ICES Journal of Marine Science

ICES Journal of Marine Science (2020), 77(7-8), 2411-2422. doi:10.1093/icesjms/fsaa094

Food for Thought

The challenges of detecting and attributing ocean acidification impacts on marine ecosystems

Steve S. Doo (D^{1,2*}, Andrea Kealoha (D^{3,4}, Andreas Andersson (D⁵, Anne L. Cohen (D⁶, Tacey L. Hicks (D³, Zackary I. Johnson (D⁷, Matthew H. Long (D⁸, Paul McElhany (D⁹, Nathaniel Mollica (D⁶, Kathryn E. F. Shamberger (D³, Nyssa J. Silbiger (D¹, Yuichiro Takeshita (D¹⁰, and D. Shallin Busch (D¹¹)

¹Department of Biology, California State University, Northridge, CA, USA

²Geoecology and Carbonate Sedimentology Group, Leibniz Centre for Tropical Marine Research, Bremen, Germany

³Department of Oceanography, Texas A&M University, College Station, TX, USA

⁴Department of Science, Technology, Engineering and Mathematics, University of Hawai'i, Maui College, HI, USA

⁵Scripps Institution of Oceanography, University of California San Diego, La Jolla, CA, USA

⁶Geology and Geophysics Department, Woods Hole Oceanographic Institution, Woods Hole, MA, USA

⁷Marine Laboratory and Biology Department, Duke University, Beaufort, NC, USA

⁸Marine Chemistry and Geochemistry Department, Woods Hole Oceanographic Institution, Woods Hole, MA, USA

⁹Conservation Biology Division, Northwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Mukilteo, WA, USA

¹⁰Monterey Bay Aquarium Research Institute, Moss Landing, CA, USA

¹¹Ocean Acidification Program, Oceanic and Atmospheric Research and Conservation Biology Division, Northwest Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, Seattle, WA, USA

*Corresponding author: tel: +49 421 238 00178; e-mail: steve.doo@leibniz-zmt.de.

Doo, S. S., Kealoha, A., Andersson, A., Cohen, A. L., Hicks, T. L., Johnson, Z. I., Long, M. H., McElhany, P., Mollica, N., Shamberger, K. E. F., Silbiger, N. J., Takeshita, Y., and Busch, D. S. The challenges of detecting and attributing ocean acidification impacts on marine ecosystems. – ICES Journal of Marine Science, 77: 2411–2422.

Received 12 December 2019; revised 9 May 2020; accepted 11 May 2020; advance access publication 9 August 2020.

A substantial body of research now exists demonstrating sensitivities of marine organisms to ocean acidification (OA) in laboratory settings. However, corresponding *in situ* observations of marine species or ecosystem changes that can be unequivocally attributed to anthropogenic OA are limited. Challenges remain in detecting and attributing OA effects in nature, in part because multiple environmental changes are cooccurring with OA, all of which have the potential to influence marine ecosystem responses. Furthermore, the change in ocean pH since the industrial revolution is small relative to the natural variability within many systems, making it difficult to detect, and in some cases, has yet to cross physiological thresholds. The small number of studies that clearly document OA impacts in nature cannot be interpreted as a lack of

© International Council for the Exploration of the Sea 2020.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/ licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

International Council for the Exploration of the Sea larger-scale attributable impacts at the present time or in the future but highlights the need for innovative research approaches and analyses. We summarize the general findings in four relatively well-studied marine groups (seagrasses, pteropods, oysters, and coral reefs) and integrate overarching themes to highlight the challenges involved in detecting and attributing the effects of OA in natural environments. We then discuss four potential strategies to better evaluate and attribute OA impacts on species and ecosystems. First, we highlight the need for work quantifying the anthropogenic input of CO_2 in coastal and open-ocean waters to understand how this increase in CO_2 interacts with other physical and chemical factors to drive organismal conditions. Second, understanding OA-induced changes in population-level demography, potentially increased sensitivities in certain life stages, and how these effects scale to ecosystem-level processes (e.g. community metabolism) will improve our ability to attribute impacts to OA among co-varying parameters. Third, there is a great need to understand the potential modulation of OA impacts through the interplay of ecology and evolution (eco–evo dynamics). Lastly, further research efforts designed to detect, quantify, and project the effects of OA on marine organisms and ecosystems utilizing a comparative approach with long-term data sets will also provide critical information for informing the management of marine ecosystems.

Keywords: anthropogenic CO₂ contribution, coral reefs, ecological-evolutionary dynamics, ecosystem trajectory, oysters, pteropods, seagrass

Introduction

A third of the anthropogenic CO₂ released to the atmosphere has been absorbed by the oceans, causing declines in ocean pH and calcium carbonate saturation state (Bindoff et al., 2019; Gruber et al., 2019). These changes are referred to as ocean acidification (OA) (Caldeira and Wickett, 2003; Doney et al., 2009; Le Quéré et al., 2018). Information from the geological record (Hönisch et al., 2012), laboratory experiments (Kroeker et al., 2013), field observations (Keller et al., 2014; Sutton et al., 2016, 2017; Henson et al., 2017; Turk et al., 2019), and numerical modelling (Marshall et al., 2017) strongly suggests that OA has the potential to alter the function of ocean ecosystems, impacting marine biota and ecosystem services (Andersson et al., 2015). However, characterizing current and future effects of OA on marine systems is challenging. While there is a general consensus that OA elicits largely negative effects on calcifying organisms and positive effects on primary producers (Kroeker et al., 2010; Busch and McElhany, 2016; Mostofa et al., 2016), these conclusions are primarily drawn from laboratory experiments in which species sensitivity is evaluated using shortterm incubations under elevated CO2 conditions. Controlled experiments have found relationships between organism responses and CO₂ conditions (Waldbusser et al., 2014) and have considered how physiological sensitivities scale to predictions of evolutionary responses (Munday et al., 2013). These types of studies offer important insight into mechanistic responses of marine organismal physiology to OA but may provide a limited assessment of population-level impacts due to the complexity of how OA impacts may cascade through ecosystems (e.g. variation in the sensitivity of individuals within a community and subsequent impacts on population dynamics; Busch et al., 2013; Busch and McElhany, 2016). In addition to characterizing existing organismal sensitivities to OA, we must document how ecologically complex in situ conditions (e.g. simulating natural variability of carbonate chemistry, food availability) vary from those observed in the laboratory to understand OA impacts and interpret ecosystem-level responses (Andersson and Mackenzie, 2012). OA sensitivities are also expected to vary widely in natural systems, for example an average 0.1 decline in pH due to OA could be enough to push some species or ecosystems over critical thresholds, or might be unimportant in systems that have natural variability ranging from 0.5 to 1 pH units within a day (Hofmann et al., 2011).

Challenges of both spatial and temporal scaling of laboratory results to *in situ* responses are compounded by the need to understand how OA interacts with other physical, chemical, and

biological forcings (Breitburg et al., 2015; Kroeker et al., 2017). While researchers generally agree that a multifaceted approach is necessary, evaluating the benefits and drawbacks of different approaches requires careful consideration (see Andersson et al., 2015; Boyd et al., 2018). For example, free ocean carbon enrichment-type experiments constrain natural variation between specific locations within an ecosystem while only manipulating CO₂ (Barry et al., 2014; Gattuso et al., 2014; Doo et al., 2019) but are difficult to scale to ecosystem-level projections of OA impacts. Furthermore, in situ large-scale pelagic mesocosms studies have been performed to document changes in plankton communities, although the community composition and trajectory (e.g. potential phytoplankton blooms in select mesocosms) are difficult to constrain (Bach et al., 2016; Algueró-Muñiz et al., 2017; Riebesell et al., 2017, 2018). Field-based observations are largely gleaned from natural CO₂ gradients (from vents/seeps and spatial pH gradients) across ecosystem scales (e.g. Hall-Spencer et al., 2008; Fabricius et al., 2011; Silbiger et al., 2014; Barkley et al., 2015; Mollica et al., 2018). However, these effects are often interwoven with other physical and chemical parameters that are difficult to disentangle (Silbiger et al., 2017). Other methods, including statistical techniques (e.g. Silbiger et al., 2014) and proxies (e.g. Mollica et al., 2018), have been used to gain insight into how ecosystems may respond to OA. Scaling between sensitivity information from laboratory settings to multi-generational and ecosystem-level responses in nature has largely been done using conceptual models (Busch et al., 2015; Edmunds et al., 2016). These models are complemented by laboratory studies that assess potential transgenerational adaptation effects, highlighting the possibility for organisms to rapidly adapt to changing CO₂ conditions (Parker et al., 2015; Putnam and Gates, 2015; Wong et al., 2018). Although both natural and laboratory experiments strongly suggest negative biological consequences in response to OA, long-term (multi-decadal) biological and ecological measurements that are unequivocally linked to anthropogenic CO2 accumulation in situ are limited to a handful of studies, mostly on planktonic foraminifera (de Moel et al., 2009; Moy et al., 2009; Fox et al., 2020; Osborne et al., 2020).

Disentangling effects of OA on marine species from natural environmental variability and other climate change drivers has been a cornerstone of OA research over the past decade (Breitburg *et al.*, 2015). The quality and abundance of ocean carbonate chemistry measurements have advanced, making progress in attributing ocean chemistry changes to anthropogenic CO_2 (Weisberg *et al.*, 2016). Although OA has been unequivocally

observed in the open ocean (Bates et al., 2014), this trend is only beginning to be documented in near-shore environments due to high natural variability and limited duration of observations (Duarte et al., 2013; Andersson et al., 2015; Reimer et al., 2017; Sutton et al., 2019). Time of emergence refers to the point at which an anthropogenic signal is detectable outside the bounds of natural variability; it has been applied with success to marine carbonate chemistry and other oceanographic measurements of CO₂ increase (Keller et al., 2014; Sutton et al., 2016, 2017; Henson et al., 2017; Turk et al., 2019), but has not been observed in some ocean environments, especially those lacking historical measurements, including many coastal regions (Sutton et al., 2019). With many marine ecosystems lacking time-series measurements of carbonate chemistry and biological indices that are longer than the time of emergence, the extent to which biological responses are attributable to OA in nature remains an open question.

