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Year: 2020

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DOI: https://doi.org/10.1016/j.hermed.2020.100337

Posted at the Zurich Open Repository and Archive, University of Zurich ZORA URL: https://doi.org/10.5167/uzh-196168 Journal Article Accepted Version



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Originally published at:

Gartenmann, Stefanie J; Steppacher, Severine L; von Weydlich, Yves; Heumann, Christian; Attin, Thomas; Schmidlin, Patrick R (2020). The Effect of Green Tea on plaque and gingival inflammation: A systematic review. Journal of Herbal Medicine, 21:100337. DOI: https://doi.org/10.1016/j.hermed.2020.100337

The Effect of Green Tea on plaque and gingival inflammation: A systematic review

Stefanie J. Gartenmann^a, Severine L. Steppacher^b, Yves von Weydlich^b, Christian Heumann^c, Thomas Attin^a, Patrick R. Schmidlin^a

^aClinic of Preventive Dentistry, Periodontology and Cariology, Center of Dental Medicine, University of Zurich, Zürich, Switzerland

^bPrivate practices in Switzerland, Switzerland

^cDepartment for Statistics, Ludwig-Maximilians-University Munich, Munich, Germany

Abstract

Green tea has been shown in individual studies to be effective in reducing plaque and against gingivitis. Therefore, the aim of this study was to to systematically review available literature on green tea catechin.

The systematic literature search was performed using electronic databases in CINAHL, Cochrane Library, MEDLINE, PubMed and Scopus until January 2017. The PRISMA criteria were applied and a research question was posed according to PICO: "In patients with gingivitis (population), what is the effect of green tea catechins-containing mouthwash (intervention and comparison) on plaque accumulation and gingival inflammation (outcome)?" Out of 187 titles identified by the search strategy, five were suitable for meta-analyses. These five studies were undertaken on a predominately Asian population. Plaque (PI) and Gingival Index (GI) were compared at endpoint and with respect to the change throughout the study (baseline-endpoint). The results from the meta-analysis indicated that green tea and chlorhexidine (CHX) resulted in lower PI compared to placebo while there was no significant difference between CHX and green tea, either at endpoint or over time. In addition, there was little evidence of side effects with green tea mouthwash.

Green tea mouthwash may be a viable alternative to CHX, especially for long-term use. However, due to the very heterogeneous data and the risk of bias, this evidence should be interpreted with caution. Further clinically controlled studies with a longer observation period are required.

1. Introduction

To prevent gingivitis, both mechanical plaque removal and chemical plaque control may be recommended. In particular, chemotherapeutics have the potential to inhibit plaque growth and reduce gingivitis (Supranoto et al., 2015). Due to a lack of compliance and manual dexterity a significant number of people do not achieve sufficient mechanical plaque removal and in turn benefit from adjunctive chemical usage (Axelsson and Lindhe, 1987; Baker, 1993; Christie et al., 1998).

Therapeutic mouthwashes contains active ingredients that help control bad breath, plaque and gingivitis. Some of the most common therapeutic ingredients in mouthwash products are essential oils (EOs), cetyl pyridinium chloride (CPC) and chlorhexidine (CHX) (Araujo et al., 2015; Van der Weijden et al., 2015).

The anti-inflammatory effect of EO is based on its antioxidant activity, which has been shown to reduce plaque and gingival inflammation through plaque penetration (Haas et al., 2016; Ouhayoun, 2003; Van der Weijden et al., 2015). However, caution should be exercised when using alcohol-based mouthwashes since their use may cause pain and/or burning and may affect the connective tissue in the oral cavity (Poggi et al., 2003).

The cationic quaternary ammonium compound of CPC interacts with the cell membrane of bacteria, interrupting cell metabolism and inhibiting cell growth, followed by cell death as a consequence (Van der Weijden et al., 2015).

Of the chemical plaque control agents, CHX is the gold standard for the prevention of dental plaque (Axelsson and Lindhe, 1987; Van der Weijden et al., 2015). Years of documented research have demonstrated that chlorhexidine digluconate may prevent and control plaque formation, thereby inhibiting and reducing the development of gingivitis (Gunsolley, 2010; Loe, 1967). However, there are reports of side effects such as discoloration of the teeth, restorations and the tongue; increased formation of supragingival calculus and impairment of taste sensation. Occasionally, mucous membrane irritation and desquamation of the oral tissues have also been associated with the use of CHX, especially with prolonged use (Van der Weijden et al., 2015).

Based on the reported side effects, alternative ingredients such as herbs or probiotics have gained in popularity and are the focus of attention in many research projects (Anand et al., 2015; Haffajee et al., 2008; Jockel-Schneider et al., 2016; Martin-Cabezas et al., 2016; Oliveira et al., 2017; Schlagenhauf et al., 2016). The search for alternative products and natural phytochemicals isolated from plants as used in herbal medicines is considered a good alternative to synthetic chemicals (Prabu et al., 2006). Especially for prolonged use and routine application, organic agents are desired as an alternative.

Among these natural phytochemicals, the health benefits of green teacatechins from the leaves of the plant *Camellia sinensis* have been shown to be beneficial in the treatment of a variety of diseases in Western medicine (Chacko et al., 2010; Khan and Mukhtar, 2007; McKay and Blumberg, 2002). The polyphenols in green tea have been found to contain bioactive ingredients with antioxidant properties that are useful in the treatment of chronic diseases (Khan and Mukhtar, 2007). The chemical composition of green tea polyphenols includes flavonols, flavandiole and phenolic acids. Most of the polyphenols are flavonols, also known as catechins (Chacko et al., 2010). These potent antioxidant catechins are epicatechin gallate (ECG), epicatechin (EC), epigallocatechin (EGC) and epigallocatechin-3-gallate (EGCG). EGCG has proven to be the most active catechin in green tea (Arab et al., 2011). The antioxidant, antimutagenic, anti-inflammatory, antiviral and antibacterial effect against gram positive bacteria of polyphenols are responsible for their health promoting effect (Chan et al., 2011; Hambire et al., 2015). Some studies have shown that polyphenols are able to inhibit the growth and adhesion of oral pathogens (Lombardo Bedran et al., 2014; Venkateswara et al., 2011).

However, little is known about the effect of green tea (as compared to CHX) in the treatment of gingivitis. Therefore, the purpose of this systematic review is to answer the following PICO question: In patients with gingivitis does mouthwash containing green tea catechins have an effect on plaque accumulation and gingival inflammation?

2. Methods

The review considered the PRISMA checklist (Moher et al., 2015) and the focused question applied the criteria of the PICO method (Miller and Forrest).

2.1. Literature search strategy and study selection

The following electronic databases were reviewed up to January 2017: CINAHL, Cochrane Library, MEDLINE, PubMed and Scopus. The search was limited to human subjects, clinical trails, English and German language.

The following search terms were used (Appendix 1):

Population: "periodontal" OR "parodontal" AND "disease" OR "loss" OR "pocket" OR "pockets" OR "abscess" OR "pericementitis" OR "periodontoses" OR "periodontosis" OR "attachment" AND "loss" OR "clinical" OR "periodontal" OR "parodontal" AND "attachment"

*Intervention: "*camellia" AND "chinensis" OR "sinensis" OR "green tea" OR "epigallocatechin" OR "gallate" OR "gallic acid" OR "Veregen" OR "Exolise"

Two reviewers (SST and YvW) performed the primary search independently and screened the titles and abstracts for inclusion. The same reviewers selected the full

manuscript of those studies meeting the inclusion criteria. Any disagreement was resolved by discussion with a third reviewer (SE).

2.2. Inclusion and exclusion criteria

Inclusion criteria for the present study was conform to PICO criteria (Miller and Forrest). Abstracts were considered if the following inclusion criteria were fulfilled.

Population: patients with gingivitis and good general health.

Intervention: the use of green tea extract in mouthwash.

Comparison: the control group contained either chlorhexidine gluconate or saline.

Outcome: plaque and gingival index (Loe, 1967; Loe and Silness, 1963) were recorded at the baseline and at the end of the trial.

Study design: randomized controlled clinical trials (RCT) or controlled clinical trails (CCT) with a minimum intervention time of 2 weeks with green tea mouthwash.

Studies were excluded for the following reasons: animal studies, in vitro studies, case reports, commentaries, only green tea and not a green tea extract was used, the study population suffered under a systemic disease (for example diabetes) (Appendix 2).

