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IMPROVEMENT OF ALGAL-ALGINATE BEAD STABILITY BY ZEOLITE MOLECULAR SIEVES 13X AND ITS APPLICATION IN BIOSORPTION

SEYED AMIREBRAHIM EMAMI MOGHADDAM

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SEYED AMIREBRAHIM EMAMI MOGHADDAM

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

September 2019



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DEDICATION

This thesis is dedicated to

My lovely family: mother, father & sister

With love, respect and a bunch of memories

Indeed, we belong to Allah and indeed to Him we will return.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

IMPROVEMENT OF ALGAL-ALGINATE BEAD STABILITY BY ZEOLITE MOLECULAR SIEVES 13X AND ITS APPLICATION IN BIOSORPTION

By

SEYED AMIREBRAHIM EMAMI MOGHADDAM

September 2019

Chair Faculty : Mohd Razif Harun, PhD : Engineering

The interest in utilizing algae for wastewater treatment has been increased due to many advantages. Algae-Wastewater treatment system offers a cost-efficient and environmentally friendly alternative to conventional treatment processes. However, the recovery of free suspended algae from the treated effluent is one of the challenges during the treatment process. Therefore, the application of immobilized algae is a good approach to resolve the harvesting issue. Up to now, most of the algal immobilization has been done using cell entrapment method in which alginate (a natural polymer) has been applied as a carrier. Although alginate provides advantages in terms of biocompatibility, nontoxicity, cost-effectiveness, etc., this material has low stability to the chelating agents and a similar charge with cell surface of microorganisms, hence, it easily contributes to the leakage of large molecules due to the open lattice structure. Therefore, this study aims to improve the stability of *Chlorella*-Alginate Beads (CABs) by zeolite molecular sieves 13X (an aluminosilicate mineral with sodium ion) and further examined the potential use of the synthesized Zeolite 13X-Algal-Alginate Beads (ZABs) for copper biosorption from aqueous solution.

The immobilization was done via the entrapment of green living microalgae, *Chlorella vulgaris* within alginate/powdered zeolite 13X hydrogels. Cross-linking was carried out using 0.1 M CaCl₂ solution. The stability of the beads was tested by immersing them in a phosphate buffer solution at pH 7 as a chelating agent. Different process variables, including ratio of zeolite/alginate, pH and volume of beads were optimized using response surface methodology (RSM) to obtain the algal beads with high stability. Dissolution time of synthesized Zeolite-Algal-Alginate Beads (ZABs) in a chelating agent revealed a significant improvement on the beads stability (78.5 \pm 0.5 min) compared to the control beads (51.5 \pm 0.5 min) under the optimum conditions of zeolite/alginate (1.5:1), pH 5 and 2% of beads. Monitoring cell growth during 5 days of incubation showed good biocompatibility of zeolite 13X. Scanning electron microscopy (SEM) indicated rough surface and spherical shapes of ZABs. Brunauer-Emmett-Teller (BET) analysis revealed higher surface area for ZABs than other ABs. Energy dispersive

X-ray spectroscopy (EDX) and Fourier transform infrared spectroscopy (FTIR) of ZABs confirmed the presence of zeolite 13X within the matrix. The zeta potential value of ZABs (-23.33 ± 0.29 mV) indicated that the beads were relatively stable.

In addition, the potential use of ZABs for copper biosorption was evaluated and compared with Blank-Alginate Beads (BABs) and *Chlorella*-Alginate Beads (CABs). Different process parameters were investigated including contact time, pH and initial metallic ion concentration. It was found that the maximum biosorption capacity of ZABs was 85.88 mg/g biosorbent achieved at 180 min, pH 5 and initial metallic ion concentration of 150 mg/l whereas the maximum biosorption capacity of 70.02 and 77.32 mg/g biosorbent was obtained for BABs and CABs, respectively. ZABs showed higher stability than BABs and CABs in biosorption-desorption cycles. The kinetic and equilibrium data were analyzed via reaction/diffusion and Langmuir/Freundlich models, respectively. Scanning electron microscopy (SEM), Energy dispersive X-ray spectroscopy (EDX) and Fourier transform infrared spectroscopy (FTIR) analyses revealed bonded metal ion to the ABs.

The findings of this research confirmed that modification of algal-alginate beads by zeolite molecular sieves 13X has the potential to improve the beads stability and their biosorption capacity.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENAMBAHBAIKAN KESTABILAN MANIK ALGA-ALGINAT OLEH PENAPIS MOLEKUL ZEOLIT 13X DAN KEGUNAANNYA DALAM PENJERAPAN BIO ION LOGAM

Oleh

SEYED AMIREBRAHIM EMAMI MOGHADDAM

September 2019

Pengerusi Fakulti : Mohd Razif Harun, PhD : Kejuruteraan

Kepentingan penggunaan alga untuk rawatan air sisa kian meningkat disebabkan oleh pelbagai manfaat. Sistem rawatan Alga-Air sisa menawarkan alernatif yang kos efektif alam berbanding dengan proses-proses rawatan konvensional. dan mesra Walaubagaimanapun, pemulihan alga terampai yang bebas daripada efluen yang telah dirawat merupakan salah satu cabaran semasa proses rawatan. Oleh itu, aplikasi alga pegun adalah pendekatan yang baik untuk mengatasi isu penuaian. Sehingga kini, kebanyakan alga pegun telah dilakukan menggunakan kaedah pemerangkapan sel dimana alginat (polimer semulajadi) telah digunakan sebagai pembawa. Walaupun alginat mempunyai banyak kelebihan dari segi biokompatibiliti, ketidaktoksikan, keberkesanan kos, dan lain lain, bahan ini mempunyai kestabilan yang rendah kepada agen pengkelat dan berkongsi cas yang sama dengan permukaan sel mikroorganisma, justeru, hal ini mudah menyumbang kepada kebocoran molekul-molekul yang besar disebabkan oleh struktur kekisi yang terbuka. Oleh itu, tujuan kajian ini adalah untuk menambah baik kestabilan manik-manik Chlorella-alginat dengan penapis molekul zeolit 13X (sejenis mineral aluminosilikat dengan ion sodium) dan untuk terus memeriksa keupayaan penggunaan manik-manik Zeolit 13X-Alga-Alginat yang telah disintesis untuk penjerapan bio kuprum daripada larutan berair.

Kaedah pegun telah dijalankan melalui pemerangkapan mikroalga hijau yang hidup, *Chlorella vulgaris* didalam gel-gel hidro alginat/serbuk zeolit 13X. Pautan silang telah dilakukan dengan menggunakan larutan CaCl₂ sebanyak 0.1 M. Kestabilan manik-manik telah diuji dengan merendamkam kesemuanya di dalam larutan penimbal fosfat pada pH 7 sebagai agen pengkelat. Proses pembolehubah-pembolehubah yang berbeza, termasuklah nisbah zeolit/alginat, pH dan bilangan manik-manik telah dioptimumkan menggunakan kedah gerak balas permukaan (RSM) untuk mendapatkan manik-manik alga yang mempunyai kestabilan yang tinggi. Masa pelarutan untuk manik-manik Zeolit-Alga-Alginat (ZABs) yang telah disintesis di dalam agen pengkelat menunjukkan penambahbaikan yang begitu signifikan untuk kestabilan manik-manik (78.5 \pm 0.5 min)

berbanding dengan manik-manik kawalan (51.5 \pm 0.5 min) pada keadaan-keadaan yang optimum untuk zeolit/alginat (1.5:1), pH 5 dan sebanyak 2% manik-manik. Pemonitoran pertumbuhan sel selama 5 hari masa inkubasi telah menunjukkan biokompatibiliti yang baik bagi zeolit 13X. Mikroskop pengimbas elektron (SEM) telah memperlihatkan permukaan yang kasar dan bentuk-bentuk sfera bagi ZABs. Analisis Brunauer-Emmett-Teller (BET) telah menunjukkan luas permukaan yang tinggi untuk ZABs berbanding dengan ABs yang lain. Spektroskopi Tenaga Penyerakan Sinar-X (EDX) dan Spektroskopi Inframerah Transformasi Fourier (FTIR) pada ZABs telah mengesahkan kehadiran zeolit 13X didalam matriks. Nilai keupayaan zeta untuk ZABs (-23.33 ± 0.29 mV) telah menunjukkan bahawa manik-manik tersebut adalah begitu stabil.

