

nostic modalities in the acute stem cell transplant period. A normal CXR does not definitively exclude a pulmonary process in this group of patients and additional CT evaluation should be considered, particularly in patients with pulmonary symptoms.

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HUMAN POLYOMAVIRUS BK AND JC INFECTION IN CHILDREN AFTER HEMATOPOIETIC STEM CELL TRANSPLANTATION: ANALYSIS OF RISK FACTORS CONTRIBUTING TO VIRUSES REACTIVATION AND HEMORRHAGIC CYSTITIS

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Polyomaviruses (HPV: BKV or JCV)-induced late-onset hemorrhagic cystitis (HC) is a significant cause of morbidity after hematopoietic stem cell transplantation (HSCT). However, issues facilitating HPV reactivation and related HC remain unclear. The aim of the study was to evaluate the incidence, risk factors and clinical implications of BKV and JCV infection in children after HSCT. One hundred forty two children (121 after allo- and 21 after auto-HSCT) were prospectively evaluated for BKV/JCV infection and HC. Viral DNA was identified in plasma and urine with qualitative PCR. The PCR products were cleaved with the enzyme BamHI to discriminate between BKV and JCV sequences. HC diagnosis was based on the clinical symptoms, the presence of hematuria and USG of the bladder. Cumulative incidence (CI) curves were created to assess the risk of BKV or JCV infection. Multivariate analysis of potential risk factors of HPV reactivation and HC among allografted patients (Age, gender, diagnosis, donor, cell source, CMV reactivation, conditioning, GvHD and its prophylaxis) was performed with Fine and Grey model. RESULTS: HPV viremia was detected in 6 (28.5%) of 21 autografted patients, in 5 BKV (23.8%) and 1 JCV (4.8%), only 1 patient developed HC. No HPV viremia was observed. Following allo-HSCT, HPV were found in 65 (53,7%) children. BKV viremia was detected in 60 (49,5%) and was accompanied by BKV viremia in 15 patients. JCV viremia was discovered in 4 (3.3%) children with concomitant JCV viremia in 1 patient. In 1 child (0.8%) parallel BKV and JCV viremia and viremia were found. The CI of HPV viremia was higher among children after allo-HSCT, however the difference was insignificant ($p=0.07$). In univariate analysis, the following significant risk factors of HPV viremia were identified: age >10 years, HSCT from MUD and ATG in the conditioning. In multivariate analysis age remained the only significant. Multivariate analysis of potential risk factors of HC in children with HPV viremia revealed that HPV viremia and chemotherapy-based conditioning increased the risk significantly, while TBI appeared to do the opposite. CONCLUSIONS: BKV viremia is common in children after allo-HSCT. The risk of HPV shedding increases with age and this indirectly indicates reactivation rather than de novo infection. HC after allo-HSCT is induced by HPV in conjunction with viremia and chemotherapy-based conditioning. However, the role of additional factors can not be excluded.

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THERAPEUTIC MONITORING OF SIROLIMUS IS ESSENTIAL IN PEDIATRIC BMT RECIPIENTS

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Limited data on sirolimus (SRL) in adult BMT patients is consistent with a long elimination half-life ($t_{1/2}$: 57-63 h) and little

effect of an occasional missed dose on overall drug exposure. Pharmacokinetic (PK) studies in pediatric organ transplant recipients suggest that SRL may be metabolized more rapidly in children. BMT patients may have variable SRL exposure due to gastrointestinal toxicity, and interactions with calcineurin inhibitors and imidazole antifungal drugs.

SRL PK profiles were evaluated in 19 pediatric BMT recipients with high-risk ALL treated on a pilot trial using SRL-based GVHD prophylaxis. Mean age at transplant was 11 y (4-21 y); 13 males, 6 females; 14 Whites, 4 Hispanic, 1 Black. Stem cell source: 12 unrelated cord blood, 6 sibling bone marrow, and 1 unrelated PBSC. Preparative regimen: TBI 1200 cGy, thiotepa 10mg/kg, and cyclophosphamide 120mg/kg. GVHD prophylaxis: continuous infusion tacrolimus from day -2, IV methotrexate (5mg/m², 4-5 doses) and oral SRL (2.5 mg/m²/d; 4mg/d max) from day -2 (1), day -1 (1), day 0 (16), and day 1 (1). Out of 119 SRL doses, 14 were followed by emesis, and 10 of these were redosed. Sixteen patients also received fluconazole prophylaxis.