2.3. Outcome measures

The primary outcome measure to assess the efficacy of mouth rinse was the reduction of PI and GI after the use of either a green tea or CHX / saline mouth rinse.

2.4. Data extraction

The following data for each study were extracted: number of subjects, chemical composition of the green tea extract (test), chemical composition of the control rinse (placebo or CHX), study period, PI at baseline and study end and GI at baseline and study end.

2.5. Data analysis and synthesis

To compare and summarize the studies, data were extracted to calculate and analyze mean and standard deviation (SD) at baseline and study end with 95% confidence intervals (CIs). To compare the PI and GI values of the different studies, two types of meta-analysis were performed, analyzing the endpoint and changes over time. Forest plots were created to illustrate the effects of the different studies in the meta-analysis. The open source software R with the package "metafor" was used for a random effects analysis according to the method of DerSimonian and Laird (DerSimonian and Laird,

1986). The statistical heterogeneity among the studies was assessed using the Q test, according to chi-square statistics and the l^2 index ($l^2 = 25\%$: low; $l^2 = 50\%$: moderate; $l^2 = 75\%$: high heterogeneity). The statistical significance was defined as a *p*-value < 0.05.

2.6. Assessment of risk of bias

The studies of interest were evaluated for quality and risk of bias with a modified version of the Cochrane Collaboration's Tool (Graziani et al., 2012) (Appendix 2).

3. Results

3.1. Study selection

Initially, 187 studies were identified by electronic data search. Once the titles and abstracts were screened sixteen potentially relevant studies were subject to full text assessment. In the end, six studies fulfilled the inclusion criteria for the systematic review (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015). The reasons for excluding the remaining studies are reported in Appendix 3. Due to missing information and the presentation of PI and GI values only, the study of Jenabian and colleagues (Jenabian et al., 2012) was also excluded. Finally, five studies (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2017; Nassameemasmaung et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015; Priya et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015) were included for meta-analysis. An outline of the selection process is provided as in a flow-chart in Figure 1.

3.2. Description of characteristics and results

The methodological characteristics of the selected studies are depicted in Table 1. The study characteristics, relevant for the specific research question, are described as follows:

Population: The studies included were performed in India (Hambire et al., 2015; Priya et al., 2015; Sarin et al., 2015), Thailand (Rassameemasmaung et al., 2013) and Iran (Radafshar et al., 2017). The study by Hambire et al. (Hambire et al., 2015) investigated only children (9-14y old). All remaining studies defined the age range between 18-60 years. All studies included patients with gingivitis but no further "periodontal disease reported". Overall, a total of 300 subjects were evaluated in the meta-analysis. In detail 110 subjects participated in the study designed to receive

saline as a negative control and 170 subjects were enrolled in the study distributing CHX in the control group.

Intervention/Comparison: Four (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017; Sarin et al., 2015) out of the five studies prepared a green tea solution by fragmenting dry leaves of *Camellia sinensis*. The final concentration was between 0.5-2% of green tea extract. Rassameemasmaung et al. (Rassameemasmaung et al., 2013) described a green tea extract containing more than 80% total catechins. Three studies (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017) used CHX as a (positive) control, whereas two studies (Rassameemasmaung et al., 2013; Sarin et al., 2015) chose to use normal saline or placebo for comparison. In four studies (Hambire et al., 2015; Radafshar et al., 2017; Sarin et al., 2015), the study population was instructed to rinse twice a day with 10 -15ml for at least 60s. Priya and co-workers (Priya et al., 2015) mentioned the administration of verbal and written oral hygiene instructions but no rinse protocol was described. All patients were advised to follow their routine oral home care, using their usual toothbrush and toothpaste. Only participants from the study by Priya et al. (Priya et al., 2015) were advised to use the modified bass technique. Study periods ranged from two to four weeks.

Outcome: All studies recorded PI (Loe, 1967) and GI (Loe and Silness, 1963) at baseline and at the end of examination. The endpoint for the study by Hambire et al. (Hambire et al., 2015) was 15 days, while all the other studies concluded after 28 days. **Study design:** All studies included a control group, which used either CHX (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017) or saline (Rassameemasmaung et al., 2013; Sarin et al., 2015). Three controlled clinical trails were double blinded (Hambire et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013), one triple blinded (Sarin et al., 2015) and one single blinded (Priya et al., 2015).

3.3. Outcome of the intervention on plaque index (PI) and gingival index (GI)

In the present systematic review, plaque Index (PI) and gingival Index (GI) endpoint measurements were compared between test (green tea) and control (CHX or placebo) groups. In addition, differences in changes between baseline and endpoint measurements for test and control groups were taken.

a) Plaque Index (PI): Test vs. CHX

Three Studies (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017) measured PI after intervention with green tea or CHX mouth rinse (Priya et al., 2015; Radafshar et al., 2017). Two of the studies using CHX as a control agent started with a higher PI in the control group, except for the study conducted by Hambire et al. (Hambire et al., 2015), who showed close to identical values (Table 2a).

b) Plaque Index (PI): Test vs. Placebo

Two studies (Rassameemasmaung et al., 2013; Sarin et al., 2015) measured mean PI after intervention of green tea and placebo mouth rinse. At baseline both the intervention group as well as the control group showed similar values in PI but not at endpoint. The test group displayed slightly better values of PI than the control group but not statistically significant (Table 2b).

c) Gingival Index (GI): Test vs. CHX

Three Studies (Hambire et al., 2015; Priya et al., 2015; Radafshar et al., 2017) measured GI after intervention of green tea or CHX mouth rinse (Table 3a). Both the test and control group show similar GI values at baseline.

d) Gingival Index (GI): Test vs. Placebo

Two studies (Rassameemasmaung et al., 2013; Sarin et al., 2015) compared GI between the green tea group and the control (saline) group. Both groups started out with similar values for GI (Table 3b). Overall, at the end of the interventions little change was seen in either the green tea or control group.

e) Meta-analysis PI

A meta- analysis was conducted to evaluate changes in PI at endpoint and over time. Further, a Forest plot was created to illustrate the data referenced in the meta- analysis (Table 4a-d). Overall the meta-analysis revealed no statistically significant changes between the test and CHX control groups, neither at endpoint nor over time (baseline – endpoint). Treatment modalities with a placebo (saline) also did not show any statistically significant data. With the exception of the study by Radafshar et al. (Radafshar et al., 2017), the Forest plot did show a tendency in favor of green tea, based on the data representing change over time.

f) Meta-analysis GI

To evaluate changes in GI at endpoint and over time, a second meta-analysis was performed (Tables 5 a-d). Data of the meta-analysis showed neither statistically significant results in GI values at endpoint nor changes over time. A weak trend in favor of green tea could be observed in changes over time (p= 0.47, - 0.03 [-0.13, 0.06]; p= 0.33, - 0.3 [- 0.94, 0.31]) (Tables 5 c-d).

g) Adverse-effects

Four Studies (Hambire et al., 2015; Radafshar et al., 2017; Rassameemasmaung et al., 2013; Sarin et al., 2015) investigated adverse effects by questionnaire; none were reported. Unpleasant or altered taste and one single epithelium desquamation was reported by Radafshar and coworkers (Radafshar et al., 2017). Priya and colleagues (Priya et al., 2015) reported no statistical significance on tooth- or tongue stain.

4. Discussion

The present study evaluated the clinical efficacy of green tea containing mouthwash and its effect on plaque and gingival inflammation. Rinsing with green tea catechins resulted in a similar PI and GI as CHX or Placebo, without the side effects commonly experienced with CHX usage. The present systematic review included five randomized single and double-blinded controlled trials using green tea catechin containing mouthwash. Overall the data showed no statistically significant changes between the different mouthwash modalities used. However, the Forest plots illustrated a trend in favor of green tea. Hence, green tea mouthwash may be a good alternative to CHX as an adjunctive product in daily home care.

Axelsson and colleagues (Axelsson et al., 2004) have demonstrated that gingivitis can be effectively prevented and treated by well-performed mechanical oral hygiene, including tooth brushing in combination with interdental cleaning. Whereas mechanical means of plaque removal have gained widespread acceptance, it is interesting to examine the adjunctive benefits of chemotherapeutic mouthwash (Afennich et al., 2011). There is evidence that chemical agents can be effective against gingivitis. While mechanical plaque removal is considered to be the most effective means of removing biofilm from tooth surfaces (Jongsma et al., 2015), chemotherapeutic mouthwashes bear consideration for their ease of use, patient acceptance and ability to reach areas patients cannot always reach with their toothbrush or where tooth brushing may be contraindicated (ie. post-surgical sites). Although there are limitations in the efficacy of mouthwashes (e.g. inability to penetrate mature plaque), there is evidence that such chemotherapeutic solutions can be effective against gingivitis (Albert-Kiszely et al., 2007; Prasad et al., 2016; Sharma et al., 2004).