Tambahan lagi, keupayaan penggunaan ZABs sebagai penjerap bio untuk menyingkirkan ion logam kuprum dari larutan berair telah pun dinilai menggunakan manik-manik kawalan kosong-Alginat (BABs) dan manik-manik *Chlorella*-Alginat (CABs) sebagai sistem-sistem kawalan. Proses-proses parameter yang berbeza telah diselidik termasuklah waktu sentuh, pH dan kepekatan awal ion logam. Hal ini menemukan kapasiti penjerapan bio paling maksimum pada ZABs ialah 85.88 mg/g yang telah dicapai pada 180 minit, pH 5 dan kepekatan awal ion logam pada 150 mg/l manakala kapasiti penjerapan bio bagi BABs dan CABs, adalah masing-masing dicapai pada penjerap bio sebanyak 70.02 mg/g dan 77.32 mg/g. ZABs telah menunjukkan kestabilan yang lebih tinggi berbanding BABs dan CABs dalam kitaran-kitaran penjerapan bio-penyahjerapan. Data kinetik dan data keseimbangan masing-masingnya telah dianalisis melalui reaksi/serapan dan model-model Langmuir/Freundlich. Analisis-analisis Mikroskopi pengimbas elektron (SEM), Spektroskopi Tenaga Penyerakan Sinar-X (EDX) dan Spektroskopi Inframerah Transformasi Fourier (FTIR) telah mendedahkan keterikatan ion logam kepada ABs.

Penemuan-penemuan melalui kajian ini telah mengesahkan bahawa pengubahsuaian bagi manik-manik alga-alginat oleh penapis molekul zeolit 13X mempunyai keupayaan untuk menambah baik kestabilan manik-manik dan kemampuan penjerapan bio mereka.

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I certify that a Thesis Examination Committee has met on 5 September 2019 to conduct the final examination of Seyed Amirebrahim Emami Moghaddam on his thesis entitled "Improvement of Algal-Alginate Bead Stability by Zeolite Molecular Sieves 13X and its Application in Biosorption" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Thomas Choong Shean Yaw, PhD Professor Ir. Faculty of Engineering Universiti Putra Malaysia (Chairman)

Siti Mazlina bt Mustapa Kamal, PhD

Associate Professor Ir. Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Wan Azlina binti Wan Ab Karim Ghani, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Azlina Harun @ Kamaruddin, PhD

Professor School of Chemical Engineering Universiti Sains Malaysia Malaysia (External Examiner)

ROBIAH BINTI YUNUS, PhD Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 22 October 2019

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Razif Harun, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairman)

Mohd Noriznan Mokhtar, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Member)

Rabitah Zakaria, PhD

Senior Lecturer Faculty of Engineering Universiti Putra Malaysia (Member)

ROBIAH BINITI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

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Signature: Name of	
Chairman of	
Supervisory	
Committee:	Assoc. Prof. Dr. Mohd Razif Harun
Signature:	
Name of	
Member of	
Supervisory	
Committee:	Assoc. Prof. Dr. Mohd Noriznan Mokhtar
Signature:	
Name of	
Member of	
Supervisory	
Committee:	Dr. Rabitah Zakaria

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LIST OF ABBREVIATIONS/GLOSSARY OF TERMS

Abbreviation	Definition
AAS	Atomic Absorption Spectrophotometer
ABs	Alginate Beads
ANOVA	Analysis of Variance
ATR	Attenuated Total Reflectance
BABs	Blank-Alginate Beads
BC	Biomass Concentration
BET	Brunauer-Emmett-Teller
CABs	Chlorella-Alginate Beads
CCD	Central Composite Design
EDX	Energy Dispersive X-ray Spectroscopy
FTIR	Fourier Transform Infrared Spectroscopy
OD	Optical Density
RSM	Response Surface Methodology
SEM	Scanning Electron Microscopy
WC	Water Content
ZABs	Zeolite 13X-Algal-Alginate Beads

	Glossary	Definition	
	А	Code of ratio of zeolite/alginate	
	b	Langmuir model Constant	(l/mg)
	В	Code of pH	
	С	Code of volume of beads	(%)
	C_0	Initial metal ion concentration	(mg/l)
	C_e	Concentration of metal ion at equilibrium	(mg/l)
	D	Weight of dried sample	(g)
	I	Diffusion model constant	
	k _I	Rate constant of pseudo-first order biosorption	(min ⁻¹)
	<i>k</i> ₂	Rate constant of pseudo-second order biosorption	[g/(mg min)]
	<i>k</i> _d	Intraparticle diffusion rate constant	[mg/(g min ^{0.5})]
	K _F	Freundlich model constant	$[(mg/g) (mg/l)^n]$
	m	Amount of biosorbent	(g)
	n	Freundlich model exponent	
	q	Biosorption capacity	(mg/g)
	q_e	Biosorption capacity at equilibrium	(mg/g)
	q_m	Maximum theoretical biosorption capacity	(mg/g)
	q_t	Biosorption capacity at time of t	(mg/g)
	R ²	Correlation coefficient	
	$R_{\rm L}$	Dimensionless separation factor	
(\mathbf{C})	Std	Standard order	
U	t	Time	(min)
	V	Solution volume	(1)

W	Weights of wet sample	(g)
Y	Code of dissolution time	(min)



CHAPTER 1

INTRODUCTION

1.1 Background of study

Water is the basic element of life on earth. The demand for water in the entire world is growing fast for domestic, agricultural and industrial activities. However, the availability and quality of water resources face severe threats due to the industrialization and rapid economic development, hence, producing a huge amount of wastewater. This wastewater contains organics, suspended solids and hazardous materials such as heavy metals which are not biodegradable. These materials tend to accumulate in organisms and potentially cause severe contamination and diseases (Inglezakis, Loizidou, & Grigoropoulou, 2003; Kwakye, McMinimy, & Aschner, 2016). Hence, wastewater treatment is essential to prevent deterioration of the environment and solve the issue of water shortage and health.

Wastewater treatment consists of removing or decreasing a number of hazardous substances such as chemicals and biological pollutants (Eroglu, Smith, & Raston, 2015). The selection of the treatment approaches is strongly influenced by their characteristics and compositions. Various wastewater treatment approaches have been used such as chemical, physical and biological (Eroglu et al., 2015). Advanced oxidation, electrocoagulation and flocculation are the most common methods of physical and chemical treatment whereas suspended or activated sludge process is an example of the biological treatment that is widely applied (Das & Adholeya, 2015). Although these conventional methods have successfully treated various wastewater sources, they have some limitations. The physical and chemical methods have the problems of high-energy requirements, incomplete removal of heavy metals, generation of secondary pollutants, complex operation, and high cost (Das & Adholeya, 2015; Krishnani, Meng, Christodoulatos, & Boddu, 2008; Quintelas et al., 2009), whereas biological method deals with the problems of the easy washout and low biomass concentration (Das & Adholeya, 2015). To overcome the issues, the use of microalgae to treat wastewater is currently of global interest due to its advantages.

Microalgal cells have the ability to uptake nutrients such as phosphorus, nitrogen and ammonium as well as heavy metals to reduce BOD in wastewater (Abdel-Raouf, Al-Homaidan, & Ibraheem, 2012; Aziz & Ng, 1992; Phang & Ong, 1988; Sydney et al., 2011) and also simultaneously capture carbon dioxide from the atmosphere during photosynthesis, hence, decreasing the greenhouse gaseousness (Zeng, Danquah, Chen, & Lu, 2011; Zeng et al., 2012). Also, the wastewater can be considered as a cheaper nutrient source for the growth of microalgae. However, at the current state, the biological nutrient removal technologies, including the use of microalgae, have not been yet to be competitive in the wastewater industries. The main issue in integrating algae with wastewater treatment contributes to the high cost of recovery of the treated effluent and

biomass using current dewatering methods such as centrifugation and filtration (Zeng et al., 2012). As an option, the existing cell suspended method can be replaced with immobilization method. Not only cell immobilization simplify the separation process, but it also offers other advantages such as higher cell density, higher productivity, better cell stability, and biomass recirculation (Das & Adholeya, 2015; Djukić-Vuković, Jokić, Kocić-Tanackov, Pejin, & Mojović, 2016; Eroglu et al., 2015; Idris & Suzana, 2006; Vasilieva, Lobakova, Lukyanov, & Solovchenko, 2016).

