

The strophomenide brachiopod *Ahtiella* Öpik in the Ordovician of Gondwana and the early history of the plectambonitoids

Juan L. Benedetto

Centro de Investigaciones en Ciencias de la Tierra (CICTERRA), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) and Universidad Nacional de Córdoba, Vélez Sarsfield 1611, X5016GCA Córdoba, Argentina (juan.benedetto@unc.edu.ar)

Abstract.—The Precordilleran species *Ahtiella argentina* Benedetto and Herrera, 1986 is redescribed and illustrated and *Monorthis coloradoensis* Benedetto, 1998 from northwestern Argentina is reassigned to the genus *Ahtiella* Öpik, 1932. *Ahtiella famatiniana* new species from volcanoclastic rocks of the Famatina range (western Argentina) and *Ahtiella tunaensis* new species from the Precordillera basin (Cuyania terrane) are proposed. Paleogeographic and stratigraphic evidence strongly suggests that *Ahtiella* originated in the Andean region of Gondwana to further migrate to Avalonia, Baltica, and Cuyania. Contrary to previous assumptions, the fossil record from the Famatina volcanoclastic succession suggests that the plectambonitoid *Ahtiella famatiniana* n. sp. evolved from the hesperonomiid orthoid *Monorthis transversa* Benedetto, 2003 that always occurs in the underlying strata. Phylogenetic analysis of *Ahtiella* species shows that *A. famatiniana* n. sp. and the Peruvian *A. zarelæ* Villas in Gutiérrez-Marco and Villas, 2007 are not only the earliest species of the genus but also are morphologically intermediate between *Monorthis* Bates, 1968 and the later and more derived species of *Ahtiella* from Baltica and Cuyania. If, as empirical evidence presented here shows, *Ahtiella* originated from *Monorthis* through a series of minor transformations, then the impressive morphological gap between orthoides and strophomenides was bridged through short-time cladogenesis events, suggesting that it might not have a definite discontinuity between the species level evolution and the origin of higher taxa (macroevolution).

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Introduction

The genus *Ahtiella* Öpik, 1932 is a distinctive resupinate and variably geniculate plectambonitoid brachiopod described originally from the Baltic region (Öpik, 1932, 1933; Hessland, 1949) but subsequently recognized in Wales (Bates, 1968) and central Newfoundland (Neuman, 1976). In South America, *Ahtiella* is very common in the lower–middle Darrivilian carbonate-ramp deposits of the Precordillera basin of west-central Argentina where it defines the uppermost of the six brachiopod biozones recognized through the San Juan Formation (Herrera and Benedetto, 1991; Benedetto, 2002, 2007). Later, *Ahtiella* was reported from the Floian-Dapingian volcanoclastic succession of the Famatina Range (Benedetto et al., 2003; Benedetto 2003a) but these specimens remain undescribed. Its presence in southern Peru (Gutiérrez-Marco and Villas, 2007), together with its record in the central Andean Basin of northwestern Argentina (this paper) and probably Bolivia (described as *Valcourea* sp. by Havlíček and Branisa, 1980), indicate that this genus not only attained a wide geographic range in South America but also experienced a significant speciation event encompassing at least five species. As Gutiérrez-Marco and Villas (2007) pointed out, the records of *Ahtiella* in the Floian of Peru and Dapingian of Famatina are the oldest known of the genus, strongly suggesting that it originated on the Andean margin of Gondwana and later migrated to other regions.

One of the objectives of this study is to update the taxonomy of the genus *Ahtiella* from the three major Ordovician basins of Argentina: Precordillera, Famatina, and Central Andes. This includes: (1) the redescription of the Precordilleran species *Ahtiella argentina* Benedetto and Herrera, 1986, on the basis of extensive collections made in the past twenty years from the upper part of the San Juan Formation, as well as the proposal of a new species of *Ahtiella* from the somewhat younger Las Chacritas Formation; (2) the first description of the *Ahtiella* specimens from volcanoclastic rocks of the Famatina Range; and (3) the reassignment to the genus *Ahtiella* of *Monorthis coloradoensis* Benedetto, 1998b, from northwestern Argentina.

Evidence presented here aims to shed light on the long-standing and not yet resolved issue of the origin of plectambonitoid brachiopods. Although parsimony analysis constitutes an indispensable tool for unravelling the phylogeny of fossil groups, the most difficult task is to corroborate in the fossil record the phyletic lineages predicted in such analyses, and even more problematic is to detect those morphological transitions leading to the origin of new taxa. According to the punctuated equilibrium hypothesis (Eldredge and Gould, 1972; Gould and Eldredge, 1977; Benton and Pearson, 2001), this can be explained by the conjunction of the rapidity as cladogenesis events occur and the relatively small size and geographic restriction of populations undergoing phenotypic change. In this respect, the continuous and richly fossiliferous volcanosedimentary succession of the

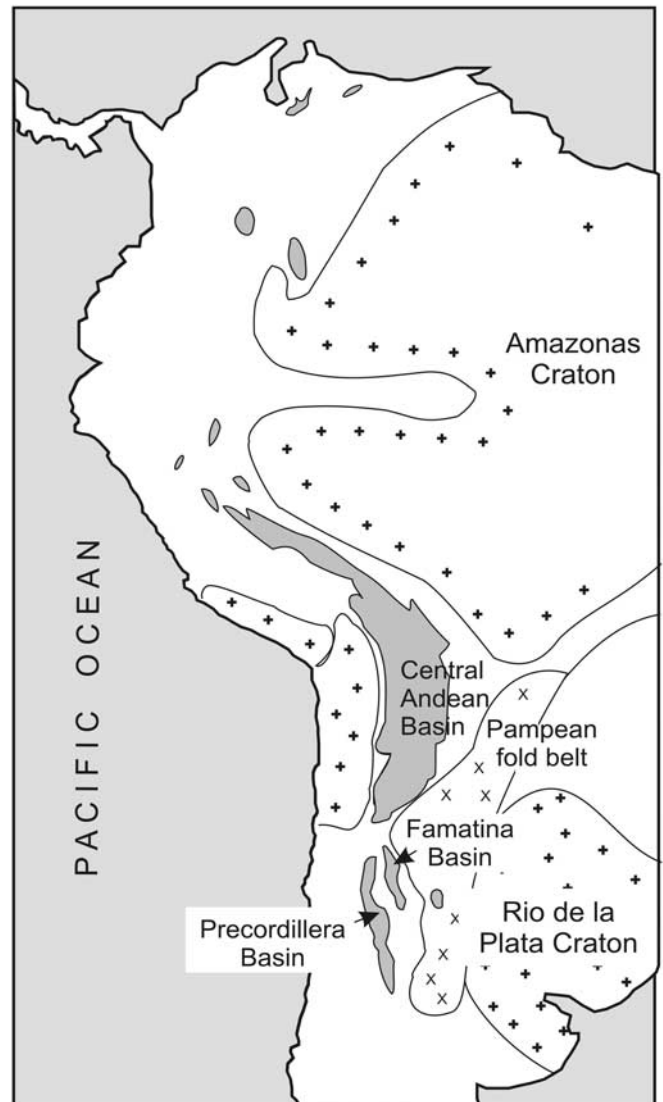
75 Famatina Range provides an invaluable frame to establish well-
 76 resolved phylogenies based on the fossil record. In this paper,
 77 evidence is presented suggesting that *Ahtiella* originated from the
 78 hesperonomiid orthoid *Monorthis transversa* Benedetto, 2003b,
 79 which always occurs in strata below those bearing *Ahtiella*
 80 *famatiniana* new species (described herein). A general trend of
 81 morphological change through time emerges from the comparative
 82 morphology of *Ahtiella* species and its putative ancestor
 83 *Monorthis*. Relevant for our phylogenetic hypothesis is the fact
 84 that the earliest species of *Ahtiella* recorded in Gondwana exhibit
 85 transitional characteristics between *Monorthis* and the typical
 86 *Ahtiella* species from younger strata of Cuyania and Baltica.
 87 Finally, a phylogenetic analysis is presented herein to investigate
 88 the evolutionary relationships among the Gondwanan species of
 89 *Ahtiella* and those from Cuyania, Baltica, and Avalonia.

90 Stratigraphic provenance and age

91 The early Paleozoic geodynamic history of southern South
 92 America involved three main sedimentary domains (Fig. 1),
 93 which were inhabited at different times by species of *Ahtiella*.
 94 They are: (1) the autochthonous Central Andean Basin developed
 95 around the Brazilian craton through Peru, Bolivia, and
 96 northwestern Argentina; (2) the volcanosedimentary Famatina
 97 Basin, which together with the Puna volcanic arc developed
 98 peripheral to the active pre-Andean Gondwana margin; and
 99 (3) the Precordillera Basin developed on the Laurentian-derived
 100 Cuyania terrane, which accreted to Gondwana during the early
 101 Paleozoic (for a comprehensive review of the Ordovician basins
 102 of Argentina, see Astini, 2003).

103 *Central Andean Basin*.—The southern part of the large Central
 104 Andean Basin is widely exposed in the Cordillera Oriental of
 105 northwestern Argentina, where the uppermost Cambrian to
 106 Lower Ordovician siliciclastic succession of the Santa Victoria
 107 Group attains 3,800 m in thickness. In the study area of Los
 108 Colorados (Fig. 2), it is overlain by a succession of reddish
 109 purple sandstones named the Alto del Cóndor Formation, which
 110 is succeeded by fossiliferous greenish mudstones and marls.
 111 This interval, which was referred to as the ‘Green Member of the
 112 Sepulturas Formation’ by Astini (1994) and as the Sepulturas
 113 Formation by Astini et al. (2004a), is well exposed at Quebrada
 114 Chamarra and Quebrada del Cardonal (Fig. 2). Brachiopods
 115 consist of *Monorthis coloradoensis* (reassigned herein to
 116 *Ahtiella*) and rare specimens of *Paralenorthis* sp., *Dinorthis*?
 117 sp., and small dalmanellids. Trilobites are represented by
 118 *Neseuretus* sp., a trinucleid of the *Anebolithus-Incaia* group
 119 (personal communication, B.G. Waisfeld, 2017), and a new
 120 species of *Hoekaspis*, the latter recorded elsewhere in the Sub-
 121 andean Ranges of northwestern Argentina from beds not older
 122 than the upper Darriwilian (Waisfeld and Vaccari, 2003).
 123 Albanesi and Astini (2002) reported from interbedded
 124 carbonate-rich layers a conodont assemblage consisting of
 125 *Erraticodon*, *Erismodus*, and *Plectodina*, as well as micro-
 126 remains of the agnathan *Sacabambaspis* considered of late
 127 Darriwilian age.

128 *Famatina Basin*.—The Famatina Range is characterized by a
 129 thick succession of sedimentary and volcanosedimentary rocks



136 **Figure 1.** Map of central-western South America showing the main
 137 Ordovician sedimentary basins discussed in the text (gray shading).

138 deposited in a retroarc basin almost synchronously with the
 139 emplacement of crust-derived magmatism along the proto-
 140 Andean margin (Pankhurst et al., 2001; Dahlquist et al., 2005).
 141 Marine intervals are well exposed to the north of the basin in the
 142 Chaschuil area (Catamarca Province), and to the south in the
 143 Cachiuyo-Saladillo rivers area (La Rioja Province) (Fig. 3). In
 144 the Chaschuil area, a regressive volcanosedimentary sequence
 145 accumulated on a high-gradient narrow platform flanking the
 146 volcanic chain (Mángano and Buatois, 1996, 1997). Its lower
 147 part, ~150 m thick, was referred to the Loma del Kilómetro
 148 Member of the Suri Formation; it has been interpreted as
 149 deposited in a storm- and mass flow-dominated shelf, whereas
 150 the upper Punta Pétreá Member is a coarse-grained volcani-
 151 clastic wedge that records the progradation of a fan delta system
 152 onto the shelf sediments. The Dapingian age of the Loma del
 153 Kilómetro Member, containing *Monorthis transversa* (dis-
 154 cussed below) and other brachiopods (Benedetto, 1994), is
 155 based on conodonts of the *Baltoniodus navis* Biozone (Albanesi
 156 and Vaccari, 1994) and the *Baltoniodus triangularis* Biozone

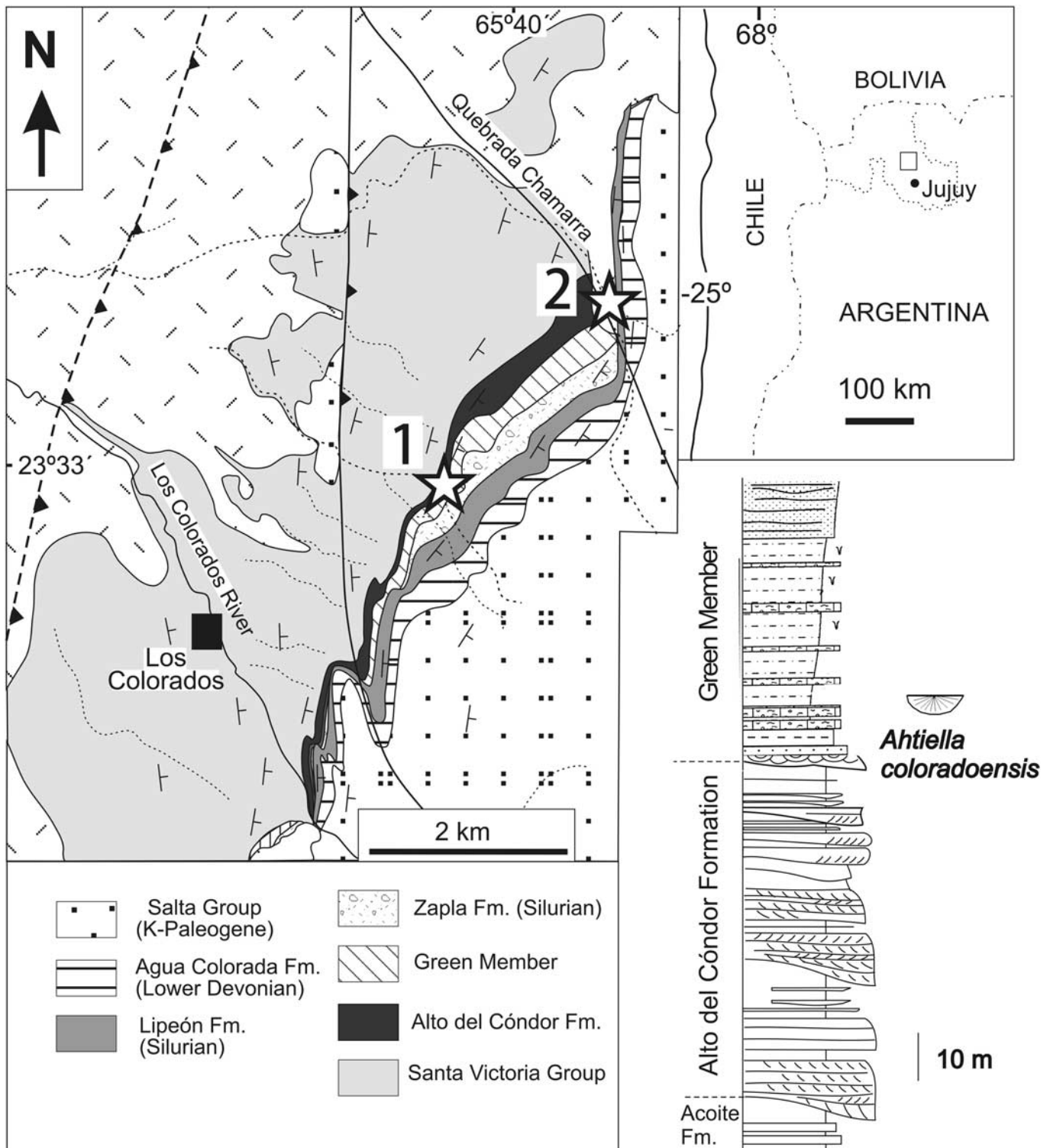


Figure 2. Geological map of the Los Colorados area of the Central Andean Basin (modified from Astini et al., 2004a) showing fossil locations (stars) and integrated stratigraphic column of the study area showing levels yielding *Ahtiella coloradoensis*. 1 = Quebrada Chamarra; 2 = Quebrada Cardonal.