In today's Western population, mouthwash is accepted as adjunctive agent for prophylaxis and the battle against gingivitis, periodontal disease and caries. An increasing level of awareness about oral microbiology and plaque has led to the development of specific strategies, utilizing the antimicrobial effects of chemical substances (Afennich et al., 2011) Therefore, the requirement of an appropriate mouth rinse is that it contains antiseptic, antiplaque and anti-caries properties. In addition, mouth rinses with no (or just few minor) side effects are desired (Baker, 1993).

Chlorhexidine gluconate is considered the gold standard of antimicrobial mouthwash for the prevention of dental plaque formation (Axelsson and Lindhe, 1987). Compared to other antiseptics, CHX has been shown to be the most effective (Gjermo et al., 1970). The ability of CHX to reduce plaque formation and gingival inflammation was demonstrated in several systematic reviews (Gunsolley, 2010; Van Strydonck et al., 2012), and most recently by Serrano and colleagues (Serrano et al., 2015). Berchier et al. considered the clinical relevance of 0,2% versus 0.12% CHX to be negligible (Berchier et al., 2010). Rinsing with CHX in addition to oral hygiene procedures results in approximately 33% less plaque and 26% less gingivitis as compared to controls (Van Strydonck et al., 2012). Gunsolley et al. evaluated 0.12% CHX and achieved a mean GI reduction of 28,7% and a mean PI reduction of up to 40,4% (Gunsolley, 2010). However due to the side effects of CHX, it is not recommended to use this agent for a prolonged time period (Supranoto et al., 2015; Van der Weijden et al., 2015; Van Strydonck et al., 2012). While reversible, local side effects such as staining and taste alteration are associated with long-term use of CHX mouth rinse (Supranoto et al., 2015). All these factors - to a varying degree - adversely affect patient compliance (Addy et al., 1995).

One of the shortcomings of this systematic review is the lack of uniformity in regards to the solutions employed in the control groups. Hence, the authors have chosen to separate both groups for evaluation of the meta-analysis. Nevertheless, the studies still revealed a high level of heterogeneity. The clinical protocol guidelines (ADA guidelines) demand statistically significant data in the reduction for both plaque and gingivitis. The present meta-analysis could not verify statistically significant data. An additional limitation is the short time frame of the studies examined, none of which fulfilled the 6-month requirement necessary for the ADA seal of acceptance in "Chemotherapeutic Products for Control of Gingivitis". A 6-month period is required to evaluate both the efficacy and safety of chemical agents as well as patients' compliance (https://www.ada.org/en/science-research/ada-seal-of-acceptance/how-to-earn-the-ada-seal/general-criteria-for-acceptance).

The studies included in this systematic review advised all participants to follow their normal oral hygiene protocol. However as past studies have shown, daily use of a toothbrush may both prohibit gingival inflammation from occurring and if present, may lead to its reversal as a stand-alone measure (Loe et al., 1965). On this basis, the effectiveness of green tea as a mouth rinse agent may not be proven. In addition, all participants were very young, in good health, had no systemic diseases and predominately encompassed a population from India, Thailand and Iran. Therefore, further studies on diverse ethnic groups are recommended. This systematic review can be considered as a good beginning point for further reports on this product application. In summary, placebo-controlled randomized clinical trials with an expanded observation time of at least 6 months are recommended and necessary to determine the beneficial effects of green tea catechins on dental plaque and gingival inflammation.

5. Conclusion

It appears that Green tea may have an antiplaque and anti-gingivitis effect without causing side effects. Due to the highly heterogeneous data and the risk of bias, this evidence needs to be interpreted with caution. Further placebo controlled clinical trials with longer observation periods are needed.

Conflict of interests

The authors declare that there are no conflicts of interest.

Acknowledgement

The authors would like to acknowledge the work of Dr. S. Steppacher for her support in the analysis of the data as part of her master thesis. We also would like to thank our colleague D. Hofer from the Clinic of Preventive Dentistry, Periodontology and Cariology, Center of Dental Medicine, University of Zurich, Switzerland for proof reading the article.

Funding sources

This systematic review did not receive any grants from funding agencies or any outside sources.

Appendix 1-3 and Supplementary data

Supplementary data associated with this article can be found, in the online version

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Messora, M.R., 2017. Benefits of Bifidobacterium animalis subsp. lactis Probiotic in Experimental Periodontitis. The Journal of periodontology 88(2), 197-208.

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- Radafshar, G., Ghotbizadeh, M., Saadat, F., Mirfarhadi, N., 2017. Effects of green tea (Camellia sinensis) mouthwash containing 1% tannin on dental plaque and chronic gingivitis: a double-blinded, randomized, controlled trial. Journal of investigative and clinical dentistry 8(1).
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Appendix 1: Pubmed search strategy

Step	Query	Hits
1	Search (((((((periodontal[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND (disease[tiab] OR diseases[tiab] OR loss[tiab] OR pocket[tiab] OR pockets[tiab] OR abscess[tiab] OR abscesses[tiab] OR index[tiab])) OR (pericementitides[tiab] OR pericementitis[tiab] OR periodontitides[tiab] OR periodontitis[tiab] OR periodontoses[tiab] OR periodontosis[tiab] OR paradontitis[tiab] OR parodontitis[tiab]) OR (attachment[tiab] AND loss[tiab]) OR ((clinical[tiab] OR periodontal[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND attachment[tiab])))) AND (((((camellia[tiab] AND (chinensis[tiab] OR sinensis[tiab]))) OR ("green tea"[tiab] OR epigallocatechin[tiab] OR gallate[tiab] OR "gallic acid"[tiab] OR Veregen[tiab] OR Exolise[tiab]))))) AND ((((inprocess[sb])) OR (publisher[sb] NOT pubstatusnihms NOT pubstatuspmcsd NOT pmcbook))	8
2	Search ((inprocess[sb])) OR (publisher[sb] NOT pubstatusnihms NOT pubstatuspmcsd NOT pmcbook)	1077566
3	Search (((((periodontal[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND (disease[tiab] OR diseases[tiab] OR loss[tiab] OR pocket[tiab] OR pockets[tiab] OR abscess[tiab] OR abscesses[tiab] OR index[tiab])) OR (pericementitides[tiab] OR pericementitis[tiab] OR periodontitides[tiab] OR periodontitis[tiab] OR periodontoses[tiab] OR periodontosis[tiab] OR paradontitis[tiab] OR parodontitis[tiab]) OR (attachment[tiab] AND loss[tiab]) OR ((clinical[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND attachment[tiab]))) AND (((((camellia[tiab] AND (chinensis[tiab] OR sinensis[tiab]))) OR ("green tea"[tiab] OR epigallocatechin[tiab] OR gallate[tiab] OR "gallic acid"[tiab] OR Veregen[tiab] OR Exolise[tiab]))))	57
4	Search ((((camellia[tiab] AND (chinensis[tiab] OR sinensis[tiab])) OR ("green tea" [tiab] OR epigallocatechin[tiab] OR gallate[tiab] OR "gallic acid"[tiab] OR Veregen[tiab] OR Exolise[tiab]))	14095
5	Search (((periodontal[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND (disease[tiab] OR diseases[tiab] OR loss[tiab] OR pocket[tiab] OR pockets[tiab]	49475

OR abscess[tiab] OR abscesses[tiab] OR index[tiab])) OR	
(pericementitides[tiab] OR pericementitis[tiab] OR periodontitides[tiab] OR periodontitis[tiab]	
OR Periodonalis[liab] ON periodonalides[liab] ON periodonalis[liab]	
periodontoses[tiab] OR periodontosis[tiab] OR paradontitis[tiab]	
ORparodontitis[tiab]) OR (attachment[tiab] AND loss[tiab]) OR	
((clinical[tiab] OR	
periodontal[tiab] OR parodontal[tiab] OR paradontal[tiab]) AND	
attachment[tiab]))	

