also found in both study sites.

Mosquito density and feeding behavior

The monthly density of An. minimus and two closely related species, An. aconitus and An. varuna, collected from the two sites during the two years of collection is shown in Table 2. Anopheles minimus was found throughout the year with the highest density during the 'summer' period, between February and April, in both sites and years. Comparatively, An. aconitus, and An. varuna were collected in much smaller densities in Mae Sot and Sop Moei, varying from 0.7% to 1.9% for An. aconitus, and from 2.7% to 1.2% for An. varuna, respectively (data not shown). Both species were highly zoophilic as all were collected on cattle-baited traps in Mae Sot, and more than 80% were attracted to cattle in Sop Moei. Indoor and outdoor feeding behaviors of *An. minimus* in Mae Sot were quite similar, with peak biting activity occurring between 21:00 and 23:00. In contrast, indoor and outdoor feeding behaviors of An. minimus from Sop Moei were markedly different, with the outdoor peak between 02:00 and 04:00 and indoor peak after 05:00 (Figure 2).

The monthly densities of An. dirus and An. baimaii collected from both study sites are shown in Table 2. Both species were found in similar proportions, with 18 and 21 specimens, respectively, in Mae Sot and 11 specimens for each species in Sop Moei. Nearly twice as many specimens were captured in Mae Sot (n=39) compared to Sop Moei (n=22). In both years, the greatest numbers of individuals were encountered in June. In Mae Sot, An. dirus and An. baimaii had a preference/attraction to humans with 97% (n=38) collected on human-bait compared to cattle. The indoor:outdoor (9:8) ratios were nearly identical, although the overall numbers were small, while An. baimaii was collected in greater number outdoors (n=16) than indoors (n=5) on humans. In Sop Moei, neither species showed any significant anthropophilic behavior. Both species were collected as frequently on humans (n=11) as on cattle (n=11). In Mae Sot, An. dirus presented an early peak of biting activity indoors (19:00-20:00) and a later peak outdoors (23:00-midnight)

Table 1. Numbers of *Anopheles* collected by species and percentage divided by subgenus in Mae Sot and Sop Moei from February, 2011 to January, 2013.

Species	Mae Sot (%)	Sop Moei (%)	Total (%)
Anopheles (Cellia)			
An. minimus	2,103 (58.6)	951 (37.0)	3,054 (49.6)
An. aconitus	16 (0.5)	19 (0.7)	35 (0.6)
An. varuna	60 (1.7)	12 (0.5)	72 (1.2)
An. dirus	18 (0.5)	11 (0.4)	29 (0.5)
An. baimaii	21 (0.6)	11 (0.4)	32 (0.5)
An. maculatus	911 (25.4)	525 (20.44)	1,436 (23.3)
An. sawadwongporni	116 (3.2)	274 (10.7)	390 (6.3)
An. pseudowillmori	4 (0.1)	51 (2.0)	55 (0.9)
An. dravidicus	29 (0.8)	16 (0.6)	45 (0.7)
An. annularis s.l.	-	8 (0.3)	8 (0.1)
An. culicifacies s.l.	2 (0.1)	83 (3.2)	85 (1.4)
An. jamesii	78 (2.2)	89 (3.5)	167 (2.7)
An. jeyporiensis	-	2 (0.1)	2 (0.0)
An. karwari	3 (0.1)	11 (0.4)	14 (0.2)
An. kochi s.l.	93 (2.6)	316 (12.3)	409 (6.6)
An. nivipes	9 (0.3)	6 (0.2)	15 (0.2)
An. philippinensis	72 (2.0)	65 (2.5)	137 (2.2)
An. splendidus	2 (0.1)	2 (0.1)	4 (0.1)
An. stephensi	1 (0.0)	1 (0.0)	2(0.0)
An. subpictus	1 (0.0)	9 (0.4)	10 (0.2)
An. tessellatus	-	38 (1.5)	38 (0.6)
An. vagus	47 (1.3)	70 (2.7)	117 (1.9)
Total (s.g. Cellia)	3,586 (58.3%)	2,570 (41.7%)	6,156
Anopheles (Anopheles)			
An. barbirostris Group	127 (36.0)	136 (21.9)	263 (27.0)
An. umbrosus Group	42 (11.8)	134 (21.6)	176 (18.1)
An. hyrcanus Group	165 (46.7)	279 (45.0)	444 (45.6)
An. peditaeniatus	19 (5.4)	71 (11.5)	90 (9.2)
Total (s.g. Anopheles)	353 (36.3%)	620 (63.7%)	973
Grand total	3,939 (55.3%)	3,190 (44.7%)	7,129

Table 2. Number, distribution and frequency of four primary Anopheles malaria vector species collected in Mae Sot and Sop Moei (I= indoor human landing, O=outdoor human landing, C= cattle baited).