The topic of scaling from laboratory-based, single-species studies to understanding OA impacts in situ has been discussed in previous perspectives and syntheses (Hennige et al., 2014; Riebesell and Gattuso, 2015; Hurd et al., 2018). Our aim here is to review our ability to detect and attribute OA impacts for four well-studied groups and to stimulate further discussion and consideration of how to improve detection and attribution as the OA research field continues moving forward. Here, we refer to OA sensitivity as any biological response (physiological change) of an organism to increasing CO2. An impact of OA is defined as a change in an *in situ* biological measurement that is attributed to in situ changes in seawater chemistry resulting from increasing anthropogenic CO₂. We focus on four groups (seagrasses, pteropods, oysters, and coral reefs), selected for their sensitivity to OA and their ecological and/or economic importance. The authors also have expertise in each of these groups. For each, we summarize the results of laboratory and field-based studies on CO₂ sensitivity and the current ability to detect and attribute change in the system to OA. The complications discussed here are not meant to criticize existing studies but to highlight the need for a greater understanding of the impacts of OA in natural ecosystems and for an improved ability to attribute and quantify these impacts.

Seagrass

Seagrasses are commonly considered potential beneficiaries of OA; they are carbon-limited under current CO₂ conditions and increase photosynthesis under higher CO₂ concentrations (Koch et al., 2013). This is in contrast to most marine autotrophs, which have developed efficient strategies for utilizing bicarbonate (HCO_3^-) , and is due to the relatively recent evolution of marine seagrasses under comparatively higher CO₂ concentrations (Beer and Koch, 1996; Zimmerman et al., 1997). Results from mesocosm and in situ manipulations of CO2 indicate increased seagrass productivity, shoot density, and biomass under elevated CO₂ conditions (Beer and Koch, 1996; Zimmerman et al., 1997; Hall-Spencer et al., 2008; Fabricius et al., 2011; Campbell and Fourqurean, 2014). However, divergent results have been found in volcanic CO₂ seep sites. Seagrasses in the Mediterranean show decreases in density and biomass (Apostolaki et al., 2014) and in Papua New Guinea have up to a fivefold biomass increase (Takahashi et al., 2016) with increasing CO2. In addition, seagrass species live in a complex environment; thus, seagrass response to OA will likely be modulated by interactions with other species. For example, a decrease in calcareous epiphytes on seagrasses at

 CO_2 seeps has been shown (Martin *et al.*, 2008), while the potential for an increase in fleshy epiphytes has also been documented (Campbell and Fourqurean, 2014). Globally, seagrass abundance has declined by ~30%, which has been attributed to coastal urbanization, rising sea surface temperatures, and water quality deg-

radation (Waycott et al., 2009). To our knowledge, no in situ study has attributed positive effects of anthropogenic OA on seagrass growth, while decreases in species density and range have been observed in response to other anthropogenic stress (e.g. pollution, warming; Koch et al., 2013). Furthermore, theoretical OA refugia created by seagrasses have not yet been observed consistently in situ and are likely dependent on site-specific factors (e.g. residence times, autotroph location relative to water advection, community composition) making successful in situ attribution of benefits to adjacent calcifiers difficult (Anthony et al., 2011, 2013; Unsworth et al., 2012; Mongin et al., 2016). In addition, although photosynthesis by seagrasses decreases CO₂ during the day, potentially equal or greater night-time respiration may counteract daytime effects by increasing CO₂, resulting in a near-zero daily balance that produces negligible effects on the progression of OA (Koweek et al., 2018; Pacella et al., 2018; Kapsenberg and Cyronak, 2019). While the theoretical benefits of OA on seagrass growth have been well documented in the laboratory, it appears that substantial negative impacts from other anthropogenic stressors may counteract any positive effects of increased CO₂ and have likely prevented the isolation and attribution of the potential beneficial responses of OA (Koch et al., 2013).

Pteropods

Pteropods were one of the first taxonomic groups identified as vulnerable to OA (Orr *et al.*, 2005). Numerous laboratory experiments have documented negative effects of exposure to elevated CO₂, including shell dissolution, reduced (or absent) calcification, altered respiration rates, decreased sinking rates, differential gene expression, delayed egg development, and increased mortality (Comeau *et al.*, 2009, 2010a, b; Lischka and Riebesell, 2012, 2017; Manno *et al.*, 2012; Seibel *et al.*, 2012; Busch *et al.*, 2014; Koh *et al.*, 2015; Maas *et al.*, 2015; Thabet *et al.*, 2015; Moya *et al.*, 2016; Johnson and Hofmann, 2017). However, the response of pteropods to high CO₂ is not uniformly negative (Maas *et al.*, 2016), and the outer organic layer of the pteropod shell offers some protection from undersaturated waters (Peck *et al.*, 2016, 2018).

OA-related pteropod field observations have focused on a variety of time scales and response metrics. Analysis of pteropod shell collections from the past 100 years in the Mediterranean show declines in shell thickness and density for two different species (Howes et al., 2017). Sediment core studies indicate some evidence for a correlation between fossil pteropod shell dissolution during life and atmospheric CO₂ (Wall-Palmer et al., 2012, 2013; Manno et al., 2017). Single-season, in situ studies have shown correlations between carbonate chemistry conditions and pteropod shell dissolution, oxidative stress, relative abundance, and vertical distribution (Bednaršek et al., 2012, 2014, 2017, 2018; Bednaršek and Ohman, 2015; Feely et al., 2016; Engström-Öst et al., 2019). Observations of shell dissolution along natural gradients in aragonite saturation state (Ω_{ar}) and snapshots of current pteropod distributions correlated with Ω_{ar} have been combined with historical reconstructions of carbonate chemistry to provide hypotheses about recent changes in pteropod abundance due to OA (Bednaršek et al., 2017). While spatial gradient studies show correlations with carbonate chemistry that provide strong evidence for a negative effect of OA on pteropod shell condition, they do not necessarily offer direct evidence of modern OA effects because they substitute space for time and make inferences about historical states without direct observations (McElhany, 2017). Available time-series analyses find no significant relationships between pteropod abundance and carbonate chemistry (Howes et al., 2015; Thibodeau et al., 2018). Recent analyses of pteropod abundance time-series from around the globe show that populations vary in trajectories with some declining, some increasing, and others showing no change; this is counter to what would be expected if the negative effects of OA now dominate population processes, suggesting that other local and regional drivers, including ocean warming, currently influence pteropods more than OA (Ohman et al., 2009; Head and Pepin, 2010; Mackas and Galbraith, 2012; Beare et al., 2013; Beaugrand et al., 2013). While both historical and modern samples suggest that pteropods are sensitive to carbonate chemistry conditions, more evidence is needed to link the progress of OA to impacts on the demographics of pteropod populations. It is possible that there are variable responses of pteropods in situ, time-series are not yet long enough to detect a directional change caused by OA, and/or the chemical thresholds at which ocean carbonate chemistry influences pteropods have not yet been crossed at the ecosystem scale.

Oysters

Impacts of elevated CO2 on oyster larvae were key in raising concerns about the implications of OA for marine ecosystems (Kelly et al., 2014). Laboratory studies have vielded a more complete understanding of the sensitivity of oysters to acidified conditions, documenting effects in the larval stage such as decreased calcification, reduced growth, delayed metamorphosis, and increased mortality (Miller et al., 2009; Talmage and Gobler, 2009; Watson et al., 2009; Parker et al., 2010, 2011; Dickinson et al., 2012; Waldbusser et al., 2013; Barton et al., 2015; Frieder et al., 2017). Laboratory research has also indicated that juvenile and adult oysters are sensitive to OA, though responses are variable. Some species and populations show changes in metabolism, calcification, and shell strength under OA conditions, with effects on juveniles sometimes carried over from larval exposure (Gazeau et al., 2007; Beniash et al., 2010; Welladsen et al., 2010; Parker et al., 2011, 2012; Hettinger et al., 2012; Sanford et al., 2014; Wright et al., 2014).

Carbonate chemistry conditions documented in shellfish hatcheries provide an example of how acidification can be linked to declines in larval performance in an artificial system (Barton et al., 2012; Ellis et al., 2017). Many oyster hatcheries now control seawater conditions (modification of carbonate chemistry, abundance of food, decrease in predation) and oyster producers have long practiced selection/breeding for performance (Barton et al., 2012; Ellis et al., 2017). Curiously, Pacific oyster recruitment still occurs in wild populations exposed to Ω_{ar} near threshold limits for calcification found in the laboratory (Ruesink et al., 2018). This apparent contradiction suggests that the influence of carbonate chemistry on oyster populations is complex and likely affected by varying and heterogeneous chemical conditions, other environmental factors, adaptation mechanisms, and/or transgenerational effects (Parker et al., 2010, 2012, 2017a, b; Dickinson et al., 2012; Hettinger et al., 2013; Ruesink et al., 2018). There is limited information about the micro-habitat carbonate chemistry conditions that natural oyster populations experience (Hales et al., 2017), though first principles suggest that they persist in a wide range of conditions given the influence of fluctuations in freshwater inputs, other dynamic physical drivers, and biological activity in their habitat. Over the last 130 years, a global decline in oyster populations has been driven by over-harvesting, competition with non-native species, disease, and other anthropogenic factors (Beck *et al.*, 2011). Any role of OA in these changes *in situ* is still unclear due to the lack of available demographic data and related carbonate chemistry time-series in coastal environments.

Tropical coral reefs

The expectation that OA will negatively affect tropical coral reef calcification is rooted in thermodynamics (e.g. Plummer and Busenberg, 1987) and early abiogenic CaCO₃ precipitation experiments that provided a quantitative framework within which to understand, predict, and interpret biological responses (Burton and Walter, 1987; Morse and Mackenzie, 1990). Subsequent experiments supported the prediction that as Ω_{ar} declines, calcification decreases (Langdon et al., 2000; Leclercq et al., 2002; Langdon and Atkinson, 2005) and CaCO₃ dissolution increases (Andersson et al., 2007; Andersson and Gledhill, 2013). Field and laboratory-based studies suggest that OA may enhance the bioerosion capabilities of borers, increasing breakdown of the calcium carbonate framework (Tribollet et al., 2009; Wisshak et al., 2012; Silbiger et al., 2014; DeCarlo et al., 2015). Field studies have found correlations between Ω_{ar} and net ecosystem calcification (NEC), the net balance of gross ecosystem calcification and dissolution. For example, manipulative short-term, in situ, pulse alkalinization (Albright et al., 2016) and pulse acidification (Albright et al., 2018) experiments across a coral reef flat documented increased and decreased NEC, respectively, providing critical information for how net calcification responds to OA at the ecosystem level. Field observations across natural Ω_{ar} gradients report declines in coral skeletal density, coral diversity, colony size, NEC, and increases in bioerosion and dissolution with declining Ω_{ar} (Silverman *et al.*, 2007; Manzello *et al.*, 2008; Fabricius et al., 2011; Shamberger et al., 2011; Enochs et al., 2016; Silbiger et al., 2016; Eyre et al., 2018; Mollica et al., 2018). However, there are notable exceptions (e.g. Shamberger et al., 2014; Barkley et al., 2015; DeCarlo et al., 2017; Silbiger et al., 2017).