Appendix 3: Excluded studies

Excluded studies

Reason for exclusion

Sinija VR, Mishra HN. Green tea: health benefits. <i>Journal of Nutritional</i> & <i>Environmental Medicine</i> . 2008;17(4):232–24211p.	Not addressing research question
Molina FD, Maniglia JV, Magalhaes FP, Dafico SR, Rezende RS. The efficacy of bismuth subgallate in tonsillectomy as hemostatic agent. <i>Rev Bras Otorrinolaringol</i> . 2000;66(3):194-197.	Not addressing research question
Magee K, Loiacono C. A review of common herbs and potential interactions. <i>Int J Dent Hygiene</i> . 2004;2(3):111–12111p.	Not addressing research question
Lerman A, Lockwood B. Nutraceuticals in veterinary medicine. <i>PharmJ</i> .2007;278(7434):51-55.	Not addressing research question
Pavel L, Pavel S. [Usefulness of micronutrients in the treatment of periodontitis]. <i>Ned Tijdschr Tandheelkd</i> . 2010;117(2):103-106.	Full text assessment
Cravotto G, Boffa L, Genzini L, Garella D. Phytotherapeutics: an evaluation of the potential of 1000 plants. <i>Journal of Clinical Pharmacy & Therapeutics</i> . 2010;35(1):11–4838p. doi:10.1111/j.1365-2710.2009.01096.x.	Not addressing research question
Priya BM, Anitha V, Shanmugam M, Ashwath B, Sylva SD, Vigneshwari SK. Efficacy of chlorhexidine and green tea mouthwashes in the management of dental plaque-induced gingivitis: A comparative clinical study. <i>Contemp.</i> 2015;6(4):505-509. doi:10.4103/0976-237X.169845. Viana GSB, Menezes SMS, Cordeiro LN, Matos FJA. Biological effects of	Included study
pomegranate (Punica granatum L.), especially its antibacterial actions, against microorganisms present in the dental plaque and other infectious processes. In: <i>Bioactive Foods in Promoting Health</i> . Elsevier Inc.; 2010:457-478. doi:10.1016/B978-0-12-374628-3.00031-1.	Not addressing research question
Isogai H, Isogai E, Takahashi K, Kurebayashi Y. Effect of catechin diet on gingivitis in cats. <i>International Journal of Applied Research in Veterinary Medicine</i> . 2008;6(2):82-86.	Animal study
Gurenlian JR, Spolarich AE. Risk assessment for clients with diabetes. <i>Access</i> . 2010;24(6):32–354p.	Not addressing research question
Jurenka J. Therapeutic applications of pomegranate (Punica granatum L.): a review. <i>Alternative Medicine Review</i> . 2008;13(2):128–14417p.	Not addressing research question
Lauten JD, Boyd L, Hanson MB, Lillie D, Gullion C, Madden TE. A clinical study: Melaleuca, Manuka, Calendula and green tea mouth rinse. <i>Phytother Res</i> . 2005;19(11):951-957.	Not addressing research question
Wang CY, Deng YT, Huang SY, Liu CM, Chang HH, Wong MY. Epigallocatechin-3-gallate inhibits lysophosphatidic acid-stimulated connective tissue growth factor via JNK and Smad3 suppression in human gingival fibroblasts. <i>J Formos Med Assoc</i> . 2014;113(1):50-55.	
doi:10.1016/j.jfma.2012.04.004.	In vitro study
Hurd L. Publisher's notes. <i>Total Health</i> . 2006;27(6):6–61p.	Not addressing research question
Ravi K, Divyashree P. Psidium guajava: A review on its potential as an adjunct in treating periodontal disease. <i>Pharmacogn Rev.</i> 2014;8(16):96-100. doi:10.4103/0973-7847.134233.	Review
Funosas ER, Martinez AB, Pignolo M, et al. Efficacy of green tea in the treatment of chronic periodontitis. <i>Av Odontoestomatol</i> . 2005;21(3):159-166.	Full text assessment
Anonymous. Green tea and oral health examined in study. <i>Br Dent J.</i> 2010;208(9):384. doi:10.1038/sj.bdj.2010.436.	Not addressing research question

Kozai K, Suzuki J, Okada M, Nagasaka N. Effect of oleanolic acid-cyclodextrin inclusion compounds on dental caries by in vitro experiment and rat-caries model. <i>Microbios</i> . 1999;97(388):179-188.	Animal study
Gardner EJ, Ruxton CHS, Leeds AR. Black tea - Helpful or harmful? A review of the evidence. <i>Eur J Clin Nutr</i> . 2007;61(1):3-18. doi:10.1038/sj.ejcn.1602489.	Review
5th Joint Meeting of the European Tissue Repair Society and the Wound Healing Society. <i>Wound Repair & Regeneration</i> . 2009;17(4):A54–871p.	Not addressing research question
Ooshima T, Minami T, Aono W, et al. Oolong tea polyphenols inhibit experimental dental caries in spf rats infected with mutans streptococci. <i>Caries Res.</i> 1993;27(2):124-129. doi:10.1159/000261529.	Animal study
Strausfogel S. Dental care to smile about. <i>Better Nutrition</i> . 2007;69(2):42-421p.	Not addressing research question
Prithi R, Geetha RV. Static effects of fruits on periodontitis. <i>Res J Pharm</i> <i>Technol</i> . 2014;7(3):365-367.	Not addressing research question
Tomczyk M, Pleszczynska M, Wiater A. Variation in Total Polyphenolics Contents of Aerial Parts of Potentilla Species and Their Anticariogenic Activity. <i>Molecules</i> . 2010;15(7):4639-4651. doi:10.3390/molecules15074639.	In vitro study
Spratt DA, Daglia M, Papetti A, et al. Evaluation of Plant and Fungal Extracts for Their Potential Antigingivitis and Anticaries Activity. <i>J Biomed Biotechnol</i> . 2012:12. doi:10.1155/2012/510198.	Not addressing research question
Zagorouiko V, Mizin V, Bogadelnikov I, Ogay U. The Dietary Grape Polyphenol Concentrate "ENOANT" Enables Protection Against Biological Agents. In: Dishovsky C, Pivovarov A, eds. <i>Counteraction to Chemical and Biological</i> <i>Terrorism in East European Countries</i> . Dordrecht: Springer; 2009:167-176. doi:10.1007/978-90-481-2342-1_21. Signoretto C, Canepari P, Pruzzo C, Gazzani G. Woodhead Publ. Food Sci. Technol. Nutr. In: Wilson M, ed. <i>Food Constituents and Oral Health: Current</i> <i>Status and Future Prospects</i> . Cambridge: Woodhead Publ Ltd; 2009:240-262.	Not addressing research question
doi:10.1533/9781845696290.2.240.	Not addressing research question
Subapriya R, Nagini S. Medicinal properties of neem leaves: A review. <i>Curr Med Chem Anti-Cancer Agents</i> . 2005;5(2):149-156. doi:10.2174/1568011053174828.	Not addressing research question
Belozerskaya GG, Makarov VA, Zhidkov EA, et al. Local hemostatics (A review). <i>Pharm Chem J.</i> 2006;40(7):353-359. doi:10.1007/s11094-006-0126-3.	Not addressing research question
Pandit S, Song K-Y, Jeon J-G. Withania somnifera Attenuates Acid Production, Acid Tolerance and Extra-Cellular Polysaccharide Formation of Streptococcus mutans Biofilms. <i>American Journal of Chinese Medicine</i> . 2014;42(1):157– 17115p. doi:10.1142/S0192415X14500116.	Not addressing research question
Hamilton-Miller JMT. Anti-cariogenic properties of tea (Camellia sinensis). <i>J Med Microbiol.</i> 2001;50(4):299-302.	Not addressing research question
Choi CH, Kim BI, Kwon HK, Hong SJ. Effects of herbal extracts on dental plaque formation and human gingival fibroblasts. In: Kim YH, Cho CS, Kang IK, Kim SY, Kwon OH, eds. <i>ASBM7: Advanced Biomaterials VII</i> . Vol 342-343. Stafa-Zurich: Trans Tech Publications Ltd; 2007:773-776.	In vitro
ASBM7: Advanced Biomaterials VII. Stafa-Zurich: Trans Tech Publications Ltd; 2007.	Not addressing research question
Patel VK, Venkatakrishna-Bhatt H. Folklore therapeutic indigenous plants in periodontal disorders in India (review, experimental and clinical approach). <i>Int J Clin Pharmacol Ther Toxicol.</i> 1988;26(4):176-184.	Not addressing research question
Corwin A, Zahorik L, Hurlbutt M. Herbal supplements: healthcare implications and considerations. <i>Journal of the California Dental Hygienists' Association</i> . 2009;24(2):7–159p.	Not addressing research question

