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Spa47 is an Oligomerization - Activated Type Three Secretion System (T3SS) ATPase from Shigella flexneri

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SPA47 IS AN OLIGOMERIZATION – ACTIVATED TYPE THREE SECRETION SYSTEM (T3SS) ATPASE FROM *SHIGELLA FLEXNERI*

by

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of

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in

Biochemistry
in the Department of Chemistry and Biochemistry

Approved:

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ABSTRACT:

Gram-negative pathogens often use conserved type three secretion systems (T3SS) for virulence. The Shigella type three secretion apparatus (T3SA) penetrates the host cell membrane and provides a unidirectional conduit for injection of effectors into host cells. The protein Spa47 localizes to the base of the apparatus and is speculated to be an ATPase that provides the energy for T3SA formation and secretion. Here, we developed an expression and purification protocol, producing active Spa47 and providing the first direct evidence that Spa47 is a bona fide ATPase. Additionally, size exclusion chromatography and analytical ultracentrifugation identified multiple oligomeric species of Spa47 with the largest greater than 8 fold more active for ATP hydrolysis than the monomer. An ATPase inactive Spa47 point mutant was then engineered by targeting a conserved Lysine within the predicted Walker A motif of Spa47. Interestingly, the mutant maintained a similar oligomerization pattern as active Spa47, but was unable to restore invasion phenotype when used to complement a spa47 null S. flexneri strain. Together, these results identify Spa47 as a Shigella T3SS ATPase and suggest that its activity is linked to oligomerization, perhaps as a regulatory mechanism as seen in some related pathogens. Additionally, Spa47 catalyzed ATP hydrolysis appears to be essential for host cell invasion, providing a strong platform for additional studies dissecting its role in virulence and providing an attractive target for anti-infective agents.

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INTRODUCTION

Shigella species cause a severe form of bacillary dysentery (shigellosis) throughout the world with an estimated 90 million annual infections responsible for greater than 100,000 deaths each year.[1] *Shigella* require as few as 10-100 organisms to cause infection and are efficiently spread through the fecal oral route and/or contaminated drinking water.[2] Most reported cases of shigellosis occur in developing regions where sanitation and access to clean drinking water are limited. Increasing reports in industrialized regions[3] and the appearance of antibiotic resistant strains further emphasize the need to better understand the mechanism of *Shigella* virulence and specifically the means by which it invades targeted host cells.[4]

Shigella invasion and subsequent spread throughout the colonic epithelium relies on a complex type three secretion system (T3SS) including effector proteins that are directly injected into host cell cytoplasm and the needle-like apparatus that facilitates effector transfer.[5] The type three secretion apparatus (T3SA) is highly conserved and resembles a nano-needle and syringe consisting of a cytoplasmic bulb, basal body that spans the inner and outer membranes of the bacterium, a hollow needle that extends beyond the thick LPS layer, and an associated tip complex that interacts with the host cell membrane (Figure 1).[6,7] Effector proteins and those that make up the T3SA tip complex are secreted through the 2.5 nm diameter channel of the MxiH needle both as the complex matures and upon activation of the

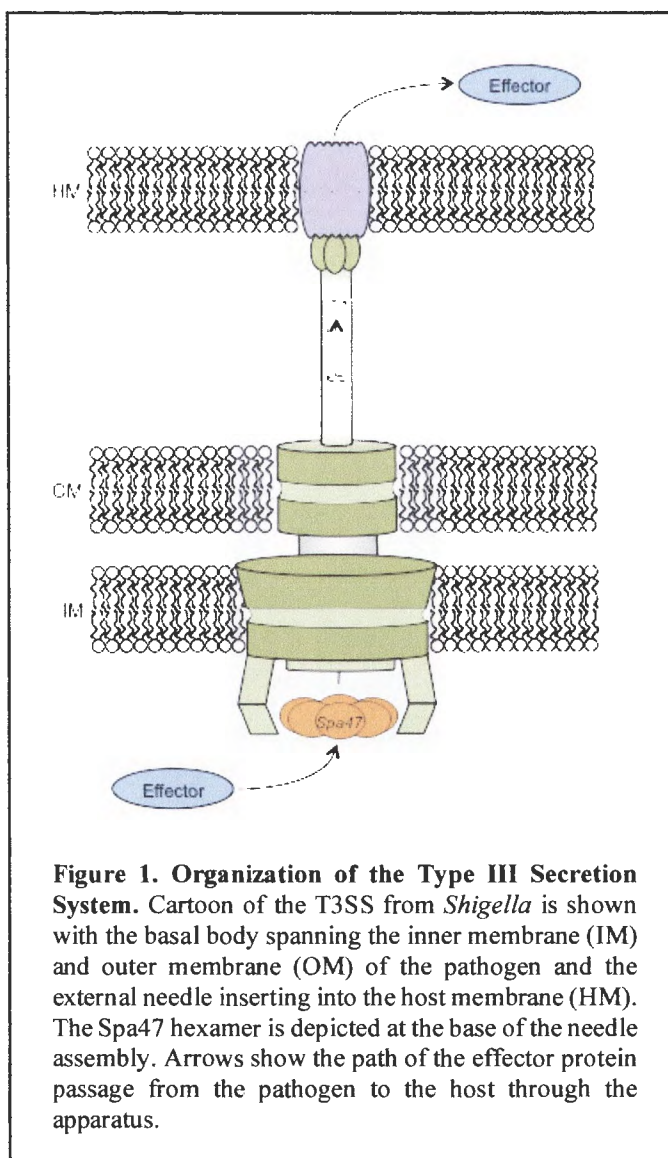**Figure 1. Organization of the Type III Secretion System.** Cartoon of the T3SS from *Shigella* is shown with the basal body spanning the inner membrane (IM) and outer membrane (OM) of the pathogen and the external needle inserting into the host membrane (HM). The Spa47 hexamer is depicted at the base of the needle assembly. Arrows show the path of the effector protein passage from the pathogen to the host through the apparatus.

system.[8-11] Proper timing and levels of secretion are essential for virulence, making identification of the trigger mechanism and driving force of secretion of great interest.[12,13] Sequence and structural similarities between the β-subunit of F_1F_o ATP synthase,[14] FliI of the bacterial flagellar system,[15] and identified ATPases associated with the translocon-associated T3SS apparatus of several pathogens[16-18] suggest that many T3SSs contain an ATPase at the base of the needle which could provide the necessary energy source for protein secretion.[19,20]