						Ma	Mae Sot											Sop Moei	ſoei					
Month	An	An. minimus	nus	7	An.dirus	ns	A	An. baimaii	паії	An.	An.maculatus	atus	An.	An. minimus	snu	A	An.dirus	15	An.	An. baimaii	ii	An.m	An.maculatus	ns
	Ι	0	С	Ι	0	С	Ι	0	С	Ι	0	С	I	0	С	Ι	0	С	Ι	0	С	Ι	0	С
Year 1																								
Feb-11	134	128	207	1	1	1	1	ı	•	1	9	10	8	40	24	ı	ı	ı			ı		1	8
Apr-11	121	1111	220	-	-	1	1	1	ı	3	3	21	21	20	108	ı	1	1	ı	ı	,	ı	7	9
Jun-11	9	106	30	9	7	П	7	2	1	6	53	39	9	15	38	1	1	1	1	1	2	2	9	20
Aug-11	ı	5	7	ı	ı	ı	1	2		3	11	ı	ı	8	10	ı	ı	ı	ı	ı	ı	ı	ı	4
Oct-11	23	26	33	ı	ı	ı	1	ı		26	32	193	2	2	2	ı	1	1	ı	ı	ı	2	4	65
Dec-11	30	36	61	1	-	1	1	1		20	20	38	1	1	95	-	-	-	-	-	-	3	9	36
Year 2																								
Feb-12	41	31	1111	ı	1	1	1	ı	•	8	8	ı		49	117	ı	ı	ı		ı	ı		ı	ı
Apr-12	29	79	183	1	1	1	1	ı	1	4	4	ı	21	20	25	1	1	1	1	,		2	1	1
Jun-12	38	37	78	П	1	1	1	2	1	17	20	114	48	87	10	1	1	7	1	4	2	3	20	149
Aug-12	1	3	10	1	1	1	3	1	ı	2	25	42	1	6	2	ı	ı	ı	1	1	1	ı	7	27
Oct-12	16	9	21	П	1	1	1	4	1	23	45	ı	4	10	12	1	1	7	1	1	1	1	7	1
Dec-12	11	24	17	•	1	1	1	١	1	30	78	1	8	8	78						1	2	2	121
Total	538	592	973	6	8	1	5	16	1	149	305	457	126	268	557	2	3	9	1	5	5	14	75	436

*Summer= February, March, April, May; Wet=May, June, July, August, September, October; Dry= November, December, January.

(Figure 3). In Sop Moei, the numbers were too low to clearly define the precise biting pattern. For *An. baimaii*, outdoor and indoor collections showed nearly identical biting peaks from 20:00 to 22:00 hrs, with a second peak for outdoor collections (04:00-05:00). In Sop Moei, the biting (landing) peak activity was from 01:00 to 02:00 (Figure 4). Neither species was found in great abundance, so that drawing any definitive conclusions about these data should be done with caution.

Four species belonging to the Maculatus Group were identified in Mae Sot and Sop Moei: An. maculatus, An. sawadwongporni, An. pseudowillmori and An. dravidicus (Tables 1, 2). Collectively, the group was found throughout the year with the highest density during the rainy period between May and October in both sites. Anopheles maculatus was the most abundant and consistent species present in both sites (n=1,436) representing 74.6% of the species group, followed by An. sawadwongporni (n=390) representing 20.2%. Anopheles pseudowillmori and An. dravidicus, were captured in much smaller proportions, 2.9% and 2.3% of total collections, respectively, and having periods (months) of little or no captures recorded. In general, all Maculatus Group members were collected in greater numbers from cow than human, except for An. sawadwongporni in Mae Sot and An. pseudowillmori in Sop Moei where higher numbers in outdoor HLC were seen (data not shown). The activity of An. maculatus in Mae Sot showed a very clear early biting peak between 18:00 and 21:00 for outdoor HLC (Figure 5). The biting activity of An. sawadwongporni from outdoor HLC in both villages showed a small peak between 19:00 to 20:00 (data not shown). Like the Dirus Complex, neither species was found in great abundance, so drawing any definitive conclusions regarding these data should be done with caution.

Non-parametric analysis of densities of *Anopheles minimus* and *An. maculatus* in different collection sites between the numbers of mosquitoes collected from indoor/outdoor human-baited collections and environmental data (temperature, relative humidity (RH) and rainfall) found no statistical association except for *An. maculatus* in Sop Moei regards adult densities and variations in mean ambient temperature.

Trophic behavior

The non-parametric Wilcoxon analysis was used to compare the hourly landing rates of the two most abundant mosquito species, An. minimus and An. maculatus, using different collection methods and study locations. For An. minimus, no significant differences (Z=-0.94, p=0.348) were observed in Mae Sot between the indoor (3.76±0.3) and outdoor (4.11±0.41) human landing. However, An. minimus showed significantly greater (Z=-2.43, p=0.02) anthropophilic (7.85±0.72) behavior compared to its attraction to cattle (6.76±0.72) (Table 5). In Sop Moei, An. minimus was significantly more exophagic (Z=-5.72, p<0.01) and zoophilic (Z=-3.99, p<0.01) (Table 5). An. maculatus showed similar and highly significant exophagic and zoophilic behavior in both study sites.

Parasite detection

Eight Anopheles species (n=1,447) from both collection sites in Mae Sot and Sop Moei were examined for parasite infection rates as follows: An. minimus (1,090, 132), An. aconitus (16, 10), An. dirus (18, 10), An. baimaii (14, 19), An. maculatus (55, 25), An. sawadwongporni (2, 39), An. pseudowillmori (2, 39) and An. dravidicus (5, 1), respectively. Only one specimen, An. minimus from Mae Sot in an outdoor HLC, 00:00-01:00 in April, 2011, was found positive for P. vivax.

DISCUSSION

This study is the first detailed description of the diversity of *Anopheles* species based on a molecular identification in the two most malaria endemic areas of Thailand, Mae Sot District in Tak Province and Sop Moei District in Mae Hong Son Province, both located in the northwestern part of the country that borders Myanmar. Results from this study provide accurate species identification as well as seasonal abundance and trophic behavior of *Anopheles minimus* and *Anopheles maculatus*, among the most important malaria vectors in northwestern Thailand.

