

REVIEW

# Available online at www.sciencedirect.com

# ScienceDirect



# Biology, invasion and management of the agricultural invader: Fall armyworm, Spodoptera frugiperda (Lepidoptera: Noctuidae)

Jing WAN<sup>1\*</sup>, HUANG Cong<sup>2\*</sup>, LI Chang-you<sup>3</sup>, ZHOU Hong-xu<sup>3</sup>, REN Yong-lin<sup>1</sup>, LI Zai-yuan<sup>4</sup>, XING Longsheng<sup>2</sup>, ZHANG Bin<sup>3</sup>, QIAO Xi<sup>2</sup>, LIU Bo<sup>2</sup>, LIU Cong-hui<sup>2</sup>, XI Yu<sup>2</sup>, LIU Wan-xue<sup>5</sup>, WANG Wen-kai<sup>4</sup>, QIAN Wan-qiang<sup>2</sup>, Simon MCKIRDY<sup>1</sup>, WAN Fang-hao<sup>1, 2, 3, 5</sup>

<sup>1</sup> Harry Butler Institute, Murdoch University, Perth, WA 6150, Australia

#### Abstract

The fall armyworm (FAW), *Spodoptera frugiperda* (J. E. Smith), is native to the Americas. It has rapidly invaded 47 African countries and 18 Asian countries since the first detection of invasion into Nigeria and Ghana in 2016. It is regarded as a super pest based on its host range (at least 353 host plants), its inherent ability to survive in a wide range of habitats, its strong migration ability, high fecundity, rapid development of resistance to insecticides/viruses and its gluttonous characteristics. The inherently superior biological characteristics of FAW contribute to its invasiveness. Integrated pest management (IPM) of FAW has relied on multiple applications of monitoring and scouting, agricultural control, chemical pesticides, viral insecticides, sex attractants, bio-control agents (parasitoids, predators and entomopathogens) and botanicals. Knowledge gaps remain to be filled to: (1) understand the invasive mechanisms of *S. frugiperda*; (2) understand how to prevent its further spread and (3) provide better management strategies. This review summarizes the biological characters of FAW, their association with its invasiveness and IPM strategies, which may provide further insights for future management.

Keywords: invasive alien species, fall armyworm, *Spodoptera frugiperda*, biological invasions, prevention, management, biosecurity

Received 24 June, 2020 Accepted 25 July, 2020 Jing WAN, E-mail: Jing.Wan@murdoch.edu.au; HUANG Cong, E-mail: huangcong16@163.com; Correspondence WAN Fanghao, E-mail: wanfanghao@caas.cn; Simon MCKIRDY, E-mail: s.mckirdy@murdoch.edu.au

# 1. Introduction

Invasive alien species (IAS) seriously threaten agricultural and forestry ecosystems, biodiversity, human health, and cause significant economic losses. The emergence and invasion of IAS are closely linked with increasing trade and have become a major global issue. It is vital to effectively manage IAS (Wan and Yang 2016).

<sup>&</sup>lt;sup>2</sup> Shenzhen Branch, Guangdong Laboratory for Lingnan Modern Agriculture, Genome Analysis Laboratory of the Ministry of Agriculture, Agricultural Genomics Institute at Shenzhen, Chinese Academy of Agricultural Sciences, Shenzhen 518120, P.R.China

<sup>&</sup>lt;sup>3</sup> College of Plant Health and Medicine, Qingdao Agricultural University, Qingdao 266109, P.R.China

<sup>&</sup>lt;sup>4</sup> Hubei Engineering Research Center for Pest Forewarning and Management, Yangtze University, Jingzhou 434025, P.R.China

<sup>&</sup>lt;sup>5</sup> Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, P.R.China

<sup>&</sup>lt;sup>\*</sup> These authors contributed equally to this study.

<sup>© 2021</sup> CAAS. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http:// creativecommons.org/licenses/by-nc-nd/4.0/). doi: 10.1016/S2095-3119(20)63367-6

Fall armyworm (FAW), Spodoptera frugiperda (J. E. Smith) (Lepidoptera: Noctuidae), is native to tropical and sub-tropical areas of the Americas (Sparks 1979). FAW has a strong migration ability and in the past three years it has invaded 47 African countries, 18 Asian countries and now Australia where it seriously threatens crop production (https://www.cabi.org/isc/datasheet/29810). FAW is polyphagous and two sympatric host-plant strains have been identified, the "corn-strain" (C-strain) feeding mostly on maize, cotton and sorghum and the "rice-strain" (R-strain) mostly associated with rice and various pasture grasses (Nagoshi and Meagher 2004). In the past few decades, FAW has developed multiple resistance and cross-resistance mechanisms against various kinds of insecticides and transgenic Bacillus thuringiensis (Bt) maize, due to the extensive use of the treatments to manage the pest. The synthesis of these biological characteristics has contributed to its spread and invasion, and increased its economical importance. The cost of controlling FAW is enormous: according to statistics from the Food and Agriculture Organization (FAO), Brazil alone spends US\$600 million each year in attempts to control FAW (Wild 2017). Due to its perniciousness and invasiveness, it was rated as one of the top ten out of 1 187 arthropod pests by the Centre for Agriculture and Biosciences International (CABI) in the report "State of the World's Plants" in 2017 (Wild 2017).

## 2. Biological characteristics

IAS are superior in terms of their life cycle, genetics and evolution when compared with related non-invasive species. These characteristics are embodied in the morphology, physiology, ecology, genetics and behavior of the species. It is presumed that FAW invasiveness is associated with its superior biological characteristics including absence of diapause, short generation time, high fecundity, high polyphagy, long-distance migration ability and formidable resistance to insecticides, viruses and Bt toxin.

# 2.1. Absence of diapause, short generation time and high fecundity

FAW is a lepidopteran insect that undergoes holometabolous metamorphosis. Its life cycle includes egg (2–3 days), larvae (total six instars, 13–14 days), pupae (7–8 days) and adults (7–21 days). FAW has a generation time of approximately 30–40 days during the warm summer months (daily temperature of ~28°C), and approximately 55 days in cooler temperatures (Prasanna *et al.* 2018; Sharanabasappa *et al.* 2018). It does not have the ability to diapause, so the number of generations occurring in an endemic area depends on environmental conditions, e.g., temperatures

and host plants (Prasanna et al. 2018). In several regions of North America, FAW occurs seasonally through migration and it dies out in cold winter months. Whereas in the invaded countries, such as most of Africa, it occurs throughout the year with overlapping generations wherever host plants are available and climatic conditions are favorable (Abrahams et al. 2017). In southern China, it has been reported that FAW occurred all year round in the winter corn fields without diapause in winter (Qi et al. 2020), however, it could not survive when the average temperature was below 10°C for 8-10 days in Anhui Province (Xie et al. 2020). Average egg production per female is about 1 500 (a maximum of over 2000) in Africa, demonstrating high fecundity (Prasanna et al. 2018). However, the egg production in India (1064 eggs per female) and China (1052-1323 eggs per female when feeding on different maize varieties) are lower than that in Africa (Prasanna et al. 2018; Sun et al. 2020). Fecundity appears to be affected by variations in biotic (different hosts) and abiotic (temperature, humidity, etc.) factors.

#### 2.2. Highly polyphagous

FAW has a wide host range of more than 353 recorded plants from 76 families, principally Poaceae (106), Asteraceae (31) and Fabaceae (31). Among them, it has strong preference for maize, rice, sorghum, cotton, pasture grasses and sugarcane (Montezano et al. 2018; Dumas et al. 2015), which are all major cultivated crops in America, Africa and Asia. Remarkably, FAW has developed two defined strains, C-strain and R-strain, which are morphologically identical but differ in host range (Groot et al. 2010), mating behaviors (Schofl et al. 2009), genetics (Dumas et al. 2015) and pheromone components (Groot et al. 2010) in natural and laboratory maintained populations (Velasquez-Velez et al. 2011; Dumas et al. 2015). The asymmetric distribution of the two strains with selective plant host preference is consistently observed. The C-strain feeds predominantly on maize, cotton, and sorghum while the R-strain feeds primarily on rice and pasture grasses (Dumas et al. 2015). However, in Nagoshi et al. (2014), a small number of individuals of one strain were found in host habitats dominated by the other strain. In addition, previous laboratory studies indicated that both strains can exploit preferred hosts of the other strain, suggesting that host preference observed in the field cannot fully be explained by differential larval feeding (Groot et al. 2010). In addition, strain-specific female oviposition associated with host-preference has been observed under laboratory conditions (Hay-Roe et al. 2011).

#### 2.3. Long-distance migration ability

FAW displays high migratory ability (over 100 km per

night), through which the moths can find a broad range of habitats within its preferred environmental conditions (Tendeng et al. 2019). Laboratory testing has shown that 3-day-old moths have the strongest flight capacity and average flight distance, flight duration and flight velocity in 24 h can be 29.21 km, 11.00 h and 2.69 km h<sup>-1</sup>, respectively (Ge et al. 2019). In its native region, FAW populations can only overwinter in southern Texas and southern Florida, which are considered the northern-most winterbreeding areas available. However, in late summer, FAW are annually detected as far north as Ontario and Québec, Canada, which are considered to be migratory populations (Westbrook et al. 2016). The ability to migrate long distances has been confirmed by radar monitoring of noctuid moth species (including FAW) in Texas which identified a 400 km migratory flight displacement in 7.8 h (Westbrook 2008). In addition, in the Caribbean, the FAW can migrate from Puerto Rico to Barbados, a distance of more than 900 km (Nagoshi et al. 2017). In its invaded region in Africa and Asia, the spread of FAW also depends on its formidable flight capacity.

In China, FAW quickly invaded almost all maize belts within a year (Jiang *et al.* 2019). There are two main migratory routes for spread and reinvasion in China, the western and eastern routes. The origin of the western route is the westerly winter-breeding region (Myanmar/Yunnan, China) *via* Guizhou and Sichuan provinces through windborne transport. The origin of the eastern route is the easterly winter-breeding region (northern Thailand, Laos, Vietnam and Guangxi and Guangdong, China) *via* east-central China before arriving in the main maize belts (the Huang-Huai-Hai and Northeast Regions) with the help of Asian monsoons (Li *et al.* 2020).

#### 2.4. Formidable adaptability to adversity

FAW has developed high resistance to a range of insecticides. In the mid-1980s, it developed resistance to carbaryl, methyl parathion, trichlorfon and diazinon in the southeastern United States (Pitre 1986). Subsequently, it developed more than 200-fold resistance to organophosphates and carbamate successively in North, Central and South Florida (Yu 1991). In 2002, two field FAW populations in Citra and Gainesville, Florida showed high resistance to carbaryl (626- and 1159-fold), and moderate resistance to parathion-methyl (30- and 39-fold) (Yu and McCord 2007). In 2016, resistance ratios (RR) of various FAW populations from Mexico and Puerto Rico to chlorpyrifos, permethrin, flubendiamide, and chlorantraniliprole were up to 500-fold; RR to methomyl, cypermethrin and deltamethrin were 20- to 48-fold; and RR to ethyl dodecycin, dodecycin, emamectin benzoate and abamectin were 7- to 14-fold (Gutierrez-Moreno *et al.* 2019). By 2017, FAW had developed resistance to at least 29 insecticides, including carbamates, organophosphorus, pyrethroids and Bt insecticidal proteins in the Americas (https://www.pesticideresistance.org/). Currently, polyfungicide is the preferred pesticide for FAW control in corn fields and is applied three times during a growing season in Brazil. FAW has developed resistance to polyfungicide in Brazil, as well as to cyhalothrin, chlorpyrifos, and guanidine, and transgenic Bt maize Cry1F and Cry1Ab (Li Y P *et al.* 2019). Some studies have shown that the FAW population that invaded China carries resistance to organophosphate and pyrethroid pesticides (Zhang *et al.* 2020).