The *Shigella* T3SS protein Spa47 shares 37.4% sequence identity and 53.6% similarity to the *Salmonella enterica* ATPase FliI which drives hook rotation responsible for flagellar motion and even higher levels of homology to identified T3SS ATPases from *Escherichia*, *Yersinia*, and *Chlamydophila* species (40.1%, 42.0% and 43.4% identity to Spa47, respectively) as determined by the alignment software Clustal Omega. Additionally, Spa47 knockout strains of *S. flexneri* do not properly secrete some T3SS proteins including MxiH which is necessary for construction of the needle.[21] As a result, Spa47 is proposed to be a *Shigella* T3SS ATPase which may play a vital role in virulence by powering the T3SA machinery. Here, we describe the development of an expression and purification methodology for Spa47, allowing us to directly evaluate its ATPase activity *in vitro*. ATPase activity assays together with biophysical characterization of Spa47 found that the oligomerization state of the enzyme significantly impacts its activity, providing a potential mechanism for *in vivo* control over the associated T3SS. Specifically, the most active form identified purifies as a stable homo-oligomer while the monomeric species are 8-fold less active. A Spa47 point mutant was generated in the predicted catalytic Walker A domain to prevent ATP binding and provide an ATPase inactive Spa47 species. The oligomeric distribution of the mutant was similar to native Spa47 though it is unable to hydrolyze ATP and when used to complement a *spa47* knockout *Shigella* strain, is unable to restore the invasion phenotype. The overall findings identify Spa47 as an oligomerization-dependent active *Shigella* T3SS ATPase and suggest that its ability to hydrolyze ATP is key to the *Shigella* invasion phenotype and ultimately virulence.

MATERIALS AND METHODS

Materials

Wild-type (WT) *S. flexneri* corresponds to the serotype 2a 2457T strain originally isolated in 1954.[48] The *S. flexneri spa47* null strain was engineered by Abdelmounaaïm Allaoui as described in Jouihri et al.[21] *E. coli* strains and 2X ligation mix were from Novagen (Madison, WI). Restriction enzymes, the pTYB21 protein expression plasmid, PCR buffer, Phusion High-Fidelity polymerase, and chitin resin were purchased from New England Biolabs (Ipswich, MA). Oligonucleotide primers and the synthesized Spa47 gene were from Integrated DNA Technologies (Coralville, IA). The Superdex 16/600 size exclusion and 5mL Q FF columns were purchased from General Electric (Pittsburgh, PA). ATP was from Sigma-Aldrich (St. Louis, MO) and α-^{32}P-ATP was from Perkin Elmer (Boston, MA). DTT and ampicillin were from Gold Biotechnology (St. Louis, MO). All other solutions and chemicals were of reagent grade.

Methods

Cloning- The Spa47 gene was purchased as a single-stranded gBlock product from Integrated DNA Technologies. A silent mutation was created within the gene to remove the native internal SapI restriction site. A PstI restriction site was included at the 3′ end of the gene immediately following the native stop codon and a SapI site followed by the sequence "CAAC" was introduced 5′ of the native start codon. The additional sequence maintains the proper reading frame and provides an N-terminal Asparagine in Spa47 which aids in efficient intein cleavage during protein purification. Double-stranded product was generated by amplifying the gBlock using complementary primers containing 5′-GGTGGT sequences to aid in digestion (5′-GGTGGTTGCTCTTCCAACATGAGCTATACAAAATTGCT-3′ and 5′-GGTGGTCTGCAGTCATTATCTAATTGTTTCACCAATA-3′). The SapI/PstI digested gene was ligated into the expression plasmid pTYB21 which encodes an N-terminal chitin binding domain (CBD) and intein linker. The ligated product was transformed into *E. coli* Nova blue cells by heat shock and screened for the presence of *spa47* via PCR. Sequences were verified by Sanger sequencing (Genewiz, Inc., South Plainfield, NJ). *spa47* was cloned into the plasmid pWPsf4 by introducing NdeI and BamHI restriction sites at the 5′ and 3′ ends,

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respectively, and ligating into the digested vector backbone. Sequences were again verified by Sanger sequencing prior to transformation into electro-competent *S. flexneri* strains via electroporation. The Spa47^{K165A} point mutation was made in both pTYB21 and pWPsf4 using inverse PCR followed by sequence verification and transformation into *E. coli* and *S. flexneri*, respectively.

Protein expression and purification- Spa47 and *spa47^{K165A}* in pTYB21 were transformed into *E. coli* Tuner (DE3) cells and were grown overnight in LB Broth (Miller) containing 0.1 mg/mL ampicillin to ensure maintenance of the plasmid. 5 mL/L of the starter culture was added to Terrific Broth media containing 0.1 mg/mL ampicillin and was grown to an OD$_{600}$ of 0.8 at 37 °C, 200 RPM. The culture was then cooled to 17 °C before induction with 1 mM IPTG for approximately 20 hours (17 °C, 200 RPM). All subsequent steps were carried out at 4 °C. Typical growths included 3 liters of Spa47 expressing E. coli culture (1L per 2.8L baffled Fernbach flask) Cells were pelleted by centrifugation, resulting in typical cell pellet masses of approximately 27 g/L, resuspended in binding buffer (20 mM Tris, 500 mM NaCl, pH 7.9), and lysed by sonication. The suspension was clarified by centrifugation and the supernatant run over a chitin affinity column (10 mL resin per liter of cultured cells) at approximately 1 mL/min followed by washing with ~20 bed volumes of Binding Buffer. The chitin beads were quickly equilibrated with 3 bed volumes of cleavage buffer (20 mM Tris, 500 mM NaCl, 50 mM DTT, pH 7.9) and incubated overnight to cleave Spa47 from the chitin binding domain (CBD). Column elutions were collected three times per day until no further elution of Spa47 was observed by SDS-PAGE (~4 days). Collected elutions were pooled and diluted to generate final buffer concentrations of 20 mM Tris, 100 mM NaCl, 5 mM DTT, pH 7.9 and further purified over a 5 mL Q Sepharose FF anion exchange column, consistently resulting in ~10 mg of Spa47 per liter of expression culture. The Q column provided a negative selection and the purified Spa47 in the flow through was concentrated using an Amicon Ultra centrifugal filter unit with a 30kDa molecular weight cut off and further purified/characterized using a Superdex 16/600 size exclusion column equilibrated with 20 mM Tris, 100 mM NaCl, 5 mM DTT, pH 7.9. Spa47 contains no Tryptophan residues, resulting in a low molar A$_{280}$ extinction coefficient that is below the limits of quantitation for the

concentrations of Spa47 obtained and used throughout the study. All Spa47 concentrations were determined using a more sensitive Bradford protein assay with bovine serum albumin as a standard after verifying that comparison to a BSA standard curve provides accurate Spa47 quantitation results. All Spa47 concentrations are reported in monomer concentration units for consistency and clarity.