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(Carlorosi et al., 2017) recovered from the upper shell beds of this member. That age is consistent with the Whitlandian British regional stage suggested by the underlying trilobite fauna (Vaccari and Waisfeld, 1994). The Punta Pétreá Member, which crops out between Agua del Médano and Puesto Chaschuil, yielded an undescribed species of *Famatinorthis* and *Ahtiella*

famatiniana n. sp. (this paper). This succession is partially interbedded with (Cisterna, 2001), or is unconformably overlain by (Astini and Dávila, 2002), the Sierra de Las Planchadas rhyolite, which provided a $^{206}\text{Pb}/^{238}\text{U}$ SHRIMP age of 468.3 ± 3.4 Ma (Baldo et al., 2003), i.e., close to the Dapingian-Darriwilian boundary.

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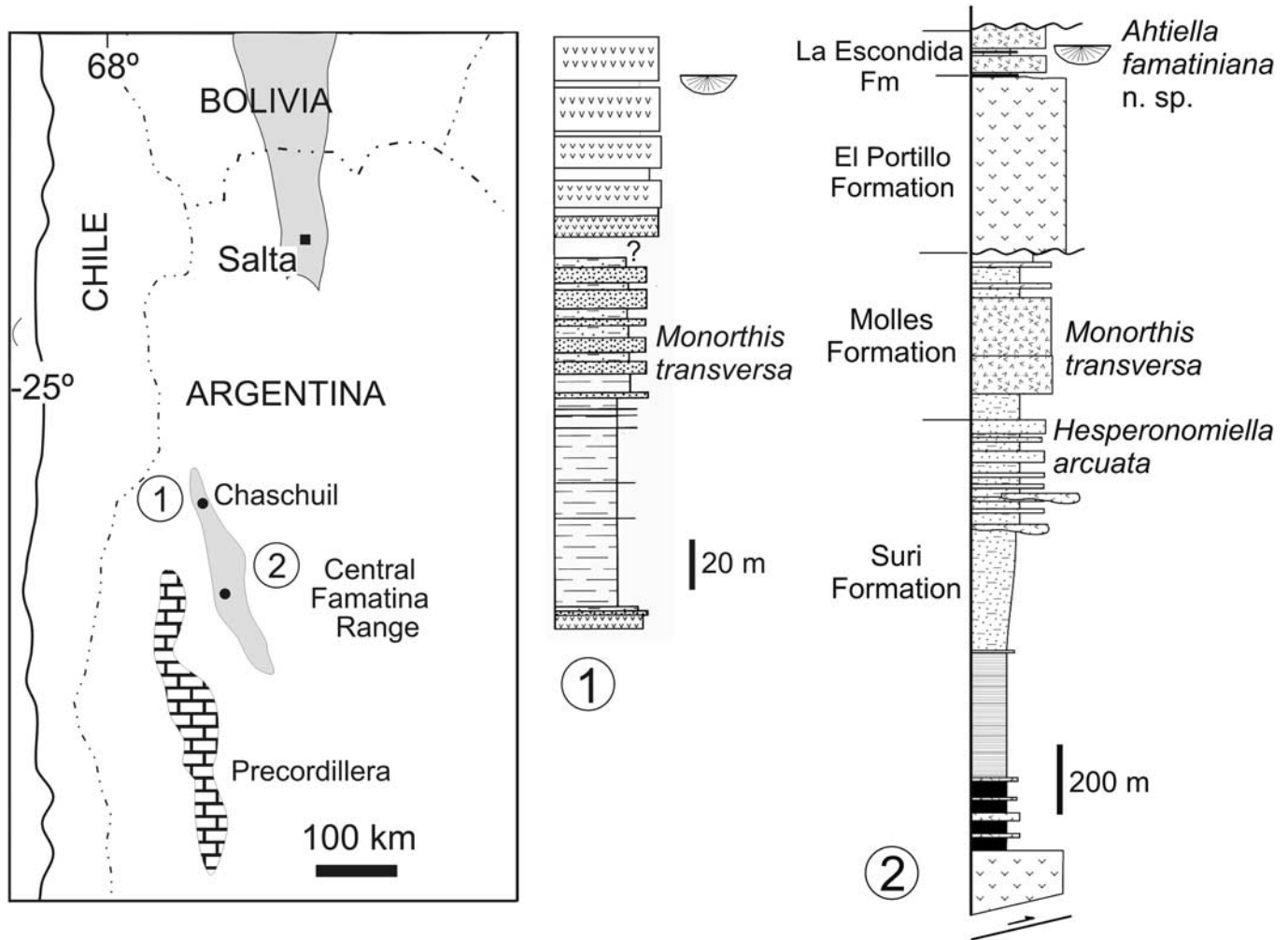


Figure 3. Location of the Famatina Range showing sampled areas (circled numbers), stratigraphic columns, and fossiliferous horizons. 1 = Chaschuil River area; 2 = Central Famatina Range (Cachiyuyo-Saladillo rivers area).

168 The thicker and more continuous volcanosedimentary
 169 succession exposed in the central Famatina Range has been
 170 referred to the Famatina Group, encompassing the Suri and
 171 Molles formations (Harrington and Leanza, 1957). Both units
 172 are profusely fossiliferous, containing rhynchonelliform
 173 (Benedetto, 2003b, 2008, 2013) and lingulate brachiopods
 174 (undescribed), bivalves (Sánchez and Babin, 1993; Sánchez,
 175 1997), trilobites (Harrington and Leanza, 1957; Vaccari and
 176 Waisfeld, 1994), and graptolites (Toro and Brussa, 1997). The
 177 up to 2,000 m thick succession reflects deposition on a high-
 178 gradient, mixed siliciclastic-volcaniclastic platform (Astini and
 179 Benedetto, 1996; Astini, 1999, 2003; Mángano et al., 2002).
 180 The Suri Formation displays a shallowing-upward trend ranging
 181 from dark shales deposited in relatively deep oxygen-deficient
 182 waters to shallow platform facies. The Molles Formation
 183 consists of fossiliferous grey mudstones alternating with
 184 silicified tuffs, volcanogenic sandstones, and reddish sandstones
 185 with evidence of tidal action. Concerning the age, the lower part
 186 of the Suri Formation yielded graptolites that indicate the Floian
 187 *Baltograptus deflexus* and *Didymograptellus bifidus* biozones
 188 (Toro and Brussa, 1997), whereas the shell beds from the top of

the Suri Formation and the base of the overlying Molles
 Formation contain conodonts referable to the upper part of the
Oepikodus evae Biozone of Floian age (late F12 stage slice of
 Bergström et al., 2009) (Albanesi and Astini, 2000). Lehnert
 et al. (1997) reported from the Molles Formation conodonts of
 the *Paroistodus originalis* Biozone of middle Dapingian age.

The Molles Formation is unconformably overlain by a
 720 m thick volcanosedimentary succession named the Cerro
 Morado Group, which starts with acidic volcanics and
 ignimbrites (Portillo Formation) and culminates with silicified
 tuffs, pyroclastic flows, and bioturbated fossiliferous green
 shales interbedded with coquina layers (La Escondida Forma-
 tion) (Astini and Dávila, 2002) (Fig. 3). The fauna is dominated
 by *Famatinorthis turneri* (Benedetto, 2013), a distinctive taxon
 of the underlying Molles Formation. A volcanosedimentary unit
 probably equivalent to the La Escondida Formation crops out to
 the north of the Cachiyuyo River at Las Pircas anticline. These
 levels yielded an undescribed *Skenidioides?* sp., *Paralenorthis*
 sp., and *Ahtiella* sp. (Benedetto et al., 2003), the last described
 herein as *A. famatiniana* n. sp. The age of this interval, in
 absence of conodont data, is still poorly constrained, but given

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210 that in the Chaschuil area the levels of *A. famatiniana* n. sp. are
211 interbedded with isotopically dated volcanic rocks (see above),
212 a middle-late Dapingian age seems most likely.

213 *Precordillera Basin (Cuyania terrane)*.—There is a general
214 agreement that the up to 2,500 m thick, passive-margin carbo-
215 nate succession started to accumulate during the Cambrian
216 Series 2 on the Laurentian continental margin, a segment of
217 which subsequently rifted from the Ouachita Embayment ori-
218 ginating the Cuyania terrane (Astini et al., 1995; Thomas and
219 Astini, 1996, 2003; Astini, 1998; Benedetto, 1998a, 2004;
220 Ramos, 2004; but see Finney, 2007, for a para-autochthonous
221 Gondwanan hypothesis). Cuyania became part of the Gond-
222 wana continent since its accretion to the proto-Andean margin
223 by Middle Ordovician or more probably Late Ordovician times.

224 Depositional environments evolved from tidal flats, shoals,
225 shallow subtidal settings, and restricted subtidal settings during
226 the Cambrian – early Tremadocian, to open shelf settings by late
227 Tremadocian–Darriwilian times (Cañas, 1999; Keller, 1999,
228 2012; Gómez and Astini, 2015). The carbonate succession
229 referred to the San Juan Formation consists of ~350 m of
230 burrowed skeletal wackestones and packstones capped by a
231 25–30 m thick interval of mid-outer ramp nodular limestones
232 bearing a rich benthic fauna dominated by rhynchonelliform
233 brachiopods and sponges, with trilobites, bryozoans, gastro-
234 pods, crinoids, and linguliforms as secondary components
235 (Carrera, 2003; Waisfeld et al., 2003; Sorrentino et al., 2009;
236 Carrera and Ernst, 2010; Lavié and Benedetto, 2016) (Fig. 4).
237 The brachiopod association from these beds encompasses the
238 *Ahtiella argentina* Biozone, which is the uppermost of the six
239 biozones recognized through the San Juan Formation (Herrera
240 and Benedetto, 1991; Benedetto, 2002, 2007). This interval is
241 particularly well exposed along the western slope of Cerro
242 Viejo, ~20 km northeast of the city of San José de Jáchal, where
243 the San Juan Formation forms a westward-dipping homoclinal
244 succession. The *A. argentina* specimens described herein were
245 collected mainly at Quebrada Los Gatos and the adjacent
246 Quebrada Honda stratigraphic sections from a 10–12 m thick
247 package of nodular limestones lying immediately below the
248 contact with the Los Azules Formation black shales (Fig. 4).
249 The age of this interval is well constrained by conodonts of the
250 *Paroistodus horridus* Subzone within the *Lenodus variabilis*
251 Biozone (Albanesi and Ortega, 2002; Ortega et al., 2007) and
252 the lower part of the succeeding *Yangtzeplacognathus crassus*
253 Biozone (Mestre and Heredia, 2013; Serra et al., 2015).
254 According to the time-slices schema proposed by Bergström
255 et al. (2009), the *A. argentina* beds fall mostly within Dw1
256 reaching the lower part of Dw2.

257 At Sierra de la Trampa, near 40 km south of the city of San
258 José de Jáchal, a ~60 m thick succession of nodular limestones
259 crops out at Quebrada Las Chacritas and Quebrada La Tuna
260 (Fig. 4). This package was originally referred by Espisúa (1968)
261 to the ‘upper member’ of the San Juan Formation, then to the
262 ‘Las Tunas calcareous unit’ by Carrera (1997), and finally to the
263 Las Chacritas Formation by Astini (1998), which was formally
264 defined by Peralta et al. (1999). The thin bedded wackestones,
265 bioclastic grainstones, and mudstones of the Las Chacritas
266 Formation have yielded rich poriferan assemblages (Carrera,
267 1997), excellently silicified trilobite larval stages (Waisfeld

et al., 2001, and references therein), and numerous brachiopods
268 not yet described, including *Skenidioides?* sp. and *Ahtiella*
269 *tunaensis* new species (this paper). Several conodont studies led
270 to the recognition the *Y. crassus* Zone in the lower part of the
271 Las Chacritas Formation, the *Eoplacognathus pseudoplanus*
272 Biozone from 36 m above the base, and the *Eoplacognathus*
273 *suecicus* Biozone near the top (Albanesi and Astini, 2000;
274 Heredia et al., 2011; Mestre and Heredia, 2012, 2013; Serra
275 et al., 2015). Accordingly, *A. tunaensis* n. sp. can be confidently
276 dated as middle Darriwilian (Dw2).
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278 **Remarks on the biogeography of *Ahtiella* and related** 279 **ahtieillins**

280 The earliest known representative of the genus is *Ahtiella zarelae*
281 Villas in Gutiérrez-Marco and Villas, 2007 from the upper Floian
282 San José Formation of southern Peru (Gutiérrez-Marco et al.,
283 2008). The slightly younger *A. famatiniana* n. sp. occurs in vol-
284 caniclastic rocks of the Famatina Range of middle-late Dapingian
285 age. In Anglesey (northwestern Wales), *Ahtiella* is represented by
286 *A. quadrata* Bates, 1969, from the *Expansograptus hirundo*
287 Biozone, which in the Atlantic Province encompasses the
288 Dapingian and the base of Darriwilian (Zalasiewicz et al., 2009),
289 and *A. concava* Bates, 1969, from the slightly younger Bob Dei-
290 niol Formation. *Ahtiella paucirugosa* Neuman, 1976 has been
291 reported from the lower Darriwilian Summerford Group of New
292 World Island, central Newfoundland. These volcanoclastic rocks
293 were interpreted as recording intra-Iapetus volcanic islands related
294 to the Avalonian paleocontinent (Neuman, 1976, 1984; Neuman
295 and Harper, 1992; Harper et al., 1996). *Monorthis coloradoensis*
296 occurs in the Cordillera Oriental of northwestern Argentina from
297 beds probably not older than mid-Darriwilian. *Ahtiella* is common
298 in the carbonate or mixed carbonate-clastic rocks of Baltica and
299 Cuyania. In Sweden and Estonia, *Ahtiella* encompasses the
300 Kunda and Asseri regional stages, the latter reaching the middle
301 Darriwilian *Pterograptus elegans* Biozone and *E. suecicus* Zone
302 (Tolmacheva et al., 2003; Suyarkova and Koren, 2009). As sta-
303 ted above, in the Precordillera basin, *Ahtiella* ranges from the
304 lower Darriwilian (*A. argentina*) to the middle Darriwilian (*A.*
305 *tunaensis* n. sp.) but does not reach the *E. suecicus* Zone. Such a
306 distribution led Gutiérrez-Marco and Villas (2007) to infer that
307 *Ahtiella* migrated eastward from the mid-latitude (~30–40°)
308 Andean region into Avalonia and Baltica, and simultaneously
309 moved into the low-latitude Cuyania taerrane. It should be noted
310 that Gutiérrez-Marco and Villas (2007, fig. 8) adopted the
311 paleogeographic reconstruction of Aceñolaza et al. (2002) and
312 Finney (2007) and placed Cuyania into the gap delimited by
313 southern South America, South Africa, and Antarctica. Because
314 neither tectonostratigraphic (Astini and Rapalini, 2003; Ramos,
315 2004; Thomas et al., 2004) nor paleontological evidence (Ben-
316 edetto, 2004) supports such a para-autochthonous Gondwanan
317 origin, Cuyania is located here fairly closer to, and at approxi-
318 mately the same paleolongitude as the Famatina-Puna volcanic
319 arc (Fig. 5). Perhaps the major weakness—but not the only
320 one—of the Finney (2007) reconstruction is the complete
321 absence in the Cambrian carbonate rocks of the Precordillera of
322 Redlichiid-realm trilobites, which, as it is known, are distinctive
323 of Australasia and Antarctica. Instead, the Cambrian-
Tremadocian trilobites from Cuyania (Astini et al., 1995;