GREEN TEA BENEFITS. <i>Explorer (08947929)</i> . 2015;41(1):2–21p.	Not addressing research question
Xu X, Zhou XD, Wu CD. Tea catechin epigallocatechin gallate inhibits Streptococcus mutans biofilm formation by suppressing gtf genes. Arch Oral	····
<i>Biol.</i> 2012;57(6):678-683. doi:10.1016/j.archoralbio.2011.10.021.	Not addressing research question
Shanbhag VK. Triphala in prevention of dental caries and as an antimicrobial in oral cavity- A review. <i>Infect Disord Drug Targets</i> . 2015;15(2):89-97.	Not addressing research question
Jaladat AM, Atarzadeh F, Rezaeizadeh H, et al. Botanicals: An alternative remedy to radiotherapy-induced dysuria. <i>Complement Ther Med.</i> 2015;23(1):90-99. doi:10.1016/j.ctim.2014.11.004.	Not addressing research question
Montbriand MJ. Herbs or natural products that increase cancer growth or recurrence: part two of a four-part series [corrected] [published erratum appears in ONCOL NURS FORUM 2006 Jul;33(4):684]. <i>Oncology Nursing Forum.</i> 2004;21(5):E00_11(5):E00_	Net oddressing
2004;31(5):E99–1151p.	Not addressing research question
Varoni EM, Lodi G, Sardella A, Carrassi A, Iriti M. Plant Polyphenols and Oral Health: Old Phytochemicals for New Fields. <i>Current Medicinal Chemistry</i> . 2012;19(11):1706-1720.	Not addressing research question
Logan EI. Dietary Influences on Periodontal Health in Dogs and Cats. <i>Vet Clin North Am Small Anim Pract.</i> 2006;36(6):1385-1401.	
doi:10.1016/j.cvsm.2006.09.002.	Not addressing research question
Awadalla HI, Ragab MH, Fayed MT, Abbas MO, Bassuoni MW. Evaluation of the effect of green tea on dental caries and composite restorations. TAF Preventive Medicine Bulletin.	
http://onlinelibrary.wiley.com/o/cochrane/clcentral/articles/483/CN- 00894483/frame.html. Published 2011. Accessed January 31, 2016.	Not addressing research question
Soref A. A swish a day. <i>Better Nutrition</i> . 2006;68(2):46–461p.	Not addressing research question
Makimura M, Hirasawa M, Kobayashi K, et al. Inhibitory effect of tea catechins on collagenase activity. <i>Journal of Periodontology</i> . 1993;64(7):630-636.	Not addressing research question
Williamson MP, Trevitt C, Noble JM. NMR-STUDIES OF DEXTRAN OLIGOMER INTERACTIONS WITH MODEL POLYPHENOLS. <i>Carbohydrate</i> <i>Research</i> . 1995;266(2):229-235. doi:10.1016/0008-6215(94)00273-i.	Not addressing research question
Hattarki SA, Pushpa SP, Bhat K. Evaluation of the efficacy of green tea catechins as an adjunct to scaling and root planing in the management of chronic periodontitis using PCR analysis: A clinical and microbiological study. <i>J Indian Soc Periodontol.</i> 2013;17(2):204-209. doi:10.4103/0972-124X.113071.	Not addressing research question
Finkel J. In the news. Green tea may strengthen teeth. <i>Life Extension</i> . 2010;16(7):20–201p.	Not addressing
	research question
Oberg E. Preconception counseling: helping patients plan for the future. <i>Integrative Medicine: A Clinician's Journal</i> . 2009;8(4):46–494p.	Not addressing research question
Yoo S, Murata RM, Duarte S. Antimicrobial Traits of Tea- and Cranberry- Derived Polyphenols against Streptococcus mutans. <i>Caries Res.</i> 2011;45(4):327-335. doi:10.1159/000329181.	Not addressing research question
Spratt DA, Daglia M, Papetti A, et al. Evaluation of Plant and Fungal Extracts for Their Potential Antigingivitis and Anticaries Activity. <i>J Biomed Biotechnol</i> . 2012;2012:1–1212p.	Not addressing research question
Daglia M, Papetti A, Mascherpa D, et al. Plant and Fungal Food Components with Potential Activity on the Development of Microbial Oral Diseases. <i>J Biomed</i>	Not addressing research question

Biotechnol. 2011:1-99p.

Abebe W. Literature review. An overview of herbal supplement utilization with particular emphasis on possible interactions with dental drugs and oral manifestations. <i>Journal of Dental Hygiene</i> . 2003;77(1):37–4610p.	Not addressing research question
Consolini AE, Ragone MI. Patterns of self-medication with medicinal plants and related adverse events - A South American survey. <i>Curr Drug Saf.</i> 2010;5(4):333-341. doi:10.2174/157488610792246019.	Not addressing research question
Cai LN, Wu CD. Compounds from Syzygium aromaticum possessing growth inhibitory activity against oral pathogens. <i>J NAT PROD</i> . 1996;59(10):987-990. doi:10.1021/np960451q	Not addressing research question
Wojtaszek C. Management of chemotherapy-induced stomatitis. <i>Clinical Journal of Oncology Nursing</i> . 2000;4(6):263–28210p.	Not addressing research question
Ruxton C. Fluoride in the UK diet. <i>Nursing Standard</i> . 2014;28(49):52–598p. doi:10.7748/ns.28.49.52.e9031.	Not addressing research question
Krahwinkel T, Willershausen B. The effect of sugar-free green tea chew candies on the degree of inflammation of the gingiva. <i>Eur J Med Res.</i> 2000;5(11):463-467.	Full text assessment
Homer KA, Manji F, Beighton D. Inhibition of protease activities of periodontopathic bacteria by extracts of plants used in Kenya as chewing sticks (mswaki). <i>Arch Oral Biol.</i> 1990;35(6):421-424. doi:10.1016/0003-9969(90)90203-M.	Not addressing research question
Radafshar G, Ghotbizadeh M, Saadat F, Mirfarhadi N. Effects of green tea (Camellia sinensis) mouthwash containing 1% tannin on dental plaque and chronic gingivitis: a double-blinded, randomized, controlled trial. <i>J Investig Clin Dent</i> . August 2017. doi:10.1111/jicd.12184.	Included study
Gadagi JS, Chava VK, Reddy VR. Green tea extract as a local drug therapy on periodontitis patients with diabetes mellitus: A randomized case-control study. <i>J Indian Soc Periodontol.</i> 2013;17(2):198-203. doi:10.4103/0972-124X.113069.	Full text assessment
Strausfogel S. Gum health guide. <i>Better Nutrition</i> . 2008;70(11):32–342p.	Not addressing research question
Liu T, Chi Y. [Experimental study on polyphenol anti-plaque effect in human]. <i>Chung Hua Kou Chiang Hsueh Tsa Chih</i> . 2000;35(5):383-384.	Not addressing research question
News from The Journal of Chinese Medicine. <i>Journal of Chinese Medicine</i> . 2006;(81):67–737p.	Not addressing research question
Awadalla HI, Ragab MH, Bassuoni MW, Fayed MT, Abbas MO. A pilot study of the role of green tea use on oral health. <i>Int J Dent Hygiene</i> . 2011;9(2):110-116. doi:10.1111/j.1601-5037.2009.00440.x.	Full text assessment
Rivai H, Rina W, Rina DY, et al. Preparation and evaluation of herbal tea and toothpaste of mulberry leaves (Morus alba L.). <i>Res J Pharm, Biol Chem Sci.</i> 2015;6(4):1672-1677.	Not addressing research question
Abstracts. Alternative Medicine Review. 2006;11(3):244–26118p.	Not addressing research question
Sirois M, Darby M, Tolle S. Understanding Muslim patients: cross-cultural dental hygiene care. <i>Int J Dent Hygiene</i> . 2013;11(2):105–11410p. doi:10.1111/j.1601-5037.2012.00559.x.	Not addressing research question
Ready, set, shop. <i>Better Nutrition</i> . 2007;69(7):14–141p.	Not addressing research question