In this study, six of the most important (primary and secondary) malaria vector species in Thailand were collected. The most abundant species in Mae Sot and Sop Moei was An. minimus (42.8%), followed by An. maculatus (20.1%) and An. sawadwongporni (5.5%). Collectively, these species are responsible to varying degrees for maintaining malaria transmission in both areas which have been listed as 'A1' areas by the Thai Ministry of Public Health, areas where malaria occurs at least 6 months of the year and longer (Chareonviriyaphap et al. 2001, BVBD 2013).

Anopheles minimus represented 43% of the total Anopheles mosquitoes recorded from both areas during the two-year collection period (53.4% in Mae Sot and 29.8% of all anophelines in Sop Moei. Anopheles maculatus and An. sawadwongporni were found in greater numbers compared to An. pseudowillmori and An. dravidicus, all species within the Maculatus Group. These findings are consistent with previous observations in Mae Sot District (Tum Sua Village) that found An. minimus to be the most abundant species (71%), followed by An. maculatus Group (28%) and few specimens of the An. dirus Complex (Tisgratog et al. 2012). Anopheles dirus and An. baimaii, closely related and the most important forestrelated malaria vectors, were found in very low densities with only a total of 39 specimens collected in Mae Sot and 22 in Sop Moei during a two-year period. These two species were slightly more common during the June collection both years with only a few specimens detected other months. Low collection numbers of the Dirus Complex complicate drawing any definitive conclusions on host-seeking behavior in this study.

In Thailand, the biting activity and behavior of specific malaria vectors, based on reliable molecular identification methods, has been studied in detail in relatively fewer instances compared to earlier studies that based all identification on morphological characters alone. For example, two species

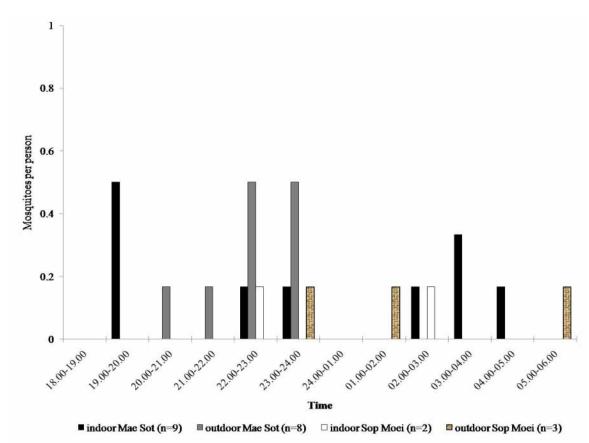


Figure 3. Mean indoor and outdoor human-landing rates by hour for *Anopheles dirus* in Mae Sot and Sop Moei.

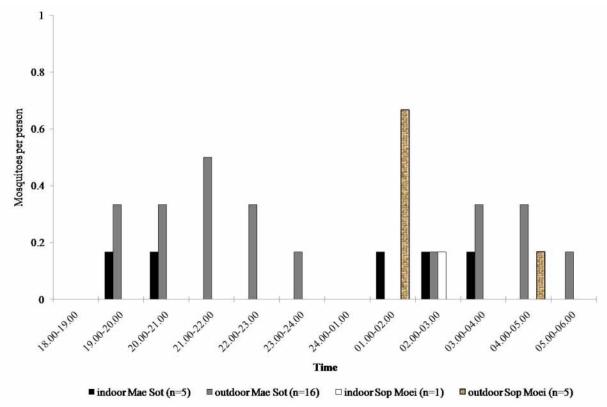
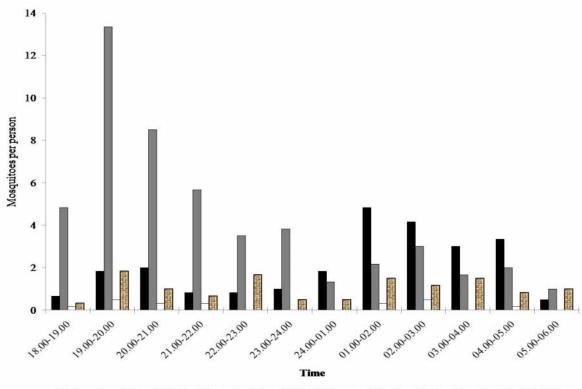


Figure 4. Mean indoor and outdoor human-landing rates by hour for *Anopheles baimaii* by hour in Mae Sot and Sop Moei.



■ indoor Mae Sot (n=149) ■ outdoor Mae Sot (n=305) □ indoor Sop Moei (n=14) ■ outdoor Sop Moei (n=75)

Figure 5. Mean indoor and outdoor human-landing rates by hour for *Anopheles maculatus* in Mae Sot and Sop Moei.

within the Minimus Complex, An. minimus and An. harrisoni were studied from western Thailand using a molecular identification assay (Sungvornyothin et al. 2006), and more recently, Kongmee et al. (2012) characterized the seasonal abundance, distribution and breeding sites of An. minimus and An. harrisoni from the same site in Sai Yok District, Kanchanaburi Province in western Thailand. Species diversity, biting activity and trophic behavior of the Minimus Complex were also observed previously in Mae Sot District (Tisgratog et al. 2012) where the two closely related species, An. minimus and An. aconitus were collected. In this study, An. varuna, another genetically related species to An. minimus, was identified for the first time in both locations. Although An. aconitus and An. varuna are not as efficient vectors compared to An. minimus, the presence of these two species, especially An. aconitus, in Mae Sot and Sop Moei is of importance as these two species could possibly play a secondary role in malaria transmission (Gould et al. 1967, Scanlon et al. 1968, Junkum et al. 2007).