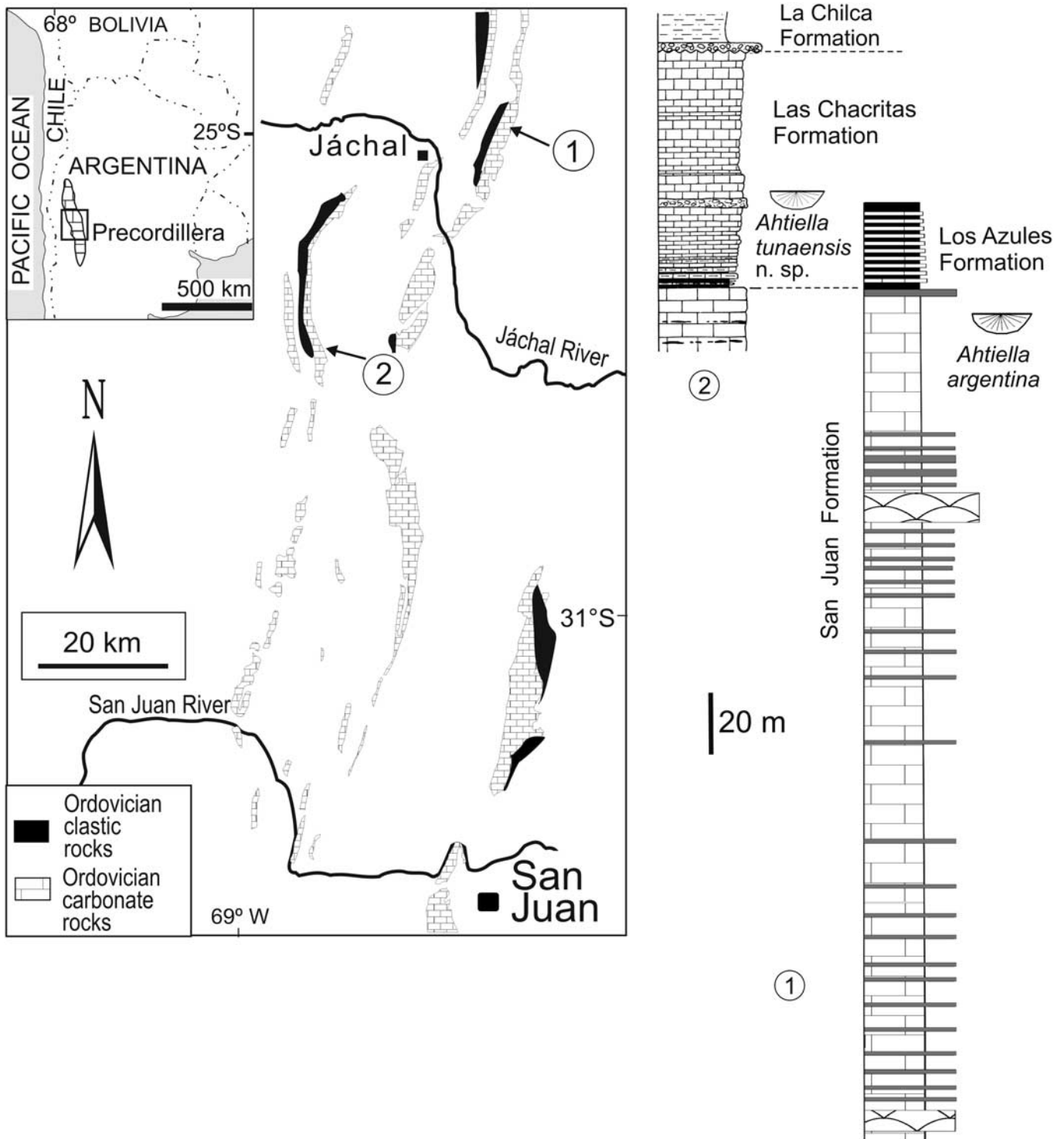


Figure 4. Outcrops of Ordovician rocks in the Precordillera mountain belt, sampled areas (circled numbers), and stratigraphic columns showing fossiliferous horizons. 1 = Cerro Viejo (Quebrada Los Gatos and Quebrada Honda); 2 = Sierra de la Trampa (Quebrada Las Chacritas and Quebrada La Tuna).

324 Vaccari, 1995; Benedetto, 2004; Benedetto et al., 2009), as well
 325 as brachiopods (Benedetto and Foglia, 2012) and other fossil
 326 groups (Carrera and Rigby, 1999; Carrera, 2003; Astini et al.,
 327 2004b) display indisputable Laurentian affinities. On the other
 328 hand, the Early Ordovician brachiopod faunas of Famatina—
 329 with a strong Celtic province signature (Benedetto, 2004; Harper
 330 et al., 2009)—share several genera (though not the same species)

with Cuyania, e.g., *Skenidiodes* Schuchert and Cooper, 1931;
Paralenorthis Havlíček and Branisa, 1980; *Productorthis*
 Kozłowski, 1927; *Monorthis* Bates, 1968; *Ffynnonia* Neuman
 and Bates, 1978; *Hesperonomia* Ulrich and Cooper, 1936;
Hesperonomiella Ulrich and Cooper, 1936; *Camerella* Billings,
 1859; and *Rugostrophia* Neuman, 1971. This indicates that by
 the Darriwilian, Cuyania was separated from Gondwana by a

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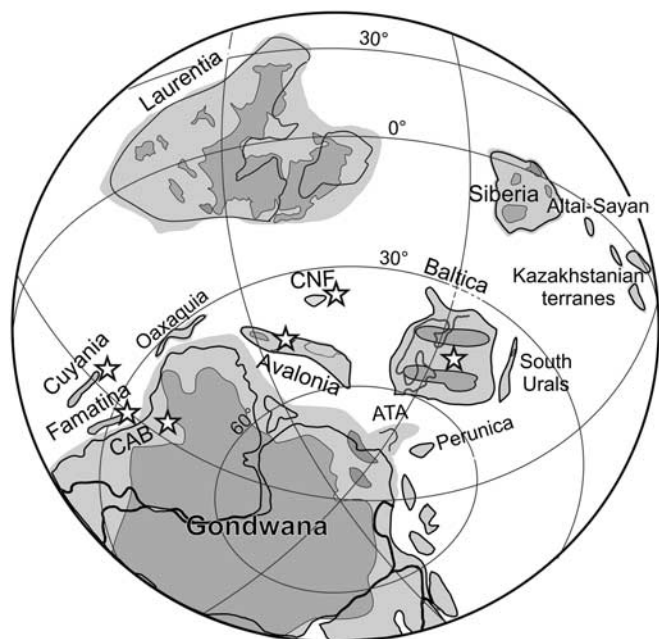


Figure 5. Early–Middle Ordovician paleogeography map (modified from Cocks and Torsvik, 2002 and Popov et al., 2009), showing global distribution of the genus *Ahtiella* (stars). Paleogeographic map. ATA = Armorican Terrane Assemblage; CAB = Central Andean Basin; CNF = Central Newfoundland (placement based on Neuman, 1984).

remnant ocean not large enough to prevent faunal dispersion (Benedetto et al., 2003; Benedetto, 2004). It seems likely that brachiopod dispersion from Famatina to Cuyania was facilitated by the gradual approximation of the Cuyania terrane to the Gondwana margin combined with a generalized sea-level rise (Carrera and Astini, 1998; Cañas, 1999; Astini, 2003).

An interesting feature is that diversification of the subfamily Ahtielinae was centered mainly in Avalonia, Cuyania, and Baltica (Fig. 5). The Welsh Treiorwerth Formation yielded *Inversella* (*Reinversella*) *monensis* Bates, 1969 (Neuman and Bates, 1978), whereas the Central Newfoundland Summerford Group contains the ahtielins *Schedophyla potteri* Neuman, 1971, *Inversella* sp., and the endemic *Guttasella gutta* Neuman, 1976. In the Cuyanian Precordillera Basin, *Ahtiella argentina* co-occurs with *I. (R.) arancibiai* Herrera and Benedetto, 1987 (Benedetto et al., 2008) and the endemic ahtielin *Sanjuanella plicata* Benedetto and Herrera, 1987. In Estonia, *Ahtiella lirata* Öpik, 1932 is approximately coeval with *I. (Inversella) borealis* Öpik, 1933. Outside the Baltic and Celtic faunal provinces, the only ahtielins reported are *Borua* Williams and Curry, 1985 from Ireland, and two species of *Schedophyla* Neuman, 1971 from southern China (Xu and Liu, 1984; Zhan et al., 2006). However, as noted below, the placement of *Schedophyla* among the ahtielins requires further confirmation. The Norwegian *Rutrumella* Harper in Bruton and Harper, 1981 is a poorly known genus that has been referred questionably to the subfamily (Cocks and Rong, 2000).

The Andean region as a center of origin

As Gutiérrez-Marco and Villas (2007) previously noted, and regardless of the chosen paleogeographic scenario, it is apparent that *Ahtiella* originated along the proto-Andean Gondwana

margin. Several recent paleontological discoveries provided evidence supporting that both the Central Andean Basin and the arc-related Puna-Famatina Basin operated simultaneously as centers of evolutionary radiation ('centers of origin') and species pump regions (sensu Harper et al., 2013) from which new taxa spread to neighboring areas (Benedetto and Sánchez, 2003; Muñoz and Benedetto, 2016; Benedetto and Muñoz, 2017). Such temperate Gondwana basins acted as sites of origination, as did the equatorial shallow-water shelves of Gondwana and peri-Gondwanan terranes, which have been identified by Bassett et al. (2002) as the main source of the precursors to the Ordovician radiation. For instance, the earliest known punctate orthide *Lipanorthis* Benedetto in Benedetto and Carrasco, 2002 from the upper Tremadocian of northwestern Argentina was not an immigrant from the tropical belt, as Harper et al. (2004) suggested, but probably originated from a *Protorthisina*-like plectorthoid ancestor inhabiting the Central Andean Basin in the latest Cambrian (Benedetto, 2013). Furthermore, based on cladistic analysis, Benedetto and Muñoz (2017) showed that plectorthoids not only underwent an important diversification in the Central Andean Basin during the Tremadocian and Floian but also could have been a source for the heterorthoids, which through the Ordovician spread along the western Gondwanan shelves (Peru, northern Africa) and peri-Gondwanan terranes (Avalonia, Armorica).

The Puna-Famatina volcanic arc (Fig. 5) was another significant center of origin during the Early to Middle Ordovician. As it has been already noted, its shelly faunas exhibit a high level of endemism, in particular bivalves (Sánchez and Babin, 1993; Sánchez, 1997) and brachiopods (Benedetto, 2003b; Benedetto and Sánchez, 2003). Volcanic islands and archipelagos have long been recognized as important evolutionary centers of modern biota (e.g., MacArthur and Wilson, 1967), but their role in promoting faunal diversification in the past was not fully acknowledged until Neuman (1984) proposed that the distinctive Celtic faunas from the Ordovician volcanoclastic rocks of the Caledonian-Appalachian folded belt inhabited intra-Iapetus volcanic islands. Also relevant was the subsequent study by Webby (1992) on the low-latitude Ordovician faunas from the volcanoclastic rocks of New South Wales. Harper et al. (2009) emphasized the role of such volcanic chains as cradles and centers of origin contributing to the increase of γ -diversity during the Great Ordovician Biodiversification Event.

Current ideas about the origin of Plectambonitoidea

The general statement that the order Strophomenida evolved from the early to middle Cambrian Nisusiidae of the class Kutorginata (Williams and Hurst, 1977) or, alternatively, from an ancestor similar to *Billingsella* Hall and Clarke, 1892 at the Cambrian-Ordovician transition, has been based essentially on the presence in all these groups of an apically perforated pseudodeltidium (Cocks and Rong, 1989; Williams et al., 1996). However, no further compelling evidence has been presented to support such ancestor-descendant relationships for all members of the order. According to Bassett et al. (2001), bilingsellides and kutorginates share the well-developed perforate pseudodeltidium and the lack of dental plates, but differ in that sockets and socket plates have a different origin in bilingsellides and

426 strophomenides, concluding that their phylogenetic links still
427 remain unclear. Subsequently, Bassett et al. (2008) and Bassett
428 and Popov (2017), based on a study of the ontogeny of the
429 orthotetide *Coolinia* Bancroft, 1949, inferred an early diver-
430 gence of strophomenates and rhynchonellate brachiopods.

431 At the superfamily level, it has been assumed that Stro-
432 phomenoidea was derived from the Plectambonitoidea during
433 the Early Ordovician. Spjeldnaes (1957) did not identify the
434 group of plectambonitoideans that gave rise to the strophome-
435 noideans and left open the possibility that the latter group is
436 polyphyletic. According to Cocks and Rong (1989, 2000),
437 strophomenoideans originated from plectambonitoideans by a
438 transformation of the cardinal process from simple to bifid,
439 suggesting as potential ancestor a leptellinid like *Apatomorpha*
440 Cooper, 1956 or *Toquimia* Ulrich and Cooper, 1936. Recent
441 discoveries demonstrated that the three basic types of stropho-
442 menoid cardinalia were already differentiated in the oldest
443 known members of the clade recorded in the Dapingian of
444 southern China (Zhan et al., 2015), supporting that the Stro-
445 phomenoidea originated in the Floian from an unknown ‘stro-
446 phomenide stem group’ shortly after the first appearance of
447 plectambonitoids. Dewing (2004) challenged the hypothesis of
448 the plectambonitoid derivation of strophomenoids based on
449 their different shell structure (laminar in the former and fibrous
450 in the latter). Unlike the a priori assumption that shell structure is
451 homoplastic (Cocks and Rong, 2000), i.e., evolved independ-
452 ently in different clades, Dewing (2004, fig. 3) proposed a
453 phylogenetic scenario in which the Strophomenoidea arose from
454 a Cambrian laminar-shelled billingselloid, whereas the common
455 ancestor of both the fibrous-shelled Plectambonitoidea and
456 Clitambonitoidea was left with interrogation. On the contrary,
457 the parsimony analysis performed by Congreve et al. (2015)
458 indicated that plectambonitoideans and strophomenoideans are
459 phylogenetically related but, in contrast to previous inferences,
460 plectambonitoideans do not constitute a monophyletic group but
461 a paraphyletic grade of the strophomenoidean clade. Signifi-
462 cantly, in the phylogeny presented by Congreve et al. (2015,
463 fig. 4), *Taffia* Butts, 1926, *Railtonella* Laurie, 1991, and
464 *Ahtiella*, all currently included in the family Taffiidae, cluster as
465 basal forms to all other Strophomenida, as Spjeldnaes (1957)
466 intuitively depicted in his phylogenetic tree sixty years earlier.

467 The absence or extreme paucity of undisputed plectambo-
468 nitoids in the Tremadocian, along with their sudden diversifi-
469 cation around the Floian-Dapingian transition, suggests that this
470 superfamily originated in the Early Ordovician rather than deep
471 in the Cambrian. In my opinion, billingselloids are too derived
472 morphologically to be considered direct ancestors of plectam-
473 bonitoids (excepting the family Plectambonitidae, as discussed
474 below). Since their first appearance in the middle Cambrian,
475 billingsellides developed a proportionally high planar ventral
476 interarea leading in the late Tremadocian to the hemipyramidal
477 shells that characterize most polytoechioids, e.g., *Protambo-*
478 *nites* Havlíček in Havlíček and Josopait, 1972 and *Tritoechia*
479 Ulrich and Cooper, 1936, which form a consistent monophyletic
480 clade (Benedetto, 2009; Topper et al., 2013). The apically per-
481 forated pseudodeltidium—the main feature linking bill-
482 ingsdelloids and plectambonitoids—could be a plesiomorphic
483 condition of basal rhynchonelliforms already present in some of
the earliest members of the clade (e.g., Nisusioidea) or,

alternatively, could be an homoplastic feature that appeared and
became lost at different times in different clades. In fact, in
certain basal plectambonitoids, e.g., *Plectella* Lamansky, 1905,
Ingria Öpik, 1930, *Aporthophyla* Ulrich and Cooper, 1936,
Tourmakeadia Williams and Curry, 1985, and *Pelonomia*
Cooper, 1956, the pseudodeltidium is rudimentary or lacking. In
any cases, this structure is not as phylogenetically informative as
previously supposed.

Noteworthy, the widely splayed, rodlike socket ridges
running almost parallel to the hinge line of billingselloids are
closely comparable to those of the family Plectambonitidae
(e.g., *Plectella*, *Plectambonites* Pander, 1830, and *Ingria*). Such
an arrangement is quite different from the typically orthoid
cardinalia seen in taffiids. Therefore, it is not surprising that in
the parsimony analysis carried out by Congreve et al. (2015),
Plectambonites appears as monophyletic only if it is excluded
from all other ‘plectambonitoids’ and placed in a separate
superfamily. In this context, it is worth noting the close resem-
blance between the billingsellide (?) *Kozhuchinella* Severgina,
1967 and the oldest known probable plectambonitoid *Akelina*
Severgina, 1967, both from the upper Tremadocian Algan For-
mation of Kuznetz-Altai, Altai Mountains, Siberia (Severgina,
1967). Despite the poor preservation of the latter (reillustrated
by Cocks and Rong, 1989, figs. 13–17), both genera share a
concavoconvex profile; parvicostellate ornamentation (typical
billingsellides are multicostellate or ramificostellate); the absence
of dental plates; a prominent dorsal median ridge; a simple
knob-like cardinal process; long, widely divergent socket rid-
ges; and a well-developed dorsal subperipheral rim. Accord-
ingly, *Akelina* and *Kozhuchinella* are likely related forms, which
could be considered either as early members of the ‘plectam-
bonitoid’ clade or, alternatively, as derived billingselloids (the
presence of pseudopunctae has not yet been demonstrated in
these genera). This raises the possibility that only the plectam-
bonitoid clade sensu stricto (the subfamily Plectambonitinae in
the current classification) evolved from a billingselloid ancestor,
and that ahtiellins (and probably other taffiids) had a different
ancestor, which should be sought among the Orthoidea, as
discussed below.