The general expectation, based on theoretical predictions and experimental results, is that OA should have already negatively affected coral reefs (Table 1). However, the current inability to confidently isolate and attribute effects of anthropogenic OA on coral reefs *in situ* suggests that either the current measurement methods are not sensitive enough to detect expected impacts, or these impacts have been mitigated by other processes or masked by covarying oceanic changes that have stronger effects. Key insights from the last decade of OA coral reef studies are as follows:

- The metabolism of coral reef organisms strongly affects coral reef seawater chemistry (e.g. Shaw *et al.*, 2012; Cyronak *et al.*, 2014; Shamberger *et al.*, 2014; DeCarlo *et al.*, 2017) and may slow or enhance the acidification of the surrounding openocean source water to the reef.
- Corals and other coral reef organisms modulate the chemistry of their calcifying fluids and may override changes in the chemistry of the seawater source to the site of calcification

Table 1. Summary of marine system responses to OA

Marine groups	Summary of experimental findings	Observations of wild populations	Data/analysis that could increase detection <i>in situ</i>
Seagrasses	Increased productivity, shoot density, and biomass; changes in community composition	No effects attributable directly to OA	Improved understanding of the interplay of the factors that drive seagrass abundance and distribution
Pteropods	Dissolution, reduced calcification, physiological and early life stage impairments, mortality	Dissolution in naturally low pH environments; no population effects attributable directly to OA	Multi-factor analyses to tease out the role of OA in driving pteropod condition and population dynamics from modern and historical samples
Oysters	Reduced calcification/growth, physiological effects, and mortality, particularly in larvae and juveniles	No effects attributable directly to OA	Condition and demography of populations living in different carbonate chemistry environments; studies of the effects of OA throughout the entire life cycle in the context of multiple interacting drivers
Coral reef ecosystems	Reduced calcification, increased dissolution, and bioerosion	Increased bioerosion and dissolution; no effects attributable directly to OA	Constrain natural spatiotemporal variability of NEC; understand response to multiple interacting drivers; long- term time-series studies of environmental and reef conditions

The expected impacts are based on laboratory/mesocosm CO_2 sensitivity experiments, and observations are based on *in situ* studies (e.g. time-series, natural pH gradients). Data or analyses that may improve the probability of detecting the impacts of OA *in situ* are suggested.

(Cohen and Holcomb, 2009; Cohen *et al.*, 2009; McCulloch *et al.*, 2012).

- Coral feeding, availability of dissolved inorganic nutrients, and energetic demands related to reproductive status can mitigate or exacerbate the impact of OA on coral calcification (Langdon and Atkinson, 2005; Cohen and Holcomb, 2009; Holcomb *et al.*, 2010; Edmunds, 2011; Drenkard *et al.*, 2013; Silbiger *et al.*, 2018; Kealoha *et al.*, 2019).
- Ocean-warming-induced coral bleaching is an important dominant driver of declines in coral growth over the 20th century (Cantin *et al.*, 2010; Courtney *et al.*, 2017; Hughes *et al.*, 2018) that may mask the influence of OA on coral growth histories.

Naturally high variability and uncertainty in NEC measurements (Courtney and Andersson, 2019) makes it difficult to determine whether changes in NEC are driven by environmental change or are within the natural variability of the system (Silverman *et al.*, 2014; Shamberger *et al.*, 2018). One consistent response of coral reef organisms and ecosystems across natural gradients in pH, in both laboratory and field experiments and observations, is an increase in bioerosion and sediment dissolution (e.g. Barkley *et al.*, 2015; DeCarlo *et al.*, 2015; Silbiger and Donahue, 2015; Enochs *et al.*, 2016; Silbiger *et al.*, 2016; Eyre *et al.*, 2018). However, these processes are also influenced by factors such as nutrient inputs and organic matter content of sediments, and deconvolving the various contributions remains challenging.

Research needs for OA attribution in biological systems

Great strides have been made to understand OA impacts. In this perspective, we highlight that laboratory-based studies have identified a variety of ways that a broad taxonomic range of marine species are sensitive to elevated CO_2 . Informed by these experimental results, progress is also being made on the detection and

attribution of anthropogenic OA impacts in wild populations (Table 1). For example, some biological impacts in situ have been correlated with carbonate chemistry and suggest attribution to OA, such as increased shell dissolution of pteropods (Bednaršek et al., 2014) and decreased shell thickness in planktic foraminifera (de Moel et al., 2009; Moy et al., 2009; Fox et al., 2020; Osborne et al., 2020). However, impacts attributable to OA have yet to be detected on ecosystem-level biological parameters such as population density, trophic interactions, or energy transfer through food webs. To improve our detection and attribution ability, research is needed to determine impacts of OA in situ. For some taxa, like oysters, studies are needed to understand how OA may influence the entire life cycle, since OA has different effects across life stages (Pandori and Sorte, 2019). Other groups discussed (seagrasses, oysters, and coral reefs) require efforts to tease out the influence of OA from other co-varying factors that drive physical and chemical conditions (Table 1). Below, we detail four avenues of research that would improve the ability to detect and attribute impacts of OA on marine ecosystems in situ.

(1) Quantify the anthropogenic contribution of CO₂ in coastal environments: a challenge for attributing change in biological systems to OA is knowledge of the chemical conditions that a species or community inhabits and how OA has altered them. The majority of long-term ocean pH/pCO₂ measurements have been made in the open-ocean, which is relatively stable chemically. Coastal oceans tend to have shorter time-series measurements of pH/pCO2, complex biogeochemical and physical processes, and a higher rate of biological activity, causing larger diel, seasonal, and episodic fluctuations in ocean chemistry (e.g. Hofmann et al., 2011; Guadayol et al., 2014; Chan et al., 2017; Silbiger and Sorte, 2018; Lowe et al., 2019). While the chemical signal of OA has already emerged in open oceans, it will take longer to emerge in coastal ecosystems (Sutton et al., 2019). Therefore, we suggest further studies that employ statistical methods to estimate anthropogenic input of CO_2 (Gruber *et al.*, 1996; Feely *et al.*, 2016; Carter *et al.*, 2017). These statistical methods will aid in quantifying chemical changes in the oceans due to OA and linking biological impacts. Global coordination of OA monitoring through the Global Ocean Acidification Observing Network will aid robust data collection and synthesis needed for estimating anthropogenic input of CO_2 (Newton *et al.*, 2019; Tilbrook *et al.*, 2019).

- (2) Attribute biological impacts to OA among other co-varying parameters: marine organisms face multiple changing and covarving physical and chemical parameters associated with climate change (e.g. OA, warming, hypoxia). Identifying specific biological traits that can be measured in situ and empirically linked to OA impacts is of crucial importance in advancing efforts to detect in situ impacts of OA. Such traits of interest to monitor in situ can be physiological (Strader et al., 2019), structural [e.g. coral skeletal density changes in Mollica et al. (2018); foraminifera test thickness changes in Moy et al. (2009)], or components of population fitness (Falkenberg et al., 2018). Importantly, there is a great need to understand how differential sensitivities to OA exist within a species' life cycle (Byrne and Przesławski, 2013). In addition, increased efforts to monitor community-level traits of interests (e.g. population density, biomass) are needed to understand ecological alterations in marine ecosystems due to OA. With all research techniques, a holistic approach of detailed characterization of both biological impacts in conjunction with physical and chemical environmental parameters are needed to achieve such an aim.
- (3) Understand how ecological-evolutionary dynamics alter OA responses in situ: feedbacks between changing conditions in marine environments and organismal adaptation potential have been highlighted with recent efforts to understand the interplay between ecology and evolution (eco-evo dynamics) in driving demographic responses (Parmesan, 2006; Chevin et al., 2013). These eco-evo dynamics on longer time scales have the potential to facilitate intra-generational adaptation to changing ocean conditions through the interplay of ecological processes such as range shifts (Sunday et al., 2012; Vergés et al., 2014; Pecl et al., 2017), alteration in phenotype such as a modification of microbiome (Botté et al., 2019), as well as epigenetic mechanisms (Putnam et al., 2016; Hofmann, 2017). It is crucial to understand how OA has the potential to alter plasticity of phenotypes, which in turn could either constrain adaptive genetic changes through the persistence of diverse genotypes within the population or promote adaptive genetic changes through allowing for persistence in extreme environments (Hendry, 2016). Phytoplankton, in particular, have been used to test the hypothesis that increased phenotypic plasticity over multiple generations will lead to increased evolution in OA conditions (Collins, 2011; Lohbeck et al., 2012; Schaum and Collins, 2014) and have found increased plasticity as a good indicator of adaptation to increasing CO₂ conditions (Schaum and Collins, 2014). Future research could expand on current studies that focus on understanding phenotypic plasticity of organismal physiology (Torda et al., 2017; Donelson et al., 2018; Ryu et al., 2018; Willoughby et al., 2018; Catullo et al., 2019) by using modelling efforts that incorporate eco-evo dynamics of both past and future OA conditions.
- (4) Characterize ecosystem trajectories through long-term monitoring: understanding how and why species are sensitive to OA has vastly improved, but this is just one aspect of understanding population and ecosystem responses in situ. For example, a species' population dynamics may be influenced more by OAinduced modifications of ecological interactions than by direct sensitivity (Marshall et al., 2017). In some instances, ecological interactions have been hypothesized to mitigate OA impacts through enhancing adaptive capacity or mitigating the effects of elevated CO₂ conditions (Kapsenberg and Cyronak, 2019). To attribute changes in species dynamics or ecological processes to OA, more work is needed to describe how OA impacts scale in situ in space and time. Insights into ecosystem environmental changes can be gained using shell geochemistry as paleo-proxies to document OA effects (Foster and Rae, 2016), and potentially how further changes in ocean conditions are linked to mass extinction and declines in biodiversity (Kiessling and Simpson, 2011; Hennige et al., 2014). Modelling exercises can help elucidate ecological processes, but they cannot replace time-series biological data. Of particular importance are long-term observational studies that pair a detectable chemical signal of OA with biological responses that account for ecological processes and patterns (e.g. yearly population growth patterns, NEC). With detailed datasets, broad comparative trends can be used to understand mechanisms of resilience to disturbance events. For example, comparative data indicate that community resilience to changing conditions can develop from various environmental drivers such as indiscriminate disturbance events of crown-of-thorns starfish in Mo'orea, French Polvnesia, and repeated thermal stress in Panama, Eastern Tropical Pacific (Edmunds et al., 2019). The variation in environmental drivers has resulted in differences in reproductive strategies of dominant reef-building corals, coral-algal symbiont communities, functional diversity of herbivorous fishes, and the reef framework (Edmunds et al., 2019), highlighting that comparative approaches can be used to understand how differing environmental drivers (such as OA) can alter ecosystem trajectories.