ATP hydrolysis activity assays- A single time point activity assay was used to observe ATP hydrolysis from collected size exclusion chromatography (SEC) fractions of Spa47 and Spa47^{K165A}. The reactions were initiated by combining protein from the SEC fractions with a prepared ATP solution resulting in a final concentration of 0.5 mM ATP, 10 mM MgCl$_2$, and 0.5 μCi (~300 nM) α-^{32}P-ATP. The reactions were allowed to proceed for 15 minutes before quenching with the addition of EDTA to a final concentration of 0.25 M. The level of ATP hydrolysis was quantified by first spotting 1 μL of the quenched reaction onto a TLC plate and developing in 0.6 M potassium phosphate buffer (pH 3.4) for 70 min. The α-^{32}P-ATP and the α-^{32}P-ADP were detected with a Storm PhosphorImager (Molecular Dynamics) and quantified using associated ImageQuant software (Molecular Dynamics). A multiple time point activity assay was used to determine apparent k_{cat} values for Spa47 species and was carried out under similar conditions to the single time point assay described above. Specifically, the purified Spa47 concentration was held constant at 1.35 μM for each species, the ATP concentration was increased to 1 mM, and samples were removed from the reaction every 30 sec for 3.5 minutes and rapidly quenched with 0.25 M EDTA prior to separation via TLC.

Far-UV Circular Dichroism (CD)- Far-UV CD spectra and thermal stability melts were obtained for isolated monomer (I) and oligomer fractions (III) of Spa47 and Spa47^{K165A}. Measurements were obtained using a JASCO model J-1500 spectropolarimeter with a Peltier temperature controller (Jasco, Easton, MD). Spectra were obtained from 200 to 260 nm at 10 °C with a 0.1 cm path length, 0.5 nm spectral resolution, 50 nm/min scan rate and a 1 sec data integration time. Secondary structure thermal stability profiles were generated by monitoring CD signal at 222 nm while the temperature was increased from 10

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to 90 °C at 0.3 °C/min. All measurements were performed on 0.3 mg/mL protein concentration. CD signals were converted to mean residue molar ellipticity.

Sedimentation velocity analytical ultracentrifugation- Sedimentation velocity analytical ultracentrifugation (SV-AUC) experiments were conducted using an Optima XL-I (Beckman Coulter, Fullerton, CA) analytical ultracentrifuge equipped with an interference optical detection system. 1.5 mg/ml Spa47 and a buffer reference were loaded into Beckman charcoal-epon two sector cells with 12 mm path lengths and sapphire windows. The samples were analyzed at 20 °C and 40,000 RPM, using interference detection and scanning until complete sedimentation was achieved. The data were regularized with a confidence interval of 0.95 and analyzed using the software program Sedfit[49] with a continuous c(s) distribution and 500 scans. The Spa47 partial specific volume (0.7422 mL/g) and the buffer density and viscosity used in the analysis (1.0031 g/mL and 0.01018 Poise respectively) were calculated using Sednterp.[50] Additional SV-AUC distributions were collected for Spa47 concentrations of 1.0, 0.66, and 0.1 mg/mL Spa47 to ensure that the assigned peaks are the result of discrete species and not an artifact resulting from a reaction boundary.

Bacterial invasion of epithelial cells-S. flexneri invasion of cultured HeLa cells was monitored by a gentamicin protection assay as previously described.[51] Sterile 24-well plates were seeded with passaged HeLa cells and grown overnight in DMEM supplemented with 10% fetal calf serum, penicillin, and streptomycin at 100% relative humidity, 37 °C, and 5% CO_2. Tested *S. flexneri* strains were streaked onto tryptic soy agar plates containing 0.025% Congo red and grown overnight at 37 °C. Small cultures containing appropriate antibiotics to maintain the transformed plasmid were inoculated from the agar plates and grown to an OD_{600} ~0.4 at 37 °C and 200 RPM. Equivalent bacterial loads were then introduced to the cultured HeLa cells as described previously.[51] The plates were centrifuged at 1000 x g to synchronize contact between the bacteria and HeLa cells, incubated at 37 °C for 30 min, and rinsed to remove most of the extracellular bacteria. *Shigella* that had not successfully invaded the HeLa cells were killed by treatment

with 50 μg/mL gentamicin. Bacteria that had invaded were visualized by lysing the host cells with 1% agarose in water and overlaying with a 2x LB agar solution. Overnight incubation at 37 °C resulted in colony formation that is quantified and used to provide relative levels of invasiveness between the tested *S. flexneri* strains.

RESULTS

Expression and purification of an active S. flexneri T3SS ATPase- T3SS ATPases from several organisms have been successfully expressed and purified, allowing them to be studied both structurally and biochemically.[16,22-26] While these findings have been key in understanding the role of ATPases in T3SSs, work with *Shigella* Spa47 has proven challenging and has limited the understanding of its role in the otherwise well-defined *Shigella* system. Here, we have overcome the hurdle of Spa47 purification by generating a CBD-tagged Spa47 fusion, resulting in *E. coli* expression of soluble Spa47 containing an N-terminal CBD attached through an intein linker

Figure 2. Purification of Spa47. (A) N-terminal chitin binding domain (CBD) and Intein tagged version of Spa47 used for overexpression and purification. The site of cleavage is shown. (B) SDS-PAGE of Spa47 at various stages during protein purification. Whole cell lysate (WC), clarified supernatant (Sup), cleaved protein after elution from the chitin column (Chitin) and, purified Spa47 from the Q-sepharose (Q) are shown. ** and * denote the corresponding positions of the uncleaved CBD-Intein-Spa47 and cleaved Spa47, respectively.