The transgenic insect resistant Bt maize has also been widely used to control FAW (Buntin *et al.* 2001). When exposed to Bt-maize toxin, FAW can develop monogenic (based on a single gene) or oligogenic (based on a few genes) resistance to these transgenic crop varieties (Huang *et al.* 2014). For example, Cry1F resistance in FAW has been detected in maize fields from Puerto Rico (Storer *et al.* 2010), southeastern mainland USA (Huang *et al.* 2014), Brazil (Farias *et al.* 2014), and Argentina (Chandrasena *et al.* 2018). Furthermore, it has developed resistance to other Bt proteins including Cry1Ac, Cry1Ab and Cry1A.105 (Storer *et al.* 2010).

Baculoviruses, the biological control agent, are widely used to control lepidopteran pests, and offers a promising alternative to chemical pesticides to avoid insecticidal resistance. However, several studies have now shown that FAW has developed resistance to the *S. frugiperda* multiple nucleopolyhedrovirus (SfMNPV) and *Autographa californica* multiple nucleopolyhedrovirus (AcMNPV) (Martinez *et al.* 2004; Haas-Stapleton *et al.* 2005).

# 3. Invasion mechanisms

Considering the abovementioned biological characteristics of FAW, and limited distribution in its native regions for more than two centuries, the sudden invasion and spread in Africa and Asia were unexpected. The fast rate of invasion has driven scientists to explore the underlying mechanisms. Genome research has greatly assisted in understanding the invasive mechanisms for alien species (Wan *et al.* 2019). With the development of genome sequencing, "Invasion Genomics" has increased investigations of invasion mechanisms as well as the prevention and management of IAS (Huang *et al.* 2019). In studies of FAW invasiveness, genomic data mining has improved our understanding of different biological characteristics and behaviors (Kakumani *et al.* 2014; Gouin *et al.* 2017; Nandakumar *et al.* 2017; Gimenez *et al.* 2020; Gui *et al.* 2020; Xiao *et al.* 2020; Zhang *et al.* 2020).

#### 3.1. High adaptability to diverse hosts

The expansion of gene families is generally believed to be associated with the invasiveness success of invasive species. That is especially the case for chemosensory and detoxification related gene families, which contribute to the polyphagy and adaptive evolution to host plants for the invasive species (Huang et al. 2019). For example, the gustatory receptor (GR) genes of invasive polyphagous moths Helicoverpa armigera, Spodoptera litura and Hyphantria cunea were significantly expanded compared to monophagous or oligophagous non-invasive moths (Cheng et al. 2017; Pearce et al. 2017; Wu N et al. 2018). The detoxification gene families cytochrome P450 (P450), UDP glucuronosyltransferase (UGT), glutathione S-transferase (GST) and carboxylesterase have expanded in the invasive species H. armigera, Bemisia tabaci and S. litura, which contribute to their adaptive success in diverse host plants (Chen et al. 2016; Cheng et al. 2017; Pearce et al. 2017). Similarly, the GR gene family has expanded dramatically in FAW, especially the recurrent tandem duplications of "bitter" receptors, compared with non-polyphagous lepidopteran species (Gouin et al. 2017; Xiao et al. 2020). In addition, P450 and GST were also expanded in genomes of both strains (Gouin et al. 2017; Gui et al. 2020). There are signatures of positive selection and copy number variation (CNV) in these genes involved in chemoreception, detoxification and digestion (Gouin et al. 2017; Gimenez et al. 2020).

#### 3.2. Insecticide resistance or tolerance

The resistance or tolerance mechanisms of FAW to insecticides compose two aspects: the detoxification metabolic mechanism and the target resistance mechanism (Zhang *et al.* 2020). The increased activity of detoxification metabolizing enzyme is an important reason for the FAW insecticide resistance (Yu *et al.* 2003). Therefore, some of the above detoxification related gene families, such as mixed function oxidases (MFO), GSTs, P450, esterases (ESTs), alkaline phosphatase, trypsin, aminopeptidase and chymotrypsin (Table 1) are also associated with insecticide resistance, which contribute to the invasiveness of FAW (McCord Jr and Yu 1987; Yu *et al.* 2003; Zhu *et al.* 2015).

Previous studies have indicated that the amino acid substitutions in acetylcholinesterase (AChE), VGSC and RyR result in resistance or tolerance to organophosphate, pyrethroid and diamide insecticides, respectively (Table 1). A genome-wide association study (GWAS) of 105 FAW samples from 16 provinces in China indicated that the single-nucleotide polymorphisms (SNPs) in AChE (AA201, AA290) contribute to its high risk of resistance to conventional pesticides and confirmed that the FAW population invading China is resistant to organophosphate and pyrethroid insecticides by scanning of resistance-related genes (Zhang *et al.* 2020) (Table 1).

#### 3.3. Bt crop resistance

The resistance mechanisms of target pests to Bt crops are composed of toxin activation, mutation of toxin receptor and regulation of the immune system (Xiao and Wu 2019). Until

Table 1 Mechanism of pesticide resistance or tolerance reported in the fall armyworm

Active ingredient	Resistance category	Mechanism of resistance or tolerance	Reference
Acetylcholinesterase (AC	ChE)		
Methyl-parathion	Detoxification metabolic	Increased activity of mixed function oxidases	McCord Jr and Yu (1987);
Carbaryl	Detoxification metabolic	(MFO), glutathione S-transferases (GSTs) and esterases (ESTs)	Yu <i>et al.</i> (2003)
Acephate	Detoxification metabolic	Increased activity of alkaline phosphatase, aminopeptidase, trypsin and chymotrypsin, P450, GSTs	Zhu <i>et al.</i> (2015)
Chlorpyrifos	Target resistance	Mutation of AChE	Carvalho et al. (2013);
		(A201S, G227A, F290V; AA201 and AA290)	Zhang <i>et al.</i> (2020)
Malathion	Target resistance	Mutation of AChE (AA201 and AA290)	Zhang <i>et al.</i> (2020)
Phoxim	Target resistance	Mutation of AChE (AA201 and AA290)	Zhang <i>et al.</i> (2020)
Ryanodine receptor (Ryl	R) allosteric modulator		
Chlorantraniliprole	Target resistance	Mutation of RyR (I4790M)	Boaventura et al. (2020)
Voltagegated sodium ch	annel (VGSC)		
Lambda-cyhalothrin	Target resistance	Mutation of VGSC	Carvalho <i>et al.</i> (2013);
		(T929I, L932F, L1014F)	Zhang et al. (2020)
Beta-cypermethrin	Target resistance	Mutation of VGSC (AA932)	Zhang <i>et al.</i> (2020)
Fenvalerate	Target resistance	Mutation of VGSC (AA932)	Zhang <i>et al.</i> (2020)
Deltamethrin	Target resistance	Mutation of VGSC (AA932)	Zhang et al. (2020)

recently, there was very little knowledge on the mechanism of FAW Bt-resistance. Some research showed that toxin activation and mutation of toxin receptors are associated with FAW resistance to Bt toxin proteins. The downregulated expression of Bt receptor alkaline phosphatase (ALP) in FAW populations was related to Cry1F resistance (Monnerat *et al.* 2015; Jakka *et al.* 2016). The mutation of ABCC2 (ATP-binding cassette sub-family C member 2), which is the receptor of both Cry1F and Cry1A.105, resulted in cross-resistance to Cry1F and Cry1A.105 (Flagel *et al.* 2018). The down-regulated expression of serine protease may reduce the FAW sensitivity to Cry1Ca1 toxin, which indicated that serine protease is involved in toxin activation (Rodriguez-Cabrera *et al.* 2010).

#### 3.4. Mechanism of migration and high fecundity

The comparative genomic analyses between FAW and the related species *S. litura*, showed that the elevated ratio of potential host adaptation genes were contributing to FAW invasiveness. Twenty-three of those invasiveness-related genes were under positive selection, including: 1) gustatory receptor (GR) and acetaldehyde oxidase, which contribute to host detection in invasion and expansion processes; 2) mitochondrial adenosine triphosphate synthase  $\beta$ -subunit and ferritin heavy chain, which contribute to long-distance migration during invasion and rapid expansion, due to enhanced locomotion and resistance; and 3) replacement in one site of chorion protein, which affects the protein function to maintain higher hatchability and ensure genetic resources for expansion (Cui *et al.* 2020).

## 4. Invasion, outbreak and damage

In the past two centuries, the distribution of FAW was limited to tropical and sub-tropical areas in the Americas, with several outbreaks at irregular intervals (Sparks 1979). However, in recent years, it has successfully invaded into Africa and Asia, and is in the process of invading Oceania (Fig. 1).

FAW was first detected in West and Central Africa in January 2016, and spread to the islands of São Tomé and Príncipe within 2 months (Goergen *et al.* 2016), followed by sudden outbreaks in 46 African countries including many countries in central, eastern and southern Africa (Fig. 1) (https://www.cabi.org/isc/datasheet/29810). Molecular identification of FAW showed that the invasive population in Africa includes both C-strain and R-strain (Assefa 2019). The invasion into India was first reported in May 2018 (Mahadevaswamy *et al.* 2018; Sharanabasappa *et al.* 2018), and then quickly spread to Sri Lanka, Thailand, Yemen, Nepal, Myanmar and Bangladesh (Farmer 2019).

Genetic diversity studies showed that the FAW population in India belongs to R-strain based on polymorphisms in the cytochrome oxidase subunit I (COI) gene (Mahadevaswamy et al. 2018) and triose-phosphate isomerase (Tpi). These findings suggest a small, shared founder population as the source of FAW in Africa and India (Nagoshi et al. 2019). FAW invaded Yunnan, China in December 2018 (Sun et al. 2021), spread rapidly and subsequently outbreaks were detected in 26 provinces (autonomous regions, municipalities) (Jiang et al. 2019). Both COI and Tpi showed that the invading populations in China were C-strain (Zhang et al. 2019). According to reports from the European and Mediterranean Plant Protection Organization (EPPO), FAW was first found in January 2020 on the islands of Saibai and Erub, in the Torres Strait and then at Bamaga, in the northern Queensland, Australia. By May of 2020, it had spread to 11 regions of Queensland, three regions of the Northern Territory, and three regions of Western Australia (https://gd.eppo.int/taxon/LAPHFR/distribution). In addition, Timor-Leste and Mauritania have also confirmed FAW in 2020 (http://www.fao.org/fall-armyworm/monitoring-tools/ faw-map/en/).

Maize yield losses have been estimated at 15 to 73% when infested with FAW (Hruska and Gould 1997). The annual economic losses in Ghana and Zambia have reached US\$177.3 million and US\$159.3 million, respectively (Abrahams *et al.* 2017). Collectively, maize, rice, sorghum and sugarcane, have suffered total economic crop losses of US\$13 billion per annum in sub-Saharan Africa (Abrahams *et al.* 2017). Estimation of the potential economic loss of maize in China caused by FAW indicates a range from US\$5.4–47 billion per annum (Qin *et al.* 2020).

#### 5. Prevention and management

#### 5.1. Monitoring and scouting in fields

For migratory invasive insects, monitoring and scouting are extremely important for timely responses to the dynamics of pest occurrence and development as well as crop health. This enables the formulation of comprehensive measures for better prevention and control. These actions must be taken based upon minimum cost parameters to keep the FAW population below the economic threshold level.