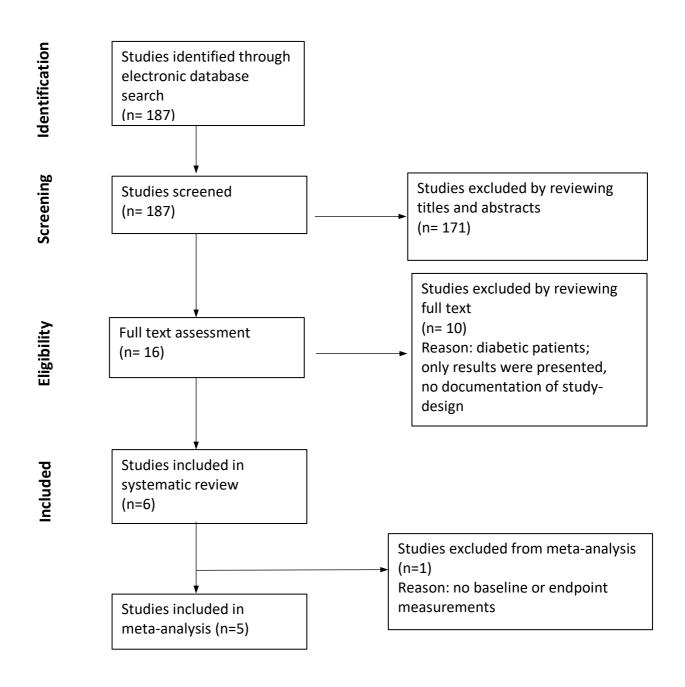
Low SB, Peak RM, Smithson CW, Perrone J, Gaddis B, Kontogiorgos E. Evaluation of a topical gel containing a novel combination of essential oils and antioxidants for reducing oral malodor in dogs. <i>American Journal of Veterinary</i> <i>Research</i> . 2014;75(7):653-657.	Animal study
Allaker RP, Douglas CWI. Novel anti-microbial therapies for dental plaque- related diseases. <i>International Journal of Antimicrobial Agents</i> . 2009;33(1):8-13. doi:10.1016/j.ijantimicag.2008.07.014.	Not addressing research question
Kaur H, Jain S, Kaur A. Comparative evaluation of the antiplaque effectiveness of green tea catechin mouthwash with chlorhexidine gluconate. <i>J Indian Soc Periodontol.</i> 2014;18(2):178-182. doi:10.4103/0972-124X.131320.	Full text assessment
Recently published abstracts. <i>Alternative Medicine Review</i> . 2010;15(4):369–38012p.	Not addressing research question
Turner L. Indulge. Revive. Glow. <i>Better Nutrition</i> . 2011;73(4):31–4010p.	Not addressing research question
Hambire CU, Jawade R, Patil A, Wani VR, Kulkarni AA, Nehete PB. Comparing the antiplaque efficacy of 0.5% Camellia sinensis extract, 0.05% sodium fluoride, and 0.2% chlorhexidine gluconate mouthwash in children. <i>J.</i> 2015;5(3):218-226. doi:10.4103/2231-0762.158016.	Included study
Hsu S-P, Liao C-S, Li C-Y, Chiou A-F. The effects of different oral care protocols on mucosal change in orally intubated patients from an intensive care unit. <i>Journal of Clinical Nursing</i> . 2011;20(7/8):1044–105310p. doi:10.1111/j.1365-2702.2010.03515.x.	Not addressing research question
Bedran TBL, Morin MP, Spolidorio DP, Grenier D. Black Tea Extract and Its Theaflavin Derivatives Inhibit the Growth of Periodontopathogens and Modulate Interleukin-8 and beta-Defensin Secretion in Oral Epithelial Cells. <i>PLoS ONE</i> . 2015;10(11):11. doi:10.1371/journal.pone.0143158. Rassameemasmaung S, Phusudsawang P, Sangalungkarn V. Effect of green tea mouthwash on oral malodor. <i>ISRN Prev Med</i> . 2013;2013:975148.	Not addressing research question
doi:10.5402/2013/975148.	Included study
Spolarich AE, Andrews L. An examination of the bleeding complications associated with herbal supplements, antiplatelet and anticoagulant medications. <i>Journal of Dental Hygiene</i> . 2007;81(3):67–671p.	Not addressing research question
Percival RS, Devine DA, Duggal MS, Chartron S, Marsh PD. The effect of cocoa polyphenols on the growth, metabolism, and biofilm formation by Streptococcus mutans and Streptococcus sanguinis. <i>European Journal of Oral Sciences</i> . 2006;114(4):343-348.	Not addressing research question
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Wang RK, Zhao PP, Zhu B, Li JY. Inhibitive effect of extracts of Galla Chinesis on caries development in rats. <i>J Sichuan Univ Med Sci Ed</i> . 2008;39(3):474-477.	Not addressing research question
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Signoretto C, Burlacchini G, Bianchi F, Cavalleri G, Canepari P. Differences in microbiological composition of saliva and dental plaque in subjects with different	Not addressing research question

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2231-20-39.	Full text assessment
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CARIOGENIC BACTERIA IN-VITRO BY PLANT FLAVANONES. Experientia. 1994;50(9):846-849. doi:10.1007/bf01956469.	Not addressing research question
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teeth extraction: a randomized controlled trial. <i>Evid Based Complement Alternat</i> <i>Med.</i> 2014;2014:857651. doi:10.1155/2014/857651.	Not addressing research question
Hrishi T, Kundapur P, Naha A, Thomas B, Kamath S, Bhat G. Effect of adjunctive use of green tea dentifrice in periodontitis patients - A Randomized Controlled Pilot Study. <i>Int J Dent Hygiene</i> . 2015. doi:10.1111/idh.12131.	Not addressing research question

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Oli MW, Otoo HN, Crowley PJ, et al. Functional amyloid formation by Streptococcus mutans. <i>Microbiology</i> . 2012;158(Pt 12):2903-2916. doi:10.1099/mic.0.060855-0.	Not addressing research question
Kukreja BJ, Dodwad V. Herbal mouthwashes - A gift of nature. <i>Intl J Pharma Bio Sci</i> . 2012;3(2):P46-P52.	Not addressing research question
Tamura M, Saito H, Kikuchi K, et al. Antimicrobial Activity of Gel-Entrapped Catechins toward Oral Microorganisms. <i>Biol Pharm Bull.</i> 2011;34(5):638-643.	Not addressing research question
Kemoli AM, van Amerongen WE, de Soet JJ. Antimicrobial and buffer capacity of crude extracts of chewing sticks (Miswaki) from Kenya. <i>J Dent Child.</i> 2001;68(3):183–8–152.	Not addressing research question
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Market basket. <i>Better Nutrition</i> . 1998;60(11):68–681p. Scherer W, Gultz J, Lee SS, Kaim J. The ability of an herbal mouthrinse to reduce gingival bleeding. <i>J Clin Dent</i> . 1998;9(4):97-100.	Not addressing research question Not addressing research question
Han KC, Wong WC, Benzie IFF. Genoprotective effects of green tea (Camellia sinensis) in human subjects: results of a controlled supplementation trial. <i>Br J Nutr</i> . 2011;105(2):171-179. doi:10.1017/S0007114510003211.	Not addressing research question



First author (year of publication)	Population characteristics (ethnici- ty, age, inclusion crite- ria)	Number of subjects	Study design, Intervention	Green tea characteristics, clinical measure- ments, study period, oral prophylax- is/instructions	Analyzed parameters
Priya (2015)	 Ethnicity: Indian 18 to 24 years Inclusion criteria: Gl≥1 (≥60% of sites) Pl≥1 PPD ≤3mm no CAL loss good medical health Exclusion criteria: systemic and/or topical steroidal drugs; NSAR; topical or systemic antibiotic treatment in the last 6 weeks; fixed or removable ortho- dontic device; oral soft tissue pathology; physical or mental handicap; no oral prophylaxis in the last 6 month 	30	 Study design: single blinded randomized controlled trial Application: na Quantity: na 	 Test: green tea extract made of fragmented leaves (<i>Camellia sinensis</i>; 0,5g extract + 10ml dis- tilled water), 5% Mouthwash solution Control: CHX 0,2% Clinical measurements: baseline, 15d, 28d (endpoint) Study period: 28 d Oral prophylaxis/instructions: modified bass technique 	 PI plaque index (Turesky-Gilmore-Glickman 1970) GI gingival Index (Löe and Silness 1963) BI bleeding Index (Ainamo & Bay TS Tongue Index (Caydon 2001) Stain index (Gründemann et al. 2000)
Radafshar (2017)	 Ethnicity: Iran 18 to 25 years Inclusion criteria: Gl≥1 minimal supragingival calculus on at least 20 teeth no clinical signs for periodontitis Exclusion criteria: fixed or removable appliances severe dental crowd- ing mouth breathing smoking and systemic disease 	40	 Study design: double blinded randomized clinical controlled trial Application: rinse twice 15mL per day for 60s 1h no coloring drinks after rinsing 	 Test: green tea extract made of fragmented leaves of <i>Camellia sinensis</i>, containing 1% of tannin, Mouthwash solution Control: CHX 0,12% Clinical measurements: baseline, 28d (endpoint) Study period: 28 d Oral prophylaxis/instructions: routine oral hygiene report of any side effects 	 PI plaque index (Turesky-Gilmore-Glickman, 1970) GI gingival Index (Löe and Silness 1963) GBI gingival bleeding Index side effects