(Figure 2A). Affinity purification with chitin resin followed by DTT-induced intein cleavage produced CBD-less Spa47 at relatively high purity (Figure 2B). Negative selection using Q Sepharose anion exchange chromatography further removed contaminants and resulted in highly soluble full length recombinant
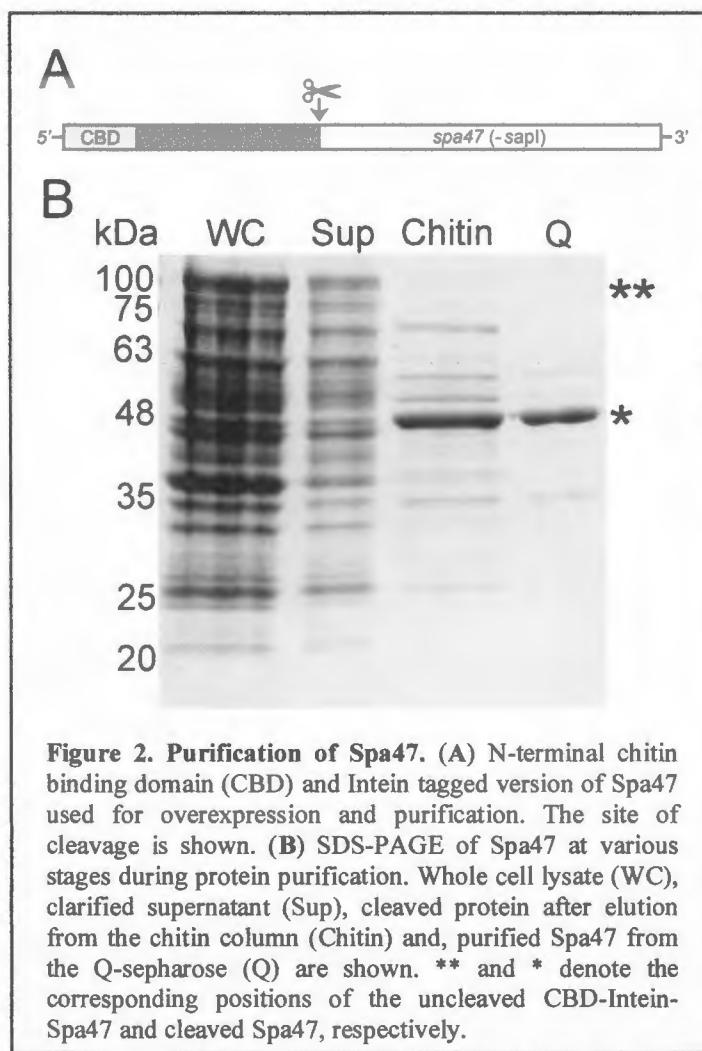
Spa47 that was ≥ 85% pure (by SDS PAGE) with identity confirmed by in gel digestion followed by tandem mass spectrometry.

Initial purification was followed by SEC to provide an additional purification step and as a means to characterize the oligomerization state of the recombinant Spa47. The elution profile identified multiple discrete species of Spa47 with the majority of the protein eluting later in the separation, suggesting that the recombinant Spa47 purified in multiple oligomeric states with the main peaks eluting at ~0.4 and 0.6 column volumes, representing species III and I, respectively (Figure 3A and B). While we were not expecting such a distribution, ATPase oligomerization is not uncommon and is known to be essential for activity of the hetero-hexameric F_1F_o ATP synthase[27] as well as suspected to play an important role in activation and control of hydrolysis for several T3SS ATPases,[22,25,26,28] providing a potential mechanism for regulation. We next tested if the purified Spa47 was active with respect to ATP hydrolysis which would not
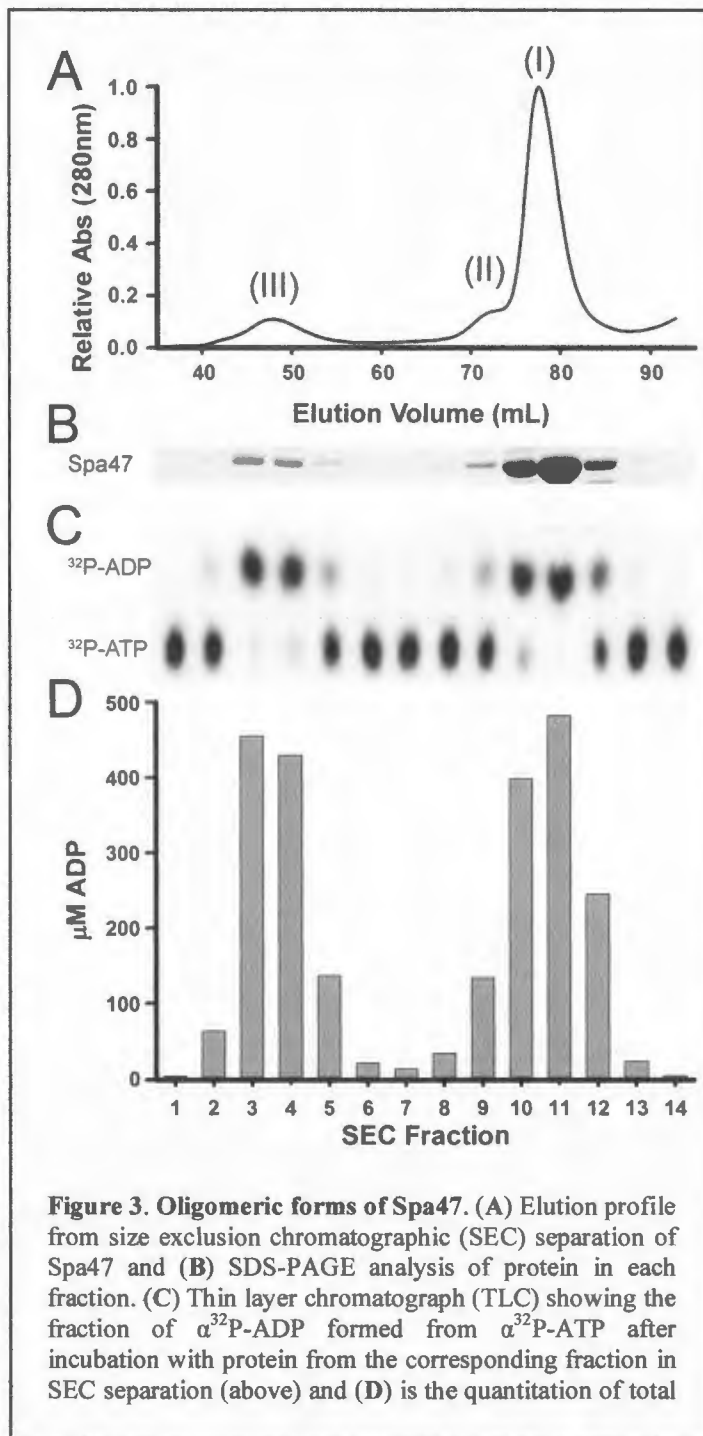
Figure 3. Oligomeric forms of Spa47. (A) Elution profile from size exclusion chromatographic (SEC) separation of Spa47 and (B) SDS-PAGE analysis of protein in each fraction. (C) Thin layer chromatograph (TLC) showing the fraction of α^{32}P-ADP formed from α^{32}P-ATP after incubation with protein from the corresponding fraction in SEC separation (above) and (D) is the quantitation of total