Current challenges in attributing large-scale OA effects on marine systems does not mean that there has been no OA effect to date nor that there will not be one in the future. We are beginning efforts to detect and attribute OA impacts *in situ*, with experimental results informing field campaigns and observational studies approaching the time of emergence for an OA signal in increasingly variable environments. Knowledge accumulated over the last decade puts us in a better position to design an observation system that could detect the emergence of impacts of OA at species and ecosystem levels. Research on species sensitivity to OA that can be scaled into projected ecosystem-level impacts in a multi-stressor ocean and verified with *in situ* detection is critical to inform the conservation and sustainable use of ocean ecosystems.

Acknowledgements

This study is a product of the Ocean Acidification Principal Investigators Meeting (17–19 February 2018), organized by the Ocean Carbon and Biogeochemistry Project Office with support from the National Science Foundation. No new data were analyzed or generated in support of this research.

Funding

SSD was funded by NSF OCE (grant # 1415268). DSB and PM were supported by the NOAA Ocean Acidification Program and Northwest Fisheries Science Center, MHL was supported by NSF OCE (grant # 1633951), ZIJ was supported by NSF OCE (grant # 1416665) and DOE EERE (grant #DE-EE008518), NJS was supported by NSF OCE (grant # 1924281), ALC was supported by NSF OCE (grant # 1737311), and AA was supported by NSF OCE (grant # 1416518). KEFS, AK, and TLH were supported by Texas A&M University. This is CSUN Marine Biology contribution (# 306).

Author contributions

All authors conceived the idea for this paper in discussion at a workshop and contributed to the writing of the manuscript. SSD and DSB led the group and contributed the most to the text.

References

- Albright, R., Caldeira, L., Hosfelt, J., Kwiatkowski, L., Maclaren, J. K., Mason, B. M., Nebuchina, Y., *et al.* 2016. Reversal of ocean acidification enhances net coral reef calcification. Nature, 531: 362–365.
- Albright, R., Takeshita, Y., Koweek, D. A., Ninokawa, A., Wolfe, K., Rivlin, T., Nebuchina, Y., *et al.* 2018. Carbon dioxide addition to coral reef waters suppresses net community calcification. Nature, 555: 516–519.
- Algueró-Muñiz, M., Alvarez-Fernandez, S., Thor, P., Bach, L. T., Esposito, M., Horn, H. G., Ecker, U., *et al.* 2017. Ocean acidification effects on mesozooplankton community development: results from a long-term mesocosm experiment. PLoS One, 12: e0175851.
- Andersson, A. J., Bates, N. R., and Mackenzie, F. T. 2007. Dissolution of carbonate sediments under rising pCO₂ and ocean acidification: observations from Devil's Hole, Bermuda. Aquatic Geochemistry, 13: 237–264.
- Andersson, A. J., and Gledhill, D. 2013. Ocean acidification and coral reefs: effects on breakdown, dissolution, and net ecosystem calcification. Annual Review of Marine Science, 5: 321–348.
- Andersson, A. J., Kline, D., Edmunds, P., Archer, S., Bednaršek, N., Carpenter, R., Chadsey, M., *et al.* 2015. Understanding ocean acidification impacts on organismal to ecological scales. Oceanography, 25: 16–27.
- Andersson, A. J., and Mackenzie, F. T. 2012. Revisiting four scientific debates in ocean acidification research. Biogeosciences, 9: 893–905.
- Anthony, K. R. N., Diaz-Pulido, G., Verlinden, N., Tilbrook, B., and Andersson, A. J. 2013. Benthic buffers and boosters of ocean acidification on coral reefs. Biogeosciences, 10: 4897–4909.
- Anthony, K. R. N., Kleypas, J. A., and Gattuso, J.-P. 2011. Coral reefs modify their seawater carbon chemistry—implications for impacts of ocean acidification. Global Change Biology, 17: 3655–3666.
- Apostolaki, E. T., Vizzini, S., Hendriks, I. E., and Olsen, Y. S. 2014. Seagrass ecosystem response to long-term high CO₂ in a Mediterranean volcanic vent. Marine Environmental Research, 99: 9–15.
- Bach, L. T., Taucher, J., Boxhammer, T., Ludwig, A.; The Kristineberg KOSMOS Consortium, Achterberg, E. P., Algueró-Muñiz, M., et al. 2016. Influence of ocean acidification on a natural winter-to-summer plankton succession: first insights from a long-term mesocosm study draw attention to periods of low nutrient concentrations. PLoS One, 11: e0159068.
- Barkley, H. C., Cohen, A. L., Golbuu, Y., Starczak, V. R., DeCarlo, T. M., and Shamberger, K. E. F. 2015. Changes in coral reef

communities across a natural gradient in seawater pH. Science Advances, 1: e1500328.

- Barry, J. P., Lovera, C., Buck, K. R., Peltzer, E. T., Taylor, J. R., Walz, P., Whaling, P. J., *et al.* 2014. Use of a free ocean CO₂ enrichment (FOCE) system to evaluate the effects of ocean acidification on the foraging behavior of a deep-sea urchin. Environmental Science & Technology, 48: 9890–9897.
- Barton, A., Hales, B., Waldbusser, G. G., Langdon, C., and Feely, R. A. 2012. The Pacific oyster, *Crassostrea gigas*, shows negative correlation to naturally elevated carbon dioxide levels: implications for near-term ocean acidification effects. Limnology and Oceanography, 57: 698–710.
- Barton, A., Waldbusser, G. G., Feely, R. A., Weisberg, S. B., Newton, J. A., Hales, B., Cudd, S., *et al.* 2015. Impacts of coastal acidification on the Pacific Northwest shellfish industry and adaptation strategies implemented in response. Oceanography, 25: 146–159.
- Bates, N., Astor, Y., Church, M., Currie, K., Dore, J., Gonaález-Dávila, M., Lorenzoni, L., *et al.* 2014. A time-series view of changing ocean chemistry due to ocean uptake of anthropogenic CO₂ and ocean acidification. Oceanography, 27: 126–141.
- Beare, D., McQuatters-Gollop, A., van der Hammen, T., Machiels, M., Teoh, S. J., and Hall-Spencer, J. M. 2013. Long-term trends in calcifying plankton and pH in the North Sea. PLoS One, 8: e61175.
- Beaugrand, G., McQuatters-Gollop, A., Edwards, M., and Goberville, E. 2013. Long-term responses of North Atlantic calcifying plankton to climate change. Nature Climate Change, 3: 263–267.
- Beck, M. W., Brumbaugh, R. D., Airoldi, L., Carranza, A., Coen, L. D., Crawford, C., Defeo, O., *et al.* 2011. Oyster reefs at risk and recommendations for conservation, restoration, and management. Bioscience, 61: 107–116.
- Bednaršek, N., Feely, R. A., Beck, M. W., Glippa, O., Kanerva, M., and Engström-Öst, J. 2018. El Niño-related thermal stress coupled with upwelling-related ocean acidification negatively impacts cellular to population-level responses in pteropods along the California current system with implications for increased bioenergetic costs. Frontiers in Marine Science, 5: 486.
- Bednaršek, N., Feely, R. A., Reum, J. C. P., Peterson, B., Menkel, J., Alin, S. R., and Hales, B. 2014. *Limacina helicina* shell dissolution as an indicator of declining habitat suitability owing to ocean acidification in the California Current Ecosystem. Proceedings of the Royal Society B: Biological Sciences, 281: 20140123.
- Bednaršek, N., Feely, R. A., Tolimieri, N., Hermann, A. J., Siedlecki, S. A., Waldbusser, G. G., McElhany, P., *et al.* 2017. Exposure history determines pteropod vulnerability to ocean acidification along the US West Coast. Scientific Reports, 7: 4526.
- Bednaršek, N., and Ohman, M. D. 2015. Changes in pteropod distributions and shell dissolution across a frontal system in the California Current System. Marine Ecology Progress Series, 523: 93–103.
- Bednaršek, N., Tarling, G. A., Bakker, D. C. E., Fielding, S., Jones, E. M., Venables, H. J., Ward, P., *et al.* 2012. Extensive dissolution of live pteropods in the Southern Ocean. Nature Geoscience, 5: 881–885.
- Beer, S., and Koch, E. 1996. Photosynthesis of marine macroalgae and seagrasses in globally changing CO₂ environments. Marine Ecology Progress Series, 141: 199–204.
- Beniash, E., Ivanina, A., Lieb, N. S., Kurochkin, I., and Sokolova, I. M. 2010. Elevated level of carbon dioxide affects metabolism and shell formation in oysters *Crassostrea virginica* (Gmelin). Marine Ecology Progress Series, 419: 95–108.
- Bindoff, N. L., Cheung, W. W. L., Kairo, J. G., Arístegui, J., Guinder, V. A., Hallberg, R., Hilmi, N., *et al.* 2019. Changing ocean, marine ecosystems, and dependent communities. *In* IPCC Special Report on the Ocean and Cryosphere in a Changing Climate. Ed. by H.-O. Pörtner, D. C. Roberts, V. Masson-Delmotte, P. Zhai, M. Tignor, E. Poloczanska, K. Mintenbeck, *et al. in press.*