One of the most important parts of the immobilization technique is selecting a suitable carrier. Literature show that most of the algal immobilization are carried out using cell entrapment method in which alginate (a natural polymer) has been applied as a carrier (Bayramoglu & Arıca, 2009; Bayramoğlu, Tuzun, Celik, Yilmaz, & Arica, 2006; da Costa & Leite, 1991; Kondo, Hirayama, & Matsumoto, 2013; Mallick & Rai, 1993; I. Moreno-Garrido, Campana, Lubián, & Blasco, 2005; Petrovič & Simonič, 2016; Rai & Mallick, 1992; Shen, Gao, & Li, 2017; Wan Maznah, Al-Fawwaz, & Surif, 2012; Wilkinson, Goulding, & Robinson, 1990). The reason that researchers tend to use alginate for the immobilization of microalgae is due to its advantages, including biocompatibility, simple preparation, cost-effectiveness, and nontoxicity (Desmet et al., 2015; I. Moreno-Garrido et al., 2005; Petrovič & Simonič, 2016). Zhang et al. synthesized a biocompatible hybrid matrix for the encapsulation of Chlamydomonas reinhardtii through modifying the composition (alginate, polycation and silica). They reported good chemical stability to chelating agents for the hybrid beads (Zhang, Wang, Charles, Rooke, & Su, 2016). In another study, Desmet et al. synthesized highly porous (Ca-alginate-SiO₂-polycation) shell:(Na-alginate-SiO₂) core hybrid beads for the encapsulation of Dunaliella tertiolecta. The authors reported durability of the beads (Desmet et al., 2015). Also, Pannier et al. developed alginate/silica hybrid materials for the immobilization of *Chlorella vulgaris*. They reported an improvement to the stability in salt-containing solutions compared to alginate gels using Ca2+-ions (Pannier, Soltmann, Soltmann, Altenburger, & Schmitt-Jansen, 2014).

Zeolite is another material that can be an appropriate candidate for the modification of algal-alginate beads. Zeolites are inorganic silica materials (Sakaguchi, Matsui, & Mizukami, 2005) classified into three groups of natural, modified and synthetic (Yuna, 2016). They are widely applied as ion exchangers, catalysts (Sakaguchi et al., 2005), and also as a support for biomass immobilization (Al-Hassan et al., 1991; Corona-González et al., 2014; Djukić-Vuković et al., 2016; Djukić-Vuković, Mojović, Jokić, Nikolić, & Pejin, 2013; Fernández et al., 2007; Figueroa-Torres et al., 2016; Lameiras, Quintelas, & Tavares, 2008; Mery et al., 2012; Pazos, Branco, Neves, Sanromán, & Tavares, 2010; Quintelas et al., 2009; Shindo, Takata, Taguchi, & Yoshimura, 2001; Weiß et al., 2010) because of their characteristics, including suitable thermostability performance, resistance to microbial degradation, and cost-effectiveness (Das & Adholeya, 2015; Zhou, Li, An, Fu, & Sheng, 2008).

Zeolite molecular sieves 13X is a type of synthetic zeolites that its structure involves silica, alumina and sodium ions. Sodium ions along with silica can help to improve the stability of the beads. Furthermore, the existence of sodium ions can make the zeolite 13X surface positively charged (Djukić-Vuković et al., 2013); hence, it can make the alginate matrix less negative compare to the common type resulting in more algal cell affinity to diffuse into the gel matrix reducing cell leak out from the matrix. Furthermore, because of its ion exchange capacity and microporous structure, zeolite 13X can attend in metal sorption process along with algal-alginate matrix resulting in an improvement in sorption capacity.

This study investigates the stability of algal-alginate beads by modifying the matrix by zeolite molecular sieves 13X. The optimization of process variables and evaluation of kinetic and equilibrium models are included in this study. Lastly, the examination of the synthesized beads in metallic biosorption process are carried out to determine its effectiveness.

1.2 Problem statement

There are some problems on the way of using alginate for cell immobilization. Alginate beads have low stability to the chelating agents, hence, they contribute to the leakage of large molecules due to the open lattice structure (Smidsrod & Skjak-Braek, 1990; Zhang et al., 2016). Furthermore, the alginate matrix has a negative charge (Pannier et al., 2014; Smidsrod & Skjak-Braek, 1990) and the cell surface of microorganisms such as *Chlorella* has also a negative charge (comes from dissociation of some groups such as uronic acid and/or sulfate groups) (Eroglu et al., 2012) that do not easily diffuse into the gel matrix. Although in case of successful immobilization, the cells tend to leak out from the matrix. In order to overcome the stability drawback, the alginate matrix has been modified with silica by researchers, and their findings showed the good potential of silica to improve the stability (Desmet et al., 2015; Pannier et al., 2014; Zhang et al., 2016).

In this study, algal-alginate beads were modified by zeolite molecular sieves 13X to overcome drawbacks of low stability and less cell affinity for the immobilization of green living microalgae, *Chlorella vulgaris*. Furthermore, the feasibility of the beads for the biosorption of Cu²⁺ metal ion from aqueous solution was investigated. The main reason for the selection of copper metal ion was due to the fact that it is among the most toxic pollutants that potentially cause serious problems such as kidney damage, anaemia, etc. when expose to humans. The main copper sources in industrial wastewaters involve petroleum, mining, fertilizers, electroplating, metal cleaning plating baths, pulp and paper industries (Al-Rub, El-Naas, Ashour, & Al-Marzouqi, 2006). Thus, treatment of water resources polluted with copper metal ion is vital.

1.3 Research objectives

This study is designed to achieve the following objectives:

- a) To investigate the stability of algal-alginate beads by modifying the matrix using zeolite molecular sieves 13X
- b) To optimize the process variables of the algal beads using response surface methodology (RSM) to obtain the beads with high stability
- c) To examine the biosorption capacity of the synthesized algal beads in metallic process
- d) To evaluate the kinetic and equilibrium models of biosorption process by the algal beads

1.4 Scope of the study

1.4.1 Investigation of the algal beads stability

The algal beads stability was investigated by immersing them in a chelating agent along with their characterizations through zeta potential, scanning electron microscopy (SEM), Brunauer-Emmett-Teller (BET), energy dispersive X-ray spectroscopy (EDX), and Fourier transform infrared spectroscopy (FTIR).

1.4.2 Optimization of process variables of the algal beads

In order to find the optimum formulation for the synthesis of the algal beads with high stability, response surface methodology (RSM) was used in which the variables were ratio of zeolite/alginate (0.5:1, 1:1 and 1.5:1), pH (5, 7 and 9) and volume of beads (2, 6 and 10 %) with the response of dissolution time in a chelating agent.

1.4.3 Examination of the biosorption capacity of the algal beads

The synthesized algal beads were applied for the copper metal biosorption from aqueous solution. The effects of different operational parameters, including contact time (0-240 min), pH (3-6) and initial metallic ion concentration (5-200 mg/l) on the biosorption capacity of the metal ion were investigated. SEM, EDX and FTIR analyses were used for the beads characterization in biosorption studies.



1.4.4 Evaluation of kinetic and equilibrium models

The kinetic and equilibrium models of the biosorption process by the algal beads were evaluated through experimental data. The kinetic data were analyzed by reaction (pseudo-first and pseudo-second order) and diffusion models. The equilibrium data were studied via Langmuir and Freundlich models.

1.5 Significance of the study

This study modifies *Chlorella*-alginate beads by zeolite molecular sieves 13X to improve the algal matrix stability, its cell affinity and the metal sorption capacity. The findings of this research have a significant impact on various applications such as municipal and/or industrial wastewater treatment processes where the improved stability and biosorption capacity of algal-alginate beads is essential.

1.6 Structure of the thesis

There are five chapters in this thesis of which each chapter explains the sequence of the research:

Chapter 1 presents the background on the subject, problem statement, research objectives, scope of the study (including investigation of the algal beads stability, optimization of process variables of the algal beads, examination of the biosorption capacity of the algal beads, and evaluation of kinetic and equilibrium models), significance of the study, and structure of the thesis.

Chapter 2 covers literature review on the subject including the following parts: immobilization (types of immobilization, operational modes of immobilization and ideal carrier for immobilization), algal immobilization (microalgae and its applications, immobilization of algal biomass and biosorption process and its mechanism), zeolite and its application in biotechnology (zeolite and its classification and applications of zeolite), and potential application of zeolite 13X to modify algal-alginate beads.