Searching for the *Ahtiella* ancestors

In his outstanding morphological study of Middle Ordovician
strophomenides from Norway, Spjeldnaes (1957, fig. 42) pre-
sented a diagrammatic evolutionary tree of Strophomenida
starting with two main branches, one of them lacking descen-
dants including *Plectambonites* and allied forms, and the other
including the ‘ahtiellinids’ (*Ahtiella*, *Inversella* Öpik, 1933, and
Ukoa Öpik, 1932), which albeit with a question mark, were
placed at the origin of the strophomenoid stock. A third, short-
lived basal branch was represented by *Taffia*. In their compre-
hensive revision of plectambonitoid classification, Cocks and
Rong (1989, p. 83, fig. 5) also placed the Taffiidae at the base of
the plectambonitoid tree “because we regard the absence of side
septa as representing a more primitive state than their presence
(as in the Plectambonitidae),” and also that the “oldest plec-
tambonitacean[s] ... are essentially indistinguishable from their
orthoide (probably billingselloid) ancestors except by their
pseudopunctate shell.” In fact, basal plectambonitoids share

with orthoids a simple, not undercut cardinal process, the absence of side septa, and the absence of a bema, a structure interpreted as a lophophore support and/or for providing muscle attachment. A more elaborated trifold, often undercut cardinal process, a variably elevated and bilobed bema, paired dorsal valve septa, hinge-line denticles, and more or less prominent internal papillae and/or septules are all features that gradually appear in different combinations in younger, more derived plectambonitoids.

Among taffiids, the ahtieillins—exemplified by the type genus *Ahtiella*—display such a combination of internal features that, in absence of pseudopunctae, they are virtually indistinguishable from certain basal orthoids (e.g., hesperonomiids). For instance, the inclusion among the ahtieillins of *Schedophyla potteri*, whose shell structure remains unknown, was cast in doubt by Cocks and Rong (1989, p. 97) who stated that “it is possible that the genus is an orthid.” Laurie (1991) also noted the orthoid dorsal cardinalia and musculature of *Schedophyla*. Recently Harper et al. (2017, p. 624) noted that “...questions remain regarding the placement of a number of groups such as the toquimiids, that apparently possess orthoid characters.” Perhaps the best example of such a difficulty is the conflicting taxonomic position of ‘*Monorthis*’ *coloradoensis*, from the Central Andean Basin, which is hardly differentiable from unquestionable species of *Ahtiella* such as *A. zarelae* Villas in Gutiérrez-Marco and Villas, 2007 and *A. famatiniana* n. sp. (described herein). Originally, the species ‘*coloradoensis*’ was ascribed to the hesperonomiid *Monorthis*, on the basis of its convexoplanar, slightly resupinate profile, carinate ventral fold, multicostellate ornamentation, and orthoid cardinalia and muscle scars (Benedetto, 1998b). Such an assignment was further supported by the lack of evidence of pseudopunctae and pseudodeltidium (Benedetto, 2003b), both considered apomorphic features of plectambonitoids. The shell structure, however, could not be verified because available shells are entirely decalcified, so that its nonpunctate condition was inferred from the lack of evidence of pseudopunctae on internal molds, even though many undisputable strophomenoids do not show internal traces of them.

Comparative morphology of *Monorthis* and *Ahtiella*

The orthoid ancestor hypothesis of *Ahtiella* finds empirical support from the comparison of the species *Monorthis transversa* (Fig. 6.1–6.12) and *Ahtiella famatiniana* n. sp. (Fig. 10.9–10.24), which occur in successive strata of the Famatina Basin.

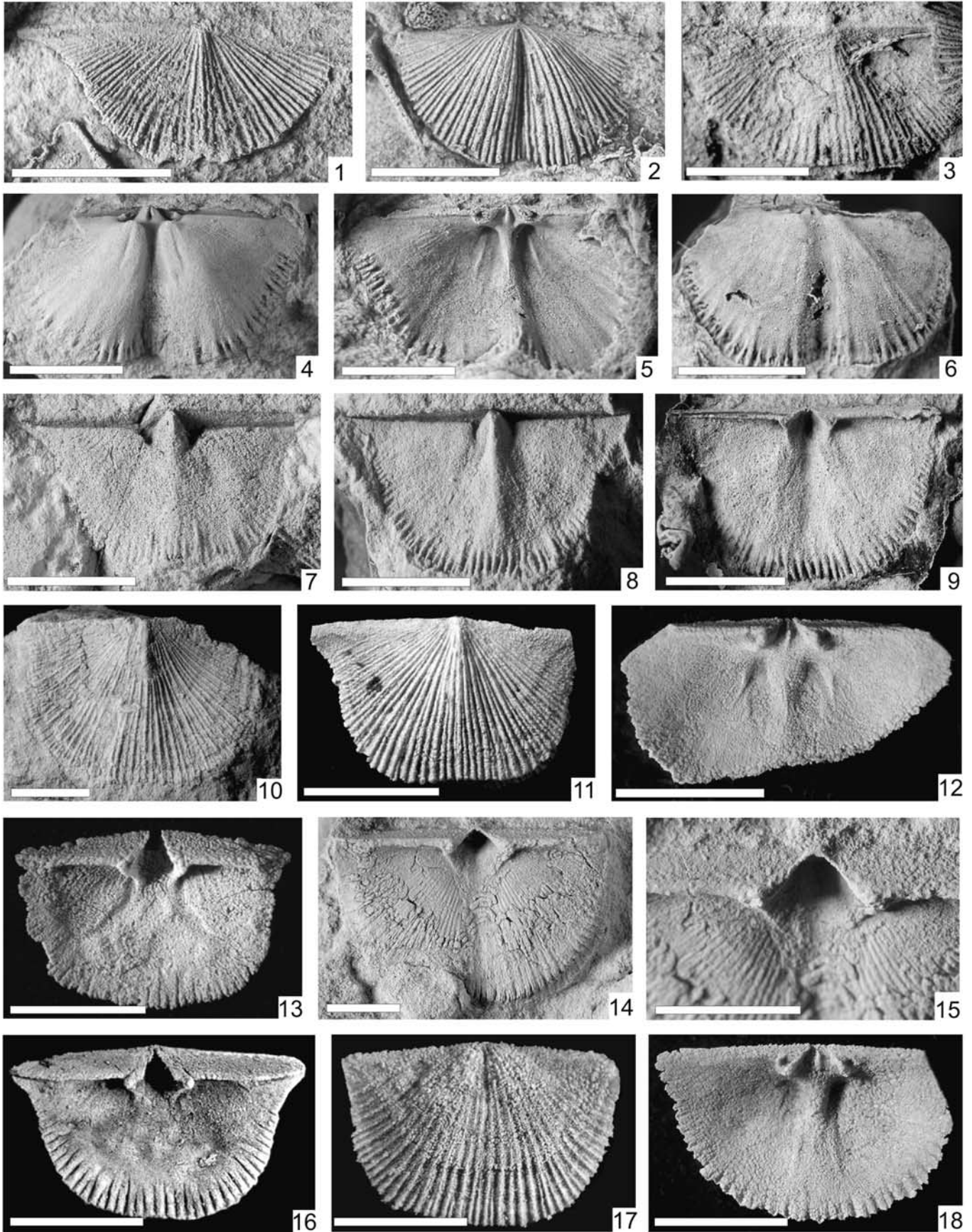
Shell shape and ornament.—Overall, the slightly resupinate, carinate, transversely elongate, alate shells of *Monorthis transversa* and *Ahtiella famatiniana* n. sp. are closely comparable. Shells of *M. transversa* are rather smaller and more transverse than *A. famatiniana*, resembling the juvenile to medium-sized specimens of the latter. Although ornamentation in *M. transversa* is slightly coarser than in *A. famatiniana* n. sp., it is subequally multicostellate in both species.

Pseudodeltidium.—Specimens of *Monorthis transversa* from Chaschuil—originally referred to as *Monorthis* aff. *M. menapiae* (Davidson, 1868) by Benedetto, 1994—and from Central

Famatina (Benedetto, 2003b) was described as having a widely open delthyrium. However, a slightly arched pseudodeltidium can be observed in a few conjoined shells (Fig. 6.1). Its absence in most specimens is likely due to postmortem breakage by hydrodynamic action, but could also reflect ecophenotypical variation within a population, ranging from absent to nearly complete according to specific (but unknown) environmental constraints. A careful revision of the Precordilleran specimens of *M. cumillangoensis* Benedetto, 2001, perhaps the better-known species of *Monorthis* worldwide (Benedetto, 2001), revealed the presence of an apparently imperforated pseudodeltidium covering the apical region of the delthyrium (Fig. 6.15). A closely comparable structure is also present in *Ahtiella coloradoensis* (Fig. 11.9). *Ahtiella zarelae* possesses a small apical pseudodeltidium that is hardly visible in the illustrated specimens (Gutiérrez-Marco and Villas, 2007, fig. 4E, F), but in *Ahtiella* sp. from slightly older strata of the same formation of Peru, it is better developed, almost attaining the delthyrium midlength. *Ahtiella famatiniana* n. sp. always possesses a well-developed pseudodeltidium covering one-half to two-thirds of the delthyrium (Fig. 10.9, 10.14). In the younger species *A. argentina* (Fig. 9.3) and *A. tunaensis* n. sp. (Fig. 10.5), the pseudodeltidium covers the pedicle opening almost entirely at all growth stages, suggesting a general trend from nearly absent to fully developed.

Muscle scars.—The ventral muscle field of *Monorthis transversa* is subtriangular and confined to the delthyrial cavity (Fig. 6.7) as in younger individuals of *Ahtiella famatiniana* n. sp. (Fig. 10.15). In mature specimens of the latter species (Fig. 10.16, 10.18), as well as in *A. coloradoensis* (Fig. 11.3, 11.8) and the Peruvian species *A. zarelae* (Fig. 11.18), the ventral muscle field becomes larger and more or less subpentagonal in outline. The Precordilleran *A. tunaensis* n. sp. is unique in having a large bilobed muscle field (Fig. 10.4). The dorsal muscle field of *M. transversa* is quadripartite, with anterior and posterior scars nearly equal in size. A persistent feature in most specimens of *M. transversa* is the presence of a pair of slightly divergent ridges bounding laterally or bisecting longitudinally the adductor field (Fig. 6.4, 6.5). Remarkably, these ridges are also present in *A. famatiniana* n. sp., *A. coloradoensis*, and *A. zarelae* (Figs. 10.23, 11.14). In the Welsh species *A. concava*, such ridges extend along the entire length of muscle scars (Bates, 1968, pl. 7, figs. 16, 19).

Cardinalia.—There are only minor differences between the cardinalia of *Monorthis* (Fig. 6.5, 6.12) and *Ahtiella* (Figs. 9.17, 10.20, 10.24, 11.10, 11.15). *Monorthis transversa* shows some degree of intraspecific variation in the cardinal process, ranging from bladelike to a ridge moderately enlarged anteriorly, occupying the entire length of a raised subtriangular notothyrial platform. The cardinalia of *M. transversa* are nearly identical to those of *A. famatiniana* n. sp. and *A. coloradoensis*, whereas in the Peruvian *A. zarelae* and *Ahtiella* sp., the cardinal process tends to be more robust and ovoid in outline. *Ahtiella argentina* is characterized by a bladelike to slightly enlarged cardinal process erected on a gently convex notothyrial platform (Fig. 9.20, 9.22). The subtriangular to suboval, anterolaterally open sockets excavated on the valve floor and partially under the



dorsal interarea of *Monorthis* and *Ahtiella*, as well as the slender, distally enlarged socket ridges, are also closely comparable.

Subperipheral rims and platforms.—Despite the redefinition of these terms by Cocks and Rong (1989), some imprecision persists in the literature, with the term ‘platform’ having been applied to both dorsal and ventral valves (not only to the dorsal one, as these authors proposed), and the two structures are often not easy to differentiate on the basis of their morphology. To avoid confusion, ‘platform’ (= ‘diaphragm’) is used here to designate a low to high, somewhat undercut elevation of the ventral and/or dorsal valve floor originating at or near the cardinal angles and not related to external geniculation, the internal disc, or any kind of valve thickening. On the other hand, following Cocks and Rong (1989), the term ‘peripheral rim’ (or ‘subperipheral rim’) is applied to a raised rim running at or near the variably thickened valve margin. When a more or less prominent peripheral rim has developed in the ventral valve, it can be mirrored in the dorsal valve by a similar structure that is often related to an internal deflection of the valve.

According to the original diagnosis of *Monorthis* (see Bates, 1968), platforms are absent in both valves. Valve margins of the Famatinan *M. transversa* are crenulated but not thickened (Fig. 6.9), whereas the large ventral valves of the Precordilleran *M. cumillangoensis* show variably thickened margins and an internal geniculation, which is nearly identical to that seen in *Ahtiella zarelae* (cf. Figs. 6.14, 11.18). The ventral valves of *A. famatiniana* n. sp. and *A. coloradoensis* exhibit a conspicuous thickening along the geniculation; in both species, however, it is absent in juvenile individuals (Fig. 10.15), suggesting that this structure developed progressively by peramorphosis. In *A. argentina*, the whole ventral valve margin is geniculated, forming a prominent internal disc deeply incised by the vascula terminalia (Fig. 9.9), like in the Baltic *A. lirata*. A corresponding discontinuous platform-like structure is usually present in the dorsal valve of *A. famatiniana* n. sp. (Fig. 10.20). On the contrary, in *A. coloradoensis*, it is poorly developed or even absent (Fig. 11.10). Unlike other Gondwanan species, the large dorsal valves of *A. argentina* display a series of curved, roughly radial ridges that can be interpreted as a platform-like structure (Fig. 9.20, 9.22). *Ahtiella baltica* Öpik, 1932, as can be seen in the specimen figured by Öpik (1933, pl. 4, fig. 6), possesses a continuous undercut platform, but it is faint or absent in *A. lirata*.

Vascular system.—Vascular trunks are not discernible in *Monorthis cumillangoensis* in part due to the strong internal impression of the external ornamentation, whereas in *M. transversa*, a series of short anastomosing canals can be present along valve margins (Fig. 6.4, 6.8). In *Ahtiella famatiniana* n. sp., the

distal portion of mantle canals is well marked on the anterior third of the ventral valve (Fig. 10.22). Adult stages of *A. argentina* and *A. paucirugosa* always display a deeply impressed mantle canal system of the saccate type, with posteriorly directed branches of vascula media enclosing large gonadal pouches (Fig. 9.10, 9.18).

Trends of morphological change.—From the above comparisons, the following trends can be recognized through the inferred *Monorthis transversa* (and its putative ancestor *Hesperonomiella arcuata* Benedetto, 2003b)—*Ahtiella argentina* lineage (Fig. 7): (1) the nongeniculated ventral valve margin of *M. transversa* progressively thickens, originating in *Ahtiella* an internal geniculation, which is low in the species from Wales (*A. quadrata*, *A. concava*) and northwestern Argentina (*A. coloradoensis*), intermediate in the Famatinan (*A. famatiniana* n. sp.) and Peruvian (*A. zarelae*) forms, and more prominent in the Darriwilian species from Cuyania and Baltica. In the latter (e.g., *A. lirata*, *A. jaanussoni* Hessland, 1949), the main trend is toward a strongly convex gibbous dorsal valve; (2) the pseudodeltidium is absent or incipient in *Monorthis*, is restricted to the apical region of delthyrium in the oldest known species (*A. zarelae*) as well as in the Welsh species, and reaches almost two-thirds in the later species from Precordillera and Baltica; (3) external ornamentation evolved from equally multicostellate in *Monorthis* (retained in the younger *A. coloradoensis*) to ramicostellate in the Floian *A. zarelae*, becoming unequally multicostellate to incipiently parvicostellate in the Dapingian *A. famatiniana*, and definitely parvicostellate in the Darriwilian species; (4) the dorsal platform is absent in *Monorthis*, is variably developed in the early species *A. zarelae* and *A. famatiniana* n. sp., and becomes more prominent in the younger Cuyanian and Baltic species; (5) the mantle canal system is indistinct or confined to the valve margin in *Monorthis*, has well-impressed distal branches on the margin of disc and trail in the Floian-Dapingian species of *Ahtiella*, and culminates in *A. argentina* and *A. paucirugosa* with a deeply impressed mantle canal system on the entire surface of adult specimens.