In China, entomological radar and vertical-pointing searchlight-traps have been used to monitor the population dynamics of migratory insects, such as *H. armigera* (Feng *et al.* 2009), *Cnaphalocrocis medinalis* (Fu *et al.* 2014), *Mythimna separata* (Zhao *et al.* 2009), *Loxostege sticticalis* (Feng *et al.* 2004), and *Spodoptera exigua* (Feng *et al.* 2003). The monitoring result of vertical-pointing searchlight-traps showed that the FAW population was first trapped in

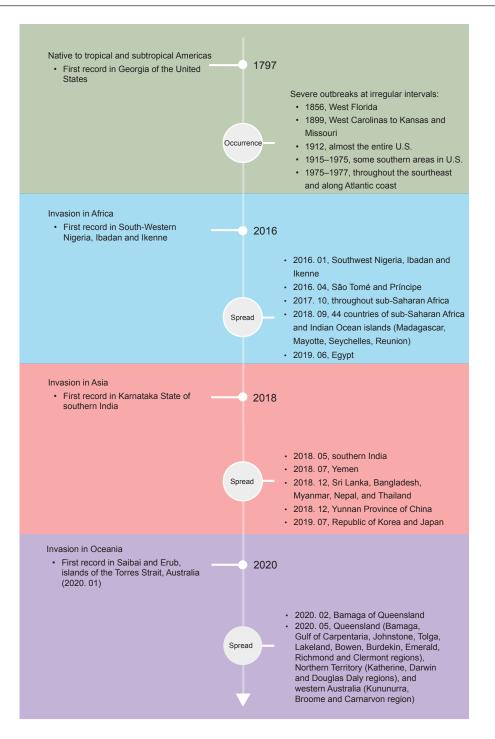


Fig. 1 Invasion and outbreak of fall armyworm (FAW) in its native and invaded regions.

June and the observation peaks appeared from August to October in eight provinces of China in 2019 (Jiang *et al.* 2020). The blacklight and commercial male traps are recommended to farmers to monitor the field population dynamics of FAW. It is recommended that the height of pheromone traps hanging should be 1.5 m above ground and the interval between two traps should be 50 m (Malo *et al.* 2013).

Farmers are recommended to scout the different plant growth stages and crop damage to determine the optimum stages for spraying insecticides based on action thresholds, which are expressed as percentages of plants with typical FAW damage/injury symptoms (Prasanna *et al.* 2018). For the early whorl stage, from vegetative emergence (VE) to 6-leaf (V6) stages, the action threshold is 10–30% of the seedlings infested as well as the tassel and silk stages, while it is 30–50% for the late whorl stage (Prasanna *et al.* 2018). For farmers, the method of scouting in the field involves randomly selecting five locations, or using a "W" or "Ladder" pattern, while avoiding edges (possibility of edge effects). Twenty plants should be examined for each location (Prasanna *et al.* 2018).

#### 5.2. Agricultural control

For smallholders, a series of low-cost agricultural control measures is an optimum option to implement as part of an effective IPM strategy against FAW. These agricultural approaches use the complex interactions between organisms and their environment to develop techniques to minimize the damage of pests on crops. In this review, a few agricultural measures that can be effective against FAW are discussed.

Traditional pre-planting, using some measures such as deep ploughing can decrease the FAW population in advance of sowing by exposing pupae to sunlight and predatory birds (Prasanna et al. 2018). Planting transgenic/ Bt insect-resistant maize varieties is also a very effective measure to decrease the damage by FAW and is an alternative method to pesticides. Bt maize is commonly used to control FAW, influencing the bioindicators of FAW including oviposition preference (Tellez-Rodriguez et al. 2014), larval dispersal (Malaquias et al. 2017), control efficacy (Horikoshi et al. 2016; Botha et al 2019) and fitness costs (Jakka et al. 2014). The use of transgenic maize expressing bacterial Bt proteins (e.g., Cry1F) has been commercially employed to control this pest since 2003 (Siebert et al. 2008). As stated above, in 2010, resistance of the FAW population to transgenic maize with Cry1F toxin was first detected in Puerto Rico (Storer et al. 2010). Developing new insecticidal targets is an urgent need due to the emergence of Bt-resistant FAW populations.

Other mechanical or physical methods are also recommended as management options, for smallholders, to reduce the economic loss caused by FAW, such as handpicking and crushing the egg masses and larvae, or using ash, sand, sawdust or dirt into whorls to desiccate young larvae (FAO 2018). The "push and pull" strategy is another very useful agroecological method to control agricultural pests such as FAW. Planting minor attractant plants or repellent plants in crop fields can decrease pest damage to major crops. Field experiments showed that maize intercropped with other plants helps to reduce the abundance of FAW. Compared with monocultured maize, intercropping with leguminous crops of bean (Phaseolus vulgaris), soybean (Glycine max) and groundnut (Vigna unguiculata) significantly decreases FAW attack by up to 40% (Hailu et al. 2018) (https://www.insectslimited.com/ history-of-pheromones). Some other plants, i.e., row intercropping with marigold (*Tagetes erecta*) and border intercropping with Napier grass (*Pennisetum purpureum*) have also been reported as effective for sustainable management of FAW (http://www.icipe.org/news/icipe-push-pull-technology-halts-fall-armyworm-rampage).

# 5.3. Divergence of sex pheromone and sex attractant application

Sex pheromones are applied worldwide for pest control as they present several advantages compared to traditional pesticides, such as nontoxicity, high specificity, and the possibility to apply minimal dosages. The first major pheromone component of FAW, (Z)-9-tetradecenyl acetate (Z9-14:OAc), was identified in 1967 (Sekul and Sparks 1967). Subsequently, other minor components were identified by analyzing the female pheromone glands and volatiles, including dodecyl acetate (12:OAc), (Z)-7dodecenyl acetate (Z7-12:OAc), 11-dodecenyl acetate (11-12:OAc), and (Z)-11-hexadecenyl acetate (Z11-16:OAc) (Tumlinson et al. 1986). The effectiveness of trapping in fields was first investigated in 1976 (Mitchell and Doolittle 1976). Since then, sex pheromones have been used to suppress and monitor FAW populations worldwide for more than 40 years and research has focused on investigating their differences and applications.

The practical effect of sex pheromones varies with geographical ranges and strains. The pheromone lures from North America and Europe were not effective against FAW in Brazil (Batista-Pereira et al. 2006), Costa Rica (Andrade et al. 2000) or Mexico (Malo et al. 2001). Some evidence points to geographic differences of the female sex pheromone blend in FAW (Batista-Pereira et al. 2006; Unbehend et al. 2014; Cruz-Esteban et al. 2018). For example, while females from Brazil (Batista-Pereira et al. 2006) produce (E)-7-dodecenyl acetate (E7-12:OAc), those from Florida, Louisiana or French Guyana do not (Tumlinson et al. 1986; Groot et al. 2008; Lima and McNeil 2009). For the Florida populations, the ratios of sex pheromone components from female glands were 4.9 (12:OAc):3.1 (Z7-12:OAc):1.7 (11-12:OAc):3.5 (Z11-16:OAc):86.9 (Z9-14:OAc). For the Brazilian populations, the sex pheromone consisted of Z7-12:OAc, E7-12:Ac, 12:OAc, (Z)-9-dodecenyl acetate (Z9-12:OAc), Z9-14:OAc, (Z)-10-tetradecenyl acetate (Z10-14:OAc), tetradecyl acetate (14:OAc)/(Z)-11-tetradecenyl acetate (Z11-14:OAc), Z11-16:OAc, and their relative proportions were 0.8:1.2:0.6:traces:82.8:0.3:1.5:12.9, respectively (Batista-Pereira et al. 2006). In addition, by adding E7-12:Ac to the major component Z9-14:OAc and critical secondary component Z7-12:OAc, the effectiveness of trapping for Brazilian populations was significantly improved (BatistaPereira et al. 2006).

Two groups have independently studied the strainspecific differences of the component concentration of sex pheromone in female FAW under laboratory and field environments. Both studies have shown that there are strain-specific differences in relative amounts of the different pheromone components (Groot et al. 2008; Lima and McNeil 2009; Unbehend et al. 2013). One group found a significantly higher relative amount of Z11-16:OAc, and lower relative amounts of Z7-12:OAc and Z9-12:OAc in the corn-strain females compared to rice-strain females in the Florida population (Groot et al. 2008). The other group found the opposite result with a significantly larger relative amount of Z9-14:OAc as well as lower relative amounts of Z7-12:OAc and Z11-16:OAc in corn-strain females compared to rice-strain females in the Louisiana population (Lima and McNeil 2009). These diametrically opposed results suggest that both geographic variation and strains contribute to the differentiation of sex pheromone composition of FAW females.

In China, the effects of four different commercial sex attractants on trapping FAW showed that the production of Shenzhen Bailebao Bio-Agricultural Technology Co., Ltd., was optimum to monitor the occurrence dynamic of FAW. The average number attracted by BLB lure was 137 individuals per trap, and the trapping performance of BLB lure was stable within the first 30 days. However, the numbers significantly decreased during the following 30 days, particularly after 50 days (Che *et al.* 2020).

#### 5.4. Chemical control

Chemical insecticides are heavily used to control FAW (Yu *et al.* 2003). Before the 1980s, insecticides, from organophosphates (methyl parathion, etc.), carbamates (carbaryl, etc.) to pyrethroids (cypermethrin, etc.), were the main method to control FAW in most countries in the Americas (Pitre 1986). Until recently, more than 57 active chemical ingredients with nine modes of action were used against FAW (Table 2). Among them, 47 active ingredients were used in the Americas in FAW native regions, while 34 and 20 active ingredients were respectively used in FAW invaded regions in Africa and Asia (Prasanna *et al.* 2018; Gutierrez-Moreno *et al.* 2019).

In the native regions in the Americas, FAW has developed resistance to more than 29 insecticides with six modes of action (Gutierrez-Moreno *et al.* 2019). Some insecticides are prohibited in the invaded regions of Africa and Asia, such as methomyl (Pitre 1988), thiodicarb (Gutierrez-Moreno *et al.* 2019), tralomethrin and fluvalinate (Leibee and Capinera 1995) (Table 2), due to the high resistance developed in FAW. To delay the development of insecticide

resistance, eight compound preparations (emamectin benzoate×indoxacarb, emamectin benzoate×hexaflumuron, emamectin benzoate×lambda-cyhalothrin, emamectin benzoate×chlorfenapyr, emamectin benzoate×lufenuron, emamectin benzoate×tebufenozide, lambda-cyhalothrin× chlorantraniliprole, and lambda-cyhalothrin×diflubenzuron) were recommended by the Ministry of Agriculture and Rural Affairs of China for emergency prevention and control of FAW as there are no currently legally registered insecticides for FAW.

Although 57 chemicals are listed in Table 2 that could be used against FAW, some of them are highly hazardous pesticides (HHPs) that are acknowledged to present particularly high levels of acute or chronic hazards to human health or the environment according to internationally accepted classification systems such as the World Health Organization (WHO) and the Globally Harmonized System of Classification and Labelling of Chemicals (GHS). If these HHPs are used, adequate precautions must be taken during application.

#### 5.5. Biological control

Biological control can reduce contamination of the environment and offer an economically and environmentally safer alternative to synthetic insecticides that are currently being used. Natural enemies include parasites, predators and entomopathogens. A great diversity of natural enemies of FAW has been reported in the Americas, Africa, and Asia (Molina-Ochoa et al. 2003; Prasanna et al. 2018; Shylesha et al. 2018). As the native regions for FAW, the Americas have the most abundant parasitoids (~150 taxa) against FAW, which have been recorded from 13 families, nine in Hymenoptera, and four in Diptera (Molina-Ochoa et al. 2003). Among them, the egg parasitoids (Trichogramma pretiosum, T. atopovirilia and Telenomus remus) (Beserra et al. 2005; Pomari et al. 2013), larval parasitoids (Campoletis sonorensis and Chelonus insularis) (Jourdie et al. 2009), and pupae parasitoids (Diapetimorpha introit and Ichneumon promissorius) (Molina-Ochoa et al. 2003) were widely used to control FAW. In Africa, eight parasitoids of FAW from three families were recovered in West, Central and East Africa, including Chelonus curvimaculatus, Chelonus cf maudae, Coccygidium luteum, Cotesia icipe, Cotesia sp., Charops ater, Charops sp., and Telenomus sp. Studies in southern India recorded five species of larval parasitoids: Coccygidium melleum, Campoletis chlorideae, Eriborus sp., Exorista sorbillans, and Odontepyris sp. (Sharanabasappa et al. 2019). In China, T. remus (Zhao et al. 2020), T. pretiosum (Zhu et al. 2020), T. dendrolimi (Tian et al. 2020), and T. chilonis (Li Z G et al. 2019) are the dominant parasitoids of FAW.