Chapter 3 involves the materials and methods describing the experimental procedure in the current research. This chapter has been divided into two sections. The first section is related to synthesize of the beads, including preparation of zeolite molecular sieves 13X, microalgae and culture conditions (preparation of medium and cultivation of *Chlorella vulgaris*), preparation of immobilized microalgae beads (preparation of *Chlorella* suspension and beads preparation), analytical procedures and characterization studies (including cell growth and the number of viable cells, stability test, surface charge, morphology of the samples, surface area of the samples and Fourier transform infrared spectroscopy (FTIR)), and experimental design for immobilization studies. The second

section is related to biosorption studies, involving preparation of metal ion stock solution, the effect of different parameters on the biosorption process, desorption and reusability, biosorption capacity, kinetic and equilibrium models, and analytical procedures and characterization studies (including determination of metal ion concentration, water content, SEM-EDX, and Fourier transform infrared spectroscopy).

Chapter 4 presents the results and discussion of the two sections mentioned in chapter 3. Section I covers characterization of the beads (including zeta potential, surface morphology, surface area and FTIR spectral analysis), statistical analysis (analysis of variance (ANOVA) and optimum conditions) and evaluation of biocompatibility of zeolite 13X. Section II describes effects of contact time, initial pH and initial metal ion concentration on metal ion biosorption. Also, biosorption-desorption cycles, biosorption kinetic models (involving reaction (pseudo-first and pseudo-second order) and diffusion based models) and biosorption equilibrium models (Langmuir and Freundlich models), and characterization of the biosorbents (including water content, SEM-EDX and FTIR spectral analyses) were presented.

Chapter 5 refers to general conclusions based on the findings achieved in the results and discussion. Also, recommendations for future studies were presented in this chapter.

REFERENCES

- Abdel-Raouf, N., Al-Homaidan, A. A., & Ibraheem, I. B. M. (2012). Microalgae and wastewater treatment. Saudi Journal of Biological Sciences, 19(3), 257–275.
- Abdel -Aty, A. M., Ammar, N. S., Abdel Ghafar, H. H., & Ali, R. K. (2013). Biosorption of cadmium and lead from aqueous solution by fresh water alga Anabaena sphaerica biomass. *Journal of Advanced Research*, 4(4), 367–374.
- Abdel Hameed, M. S., & Ebrahim, O. H. (2007). Biotechnological Potential Uses of Immobilized Algae. International Journal of Agriculture & Biology, 9(1), 183– 192.
- Abe, K., Matsumura, I., Imamaki, A., & Hirano, M. (2003). Removal of inorganic nitrogen sources from water by the algal biofilm of the aerial microalga Trentepohlia aurea. *World Journal of Microbiology and Biotechnology*, 19(3), 325–328.
- Ahluwalia, S. S., & Goyal, D. (2007). Microbial and plant derived biomass for removal of heavy metals from wastewater. *Bioresource Technology*, 98(12), 2243–2257.
- Ahmad, A., Bhat, A. H., & Buang, A. (2018). Biosorption of Transition metals by freely suspended and Ca-alginate immobilised with Chlorella vulgaris: Kinetic and Equilibrium Modeling. *Journal of Cleaner Production*, 171, 1361–1375.
- Ahmad, A., Shah, S. M. U., Othman, M. F., & Abdullah, M. A. (2014). Enhanced palm oil mill effluent treatment and biomethane production by co-digestion of oil palm empty fruit bunches with Chlorella Sp. *The Canadian Journal of Chemical Engineering*, 92(9), 1636–1642.
- Akhtar, N, Iqbal, J., & Iqbal, M. (2003). Microalgal-luffa sponge immobilized disc: a new efficient biosorbent for the removal of Ni (II) from aqueous solution. *The Society for Applied Microbiology, Letters in Applied Microbiology*, 37, 149–153.
- Akhtar, N, Iqbal, J., & Iqbal, M. (2004). Removal and recovery of nickel (II) from aqueous solution by loofa sponge-immobilized biomass of Chlorella sorokiniana: characterization studies. *Journal of Hazardous Materials*, *108*, 85–94.
- Akhtar, Nasreen, Saeed, A., & Iqbal, M. (2003). Chlorella sorokiniana immobilized on the biomatrix of vegetable sponge of Luffa cylindrica: a new system to remove cadmium from contaminated aqueous medium. *Bioresource Technology*, 88, 163–165.
- Aksu, Z., Gönen, F., & Demircan, Z. (2002). Biosorption of chromium(VI) ions by Mowital®B30H resin immobilized activated sludge in a packed bed: Comparison with granular activated carbon. *Process Biochemistry*, *38*(2), 175–186.
- Al-Hassan, Z., Ivanova, V., Dobreva, E., Penchev, I., Hristov, J., Rachev, R., & Petrov, R. (1991). Non-porous magnetic supports for cell immobilization. *Journal of Fermentation and Bioengineering*, 71(2), 114–117.

- Al-Rub, F. A. A., El-Naas, M. H., Ashour, I., & Al-Marzouqi, M. (2006). Biosorption of copper on Chlorella vulgaris from single, binary and ternary metal aqueous solutions. *Process Biochemistry*, 41(2), 457–464.
- Al-Rub, F. A. A., El-Naas, M. H., Benyahia, F., & Ashour, I. (2004). Biosorption of nickel on blank alginate beads, free and immobilized algal cells. *Process Biochemistry*, 39, 1767–1773.
- Alpat, S. K., Özbayrak, Ö., Alpat, Ş., & Akçay, H. (2008). The adsorption kinetics and removal of cationic dye, Toluidine Blue O, from aqueous solution with Turkish zeolite. *Journal of Hazardous Materials*, 151(1), 213–220.
- Anastopoulos, I., & Kyzas, G. Z. (2015). Progress in batch biosorption of heavy metals onto algae. *Journal of Molecular Liquids*, 209, 77–86.
- Australian National Algae Culture Collection. (2017). Retrieved November 14, 2017, from https://www.csiro.au/en/Research/Collections/ANACC
- Aziz, M. A., & Ng, W. J. (1992). Feasibility of wastewater treatment using the activatedalgae process. *Bioresource Technology*, 40(3), 205–208.
- Babel, S., & Kurniawan, T. A. (2003). Low-cost adsorbents for heavy metals uptake from contaminated water: a review. *Journal of Hazardous Materials*, 97, 219–243.
- Badel, S., Bernardi, T., & Michaud, P. (2011). New perspectives for Lactobacilli exopolysaccharides. *Biotechnology Advances*, 29(1), 54–66.
- Bailey, S. E., Olin, T. J., Bricka, R. M., & Adrian, D. D. (1999). A review of potentially low-cost sorbents for heavy metals. *Water Research*, 33(11), 2469–2479.
- Barquist, K., & Larsen, S. C. (2010). Chromate adsorption on bifunctional, magnetic zeolite composites. *Microporous and Mesoporous Materials*, 130(1-3), 197-202.
- Bayramoglu, G., & Arica, M. Y. (2009). Construction a hybrid biosorbent using Scenedesmus quadricauda and Ca-alginate for biosorption of Cu(II), Zn(II) and Ni(II): kinetics and equilibrium studies. *Bioresource Technology*, 100, 186–193.
- Bayramoğlu, G., Tuzun, I., Celik, G., Yilmaz, M., & Arica, M. Y. (2006). Biosorption of mercury(II), cadmium(II) and lead(II) ions from aqueous system by microalgae Chlamydomonas reinhardtii immobilized in alginate beads. *International Journal* of Mineral Processing, 81(1), 35–43.
- Bekiroğullari, M., Kaya, M., & Saka, C. (2019). Highly efficient Co-B catalysts with Chlorella Vulgaris microalgal strain modified using hydrochloric acid as a new support material for hydrogen production from methanolysis of sodium borohydride. *International Journal of Hydrogen Energy*, 44(14), 7262–7275.
- Blanchard, G., Maunaye, M., & Martin, G. (1984). Removal of heavy metals from waters by means of natural zeolites. *Water Research*, 18(12), 1501–1507.