Phylogenetic analysis

Cladistic analysis of Ahtiella species.—Comparative morphology makes evident that differences between *Monorthis* and basal species of *Ahtiella* are subtle, which makes them difficult to distinguish from each other. The question is whether such similarities reflect homologies and therefore reveal phylogenetic affinities or, on the contrary, they can be viewed as cases of extreme morphological convergence along two independent lineages. Given the striking resemblance in multiple external

Figure 6. (1–9) *Monorthis transversa* Benedetto, 2003b; Loma del Kilometro Member of the Suri Formation (Chaschuil) and Molles Formation, Famatina Range: (1) latex cast of ventral valve exterior, CEGH-UNC 19628a; (2) latex cast of dorsal valve exterior, CEGH-UNC 19628b; (3) latex cast of ventral valve, CEGH-UNC 19628a, showing incipient delthyrium cover; (4, 5) internal mold (4) and latex cast (5) of dorsal valve, CEGH-UNC 19635; (6) internal mold of dorsal valve, CEGH-UNC 19632; (7) internal mold of ventral valve, CEGH-UNC 19625; (8, 9) internal mold of ventral valve (8) and latex cast (9), CEGH-UNC 10962. (10–18) *Monorthis cumillangoensis* Benedetto, 2001; San Juan Formation, silicified specimens from Cerro Cumillango and Cerro La Chilca, Precordillera: (10) ventral valve exterior, CEGH-UNC 17915; (11) ventral valve exterior, CEGH-UNC 17917; (12) dorsal valve interior, CEGH-UNC 17920; (13) ventral valve interior, CEGH-UNC 17933; (14, 15) ventral valve interior (14) and detail of incipient delthyrium cover (15), CEGH-UNC 17948; (16) ventral valve interior, CEGH-UNC 21152; (17) dorsal valve exterior, CEGH-UNC 21153; (18) dorsal valve interior, CEGH-UNC 17942. All specimens dusted with ammonium chloride. Scale bars = 5 mm.

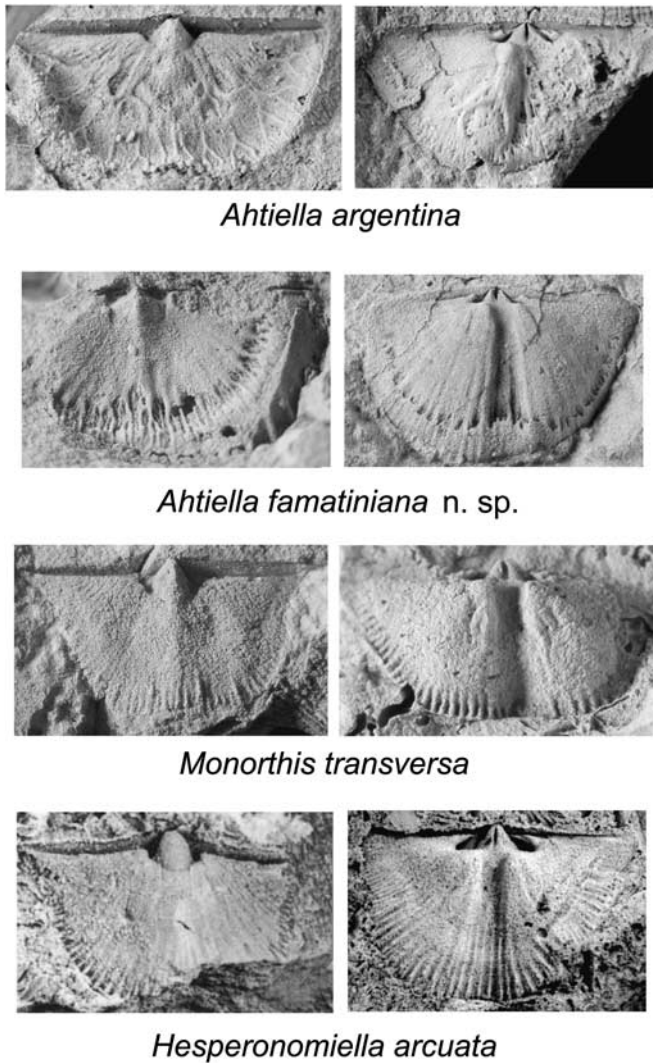


Figure 7. Generalized trend of morphological change through selected taxa from the Floian-Dapingian volcanosedimentary succession of Famatina. Ventral internal molds at left, dorsal internal molds at right. *Hesperonomiella arcuata* Benedetto, 2003b): left, CEGH-UNC 15740; right, CEGH-UNC 19078. *Monorthis transversa* Benedetto, 2003b: left, CEGH-UNC 19095; right, CEGH-UNC 19623. *Ahtiella famatiniana* n. sp.: left, CEGH-UNC 27140; right, CEGH-UNC 27135b. At the top of the succession, *A. argentina* Benedetto and Herrera, 1986 illustrates a more derived species of *Ahtiella* from the Darriwilian of Precordillera (left, CEGH-UNC 27111; right, CEGH-UNC 21118b).

747 and internal details along with gradation in some features, convergence seems highly improbable. To investigate whether the trends deduced from comparative morphology are phylogenetically significant, a cladistic analysis of *Ahtiella* species was performed using TNT (Tree Analysis Using New Technology) version 1.5 (Goloboff and Catalano, 2016), selecting the heuristic search option with multiple random addition sequences and the tree bisection reconnection (TBR) branch-swapping algorithm holding 10 trees in each addition sequence. A total of 23 characters comprising internal and external features were included within the *Ahtiella* analysis (Table 1). The 23-character matrix was analyzed for 10 taxa (Table 2). The Welsh species *A. concava* was not considered because it possesses some features atypical for the genus (e.g., apparently smooth exterior, elongate oval anterior adductor scars each flanked by prominent

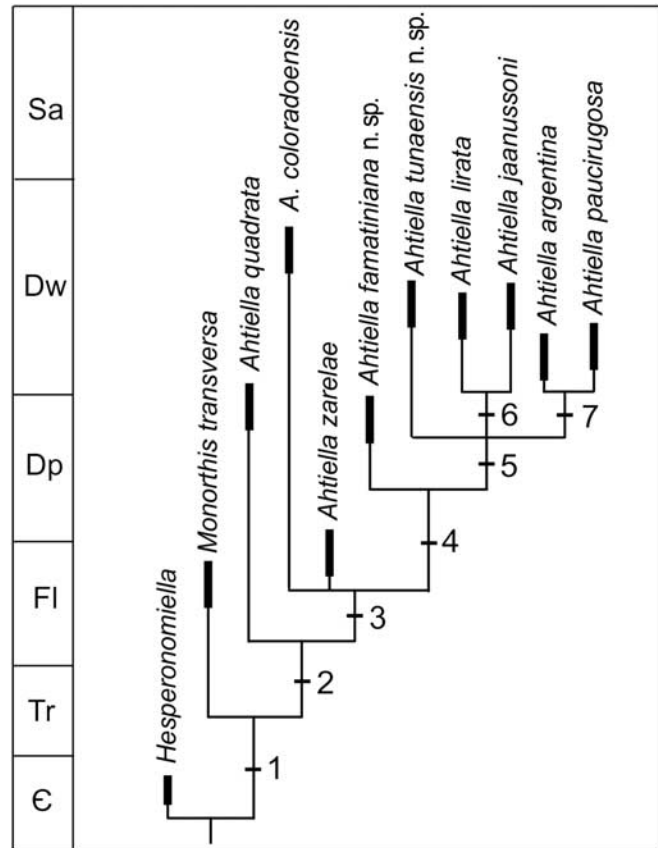


Figure 8. Phylogenetic relationships of taxa analyzed calibrated to chronostratigraphic scale. Apomorphies defining numbered nodes are discussed in the text.

762 septa), resulting in destabilization of the relationships among
 763 other taxa. *Hesperonomiella* was chosen as the outgroup for
 764 rooting the phylogenetic tree because it exhibits an ensemble of
 765 nonderived internal features, and lacks the majority of apo-
 766 morphies present in the group analyzed here. This, along with its
 767 first appearance in the middle Cambrian, suggests it as a
 768 potential ancestor of the taxa considered in this study. A heur-
 769 istic search of the data matrix in which all characters were
 770 unordered and equally weighted produced four minimal length
 771 trees 52 steps long, with consistency index of 0.692 and reten-
 772 tion index of 0.754. The strict consensus tree (Fig. 8) is pre-
 773 sented here with branch length calibrated to the age of the first
 774 appearance datum of each taxon. As the phylogenetic tree
 775 shows, the basal member of the *Ahtiella* clade is *Monorthis*
 776 *transversa*, which possesses ancestral features such as multi-
 777 costellate ornamentation, absence of a dorsal platform, an
 778 unthickened periphery of the ventral valve, an open delthyrium,
 779 and an unmodified notothyrium. *Monorthis transversa* has in
 780 common with the *Ahtiella* species a resupinate and alate shell
 781 (node 1), and cardinalia and muscle scars of the orthoid type.
 782 The Welsh species *A. quadrata* is expressed as the most basal
 783 member of the *Ahtiella* clade displaying vestigial or incipient
 784 pseudodeltidium and chilidium and a slightly thickened ventral
 785 valve margin (node 2). The apomorphy that defines the
 786 remaining *Ahtiella* species (node 3) is the presence of a dorsal
 787 platform, which varies from low and discontinuous in the older
 788 species to variably developed in the younger species. The

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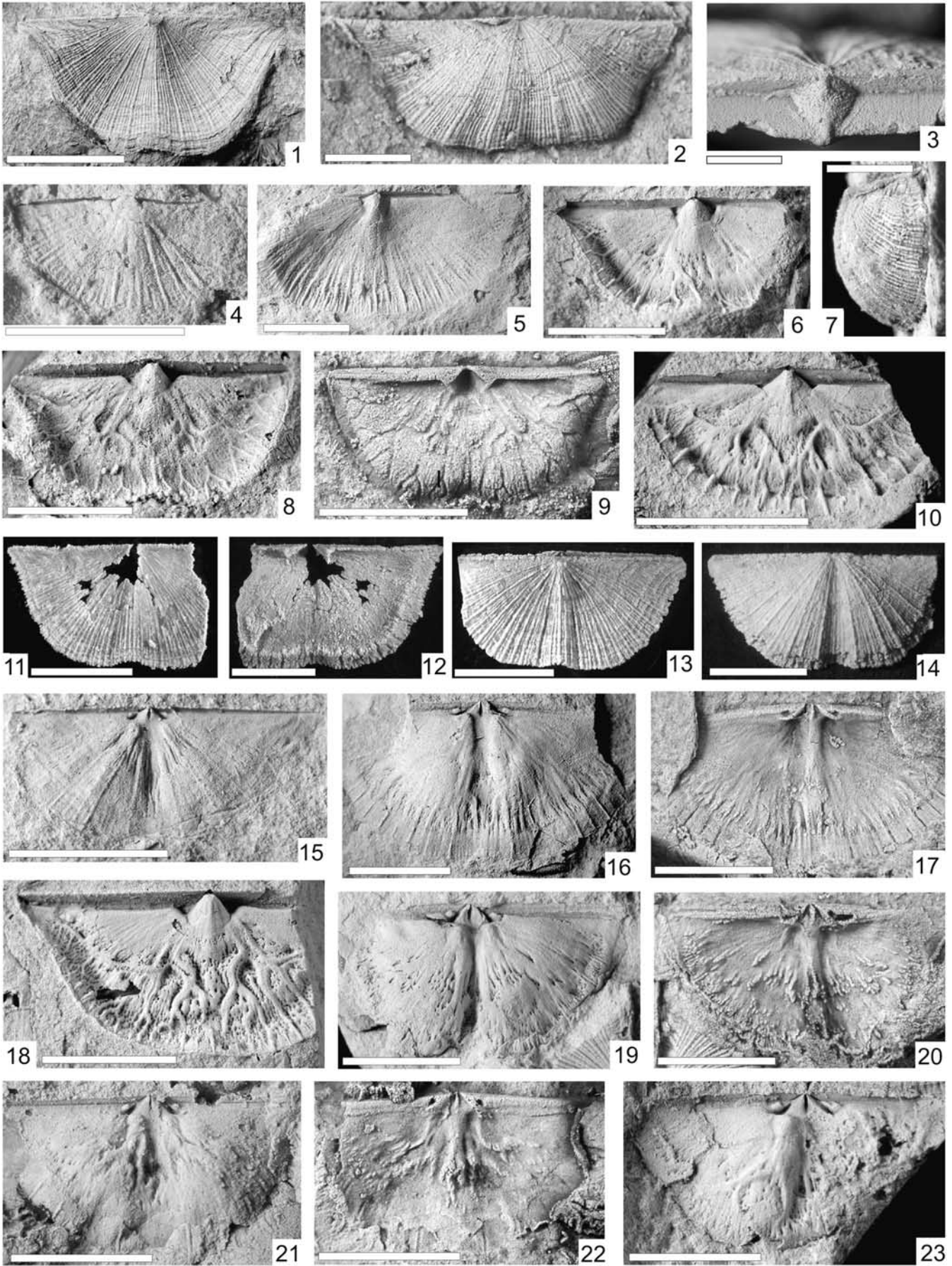
Gondwanan species *A. coloradoensis*, *A. zarelae*, and *A. famatiniana* n. sp., which each retain ancestral uniform or nearly uniform radial ornamentation and an incipient apical pseudodeltidium, appear as basal to the more derived species from Baltica, Cuyania, and Newfoundland. In addition, it should be noted that the presence of pseudopunctae has not been demonstrated in any of these Gondwanan species. Among them, *A. famatiniana* n. sp. is slightly more advanced by having a comparatively larger pseudodeltidium and chilidium, a more prominent dorsal platform, a deeply impressed vascular system on the disc/trail deflection, and ornamentation tending to be incipiently parvicostellate. The Famatinan species clusters with the group that includes *A. tunaensis* n. sp., *A. lirata*, *A. jaanussoni*, *A. argentina*, and *A. paucirugosa*, characterized by a well-defined disc and a dorsally directed trail in the interior of ventral valve (node 4). This group shares unequally parvicostellate ribbing, a well-developed pseudodeltidium and chilidium, and a relatively large pseudopunctae (node 5). *Ahtiella jaanussoni* and *A. lirata* form a cluster based on a gibbous shell profile and prominent rugae or corrugations covering most of the valve surface, and likely reflect local radiation of the genus in the Baltica paleocontinent (node 6). This group of species served originally to define the genus *Ahtiella*. On the other hand, *A. argentina* and *A. paucirugosa* form a sister group of Baltic species by sharing deeply impressed mantle canals on the entire valve interior and curved ridges in the floor of dorsal valve (node 7).

Our species-level phylogenetic tree shows that since its origin in the Floian, the genus *Ahtiella* underwent successive speciation events along the Andean margin of Gondwana where the common ancestor would likely have inhabited, and subsequently dispersed and continued speciating as new geographic areas were colonized. The diversification of the *Ahtiella* clade appears to have occurred by cladogenesis because *A. coloradoensis*, a conservative species closely related morphologically to *Monorthis*, persisted in the Central Andean Basin until the late Darriwilian. Perhaps the main conflict posed by the phylogenetic hypothesis of Figure 7 is that *A. quadrata* is shown as a basal member of the clade because the Welsh species retains some traits ancestral to *A. zarelae* and *A. famatiniana* n. sp. A possible explanation is that *A. quadrata* originated from a Gondwanan ancestor and then migrated along Gondwanan shelves to reach the colder Avalonian waters. However, to date no records of morphologically related forms are known from Gondwana, which could be due either to a lack of extensive sampling in the still poorly known Bolivian and Peruvian sectors of the Central Andean Basin or to the absence of this species in the Andean region. An alternative interpretation is that *A. quadrata* evolved independently in the Avalonian paleocontinent from a local species of *Monorthis*. The possibility of parallel evolution at the generic level as well as its possible causes were discussed in a previous paper (Benedetto, 2008) to account for the nearly simultaneous record of the genus *Productorthis* in Baltica and the Famatinan volcanic arc, with underlying strata in both regions of the ancestral genus *Panderina* Schuchert and Cooper, 1931. In our case, the Welsh species *M. menapiae* of 'lower Arenigian' (Floian) age (Bates, 1969) closely resembles the Famatinan species *M. transversa* and could be a potential ancestor of *A.*

quadrata and, eventually, of the 'anomalous' species *A. concava*. It seems likely that cases of parallel evolution in brachiopods, and thus the existence of paraphyletic genera, might be more frequent than previously thought.

In summary, available evidence from the Gondwanan material supports, contrary to previous assumptions, that the basal plectambonitoid *Ahtiella* could have evolved from the hesperonomiid orthoid *Monorthis transversa*, and that *A. famatiniana* n. sp. and *A. zarelae* are not only the earliest species of the genus, but also are morphologically intermediate between *M. transversa* and the more derived species of *Ahtiella* from the Darriwilian of Cuyania and Baltica (Fig. 8).