Active ingredient	Applied region	Active ingredient	Applied region		
Acetylcholinesterase (AChE) inhibitors		Nicotine acetylcholine receptor (nAChR) allosteric modulators			
Chlorpyrifos	America, Africa	Spinetoram	America, Asia		
Methomyl	America	Spinosad	America, Africa		
Thiodicarb	America	Acetamiprid	America, Africa, Asia		
Acephate	America, Africa, Asia	Cartap	Asia		
Carbaryl	America, Africa	Thiamethoxam	America, Africa, Asia		
Carbosulfan	Africa	Thiacloprid	America		
Trichlorfon	America	Imidacloprid	Africa		
Profenofos	Africa	Inhibitor of chitin biosynthesis			
Phenthoate	America	Triflumuron	America, Africa		
Methyl-parathion	Africa	Chlorfluazuron	America, Africa		
Methamidophos	America	Teflubenzuron	America		
Malathion	America, Africa	Novaluron	America		
Fenitrothion	America, Africa, Asia	Lufenuron	America, Africa, Asia		
Diazinon	America, Africa	Diflubenzuron	America, Asia		
Dimethoate	America, Africa	Hexaflumuron	Asia		
Sodium channel modulators		Ryanodine receptor (RyR) allo	osteric		
Permethrin	America, Africa	modulator			
Zeta-cypermethrin	Africa	Flubendiamide	America, Africa		
Deltamethrin	America, Africa, Asia	Chlorantraniliprole	America, Africa, Asia		
Alpha-cypermethrin	America, Africa	Cyantraniliprole	America, Africa, Asia		
Beta-cyfluthrin	America, Africa	Tetrachlorantraniliprole	Asia		
Beta-cypermethrin	America	Ecdysone agonists/moulting disruptors			
Bifenthrin	America, Africa	Chromafenozide	America		
Cyfluthrin	Asia	Tebufenozide	America, Asia		
Cypermethrin	America, Africa	Methoxyfenozide	America		
Fenpropathrin	America, Asia	Glutamate-gated chloride channel (GLUCL)			
Gamma-cyhalothrin	America	allosteric modulators			
Lambda-cyhalothrin	America, Africa, Asia	Emamectin benzoate	America, Africa, Asia		
Tralomethrin	America	Abamectin	Africa		
Pyrethrum	America, Africa	Uncouplers of oxidative phosp	Uncouplers of oxidative phosphorylation		
Fluvalinate	America	via disruption of proton gradie	nt		
Etofenprox	America	Chlorfenapyr	America, Africa, Asia		
Esfenvalerate	America, Africa	Voltage-dependent sodium ch	annel blockers		
		Indoxacarb	America, Africa, Asia		

Table 2 Chemical insecticides used against the fall armyworm

The presence of insect predators for both eggs and larvae is important to keep FAW populations under the control. The earwigs *Doru lineare* and *D. luteipes* prey on FAW eggs and larvae (Pasini *et al.* 2007; Sueldo *et al.* 2010) and the predators *Picromerus lewisi* and *Arma chinensis* mainly prey on 6th instar larvae of FAW (Tang *et al.* 2019a, b). Two species of predacious bugs, *Eocanthecona furcellata* (Wolff) and *Andrallus spinidens* (Fabr.) (Hemiptera: Pentatomidae) were found to effectively prey on FAW (Shylesha and Sravika 2018).

Several reviews have summarized the entomopathogen resources or potential biopesticide options of FAW and their application status (Bateman *et al.* 2018; Chen *et al.* 2019). Bt is a common biopesticide used to control pests including FAW. The soil bacterium *B. thuringiensis* produces multiple crystal (Cry) proteins or vegetative insecticidal proteins (Vip3A) that are toxic to FAW (Singh

*et al.* 2010). In addition, the fungi *Beauveria bassiana*, *B. rongniartii, Metarhizium anisopliae*, *M. rileyi, Nomuraea rileyi* and *Paecilomyces fumosoroseus* have been studied as potential entomopathogens for the control of FAW (Altre and Vandenberg 2001; Carneiro *et al.* 2008; Grijalba *et al.* 2018). The nematodes *Heterorhabditis* and *Steinernema* also effectively control FAW (Garcia *et al.* 2008).

#### 5.6. Viruses associated with S. frugiperda

SfMNPV is a member of the Group II *Alphabaculovirus* of the Baculoviridae family, which can cause FAW larval mortality rates of more than 90% (Castillejos *et al.* 2002; Simon *et al.* 2012). Different isolates of SfMNPV have been isolated in North, Central and South America (Berretta *et al.* 1998; Simon *et al.* 2012; Barrera *et al.* 2015). SfMNPV was first studied as a potential bioinsecticide for management of

FAW in 1999. Spraying with  $1.5 \times 10^{12}$  viral occlusion bodies (OBs) per ha caused approximately 40% mortality of FAW larvae at two days post application (Williams *et al.* 1999).

As a biological insecticide, the efficacy of SfMNPV and its speed of killing insects are affected by many factors, such as virulence of different isolates, larval instars, the amount of feeding viral OBs, formulation applied, and environmental conditions. Some studies indicated that diverse isolates had different efficacies: 3AP2 is a fastkilling isolate compared to the wild-type isolate Sf3, and the  $LT_{50}$  of the 3AP2 isolate was at least 30 h less than Sf3 when applied in the greenhouse and in the field (Behle and Popham 2012). There is a higher mortality of FAW and longer persistence on crop foliage caused by granular formulation than the aqueous spray application (Castillejos et al. 2002). To improve the efficacy of SfMNPV, a variety of SfMNPV formulations were produced for biological control of FAW. Recombinant baculoviruses containing two proteases with insecticidal activity decreased the time to kill insects, thus showing great potential for application in IPM programs (Gramkow et al. 2010). Nearly 90% of FAW was controlled by combining SfMNPV with 3 mg L<sup>-1</sup> Spinosad, which was 12.5–32% greater than the treatment with SfMNPV alone in a maize field (Mendez et al. 2002). Some studies indicated that microencapsulated SfMNPV also has the potential for improving FAW management (Gomez et al. 2013). A Colombian SfMNPV was microencapsulated by spray drying with a pH dependent polymer. Viral insecticide activity was not affected by microencapsulation, and the process provided effective protection from UVB radiation (Kurmen et al. 2015). Wettable powder formulations utilizing microencapsulation of SfMNPV OBs provide useful advantages related to half-life and photostability of viruses and retain the same efficacy under field conditions. In addition, adding 1% boric acid increased the mortality induced by the virus compared to application of granules containing virus alone in a field trial (Cisneros et al. 2002). Importantly, it was reported that a leading biopesticide company, Certis, USA, has obtained the license to develop and manufacture a commercial biopesticide product for field application in selected countries around the world based on Corpoica's SfMNPV strain NPV003 and formulation technology.

Spodoptera frugiperda granulovirus (SfGV) is a member of *Betabaculovirus* of the Baculoviridae family. A granulovirus of FAW in Columbia, South America, was first reported by Steinhaus (1957). SfGV attacks only the fat body, causes a proliferation of cells, and requires a relatively long time to produce mortality. One SfGV isolate was evaluated in a co-infection process and was demonstrated to enhance the insecticidal activity of *Lymantria dispar* NPV (Lepidoptera: Lymantriidae), reducing its mean lethal concentration by 13-fold (Shapiro 2000). Other studies obtained a similar synergic effect in co-infection of GV and NPV, due to the enhancins of baculovirus isolates (Hoover *et al.* 2010; Mukawa and Goto 2011). SfGV has been poorly studied compared to SfMNPV, with relatively few SfGV isolates being characterized.

#### 5.7. Botanicals for FAW management

Some plant derived-pesticides, referred to as botanicals, display good performance in insecticidal activity. They have diverse biological activities resulting in high mortality, extended larval duration, decreased pupa weight, insecticidal effects, growth inhibition, antifeedant effects, reduced fecundity, as well as sublethal and acute toxicity. Rioba and Stevenson (2020) have reviewed the opportunities and scope for botanical extracts and products for the management of FAW in Africa (Rioba and Stevenson 2020). They summarized the efficacy and potential of 69 plant species from 31 families including Azadirachta indica, Schinnus molle, and Phytolacca dodecandra. In China, Lin et al. (2020) estimated indoor toxicity and control effect of azadirachtin in a maize field for FAW. Azadirachtin has good toxicity and antifeedant activity on FAW, and the control effect reached a peak at seven days after treatment (Lin et al. 2020).

## 6. Future outlooks

FAW invaded Africa, Asia and Oceania extremely rapidly due to its strong flight capability, polyphagy, lack of diapause and quick development of insecticide/virus-resistance. Several biological characteristics associated with its invasiveness and the IPM strategies are summarized in this review, which provides some useful information for the future study of FAW. Furthermore, the five following aspects are worth studying not only for FAW, but also for all IAS.

#### 6.1. Improving monitoring by deep learning

Image recognition by deep learning presents good performance in monitoring alien invasive plants (Qiao *et al.* 2020). It provides researchers a new perspective to monitor IAS including invasive alien insects. Multiple Apps have emerged based on deep learning to identify FAW (Chiwamba *et al.* 2019a; Chulu *et al.* 2019). A system to automate FAW pheromone trapping has also been developed based on machine learning (Chiwamba *et al.* 2019b). There is a significant opportunity for researchers to further develop new monitoring techniques based on deep learning.

#### 6.2. Research on invasion mechanism

FAW has outbreaks with irregular intervals in its native

regions for two centuries, before its successful invasion in Africa and Asia. A similar phenomenon, which is called lag-time, has been found in other invasive species, such as the Brazilian pepper (*Schinus terebinthifolius*) which was present as a restricted ornamental for at least 50 years before its rapid invasion (Simberloff and Gibbons 2004). Thus, emerging questions as to why these invasive species have long invasion lag times, what facilitated invasions, and how many species are potential invasive need answering. There is a need to clarify the invasion mechanism to better prevent and control IAS including FAW. The flood of genomic data could provide opportunities for researchers to reveal the respective invasion mechanisms (Huang *et al.* 2019). to insecticides, viruses and other environment stresses (Wan *et al.* 2019). The resistance of FAW to transgenic crops, is considered by most researchers to be the result of pyramiding multiple transgenes (in the same plant). This is more effective in terms of FAW control and insect resistance management (IRM) than single-gene-based resistance (Huang *et al.* 2014; Horikoshi *et al.* 2016). Similarly, for resistance to insecticides, pesticides should be applied at the recommended rates, intervals, and seasonal totals according to instructions, which are designed to slow down the development of pesticide resistance for a FAW population (Prasanna *et al.* 2018). In addition, the IPM strategy shown in Table 3 can be used to achieve population control.