Timed multiple blood samples (8-9) were drawn after sixth (14), seventh (5), or eighth (1) dose of SRL. Whole blood SRL concentrations were measured by liquid chromatography with tandem-mass spectrometry. Mean SRL dose was 2.6 mg/day (1-4); 2.3 mg/day/m² (1.1-4). In 20 PK studies, SRL concentrations normalized to dose (mean \pm SD, ng/mL/mg) were: trough level before the dose (C_0) 3.50 ± 2.40 , maximum (C_{max}) 6.74 ± 5.37 , and 24 hours later (C_{24}) 3.29 ± 2.38 . C_0 and C_{24} were similar and correlated well ($r^2 = 0.905$), indicating achievement of steady state. SRL dose did not correlate with AUC ($r^2 = 0.12$), while C_0 and C_{24} correlated well with AUC ($r^2 = 0.918$; 0.929 , respectively). SRL apparent oral clearance was 4.1 ± 4.5 ml/min/kg (0.87-20) and $t_{1/2}$ 25 ± 13 h (9-71 h). SRL $t_{1/2}$ in our patient group were markedly lower than those reported in adults, and comparable to those observed in pediatric liver and small bowel transplant recipients on tacrolimus and SRL (19.3 to 21.2 h). In conclusion, we show that GVHD prophylaxis with SRL is feasible even in young children, and PK data supports 24 h dosing interval; wide individual variability and lack of correlation of dose with AUC supports the need for SRL therapeutic monitoring and dose adjustments based on steady state blood levels.

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DEFIBROTIDE FOR THE TREATMENT OF SINUSOIDAL OBSTRUCTION SYNDROME (SOS) FOLLOWING HEMATOPOIETIC STEM CELL TRANSPLANT IN PEDIATRIC PATIENTS

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Sinusoidal Obstruction Syndrome (SOS) is characterized by endothelial cell damage in the liver and associated with significant morbidity and mortality. It is a well documented complication of abdominal irradiation, specific chemotherapeutic agents, monoclonal antibodies and hematopoietic stem cell transplant (HSCT). Defibrotide is an experimental polydeoxyribonucleotide agent with liver endothelial cell activity that has clinically encouraging results in the treatment of SOS. In this study, we report a single institution retrospective analysis of the efficacy of defibrotide in treating SOS in a post-HSCT pediatric population. A total of 11 patients were treated with defibrotide after being clinically diagnosed with severe SOS having a >30% likelihood of mortality based on the Bearman model of life expectancy. The protocol was IRB approved and each patient was individually assigned an FDA IND number prior to treatment. All patients had received ursodiol for SOS prophylaxis. Seven of 11 patients received TBI-based conditioning and 4 patients received ablative chemotherapy alone. The median age was 6.6 years at time of treatment (range, 5 months to 18 yrs). All patients were Caucasian, with 4 males and 7 females. The stem cell source was unrelated cord blood in 4 patients, unrelated bone marrow in 2, autologous peripheral blood in 2, and related bone marrow in 3. Engraftment (ANC >500) occurred on average 23.2 days post-SCT (mean: Cord = 30 days, BM = 22.6 days, autologous = 11 days). 10 of 11 patients had primary malignancies (AML, ALL, neuroblastoma, pineoblastoma) while 1 patient had

Wolman's Syndrome. SOS was clinically diagnosed and treatment began between 7 and 37 days post-HSCT (mean=17 days). Patients were treated per protocol with a starting dose of 10 mg/kg/day with subsequent increases of 10 mg/kg/day until clinically therapeutic to a max of 60 mg/kg/day (exception: < 2 y/o population allowed a maximum of 100 mg/kg/day, n=1). The duration of defibrotide treatment ranged from 13 to 31 days (mean=18 days). There were no significant complications related to the defibrotide infusions or therapy. Seven of 11 (63.6%) patients showed complete response to the therapy, while 4 patients were classified as non-responders. The 100 day post-HSCT survival rate was 54.5% (6/11) for all patients. In conclusion, this study provides further evidence that defibrotide is an efficacious and safe treatment for pediatric SOS occurring after HSCT.

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VIRUS-SPECIFIC T CELLS ENGINEERED TO CO-EXPRESS TUMOR-SPECIFIC RECEPTORS; EFFECTS IN PATIENTS WITH NEUROBLASTOMA

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Long-term survival for children with neuroblastoma remains poor despite intensification of therapy. Neuroblastoma cells express a number of potential target antigens for the immune response including GD2 and T cells engineered to express chimeric antigen receptors (CAR) for these antigens might supplement conventional therapeutics. Unfortunately, modification of primary T cells with CAR has demonstrated limited functionality and persistence in vivo, likely because the target tumor cells lack the co-stimulatory molecules required for T cell activation. We have previously shown that Epstein Barr Virus specific cytotoxic T lymphocytes (EBV-CTLs) persist and function in vivo long term, in part because they receive co-stimulation from EBV infected B lymphocytes. We reasoned that EBV-CTLs engineered to express GD2-CAR would also receive such co-stimulatory signals following engagement of their native (EBV-specific) receptors, and hence would persist longer than primary T cells expressing the same CAR. We therefore developed a Phase I clinical study to directly compare EBV CTL-CAR and T-cell CAR. We used two retroviral vectors distinguishable by a non-expressed marker sequence so that signal detected in vivo could be assigned to the CTL or the T cells. Vector use was alternated between the two cell populations. To date 3 patients received 2×10^7 /m² of each cell population, while 4 have received 1×10^8 /m² of each product. We observed no adverse effects. Even at the low cell doses used to date, signal could be detected for 6 weeks in peripheral blood. Differential quantitative PCR analysis of the signal showed that EBV-CTL CAR were present at a higher level and persisted longer than primary T cell associated CAR. Clinically there has been one mixed response and two with stable disease, while one patient treated in CR remains disease free at 12 months. 4 patients have died of progressive disease. Expressing chimeric antigen receptors in virus specific T cells appears to improve lymphocyte survival after adoptive transfer compared to CAR expression in primary T cells, and may thereby increase the therapeutic potential of these cells. It will be of interest to use this approach early after stem cell transplantation, when residual malignant cells are at their lowest level, and homeostatic signals will favor expansion of adoptively transferred engineered CTL.