Tak Province is one of the most malaria-endemic areas of Thailand, accounting for around 23% of total malaria cases in the country in 2012 (5,199 cases) and 2013 (4,977 cases) (BVBD 2013). Six important malaria vectors were identified in the study area using AS-PCR molecular assays. Our findings on the blood-feeding habits of *An. minimus* in Mae Sot showed both zoophilic and anthropophilic behaviors with no strong preference for one host over the other which is similar to previous findings elsewhere in Mae Sot and Kanchanaburi (Sungvornyothin et al. 2006, Tananchai et al. 2012, Tisgratog

et al. 2012). In Sop Moei, *An. minimus* demonstrated a stronger zoophilic tendency as more mosquitoes feed on cattle located outside houses. Generally, zoophilic behaviors are considered less conductive for efficient malaria transmission as other animal hosts serve as alternative blood sources, thereby reducing direct contact between vector and human, and thus overall vector capacity. In Pu Teuy, Kanchanaburi Province, *An. minimus* and *An. harrisoni* are present, the latter species being predominant and with generally distinct zoophilic behavior that is believed partly responsible for the typically low levels of malaria transmission in this area (Sungvornyothin et al. 2006). *Anopheles minimus*, *An. dirus* and *An. baimaii*, found in both study areas demonstrated exophagic and zoophilic behaviors which are in agreement with previous observations by Tananchai et al. (2012).

Few studies on species of the Maculatus Group in Thailand have been published (Takai et al. 1987, Upatham et al. 1988, Green et al. 1992, Rongnoparut et al. 1996, 1999, Walton et al. 2007, Muenworn et al. 2009). Studies on this group have been hampered by the lack of reliable tools to identify cryptic species, resulting in misidentification. From our study, four species within this group, An. maculatus, An. sawadwongporni, An. pseudowillmori and An. dravidicus were identified molecularly. The most common representative was An. maculatus, followed by An. sawadwongporni, both malaria vectors in the southern Thailand (Rattanarithikul et al. 1996b, Sinka et al. 2011). Our findings showed that these two important putative vectors were present in study locations in fair proportions, 20.1% and 5.5%, respectively,

compared to all other anophelines identified. The biology of these two malaria vectors, including trophic behavior, infection rate, and parasite susceptibility (vector competence) deserves further study to determine their respective role, if any, in malaria transmission in northwestern Thailand.

Only one mosquito positive for the malaria parasite was detected during the dry season, accounting for a 0.092% (1/1,090) infection rate for An. minimus. This mosquito was collected from outdoor HLC from Mae Sot in the early part of the study (April 2011). There was no evidence of malaria infection with P. falciparum, P. malariae, P. knowlesi or the filarial parasite, W. bancrofti. The one positive An. minimus was captured during the normal malaria peak in Mae Sot in the late summer months (April-May). The explanation for the low infection rate in mosquitoes tested is unclear, but the probability of finding an infection will have naturally decreased as the overall malaria incidence has been declining year to year in both provinces and where the study took place. For instance, there was a slight decline in reported malaria cases from 2012 to 2013 in Tak Province (0.98 to 0.93 per 1,000 population) and a 38% decline in Mae Hong Son Province (0.52 to 0.32/1000) (BVBD 2013).

It is clear that environmental factors directly influence the distribution and behavior of the malaria vectors (Manguin et al. 2008). For example, the An. dirus complex typically occurs in native forest type habitats but also has an ability to adapt to changing environmental conditions that allow it to successfully invade cultivated areas that simulate favored natural forest conditions. Each species within the Dirus Complex is dependent on forest cover (shading) to some degree, and commonly occupies forest-fringe areas where it can interact with human populations to transmit malaria parasites. Habitats include natural dense forest (An. dirus), rubber plantations (An. dirus and An. baimaii) and secondary forest (An. latens) (Gingrich et al. 1990, Rosenberg et al. 1990, Suwonkerd et al. 2002). Vanwambeke et al. (2007a,b) found that land-use changes could selectively influence the diversity and likely density of mosquito species, thus having a direct impact on pathogen transmission.

Deforestation, the result of a wide variety of human including land clearing for agricultural development, logging, population resettlement programs, road construction, mining and hydropower development, is one of the most potent factors either promoting or reducing infectious diseases like malaria and dengue in the southeast Asian region, including Thailand (Guerra et al. 2006, Vanwambeke et al. 2007a,b, Yasuoka and Levins 2007). Dramatic changes in environmental conditions are the direct result of modified land use, such as conversion of rice fields to rubber plantations or forest to urbanized zones. The extensive clearing of native and secondary forests has had enormous impacts on local ecosystems, in particular the critical microclimates of mosquitoes by reducing shade cover, changing the humidity regimen, and altering rainfall patterns (Reiter 2001, Overgaard et al. 2002). For example, anopheline species that prefer to use shaded water bodies for oviposition, deforestation can reduce dramatically breeding habitats, thus affecting their propagation (Overgaard et al. 2002). In the future, the inevitable developmental changes that will occur to the landscape of northwestern Thailand will need to be monitored carefully to assess their impacts on potential malaria transmission.

