



Differential acute phase protein response induced in mice experimentally infected with *Brucella melitensis* 16M or with the vaccine strain Rev1.

Conde Bosque, L.^{1,2}, Iñarrea Lasheras, P.¹, Álava Martínez de Contrasta, M.A.^{1,2}, Jiménez de Bagüés Picazo, M.P.³, Iturralde Navarro, M.^{1,2*}

¹Dpto. Bioquímica y Biología Molecular y Celular, Facultad de Ciencias, ²Centro de Investigación Biomédica de Aragón (CIBA), ISS. Universidad de Zaragoza, 50009 Zaragoza, Spain. ³Unidad de Producción y Sanidad Animal, Centro de Investigación y Tecnología Agroalimentaria (CITA), Instituto Agroalimentario de Aragón – IA2 (CITA-Universidad de Zaragoza), 50059 Zaragoza, Spain. *Corresponding author: María Iturralde miturral@unizar.es.

This work was supported by the I.N.I.A Grants RTA 2013-00065-C02-01 and 02 from Spain.

Brucellosis affects at different species of animals and is zoonotic for humans. Vaccines are based on live attenuated bacteria and animals develop the same antibodies being impossible to distinguish serologically vaccinated animals from infected¹. Acute phase proteins (APPs), used as biomarkers², could be useful to solve this problem. THE AIM OF THIS WORK is to characterize the APPs response in mice infected with *B. melitensis* 16M or vaccinated with Rev1 to detect biomarkers differentiating between two conditions.

MATERIALS and METHODS

Kinetics of infection. Female balb/c mice (4 per lot) were intraperitoneally (ip) injected with 10⁴ colony forming units (CFU) of *B. melitensis* 16M or with the vaccine strain Rev1. Rev1 was also subcutaneously (sc) injected at 10⁵ CFU since it is the standard route and dose for vaccine administration. Control mice were ip injected with PBS. Weight and CFU from liver and spleen were determined at 7, 14, 21, 28 and 56 days. **In vitro studies.** Serum concentration of serum amyloid A (SAA), α-1 glycoprotein acid (AGP), haptoglobin (HPT) and hemopexin (HPX) were measured by ELISA. Liver and spleen mRNA relative units (RU) of this APPs were determined by qPCR. All data are represented as Mean±SEM and were analysed using independent samples t-Test with SPSS 16.0. *p<0.05, **p<0.01, ***p<0.001.

RESULTS and DISCUSSION

Differences were found in APPs serum concentration among mice injected with *B. melitensis* 16M, Rev1 ip and Rev1 sc. These differences were also reflected in liver mRNA levels of these proteins.

Peaking on day 14, AGP, HPT and HPX serum concentration were greater in mice injected with Rev1 ip, intermediate with *B. melitensis* 16M and similar to control with Rev1 sc. The peak of SAA was also reached on day 14 for mice inoculated with Rev1 ip, but did it earlier (day 7) for mice inoculated with *B. melitensis* 16M. These SAA values were higher than those for Rev1 on day 3 (sc) and 7 (ip or sc) (Figure 1).

APPs serum concentration values were in agreement with spleen and liver inflammation (weight) caused by any of these inoculations. Maximum CFU values were reached before of those of inflammation, being 1.5 fold lower for Rev1 sc (Table 1).

APPs gene expression was upregulated in liver and downregulated in spleen, except for SAA in the latter. Time-course of liver APPs gene expression was peaked on day 14 (or day 7 for *B. melitensis* 16M). Mice inoculated with Rev1 sc showed a high level of APPs mRNA that did not correspond to the protein concentration in serum. This effect was more evident for HPX (Figure 2). Thus, Rev1 sc inoculation could cause protein malformations or inhibition of the APPs secretion mechanism by liver. Spleen did not contribute to mRNA synthesis of APPs except for SAA. AGP, HPT and HPX were downregulated on day 7 post-inoculation. A recovery of their expression was observed at 14-21 days, except for those injected with *B. melitensis* 16M (Figure 3).

Table 1. Number of *Brucella* (CFU) and liver and spleen weight (g). Mean±SEM; n=4.