- Botté, E. S., Nielsen, S., Abdul Wahab, M. A., Webster, J., Robbins, S., Thomas, T., and Webster, N. S. 2019. Changes in the metabolic potential of the sponge microbiome under ocean acidification. Nature Communications, 10: 4134.
- Boyd, P. W., Collins, S., Dupont, S., Fabricius, K., Gattuso, J.-P., Havenhand, J., Hutchins, D. A., *et al.* 2018. Experimental strategies to assess the biological ramifications of multiple drivers of global ocean change—a review. Global Change Biology, 24: 2239–2261.
- Breitburg, D. L., Hondorp, D., Audemard, C., Carnegie, R. B., Burrell, R. B., Trice, M., and Clark, V. 2015. Landscape-level variation in disease susceptibility related to shallow-water hypoxia. PLoS One, 10: e0116223.
- Burton, E. A., and Walter, L. M. 1987. Relative precipitation rates of aragonite and Mg calcite from seawater: temperature or carbonate ion control? Geology, 15: 111.
- Busch, D. S., Harvey, C. J., and McElhany, P. 2013. Potential impacts of ocean acidification on the Puget Sound food web. ICES Journal of Marine Science, 70: 823–833.
- Busch, D. S., and McElhany, P. 2016. Estimates of the direct effect of seawater pH on the survival rate of species groups in the California Current Ecosystem. PLoS One, 11: e0160669.
- Busch, D. S., O'Donnell, M., Hauri, C., Mach, K., Poach, M., Doney, S., Signorini, S., *et al.* 2015. Understanding, characterizing, and communicating responses to ocean acidification: challenges and uncertainties. Oceanography, 25: 30–39.
- Busch, D. S., Maher, M., Thibodeau, P., and McElhany, P. 2014. Shell condition and survival of Puget Sound pteropods are impaired by ocean acidification conditions. PLoS One, 9: e105884.
- Byrne, M., and Przesławski, R. 2013. Multistressor impacts of warming and acidification of the ocean on marine invertebrates' life histories. Integrative and Comparative Biology, 53: 582–596.
- Caldeira, K., and Wickett, M. E. 2003. Oceanography: anthropogenic carbon and ocean pH. Nature, 425: 365–365.
- Campbell, J. E., and Fourqurean, J. W. 2014. Ocean acidification outweighs nutrient effects in structuring seagrass epiphyte communities. The Journal of Ecology, 102: 730–737.
- Cantin, N. E., Cohen, A. L., Karnauskas, K. B., Tarrant, A. M., and McCorkle, D. C. 2010. Ocean warming slows coral growth in the central Red Sea. Science, 329: 322–325.
- Carter, B. R., Feely, R. A., Mecking, S., Cross, J. N., Macdonald, A. M., Siedlecki, S. A., Talley, L. D., *et al.* 2017. Two decades of pacific anthropogenic carbon storage and ocean acidification along global ocean ship-based hydrographic investigations program sections P16 and P02, Global Biogeochemical Cycles, 31: 306–327.
- Catullo, R. A., Llewelyn, J., Phillips, B. L., and Moritz, C. C. 2019. The potential for rapid evolution under anthropogenic climate change. Current Biology, 29: R996–R1007.
- Chan, F., Barth, J. A., Blanchette, C. A., Byrne, R. H., Chavez, F., Cheriton, O., Feely, R. A., *et al.* 2017. Persistent spatial structuring of coastal ocean acidification in the California Current System. Scientific Reports, 7: 2526.
- Chevin, L.-M., Gallet, R., Gomulkiewicz, R., Holt, R. D., and Fellous, S. 2013. Phenotypic plasticity in evolutionary rescue experiments. Philosophical Transactions of the Royal Society of London Series B, Biological Sciences, 368:20120089.
- Cohen, A. L., and Holcomb, M. 2009. Why corals care about ocean acidification: uncovering the mechanism. Oceanography, 22: 118–127.
- Cohen, A. L., McCorkle, D. C., de Putron, S., Gaetani, G. A., and Rose, K. A. 2009. Morphological and compositional changes in the skeletons of new coral recruits reared in acidified seawater: insights into the biomineralization response to ocean acidification. Geochemistry, Geophysics, Geosystems, 10: Q07005.
- Collins, S. 2011. Competition limits adaptation and productivity in a photosynthetic alga at elevated CO₂. Proceedings of the Royal Society B: Biological Sciences, 278: 247–255.

- Comeau, S., Gorsky, G., Alliouane, S., and Gattuso, J.-P. 2010a. Larvae of the pteropod *Cavolinia inflexa* exposed to aragonite undersaturation are viable but shell-less. Marine Biology, 157: 2341–2345.
- Comeau, S., Gorsky, G., Jeffree, R., Teyssié, J.-L., and Gattuso, J.-P. 2009. Impact of ocean acidification on a key Arctic pelagic mollusc (*Limacina helicina*). Biogeosciences, 6: 1877–1882.
- Comeau, S., Jeffree, R., Teyssié, J.-L., and Gattuso, J.-P. 2010b. Response of the Arctic pteropod *Limacina helicina* to projected future environmental conditions. PLoS One, 5: e11362.
- Courtney, T. A., and Andersson, A. J., 2019. Evaluating measurements of coral reef net ecosystem calcification rates. Coral Reefs, 38: 997–1006.
- Courtney, T. A., Lebrato, M., Bates, N. R., Collins, A., de Putron, S. J., Garley, R., Johnson, R., *et al.* 2017. Environmental controls on modern scleractinian coral and reef-scale calcification. Science Advances, 3: e1701356.
- Cyronak, T., Santos, I. R., Erler, D. V., Maher, D. T., and Eyre, B. D. 2014. Drivers of pCO₂ variability in two contrasting coral reef lagoons: the influence of submarine groundwater discharge. Global Biogeochemical Cycles, 28: 398–414.
- DeCarlo, T. M., Cohen, A. L., Barkley, H. C., Cobban, Q., Young, C., Shamberger, K. E., Brainard, R. E., *et al.* 2015. Coral macrobioerosion is accelerated by ocean acidification and nutrients. Geology, 43: 7–10.
- DeCarlo, T. M., Cohen, A. L., Wong, G. T. F., Shiah, F.-K., Lentz, S. J., Davis, K. A., Shamberger, K. E. F., *et al.* 2017. Community production modulates coral reef pH and the sensitivity of ecosystem calcification to ocean acidification. Journal of Geophysical Research: Oceans, 122: 745–761.
- de Moel, H., Ganssen, G. M., Peeters, F. J. C., Jung, S. J. A., Kroon, D., Brummer, G. J. A., and Zeebe, R. E. 2009. Planktic foraminiferal shell thinning in the Arabian Sea due to anthropogenic ocean acidification? Biogeosciences, 6: 1917–1925.
- Dickinson, G. H., Ivanina, A. V., Matoo, O. B., Pörtner, H. O., Lannig, G., Bock, C., Beniash, E., *et al.* 2012. Interactive effects of salinity and elevated CO_2 levels on juvenile eastern oysters, *Crassostrea virginica*. The Journal of Experimental Biology, 215: 29–43.
- Donelson, J. M., Salinas, S., Munday, P. L., and Shama, L. N. S. 2018. Transgenerational plasticity and climate change experiments: where do we go from here? Global Change Policy, 24: 13–34.
- Doney, S. C., Fabry, V. J., Feely, R. A., and Kleypas, J. A. 2009. Ocean acidification: the other CO₂ problem. Annual Review of Marine Science, 1: 169–192.
- Doo, S. S., Edmunds, P. J., and Carpenter, R. C. 2019. Ocean acidification effects on *in situ* coral reef metabolism. Scientific Reports, 9: 12067.
- Drenkard, E. J., Cohen, A. L., McCorkle, D. C., de Putron, S. J., Starczak, V. R., and Zicht, A. E. 2013. Calcification by juvenile corals under heterotrophy and elevated CO₂. Coral Reefs, 32: 727–735.
- Duarte, C. M., Hendriks, I. E., Moore, T. S., Olsen, Y. S., Steckbauer, A., Ramajo, L., Carstensen, J., *et al.* 2013. Is ocean acidification an open-ocean syndrome? Understanding anthropogenic impacts on seawater pH. Estuaries and Coasts, 36: 221–236.
- Edmunds, P. J. 2011. Zooplanktivory ameliorates the effects of ocean acidification on the reef coral *Porites* spp. Limnology and Oceanography, 56: 2402–2410.
- Edmunds, P. J., Adam, T. C., Baker, A. C., Doo, S. S., Glynn, P. W., Manzello, D. P., Silbiger, N. J., *et al.* 2019. Why more comparative approaches are required in time-series analyses of coral reef ecosystems. Marine Ecology Progress Series, 608: 297–306.
- Edmunds, P. J., Comeau, S., Lantz, C., Andersson, A., Briggs, C., Cohen, A., Gattuso, J.-P., *et al.* 2016. Integrating the effects of ocean acidification across functional scales on tropical coral reefs. Bioscience, 66: 350–362.

- Ellis, R. P., Urbina, M. A., and Wilson, R. W. 2017. Lessons from two high CO₂ worlds—future oceans and intensive aquaculture. Global Change Biology, 23: 2141–2148.
- Engström-Öst, J., Glippa, O., Feely, R. A., Kanerva, M., Keister, J. E., Alin, S. R., Carter, B. R., *et al.* 2019. Eco-physiological responses of copepods and pteropods to ocean warming and acidification. Scientific Reports, 9: 4748.
- Enochs, I. C., Manzello, D. P., Kolodziej, G., Noonan, S. H. C., Valentino, L., and Fabricius, K. E. 2016. Enhanced macroboring and depressed calcification drive net dissolution at high-CO₂ coral reefs. Proceedings of the Royal Society B: Biological Sciences, 283: 1–8.
- Eyre, B. D., Cyronak, T., Drupp, P., De Carlo, E. H., Sachs, J. P., and Andersson, A. J. 2018. Coral reefs will transition to net dissolving before end of century. Science, 359: 908–911.
- Fabricius, K. E., Langdon, C., Uthicke, S., Humphrey, C., Noonan, S., De'ath, G., Okazaki, R., *et al.* 2011. Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. Nature Climate Change, 1: 165–169.
- Falkenberg, L. J., Dupont, S., and Bellerby, R. G. J. 2018. Approaches to reconsider literature on physiological effects of environmental change: examples from ocean acidification research. Frontiers of Marine Sciences, 5: 453.
- Feely, R. A., Alin, S. R., Carter, B., Bednaršek, N., Hales, B., Chan, F., Hill, T. M., *et al.* 2016. Chemical and biological impacts of ocean acidification along the west coast of North America. Estuarine, Coastal and Shelf Science, 183: 260–270.
- Foster, G. L., and Rae, J. W. B. 2016. Reconstructing ocean pH with boron isotopes in foraminifera. Annual Review of Earth and Planetary Sciences, 44: 207–237.
- Fox, L., Stukins, S., Hill, T., and Miller, C. G. 2020. Quantifying the effect of anthropogenic climate change on calcifying plankton. Scientific Reports, 10: 1620.
- Frieder, C. A., Applebaum, S. L., Pan, T.-C. F., Hedgecock, D., and Manahan, D. T. 2017. Metabolic cost of calcification in bivalve larvae under experimental ocean acidification. ICES Journal of Marine Science, 74: 941–954.
- Gattuso, J.-P., Kirkwood, W., Barry, J. P., Cox, E., Gazeau, F., Hansson, L., Hendriks, I., *et al.* 2014. Free ocean CO₂ enrichment (FOCE) systems: present status and future developments. Biogeosciences Discussions, 11: 4001–4046.
- Gazeau, F., Quiblier, C., Jansen, J. M., Gattuso, J.-P., Middelburg, J. J., and Heip, C. H. R. 2007. Impact of elevated CO₂ on shellfish calcification. Geophysical Research Letters, 34: 181.
- Gruber, N., Clement, D., Carter, B. R., Feely, R. A., van Heuven, S., Hoppema, M., Ishii, M., *et al.* 2019. The oceanic sink for anthropogenic CO₂ from 1994 to 2007. Science, 363: 1193–1199.
- Gruber, N., Sarmiento, J. L., and Stocker, T. F. 1996. An improved method for detecting anthropogenic CO₂ in the oceans. Global Biogeochemical Cycles, 10: 809–837.
- Guadayol, Ò., Silbiger, N. J., Donahue, M. J., and Thomas, F. I. M. 2014. Patterns in temporal variability of temperature, oxygen and pH along an environmental gradient in a coral reef. PLoS One, 9: e85213.
- Hales, B., Suhrbier, A., Waldbusser, G. G., Feely, R. A., and Newton, J. A. 2017. The carbonate chemistry of the 'Fattening Line,' Willapa Bay, 2011–2014. Estuaries and Coasts, 40: 173–186.
- Hall-Spencer, J. M., Rodolfo-Metalpa, R., Martin, S., Ransome, E., Fine, M., Turner, S. M., Rowley, S. J., et al. 2008. Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. Nature, 454: 96–99.
- Head, E. J. H., and Pepin, P. 2010. Spatial and inter-decadal variability in plankton abundance and composition in the Northwest Atlantic (1958–2006). Journal of Plankton Research, 32: 1633–1648.