only validate the expression/purification method, but also confirm the hypothesis that Spa47 is a T3SS ATPase. As a first approach, the ATPase activity of each of the SEC separated Spa47 oligomers was tested.

Radiolabeled ATP was mixed with a fixed volume of protein from each fraction and the formation of ADP was monitored. The results show a bimodal distribution of activity that precisely follows the protein elution profile, turning over nearly all of the 500µM ATP in each of the peak fractions under the conditions tested (Figure 3C and D). Together, these results suggest that the recombinant Spa47 is not only active, but that it probably exists in multiple oligomeric forms.

Mass spectrometry identified the purified protein as *Shigella* Spa47 (data not shown), however since Spa47 has not previously been directly tested for activity, we felt it was important to generate an ATPase inactive Spa47 mutant as a negative control to ensure that the observed ATPase activity was in fact coming from Spa47 and not a co-purifying contaminant. A Spa47 point mutant targeting the predicted Walker A motif was engineered by substituting an Alanine for Lysine 165. This residue was chosen based on sequence conservation and previous studies showing that the corresponding residues in FliI (*Salmonella*)[29] and EscN (*E. coli*)[26] are responsible for stabilizing the negative charge of ATP, supporting substrate interaction within the enzyme active site. The Spa47^{K165A} mutant was expressed and purified exactly as was done for the native Spa47 construct with SEC analysis of the Spa47^{K165A} maintaining a nearly identical oligomerization profile as native Spa47 (Figure 4) with monomeric (I) and oligomeric (III) species evident in the chromatogram. When tested for ATP hydrolysis activity, Spa47^{K165A} was inactive, with less than 4 µM total ADP observed following any reaction (Figure 4 insert), compared to nearly 500 µM ADP for wild-type Spa47. These data not only suggest that Spa47 belongs to the Walker type ATPase family,[30] but also served as a control confirming that observed activity levels for native Spa47 were the direct result of Spa47 catalyzed ATP hydrolysis. Additionally, these results suggest that
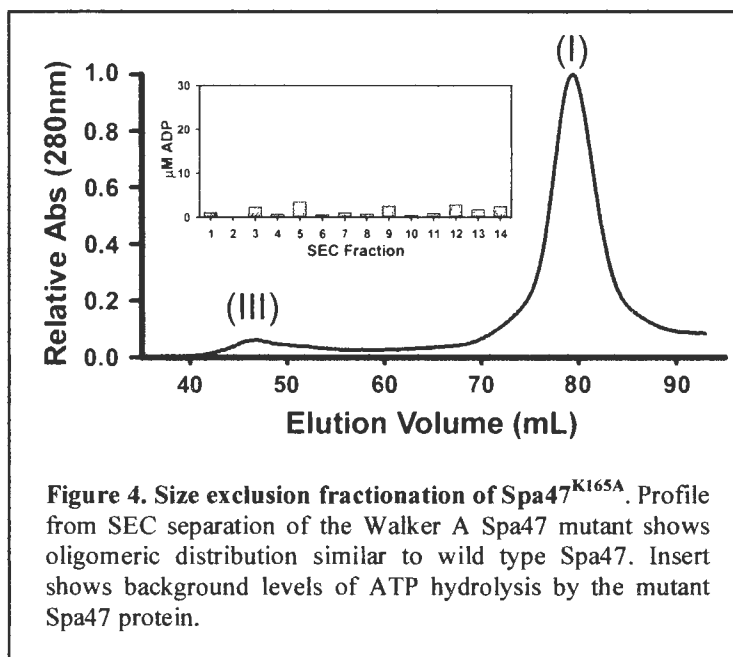

Figure 4. Size exclusion fractionation of Spa47^{K165A}. Profile from SEC separation of the Walker A Spa47 mutant shows oligomeric distribution similar to wild type Spa47. Insert shows background levels of ATP hydrolysis by the mutant Spa47 protein.

ATP hydrolysis by Spa47 is not a requirement for oligomerization however, we cannot rule out the possibility that perhaps transient higher order oligomers are driven by ATP hydrolysis.