Spleen	Time post-inoculation	3 days	7 days	14 days	21 days	28 days	56 days
	log CFU						
g	<i>B. melitensis</i> 16M	5.19±0.07	6.27±0.07	5.00±0.11	4.73±0.12	4.66±0.39	3.55±0.52
	Rev1 ip	4.71±0.12	6.53±0.06	5.65±0.16	4.87±0.18	3.42±0.36	2.93±0.61
	Rev1 sc	1.20±0.71	3.45±0.58	4.40±0.42	3.67±0.72	2.69±0.69	0.00±0.00
g	<i>B. melitensis</i> 16M	0.14±0.01	0.23±0.02	0.31±0.04	0.25±0.02	0.22±0.02	0.15±0.01
	Rev1 ip	0.11±0.01	0.12±0.01	0.45±0.04	0.33±0.06	0.21±0.01	0.20±0.01
	Rev1 sc	0.11±0.11	0.14±0.14	0.20±0.03	0.17±0.02	0.17±0.02	0.13±0.01
Liver	Time post-inoculation	3 days	7 days	14 days	21 days	28 days	56 days
	log CFU						
g	<i>B. melitensis</i> 16M	4.72±0.06	4.79±0.07	3.15±0.16	2.54±0.29	2.25±0.79	0.99±0.57
	Rev1 ip	4.78±0.07	4.81±0.17	3.02±0.26	2.34±0.78	1.43±0.52	0.36±0.36
	Rev1 sc	0.48±0.48	2.64±0.29	2.90±0.14	1.72±0.62	1.48±0.60	0.56±0.56
g	<i>B. melitensis</i> 16M	1.10±0.09	1.41±0.05	1.20±0.11	1.38±0.06	1.36±0.05	1.34±0.14
	Rev1 ip	1.09±0.08	1.06±0.03	1.52±0.06	1.40±0.09	1.24±0.06	1.33±0.07
	Rev1 sc	1.14±0.07	1.11±0.05	1.20±0.07	0.94±0.11	1.00±0.06	1.22±0.06

Figure 1. Time-course of serum concentration of SAA, AGP, HPT and HPX. Mean±SEM; n=4. ***/**/* vs Rev1 ip; */**/* vs Rev1 sc.