#### 6.3. Resistance management

One important reason for the successful invasion of invasive insects is their rapid development of resistance

#### 6.4. Development of new control techniques for IAS

In recent years, some new techniques have emerged for managing pests, including RNAi, CRISPR/Cas9, and

Table 3	Integrated	pest management measures f	for fall arm	worm, Spa	odoptera frugiperda

Management	Method <sup>1)</sup>	Pest stage	Corn growth period
Monitoring & Scouting			
Migration monitoring	Entomological radar, vertical-pointing searchlight-raps	Adult	Whole growth
Light traps	Blacklight	Adult	Whole growth
Pheromone traps	Commercial male trap, 50 m interval between two traps, traps hung at a height of 1.5 m above ground	Adult	Whole growth
Sampling	Random sampling of 20 plants in five locations	Egg and larva	Whorl stage
Agricultural control			
Insect-resistant corn	Transgenic/Bt maize varieties	Larva	Pre-planting
Cultural control	Intercropping with bean or sunflower; trap cropping with	Egg	Pre-planting
(push and pull)	castor plant or young corn plants		
Mechanical control	Handpicking egg masses and larvae	Egg and larva	Whorl stage
Physical control	Deep plowing to kill pupae in the soil/Placing sand or ash in the whorls	Larva and pupa	Pre-planting/Whorl stage
Biological control			
Enemy insects	Egg parasitoids: <i>Trichogramma pretiosum</i> and <i>Trichogramma atopovirilia</i> , etc.	Whole growth	
	Larval parasitoids: <i>Chelonus insularis</i> , Campoletis sonorensis and <i>Cotesia marginiventris</i> , etc.		
	Pupal parasitoids: Diapetimorpha introit and Ichneumon		
	promissorius Predators: Doru lineare and Podisus nigrispinus, etc.		
Dianastisidas	Virus: SfGV and SfMNPV	Larva	M/bala growth
Biopesticides	Fungus: Metarhizium anisopliae, Beauveria bassiana, seed	Laiva	Whole growth
	treatment with <i>Trichoderma</i> induces defense		
	Bacteria: Bacillus thuringiensis		
	Nematode: Heterorhabditis bacteriophora, and		
	Heterorhabditis indica, etc.		
	Botanical: Azadirachta indica, Schinnus molle, and		
	Phytolacca dodecandra, etc.		
Chemical control			
Sex attractants	Z7-12:Ac+E7-12:Ac+Z9-14:Ac (0.01:0.01:1.00 mg), or other efficient composite	Adult	Whole growth
Insecticides	A total of 20 insecticides were recommended by Ministry of Agriculture and Rural Affairs, China	Egg and larva	Pesticide sprays at VT (Vegetative–Tassel) stage afterward

<sup>1)</sup> SfGV, Spodoptera frugiperda granulovirus; SfMNPV, Spodoptera frugiperda multiple nucleopolyhedrovirus.

nanopesticides. One new technique combines RNAi and a nanocarrier to develop a novel, stable and safe strategy that may greatly improve pest management (Ma et al. 2020; Yan et al. 2020). For FAW, some scientists have focused on the potential of CRISPR/Cas9 in control programs. Wu K et al. (2018) explored the possibility of using the CRISPR/ Cas9 system to modify the abdominal-A (Sfabd-A) gene to explore new control strategies. Jin et al. (2021) generated a SfABCC2 knockout strain of FAW using the CRISPR/ Cas9 system to provide further functional evidence of the role of this gene in susceptibility and resistance to Cry1F (Jin et al. 2021). In addition, one study discussed the prospect of studying ORCO using CRISPR techniques in FAW. Due to the efficiency of targeting specific olfactory genes, it is possible to develop new alternative strategies using insecticides and/or microbial sprays to control FAW (Ayra-Pardo and Borras-Hidalgo 2019).

#### 6.5. Global collaboration for biosecurity

Invasive alien species, such as FAW, have serious negative consequences for the environment, economies, and human health and wellbeing, and with the accelerated development of global trade, these species have become a global issue (Bradshaw *et al.* 2016). There is a strong argument for strengthening global collaboration to improve individual country biosecurity defenses to prevent IAS invasions in order to protect food security, biodiversity and human health.

# Acknowledgements

This work was supported by the Australia-China Joint Center for the Prevention and Management of Exotic Invasive Species, the Harry Butler Institute, Murdoch University, WA, Australia and the Shenzhen Science and Technology Program, China (KQTD20180411143628272).

# **Declaration of competing interest**

The authors declare that they have no conflict of interest.

## References

- Abrahams P, Bateman M, Beale T, Clottey V, Cock M, Colmenarez Y, Corniani N, Day R, Early R, Godwin J, Gomez J, Moreno P G, Murphy S T, Oppong-Mensah B, Phiri N, Pratt C, Richards G, Silvestri S, Witt A. 2017. *Fall Armyworm: Impacts and Implications for Africa*. Commonwealth Agricultural Bureaux International, Wallingford.
- Altre J A, Vandenberg J D. 2001. Comparison of blastospores of two *Paecilomyces fumosoroseus* isolates: *in vitro* traits

and virulence when injected into fall armyworm, *Spodoptera frugiperda*. *Journal of Invertebrate Pathology*, **78**, 170–175.

- Andrade R, Rodriguez C, Oehlschlager A C. 2000. Optimization of a pheromone lure for *Spodoptera frugiperda* (Smith) in Central America. *Journal of the Brazilian Chemical Society*, **11**, 609–613.
- Assefa Y. 2019. Molecular identification of the invasive strain of Spodoptera frugiperda (JE smith) (Lepidoptera: Noctuidae) in Swaziland. International Journal of Tropical Insect Science, **39**, 73–78.
- Ayra-Pardo C, Borras-Hidalgo O. 2019. Fall armyworm (FAW; Lepidoptera: Noctuidae): Moth oviposition and crop protection. In: Picimbon J F, ed., Olfactory Concepts of Insect Control-Alternative to Insecticides. Springer, Switzerland. pp. 93–116.
- Banerjee R, Hasler J, Meagher R, Nagoshi R, Hietala L, Huang F, Narva K, Jurat-Fuentes J L. 2017. Mechanism and DNAbased detection of field-evolved resistance to transgenic Bt corn in fall armyworm (*Spodoptera frugiperda*). *Scientific Reports*, **7**, 10877.
- Barrera G P, Belaich M N, Patarroyo M A, Villamizar L F, Ghiringhelli P D. 2015. Evidence of recent interspecies horizontal gene transfer regarding nucleopolyhedrovirus infection of *Spodoptera frugiperda*. *BMC Genomics*, **16**, 1008.
- Bateman M L, Day R K, Luke B, Edgington S, Kuhlmann U, Cock M J W. 2018. Assessment of potential biopesticide options for managing fall armyworm (*Spodoptera frugiperda*) in Africa. *Journal of Applied Entomology*, **142**, 805–819.
- Batista-Pereira L G, Stein K, de Paula A F, Moreira J A, Cruz I, de Lourdes C Figueiredo M, Perri Jr J, Correa A G. 2006. Isolation, identification, synthesis, and field evaluation of the sex pheromone of the Brazilian population of *Spodoptera frugiperda*. *Journal of Chemical Ecology*, **32**, 1085–1099.
- Behle R W, Popham H J R. 2012. Laboratory and field evaluations of the efficacy of a fast-killing baculovirus isolate from *Spodoptera frugiperda*. *Journal of Invertebrate Pathology*, **109**, 194–200.
- Berretta M F, Rios M L, de Cap A S. 1998. Characterization of a nuclear polyhedrosis virus of *Spodoptera frugiperda* from Argentina. *Journal of Invertebrate Pathology*, **71**, 280–282.
- Beserra E B, Dias C T, Parra J R. 2005. Behavior of *Trichogramma atopovirilia* Oatman & Platner and *T. pretiosum* Riley (Hymenoptera: Trichogrammatidae) on *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) egg masses. *Brazilian Journal of Biology*, **65**, 9–17.
- Boaventura D, Bolzan A, Padovez F E O, Okuma D M, Omoto C, Nauen R. 2020. Detection of a ryanodine receptor target-site mutation in diamide insecticide resistant fall armyworm, Spodoptera frugiperda. Pest Management Science, **76**, 47–54.
- Botha A S, Erasmus A, du Plessis H, Van den Berg J. 2019. Efficacy of Bt maize for control of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in South Africa. *Journal of Economic Entomology*, **112**, 1260–1266.

- Bradshaw C J A, Leroy B, Bellard C, Roiz D, Albert C, Fournier A, Barbet-Massin M, Salles J M, Simard F, Courchamp F. 2016. Massive yet grossly underestimated global costs of invasive insects. *Nature Communications*, 7, 12986.
- Buntin G D, Lee R D, Wilson D M, McPherson R M. 2001. Evaluation of yieldgard transgenic resistance for control of fall armyworm and corn earworm (Lepidoptera : Noctuidae) on corn. *Florida Entomologist*, **84**, 37–42.
- Carneiro A A, Gomes E A, Guimaraes C T, Fernandes F T, Carneiro N P, Cruz I. 2008. Molecular characterization and pathogenicity of isolates of *Beauveria* spp. to fall armyworm. *Pesquisa Agropecuaria Brasileira*, **43**, 513–520.
- Carvalho R A, Omoto C, Field L M, Williamson M S, Bass C. 2013. Investigating the molecular mechanisms of organophosphate and pyrethroid resistance in the fall armyworm Spodoptera frugiperda. PLoS ONE, 8, e62268.
- Castillejos V, Trujillo J, Ortega L D, Santizo J A, Cisneros J, Penagos D I, Valle J, Williams T. 2002. Granular phagostimulant nucleopolyhedrovirus formulations for control of *Spodoptera frugiperda* in maize. *Biological Control*, 24, 300–310.
- Chandrasena D I, Signorini A M, Abratti G, Storer N P, Olaciregui M L, Alves A P, Pilcher C D. 2018. Characterization of fieldevolved resistance to *Bacillus thuringiensis*-derived Cry1F delta-endotoxin in *Spodoptera frugiperda* populations from Argentina. *Pest Management Science*, **74**, 746–754.
- Che J Y, Chen H, Chen Y M, Wang F L, Lu P, Zhu J P, Zhang H B, Xiao L B, Jiang C Y, Jin L, Zhu Z F, Zhang F. 2020. Trapping effect of four different sex attractants on *Spodoptera frugiperda. Plant Protection*, **46**, 261–266. (in Chinese)
- Chen W, Hasegawa D K, Kaur N, Kliot A, Pinheiro P V, Luan J, Stensmyr M C, Zheng Y, Liu W, Sun H, Xu Y, Luo Y, Kruse A, Yang X, Kontsedalov S, Lebedev G, Fisher T W, Nelson D R, Hunter W B, Brown J K, *et al.* 2016. The draft genome of whitefly *Bemisia tabaci* MEAM1, a global crop pest, provides novel insights into virus transmission, host adaptation, and insecticide resistance. *BMC Biology*, **14**, 110.
- Chen W B, Li Y Y, Wang M Q, Liu C X, Mao J J, Chen H Y, Zhang L S. 2019. Entomopathogen resources of fall armyworm *Spodoptera frugiperda*, and their application status. *Plant Protection*, **45**, 1–9. (in Chinese)
- Cheng T, Wu J, Wu Y, Chilukuri R V, Huang L, Yamamoto K, Feng L, Li W, Chen Z, Guo H, Liu J, Li S, Wang X, Peng L, Liu D, Guo Y, Fu B, Li Z, Liu C, Chen Y, *et al.* 2017. Genomic adaptation to polyphagy and insecticides in a major East Asian noctuid pest. *Nature Ecology & Evolution*, **1**, 1747–1756.
- Chiwamba S H, Phiri J, Nkunika P O Y, Nyirenda M, Kabemba M M, Sohati P H. 2019a. Machine learning algorithms for automated image capture and identification of fall armyworm (FAW) moths. *Zambia Information Communication Technology* (ICT) *Journal*, **3**, 1–4.
- Chiwamba S H, Phiri J, Nkunika P O Y, Sikasote C, Kabemba M M, Moonga M N. 2019b. Automated fall armyworm (*Spodoptera frugiperda*, J.E. Smith) pheromone trap based

on machine learning. *Journal of Computer Science*, **15**, 1759–1779.