Recombinant Spa47 exists in discrete oligomeric states-SEC elution profiles (Figure 3A) suggest that Spa47 exists as a monomer (I) and consistent homo-oligomer (III); though a strong dependence of SEC on protein shape makes stoichiometry determination difficult. Sedimentation velocity analytical ultracentrifugation (SV-AUC) measures the sedimentation coefficient for each species in a solution. The sedimentation coefficient is a function of both size and shape, and thus interpreting molecular weights from the peaks of a sedimentation coefficient distribution requires prior knowledge of the shape of each species. However, with the monomer peak identified, stoichiometric assignments can be made by comparing the sedimentation coefficient of the oligomer to that of the monomer. The sedimentation coefficient ratios expected for geometric oligomers up to octamer are outlined,[31] and because sedimentation coefficients are compared directly to other sedimentation coefficients this analysis requires no knowledge of or deconvolution of the contribution of shape to the sedimentation coefficient. AUC analysis of purified recombinant Spa47 found two distinct peaks with
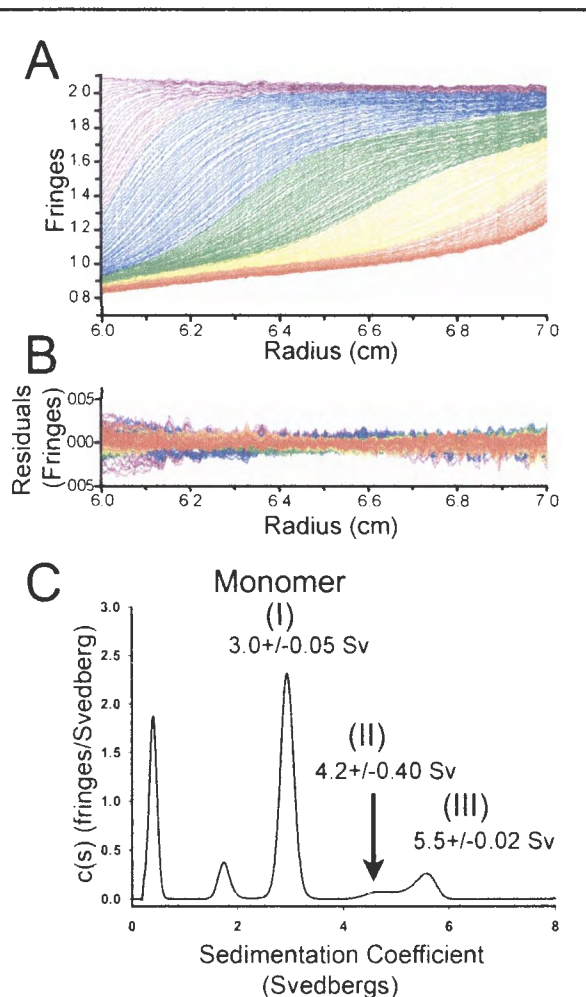
Figure 5. Sedimentation velocity analytical ultracentrifugation analysis of Spa47 oligomerization. (A) Representative interference scans of purified Spa47 monitored during sedimentation velocity analytical ultracentrifugation (SV-AUC) and **(B)** representative residuals from fitting the data to a continuous c(s) distribution model as described in the methods. **(C)** Representative sedimentation coefficient distribution (c(s) versus S) showing the presence of distinct Spa47 species with sedimentation coefficients of 3.0 ± 0.05, 4.2 ± 0.40, and 5.5 ± 0.02 Svedbergs. Sedimentation coefficients represent mean ± SD from 3 independent measurements.

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sedimentation coefficients of 3.0 ± 0.05 and 5.5 ± 0.02 Svedbergs (Figure 5). The 3.0 s peak was the most abundant, and the sedimentation coefficient is consistent with Spa47 monomer (I) with a moderately elongated shape (frictional ratio of 1.57). If the 3.0 s peak is Spa47 monomer, comparing its sedimentation coefficient to that of the 5.5 s peak yields a ratio of 1.83. As a ratio of 1.75 is expected of a linear trimer, and a ratio of 1.86 is expected of a triangular trimer, the 5.5s peak can be tentatively assigned as trimeric homo-oligomers (III). A minor peak was also observed at 4.2 ± 0.40 (II) that is consistent with an intermediate species observed in the SEC profile (Figure 3). Two additional peaks were observed in the sedimentation profiles and likely represent small contaminants that had not yet been removed as the AUC analysis was run prior to SEC to best determine species distribution in the population. The AUC results are consistent with the presence of three primary Spa47 species identified as a monomer (I) and two higher order oligomers (II and III) as seen by SEC. AUC sedimentation coefficient distributions were additionally obtained for Spa47 ranging in concentration from 0.1 to 1.0 mg/ml (Figure 6), finding similar sedimentation profiles as observed in Figure 5. In these experiments the species previously identified as monomer sedimented consistently at approximately 3.0
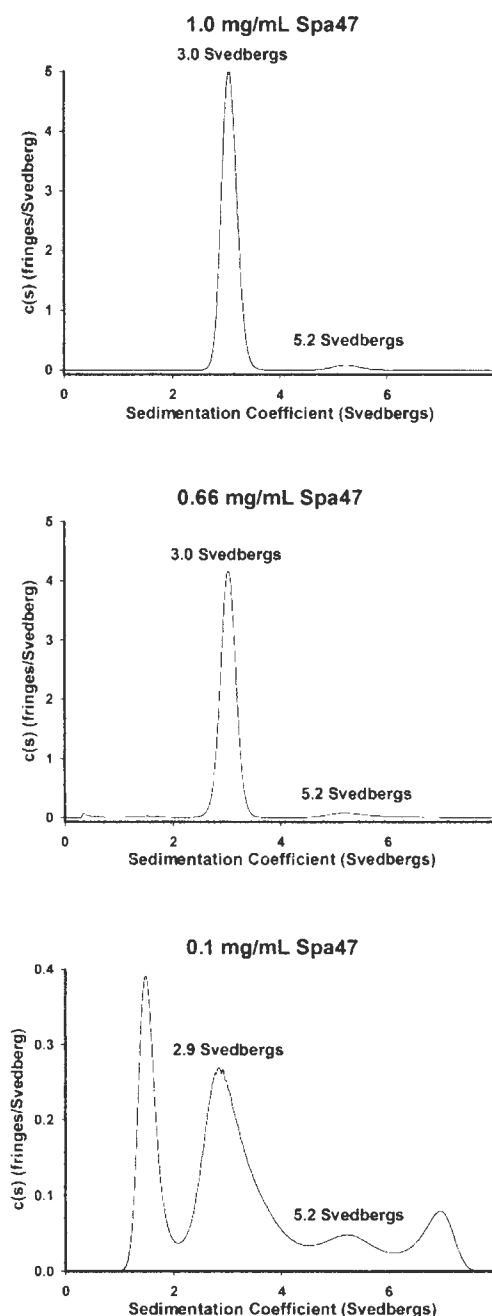
Figure 6. Concentration-dependent SV-AUC analysis of Spa47 oligomerization. Sedimentation coefficient distributions (c(s) versus S) of Spa47 solutions at 1.0, 0.66, and 0.1 mg/mL Spa47 maintain monomer and trimer peaks at approximately 3.0 and 5.2 Svedbergs, respectively. The consistency of monomer and trimer peak distribution across this range of protein concentrations supports the assignment of Spa47 species stoichiometries and ensures that the assigned peaks are not the result of a reaction boundary.

s and the species we believe to be a trimer consistently sedimented at 5.2 s, regardless of concentration, ensuring that the assigned peaks are the result of discrete species and not an artifact resulting from a reaction boundary.