Table 1: Study characteristics of included studies to the review and meta- analysis

First author (year of publication)	Population characteristics (ethnici- ty, age, inclusion crite- ria)	Number of subjects	Study design, Intervention	Green tea characteristics, clinical measure- ments, study period, oral prophylax- is/instructions	Analyzed parameters
Hambire (2015)	 Ethnicity: Indian 9 to 14 years Inclusion criteria: normal occlusion, absence of caries and/or restauration, healthy periodontium, oral health Exclusion criteria: history of antibiotics, topical fluorid treat- ment with 4 week prior to baseline, regular use of xylitol chewing gum, tea, coffee, sys- temic diseases, ortho- dontic appliances 	30	 Study design: double blinded controlled trial Application: rinse twice a day for 60s (no quantity) no beverage or food for 1h 	 Test: green tea leaves with mineral water, steeped, refrigderated. 0,5% solution of green tea mouthwash Control: CHX 0,2% Sodium fluoride 0,05% Clinical measurements: baseline, 14d (endpoint) Study period: 14d Oral prophylaxis/instructions: routine oral hygiene at baseline professional oral hygiene with scaling and root planning and polishing 	PI plaque index (keine Angabe!) GI gingival index (keine Angabe!) Adverse effects by questionnaire Salivary pH (keine Angabe)
Ras- sameemasma ung (2013)	 Ethnicity: Thailand 18 to 55 years Inclusion criteria: patients with gingivitis >80% ppb of VSC Exclusion criteria: systemic complicating factors, oral mucosal lesions, smoker, den- ture wearers, antibiotic < 1month prior to the study 	60	 Study design: double blinded placebo controlled clinical trial Application: rinse twice with 15mL for 60s daily 30min after no beverage or food 	 Test : green tea hydroalcoholic brownie solution, contains green tea extract, propylene glycol, parabenes, Saccharin, mint flavor, 80% of total catechins Control: placebo (hydroalcoholic brownie solution of propylene glycol, parabenes, saccharin, mint flavor) Clinical measurements: baseline, 14d, 28d (endpoint) Study period: 28d Oral prophylaxis/instructions: routine oral hygiene, no other mouthwash, assigned toothbrush and toothpaste 	 PI plaque index (Silness& Löe 1964) VSC PBI adverse effects by questionnaire

First author (year of publication)	Population characteristics (ethnici- ty, age, inclusion crite- ria)	Number of subjects	Study design, Intervention	Green tea characteristics, clinical measure- ments, study period, oral prophylax- is/instructions	Analyzed parameters
Sarin (2015)	 Ethnicity: India 18 to 60 years Inclusion criteria: >20 natural teeth, mean Pl of at least 1.5, mean Gl of at least 1.0, no periodontal treatment for the last 3 month Exclusion criteria: periotontal pockets ≥5mm, antibiotics >3month, antimicrobial mounthrinse, smoking habits, periodontal therapy during the last 3 month 	110	 Study design: Triple-blinded, randomized controlled clinical trial Application: rinse with 10mL twice a day for 30s 	 Test : extract of leaves of <i>Camellia sinensis</i>, dried, crushed, brewed to attain required formulation (2%) of green tea mounthrinse Control group: placebo (destilled water coloured and lacked odor to match green tea mouthrinse) Clinical measurements: baseline, 28d (endpoint) Study period: 28d Oral prophylaxis/instructions: NA 	 PI plaque index (Turesky-Gilmore-Glickman, 1970)) GI gingival Index (Loe and Silness 1963) adverse effects by questionnaire, no adverse effects such as irritation, burning sensation, vesicle formation or mucosal disturbance.

Table 2a: Results of plaque index / test vs. CHX

Reference (year of publication)	Pl _{⊺est} Baseline	PI _{⊺est} End	Pl _{Control} Baseline	Pl _{Control} End	Intergroup ∆ PI _{Test}	Intergroup ∆ Pl _{Control}
Priya BM (2015)	2.20 ± 0.40	1.20± 0.30	2.19 ± 0.30	1.30 ± 0.20	1.00 ± 0.5	0.89 ± 0.36
Radafshar (2017)	1.61 ± 0.29	1.26 ± 0.23	1.72 ± 0.38	1.25 ± 0.27	0.35 ± 0.37	0.47 ± 0.47
Hambire (2015)	1.52 ± 0.04	0.56 ± 0.40	1.52 ± 0.05	0.64 ± 0.46	0.96 ± 0.40	0.88 ± 0.46

Table 2b: Results of plaque index / test vs. placebo

Reference (year of publication)	Pl _{⊺est} Baseline	PI _{⊺est} End	Pl _{Control} Baseline	PI _{Control} End	Intergroup ∆ PI _{Test}	Intergroup ∆ Pl _{Control}
Rassameemasmaung (2012)	1.29 ± 0.30	0.97± 0.24	1.17± 0.27	1.02 ± 0.25	0.32 ± 0.38	0.15 ± 0.36
Sarin (2015)	3.43 ± 0.99	1.77 ± 0.57	3.59 ± 1.01	3.46 ± 1.00	1.66 ± 1.14	0.13 ± 1.42

Table 3a: Results of gingiva index/ test vs. CHX

Reference (year of publication)	GI _{⊺est} Baseline	GI _{Test} End	GI _{Control} Baseline	GI _{Control} End	Intergroup ∆ GI _{Test}	Intergroup ∆ GI _{Control}
Priya BM (2015)	2.01 ± 0.40	1.43 ± 0.40	2.06 ± 0.10	1.53 ± 0.20	0.58 ± 0.57	0.53 ± 0.22
Radafshar (2017)	1.47 ± 0.14	1.16 ± 0.11	1.41 ± 0.10	1.14 ± 0.09	0.31 ± 0.18	0.30 ± 0.13
Hambire (2015)	2.34 ± 0.65	1.10 ± 0.50	2.68 ± 1.00	1.17 ± 0.45	1.24 ± 0.82	1.51 ± 1.09

Table 3b: Results of gingiva index/ test vs. placebo

Reference (year of publication)	GI _{Test} Baseline	GI _{⊺est} End	GI _{Control} Baseline	GI _{Control} End	Intergroup ∆ GI⊤est	Intergroup ∆ GI _{Control}
Rassameemasmaung (2012)	0.84 ± 0.24	0.76± 0.25	0.82 ± 0.35	0.73 ± 0.29	0.08 ± 0.35	0.09 ± 0.45
Sarin (2015)	1.50 ± 0.34	0.82 ± 0.24	1.47 ± 0.30	1.42 ± 0.35	0.68 ± 0.42	0.05 ± 46

Table 4 a. Meta-analysis PI Test vs. CHX: Endpoint

Author (year of publication)	PI _{Test} End	PI _{Control} End	Patients [n]	Study- period Mean d [days]	ifference $\Delta PI_{Control} - \Delta PI_{Test}$, <i>p</i> - value	95 % CI
Priya (2015)	1.20± 0.30	1.30 ± 0.20	30	28		0.10 [-0.08, 0.28]
Radafshar (2017)	1.26 ± 0.23	1.25 ± 0.27	40	28	⊢	-0.01 [-0.17, 0.15]
Hambire (2015)	0.56 ± 0.40	0.64 ± 0.46	40	14	••	0.08 [-0.19, 0.35]
				RE Model	0.43	0.04 [-0.06, 0.15]
	² = 0, SE = 0.00 =2), p-val = 0.63	99 , τ = 0 , I ² = 0%, I 92	-0.2 0 0.2 0.4			
			Observed Outcome			