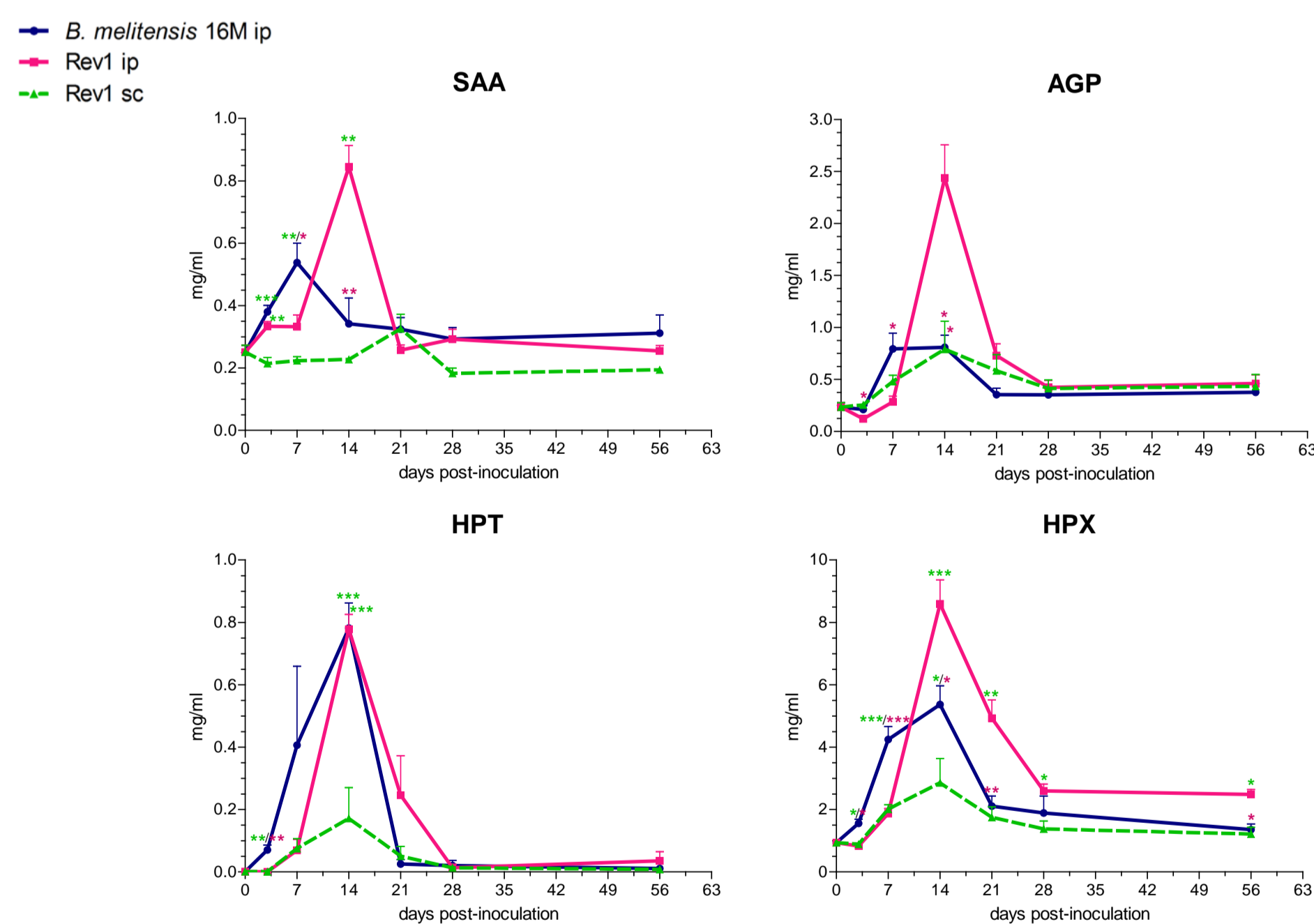


Figure 2. Time-course of liver mRNA level of SAA, AGP, HPT and HPX. Mean±SEM; n=2.