Some macroevolutionary implications.—There is a general consensus that the peculiar strophomenide shell architecture and concomitant anatomical and physiological changes were adaptations (key innovations) allowing invasion of a new ecological niche or adaptive zone, which was essentially the acquisition of an ambitopic or permanent liberossessile life strategy linked to the colonization of low-energy, offshore marine environments (Bassett, 1984). Over time, such morphological changes became so marked that they led to the recognition of a separate higher taxon, the order Strophomenida, which together with other groups was lumped into the large and quite heterogeneous class Strophomenata. Although higher taxa are often viewed as artificial, nonmonophyletic, subjective entities, a number of quantitative studies have confirmed the taxic metrics as an adequate proxy for assessing morphological disparity (Erwin, 2007, p. 59). If, as evidence presented here suggests, *Ahtiella* originated from *Monorthis* through a series of minor transformations, then the impressive morphological gap among 'typical' orthides and strophomenides was bridged through a brief cladogenetic event. At first, such a transition indicates that there is not a definite discontinuity between species-level evolution (processes that occur within a species or lead to a new species) and the origin of higher taxa (macroevolution). Central to this statement is the assumption that higher taxa are evolutionary entities characterized by a significant morphological disparity achieved over a long period of time, then the greater the time elapsed since their origin from a common ancestor, the larger morphological disparity. In the present case study, it would be expected that the highly plesiomorphic basal forms (i.e., ahtiellins) of a given higher taxon (i.e., strophomenides) are more similar to their putative ancestors (i.e., hesperonomiids) than they are to the more derived (apomorphic) end-members of the same clade (i.e., sowerbyellids, aegiromenids). In other words, morphological discontinuity becomes minimal at a point closer to the initial divergence of two phylogenetically related higher taxa. A consequence of this is that assignation of basal forms to one or another higher taxon can be difficult in the lack of a well-supported phylogeny. The need of a 'shoehorn' to classify such earliest members of a given higher taxon into a specified order or suborder has also been noted in other groups of marine benthic organisms. For instance, the clade Bivalvia includes a number of early representatives of Tremadocian-Floian age that lack certain apomorphies defining more derived crown groups. In this respect, it has been suggested that such basal taxa can be classified as plesions, i.e., paraphyletic groups having a number



906 of symplesiomorphic traits but morphologically close to a given
907 higher taxon (Fang and Sánchez, 2012). In the case of brachiopods,
908 as Carlson (2016, p. 421) stated, numerous higher taxa had been
909 thought to represent grade-level taxa, i.e., not clades, and our
910 evidence indicates that this could be the case of strophomenides.
911

912 Materials and methods

913 *Repository and institutional abbreviations.*—CEGH-UNC,
914 Centro de Investigaciones en Ciencias de la Tierra CONICET
915 and Universidad Nacional de Córdoba, Argentina; CORD-PZ,
916 Museo de Paleontología, Universidad Nacional de Córdoba,
917 Argentina; MGM, Museo Geominero, Madrid, Spain.

918 Systematic paleontology

919 The systematic classification follows that of the Treatise on
920 Invertebrate Paleontology (Cocks and Rong, 2000). Following
921 Congreve et al. (2015), the genus *Ahtiella* is referred to the
922 ‘Plectambonitoidea’ with the quotation marks denoting that the
923 superfamily is paraphyletic (Wiley, 1979).

924 Order Strophomenida Öpik, 1934
925 Superfamily ‘Plectambonitoidea’ Jones, 1928
926 Family Taffiidae Schuchert and Cooper, 1931
927 Subfamily Ahtielinae Öpik, 1933
928 Genus *Ahtiella* Öpik, 1932

929 *Type species.*—*Ahtiella lirata* Öpik, 1932 from the Darriwilian
930 (Middle Ordovician) of Tsitri, Estonia, by original designation.

931 *Diagnosis (emended).*—Shell resupinate, variably geniculate;
932 ornamentation subequally multicostellate, ramicostellate or parvi-
933 costellate, often with posterolateral rugae. Dental plates widely
934 spaced converging to valve floor, enclosing posteriorly sub-
935 triangular to subquadrate ventral muscle field. Interior of ventral
936 valve with thickened margin defining disc and trail. Cardinal pro-
937 cess simple. Dorsal median ridge broad, rounded. Dorsal platform
938 variably developed. Saccate mantle canal system usually strongly
939 impressed either along shell margin or whole interior.

940 *Ahtiella argentina* Benedetto and Herrera, 1986
941 Figure 9.1–9.23

943 1986 *Ahtiella argentina* Benedetto and Herrera, p. 114, pl. 1,
944 figs. 4–21.

945 2002 *Ahtiella argentina*; Benedetto, p. 114, pl. 2, figs. 20, 21.

946 2003a *Ahtiella argentina*; Benedetto, p. 201, pl. 9, figs. 9–12.

2009 *Ahtiella argentina*; Benedetto et al., fig. 9f. 947

Holotype.—CORD-PZ 8283, a conjoined specimen from the 948
upper San Juan Formation, Quebrada Los Gatos, Cerro Viejo, 949
San Juan Province, Argentina (Benedetto and Herrera, 1986). 950

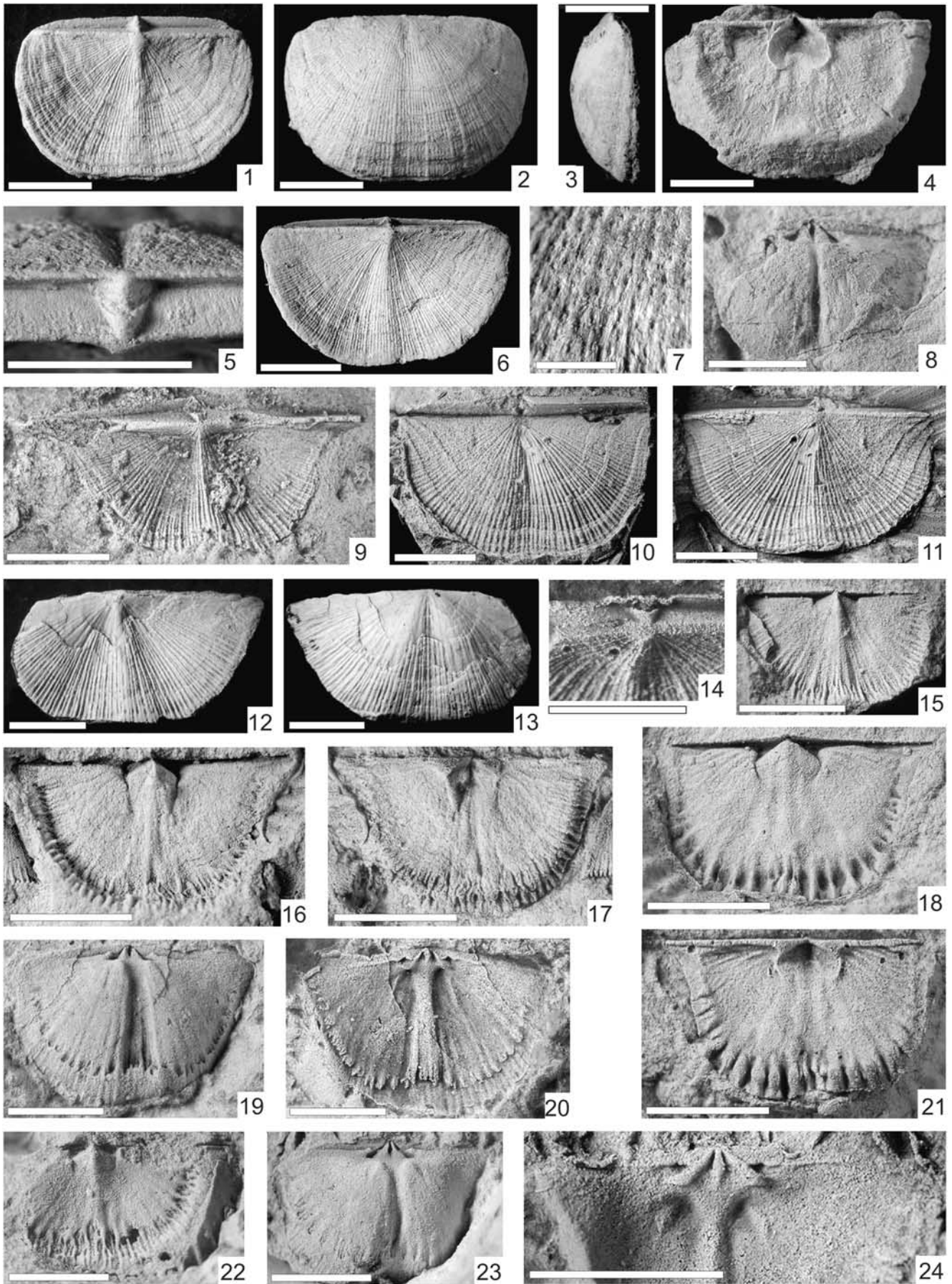
Diagnosis.—Shell transversely semielliptical (mean length/ 951
width ratio 0.39) with acute cardinal extremities; profile variably 952
resupinate with carinate ventral median fold and corresponding 953
subangular dorsal sulcus reaching anterior margin. Radial 954
ornamentation unequally parvicostellate. Ventral muscle field 955
short, triangular, extending up to 25% of valve length. Ventral 956
valve strongly thickened marginally, defining internal disc and 957
trail. Dorsal valve with strong rounded median ridge originating 958
in front of notothyrial platform. Internal surface with elongated 959
pustules coalescing to form irregular, roughly radial ridges. 960
Both valves with deeply impressed saccate mantle canal system. 961

Occurrence.—Uppermost levels of the San Juan Formation, 962
Precordillera, San Juan Province, Argentina. Darriwilian (Dw1– 963
Dw2). 964

Description.—Shell to 23 mm width, transversely semielliptical 965
in outline; mean length/width ratio 0.39 (N=18); profile 966
strongly concavoconvex, mature specimens up to 45% as deep 967
as long. Maximum width at hinge line. Cardinal extremities 968
auriculate or alate in juvenile to medium-sized specimens, 969
becoming acute (40–45°) in mature specimens. Ventral valve 970
resupinate, gently convex at umbonal area, becoming broadly 971
concave anteriorly; ventral median fold initially carinate, 972
becoming broader and rounded toward anterior margin where it 973
attains 15–20% of valve width, flanked by strongly concave 974
areas. Ventral interarea planar, steeply apsacline to almost 975
catacline, attaining height equivalent to 15% of shell length. 976
Pseudodeltidium well developed, semiconical, covering near 977
two-thirds of delthyrium. Dorsal valve moderately to strongly 978
convex with deep angular median sulcus becoming wide and 979
shallow anteriorly. Dorsal interarea anacline, planar, three to 980
four times lower than ventral. Notothyrium covered almost 981
entirely by large convex chilidium. Ornamentation unequally 982
parvicostellate, with 6–8 first-order rounded costellae originat- 983
ing at umbonal area and 4–8 subequal finer parvicostellae 984
intercalated between them, some of which can attain size equal 985
to primaries on anterior third of valve; costellae 10–12 per 2 mm 986
(counted at 5 mm growth stage). Fine, evenly spaced, concentric 987
fila (~18 per mm). Posterolateral margins with up to six faint 988
rugae strongly oblique to hinge line. 989

Ventral interior with small subtriangular teeth bearing well- 990
developed, slit-like crural fossettes on medial faces, supported 991
by short thick dental plates slightly converging to valve floor 992

←
Figure 9. *Ahtiella argentina* Benedetto and Herrera, 1986; Upper San Juan Formation, Cerro Viejo, Precordillera: (1) latex cast of ventral valve exterior, CEGH-UNC 27121; (2, 7) latex cast of dorsal valve exterior (2) and profile view (7), CEGH-UNC 27119; (3) posterior view of conjoined specimen showing pseudodeltidium and chilidium, CEGH-UNC 16328; (4) early juvenile ventral internal mold, CEGH-UNC 27118; (5) juvenile ventral internal mold, CEGH-UNC 27153; (6) internal mold of small adult ventral valve, CEGH-UNC 27072; (8, 9) internal mold (8) and latex cast (9) of ventral valve, CEGH-UNC 27111; (10) internal mold of ventral valve, CEGH-UNC 27181; (11, 12) external (11) and internal (12) views of silicified ventral valve, CEGH-UNC 21963; (13, 14) ventral (13) and dorsal (14) views of silicified conjoined specimen, CEGH-UNC 21964; (15) internal mold of juvenile dorsal valve, CEGH-UNC 27160; (16, 17) internal mold (16) and latex cast (17) of dorsal valve, CEGH-UNC 27108; (18) internal mold of ventral valve, CEGH-UNC 21995; (19, 20) internal mold (19) and latex cast (20) of dorsal valve, CORD-PZ 8295; (21, 22) internal mold (21) and latex cast (22) of dorsal valve, CEGH-UNC 27075; (23) dorsal internal mold, CEGH-UNC 21118b. All specimens dusted with ammonium chloride. Scale bars = 3 mm (3, 4); 5 mm (remainder).



and diverging from each other at 80–90°. Lateral delthyrial cavities of adult specimens partially to completely filled with secondary deposits. Ventral muscle field short, triangular in outline, extending ~20–25% of valve length, enclosed posterolaterally by dental plates, delimited anterolaterally by low arcuate ridges; diductor scars elongate, almost three times longer than wide, not clearly differentiated from wider subtriangular adductor field. Mantle canal system saccate, deeply impressed in mature individuals; vascula media broad, straight, diverging forward at 60°, extending from anterior ends of diductors to approximate valve midlength where they bifurcate, with one branch directed posterolaterally and the other directed medially, each becoming repeatedly dichotomized at disc margin (Fig. 9.10, 9.18). Surface between vascular trunks covered by pustules. Gonadal areas subtriangular to suboval in outline, faintly striated radially, variably pitted. Valve margin strongly thickened, geniculated, with well-defined disc and trail deflected dorsally, forming angle of ~40° with respect to disc surface. Low rounded to subangular subperipheral rim occasionally present along entire disc margin. External ribbing not reflected on valve interior except in juvenile individuals.

Dorsal interior with small cardinalia extending ~10% of valve length. Cardinal process high, bladelikey, slightly enlarged anteriorly, erected on moderately elevated slightly convex notothyrial platform merged anteriorly with strong, rounded median ridge, usually extending to valve midlength or slightly beyond, but reaching disc margin in some specimens. Socket ridges rodlike, free distally, diverging from each other at 95–100°, bounding deep semiconical sockets excavated below and partially on interarea. Muscle field faintly impressed, quadripartite, with subcircular posterior adductor scars and larger suboval anterior adductor scars. Saccate dorsal mantle canal system with broad, well-impressed vascula media divided immediately in front of anterior extremity of muscle field in two branches, one of them running subparallel to median ridge, the other directed anterolaterally, both originating numerous small canals perpendicular to valve margin. Valve interior with elongated papillae, in large specimens coalescing to form irregular roughly radial ridges, which are more conspicuous at valve midlength on both sides and at end of median ridge. Small, closely spaced papillae between peripheral vascular trunks.