Vector control remains an important component of successful integrated malaria control programs. As demonstrated in this study, a better understanding of the bionomics of a specific vector species, its vector capacity, and epidemiological importance, hinges on the accurate identification of sympatric sibling species in a given area. More investigations are needed in all remaining malaria endemic areas in Thailand so as to develop more cost-effective and targeted vector control strategies based on evidence derived from well-design field studies.

Acknowledgments

This study was supported by the Thailand Research Fund Organization through the Thai grant of the Royal Golden Jubilee Ph.D. Program (PHD/0156/2552), the Senior Research Scholar Program (RTA 5580002), Center of Excellence on Agricultural Biotechnology, Science and Technology Postgraduate Education and Research Development Office, Office of Higher Education Commission, Ministry of Education (AG-BIO/PERDO-CHE), The Capacity Building for KU Students on Internationalization Program, Kasetsart University, the French Government's contribution to the Royal Golden Jubilee Project for Year 2011, and Institut de Recherche pour le Développement (Baird), UMR-MD3, France for molecular training and laboratory expenses.

REFERENCES CITED

Apiwathnasor, C., S. Prommongkol, Y. Samung, D. Limrat, and B. Rojruthai. 2002. Potential for *Anopheles campestris* (Diptera: Culicidae) to transmit malaria parasites in Pa Rai subdistrict (Aranyaprathet, Sa Kaeo Province), Thailand. J. Med. Entomol. 39: 583-586.

Baimai, V., C.A. Green, R.G. Andre, B.A. Harrison, and E.L. Peyton. 1984. Cytogenetic studies of some species complexes of *Anopheles* in Thailand and Southeast Asia. *Southeast* Asian J. Trop. Med. Publ. Hlth. 15: 536-456.

Baimai, V., R.E. Harbach, and U. Kijchalao. 1988. Cytogenetic evidence for a fifth species within the taxon *Anopheles dirus* in Thailand. J. Am. Mosq. Contr. Assoc. 4: 333-338.

Baimai, V. 1989. Speciation and species complexes of the *Anopheles* malaria vectors in Thailand, pp. 146-162, Third Conference on Malaria Research. Malaria Division, Department of Communicable Disease Control, Ministry of Public Health, Thailand.

Bhumiratana, A., B. Wattanakull, S. Koyadun, S. Suvannadabba, J. Rojanapremsuk, and W. Tantiwattanasup. 2002. Relationship between male hydrocele and infection prevalences in clustered communities with uncertain transmission of *Wuchereria bancrofti* on the Thailand-Myanmar border. *Southeast* Asian J. Trop. Med. Publ. Hlth. 33: 7-17.

Bhumiratana, A., S. Koyadun, M. Srisuphanunt, P.

- Satitvipawee, N. Limpairojn, and G. Gaewchaiyo. 2005. Border and imported bancroftian filariases: baseline seroprevalence in sentinel populations exposed to infections with *Wuchereria bancrofti* and concomitant HIV at the start of diethylcarbamazine mass treatment in Thailand. *Southeast* Asian J. Trop. Med. Publ. Hlth. 36: 390-407.
- Bhumiratana, A., A. Intarapuk, P. Sorosjinda-Nunthawarasilp, P. Maneekan, and S. Koyadun. 2013. Border malaria associated with multidrug resistance on Thailand-Myanmar and Thailand-Cambodia borders: transmission dynamic, vulnerability, and surveillance. Biomed. Res. Int. 2013: 363417.
- BVBD. 2013. Annual report 2013 pp. 143, Malaria situation 2013. Bureau of Vector Borne Disease, Department of Disease Control, Ministry of Public Health, Nonthaburi.
- Chareonviriyaphap, T., S. Sungvornyothin, S. Ratanatham, and A. Prabaripai. 2001. Insecticide-induced behavioral responses of *Anopheles minimus*, a malaria vector in Thailand. J. Am. Mosq. Contr. Assoc. 17: 13-22.
- Chareonviriyaphap, T., A. Prabaripai, M. J. Bangs, and B. Aum-Aung. 2003. Seasonal abundance and blood feeding activity of *Anopheles minimus* Theobald (Diptera: Culicidae) in Thailand. J. Med. Entomol. 40: 876-881.
- Corbel, V., F. Nosten, K. Thanispong, C. Luxemburger, M. Kongmee, and T. Chareonviriyaphap. 2013. Challenges and prospects for dengue and malaria control in Thailand, Southeast Asia. Trends Parasitol. 29: 623-633.
- Divis, P.C., S.E. Shokoples, B. Singh, and S.K. Yanow. 2010. A TaqMan real-time PCR assay for the detection and quantitation of *Plasmodium knowlesi*. Malar. J. 9: 344.
- Dusfour, I., J.R. Michaux, R.E. Harbach, and S. Manguin. 2007. Speciation and phylogeography of the Southeast Asian *Anopheles sundaicus* complex. Infect. Genet. Evol. 7: 484-493.
- Eamkum, P., S. Sungvornyothin, O. Kritpetcharat, J. Daduang, U. Lek-Uthai, L. Charerntanyarak, and P. Kritpetcharat. 2014. A single-round multiplex PCR assay for the identification of *Anopheles minimus* related species infected with *Plasmodium falciparum* and *Plasmodium vivax*. Parasitol. Int. 63: 442-449.
- Garros, C., L. L. Koekemoer, M. Coetzee, M. Coosemans, and S. Manguin. 2004. A single multiplex assay to identify major malaria vectors within the African *Anopheles* funestus and the Oriental *An. minimus* groups. Am. J. Trop. Med. Hyg. 70: 583-590.
- Gingrich, J. B., A. Weatherhead, J. Sattabongkot, C. Pilakasiri, and R. A. Wirtz. 1990. Hyperendemic malaria in a Thai village: dependence of year-round transmission on focal and seasonally circumscribed mosquito (Diptera: Culicidae) habitats. J. Med. Entomol. 27: 1016-1026.
- Gould, D. J., S. Esah, and U. Pranith. 1967. Relation of *Anopheles aconitus* to malaria transmission in the central plain of Thailand. Trans. R. Soc. Trop. Med. Hyg. 61: 441-442
- Green, C. A., R. F. Gass, L. E. Munstermann, and V. Baimai. 1990. Population-genetic evidence for two species in *Anopheles minimus* in Thailand. Med. Vet. Entomol. 4:

- 25-34.
- Green, C. A., R. R., P. S., S. P., and B. V. 1991. A newly-recognized vector of human malarial parasites in the Oriental region, *Anopheles (Cellia) pseudowillmori* (Theobald, 1910). Trans. R. Soc. Trop. Med. Hyg. 85: 35-36
- Green, C. A., L. E. Munstermann, S. G. Tan, S. Panyim, and V. Baimai. 1992. Population genetic evidence for species A, B, C and D of the *Anopheles dirus* complex in Thailand and enzyme electromorphs for their identification. Med. Vet. Entomol. 6: 29-36.
- Guerra, C. A., R. W. Snow, and S. I. Hay. 2006. A global assessment of closed forests, deforestation and malaria risk. Ann. Trop. Med. Parasitol. 100: 189-204.
- Harbach, R. E., V. Baimai, and S. Sukowati. 1987. Some observations on sympatric populations of the malaria vectors Anopheles leucosphyrus and Anopheles balabacensis in a village-forest setting in South Kalimantan. Southeast Asian J. Trop. Med. Publ. Hlth. 18: 241-247.
- Ismail, I. A., V. Notananda, and J. Schepens. 1974. Studies on malaria and responses of Anopheles balabacensis balabacensis and Anopheles minimus to DDT residual spraying in Thailand I. Pre-spraying observations. Acta. Trop. 31: 129-164.
- Jitpakdi, A., W. Choochote, P. Panart, B. Tookyang, K. Panart, and S. Prajakwong. 1998. Possible transmission of two types of Wuchereria bancrofti in Muang District, Chiang Mai, northern Thailand. Southeast Asian J. Trop. Med. Publ. Hlth. 29: 141-143.
- Junkum, A., B. Pitasawat, B. Tuetun, A. Saeung, E. Rattanachanpichai, N. Jariyapan, N. Komalamisra, M. Mogi, U. Chaithong, and W. Choochote. 2007. Seasonal abundance and biting activity of Anopheles aconitus (Diptera: Culicidae) in Chiang Mai, northern Thailand. Southeast Asian J. Trop. Med. Publ. Hlth. 38: 215-223.
- Khamboonruang, C., P. Thitasut, S. Pan-In, N. Morakote, W. Choochote, P. Somboon, and P. Keha. 1987. Filariasis in Tak Province, northwest Thailand: the presence of subperiodic variant Wuchereria bancrofti. Southeast Asian J. Trop. Med. Publ. Hlth. 18: 218-222.
- Kongmee, M., N. L. Achee, K. Lerdthusnee, M. J. Bangs, S. Chowpongpang, A. Prabaripai, and T. Charoenviriyaphap. 2012. Seasonal abundance and distribution of Anopheles larvae in a riparian malaria endemic area of western Thailand. Southeast Asian J. Trop. Med. Publ. Hlth. 43: 601-613.
- Linton, Y. M., R. E. Harbach, S. Chang Moh, T. G. Anthony, and A. Matusop. 2001. Morphological and molecular identity of Anopheles (Cellia) sundaicus (Diptera: Culicidae), the nominotypical member of a malaria vector species complex in Southeast Asia. Syst. Entomol. 26: 357-366.
- Manguin, S., P. Kengne, L. Sonnier, R. E. Harbach, V. Baimai, H. D. Trung, and M. Coosemans. 2002. SCAR markers and multiplex PCR-based identification of isomorphic species in the *Anopheles dirus* complex in Southeast Asia. Med. Vet. Entomol. 16: 46-54.