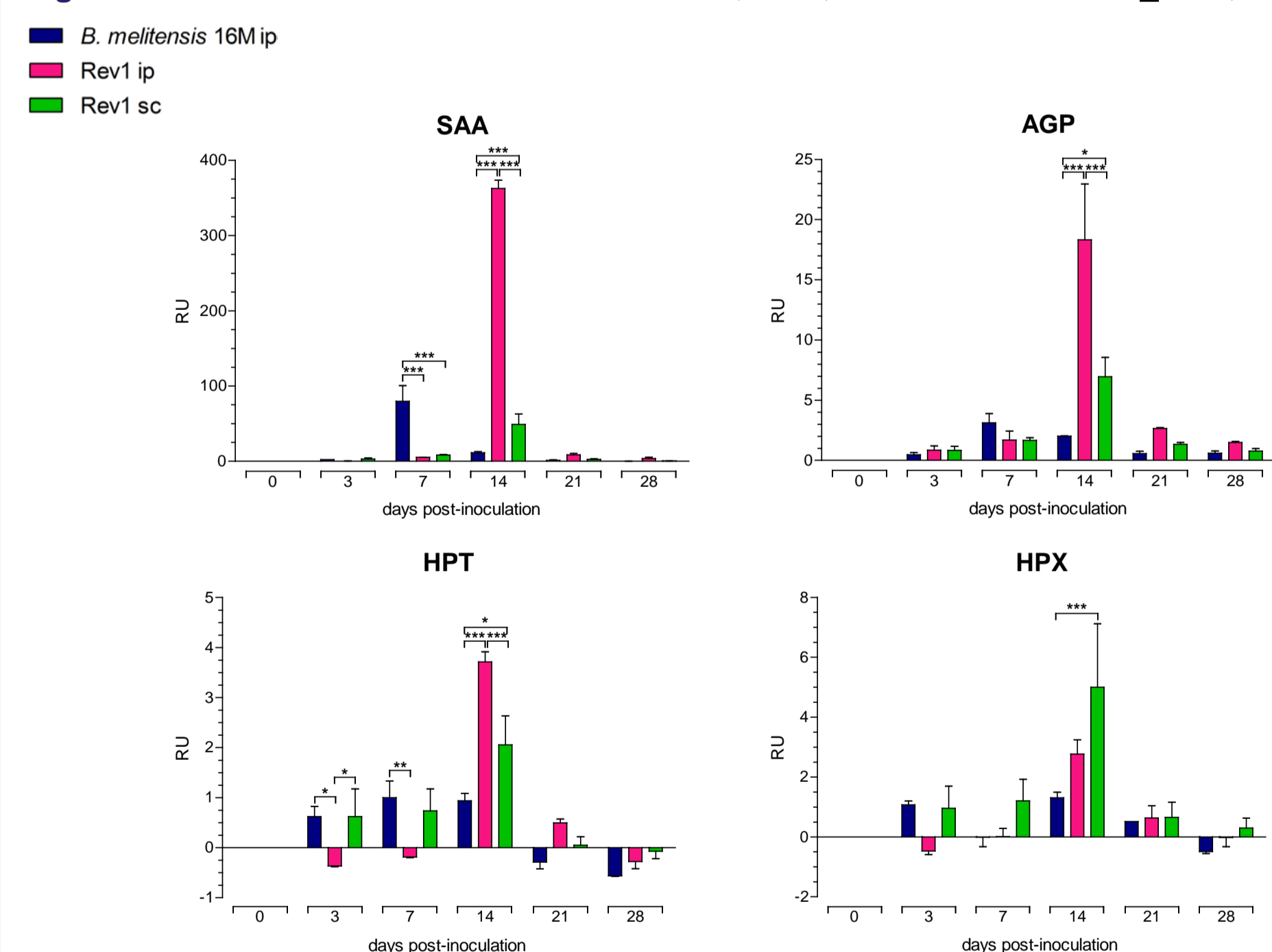
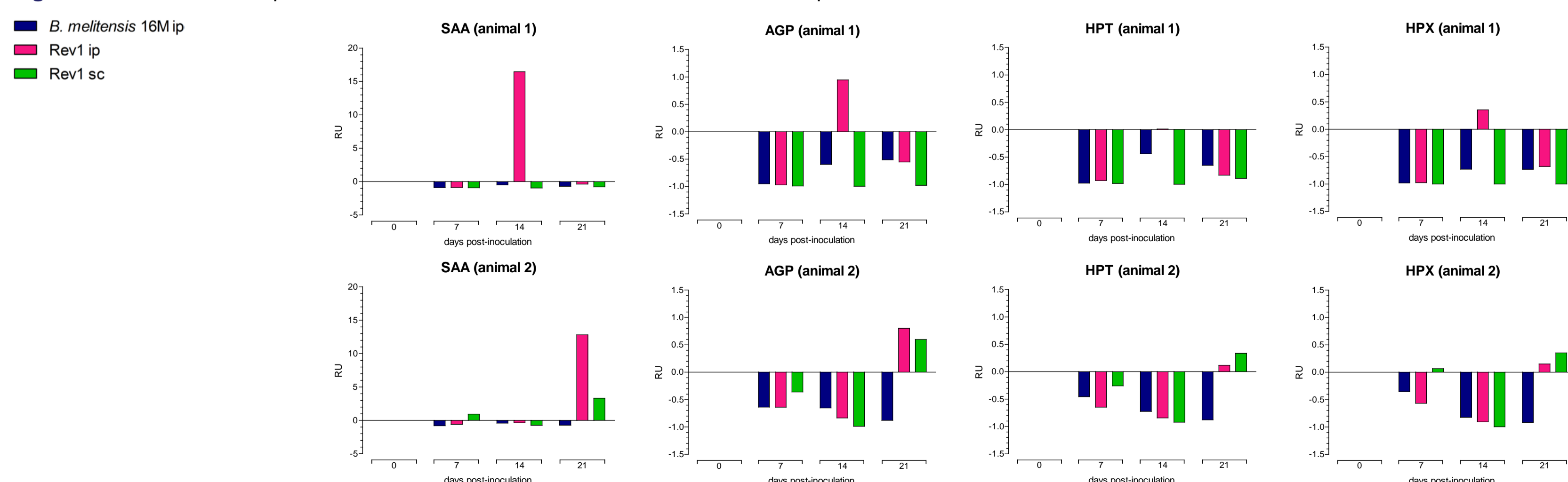


Figure 3. Time-course of spleen mRNA level of SAA, AGP, HPT and HPX. Two separate animals.



CONCLUSION: APPs show great promise as biomarkers to differentiate between *Brucella melitensis* infected and vaccinated animals. Specifically, HPT and HPX during medium term and SAA during the early stage of *Brucella*-induced inflammation.

REFERENCES

- Jiménez de Bagüés MP, Elzer PH, Jones SM, Blasco JM, Enright FM, Schurig GG, Winter AJ. Vaccination with *Brucella abortus* rough mutant RB51 protects BALB/c mice against virulent strains of *Brucella abortus*, *Brucella melitensis*, and *Brucella ovis*. Infect Immun. 1994 Nov;62(11):4990-6
- Carpintero R, Alonso C, Piñeiro M, Iturralde M, Andrés M, Le Potier MF, Madec F, Alava MA, Piñeiro A, Lampreave F. Pig major acute-phase protein and apolipoprotein A-I responses correlate with the clinical course of experimentally induced African Swine Fever and Aujeszky's disease. Vet Res. 2007 Sep-Oct;38(5):741-53