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EFFECTIVE IMMUNOTHERAPY FOR NEUROBLASTOMA REQUIRES HSCT AND T CELL TRANSFER

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We have employed a mouse model for neuroblastoma, AGN2a cells injected subcutaneously in to strain A/J mice, to demonstrate that transfection of AGN2a with CD80, CD86, CD54, and CD137L transforms this lethal cell line in to an effective cell-based vaccine (AGN2a-4P). When tumor vaccines are administered in the context of Treg blockade (with anti-CD25 antibody), the anti-tumor effect is heightened. Our previous work also demonstrated that the inclusion of CD137L in the vaccine preparation is uniquely responsible for a strong anti-tumor T cell response. However, when we switched from tumor-challenge to tumor-bearing regimens the vaccine was ineffective. Even the inclusion of gene expression vectors encoding GM-CSF, IL-15, lymphotactin, or SLC did not inhibit tumor progression. In order to determine if any impact could be made in tumor-bearing animals, we initiated a tumor-bearing model system featuring HSCT. Analysis of lymphoid reconstitution post-HSCT revealed that up to day 21, mice remained severely lymphopenic. However, it was during this time period that vaccination with AGN2a-4P proved most effective in a tumor-challenge experiment. Moreover, vaccination early post-HSCT was markedly improved when T cells were adoptively transferred 3 days after HSCT, just prior to the initiation of AGN2a-4P vaccination. ELISPOT data with purified CD4 and CD8 cells indicated that adoptively transferred lymphocytes early post-conditioning generated an IFN- γ -producing tumor-specific effector population. Furthermore, splenic reconstitution studies at day 21 clearly indicated that animals given lymphocyte adoptive transfer had greater percentages of donor (as opposed to residual host) T cells in their spleens. This indicates that combining adoptive T cell transfer with HSCT alters the lymphocyte populations present early post-HSCT, and that these populations are crucial to the generation of anti-tumor effector cells. We then tested our post-HSCT vaccine strategy in a tumor-bearing model. 1×10^5 live tumor cells were given on day -8, TBI on day -1, HSCT consisting of bone marrow plus 6×10^6 T cells on day 0, and irradiated AGN2a-4P vaccine on days 2 and 7. Effective neuroblastoma therapy required HSCT, T cell transfer and AGN2a-4P vaccination. Our results support the current practice of autologous HSCT for neuroblastoma therapy, and suggest that the addition of vaccination and adoptive immunotherapy (T cell transfer) to these regimens may improve outcomes.

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LOW RATES OF TOXICITY AND LONG-TERM RESPONSES AFTER BU/FLU/ATG RIC ALLOGENEIC TRANSPLANTATION IN VERY HIGH RISK PEDIATRIC PATIENTS INELIGIBLE FOR MYELOBLASTIC THERAPY: A PEDIATRIC BLOOD AND MARROW TRANSPLANT CONSORTIUM STUDY
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The role of reduced intensity conditioning (RIC) regimens in pediatric pts is unclear. To define the feasibility and toxicity of this approach using multiple stem cell sources in pediatric pts ineligible for myeloablative transplant, we conducted a trial at 25 centers participating in the Pediatric Blood and Marrow Transplant Consortium (PBMTTC). Thirty two pediatric pts (age 2-20yrs, med 12) have been enrolled to date with the following stem cell sources: 7 related donor (RD) BM (2 mismatched); 5 RD-PBSC; 9 unrelated donor (UD) BM; 4UD-PBSC; 7 UD-CB. Qualifying indications included a previous allogeneic (n=15) or autologous (n=4) transplant, severe organ toxicity (n=5), invasive fungal infection (n=2), and other comorbidities, (n= 6 pts, 4 with Down syndrome). Diagnoses included ALL (4 CR2; 9 CR3), AML (6 CR2; 3 CR3; 1 secondary), MLL (1 CR3), HD (2 CR3), B-NHL (1 PR3), MDS (2 RA;