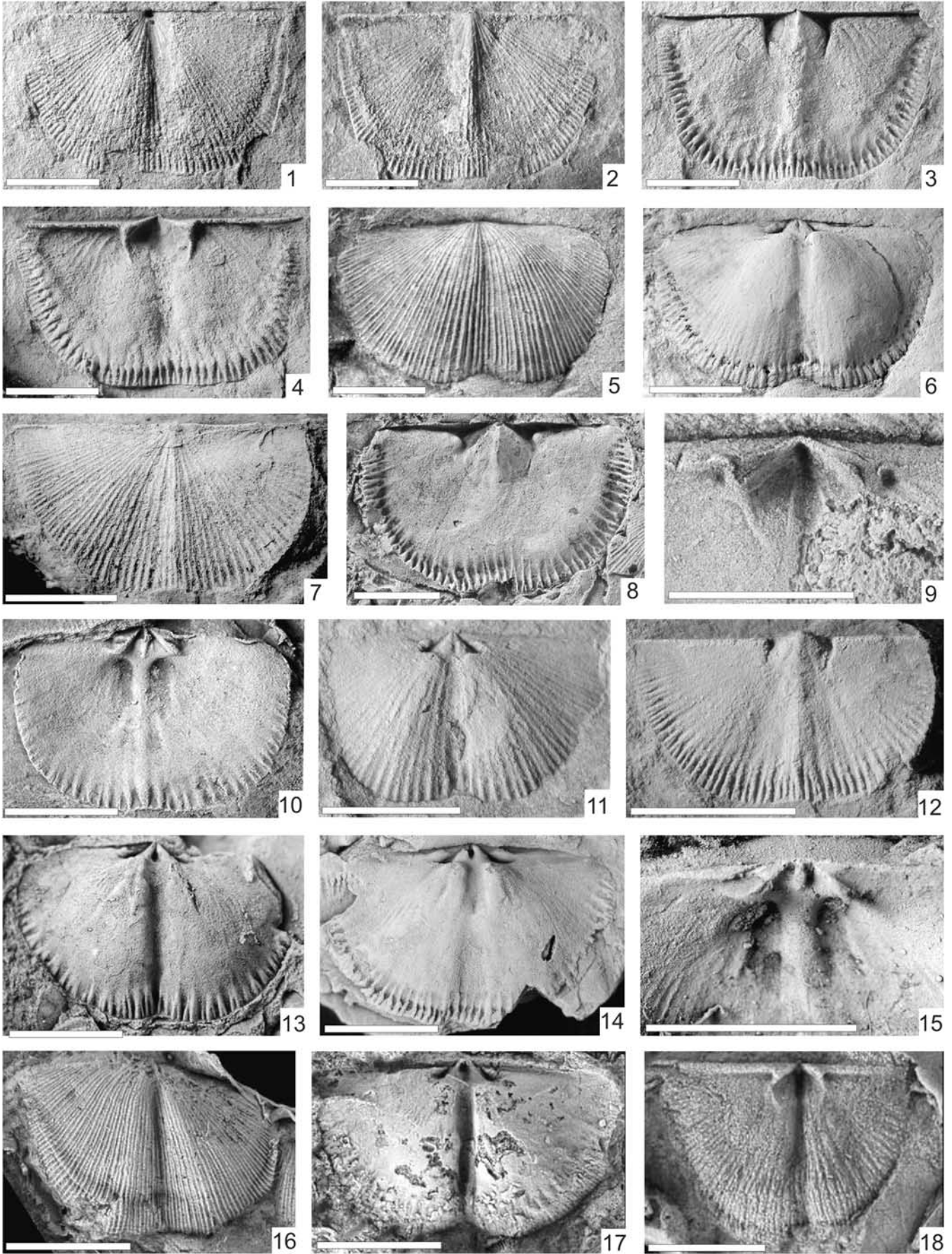
Materials.—In addition to the material listed and illustrated by Benedetto and Herrera (1986), the following new specimens have been included in the present study: CEGH-UNC 21960–21965, 21981, and 21994–21995 from Cerro Viejo (Quebrada Honda); CEGH-UNC 10561–10564, 16319–16321, 22070–

22075, 27104–27130, and 27160–27162 from Cerro Viejo (Quebrada Los Gato).

Remarks.—This species clearly differs from *Ahtiella lirata* in its more transverse shell outline and less convex, uniformly curved shell profile lacking abrupt geniculation. Moreover in *A. lirata*, the dorsal sulcus is very shallow and confined to the posterior region of the valve and the corresponding ventral fold is inconspicuous, whereas in *A. argentina*, the sulcus is deeper and always attains the anterior margin, and the ventral fold is well defined and carinate posteriorly. Internally, the Precordilleran species can be distinguished by having smaller subtriangular ventral muscle field, which in the type species tends to be subquadrate and extends to approximately one-third of the valve length; there are deeply impressed vascular trunks on the internal surface of both valves, but especially in the ventral one. The slightly older species *A. baltica*, from the Rõgo Sandstone of Estonia, resembles *A. argentina* in its more transverse shell outline but differs in having a longer bilobed ventral muscle field of *lirata*-type. The dorsal interior, even though exhibiting features of the genus, shows a quite atypical continuous, anteriorly free dorsal platform (Öpik, 1933, pl. 4, fig. 6). *Ahtiella arenaria* Öpik, 1933 is a poorly known Estonian species (only a dorsal exterior and a ventral interior have been illustrated, by Öpik, 1933, pl. 4, figs. 7, 8) recovered from the same beds and localities as *A. baltica* to which it strongly resembles. Of the species from Sweden (Island of Öland) described by Hessland (1949), only *A. jaanussoni* is known from dorsal interiors (ventral interiors are unknown). Its dorsal valve is reminiscent of *A. lirata*, but the shell is somewhat more transverse, the posterolateral rugae are more pronounced, and the dorsal interior possesses a long prominent median ridge. As Cocks and Rong (1989) stated, the rest of the *Ahtiella* species described by Hessland (1949) can only be questionably assigned to the genus because internal morphology remains unknown. Among them, *A. ? oelandica* Hessland, 1949 is the most similar to *A. argentina* in its transverse shell outline, uniformly curved dorsal valve profile, and especially in the presence of a well-developed dorsal median sulcus reaching the anterior margin. The main external difference lies in the coarser and more prominent rugae in the Swedish species. It is unclear whether such differences in size, shell outline, definition of disc and trail, and strength of posterolateral rugae are taxonomically significant because these features show gradation between species and are quite variable even in individuals of the same species. Clearly, a revision of the Baltic species is needed to clarify this issue.

Ahtiella concava from the Bob Deiniol Formation of Anglesey (Wales) resembles *A. argentina* in its moderately convex and uniformly curved dorsal valve profile, acute cardinal

Figure 10. (1–8) *Ahtiella tunaensis* n. sp.; Las Chacritas Formation, Sierra de la Trampa, Precordillera: (1–3, 5) paratype, conjoined specimen, CEGH-UNC 21084, in ventral (1), dorsal (2), and profile (3) views, and detail of interarea (5) showing pseudodeltidium and chilidium; (4) holotype, interior of ventral valve, CEGH-UNC 21083; (6) paratype, ventral view of conjoined specimen, CEGH-UNC 27172; (7) detail of ventral valve, CEGH-UNC 27174, showing pseudopunctae; (8) paratype, dorsal internal mold, CEGH-UNC 27171. (9–24) *Ahtiella famatiniana* n. sp.; Punta Pétreá Member of the ‘Suri’ Formation (Chaschuil), and volcanoclastic beds exposed at Las Pirca anticline (Central Famatina Range): (9) paratype, latex cast of ventral valve exterior, CEGH-UNC 27149; (10, 11) external mold (10) and latex cast (11) of ventral valve, CEGH-UNC 27131; (12–14) ventral (12) and dorsal (13) views of conjoined specimen, CEGH-UNC 27163, and detail of pseudodeltidium (14); (15) juvenile ventral internal mold, CEGH-UNC 27158; (16, 17) paratype, internal mold (16) and latex cast (17) of ventral valve, CEGH-UNC 27135a; (18, 21) holotype, internal mold (18) and latex cast (21) of ventral valve, CEGH-UNC 27137; (19, 20) internal mold (19) and latex cast (20) of dorsal valve, CEGH-UNC 27135b; (22) internal mold of ventral valve, CEGH-UNC 27140; (23, 24) internal mold (23) and latex cast (24) of dorsal valve, CEGH-UNC 27141, showing detail of cardinalium. All specimens dusted with ammonium chloride. Scale bars = 1 mm (7); 3 mm (5, 14); 5 mm (remainder).



1089 angles, and well-developed, carinate ventral fold, but differs
1090 from the Precordilleran species in having a catacline to procline
1091 ventral interarea and a less transverse shell; the width/length
1092 ratio in the specimens measured by Bates (1968, p. 168) is 0.63.
1093 According to Bates (1968, p. 167), the ornamentation in the
1094 Welsh specimens is “too fine to be observed,” and posterolateral
1095 rugae are not evident. Internally, *A. concava* has a subquadrate
1096 rather than subtriangular ventral muscle field. The dorsal valve
1097 shares with *A. argentina* a high rounded median dorsal ridge
1098 extending beyond the valve midlength, but in the Welsh species,
1099 the muscle field is strongly elongated and bounded by a pair of
1100 conspicuous ridges, a feature atypical of the genus. *Ahtiella*
1101 *quadrata* from the Torllwyn Formation of Anglesey is
1102 represented by fragmentary material, but judging from the two
1103 illustrated complete ventral valves (Bates, 1968, pl. 8, figs. 2, 3),
1104 the shell is slightly wider than long and coarsely costellate
1105 (~6 costellae per 2 mm) than *A. argentina*.

1106 *Ahtiella zarelae* from the San José Formation of Peru can
1107 readily be distinguished from *A. argentina* by its less transverse,
1108 nonauriculate shell, and its ramicostellate radial ornamentation.
1109 Internally, the Peruvian species possesses a subpentagonal
1110 rather than triangular ventral muscle field, and a higher, septum-
1111 like dorsal median ridge. In addition, the inner surface of both
1112 valves is almost entirely covered by radial ribs lacking vascular
1113 impressions.

1114 Most similar to the Precordilleran species is *Ahtiella*
1115 *paucirugosa* from the volcanoclastic Summerford Group of
1116 Newfoundland. Given that the North American specimens are
1117 tectonically distorted, shell outline is difficult to compare
1118 (*A. paucirugosa* seems to be approximately twice as wide as
1119 long). However, they have in common such external features as
1120 a well-defined dorsal sinus and ventral fold, and moderate shell
1121 convexity, although posterolateral rugae are less marked in the
1122 North American species. Internally, the dorsal valve resembles
1123 that of *A. argentina* in having a broad median ridge almost
1124 reaching the margin, rows of elongate pustules and septules
1125 forming radial crests, and deeply marked mantle canals of the
1126 saccate type. The main difference lies in the ventral muscle field
1127 of *A. paucirugosa*, which is bilobed with longer diductor scars
1128 extending approximately to the valve midlength.

1129 *Ahtiella tunaensis* new species

1130 urn:lsid:zoobank.org:act:C9CF5CB8-E169-4777-8C0B-
1131 ADEF34A52181

1132 Figure 10.1–10.8

1133 2003a *Ahtiella* n. sp.; Benedetto, p. 201, pl. 9, figs. 16–18.

1134
1135 *Type specimens*.—Holotype, CEGH-UNC 21083, a ventral
1136 valve. Paratypes: CEGH-UNC 27171, an internal mold of a

dorsal valve; CEGH-UNC 21084 and 27172, two conjoined
specimens. 1137 1138

1139 *Diagnosis*.—Shell semielliptical to subrectangular, moderately
1140 to strongly convexoconcave (mean length/width ratio 0.63) with
1141 subrectangular cardinal extremities; ventral median fold low,
1142 almost indistinct near commissure; dorsal sulcus shallow pos-
1143 teriorly, impersistent at anterior margin. Radial ribbing sub-
1144 equally to unequally parvicostellate. Ventral muscle field
1145 bilobed with suboval diductor scars extending 40–45% of valve
1146 length. Valve margin strongly thickened with peripheral rim
1147 separating disc and trail. Dorsal median ridge initially low and
1148 highest at approximately two-thirds valve length.

1149 *Occurrence*.—Type specimens and other materials considered
1150 in this study come from the Las Chacritas Formation exposed at
1151 Quebrada La Tuna, Cordón de La Trampa, San Juan Province,
1152 Argentina; middle Darriwilian.

1153 *Description*.—Shell to 16.5 mm in width, semielliptical to sub-
1154 rectangular in outline, with mean length/width ratio 0.63
1155 (N = 15); lateral profile moderately convexoconcave, slightly
1156 resupinate, typically 38% as deep as long. Cardinal extremities
1157 subrectangular, slightly acute in juvenile specimens. Ventral
1158 valve broadly concave with low carinate median fold originat-
1159 ing at umbonal area, becoming almost indistinct near commis-
1160 sure that is slightly sulcate or rectimarginate. Ventral interarea
1161 planar, catacline to gently procline, twice as high as dorsal.
1162 Arched imperforated pseudodeltidium covering one-half to two-
1163 thirds of delthyrium. Dorsal valve moderately to strongly con-
1164 vex, uniformly curved in lateral profile, with broad rounded
1165 median sulcus on its posterior third, becoming shallow to
1166 impersistent at anterior margin. Dorsal interarea planar, steeply
1167 anacline to orthocline. Notothyrium entirely covered by
1168 strongly convex chlidium. Ornamentation varying from sub-
1169 equally parvicostellate to unequally parvicostellate; accentuated
1170 costellae better defined on anterior half of largest individuals,
1171 with 6–8 finer parvicostellae between them; costellae 11 or 12
1172 per 2 mm (counted at 5 mm growth stage). Fine, evenly spaced
1173 concentric fila present on entire shell surface. Posterolateral
1174 rugae poorly defined or absent.

1175 Ventral interior with small, transversely triangular teeth
1176 supported by short strongly diverging dental plates partially
1177 masked by secondary deposits. Muscle field bilobed extending
1178 anteriorly near 40–45% of valve length; diductor scars suboval,
1179 bounded by ridges arising from anterior ends of dental plates;
1180 adductor field shorter than diductors, not raised, unbounded
1181 anteriorly. Valve margin strongly thickened with rounded
1182 subperipheral rim separating disc and trail. Vascula media
1183 straight, initially subparallel, subdivided toward valve margin.

←
Figure 11. (1–15) *Ahtiella coloradoensis* (Benedetto, 1998); ‘Green Member of the Sepulturas Formation,’ Los Colorados, northwestern Argentina: (1–4) holotype, external mold (1), latex cast (2), internal mold (3), and latex cast (4) of ventral valve, CEGH-UNC 13780; (5, 6) latex cast of exterior (5) and internal mold (6) of dorsal valve, CEGH-UNC 13817; (7) latex cast of ventral exterior, CEGH-UNC 13797; (8) ventral internal mold, CEGH-UNC 13802; (9) latex cast of ventral valve, CEGH-UNC 27187, showing incipient delthyrial cover; (10) latex cast of dorsal interior, CEGH-UNC 13808; (11) internal mold of juvenile dorsal valve, CEGH-UNC 13786; (12) internal mold of juvenile ventral valve, CEGH-UNC 13795; (13) dorsal internal mold, CEGH-UNC 13826; (14, 15) dorsal internal mold (14) and latex cast (15) of dorsal valve, CEGH-UNC 13823, showing detail of cardinalium. (16–18) *Ahtiella zarelae* Villas in Gutiérrez-Marco and Villas, 2007; San José Formation, Inambari River, Peru (illustrations from Gutiérrez-Marco and Villas, 2007, p. 552, figs. 4D, F, and J, reproduced under the Creative Commons Attribution License CC BY 4.0): (16) latex cast of dorsal valve exterior, MGM 5944X; (17) dorsal internal mold, MGM 5945X; (18) latex cast of ventral interior, MGM 5926X. All specimens dusted with ammonium chloride. Scale bars = 5 mm.

Table 1. Characters utilized in phylogenetic analysis.

1. Shell profile. 0 biconvex; 1 convexo-concave; 2 resupinate; 3 concavo-convex or planoconvex; 4 dorsal valve strongly arcuate, gibbous.
2. Outline. 0 semilelliptical; 1 transverse; 2 auriculate or alate.
3. Genuculation. 0 absent; 1 present; 2 strong defining disc and trail.
4. Radial ornamentation. 0 multicostellate or ramicostellate; 1 equally parvicostellate; 2 unequally parvicostellate.
5. Concentric fila. 0 absent; 1 present.
6. Rugae. 0 absent, 1 present on posterolateral areas; 2 strong on the entire surface.
7. Ventral fold. 0 absent; 1 present.
8. Dorsal sulcus. 0 absent; 1 present.
9. Ventral interarea. 0 apsacline; 1 catacline to procline.
10. Dorsal interarea. 0 anacline to orthocline; 1 catacline to hypercline
11. Chilidium. 0 absent; 1 short; 2 large.
12. Delthyrial cover: 0 absent; 1 separated into delthyrial plates; 2 pseudodeltidium.
13. Coverage of pseudodeltidium. 0 absent; 1 apical; 2 nearly complete.
14. Dental plates. 0 absent; 1 short recessive; 2 long well defined.
15. Ventral subperipheral rim. 0 absent; 1 low discontinuous; 2 strongly thickened; 3 defining a disc and trail.
16. Dorsal platform. 0 absent; 1 weak or discontinuous; 2 strong complete; 3 coalescing septules forming curved ridges.
17. Ventral muscle field. 0 short confined delthyrial cavity; 1 long and narrow; 2 small bilobed; 3 large subquadrate or subpentagonal; 4 large bilobed.
18. Ventral muscle bounding ridges. 0 absent; 1 present.
19. Dorsal muscle field. 0 indistinct; 1 quadripartite, orthoid.
20. Dorsal median ridge or myophragm. 0 absent or short; 1 prominent, long and rounded; 2 high bladelike.
21. Cardinal process. 0 simple ridgelike; 1 prominent or bulbous.
22. Ventral mantle canal system. 0 indistinct; 1 impressed on the valve margin; 2 strongly impressed on the entire surface.
23. Shell structure. 0 impunctate; 1 pseudopunctate.