- Bolch, C. J. S., & Blackburn, S. I. (1996). Isolation and purification of Australian isolates of the toxic cyanobacterium Microcystis aeruginosa Kütz. *Journal of Applied Phycology*, 8(1), 5–13.
- Bulgariu, D., & Bulgariu, L. (2012). Equilibrium and kinetics studies of heavy metal ions biosorption on green algae waste biomass. *Bioresource Technology*, 103(1), 489– 493.
- Cai, T., Park, S. Y., & Li, Y. (2013). Nutrient recovery from wastewater streams by microalgae: Status and prospects. *Renewable and Sustainable Energy Reviews*, 19, 360–369.
- Cassidy, M. B., Lee, H., & Trevors, J. T. (1996). Environmental applications of immobilized microbial cells: A review. *Journal of Industrial Microbiology*, 16(2), 79–101.
- Chan, A., Salsali, H., & McBean, E. (2014). Heavy Metal Removal (Copper and Zinc) in Secondary Effluent from Wastewater Treatment Plants by Microalgae. ACS Sustainable Chemistry & Engineering, 2(2), 130–137.
- Chen, Z., Ma, W., & Han, M. (2008). Biosorption of nickel and copper onto treated alga (Undaria pinnatifida): Application of isotherm and kinetic models. *Journal of Hazardous Materials*, 155(1-2), 327-333.
- Corona-González, R. I., Miramontes-Murillo, R., Arriola-Guevara, E., Guatemala-Morales, G., Toriz, G., & Pelayo-Ortiz, C. (2014). Immobilization of Actinobacillus succinogenes by adhesion or entrapment for the production of succinic acid. *Bioresource Technology*, 164, 113–118.
- da Costa, A. C. A., & Leite, S. G. F. (1991). Metals biosorption by sodium alginate immobilized chlorella homosphaera cells. *Biotechnology Letters*, 13(8), 559–562.
- Darnall, D. W., Greene, B., Henzl, M. T., Hosea, J. M., McPherson, R. A., Sneddon, J., & Alexander, M. D. (1986). Selective recovery of gold and other metal ions from an algal biomass. *Environmental Science & Technology*, 20(2), 206–208.
- Das, M., & Adholeya, A. (2015). Potential uses of immobilized bacteria, fungi, algae, and their aggregates for treatment of organic and inorganic pollutants in wastewater. In ACS Symposium Series (pp. 319–337).
- De-Bashan, L. E., & Bashan, Y. (2010). Immobilized microalgae for removing pollutants: Review of practical aspects. *Bioresource Technology*, 101(6), 1611– 1627.
- Desmet, J., Meunier, C., Danloy, E., Duprez, M. E., Lox, F., Thomas, D., ... Su, B. L. (2015). Highly efficient, long life, reusable and robust photosynthetic hybrid coreshell beads for the sustainable production of high value compounds. *Journal of Colloid and Interface Science*, 448, 79–87.
- Djukić-Vuković, A. P., Jokić, B. M., Kocić-Tanackov, S. D., Pejin, J. D., & Mojović, L. V. (2016). Mg-modified zeolite as a carrier for Lactobacillus rhamnosus in L(+)

lactic acid production on distillery wastewater. *Journal of the Taiwan Institute of Chemical Engineers*, 59, 262–266.

- Djukić-Vuković, A. P., Mojović, L. V., Jokić, B. M., Nikolić, S. B., & Pejin, J. D. (2013). Lactic acid production on liquid distillery stillage by Lactobacillus rhamnosus immobilized onto zeolite. *Bioresource Technology*, 135, 454–458.
- Doula, M. K. (2006). Removal of Mn2+ ions from drinking water by using Clinoptilolite and a Clinoptilolite-Fe oxide system. *Water Research*, 40, 3167–3176.
- Durham, D. R., Marshall, L. C., Miller, J. G., & Chmurny, A. B. (1994). New composite biocarriers engineered to contain adsorptive and ion- exchange properties improve immobilized-cell bioreactor process dependability. *Applied and Environmental Microbiology*, 60(11), 4178–4181.
- Duygu, D. (Yalcin), Udoh, A. U., Ozer, T. (Baykal), Akbulut, A., Erkaya, Ii. (Acikoz), Yildiz, K., & Guler, D. (2012). Fourier transform infrared (FTIR) spectroscopy for identification of Chlorella vulgaris Beijerinck 1890 and Scenedesmus obliquus (Turpin) Kützing 1833. Afr. J. Biotechnol, 11(16), 3817–3824.
- Dwivedi, S. (2012). Bioremediation of heavy metal by algae: current and future perspective. *Journal of Advance Laboratory Research in Biology*, *III*(III), 195–199.
- Erdem, E., Karapinar, N., & Donat, R. (2004). The removal of heavy metal cations by natural zeolites. *Journal of Colloid and Interface Science*, 280, 309–314.
- Eroglu, E., Agarwal, V., Bradshaw, M., Chen, X., Smith, S. M., Raston, C. L., & Swaminathan Iyer, K. (2012). Nitrate removal from liquid effluents using microalgae immobilized on chitosan nanofiber mats. *Green Chemistry*, 14(10), 2682–2685.
- Eroglu, E., Smith, S. M., & Raston, C. L. (2015). Application of various immobilization techniques for algal bioprocesses. In N. R. Moheimani, M. P. McHenry, K. de Boer, & P. A. Bahri (Eds.), *Biomass and Biofuels from Microalgae* (Vol. 2, pp. 19–44). Springer.
- Fernández, N., Montalvo, S., Fernández-Polanco, F., Guerrero, L., Cortés, I., Borja, R., ... Travieso, L. (2007). Real evidence about zeolite as microorganisms immobilizer in anaerobic fluidized bed reactors. *Process Biochemistry*, 42(4), 721–728.
- Fierro, S., del Pilar Sánchez-Saavedra, M., & Copalcúa, C. (2008). Nitrate and phosphate removal by chitosan immobilized Scenedesmus. *Bioresource Technology*, 99(5), 1274–1279.
- Figueroa-Torres, G. M., Certucha-Barragan, M. T., Acedo-Felix, E., Monge-Amaya, O., Almendariz-Tapia, F. J., & Gasca-Estefania, L. A. (2016). Kinetic studies of heavy metals biosorption by acidogenic biomass immobilized in clinoptilolite. *Journal of the Taiwan Institute of Chemical Engineers*, 61, 241–246.

- Foo, K. Y., & Hameed, B. H. (2010). Insights into the modeling of adsorption isotherm systems. *Chemical Engineering Journal*, 156, 2–10.
- Gekeler, W., Grill, E., Winnacker, E. L., & Zenk, M. H. (1988). Algae sequester heavy metals via synthesis of phytochelatin complexes. *Archives of Microbiology*, 150(2), 197–202.
- Godlewska-Żyłkiewicz, B. (2003). Biosorption of platinum and palladium for their separation/preconcentration prior to graphite furnace atomic absorption spectrometric determination. *Spectrochimica Acta Part B*, 58(8), 1531–1540.
- Hanim, S. A. M., Malek, N. A. N. N., & Ibrahim, Z. (2016). Amine-functionalized, silver-exchanged zeolite NaY: preparation, characterization and antibacterial activity. *Applied Surface Science*, 360, 121–130.
- He, J., & Chen, J. P. (2014). A comprehensive review on biosorption of heavy metals by algal biomass: Materials, performances, chemistry, and modeling simulation tools. *Bioresource Technology*, 160, 67–78.
- Hedström, A., & Rastas Amofah, L. (2008). Adsorption and desorption of ammonium by clinoptilolite adsorbent in municipal wastewater treatment systems. *Journal of Environmental Engineering and Science*, 7(1), 53–61.
- Hernandez, J. P., De-Bashan, L. E., & Bashan, Y. (2006). Starvation enhances phosphorus removal from wastewater by the microalga Chlorella spp. coimmobilized with Azospirillum brasilense. *Enzyme and Microbial Technology*, 38(1–2), 190–198.
- Hrenovic, J., Ivankovic, T., & Tibljas, D. (2009). The effect of mineral carrier composition on phosphate-accumulating bacteria immobilization. *Journal of Hazardous Materials*, 166(2–3), 1377–1382.
- Hrenovic, J., Kovacevic, D., Ivankovic, T., & Tibljas, D. (2011). Selective immobilization of Acinetobacter junii on the natural zeolitized tuff in municipal wastewater. *Colloids and Surfaces B: Biointerfaces*, 88(1), 208–214.
- Hrenovic, J., Rozic, M., Sekovanic, L., & Anic-Vucinic, A. (2008). Interaction of surfactant-modified zeolites and phosphate accumulating bacteria. *Journal of Hazardous Materials*, 156(1–3), 576–582.
- Idris, A., & Suzana, W. (2006). Effect of sodium alginate concentration, bead diameter, initial pH and temperature on lactic acid production from pineapple waste using immobilized Lactobacillus delbrueckii. *Process Biochemistry*, 41(5), 1117–1123.
- Inglezakis, V. J., Loizidou, M. D., & Grigoropoulou, H. P. (2003). Ion exchange of Pb2+, Cu2+, Fe3+, and Cr3+ on natural clinoptilolite: Selectivity determination and influence of acidity on metal uptake. *Journal of Colloid and Interface Science*, 261, 49–54.
- Jami, M. S., Rosli, N., & Amosa, M. K. (2016). Optimization of manganese reduction in biotreated POME onto 3A molecular sieve and clinoptilolite zeolites. *Water*

Environment Research, 88(16), 566–576.