SEC purification isolated the predominant monomeric (I) and oligomeric (III) Spa47 species allowing further characterization of the individual oligomers. CD analysis was performed on the isolated species of Spa47 and Spa47^{K165A} (Figure 7). The Far-UV CD spectra are nearly identical for each suggesting that major differences in structure are not driving the formation of oligomers or responsible for the observed increase in ATPase activity. Furthermore, the accompanying CD thermal unfolding curves exhibit very similar transition temperatures, suggesting that the protein structure is not significantly different between the two tested species. The monomeric species for both proteins do, however, exhibit sharper transitions than the oligomers (III), suggesting that the monomer protein-protein interactions reduce the cooperativity of unfolding. Interestingly, the isolated species were both found to primarily maintain their original oligomeric state and activity levels after nearly one month at 4 °C, suggesting that even though CD found their structure content to be similar, they remain stable under the tested *in vitro* storage conditions (data not shown).

Figure 7. Circular dichroism (CD) analysis of Spa47. (A) Far-UV CD spectra for monomeric (I) and oligomeric (III) forms of wild-type Spa47 and Spa47^{K165A} exhibit minima at 208 nm and 222 nm, characteristic of proteins containing high α-helical secondary structure content. (B) Thermal unfolding profiles were collected by monitoring CD signal at 222 nm as the sample temperature increased from 10 °C to 90 °C. The transition temperatures are similar for each species while the oligomeric (III) forms exhibit more gradual transitions compared to the monomers.

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Spa47 ATPase activity is oligomer-dependent-Together, the SEC and AUC profiles indicate that recombinant Spa47 exists in multiple states with monomer being the predominant species with lesser populations of homo-oligomers (Figures 3 and 5). While monomers, hexamers, and dodecamers of similar proteins have been captured before, the AUC results found that the largest observed Spa47 species (III) exhibits a sedimentation coefficient that is much lower than would be observed for a dodecamer or even homo-hexamer of Spa47 warranting further characterization. A single time-point activity assay of SEC fractions containing isolated Spa47 oligomers found that each of the oligomer species was able to hydrolyze ATP (Figures 3C and D) although activity levels were not proportional to the quantity of protein in each fraction, suggesting that the formation of the oligomeric complex (III) leads to higher activity levels than seen in the monomeric and complex (II) species (Figure 3B). Reaction conditions were explored and it was found that consistent activity levels were observed at room temperature (21 °C) and slightly basic pH (7.9) allowing a direct comparison of the SEC isolated wild-type Spa47 oligomeric species as well as those for the Spa47^{K165A} mutant. Reaction rates were determined by monitoring ATP hydrolysis through ADP product formation under equivalent enzyme and substrate concentrations (1.35 μM and 1.0 mM, respectively) as a function of time (Figure 8). The results show that the Spa47^{K165A} mutant essentially lacks the ability to hydrolyze ATP even after several minutes under these reaction conditions (k_{cat} values ≤ 0.03 s^{-1}) and identified significant differences in hydrolysis activity for the native Spa47 oligomers. Specifically, the Spa47 monomer and predicted homo-dimeric species (II)
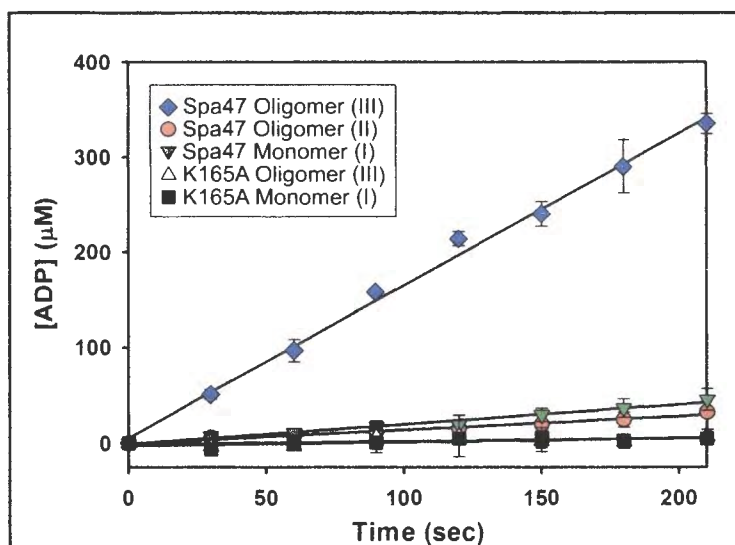
Figure 8. ATPase activity of Spa47. Kinetics of ATP hydrolysis by various oligomeric forms of Spa47. The active oligomeric form (III) of Spa47 hydrolyzes ATP with a k_{cat} of 1.18 ± 0.03 s^{-1}, which is 8-10 fold faster than the monomer and smaller oligomeric form (II) ($k_{cat} = 0.15 \pm 0.01$ s^{-1} and 0.11 ± 0.01 s^{-1} respectively). All forms of the Spa47^{K165A} mutant protein are essentially inactive for ATP hydrolysis with k_{cat} values $\leq 0.03 \pm 0.01$ s^{-1}. Each data point represents the mean $\pm$ SD of 3 independent measurements from 2 separate protein preparations.

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hydrolyze ATP at rates of 0.21 ± 0.01 μM/sec and 0.14 ± 0.01 μM/sec, respectively, while an equivalent

concentration of oligomer (III) predicted to be a trimeric exhibits a rate of 1.60 ± 0.05 μM/sec. This

corresponds to an apparent k_{cat} of 1.18 ± 0.03 s^{-1} for (III) which is similar to that of other T3SS ATPases

including CdsN and FliI,[23,24,29,32] suggesting that Spa47 oligomerization plays a key role in its ATPase

activity.

*ATPase active Spa47 is necessary for cellular invasion-*The full spectrum of T3SS ATPases

functions remains unclear though several key studies have demonstrated their importance *in vivo* and have

provided insight into some specific functions.[21,33,34] While this represents the first direct evidence that Spa47

is an ATPase, its importance in *Shigella* virulence was shown previously when a knockout strain failed to

properly secrete several essential T3SA structural proteins as well as effectors associated with virulence.[21]

We suspected that this secretion handicap would result in a loss of ability for *S. flexneri* to invade cultured

HeLa cells and directly tested this using a gentamycin protection assay. As expected, we found a *spa47* null

S. flexneri strain to be essentially non-invasive, exhibiting only 2% invasion compared to the wild-type

strain 2457T. Complementation of the *spa47* null strain with a pWPsf4 vector containing the native *spa47*

gene restored invasion properties to greater than 90% of wild-type, showing that the presence of Spa47 is

essential for cellular invasion and ultimately the ability of *S. flexneri* to cause infection (Table 1). Because