- Chulu F, Phiri J, Nkunika P O Y, Nyirenda M, Kabemba M M, Sohati P H. 2019. A convolutional neural network for automatic identification and classification of fall armyworm moth. *International Journal of Advanced Computer Science and Applications*, **10**, 112–118.
- Cisneros J, Pérez J A, Penagos D I, Ruiz J, Goulson D, Caballero P, Cave R D, Williams T. 2002. Formulation of a nucleopolyhedrovirus with boric acid for control of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in maize. *Biological Control*, **23**, 87–95.
- Cruz-Esteban S, Rojas J C, Sanchez-Guillen D, Cruz-Lopez L, Malo E A. 2018. Geographic variation in pheromone component ratio and antennal responses, but not in attraction, to sex pheromones among fall armyworm populations infesting corn in Mexico. *Journal of Pest Science*, **91**, 973–983.
- Cui Y, Ren Y D, Lyu M, Zheng S C, Feng Q L, Xiang H. 2020. Genomic divergences between the two polyphagous *Spodoptera* relatives provide cues for successful invasion of the fall armyworm. *Insect Science*, **27**, 1257–1265.
- Dumas P, Legeai F, Lemaitre C, Scaon E, Orsucci M, Labadie K, Gimenez S, Clamens A L, Henri H, Vavre F, Aury J M, Fournier P, Kergoat G J, d'Alencon E. 2015. *Spodoptera frugiperda* (Lepidoptera: Noctuidae) host-plant variants: Two host strains or two distinct species? *Genetica*, **143**, 305–316.
- FAO. 2018. Integrated management of the fall armyworm on maize: a guide for farmer field schools in Africa. Food and Agriculture Organnization, Rome.
- Farias J R, Andow D A, Horikoshi R J, Sorgatto R J, Fresia P, dos Santos A C, Omoto C. 2014. Field-evolved resistance to Cry1F maize by *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in Brazil. *Crop Protection*, **64**, 150–158.
- Farmer B. 2019. Fall armyworm marches on as pest that devastated African crops spreads in Asia. [2019-01-09]. https://www.telegraph.co.uk/global-health/climate-andpeople/fall-armyworm-marches-pest-devastated-africancrops-spreads/
- Feng H Q, Wu K M, Cheng D F, Guo Y Y. 2003. Radar observations of the autumn migration of the beet armyworm *Spodoptera exigua* (Lepidoptera: Noctuidae) and other moths in northern China. *Bulletin of Entomological Research*, **93**, 115–124.
- Feng H Q, Wu K M, Cheng D F, Guo Y Y. 2004. Spring migration and summer dispersal of *Loxostege sticticalis* (Lepidoptera: Pyralidae) and other insects observed with radar in Northern China. *Environmental Entomology*, **33**, 1253–1265.
- Feng H Q, Wu X F, Wu B, Wu K M. 2009. Seasonal migration of *Helicoverpa armigera* (Lepidoptera: Noctuidae) over the Bohai Sea. *Journal of Economic Entomology*, **102**, 95–104.
- Flagel L, Lee Y W, Wanjugi H, Swarup S, Brown A, Wang J, Kraft E, Greenplate J, Simmons J, Adams N, Wang Y, Martinelli S, Haas J A, Gowda A, Head G. 2018. Mutational disruption of the ABCC2 gene in fall armyworm, Spodoptera

*frugiperda*, confers resistance to the Cry1Fa and Cry1A.105 insecticidal proteins. *Scientific Reports*, **8**, 7255.

- Fu X W, Li C, Feng H Q, Liu Z F, Chapman J W, Reynolds D R, Wu K M. 2014. Seasonal migration of *Cnaphalocrocis medinalis* (Lepidoptera: Crambidae) over the Bohai Sea in northern China. *Bulletin of Entomological Research*, **104**, 601–609.
- Garcia L C, Raetano C G, Leite L G. 2008. Application technology for the entomopathogenic nematodes *Heterorhabditis indica* and *Steinernema* sp. (Rhabditida: Heterorhabditidae and Steinernematidae) to control *Spodoptera frugiperda* (Smith) (Lepidoptera: Noctuidae) in corn. *Neotropical Entomology*, **37**, 305–311.
- Ge S S, He L M, He W, Xu R B, Sun X T, Wu K M. 2019. Determination on moth flight capacity of *Spodoptera frugiperda*. *Plant Protection*, **45**, 28–33. (in Chinese)
- Gimenez S, Abdelgaffar H, Goff G L, Hilliou F, Blanco C A, Hanniger S, Bretaudeau A, Legeai F, Negre N, Jurat-Fuentes J L, d'Alencon E, Nam K. 2020. Adaptation by copy number variation increases insecticide resistance in the fall armyworm. *Communications Biology*, **3**, 664.
- Goergen G, Kumar P L, Sankung S B, Togola A, Tamo M. 2016. First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (J E Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in West and Central Africa. *PLoS ONE*, **11**, e0165632.
- Gomez J, Guevara J, Cuartas P, Espinel C, Villamizar L. 2013. Microencapsulated Spodoptera frugiperda nucleopolyhedrovirus: Insecticidal activity and effect on arthropod populations in maize. *Biocontrol Science and Technology*, 23, 829–846.
- Gouin A, Bretaudeau A, Nam K, Gimenez S, Aury J M, Duvic B, Hilliou F, Durand N, Montagne N, Darboux I, Kuwar S, Chertemps T, Siaussat D, Bretschneider A, Mone Y, Ahn S J, Hanniger S, Grenet A G, Neunemann D, Maumus F, *et al.* 2017. Two genomes of highly polyphagous lepidopteran pests (*Spodoptera frugiperda*, Noctuidae) with different host-plant ranges. *Scientific Reports*, **7**, 11816.
- Gramkow A W, Perecmanis S, Sousa R L, Noronha E F, Felix C R, Nagata T, Ribeiro B M. 2010. Insecticidal activity of two proteases against *Spodoptera frugiperda* larvae infected with recombinant baculoviruses. *Virology Journal*, 7, 143.
- Grijalba E P, Espinel C, Cuartas P E, Chaparro M L, Villamizar L F. 2018. *Metarhizium rileyi* biopesticide to control *Spodoptera frugiperda*: stability and insecticidal activity under glasshouse conditions. *Fungal Biology*, **122**, 1069–1076.
- Groot A T, Marr M, Heckel D G, Schofl G. 2010. The roles and interactions of reproductive isolation mechanisms in fall armyworm (Lepidoptera: Noctuidae) host strains. *Ecological Entomology*, **35**, 105–118.
- Groot A T, Marr M, Schofl G, Lorenz S, Svatos A, Heckel D G. 2008. Host strain specific sex pheromone variation in *Spodoptera frugiperda. Frontiers in Zoology*, **5**, 20.
- Gui F R, Lan T M, Zhao Y, Guo W, Dong Y, Fang D M, Liu H, Li H M, Wang H L, Hao R S, Cheng X F, Li Y H, Yang P

C, Sahu S K, Chen Y P, Cheng L, He S Q, Liu P, Fan G Y, Lu H R, *et al.* 2020. Genomic and transcriptomic analysis unveils population evolution and development of pesticide resistance in fall armyworm *Spodoptera frugiperda*. *Protein* & *Cell*, doi: 10.1007/s13238-020-00795-7.

- Gutierrez-Moreno R, Mota-Sanchez D, Blanco C A, Whalon M E, Teran-Santofimio H, Rodriguez-Maciel J C, DiFonzo C. 2019. Field-evolved resistance of the fall armyworm (Lepidoptera: Noctuidae) to synthetic insecticides in Puerto Rico and Mexico. *Journal of Economic Entomology*, **112**, 792–802.
- Haas-Stapleton E J, Washburn J O, Volkman L E. 2005. Spodoptera frugiperda resistance to oral infection by Autographa californica multiple nucleopolyhedrovirus linked to aberrant occlusion-derived virus binding in the midgut. Journal of General Virology, 86, 1349–1355.
- Hailu G, Niassy S, Zeyaur K R, Ochatum N, Subramanian S. 2018. Maize-legume intercropping and push-pull for management of fall armyworm, stemborers, and striga in Uganda. *Agronomy Journal*, **110**, 2513–2522.
- Hay-Roe M M, Meagher R L, Nagoshi R N. 2011. Effects of cyanogenic plants on fitness in two host strains of the fall armyworm (*Spodoptera frugiperda*). *Journal of Chemical Ecology*, **37**, 1314–1322.
- Hoover K, Humphries M A, Gendron A R, Slavicek J M. 2010. Impact of viral enhancin genes on potency of *Lymantria dispar* multiple nucleopolyhedrovirus in *L. dispar* following disruption of the peritrophic matrix. *Journal of Invertebrate Pathology*, **104**, 150–152.
- Horikoshi R J, Bernardi D, Bernardi O, Malaquias J B, Okuma D M, Miraldo L L, Amaral F S, Omoto C. 2016. Effective dominance of resistance of *Spodoptera frugiperda* to Bt maize and cotton varieties: implications for resistance management. *Scientific Reports*, **6**, 34864.
- Hruska A J, Gould F. 1997. Fall armyworm (Lepidoptera: Noctuidae) and *Diatraea lineolata* (Lepidoptera: Pyralidae): Impact of larval population level and temporal occurrence on maize yield in Nicaragua. *Journal of Economic Entomology*, **90**, 611–622.
- Huang C, Li Y Z, Yang N W, Wu Q, Xing L S, Qian W Q, Xi Y, Li F, Wan F H. 2019. Progresses in invasive insect genomics. *Plant Protection*, **45**, 112–120. (in Chinese)
- Huang F, Qureshi J A, Meagher Jr R L, Reisig D D, Head G P, Andow D A, Ni X, Kerns D, Buntin G D, Niu Y, Yang F, Dangal V. 2014. Cry1F resistance in fall armyworm *Spodoptera frugiperda*: Single gene versus pyramided Bt maize. *PLoS ONE*, **9**, e112958.
- Jakka S R, Knight V R, Jurat-Fuentes J L. 2014. Fitness costs associated with field-evolved resistance to Bt maize in *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Journal* of Economic Entomology, **107**, 342–351.
- Jakka S R K, Gong L, Hasler J, Banerjee R, Sheets J J, Narva K, Blanco C A, Jurat-Fuentes J L. 2016. Field-evolved mode 1 resistance of the fall armyworm to transgenic Cry1Fa-expressing corn associated with reduced Cry1Fa toxin binding and midgut alkaline phosphatase expression.

Applied and Environmental Microbiology, **82**, 1023–1034.