- Manguin, S., C. Garros, I. Dusfour, R.E. Harbach, and M. Coosemans. 2008. Bionomics, taxonomy, and distribution of the major malaria vector taxa of *Anopheles* subgenus *Cellia* in Southeast Asia: an updated review. Infect. Genet. Evol. 8: 489-503.
- Muenworn, V., S. Sungvornyothin, M. Kongmee, S. Polsomboon, M.J. Bangs, P. Akrathanakul, S. Tanasinchayakul, A. Prabaripai, and T. Chareonviriyaphap. 2009. Biting activity and host preference of the malaria vectors *Anopheles maculatus* and *Anopheles sawadwongporni* (Diptera: Culicidae) in Thailand. J. Vector Ecol. 34: 62-69.
- Overgaard, H., Y. Tsuda, W. Suwonkerd, and M. Takagi. 2002. Characteristics of *Anopheles minimus* (Diptera: Culicidae) larval habitat in northern Thailand. Environ. Entomol. 31: 134-141.
- Perandin, F., N. Manca, A. Calderaro, G. Piccolo, L. Galati, L. Ricci, M.C. Medici, M.C. Arcangeletti, G. Snounou, G. Dettori, and C. Chezzi. 2004. Development of a real-time PCR assay for detection of *Plasmodium falciparum*, *Plasmodium vivax*, and *Plasmodium ovale* for routine clinical diagnosis. J. Clin. Microbiol. 42: 1214-1219.
- Poolprasert, P., S. Manguin, M.J. Bangs, S. Sukhontabhirom, S. Poolsomboon, P. Akaratanakul, and T. Chareonviriyaphap. 2008. Genetic structure and gene flow of *Anopheles minimus* and *Anopheles harrisoni* in Kanchanaburi Province, Thailand. J. Vector Ecol. 33: 158-165.
- Poopittayasataporn, A. and V. Baimai. 1995. Polytene chromosome relationships of five species of the *Anopheles dirus* complex in Thailand. Genome. 38: 426-434.
- Pothikasikorn, J., M.J. Bangs, R. Boonplueang, and T. Chareonviriyaphap. 2008. Susceptibility of various mosquitoes of Thailand to nocturnal subperiodic *Wuchereria bancrofti*. J. Vector Ecol. 33: 313-320.
- Rao, R.U., L.J. Atkinson, R.M. Ramzy, H. Helmy, H.A. Farid, M.J. Bockarie, M. Susapu, S. J. Laney, S.A. Williams, and G.J. Weil. 2006. A real-time PCR-based assay for detection of *Wuchereria bancrofti* DNA in blood and mosquitoes. Am. J. Trop. Med. Hyg. 74: 826-832.
- Ratanatham, S., E.S. Upatham, C. Prasittisuk, W. Rojanasunan, N. Theerasilp, A. Tremongkol, and V. Viyanant. 1988. Bionomics of *Anopheles minimus* and its role in malaria transmission in Thailand. Southeast Asian J. Trop. Med. Publ. Hlth. 19: 283-289.
- Rattanarithikul, R., K.J. Linthicum, and E. Konishi. 1996a. Seasonal abundance and parity rates of *Anopheles* species in southern Thailand. Am. Mosq. Contr. Assoc. 12: 75-83.
- Rattanarithikul, R., E. Konishi, and K.J. Linthicum. 1996b. Observations on the nocturnal biting activity and host preference of anophelines collected in southern Thailand. J. Am. Mosq. Contr. Assoc. 12: 52-57.
- Rattanarithikul, R., B.A. Harrison, R.E. Harbach, P. Panthusiri, R.E. Coleman, and P. Panthusiri. 2006. Illustrated keys to the mosquitoes of Thailand. IV. *Anopheles*. Southeast Asian J. Trop. Med. Publ. Hlth. 37 Suppl 2: 1-128.
- Reiter, P. 2001. Climate change and mosquito-borne disease.

- Environ. Hlth. Perspect. 109 Suppl 1: 141-161.
- Rongnoparut, P., S. Yaicharoen, N. Sirichotpakorn, R. Rattanarithikul, G.C. Lanzaro, and K.J. Linthicum. 1996. Microsatellite polymorphism in *Anopheles maculatus*, a malaria vector in Thailand. Am. J. Trop. Med. Hyg. 55: 589-594.
- Rongnoparut, P., N. Sirichotpakorn, R. Rattanarithikul, S. Yaicharoen, and K.J. Linthicum. 1999. Estimates of gene flow among *Anopheles maculatus* populations in Thailand using microsatellite analysis. Am. J. Trop. Med. Hyg. 60: 508-515.
- Rosenberg, R., R.G. Andre, and L. Somchit. 1990. Highly efficient dry season transmission of malaria in Thailand. Trans. R. Soc. Trop. Med. Hyg. 84: 22-28.
- Rougemont, M., M. Van Saanen, R. Sahli, H.P. Hinrikson, J. Bille, and K. Jaton. 2004. Detection of four *Plasmodium* species in blood from humans by 18S rRNA gene subunit-based and species-specific real-time PCR assays. J. Clin. Microbiol. 42: 5636-5643.
- Saeung, A., V. Baimai, Y. Otsuka, R. Rattanarithikul, P. Somboon, A. Junkum, B. Tuetun, H. Takaoka, and W. Choochote. 2008. Molecular and cytogenetic evidence of three sibling species of the *Anopheles barbirostris* Form A (Diptera:Culicidae) in Thailand. Parasitol. Res. 102: 499-507.
- Saeung, A. 2012. *Anopheles* (Diptera:Culicidae) species complex in Thailand: identification, distribution, bionomics and malaria-vector importance. Int. J. Parasitol. 4: 75-82.
- Scanlon, J.E., E.L. Peyton, and D.J. Gould. 1968. An annotated checklist of the *Anopheles* of Thailand. Thai Natl. Sci. Pap. Fauna Ser. 2: 1-35.
- Sinka, M.E., M.J. Bangs, S. Manguin, T. Chareonviriyaphap, A.P. Patil, W.H. Temperley, P.W. Gething, I.R. Elyazar, C.W. Kabaria, R.E. Harbach, and S.I. Hay. 2011. The dominant *Anopheles* vectors of human malaria in the Asia-Pacific region: occurrence data, distribution maps and bionomic precis. Parasit. Vectors 4: 89.
- Somboon, P., D. Thongwat, and R.E. Harbach. 2011. *Anopheles (Cellia) rampae* n. sp., alias chromosomal form K of the Oriental Maculatus Group (Diptera: Culicidae) in Southeast Asia. Zootaxa 2810: 47–55.
- Sungvornyothin, S., V. Muenvorn, C. Garros, S. Manguin, A. Prabaripai, M.J. Bangs, and T. Chareonviriyaphap. 2006. Trophic behavior and biting activity of the two sibling species of the *Anopheles minimus* complex in western Thailand. J. Vector Ecol. 31: 252-261.
- Suwonkerd, W., H.J. Overgaard, Y. Tsuda, P.S., and M. Takagi. 2002. Malaria vector densities in transmission and non-transmission areas during 23 years and land use in Chiang Mai province, Northern Thailand. Basic Appl. Ecol. 3: 197-207.
- Takai, K., T. Kanda, K. Ogawa, and S. Sucharit. 1987. Morphological differentiation in *Anopheles maculatus* of Thailand accompanied with genetical divergence assessed by hybridization. J. Am. Mosq. Contr. Assoc. 3: 148-153.
- Takken, W. and N.O. Verhulst. 2013. Host preferences of