- Hendry, A. P. 2016. Key questions on the role of phenotypic plasticity in eco-evolutionary dynamics. The Journal of Heredity, 107: 25–41.
- Hennige, S., Roberts, J. M., and Williamson, P. (Eds). 2014. An Updated Synthesis of the Impacts of Ocean Acidification on Marine Biodiversity. Technical Series No. 75. Montreal: Secretariat of the Convention on Biological Diversity. 99 pp.
- Henson, S. A., Beaulieu, C., Ilyina, T., John, J. G., Long, M., Séférian, R., Tjiputra, J., *et al.* 2017. Rapid emergence of climate change in environmental drivers of marine ecosystems. Nature Communications, 8: 14682.
- Hettinger, A., Sanford, E., Hill, T. M., Hosfelt, J. D., Russell, A. D., and Gaylord, B. 2013. The influence of food supply on the response of Olympia oyster larvae to ocean acidification. Biogeosciences, 10: 6629–6638.
- Hettinger, A., Sanford, E., Hill, T. M., Russell, A. D., Sato, K. N. S., Hoey, J., Forsch, M., *et al.* 2012. Persistent carry-over effects of planktonic exposure to ocean acidification in the Olympia oyster. Ecology, 93: 2758–2768.
- Hofmann, G. E. 2017. Ecological epigenetics in marine metazoans. Frontiers in Marine Science, 4: 4.
- Hofmann, G. E., Smith, J. E., Johnson, K. S., Send, U., Levin, L. A., Micheli, F., Paytan, A., *et al.* 2011. High-frequency dynamics of ocean pH: a multi-ecosystem comparison. PLoS One, 6: e28983.
- Holcomb, M., McCorkle, D. C., and Cohen, A. L. 2010. Long-term effects of nutrient and CO₂ enrichment on the temperate coral *Astrangia poculata* (Ellis and Solander, 1786). Journal of Experimental Marine Biology and Ecology, 386: 27–33.
- Hönisch, B., Ridgwell, A., Schmidt, D. N., Thomas, E., Gibbs, S. J., Sluijs, A., Zeebe, R., *et al.* 2012. The geological record of ocean acidification. Science, 335: 1058–1063.
- Howes, E. L., Eagle, R. A., Gattuso, J.-P., and Bijma, J. 2017. Comparison of Mediterranean pteropod shell biometrics and ultrastructure from historical (1910 and 1921) and present day (2012) samples provides baseline for monitoring effects of global change. PLoS One, 12: e0167891.
- Howes, E. L., Stemmann, L., Assailly, C., Irisson, J. O., Dima, M., Bijma, J., and Gattuso, J. P. 2015. Pteropod time series from the North Western Mediterranean (1967-2003): impacts of pH and climate variability. Marine Ecology Progress Series, 531: 193–206.
- Hughes, T. P., Anderson, K. D., Connolly, S. R., Heron, S. F., Kerry, J. T., Lough, J. M., Baird, A. H., *et al.* 2018. Spatial and temporal patterns of mass bleaching of corals in the Anthropocene. Science, 359: 80–83.
- Hurd, C. L., Lenton, A., Tilbrook, B., and Boyd, P. W. 2018. Current understanding and challenges for oceans in a higher-CO₂ world. Nature Climate Change, 8, 686–694.
- Johnson, K. M., and Hofmann, G. E. 2017. Transcriptomic response of the Antarctic pteropod *Limacina helicina* antarctica to ocean acidification. BMC Genomics, 18: 812.
- Kapsenberg, L., and Cyronak, T. 2019. Ocean acidification refugia in variable environments. Global Change Biology, 25: 3201–3214.
- Kealoha, A. K., Shamberger, K. E. F., Reid, E. C., Davis, K. A., Lentz, S. J., Brainard, R. E., Oliver, T. A., *et al.* 2019. Heterotrophy of oceanic particulate organic matter elevates net ecosystem calcification. Geophysical Research Letters, 46: 9851–9860.
- Keller, K. M., Joos, F., and Raible, C. C. 2014. Time of emergence of trends in ocean biogeochemistry. Biogeosciences, 11: 3647–3659.
- Kelly, R. P., Cooley, S. R., and Klinger, T. 2014. Narratives can motivate environmental action: the Whiskey Creek ocean acidification story. Ambio, 43: 592–599.
- Kiessling, W., and Simpson, C. 2011. On the potential for ocean acidification to be a general cause of ancient reef crises. Global Change Biology, 17: 56–57
- Koch, M., Bowes, G., Ross, C., and Zhang, X.-H. 2013. Climate change and ocean acidification effects on seagrasses and marine macroalgae. Global Change Biology, 19: 103–132.

- Koh, H. Y., Lee, J. H., Han, S. J., Park, H., Shin, S. C., and Lee, S. G. 2015. A transcriptomic analysis of the response of the Arctic pteropod *Limacina helicina* to carbon dioxide-driven seawater acidification. Polar Biology, 38: 1727–1740.
- Koweek, D. A., Zimmerman, R. C., Hewett, K. M., Gaylord, B., Giddings, S. N., Nickols, K. J., Ruesink, J. L., *et al.* 2018. Expected limits on the ocean acidification buffering potential of a temperate seagrass meadow. Ecological Applications, 28: 1694–1714.
- Kroeker, K. J., Kordas, R. L., Crim, R. N., and Singh, G. G. 2010. Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. Ecology Letters, 13: 1419–1434.
- Kroeker, K. J., Kordas, R. L., Crim, R., Hendriks, I. E., Ramajo, L., Singh, G. S., Duarte, C. M., *et al.* 2013. Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. Global Change Biology, 19: 1884–1896.
- Kroeker, K. J., Kordas, R. L., and Harley, C. D. G. 2017. Embracing interactions in ocean acidification research: confronting multiple stressor scenarios and context dependence. Biology Letters, 13: 20160802.
- Langdon, C., and Atkinson, M. J. 2005. Effect of elevated pCO₂ on photosynthesis and calcification of corals and interactions with seasonal change in temperature/irradiance and nutrient enrichment. Journal of Geophysical Research: Oceans, 110: C09S07.
- Langdon, C., Takahashi, T., Sweeney, C., Chipman, D., Goddard, J., Marubini, F., Aceves, H., *et al.* 2000. Effect of calcium carbonate saturation state on the calcification rate of an experimental coral reef. Global Biogeochemical Cycles, 14: 639–654.
- Leclercq, N., Gattuso, J.-P., and Jaubert, J. 2002. Primary production, respiration, and calcification of a coral reef mesocosm under increased CO₂ partial pressure. Limnology and Oceanography, 47: 558–564.
- Le Quéré, C., Andrew, R. M., Friedlingstein, P., Sitch, S., Pongratz, J., Manning, A. C., Korsbakken, J. I., *et al.* 2018. Global carbon budget. Earth System Science Data, 10: 405–448.
- Lischka, S., and Riebesell, U. 2012. Synergistic effects of ocean acidification and warming on overwintering pteropods in the Arctic. Global Change Biology, 18: 3517–3528.
- Lischka, S., and Riebesell, U. 2017. Metabolic response of Arctic pteropods to ocean acidification and warming during the polar night/twilight phase in Kongsfjord (Spitsbergen). Polar Biology, 40: 1211–1227.
- Lohbeck, K. T., Riebesell, U., and Reusch, T. B. H. 2012. Adaptive evolution of a key phytoplankton species to ocean acidification, Nature Geoscience, 5: 346–351.
- Lowe, A. T., Bos, J., and Ruesink, J. 2019. Ecosystem metabolism drives pH variability and modulates long-term ocean acidification in the northeast pacific coastal ocean. Scientific Reports, 9: 963.
- Maas, A. E., Lawson, G. L., and Tarrant, A. M. 2015. Transcriptome-wide analysis of the response of the thecosome pteropod *Clio pyramidata* to short-term CO₂ exposure. Comparative biochemistry and physiology. Part D. Genomics & Proteomics, 16: 1–9.
- Maas, A. E., Lawson, G. L., and Wang, Z. A. 2016. The metabolic response of thecosome pteropods from the North Atlantic and North Pacific Oceans to high CO₂ and low O₂. Biogeosciences Discussions, 13: 6191–6143.
- Mackas, D. L., and Galbraith, M. D. 2012. Pteropod time-series from the NE Pacific. ICES Journal of Marine Science, 69: 448–459.
- Manno, C., Bednaršek, N., Tarling, G. A., Peck, V. L., Comeau, S., Adhikari, D., Bakker, D. C. E., *et al.* 2017. Shelled pteropods in peril: assessing vulnerability in a high CO₂ ocean. Earth-Science Reviews, 169: 132–145.
- Manno, C., Morata, N., and Primicerio, R. 2012. *Limacina retrover*sa's response to combined effects of ocean acidification and sea water freshening. Estuarine, Coastal and Shelf Science, 113: 163–171.