Author (year of publication)	Mean difference Δ PI _{Test} [mm]	Mean difference ∆ Pl _{Control}	Patients [n]	Study- period [days]	Mean dif	ference $\Delta PI_{Control} - \Delta PI_{Test}$,	<i>p</i> - value	95 % CI
Priya (2015)	1.00 ± 0.10	0.89 ± 0.10	30	28		F		-0.11 [-0.42, 0.20]
Radafshar (2017)	0.35 ± 0.06	0.47 ± 0.11	40	28				0.12 [-0.14, 0.38]
Hambire (2015)	0.96 ± 0.36	0.88 ± NA	40	14				-0.08 [-0.35, 0.19]
				R	E Model		0.88	-0.01 [-0.17, 0.15]
Heterogeneity Q = 1.6088 (d	03 , τ = 0 , I ² = 0%, H 74	² = 1.00			-0.6 -0.2 0.2			
a					Observed Outcome			

Table 4 b. Meta-analysis PI Test vs, CHX: Change

Author (year of publication)	PI _{Test} End	PI _{Control} End	Patients [n]	Study- period [days]	Mean difference $\Delta PI_{Test} - \Delta PI_{Control}$,	<i>p</i> - value	95 % CI
Rassamee masmaung (2015)	0.97± 0.24	1.02 ± 0.25	60	28	a n a n an		0.05 [-0.07, 0.17]
Sarin (2015)	1.77 ± 0.57	3.46 ± 1.00	110	28			<u>1.69 [1.39, 1.99]</u>
				RE M	odel	0.29	0.86 [-0.75, 2.47]
	y τ² = 1.3308, SE = (df=1), p-val < 0.00	= 1.9018 , τ = 1.1536 001	, I ² = 98.96°	%, H ² = 95.7	4 -0.5 0.5 1 1.5 2 Observed Outcome		

Table 4c. Meta-analysis PI Test vs, Placebo: Endpoint

Table 4d. Meta-analysis PI Test vs, Placebo: Change

Author (year of publication)	Mean difference ΔPI_{Test}	Mean difference $\Delta \operatorname{Pl}_{\operatorname{Control}}$	Patients [n]	Study- period [days]	Mean dif	ference Δ PI _{Test} - Δ PI _{Co}	ontrol,	<i>p</i> - value	95 % CI
Rassamee masmaung (2015)	0.32 ± 0.06	0.15 ± 0.02	60	28		- 			-0.17 [-0.36, 0.02]
Sarin (2015)	1.66 ± 0.42	0.13 ± 0.01	110	28		·			<u>-1.53 [-2.01, -1.05]</u>
				RE M	Nodel			0.22	-0.83 [-2.16, 0.50]
Heterogeneity τ^2 = 0.8899, SE = 1.3079 , τ = 0.9433 , I² = 96.22%, H² = 26.46 Q = 26.4646 (df=1), p-val < 0.0001						-2.5 -1.5 -0.5	0.5		

Table 5a. Meta-analysis GI Test vs, CHX: Endpoint

Author (year of publication)	GI_{Test} End	GI _{Control} End	Patients [n]	Study- period [days]	Mean difference $\Delta \operatorname{GI}_{\operatorname{Test}}$ - $\Delta \operatorname{GI}_{\operatorname{Control}}$, <i>p</i> - value	95 % CI
Priya (2015)	1.43 ± 0.40	1.53 ± 0.20	30	28	<u>⊢</u>	0.10 [-0.13, 0.33]
Radafshar (2017)	1.16 ± 0.11	1.14 ± 0.09	40	28	⊢≣ -1	-0.02 [-0.08, 0.04]
Hambire (2015)	1.10 ± 0.50	1.17 ± 0.45	40	14		0.07 [-0.22, 0.36]
				RE Mode	0.78	-0.01 [-0.07, 0.05]
Heterogeneity $\tau^2 = 0$	0, SE = 0.0091 ,	$\tau = 0$, $I^2 = 0\%$, I	H ² = 1.00		-0.4 -0.2 0 0.2 0.4	
Q = 1.2862 (df=2),		. ,			Observed Outcome	

Author (year of publication)	Mean difference Δ GI _{Test}	Mean difference Δ GI _{Control}	Patients [n]	Study- period [days]	Mean difference $\Delta \operatorname{GI}_{\text{Test}}$ - $\Delta \operatorname{GI}_{\text{Control}}$,	<i>p</i> - value	95 % CI
Priya (2015)	0.58 ± 0.00	0.53 ± nr	30	28	<u> </u>		-0.05 [-0.36, 0.26]
Radafshar (2015)	0.31 ± 0.03	0.30 ± 0.01	40	28	• ∎•		-0.04 [-0.14, 0.06]
Hambire (2015)	1.24 ± 0.15	1.51 ± 0.55	40	14			0.27 [-0.33, 0.87]
				RE	Model	0.47	-0.03 [-0.13, 0.06]
Heterogeneity $\tau^2 = 0$, Q = 1.0105 (df=2), p-		= 0 , I ² = 0%, H ²	² = 1.00		–0.5 0 0.5 1 Observed Outcome		

Table 5b. Meta-analysis GI Test vs, CHX: Change

Table 5c. Meta-analysis GI Test vs, Placebo: Endpoint

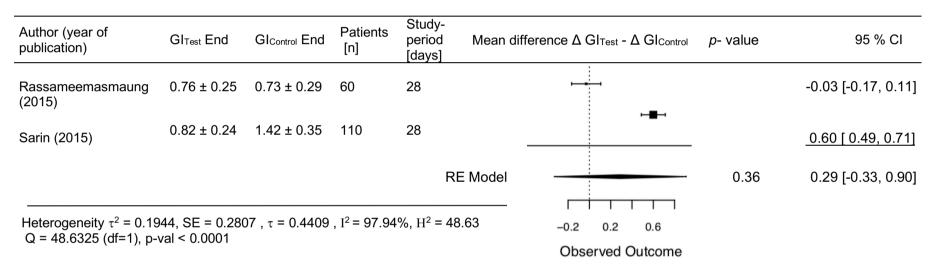


Table 5d. Meta-analysis GI Test vs, Placebo: Change

Author (year of publication)	Mean difference Δ GI _{Test}	Mean difference Δ GI _{Control}	Patients [n]	Study- period [days]	Mean difference ΔGI_{Test} - $\Delta GI_{\text{Control}}$	<i>p</i> - value	95 % CI
Rassameemasmaung (2015)	0.08 ± nr	0.09 ± 0.06	60	28			0.01 [-0.19, 0.21]
Sarin (2015)	0.68 ± 0.10	0.05 ± nr	110	28			<u>-0.63 [-0.79, -0.47]</u>
				REI	Model	0.33	-0.31 [-0.94, 0.31]
Heterogeneity $\tau^2 = 0.19$ Q = 22.8792 (df=1), p-v		6 , τ = 0.4425	-0.8 -0.4 0 0.4 Observed Outcome				

PRISMA Checklist

Section/topic	#	Checklist item	Reported on page #
TITLE			
Title	1	Identify the report as a systematic review, meta-analysis, or both.	1
ABSTRACT			
Structured summary	2	Provide a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings; systematic review registration number.	2
INTRODUCTION			
Rationale	3	Describe the rationale for the review in the context of what is	4, 5
Objectives	4	already known. Provide an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	6
METHODS			
Protocol and registration	5	Indicate if a review protocol exists, if and where it can be accessed (e.g., Web address), and, if available, provide registration information including registration number.	5
Eligibility criteria	6	Specify study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	6
Information sources	7	Describe all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	6
Search	8	Present full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	6
Study selection	9	State the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable,	6, 7
Data collection process	10	included in the meta-analysis). Describe method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	7
Data items	11	List and define all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	7, 8
Risk of bias in individual studies	12	Describe methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level), and how this information is to be used in any data synthesis.	7
Summary	13	State the principal summary measures (e.g., risk ratio,	7
measures Synthesis of results	14	difference in means). describe the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., l ²)	7,8
Risk of bias across studies	15	for each meta-analysis. Specify any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	7
Additional analyses	16	Describe methods of additional analyses (e.g., sensitivity or subgroup analyses, meta-regression), if done, indicating which were pre-specified.	7,8