- Jiang Y Y, Liu J, Xie M C, Li Y H, Yang J J, Zhang M L, Qiu K. 2019. Observation on law of diffusion damage of *Spodoptera frugiperda* in China in 2019. *Plant Protection*, **45**, 10–19. (in Chinese)
- Jiang Y Y, Liu J, Yang J J, Zhao W X, Yin L, Liu Y, Ye S F, Qin B Q, Song L D. 2020. Trapping effect of searchlight-trap and light trap for the moth of *Spodoptera frugiperda* in 2019. *Plant Protection*, **46**, 118–122. (in Chinese)
- Jin M H, Tao J H, Li Q, Cheng Y, Sun X X, Wu K M, Xiao Y T. 2021. Genome editing of the *SfABCC2* gene confers resistance to Cry1F toxin from *Bacillus thuringiensis* in *Spodoptera frugiperda. Journal of Integrative Agriculture*, **20**, 815–820.
- Jourdie V, Alvarez N, Turlings T C, Franck P. 2009. Isolation and characterization of polymorphic microsatellite loci in two primary parasitoids of the noctuid *Spodoptera frugiperda*: *Chelonus insularis* and *Campoletis sonorensis* (Hymenoptera). *Molecular Ecology Resources*, **9**, 171–173.
- Kakumani P K, Malhotra P, Mukherjee S K, Bhatnagar RK. 2014. A draft genome assembly of the army worm, *Spodoptera frugiperda. Genomics*, **104**, 134–143.
- Kurmen C, Elena J, Gómez Alvarez M I, Villamizar Rivero L F. 2015. Microencapsulation of a colombian *Spodoptera frugiperda* nucleopolyhedrovirus with Eudragit<sup>®</sup> S100 by spray drying. *Brazilian Archives of Biology and Technology*, 58, 468–476.
- Leibee G L, Capinera J L. 1995. Pesticide resistance in Florida insects limits management options. *Florida Entomologist*, 78, 386–399.
- Li X J, Wu M F, Ma J, Gao B Y, Wu Q L, Chen A D, Liu J, Jian Y Y, Zhai B P, Early R, Chapman J W, Hu G. 2020. Prediction of migratory routes of the invasive fall armyworm in eastern China using a trajectory analytical approach. *Pest Management Science*, **76**, 454–463.
- Li Y P, Zhang S, Wang X J, Xie X P, Liang P, Zhang L, Gu S H, Gao X W. 2019. Current status of insecticide resistance in *Spodoptera frugiperda* and strategies for its chemical control. *Plant Protection*, **45**, 14–19. (in Chinese)
- Li Z G, Lv X, Ya Y K, Xu Q Y, Ye J W, Han S C, Zhang C L, Li J. 2019. The parasitism of *Telenomus remus* and *Trichogramma chilonis* on *Spodoptera frugiperda* found in the fields of Guangzhou and Hong Kong. *Journal of Environmental Entomology*, **41**, 760–765.
- Lima E R, McNeil J N. 2009. Female sex pheromones in the host races and hybrids of the fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae). *Chemoecology*, **19**, 29–36.
- Lin S K, Liu K H, Wang R F, Liu B J, Zhang Y, Wu J Y Z, Cheng D M, Xu H H, Zhang Z X. 2020. Indoor toxicity of azadirachtin to Spodoptera frugiperda and its control effect in field. Journal of South China Agricultural University, 41, 22–27. (in Chinese)
- Ma Z Z, Zhou H, Wei Y L, Yan S, Shen J. 2020. A novel plasmid-Escherichia coli system produces large batch dsRNAs for insect gene silencing. *Pest Management Science*, **76**,

2505-2512.

- Mahadevaswamy H M, Asokan R, Kalleshwaraswamy C M, Sharanabasappa D, Prasad Y G, Maruthi M S, Shashank P R, Naorem I D, Surakasula A, Adarsha S, Srinivas A, Rao S, Sekhar V, Raju M S, Reddy G S S, Nagesh S N. 2018. Prevalence of 'R' strain and molecular diversity of fall army worm *Spodoptera frugiperda* (J.E.Smith) (Lepidoptera: Noctuidae) in India. *Indian Journal of Entomology*, **80**, 544–553.
- Malaquias J B, Godoy W A C, Garcia A G, Ramalho F S, Omoto C. 2017. Larval dispersal of *Spodoptera frugiperda* strains on Bt cotton: A model for understanding resistance evolution and consequences for its management. *Scientific Reports*, 7, 16109.
- Malo E A, Cruz-Lopez L, Valle-Mora J, Virgen A, Sanchez J A, Rojas J C. 2001. Evaluation of commercial pheromone lures and traps for monitoring male fall armyworm (Lepidoptera: Noctuidae) in the coastal region of Chiapas, Mexico. *Florida Entomologist*, 84, 659–664.
- Malo E A, Rojas J C, Gago R, Guerrero Á. 2013. Inhibition of the responses to sex pheromone of the fall armyworm, Spodoptera frugiperda. Journal of Insect Science, 13, 134.
- Martinez A M, Caballero P, Villanueva M, Miralles N, San Martin I, Lopez E, Williams T. 2004. Formulation with an optical brightener does not increase probability of developing resistance to Spodoptera frugiperda nucleopolyhedrovirus in the laboratory. *Journal of Economic Entomology*, **97**, 1202–1208.
- McCord Jr E, Yu S. 1987. The mechanisms of carbaryl resistance in the fall armyworm, *Spodoptera frugiperda* (JE Smith). *Pesticide Biochemistry and Physiology*, **27**, 114–122.
- Mendez W A, Valle J, Ibarra J E, Cisneros J, Penagos D I, Williams T. 2002. Spinosad and nucleopolyhedrovirus mixtures for control of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in maize. *Biological Control*, **25**, 195–206.
- Mitchell E, Doolittle R. 1976. Sex pheromones of Spodoptera exigua, S. eridania, and S. frugiperda: bioassay for field activity. Journal of Economic Entomology, 69, 324–326.
- Molina-Ochoa J, Carpenter J E, Heinrichs E A, Foster J E. 2003. Parasitoids and parasites of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Americas and Caribbean Basin: An inventory. *Florida Entomologist*, **86**, 254–289.
- Monnerat R, Martins E, Macedo C, Queiroz P, Praca L, Soares C M, Moreira H, Grisi I, Silva J, Soberon M, Bravo A. 2015. Evidence of field-evolved resistance of *Spodoptera frugiperda* to Bt corn expressing Cry1F in Brazil that is still sensitive to modified Bt toxins. *PLoS ONE*, **10**, e0119544.
- Montezano D G, Specht A, Sosa-Gómez D R, Roque-Specht V F, Sousa-Silva J C, Paula-Moraes S V D, Peterson J A, Hunt T. 2018. Host plants of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) in the Americas. *African Entomology*, 26, 286–300.
- Mukawa S, Goto C. 2011. Enhancing effect of proteins derived from Xestia c-nigrum granulovirus on Mamestra brassicae nucleopolyhedrovirus infection in larvae of Autographa

nigrisigna (Lepidoptera: Noctuidae) on cabbage. *Applied Entomology and Zoology*, **46**, 55–63.

- Nagoshi R N, Adamczyk J J, Meagher R L, Gore J, Jackson R. 2014. Using stable isotope analysis to examine fall armyworm (Lepidoptera: Noctuidae) host strains in a cotton habitat. *Journal of Economic Entomology*, **100**, 1569–1576.
- Nagoshi R N, Dhanani I, Asokan R, Mahadevaswamy H M, Kalleshwaraswamy C M, Sharanabasappa D, Meagher R L. 2019. Genetic characterization of fall armyworm infesting South Africa and India indicate recent introduction from a common source population. *PLoS ONE*, **14**, e0217755.
- Nagoshi R N, Fleischer S, Meagher R L, Hay-Roe M, Khan A, Murua M G, Silvie P, Vergara C, Westbrook J. 2017. Fall armyworm migration across the Lesser Antilles and the potential for genetic exchanges between North and South American populations. *PLoS ONE*, **12**, e0171743.
- Nagoshi R N, Meagher R L. 2004. Behavior and distribution of the two fall armyworm host strains in Florida. *Florida Entomologist*, **87**, 440–449.
- Nandakumar S, Ma H, Khan A S. 2017. Whole-genome sequence of the *Spodoptera frugiperda* Sf9 insect cell line. *Genome Announcements*, **5**, e00829–e00846.
- Pasini A, Parra J R, Lopes J M. 2007. Artificial diet for rearing Doru luteipes (Scudder) (Dermaptera: Forficulidae), a predator of the fall armyworm, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae). Neotropical Entomology, 36, 308–311.
- Pearce S L, Clarke D F, East P D, Elfekih S, Gordon K H J, Jermiin L S, McGaughran A, Oakeshott J G, Papanicolaou A, Perera O P, Rane R V, Richards S, Tay W T, Walsh T K, Anderson A, Anderson C J, Asgari S, Board P G, Bretschneider A, Campbell P M, *et al.* 2017. Genomic innovations, transcriptional plasticity and gene loss underlying the evolution and divergence of two highly polyphagous and invasive *Helicoverpa* pest species. *BMC Biology*, **15**, 63.
- Pitre H N. 1986. Chemical control of the fall armyworm (Lepidoptera: Noctuidae): An update. *Florida Entomologist*, **69**, 570–578.
- Pitre H N. 1988. Relationship of fall armyworm (Lepidoptera: Noctuidae) from Florida, Honduras, Jamaica, and Mississippi: Susceptibility to insecticides with reference to migration. *Florida Entomologist*, **71**, 56–61.
- Pomari A F, Bueno A D, Bueno R, Junior A, Fonseca A. 2013. Releasing number of *Telenomus remus* (Nixon) (Hymenoptera: Platygastridae) against *Spodoptera frugiperda* Smith (Lepidoptera: Noctuidae) in corn, cotton and soybean. *Ciencia Rural*, **43**, 377–382.
- Prasanna B, Huesing J, Eddy R, Peschke V. 2018. *Fall Armyworm in Africa: A Guide for Integrated Pest Management*. CAB International, Wallingford. pp. 1–109.
- Qi G J, Huang D C, Wang L, Zhang Y P, Xiao H X, Shi Q X, Xiao Y, Su X N, Huang S H, Zou S F, Chen K W, Zhou Z B, Zhong B Y, Zheng J J, Zhang Z X, Jiang T H, Lv L H, Lu Y Y, Zhang Z F. 2020. The occurrence characteristic in winter and year-round breeding region of the fall armyworm,

Spodoptera frugiperda (J. E. Smith) in Guangdong Province. Journal of Environmental Entomology, **42**, 573–582. (in Chinese)

- Qiao X, Li Y Z, Su G Y, Tian H K, Zhang S, Sun Z Y, Yang L, Wan F H, Qian W Q. 2020. MmNet: Identifying *Mikania micrantha* Kunth in the wild via a deep Convolutional Neural Network. *Journal of Integrative Agriculture*, **19**, 1292–1300.
- Qin Y J, Yamng D C, Kang D L, Zhao Z H, Zhao Z H, Yang P Y, Li Z H. 2020. Potential economic loss assessment of maize industry caused by fall armyworm (*Spodoptera frugiperda*) in China. *Plant Protection*, **46**, 69–73. (in Chinese)
- Rioba N B, Stevenson P C. 2020. Opportunities and scope for botanical extracts and products for the management of fall armyworm (*Spodoptera frugiperda*) for smallholders in Africa. *Plants*, **9**, 207.
- Rodriguez-Cabrera L, Trujillo-Bacallao D, Borras-Hidalgo O, Wright D J, Ayra-Pardo C. 2010. RNAi-mediated knockdown of a Spodoptera frugiperda trypsin-like serine-protease gene reduces susceptibility to a Bacillus thuringiensis Cry1Ca1 protoxin. Environmental Microbiology, **12**, 2894–2903.
- Schofl G, Heckel D G, Groot A T. 2009. Time-shifted reproductive behaviours among fall armyworm (Noctuidae: *Spodoptera frugiperda*) host strains: evidence for differing modes of inheritance. *Journal of Evolutionary Biology*, 22, 1447–1459.
- Sekul A A, Sparks A N. 1967. Sex pheromone of the fall armyworm moth: isolation, identification, and synthesis. *Journal of Economic Entomology*, **60**, 1270–1272.
- Shapiro M. 2000. Effect of two granulosis viruses on the activity of the gypsy moth (Lepidoptera: Lymantriidae) nuclear polyhedrosis virus. *Journal of Economic Entomology*, **93**, 1633–1637.
- Sharanabasappa D, Kalleshwaraswamy C M, Maruthi M S, Pavithra H B. 2018. Biology of invasive fall army worm *Spodoptera frugiperda* (J.E. Smith) (Lepidoptera: Noctuidae) on maize. *Indian Journal of Entomology*, **80**, 540–543.
- Sharanabasappa D, Kalleshwaraswamy C M, Poorani J, Maruthi M S, Pavithra H B. 2019. Natural enemies of Spodoptera frugiperda (J. E. Smith) (Lepidoptera: Noctuidae), a recent invasive pest on maize in South India. Florida Entomologist, 102, 619–623.
- Shylesha A N, Jalali S K, Gupta A, Varshney R, Venkatesan T, Shetty P, Ojha R, Ganiger P C, Navik O, Subaharan K, Bakthavatsalam N, Ballal C R, Raghavendra A. 2018.
  Studies on new invasive pest *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) and its natural enemies. *Journal of Biological Control*, **32**, 1–7.
- Shylesha A N, Sravika A. 2018. Natural occurrence of predatory bugs, *Eocanthecona furcellata* (Wolff) and *Andrallus spinidens* (Fabr.) on *Spodoptera frugiperd* (Smith) (Hemiptera: Pentatomidae) in maize and their potential in management of fall armyworm. *Journal of Biological Control*, **32**, 209–211.
- Siebert M W, Babock J M, Nolting S, Santos A C, Adamczyk J J, Neese P A, King J E, Jenkins J N, McCarty J, Lorenz

G M, Fromme D D, Lassiter R B. 2008. Efficacy of Cry1f insecticidal protein in maize and cotton for control of fall armyworm (Lepidoptera: Noctuidae). *Florida Entomologist*, **91**, 555–565.