- Jiménez-Pérez, M. V., Sánchez-Castillo, P., Romera, O., Fernández-Moreno, D., & Pérez-Martinez, C. (2004). Growth and nutrient removal in free and immobilized planktonic green algae isolated from pig manure. *Enzyme and Microbial Technology*, 34(5), 392–398.
- Kaewsarn, P. (2002). Biosorption of copper(II) from aqueous solutions by pre-treated biomass of marine algae Padina sp. *Chemosphere*, 47(10), 1081–1085.
- Kamyab, H., Md Din, M. F., Tin, C. L., Ponraj, M., Soltani, M., Mohamad, S. E., & Roudi, A. M. (2014). Micro-macro algal mixture as a promising agent for treating POME discharge and its potential use as animal feed stock enhancer. *Jurnal Teknologi (Sciences and Engineering)*, 68(5), 1–4.
- Kaparapu, J., Narasimha, M., & Geddada, R. (2016). Applications of immobilized algae. *Journal of Algal Biomass Utilization*, 7(2), 122–128.
- Kaplan, D. (2013). Absorption and Adsorption of Heavy Metals by Microalgae. In A. Richmond & Q. Hu. (Eds.), *Handbook of Microalgal Culture: Applied Phycology* and Biotechnology (Second Edi, pp. 602–611).
- Karthikeyan, S., Balasubramanian, R., & Iyer, C. S. P. (2007). Evaluation of the marine algae Ulva fasciata and Sargassum sp. for the biosorption of Cu(II) from aqueous solutions. *Bioresource Technology*, 98(2), 452–455.
- Kexun, L., Shun, L., & Xianhua, L. (2014). An overview of algae bioethanol production. International Journal of Energy Research, 38, 965–977.
- Kondo, K., Hirayama, K., & Matsumoto, M. (2013). Adsorption of metal ions from aqueous solution onto microalga entrapped into Ca-alginate gel bead. *Desalination* and Water Treatment, 51, 4675–4683.
- Kosobucki, P., Kruk, M., & Buszewski, B. (2008). Immobilization of selected heavy metals in sewage sludge by natural zeolites. *Bioresource Technology*, *99*, 5972–5976.
- Krishnani, K. K., Meng, X., Christodoulatos, C., & Boddu, V. M. (2008). Biosorption mechanism of nine different heavy metals onto biomatrix from rice husk. *Journal* of Hazardous Materials, 153(3), 1222–1234.
- Kubota, M., Nakabayashi, T., Matsumoto, Y., Shiomi, T., Yamada, Y., Ino, K., ... Sakaguchi, K. (2008). Selective adsorption of bacterial cells onto zeolites. *Colloids* and Surfaces B: Biointerfaces, 64(1), 88–97.
- Kwakye, G. F., McMinimy, R. A., & Aschner, M. (2016). Disease-Toxicant Interactions in Parkinson's Disease Neuropathology. *Neurochemical Research*, 1–15.
- Kyzioł, A., Mazgała, A., Michna, J., Regiel-Futyra, A., & Sebastian, V. (2017). Preparation and characterization of alginate/chitosan formulations for ciprofloxacin-controlled delivery. *Journal of Biomaterials Applications*.

- Lameiras, S., Quintelas, C., & Tavares, T. (2008). Biosorption of Cr (VI) using a bacterial biofilm supported on granular activated carbon and on zeolite. *Bioresource Technology*, 99(4), 801–806.
- Leenen, E. J. T. M., Dos Santos, V. A. P., Grolle, K. C. F., Tramper, J., & Wijffels, R. H. (1996). Characteristics of and selection criteria for support materials for cell immobilization in wastewater treatment. *Water Research*, 30(12), 2985–2996.
- Li, Y., Horsman, M., Wu, N., Lan, C. Q., & Dubois-calero, N. (2008). Biofuels from Microalgae. *Biotechnology Progress*, 24, 815–820.
- Liu, H., Peng, S., Shu, L., Chen, T., Bao, T., & Frost, R. L. (2013a). Effect of Fe 3O 4 addition on removal of ammonium by zeolite NaA. *Journal of Colloid and Interface Science*, *390*(1), 204–210.
- Liu, H., Peng, S., Shu, L., Chen, T., Bao, T., & Frost, R. L. (2013b). Magnetic zeolite NaA: Synthesis, characterization based on metakaolin and its application for the removal of Cu2+, Pb2+. *Chemosphere*, 91(11), 1539–1546.
- Liu, Y., Rafailovich, M. H., Malal, R., Cohn, D., & Chidambaram, D. (2009). Engineering of bio-hybrid materials by electrospinning polymer-microbe fibers. *Proceedings of the National Academy of Sciences of the United States of America*, 106(34), 14201–14206.
- López, A., Lázaro, N., & Marqués, A. M. (1997). The interphase technique: A simple method of cell immobilization in gel-beads. *Journal of Microbiological Methods*, 30(3), 231–234.
- MacArio, A., Giordano, G., Setti, L., Parise, A., Campelo, J. M., Marinas, J. M., & Luna, D. (2007). Study of lipase immobilization on zeolitic support and transesterification reaction in a solvent free-system. *Biocatal. Biotransform.*, 25(2), 328–335.
- Madadi, M., & Rahimi, R. (2012). Zeolite-immobilized Mn(III), Fe(III) and Co(III) complexes with 5,10,15,20-tetra(4-methoxyphenyl)porphyrin as heterogeneous catalysts for the epoxidation of (R)-(+)-limonene: Synthesis, characterization and catalytic activity. *Reaction Kinetics, Mechanisms and Catalysis*, 107(1), 215–229.
- Malik, N. (2002). Biotechnological potential of immobilised algae for wastewater N, P and metal removal: a review. *BioMetals*, *15*, 377–390.
- Mallick, N., & Rai, L. C. (1993). Influence of culture density, pH, organic acids and divalent cations on the removal of nutrients and metals by immobilized Anabaena doliolum and Chlorella vulgaris. *World Journal of Microbiology and Biotechnology*, *9*, 196–201.
- Mallick, N., & Rai, L. C. (1994). Removal of inorganic ions from wastewaters by immobilized microalgae. World Journal of Microbiology & Biotechnology, 10(4), 439–443.

- Margeta, K., Logar, N. Z., Šiljeg, M., & Farkaš, A. (2013). Natural Zeolites in Water Treatment – How Effective is Their Use. In W. Elshorbagy (Ed.), *Water Treatment*. InTech.
- Mehta, S. K., & Gaur, J. P. (2005). Use of algae for removing heavy metal ions from wastewater: progress and prospects. *Critical Reviews in Biotechnology*, 25(3), 113–152.
- Mery, C., Guerrero, L., Alonso-Gutiérrez, J., Figueroa, M., Lema, J. M., Montalvo, S., & Borja, R. (2012). Evaluation of natural zeolite as microorganism support medium in nitrifying batch reactors: influence of zeolite particle size. J. Environ. Sci. Health A Tox. Hazard. Subst. Environ. Eng, 47(November 2014), 420–427.
- Milán, Z., Villa, P., Sánchez, E., Montalvo, S., Borja, R., Ilangovan, K., & Briones, R. (2003). Effect of natural and modified zeolite addition on anaerobic digestion of piggery waste. *Water Science and Technology*, 48(6), 263–269.
- Misaelides, P. (2011). Application of natural zeolites in environmental remediation: A short review. *Microporous and Mesoporous Materials*, 144(1-3), 15–18.
- Molecular Sieves Technical Information Bulletin | Sigma-Aldrich. (2017). Retrieved November 14, 2017, from http://www.sigmaaldrich.com/chemistry/chemicalsynthesis/learning-center/technical-bulletins/al-1430/molecular-sieves.html
- Monge-Amaya, O., Valenzuela-García, J. L., Félix, E. A., Certucha-Barragán, M. T., Leal-Cruz, A. L., & Almendariz-Tapia, F. J. (2013). Biosorptive Behavior of Aerobic Biomass Biofilm Supported on Clinoptilolite Zeolite for the Removal of Copper. *Mineral Processing and Extractive Metallurgy Review*, 34(6), 422–428.
- Moreno-garrido, I., Blasco, J., González-delvalle, M. A., & Lubián, L. M. (1998). Differences in copper accumulation by the marine microalga Nannochloropsis gaditana Lubián, submitted to two different thermal treatments. *Ecotoxicol Environ Restor*, 1(1), 43–47.
- Moreno-Garrido, I., Campana, O., Lubián, L. M., & Blasco, J. (2005). Calcium alginate immobilized marine microalgae: Experiments on growth and short-term heavy metal accumulation. In *Marine Pollution Bulletin* (Vol. 51, pp. 823–829).
- Moreno-Garrido, Ignacio. (2008). Microalgae immobilization: Current techniques and uses. *Bioresource Technology*, 99(10), 3949–3964.
- Motsi, T., Rowson, N. A., & Simmons, M. J. H. (2009). Adsorption of heavy metals from acid mine drainage by natural zeolite. *International Journal of Mineral Processing*, 92, 42–48.
- Mthombeni, N. H., Onyango, M. S., & Aoyi, O. (2015). Adsorption of hexavalent chromium onto magnetic natural zeolite-polymer composite. *Journal of the Taiwan Institute of Chemical Engineers*, 50, 242–251.
- Mujtaba, G., Rizwan, M., & Lee, K. (2017). Removal of nutrients and COD from wastewater using symbiotic co-culture of bacterium Pseudomonas putida and