1184 Gonadal areas poorly defined, striated. Areas between vascular
 1185 trunks finely pustulose. External ribbing not reflected on valve
 1186 interior.

1187 Dorsal interior with high, bladelike cardinal process erected
 1188 on moderately elevated subtriangular notothyrial platform. Socket
 1189 ridges thickened distally, diverging anteriorly from each other at
 1190 ~100°, bounding semiconical sockets. Median ridge initially low,
 1191 becoming highest at approximately two-thirds valve length, fading
 1192 near anterior margin. Muscle field indistinct in single available
 1193 internal mold. Dorsal mantle canal system faintly impressed with
 1194 straight inner branches of vascula media almost subparallel to
 1195 median ridge. Internal surface with coarse papillae near margin.

1196 *Etymology.*—Named after the original locality, Quebrada
 1197 La Tuna.

1198 *Materials.*—CEGH-UNC 27172–27183, 12 conjoined speci-
 1199 mens; CEGH-UNC 27168–27170, three fragmentary ventral
 1200 valves.

1201 *Remarks.*—This new species is readily distinguished from
 1202 *Ahtiella argentina* by the less transverse subrectangular shell
 1203 outline, nonalate cardinal extremities, catacline to procline ventral
 1204 interarea, and larger bilobed ventral muscle field. In addition, the
 1205 mantle vascular system in *A. tunaensis* n. sp. is barely impressed,
 1206 although this could be due to the lack of well-preserved internal

molds. Its shell outline is reminiscent of *A. quadrata*, from which
 it differs mainly in its much finer radial ribbing. The large bilobed
 ventral muscle field of *A. tunaensis* n. sp. is comparable to that of
 the Baltic species *A. lirata*, but differs from the latter species in its
 less convex and smoothly geniculated profile, better defined
 ventral fold and dorsal sinus, and indistinct rugae. The elongate
 oval ventral diductors of the Precordilleran species, although
 rather shorter, resemble those of *A. paucirugosa*. However, the
 Precordilleran species can be distinguished by its subrectangular
 shell, broad dorsal median ridge, and more deeply impressed
 mantle canals. The Peruvian species *A. zarelae* differs in having
 uniformly costellate radial ribbing, a more prominent ventral fold
 and deeper dorsal sulcus, a proportionally shorter and wider
 subpentagonal ventral muscle field, and strongly marked internal
 radial ornamentation at all growth stages.

Ahtiella famatiniana new species
 urn:lsid:zoobank.org:act:9AB173B1-73FE-4142-A2AF-
 A2DF1FD3ED9C
 Figure 10.9–10.24

2003a *Ahtiella* sp.; Benedetto, p. 210, pl. 18, figs. 18–20.

Type specimens.—Holotype, CEGH-UNC 27137, internal mold
 of ventral valve. Paratypes: CEGH-UNC 27135, internal mold
 of dorsal valve; CEGH-UNC 27149, conjoined specimen.

Q3 Table 2. Character state distribution for taxa included in phylogenetic analysis.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
<i>Hesperonomiella</i>	0	01	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	1	1	0	0	0	0
<i>M. transversa</i>	2	2	0	0	0	0	1	1	0	0	0	0	0	1	0	0	0	01	0	0	01	0	0
<i>A. lirata</i>	4	2	2	2	1	2	0	0	1	1	2	2	2	2	3	1	3	1	0	1	0	0	1
<i>A. jaanussoni</i>	4	2	2	2	1	2	0	1	1	1	1	?	?	?	?	1	?	?	1	1	0	0	1
<i>A. quadrata</i>	2	2	1	0	0	01	1	1	01	0	1	2	1	1	2	0	3	1	1	1	0	0	?
<i>A. paucirugosa</i>	2	2	2	1	0	1	1	1	1	0	1	2	1	01	3	3	4	1	01	1	01	2	?
<i>A. argentina</i>	2	2	2	2	1	1	1	1	0	0	12	2	2	1	3	3	0	0	01	1	01	2	1
<i>A. tunaensis</i>	12	0	2	12	1	0	1	1	0	0	2	2	2	2	3	1	4	1	0	1	0	01	1
<i>A. famatiniana</i>	2	22	1	0	0	01	1	1	1	1	1	2	2	2	3	1	3	1	0	1	0	1	?
<i>A. zarelae</i>	2	01	1	0	0	01	1	1	01	0	0	2	1	1	3	1	3	1	0	1	0	0	1
<i>A. coloradoensis</i>	2	01	1	0	0	0	1	1	1	0	0	?	01	1	2	1	3	1	01	1	0	01	?

- 1231 *Diagnosis*.—Shell transversely semielliptical (mean length/
1232 width ratio 0.48), alate to mucronate; ventral valve with strongly
1233 concave posterolateral areas and prominent carinate median
1234 fold; corresponding dorsal sulcus narrow and deep posteriorly,
1235 broader and wider at anterior margin. Ornamentation subequally
1236 multicostellate to incipiently parvicostellate, with primary
1237 costellae slightly more accentuated. Ventral muscle field almost
1238 pentagonal, extending to ~32% of valve length, with elongate
1239 diductor scars bounded laterally by curved ridges and shorter
1240 adductor field. Broad rounded dorsal median ridge fading
1241 anteriorly. Usually well-defined discontinuous dorsal platform.
1242 Vascular system deeply impressed only on disc margin and trail.
- 1243 *Occurrence*.—Cerro Morado Group, Las Pircas anticline, cen-
1244 tral Famatina Range, La Rioja Province. Northern margin of
1245 Chaschuil River (Loma del Médano), Catamarca Province.
1246 Dapingian.
- 1247 *Description*.—Shell transversely semielliptical, resupinate in
1248 lateral profile, up to 18 mm in width (mean 13 mm, N = 26),
1249 mean length/width ratio 0.48 (N = 20). Cardinal extremities
1250 alate, often projecting in rodlike mucrons (usually incompletely
1251 preserved). Ventral valve strongly concave with greatest con-
1252 cavity at posterolateral areas; carinate median fold stronger
1253 posteriorly, becoming wider and rounded anteriorly; anterior
1254 commissure broadly sulcate. Ventral interarea planar, procline
1255 or occasionally catacline, approximately twice as high as dor-
1256 sal. Delthyrium covered in apical two-thirds by semiconical
1257 pseudodeltidium. Dorsal valve moderately convex, 30–35% as
1258 deep as long, uniformly curved (nongeniculated) in lateral
1259 profile, with median sulcus initially narrow and deep, becoming
1260 broader and wider at anterior margin where it attains ~25% of
1261 valve width. Dorsal interarea planar, orthocline to steeply
1262 anacline, with triangular notothyrium covered posteriorly by
1263 convex chilidium. Radial ornamentation subequally multi-
1264 costellate, with rounded primary costellae often slightly more
1265 accentuated than second order ones giving ribbing a parvicos-
1266 tellate aspect; costellae increasing in number mainly by inter-
1267 polation in dorsal valve and by interpolation and dichotomy in
1268 ventral valve, typically 9 per 2 mm (counted at 5 mm growth
1269 stage). Ribs crossed by prominent, closely and evenly spaced
1270 fila (~20 per mm) and by few prominent growth discontinuities.
1271 Posterolateral rugae weakly defined or absent. Shell substance
1272 apparently impunctate.
- 1273 Ventral interior with strong triangular teeth supported by
1274 short dental plates diverging at ~120°; umbonal cavities
1275 partially filled by secondary deposits. Muscle field proportion-
1276 ally large, subpentagonal in outline, generally wider than long,
1277 typically extending 32% of valve length (maximum 36%);
1278 diductor scars elongate subtriangular, enclosed laterally and
1279 anterolaterally by curved ridges originating in front of dental
1280 plates; adductor field shorter and more deeply impressed than
1281 diductors, suboval to subrectangular in outline, not enclosed
1282 anteriorly by diductors. Valve margin strongly thickened,
1283 defining internal disc and trail separated by low, rounded
1284 peripheral rim. Vascula media broad, weakly impressed, slightly
1285 divergent; distal region of vascular system consisting of
1286 numerous radially arranged distal branches deeply cutting disc
1287 margin and prolonged on trail; small papillae between vascular
trunks. External ribbing weakly impressed only on juvenile
interiors. Gonadal areas large, occupying most posterolateral
areas, sculptured by roughly radial anastomosing ridges.
- Dorsal interior with simple bladeliike cardinal process
slightly enlarged anteriorly, erected on elevated subtriangular
notothyrial platform. Socket ridges strong, slightly thickened
and free distally, divergent anterolaterally at ~80°, bounding
deep semiconical sockets partially excavated under dorsal
interarea. Broad, rounded median ridge arising in front of
notothyrial platform, fading anteriorly at intersection with
peripheral rim. Muscle field weakly impressed, slightly wider
than long, extending anteriorly for 30% of valve length,
bounded laterally by low ridges; posterior adductor scars
subcircular, smaller than suboval anterior pair. Dorsal mantle
canal system indistinct or feebly impressed; vascula media
broad, divergent, branching toward valve margin. Coarse ridge-
like pustules or endospines coalescing to form well-defined
discontinuous platform, which extends along entire valve
margin except posterolateral extremities.
- Etymology*.—Named after one of the original localities, the
Famatina Range.
- Materials*.—CEGH-UNC 27131–17159, 29 specimens from
the volcanosedimentary unit exposed at Anticlinal Las Pircas,
north of Cachiyuyo River, central Famatina Range, La Rioja
Province; CEGH-UNC 27163–27167, five specimens from the
Chaschuil River, north Famatina Range, Catamarca Province.
- Remarks*.—In its size and shell convexity, nearly uniform
costellate ornamentation, high carinate ventral fold, and well-
developed dorsal peripheral rim, this species strongly resembles
A. zarelae from the upper Floian of Peru, but differs in having a
proportionally larger ventral muscle field reaching approxi-
mately one-third of the valve length; broader, rounded, non-
septiform dorsal median ridge; and vascular trunks strongly
impressed on the margin of the ventral disc and especially on the
trail. The Peruvian *Ahtiella* sp., which is known only by its
dorsal valve, can be distinguished from the Famatina species by
its finer and uniform ribbing, higher and shorter median ridge,
and more continuous, anteriorly excavated platform. *Ahtiella*
argentina can readily be distinguished from *A. famatiniana* n.
sp. by having a more transverse shell outline, unequally parvi-
costellate ornamentation, a smaller triangular ventral muscle
field, and a strongly impressed vascular system. The ventral
muscle field of *A. famatiniana* n. sp. is reminiscent of that of *A.*
tunaensis n. sp., but in the latter it is more definitely bilobed.
Moreover, the Precordilleran species differs in having sub-
rectangular cardinal extremities, a less pronounced ventral
median fold, and unequally parvicostellate ornamentation.
Ahtiella lirata resembles the Famatinan species in its alate or
mucronate cardinal extremities and its large subrectangular
ventral muscle field, but can be distinguished by its unequally
parvicostellate ornamentation, poorly defined (or absent) dorsal
sulcus and corresponding ventral fold, and a strongly convex
gibbous dorsal valve. *Ahtiella paucirugosa* externally resembles
A. famatiniana n. sp. in having a well-developed dorsal sinus
and ventral fold, moderate shell convexity, and almost indistinct
posterolateral rugae, but differs internally in its elongate ventral

1344 diductor scars extending to near the valve midlength, its deeply
1345 impressed mantle canals, and its high, septum-like dorsal median
1346 ridge. As far as can be judged from the incomplete material,
1347 *A. quadrata* resembles *A. famatiniana* n. sp. in its shell outline
1348 and morphology of its ventral muscle field, but the Welsh species
1349 differs in its coarser radial ornamentation. *Ahtiella concava*
1350 clearly differs in having a strongly impressed and longer dorsal
1351 muscle field and impersistent (much finer? smooth?) ribbing.

1352 *Ahtiella coloradoensis* (Benedetto, 1998)

1353 Figure 11.1–11.15

1355 ?1980 *Valcourea* sp.; Havlíček and Branisa, p. 23, pl. 2, fig. 7.

1356 1998b *Monorthis coloradoensis* Benedetto, p. 11, pl. 2, figs.

1357 15–28.

1358 2003a *Monorthis coloradoensis*; Benedetto, p. 211, pl. 24,

1359 figs. 5–9.

1360 **Holotype**.—CEGH-UNC 13780, internal mold of ventral valve
1361 from the ‘Green Member of the Sepulturas Formation,’ north-
1362 west of Los Colorados village, Cordillera Oriental, Jujuy Pro-
1363 vince, Argentina.

1364 **Diagnosis**.—Shell semielliptical, convexoplar to gently con-
1365 vexoconcave (mean length/width ratio 0.63) with right-angled
1366 cardinal extremities in adult specimens; ventral valve almost
1367 planar with low median fold and well-defined corresponding
1368 dorsal sulcus. Ornamentation equally multicostellate, occa-
1369 sionally ramicostellate in ventral valve; ribs 10–12 per 2 mm.
1370 Ventral muscle field subrectangular to subpentagonal, slightly
1371 wider than long, extending to ~31% of valve length, bounded
1372 by strong ridges in adult specimens; variably developed plat-
1373 form in dorsal valve. Vascular system indistinct excepting on
1374 periphery of each valve.

1375 **Occurrence**.—‘Green Member of the Sepulturas Formation’
1376 (sensu Astini et al., 2004a), Los Colorados area, western slope
1377 of Cordillera Oriental, Jujuy Province, Argentina. Middle-late?
1378 Darriwilian.

1379 **Description**.—See Benedetto (1998b).

1380 **Materials**.—In addition to the type material listed by Benedetto
1381 (1998b), new specimens CEGH-UNC 27187–27190 were col-
1382 lected from Quebrada Chamarra, northwest of Los Colorados
1383 village, Jujuy Province, Argentina.

1384 **Remarks**.—The overall morphology of the Los Colorados spe-
1385 cies, in particular its convexoplane shell profile, carinate ventral
1386 fold, equally multicostellate ornamentation, acute cardinal
1387 extremities, and orthoid cardinalia led Benedetto (1998b) to
1388 refer it to the hesperonomiid genus *Monorthis*. This assignment
1389 was further supported by the lack of evidence of pseudopunctae
1390 and pseudodeltidium, both considered diagnostic features of the
1391 plectambonitoids. As discussed previously, however, the dif-
1392 ferences between *Monorthis* and the basal species of *Ahtiella* are
1393 subtle and exhibit transitional characteristics, so that assigne-
1394 ment to one or the other genus depends largely on the weight

accorded to each character. The species *M. coloradoensis* is
reassigned here to *Ahtiella* mainly because of the incipient
internal geniculation in the ventral valve defining a low disc and
a trail, the large subquadrate ventral muscle field bounded by
widely splayed dental plates, and the presence of a faint dis-
continuous platform in the dorsal valve. In addition, a revision
of the type material and newly collected specimens of *M. col-
oradoensis* revealed an incipient apical delthyrial cover, like
some specimens of *M. cumillangoensis* (Figs. 6.15 and 11.9).

Havlíček and Branisa (1980) referred to *Valcourea* sp. a
few ventral valves from supposedly Darriwilian sandstones
exposed along the road Sucre-Potosí at Tambo Acachila,
Bolivia. The single illustrated interior is nearly indistinguishable
from *Ahtiella coloradoensis* but without dorsal valves and the
lack of information of external ornamentation, assignment to
this species must remain provisional.

Ahtiella coloradoensis resembles *A. famatiniana* n. sp. in
its large subrectangular (sometimes subpentagonal) ventral
muscle field extending anteriorly to near one-third of the valve
length, its well-defined peripheral ventral thickening, and its
incipient dorsal platform. Ornamentation is multicostellate in
both species but in *A. coloradoensis* it is more uniform tending
to be ramicostellate in the ventral valve. *Ahtiella coloradoensis*
mainly differs from the Famatinian species in its planar to gently
concave ventral valve profile, its less transverse shell outline, its
rectangular cardinal extremities, and its indistinct pseudodelti-
dium. It can be distinguished from *A. zarelae* by its lower and
rounded ventral fold, less pronounced geniculation, larger
ventral muscle field, and the presence of a conspicuous
peripheral rim in the ventral valve. In addition, internal ribbing
in the Peruvian species is strongly marked.

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