- blood-feeding mosquitoes. Annu. Rev. Entomol. 58: 433-453.
- Tananchai, C., R. Tisgratog, W. Juntarajumnong, J.P. Grieco, S. Manguin, A. Prabaripai, and T. Chareonviriyaphap. 2012. Species diversity and biting activity of *Anopheles dirus* and *Anopheles baimaii* (Diptera: Culicidae) in a malaria prone area of western Thailand. Parasit. Vectors 5: 211.
- Thongsahuan, S., V. Baimai, Y. Otsuka, A. Saeung, B. Tuetun, N. Jariyapan, S. Suwannamit, P. Somboon, A. Jitpakdi, H. Takaoka, and W. Choochote. 2009. Karyotypic variation and geographic distribution of *Anopheles campestris*-like (Diptera: Culicidae) in Thailand. Mem. Inst. Oswaldo Cruz. 104: 558-566.
- Tisgratog, R., C. Tananchai, W. Juntarajumnong, S. Tuntakom, M.J. Bangs, V. Corbel, and T. Chareonviriyaphap. 2012. Host feeding patterns and preference of *Anopheles minimus* (Diptera: Culicidae) in a malaria endemic area of western Thailand: baseline site description. Parasit. Vectors 5: 114.
- Upatham, E.S., C. Prasittisuk, S. Ratanatham, C.A. Green, W. Rojanasunan, P. Setakana, N. Theerasilp, A. Tremongkol, V. Viyanant, S. Pantuwatana, and R.G. Andre. 1988. Bionomics of *Anopheles maculatus* complex and their role in malaria transmission in Thailand. Southeast Asian J. Trop. Med. Publ. Hlth. 19: 259-269.

- Vanwambeke, S.O., P. Somboon, R.E. Harbach, M. Isenstadt, E.F. Lambin, C. Walton, and R.K. Butlin. 2007a. Landscape and land cover factors influence the presence of Aedes and *Anopheles* larvae. J. Med. Entomol. 44: 133-144.
- Vanwambeke, S.O., E.F. Lambin, M.P. Eichhorn, S.P. Flasse, R.E. Harbach, L. Oskam, P. Somboon, S.V. Beers, B.H.B. Van Benthem, C. Walton, and R.K. Butlin. 2007b. Impact of land-use change on dengue and malaria in northern Thailand. Eco. Hlth. 4: 37–51.
- Walton, C., J.M. Handley, C. Kuvangkadilok, F.H. Collins, R.E. Harbach, V. Baimai, and R.K. Butlin. 1999. Identification of five species of the *Anopheles dirus* complex from Thailand, using allele-specific polymerase chain reaction. Med. Vet. Entomol. 13: 24-32.
- Walton, C., P. Somboon, S.M. O'Loughlin, S. Zhang, R.E. Harbach, Y.M. Linton, B. Chen, K. Nolan, S. Duong, M.Y. Fong, I. Vythilingum, Z.D. Mohammed, H.D. Trung, and R.K. Butlin. 2007. Genetic diversity and molecular identification of mosquito species in the *Anopheles maculatus* group using the ITS2 region of rDNA. Infect. Genet. Evol. 7: 93-102.
- Yasuoka, J., and R. Levins. 2007. Impact of deforestation and agricultural development on anopheline ecology and malaria epidemiology. Am. J. Trop. Med. Hyg. 76: 450-460.



Diversity of *Anopheles* Species and Trophic Behavior of Putative Malaria Vectors in Two Malaria Endemic Areas of Northwestern Thailand

Author(s): Krajana Tainchum, Wanapa Ritthison, Thipwara Chuaycharoensuk, Michael J. Bangs, Sylvie

Manguin, and Theeraphap Chareonviriyaphap

Source: Journal of Vector Ecology, 39(2):424-436. 2014.

Published By: Society for Vector Ecology

URL: http://www.bioone.org/doi/full/10.3376/i1081-1710-39-424

BioOne (<u>www.bioone.org</u>) is a nonprofit, online aggregation of core research in the biological, ecological, and environmental sciences. BioOne provides a sustainable online platform for over 170 journals and books published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Web site, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/page/terms_of_use.

Usage of BioOne content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.