- Simberloff D, Gibbons L. 2004. Now you see them, now you don't – population crashes of established introduced species. *Biological Invasions*, 6, 161–172.
- Simon O, Palma L, Williams T, Lopez-Ferber M, Caballero P. 2012. Analysis of a naturally-occurring deletion mutant of *Spodoptera frugiperda* multiple nucleopolyhedrovirus reveals *sf58* as a new *per os* infectivity factor of lepidopteraninfecting baculoviruses. *Journal of Invertebrate Pathology*, **109**, 117–126.
- Singh G, Sachdev B, Sharma N, Seth R, Bhatnagar R K. 2010. Interaction of *Bacillus thuringiensis* vegetative insecticidal protein with ribosomal S2 protein triggers larvicidal activity in *Spodoptera frugiperda*. *Applied and Environmental Microbiology*, **76**, 7202–7209.
- Sparks A N. 1979. A review of the biology of the fall armyworm. The Florida Entomologist, **62**, 82–87.
- Steinhaus E A. 1957. New records of insect-virus diseases. *Hilgardia*, **26**, 417–430.
- Storer N P, Babcock J M, Schlenz M, Meade T, Thompson G D, Bing J W, Huckaba R M. 2010. Discovery and characterization of field resistance to Bt maize: Spodoptera frugiperda (Lepidoptera: Noctuidae) in Puerto Rico. Journal of Economic Entomology, **103**, 1031–1038.
- Sueldo M R, Bruzzone O A, Virla E G. 2010. Characterization of the earwig, *Doru lineare*, as a predator of larvae of the fall armyworm, *Spodoptera frugiperda*: A functional response study. *Journal of Insect Science*, **10**, 38.
- Sun X X, Hu C X, Jia H R, Wu Q L, Shen X J, Zhao S Y, Jiang Y Y, Wu K M. 2021. Case study on the first immigration of fall armyworm, *Spodoptera frugiperda* invading into China. *Journal of Integrative Agriculture*, **20**, 664–672.
- Sun Y, Liu X G, Lu G Q, Hao X Z, Li S H, Li G P, Feng H Q. 2020. Comparison of population fitness of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) feeding on wheat and different varieties of maize. *Plant Protection*, **46**, 126–131. (in Chinese)
- Tang Y T, Li Y Y, Liu C X, Mao J J, Chen H Y, Zhang L S, Wang M Q. 2019a. Predation and behavior of *Arma chinensis* (Fallou) to *Spodoptera frugiperda* (J. E. Smith). *Plant Protection*, **45**, 65–68. (in Chinese)
- Tang Y T, Wang M Q, Chen H Y, Wang Y, Zhang H M, Chen F S, Zhao X Q, Zhang L S. 2019b. Predation and behavior of *Picromerus lewisi* Scott to *Spodoptera frugiperda* (J. E. Smith). *Chinese Journal of Biological Control*, **35**, 698–703. (in Chinese)
- Tellez-Rodriguez P, Raymond B, Moran-Bertot I, Rodriguez-Cabrera L, Wright D J, Borroto C G, Ayra-Pardo C. 2014. Strong oviposition preference for Bt over non-Bt maize in *Spodoptera frugiperda* and its implications for the evolution of resistance. *BMC Biology*, **12**, 48.
- Tendeng E, Labou B, Diatte M, Djiba S, Diarra K. 2019. The fall armyworm *Spodoptera frugiperda* (JE Smith), a new pest

of maize in Africa: Biology and first native natural enemies detected. *International Journal of Biological and Chemical Sciences*, **13**, 1011–1026.

- Tian J C, Lu Y H, Wang G R, Zheng X S, Yang Y J, Xu H X, Zang L S, Lv Z X. 2020. The parasitic capability of five *Trichogramma* species on eggs of fall armyworm *Spodoptera frugiperda*. *Chinese Journal of Biological Control*, **36**, 485–490. (in Chinese)
- Tumlinson J H, Mitchell E R, Teal P E, Heath R R, Mengelkoch L J. 1986. Sex pheromone of fall armyworm, *Spodoptera frugiperda* (J.E. Smith): Identification of components critical to attraction in the field. *Journal of Chemical Ecology*, **12**, 1909–1926.
- Unbehend M, Hanniger S, Meagher R L, Heckel D G, Groot A T. 2013. Pheromonal divergence between two strains of *Spodoptera frugiperda*. *Journal of Chemical Ecology*, **39**, 364–376.
- Unbehend M, Hanniger S, Vasquez G M, Juarez M L, Reisig D, McNeil J N, Meagher R L, Jenkins D A, Heckel D G, Groot A T. 2014. Geographic variation in sexual attraction of *Spodoptera frugiperda* corn- and rice-strain males to pheromone lures. *PLoS ONE*, **9**, e89255.
- Velasquez-Velez M I, Saldamando-Benjumea C I, Rios-Diez J D. 2011. Reproductive isolation between two populations of Spodoptera frugiperda (Lepidoptera: Noctuidae) collected in corn and rice fields from central Colombia. Annals of the Entomological Society of America, **104**, 826–833.
- Wan F H, Yin C L, Tang R, Chen M H, Wu Q, Huang C, Qian W Q, Rota-Stabelli O, Yang N W, Wang S P, Wang G R, Zhang G F, Guo J Y, Gu L Q, Chen L F, Xing L S, Xi Y, Liu F L, Lin K J, Guo M B, *et al.* 2019. A chromosome-level genome assembly of *Cydia pomonella* provides insights into chemical ecology and insecticide resistance. *Nature Communications*, **10**, 4237.
- Wan F H, Yang N W. 2016. Invasion and management of agricultural alien insects in China. Annual Review of Entomology, 61, 77–98.
- Westbrook J K. 2008. Noctuid migration in Texas within the nocturnal aeroecological boundary layer. *Integrative and Comparative Biology*, **48**, 99–106.
- Westbrook J K, Nagoshi R N, Meagher R L, Fleischer S J, Jairam S. 2016. Modeling seasonal migration of fall armyworm moths. *International Journal of Biometeorology*, **60**, 255–267.
- Wild S. 2017. African countries mobilize to battle invasive caterpillar. *Nature*, **543**, 13–14.
- Williams T, Goulson D, Caballero P, Cisneros J, Martinez A M, Chapman J W, Roman D X, Cave R D. 1999. Evaluation of a baculovirus bioinsecticide for small-scale maize growers in Latin America. *Biological Control*, **14**, 67–75.
- Wu K, Shirk P D, Taylor C E, Furlong R B, Shirk B D, Pinheiro D H, Siegfried B D. 2018. CRISPR/Cas9 mediated knockout of the abdominal — A homeotic gene in fall armyworm moth (Spodoptera frugiperda). PLoS ONE, **13**, e0208647.
- Wu N, Zhang S, Li X, Cao Y, Liu X, Wang Q, Liu Q, Liu H, Hu X, Zhou X J. 2018. Fall webworm genomes yield insights

into rapid adaptation of invasive species. *Nature Ecology* & *Evolution*, **3**, 105–115.

- Xiao H, Ye X, Xu H, Mei Y, Yang Y, Chen X, Yang Y, Liu T, Yu Y, Yang W, Lu Z, Li F. 2020. The genetic adaptations of fall armyworm *Spodoptera frugiperda* facilitated its rapid global dispersal and invasion. *Molecular Ecology Resources*, **20**, 1050–1068.
- Xiao Y T, Wu K M. 2019. Recent progress on the interaction between insects and *Bacillus thuringiensis* crops. *Philosophical Transactions of the Royal Society* (B: Biological Sciences), **374**, 20180316.
- Xie M H, Zhong Y Z, Chen H L, Lin L L, Zhang G L, Xu L N, Wang Z Y, Zhang J P, Zhang F, Su W H. 2020. Potential overwintering ability of fall armyworm *Spodoptera frugiperda* (J.E. Smith) in Anhui Province. *Plant Protection*, **46**, 236–241. (in Chinese)
- Yan S, Qian J, Cai C, Ma Z, Li J, Yin M, Ren B, Shen J. 2020. Spray method application of transdermal dsRNA delivery system for efficient gene silencing and pest control on soybean aphid *Aphis glycines*. *Journal of Pest Science*, **93**, 449–459.
- Yu S. 1991. Insecticide resistance in the fall armyworm, Spodoptera frugiperda (JE Smith). Pesticide Biochemistry and Physiology, 39, 84–91.
- Yu S J, McCord Jr E. 2007. Lack of cross-resistance to indoxacarb in insecticide-resistant Spodoptera frugiperda (Lepidoptera: Noctuidae) and Plutella xylostella (Lepidoptera: Yponomeutidae). Pest Management Science, 63, 63–67.
- Yu S J, Nguyen S N, Abo-Elghar G E. 2003. Biochemical characteristics of insecticide resistance in the fall armyworm, Spodoptera frugiperda (J.E. Smith). Pesticide Biochemistry

and Physiology, 77, 1–11.

- Zhang L, Jin M H, Zhang D D, Jiang Y Y, Liu J, Wu K M, Xiao Y T. 2019. Molecular identification of invasive fall armyworm *Spodoptera frugiperda* in Yunnan Province. *Plant Protection*, **45**, 19–24. (in Chinese)
- Zhang L, Liu B, Zheng W G, Liu C H, Zhang D D, Zhao S Y, Li Z Y, Xu P J, Wilson K, Withers A, Jones C M, Smith J A, Chipabika G, Kachigamba D L, Nam K, d'Alençon E, Liu B, Liang X Y, Jin M H, Wu C, *et al.* 2020. Genetic structure and insecticide resistance characteristics of fall armyworm populations invading China. *Molecular Ecology Resources*, **20**, 682–1696.
- Zhao X, Zhu K H, Zhang Z T, He K L, Zhang L S, Zhou J C, Dong H. 2020. Preliminary evaluation of the control efficacy of *Telenomus remus* against *Spodoptera frugiperda* in field conditions. *Plant Protection*, **46**, 74–77. (in Chinese)
- Zhao X C, Feng H Q, Wu B, Wu X F, Liu Z F, Wu K M, McNeil J N. 2009. Does the onset of sexual maturation terminate the expression of migratory behaviour in moths? A study of the oriental armyworm, *Mythimna separata*. *Journal of Insect Physiology*, **55**, 1039–1043.
- Zhu K H, Zhou J C, Zhang Z T, He K L, Zhang L S, Dong H. 2020. Preliminary evaluation of field competition between *Trichogramma pretiosum* and *Telenomus remus* in the control of *Spodoptera frugiperda*. *Plant Protection*, **46**, 267–271. (in Chinese)
- Zhu Y C, Blanco C A, Portilla M, Adamczyk J, Luttrell R, Huang F. 2015. Evidence of multiple/cross resistance to Bt and organophosphate insecticides in Puerto Rico population of the fall armyworm, *Spodoptera frugiperda*. *Pesticide Biochemistry and Physiology*, **122**, 15–21.

Managing Editor SUN Lu-juan