- Manno, C., Peck, V. L., and Tarling, G. A. 2016. Pteropod eggs released at high pCO₂ lack resilience to ocean acidification. Scientific Reports, 6: 25752.
- Manzello, D. P., Kleypas, J. A., Budd, D. A., Eakin, C. M., Glynn, P. W., and Langdon, C. 2008. Poorly cemented coral reefs of the eastern tropical Pacific: possible insights into reef development in a high-CO₂ world. Proceedings of the National Academy of Sciences, 105: 10450–10455.
- Marshall, K. N., Kaplan, I. C., Hodgson, E. E., Hermann, A., Busch, D. S., McElhany, P., Essington, T. E., *et al.* 2017. Risks of ocean acidification in the California current food web and fisheries: ecosystem model projections. Global Change Biology, 23: 1525–1539.
- Martin, S., Rodolfo-Metalpa, R., Ransome, E., Rowley, S., Buia, M.-C., Gattuso, J.-P., and Hall-Spencer, J. 2008. Effects of naturally acidified seawater on seagrass calcareous epibionts. Biology Letters, 4: 689–692.
- McCulloch, M., Falter, J., Trotter, J., and Montagna, P. 2012. Coral resilience to ocean acidification and global warming through pH up-regulation. Nature Climate Change, 2: 623–627.
- McElhany, P. 2017. CO₂ sensitivity experiments are not sufficient to show an effect of ocean acidification. ICES Journal of Marine Science, 74: 926–928.
- Miller, A. W., Reynolds, A. C., Sobrino, C., and Riedel, G. F. 2009. Shellfish face uncertain future in high CO₂ world: influence of acidification on oyster larvae calcification and growth in estuaries. PLoS One, 4: e5661.
- Mollica, N. R., Guo, W., Cohen, A. L., Huang, K.-F., Foster, G. L., Donald, H. K., and Solow, A. R. 2018. Ocean acidification affects coral growth by reducing skeletal density. Proceedings of the National Academy of Sciences of the United States of America, 115: 1754–1759.
- Mongin, M., Baird, M. E., Hadley, S., and Lenton, A. 2016. Optimising reef-scale CO_2 removal by seaweed to buffer ocean acidification. Environmental Research Letters, 11: 034023.
- Morse, J. W., and Mackenzie, F. T. 1990. Geochemistry of Sedimentary Carbonates. Elsevier, New York. 707 pp.
- Mostofa, K. M. G., Liu, C.-Q., Zhai, W., Minella, M., Vione, D., Gao, K., Minakata, D., *et al.* 2016. Reviews and Syntheses: ocean acidification and its potential impacts on marine ecosystems. Biogeosciences, 13: 1767–1786.
- Moy, A. D., Howard, W. R., Bray, S. G., and Trull, T. W. 2009. Reduced calcification in modern Southern Ocean planktonic foraminifera. Nature Geoscience, 2: 276–280.
- Moya, A., Howes, E. L., Lacoue-Labarthe, T., Forêt, S., Hanna, B., Medina, M., Munday, P. L., *et al.* 2016. Near-future pH conditions severely impact calcification, metabolism and the nervous system in the pteropod *Heliconoides inflatus*. Global Change Biology, 22: 3888–3900.
- Munday, P. L., Warner, R. R., Monro, K., Pandolfi, J. M., and Marshall, D. J. 2013. Predicting evolutionary responses to climate change in the sea. Ecology Letters, 16: 1488–1500.
- Newton, J., Chai, F., and Dai, M. 2019. Progress and planning in understanding ocean acidification. Eos, 100: doi: 10.1029/2019eo128617.
- Ohman, M. D., Lavaniegos, B. E., and Townsend, A. W. 2009. Multi-decadal variations in calcareous holozooplankton in the California Current System: thecosome pteropods, heteropods, and foraminifera. Geophysical Research Letters, 36: C03038.
- Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. A., Gnanadesikan, A., *et al.* 2005. Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. Nature, 437: 681–686.
- Osborne, E. B., Thunell, R. C., Gruber, N., Feely, R. A., and Benitez-Nelson, C. R. 2020. Decadal variability in twentieth-century ocean acidification in the California Current Ecosystem. Nature Geoscience, 13, 43–49.

- Pacella, S. R., Brown, C. A., Waldbusser, G. G., Labiosa, R. G., and Hales, B. 2018. Seagrass habitat metabolism increases short-term extremes and long-term offset of CO₂ under future ocean acidification. Proceedings of the National Academy of Sciences of the United States of America, 115: 3870–3875.
- Pandori, L. L. M., and Sorte, C. J. B. 2019. The weakest link: sensitivity to climate extremes across life stages of marine invertebrates. Oikos, 128: 621–629.
- Parker, L. M., O'Connor, W. A., Byrne, M., Coleman, R. A., Virtue, P., Dove, M., Gibbs, M., et al. 2017a. Adult exposure to ocean acidification is maladaptive for larvae of the Sydney rock oyster *Saccostrea glomerata* in the presence of multiple stressors. Biology Letters, 13: 20160798.
- Parker, L. M., O'Connor, W. A., Raftos, D. A., Pörtner, H.-O., and Ross, P. M. 2015. Persistence of positive carryover effects in the oyster, *Saccostrea glomerata*, following transgenerational exposure to ocean acidification. PLoS One, 10: e0132276.
- Parker, L. M., Ross, P. M., and O'Connor, W. A. 2010. Comparing the effect of elevated pCO₂ and temperature on the fertilization and early development of two species of oysters. Marine Biology, 157: 2435–2452.
- Parker, L. M., Ross, P. M., and O'Connor, W. A. 2011. Populations of the Sydney rock oyster, *Saccostrea glomerata*, vary in response to ocean acidification, Marine Biology, 158: 689–697.
- Parker, L. M., Ross, P. M., O'Connor, W. A., Borysko, L., Raftos, D. A., and Pörtner, H.-O. 2012. Adult exposure influences offspring response to ocean acidification in oysters. Global Change Biology, 18: 82–92.
- Parker, L. M., Scanes, E., O'Connor, W. A., Coleman, R. A., Byrne, M., Pörtner, H.-O., and Ross, P. M. 2017b. Ocean acidification narrows the acute thermal and salinity tolerance of the Sydney rock oyster *Saccostrea glomerata*. Marine Pollution Bulletin, 122: 263–271.
- Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. Annual Review of Ecology, Evolution, and Systematics, 37: 637–669.
- Peck, V. L., Oakes, R. L., Harper, E. M., Manno, C., and Tarling, G. A. 2018. Pteropods counter mechanical damage and dissolution through extensive shell repair. Nature Communications, 9: 264.
- Peck, V. L., Tarling, G. A., Manno, C., Harper, E. M., and Tynan, E. 2016. Outer organic layer and internal repair mechanism protects pteropod *Limacina helicina* from ocean acidification. Deep Sea Research Part II: Topical Studies in Oceanography, 127: 41–52.
- Pecl, G. T., Araújo, M. B., Bell, J. D., Blanchard, J., Bonebrake, T. C., Chen, I.-C., Clark, T. D., *et al.* 2017. Biodiversity redistribution under climate change: impacts on ecosystems and human wellbeing. Science, 355: eaai9214.
- Plummer, L. N., and Busenberg, E. 1987. Thermodynamics of aragonite-strontianite solid solutions: results from stoichiometric solubility at 25 and 76°C. Geochimica et Cosmochimica Acta, 51: 1393–1411.
- Putnam, H. M., Davidson, J. M., and Gates, R. D. 2016. Ocean acidification influences host DNA methylation and phenotypic plasticity in environmentally susceptible corals. Evolutionary Applications, 9: 1165–1178.
- Putnam, H. M., and Gates, R. D. 2015. Preconditioning in the reef-building coral *Pocillopora damicornis* and the potential for trans-generational acclimatization in coral larvae under future climate change conditions. The Journal of Experimental Biology, 218: 2365–2372.
- Reimer, J. J., Wang, H., Vargas, R., and Cai, W.-J. 2017. Multidecadal fCO₂ increase along the United States southeast coastal margin. Journal of Geophysical Research: Oceans, 122: 10061–10072.
- Riebesell, U., Aberle-Malzahn, N., Achterberg, E. P., Algueró-Muñiz, M., Alvarez-Fernandez, S., Arístegui, J., Bach, L. T., *et al.* 2018. Toxic algal bloom induced by ocean acidification disrupts the pelagic food web. Nature Climate Change, 8: 1082–1086.