immobilized microalga Chlorella vulgaris. *Journal of Industrial and Engineering Chemistry*, 49, 145–151.

- Mulbry, W., Westhead, E. K., Pizarro, C., & Sikora, L. (2005). Recycling of manure nutrients: Use of algal biomass from dairy manure treatment as a slow release fertilizer. *Bioresource Technology*, 96(4), 451–458.
- Nakajima, A., Horikoshi, T., & Sakaguchi, T. (1982). Recovery of Uranium by Immobilized Microorganisms. *European Journal of Applied Microbilogy and Biotechnology*, 16, 88–91.
- Nasreen, A., Muhammad, I., Iqbal, Z. S., & Javed, I. (2008). Biosorption characteristics of unicellular green alga Chlorella sorokiniana immobilized in loofa sponge for removal of Cr(III). *Journal of Environmental Sciences*, 20, 231–239.
- Nastaj, J., Przewłocka, A., & Rajkowska-Myśliwiec, M. (2016). Biosorption of Ni (II), Pb (II) and Zn (II) on calcium alginate beads: equilibrium, kinetic and mechanism studies. *Pol. J. Chem. Technol*, 18(3), 81–87.
- Oliveira, L. C. A., Petkowicz, D. I., Smaniotto, A., & Pergher, S. B. C. (2004). Magnetic zeolites: A new adsorbent for removal of metallic contaminants from water. *Water Research*, *38*(17), 3699–3704.
- Onyancha, D., Mavura, W., Ngila, J. C., Ongoma, P., & Chacha, J. (2008). Studies of chromium removal from tannery wastewaters by algae biosorbents, Spirogyra condensata and Rhizoclonium hieroglyphicum. *Journal of Hazardous Materials*, 158, 605–614.
- Ouki, S. K., & Kavannagh, M. (1999). Treatment of metals-contaminated wastewaters by use of natural zeolites. *Water Science and Technology*, *39*(10–11), 115–122.
- Pannier, A., Soltmann, U., Soltmann, B., Altenburger, R., & Schmitt-Jansen, M. (2014). Alginate/silica hybrid materials for immobilization of green microalgae Chlorella vulgaris for cell-based sensor arrays. J. Mater. Chem. B, 2, 7896–7909.
- Pazos, M., Branco, M., Neves, I. C., Sanromán, M. A., & Tavares, T. (2010). Removal of Cr(VI) from aqueous solutions by a bacterial biofilm supported on zeolite: optimisation of the operational conditions and scale-up of the bioreactor. *Chemical Engineering and Technology*, *33*(12), 2008–2014.
- Petrovič, A., & Simonič, M. (2016). Removal of heavy metal ions from drinking water by alginate-immobilised Chlorella sorokiniana. *International Journal of Environmental Science and Technology*, 13, 1761–1780.
- Phang, S.-M., & Ong, K.-C. (1988). Algal Biomass Production in Digested Palm Oil Mill Effluent. *Biological Wastes*, 25, 177–191.
- Pitcher, S. K., Slade, R. C. T., & Ward, N. I. (2004). Heavy metal removal from motorway stormwater using zeolites. *Science of the Total Environment*, 334–335, 161–166.

- Quintelas, C., Rocha, Z., Silva, B., Fonseca, B., Figueiredo, H., & Tavares, T. (2009). Biosorptive performance of an Escherichia coli biofilm supported on zeolite NaY for the removal of Cr(VI), Cd(II), Fe(III) and Ni(II). *Chemical Engineering Journal*, 152(1), 110–115.
- Rai, L. C., & Mallick, N. (1992). Removal and assessment of toxicity of Cu and Fe to Anabaena doliolum and Chlorella vulgaris using free and immobilized cells. *World Journal of Microbiology and Biotechnology*, 8(2), 110–114.
- Romera, E., González, F., Ballester, A., Blázquez, M. L., & Muñoz, J. A. (2007). Comparative study of biosorption of heavy metals using different types of algae. *Bioresource Technology*, 98, 3344–3353.
- Ruiz-Marin, A., Mendoza-Espinosa, L. G., & Stephenson, T. (2010). Growth and nutrient removal in free and immobilized green algae in batch and semi-continuous cultures treating real wastewater. *Bioresource Technology*, *101*(1), 58–64.
- Safak Boroglu, M., & Ali Gurkaynak, M. (2011). Fabrication and characterization of silica modified polyimide-zeolite mixed matrix membranes for gas separation properties. *Polymer Bulletin*, 66, 463–478.
- Safonova, B. E., Kvitko, K. V, Iankevitch, M. I., Surgko, L. F., Afti, I. A., & Reisser, W. (2004). Biotreatment of Industrial Wastewater by Selected Algal-Bacterial Consortia. *Engineering in Life Sciences*, 4(4), 347–353.
- Sakaguchi, K., Matsui, M., & Mizukami, F. (2005). Applications of zeolite inorganic composites in biotechnology: current state and perspectives. *Applied Microbiology* and Biotechnology, 67(3), 306–311.
- Saleh, T. A. (2015). Isotherm, kinetic, and thermodynamic studies on Hg(II) adsorption from aqueous solution by silica- multiwall carbon nanotubes. *Environmental Science and Pollution Research*.
- Sari, A., & Tuzen, M. (2008). Biosorption of Pb(II) and Cd(II) from aqueous solution using green alga (Ulva lactuca) biomass. *Journal of Hazardous Materials*, 152(1), 302–308.
- Shavandi, M. A., Haddadian, Z., Ismail, M. H. S., Abdullah, N., & Abidin, Z. Z. (2012). Removal of Fe(III), Mn(II) and Zn(II) from palm oil mill effluent (POME) by natural zeolite. *Journal of the Taiwan Institute of Chemical Engineers*, 43, 750– 759.
- Shavandi, Mohammad Amin, Haddadian, Z., Ismail, M. H. S., & Abdullah, N. (2012). Continuous metal and residual oil removal from palm oil mill effluent using natural zeolite-packed column. *Journal of the Taiwan Institute of Chemical Engineers*, 43(6), 934–941.
- Sheehan, J., Dunahay, T., Benemann, J., & Roessler, P. (1998). A look back at the U.S. Department of Energy's aquatic species program: biodiesel from algae. Golden, Colorado.

- Shen, Y., Gao, J., & Li, L. (2017). Municipal wastewater treatment via co-immobilized microalgal-bacterial symbiosis: microorganism growth and nutrients removal. *Bioresource Technology*, 243, 905–913.
- Shi, J., Podola, B., & Melkonian, M. (2007). Removal of nitrogen and phosphorus from wastewater using microalgae immobilized on twin layers: An experimental study. *Journal of Applied Phycology*, 19(5), 417–423.
- Shindo, S., Takata, S., Taguchi, H., & Yoshimura, N. (2001). Development of novel carrier using natural zeolite and continuous ethanol fermentation with immobilized Saccharomyces cerevisiae in a bioreactor. *Biotechnology Letters*, 23(24), 2001–2004.
- Smidsrod, O., & Skjak-Braek, G. (1990). Alginate as immobilization matrix for cells. *Trends Biotechnol*, 8, 71–78.
- Sugawara, T., Matsuura, Y., Anzai, T., & Miura, O. (2016). Removal of Ammonia Nitrogen From Water by Magnetic Separation. *IEEE Transactions on Applied Superconductivity*, 26(4).
- Svilović, S., Rušić, D., & Bašić, A. (2010). Investigations of different kinetic models of copper ions sorption on zeolite 13X. *Desalination*, 259(1–3), 71–75.
- Sydney, E. B., da Silva, T. E., Tokarski, A., Novak, A. C., de Carvalho, J. C., Woiciecohwski, A. L., ... Soccol, C. R. (2011). Screening of microalgae with potential for biodiesel production and nutrient removal from treated domestic sewage. *Applied Energy*, 88(10), 3291–3294.
- Tam, N. F. Y., & Wong, Y. S. (2000). Effect of immobilized microalgal bead concentrations on wastewater nutrient removal. *Environmental Pollution*, 107(1), 145–151.
- Thomas, W. J., & Crittenden, B. D. (1998). *Adsorption Technology and Design*. Oxford, Boston: Butterworth-Heinemann.
- Travieso, L., Cañizares, R. O., Borja, R., Benítez, F., Domínguez, A. R., Dupeyron, R., & Valiente, y V. (1999). Heavy Metal Removal by Microalgae. *Bull. Environ. Contam. Toxicol.*, 62, 144–151.
- Vaca Mier, M., Lopez Callejas, R., Gehr, R., Jimenez Cisneros, B. E., & Alvarez, P. J. J. (2001). Heavy metal removal with mexican clinoptilolite: Multi-component ionic exchange. *Water Research*, 35(2), 373–378.
- Vasilieva, S. G., Lobakova, E. S., Lukyanov, A. A., & Solovchenko, A. E. (2016). Immobilized Microalgae in Biotechnology. *Ecology*, *71*(3), 170–176.
- Wan Maznah, W. O., Al-Fawwaz, A. T., & Surif, M. (2012). Biosorption of copper and zinc by immobilised and free algal biomass, and the effects of metal biosorption on the growth and cellular structure of Chlorella sp. and Chlamydomonas sp. isolated from rivers in Penang, Malaysia. *Journal of Environmental Sciences*, 24(8), 1386–1393.

- Wang, J., & Chen, C. (2009). Biosorbents for heavy metals removal and their future. *Biotechnology Advances*, 27(2), 195–226.
- Wang, S., & Peng, Y. (2010). Natural zeolites as effective adsorbents in water and wastewater treatment. *Chemical Engineering Journal*, 156(1), 11–24.
- Weiß, S., Lebuhn, M., Andrade, D., Zankel, A., Cardinale, M., Birner-Gruenberger, R., ... Guebitz, G. M. (2013). Activated zeolite - Suitable carriers for microorganisms in anaerobic digestion processes? *Appl Microbiol Biotechnol*, 97(7), 3225–3238.
- Weiß, S., Tauber, M., Somitsch, W., Meincke, R., Müller, H., Berg, G., & Guebitz, G. M. (2010). Enhancement of biogas production by addition of hemicellulolytic bacteria immobilised on activated zeolite. *Water Research*, 44(6), 1970–1980.
- Wilke, A., Buchholz, R., & Bunke, G. (2006). Selective biosorption of heavy metals by algae. *Environmental Biotechnology*, 2, 47–56.
- Wilkinson, S. C., Goulding, K. H., & Robinson, P. K. (1990). Mercury removal by immobilized algae in batch culture systems. *Journal of Applied Phycology*, 2, 223– 230.
- Wingenfelder, U., Hansen, C., Furrer, G., & Schulin, R. (2005). Removal of heavy metals from mine waters by natural zeolites. *Environmental Science & Technology*, 39, 4606–4613.
- Wu, Y., Li, T., & Yang, L. (2012). Mechanisms of removing pollutants from aqueous solutions by microorganisms and their aggregates: A review. *Bioresource Technology*, 107, 10–18.
- Yuna, Z. (2016). Review of the natural, modified, and synthetic zeolites for heavy metals removal from wastewater. *Environmental Engineering Science*.
- Zamzow, M. J., Eichbaum, B. R., Sandgren, K. R., & Shanks, D. E. (1990). Removal of Heavy Metals and Other Cations from Wastewater Using Zeolites. *Separation Science and Technology*, 25(13–15), 1555–1569.
- Zeng, X., Danquah, M. K., Chen, X. D., & Lu, Y. (2011). Microalgae bioengineering: From CO2 fixation to biofuel production. *Renewable and Sustainable Energy Reviews*, 15(6), 3252–3260.
- Zeng, X., Danquah, M. K., Halim, R., Yang, S., Chen, X. D., & Lu, Y. (2013). Comparative physicochemical analysis of suspended and immobilized cultivation of Chlorella sp. J. Chem. Technol. Biotechnol, 88(2), 247–254.
- Zeng, X., Danquah, M. K., Potumarthi, R., Cao, J., Chen, X. D., & Lu, Y. (2013). Characterization of sodium cellulose sulphate/poly-dimethyl-diallyl-ammonium chloride biological capsules for immobilized cultivation of microalgae. *J Chem Technol Biotechnol*, 88(4), 599–605.

Zeng, X., Danquah, M. K., Zheng, C., Potumarthi, R., Chen, X. D., & Lu, Y. (2012).

NaCS-PDMDAAC immobilized autotrophic cultivation of Chlorella sp. for wastewater nitrogen and phosphate removal. *Chemical Engineering Journal*, *187*, 185–192.

- Zeraatkar, A. K., Ahmadzadeh, H., Talebi, A. F., Moheimani, N. R., & McHenry, M. P. (2016). Potential use of algae for heavy metal bioremediation, a critical review. *Journal of Environmental Management*, 181, 817–831.
- Zhang, B. B., Wang, L., Charles, V., Rooke, J. C., & Su, B. L. (2016). Robust and biocompatible hybrid matrix with controllable permeability for microalgae encapsulation. *ACS Appl. Mater. Interfaces*, 8(14), 8939–8946.
- Zhao, M., Xu, Y., Zhang, C., Rong, H., & Zeng, G. (2016). New trends in removing heavy metals from wastewater. *Applied Microbiology and Biotechnology*.
- Zhou, L., Li, G., An, T., Fu, J., & Sheng, G. (2008). Recent Patents on Immobilized Microorganism Technology and Its Engineering Application in Wastewater Treatment. *Recent Patents on Engineering*.

BIODATA OF STUDENT

Seyed Amirebrahim Emami Moghaddam was born on 6th July 1982 in Mashhad, Iran. He attended Islamic Azad University (IAU) and graduated with Bachelor's degree in Chemical Engineering - Food Technology in 2005. Then, he obtained Master's degree in Chemical Engineering - Biotechnology in Iran University of Science and Technology (IUST) in 2009. He joined Universiti Putra Malaysia (UPM) in 2015 to pursue PhD in Chemical Engineering.



LIST OF PUBLICATIONS

Publications

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2019). Kinetic and equilibrium modeling for the biosorption of metal ion by Zeolite 13X-Algal-Alginate Beads (ZABs), *Journal of Water Process Engineering*, Under Review.

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2019). Stability improvement of algal-alginate beads by zeolite molecular sieves 13X. *International Journal of Biological Macromolecules*, 132, 592–599.

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2019). Preliminary study on zeolite 13X as a potential carrier for algal immobilization. *Journal of Advanced Research in Materials Science*, 53(1), 1–5.

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2018). potential of zeolite and algae in biomass immobilization. *BioMed Research International*, 2018, 15.

Conferences

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2019). Stability improvement of algal-alginate beads by zeolite 13X: A preliminary research. The 9th International Conference on Bioscience, Biochemistry and Bioinformatics (ICBBB 2019), 7th – 9th Jan 2019, National University of Singapore (NUS), Singapore.

Emami Moghaddam, S. A., Harun, R., Mokhtar, M. N., & Zakaria, R. (2018). Preliminary study on zeolite 13X as a potential carrier for algal immobilization. The 5th International Conference on Biotechnology Engineering (ICBioE 2018), 19th – 20th Sep 2018, International Islamic University Malaysia (IIUM), Kuala Lumpur, Malaysia.



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