- Riebesell, U., Bach, L. T., Bellerby, R. G. J., Rafael Bermúdez Monsalve, J., Boxhammer, T., Czerny, J., Larsen, A., *et al.* 2017. Competitive fitness of a predominant pelagic calcifier impaired by ocean acidification. Nature Geoscience, 10: 19–23.
- Riebesell, U., and Gattuso, J.-P. 2015. Lessons learned from ocean acidification research. Nature Climate Change, 5: 12–14.
- Ruesink, J. L., Sarich, A., and Trimble, A. C. 2018. Similar oyster reproduction across estuarine regions differing in carbonate chemistry. ICES Journal of Marine Science, 75: 340–350.
- Ryu, T., Veilleux, H. D., Donelson, J. M., Munday, P. L., and Ravasi, T. 2018. The epigenetic landscape of transgenerational acclimation to ocean warming. Nature Climate Change, 8:504–509.
- Sanford, E., Gaylord, B., Hettinger, A., Lenz, E. A., Meyer, K., and Hill, T. M. 2014. Ocean acidification increases the vulnerability of native oysters to predation by invasive snails. Proceedings of the Royal Society B: Biological Sciences, 281: 20132681.
- Schaum, C. E., and Collins, S. 2014. Plasticity predicts evolution in a marine alga. Proceedings of the Royal Society B: Biological Sciences, 281: 20141486.
- Seibel, B. A., Maas, A. E., and Dierssen, H. M. 2012. Energetic plasticity underlies a variable response to ocean acidification in the pteropod, *Limacina helicina antarctica*. PLoS One, 7: e30464.
- Shamberger, K. E. F., Cohen, A. L., Golbuu, Y., McCorkle, D. C., Lentz, S. J., and Barkley, H. C. 2014. Diverse coral communities in naturally acidified waters of a Western Pacific Reef. Geophysical Research Letters, 41: 499–504.
- Shamberger, K. E. F., Feely, R. A., Sabine, C. L., Atkinson, M. J., DeCarlo, E. H., Mackenzie, F. T., Drupp, P. S., *et al.* 2011. Calcification and organic production on a Hawaiian coral reef. Marine Chemistry, 127: 64–75.
- Shamberger, K. E. F., Lentz, S. J., and Cohen, A. L. 2018. Low and variable ecosystem calcification in a coral reef lagoon under natural acidification. Limnology and Oceanography, 63: 714–730.
- Shaw, E. C., McNeil, B. I., and Tilbrook, B. 2012. Impacts of ocean acidification in naturally variable coral reef flat ecosystems. Journal of Geophysical Research: Oceans, 117: 1–14.
- Silbiger, N. J., and Donahue, M. J. 2015. Secondary calcification and dissolution respond differently to future ocean conditions. Biogeosciences, 12: 567–578.
- Silbiger, N. J., Donahue, M. J., and Brainard, R. E. 2017. Environmental drivers of coral reef carbonate production and bioerosion: a multi-scale analysis. Ecology, 98: 2547–2560.
- Silbiger, N. J., Guadayol, Ò., Thomas, F. I. M., and Donahue, M. J. 2014. Reefs shift from net accretion to net erosion along a natural environmental gradient. Marine Ecology Progress Series, 515: 33–44.
- Silbiger, N. J., Guadayol, O., Thomas, F. I. M., and Donahue, M. J. 2016. A novel μCT analysis reveals different responses of bioerosion and secondary accretion to environmental variability. PLoS One, 11: e0153058.
- Silbiger, N. J., Nelson, C. E., Remple, K., Sevilla, J. K., Quinlan, Z. A., Putnam, H. M., Fox, M. D., *et al.* 2018. Nutrient pollution disrupts key ecosystem functions on coral reefs. Proceedings of the Royal Society B: Biological Sciences, 285: 20172718.
- Silbiger, N. J., and Sorte, C. J. B. 2018. Biophysical feedbacks mediate carbonate chemistry in coastal ecosystems across spatiotemporal gradients. Scientific Reports, 8: 796.
- Silverman, J., Lazar, B., and Erez, J. 2007. Effect of aragonite saturation, temperature, and nutrients on the community calcification rate of a coral reef. Journal of Geophysical Research, 112: C05004.
- Silverman, J., Schneider, K., Kline, D. I., Rivlin, T., Rivlin, A., Hamylton, S., Lazar, B., *et al.* 2014. Community calcification in Lizard Island, Great Barrier Reef: a 33 year perspective. Geochimica et Cosmochimica Acta, 144: 72–81.
- Strader, M. E., Wong, J. M., Kozal, L. C., Leach, T. S., and Hofmann, G. E. 2019. Parental environments alter DNA methylation in

offspring of the purple sea urchin, *Strongylocentrotus purpuratus*. Journal of Experimental Marine Biology and Ecology, 517: 54–64.

- Sunday, J. M., Bates, A. E., and Dulvy, N. K. 2012. Thermal tolerance and the global redistribution of animals. Nature Climate Change, 2, 686–690.
- Sutton, A. J., Feely, R. A., Maenner-Jones, S., Musielwicz, S., Osborne, J., Dietrich, C., Monacci, N., *et al.* 2019. Autonomous seawater pCO₂ and pH time series from 40 surface buoys and the emergence of anthropogenic trends. Earth System Science Data, 11: 421–439. Copernicus GmbH.
- Sutton, A. J., Sabine, C. L., Feely, R. A., Cai, W.-J., Cronin, M. F., McPhaden, M. J., Morell, J. M., *et al.* 2016. Using present-day observations to detect when anthropogenic change forces surface ocean carbonate chemistry outside preindustrial bounds. Biogeosciences, 13: 5065–5083.
- Sutton, A. J., Wanninkhof, R., Sabine, C. L., Feely, R. A., Cronin, M. F., and Weller, R. A. 2017. Variability and trends in surface seawater pCO_2 and CO_2 flux in the Pacific Ocean. Geophysical Research Letters, 44: 5627–5636.
- Takahashi, M., Noonan, S. H. C., Fabricius, K. E., and Collier, C. J. 2016. The effects of long-term in situ CO₂ enrichment on tropical seagrass communities at volcanic vents. ICES Journal of Marine Science, 73: 876–886.
- Talmage, S. C., and Gobler, C. J. 2009. The effects of elevated carbon dioxide concentrations on the metamorphosis, size, and survival of larval hard clams (*Mercenaria mercenaria*), bay scallops (*Argopecten irradians*), and Eastern oysters (*Crassostrea virginica*). Limnology and Oceanography, 54: 2072–2080.
- Thabet, A. A., Maas, A. E., Lawson, G. L., and Tarrant, A. M. 2015. Life cycle and early development of the thecosomatous pteropod *Limacina retroversa* in the Gulf of Maine, including the effect of elevated CO₂ levels. Marine Biology, 162: 2235–2249.
- Thibodeau, P. S., Steinberg, D. K., Stammerjohn, S. E., and Hauri, C. 2018. Environmental controls on pteropod biogeography along the Western Antarctic Peninsula. Limnology and Oceanography, 64: S240–S256.
- Tilbrook, B., Jewett, E. B., DeGrandpre, M. D., Hernandez-Ayon, J. M., Feely, R. A., Gledhill, D. K., Hansson, L., *et al.* 2019. An enhanced ocean acidification observing network: from people to technology to data synthesis and information exchange. Frontiers in Marine Science, 6: 337.
- Torda, G., Donelson, J. M., Aranda, M., Barshis, D. J., Bay, L., Berumen, M. L., Bourne, D. G., *et al.* 2017. Rapid adaptive responses to climate change in corals. Nature Climate Change, 7: 627–636.
- Tribollet, A., Godinot, C., Atkinson, M., and Langdon, C. 2009. Effects of elevated pCO_2 on dissolution of coral carbonates by microbial euendoliths. Global Biogeochemical Cycles, 23: doi: 10.1029/2008gb003286.
- Turk, D., Wang, H., Hu, X., Gledhill, D. K., Wang, Z. A., Jiang, L., and Cai, W.-J. 2019. Time of emergence of surface ocean carbon dioxide trends in the North American coastal margins in support of ocean acidification observing system design. Frontiers in Marine Science, 6: 91.
- Unsworth, R. K. F., Collier, C. J., Henderson, G. M., and McKenzie, L. J. 2012. Tropical seagrass meadows modify seawater carbon chemistry: implications for coral reefs impacted by ocean acidification. Environmental Research Letters, 7: 024026.

- Vergés, A., Steinberg, P. D., Hay, M. E., Poore, A. G. B., Campbell, A. H., Ballesteros, E., Heck, K. L., *et al.* 2014. The tropicalization of temperate marine ecosystems: climate-mediated changes in herbivory and community phase shifts. Proceedings of the Royal Society B: Biological Sciences, 281: 20140846.
- Waldbusser, G. G., Brunner, E. L., Haley, B. A., Hales, B., Langdon, C. J., and Prahl, F. G. 2013. A developmental and energetic basis linking larval oyster shell formation to acidification sensitivity: Larval shell and acidification. Geophysical Research Letters, 40: 2171–2176.
- Waldbusser, G. G., Hales, B., Langdon, C. J., Haley, B. A., Schrader, P., Brunner, E. L., Gray, M. W., *et al.* 2014. Saturation-state sensitivity of marine bivalve larvae to ocean acidification. Nature Climate Change, 5: 273–280.
- Wall-Palmer, D., Hart, M. B., Smart, C. W., Sparks, R. S. J., Le Friant, A., Boudon, G., Deplus, C., *et al.* 2012. Pteropods from the Caribbean Sea: variations in calcification as an indicator of past ocean carbonate saturation. Biogeosciences, 9: 309–315.
- Wall-Palmer, D., Smart, C. W., and Hart, M. B. 2013. In-life pteropod shell dissolution as an indicator of past ocean carbonate saturation. Quaternary Science Reviews, 81: 29–34.
- Watson, S.-A., Southgate, P. C., Tyler, P. A., and Peck, L. S. 2009. Early larval development of the Sydney rock oyster *Saccostrea glomerata* under near-future predictions of CO₂-driven ocean acidification. Journal of Shellfish Research, 28: 431–437.
- Waycott, M., Duarte, C. M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., Calladine, A., *et al.* 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. Proceedings of the National Academy of Sciences of the United States of America, 106: 12377–12381.
- Weisberg, S. B., Bednaršek, N., Feely, R. A., Chan, F., Boehm, A. B., Sutula, M., Ruesink, J. L., *et al.* 2016. Water quality criteria for an acidifying ocean: challenges and opportunities for improvement. Ocean & Coastal Management, 126: 31–41.
- Welladsen, H. M., Southgate, P. C., and Heimann, K. 2010. The effects of exposure to near-future levels of ocean acidification on shell characteristics of *Pinctada fucata* (Bivalvia: Pteriidae). Molluscan Research, 30: 125–130.
- Willoughby, J. R., Harder, A. M., Tennessen, J. A., Scribner, K. T., and Christie, M. R. 2018. Rapid genetic adaptation to a novel environment despite a genome-wide reduction in genetic diversity. Molecular Ecology, 27: 4041–4051.
- Wisshak, M., Schönberg, C. H. L., Form, A., and Freiwald, A. 2012. Ocean acidification accelerates reef bioerosion. PLoS One, 7: e45124.
- Wong, J. M., Johnson, K. M., Kelly, M. W., and Hofmann, G. E. 2018. Transcriptomics reveal transgenerational effects in purple sea urchin embryos: adult acclimation to upwelling conditions alters the response of their progeny to differential pCO₂ levels. Molecular Ecology, 27: 1120–1137.
- Wright, J. M., Parker, L. M., O'Connor, W. A., Williams, M., Kube, P., and Ross, P. M. 2014. Populations of pacific oysters *Crassostrea gigas* respond variably to elevated CO₂ and predation by *Morula marginalba*. Biological Bulletin, 226: 269–281.
- Zimmerman, R. C., Kohrs, D. G., Steller, D. L., and Alberte, R. S. 1997. Impacts of CO₂ enrichment on productivity and light requirements of eelgrass. Plant Physiology, 115: 599–607.

Handling editor: Howard Browman