Spa47 has been shown to interact with several T3SA proteins within the basal body of the apparatus,[21,35-37]

the observed phenotype for *spa47* null strains may result from a lack of necessary structural responsibilities and/or work directly provided by ATP hydrolysis. We used Spa47[K165A] developed and characterized in this study to provide an ATPase inactive form of Spa47 to complement the *spa47* null strain. This strain containing a single Spa47 point mutation exhibited an identical non-invasive phenotype like the *spa47* null strain, suggesting that Spa47 ATPase activity is

Table 1. Invasion Phenotype of Spa47 mutants.		
S. flexneri Strain	Complementation	Relative invasion (% ± SD)
2457T (WT)	none	100 ± 0
spa47 null	none	2 ± 3
spa47 null	spa47/pWPsf4	91 ± 14
spa47 null	spa47[K165A]/pWPsf4	2 ± 1

Table 1. Gentamycin protection data outlining the effects of Spa47 mutations on virulence. This assay measures the ability of *S. flexneri* to invade host cells. The results represent a percentage of colonies formed relative to a wild-type 2457T *S. flexneri* strain ± S.D. n=3 independent experiments each performed in triplicate with an average of 182 colonies for WT.

necessary for cellular invasion. This does not, however, rule out the possibility that Spa47 contributes key structural aspects to the basal body. However, uncoupling the structural and functional roles of Spa47 will require a dedicated set of phenotype and structure studies involving Spa47 mutants such as the ATPase deficient Spa47^{K165A} mutant described in this study.

DISCUSSION

Shigella spp. rely on a T3SS to promote uptake by the epithelial cells lining the large intestine of their human hosts, initiate apoptosis of resident macrophages, escape endocytic vacuoles, and provide actin-based motility.[5,38,39] Providing these critical roles at the correct point during infection requires that the T3SS is under strict control at many levels including transcriptional and translational regulation as well as control over the maturation and activation of the injectosome itself.[13] The secretion of T3SS translocator proteins and their incorporation into the maturing T3SA have been linked to environmental stimuli by small molecules such as the bile salt deoxycholate[38,40,41] and lipid membrane components including cholesterol and sphingomyelin.[11] This stimulation has been suggested to provide signals which essentially monitor progress as the pathogen enters the host and passes through the gastrointestinal tract, preparing the *Shigella* T3SA for invasion of the colonic epithelium. What has remained unclear, however, is what provides the driving force for protein secretion through the *Shigella* T3SA and ultimately how secretion is controlled.

Sequence homology between the *Shigella* protein Spa47 and identified ATPases including the β-subunit of the hetero-hexameric F_1F_o ATP synthase, FliI of the bacterial flagellar system, and ATPases associated with the translocon T3SS of several pathogens suggests that Spa47 is a T3SS-associated ATPase in *Shigella* and that it may provide the energy necessary for T3SS-associated secretion. Much of this stems from work showing that a *spa47* null *Shigella* strain is unable to construct the T3SA and as a result fails to secrete T3SS proteins, including several Ipas and the phosphatase effector IpgD,[21] though challenges purifying Spa47 have prevented a direct correlation between *Shigella* T3SS activity and Spa47 ATP hydrolysis. In this study, we developed an expression and purification method for recombinant Spa47 that allowed us to provide the first direct characterization of its enzymatic activity and identify a link between

Spa47 oligomer state and ATPase activity. Furthermore, we captured and characterized the active Spa47 homo-oligomer (III), finding that it likely represents a smaller T3SS ATPase complex than has been observed for other systems and provide evidence suggesting that *Shigella* invasion of host cells requires Spa47 catalyzed ATP hydrolysis.

Size exclusion chromatography of recombinant Spa47 resulted in multiple resolved species, suggesting it exists in discrete oligomeric states (Figure 3A and B) which were determined by AUC to be primarily Spa47 monomer (I) with a lesser population of oligomers (II and III) which we predict to be Spa47 homo-dimers and homo-trimers, respectively, based on sedimentation coefficients obtained by AUC (Figure 5). While the implication of this distribution was initially unclear, Spa47 activity was found to be highly dependent on oligomeric state, suggesting it may be an important component of a regulation mechanism. Many ATPases including members of the AAA and AAA$^+$ classes undergo oligomerization to form active complexes, often serving as a means of regulation and providing an amplification of the mechanical force generated by ATP hydrolysis.[42] One such example is the highly studied flagellar ATPase FliI (Spa47 homolog) whose oligomerization to an active hexamer is negatively influenced by interaction with the regulatory protein FliH and is stimulated by ATP and *E. coli* phospholipid binding.[43,44] Regulation of the ATPase FliI through oligomerization provides tight control over export of flagellar proteins and may serve a similar role in the evolutionarily related *Shigella* T3SS.

Activity profiles of the isolated Spa47 oligomers found that the activated oligomer (III) exhibited 8-fold higher activity levels than the monomer (I) (k_{cat} = 1.18 ± 0.03 s^{-1}) (Figure 8), suggesting that Spa47 activation may be controlled by oligomeric state and specifically formation of the observed complex. In contrast to FliI, however, Spa47 formed oligomers in the absence of ATP (Figure 3 and Figure 5) as has been demonstrated for the T3SS ATPase HrcN from the plant pathogen *Pseudomonas syringae*.[26] Although active hexameric, dodecameric, and even higher order oligomeric states have been reported, we believe that the observed active Spa47 complex (III) studied here represents a novel trimeric T3SS ATPase though additional work is required to confirm the precise stoichiometry of the active species. Additionally, while we cannot rule out the possibility of ATP driven higher order oligomerization of the active oligomer (III)

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(eg. hexamer as seen in the transmission electron microscopy generated structure),[35] the ATP hydrolysis deficient Spa47^{K165A} mutant displayed oligomeric behavior similar to wild-type Spa47 suggesting that ATP is not required for Spa47 oligomer formation as the equivalent lysine in FliI and EscN was needed to stabilize ATP binding.[26,29]

CD analysis of the monomeric and oligomeric (III) forms of wild-type and the K165A Spa47 mutant found that each result in spectra characteristic of highly α-helical proteins (Figure 7) consistent with the known structures of related T3SS ATPases.[26,33,45] Together these results suggest that neither the oligomeric state nor the K165A mutation drastically affect the protein secondary structure. Additionally, the thermal unfolding curves exhibit similar transition temperatures while the active species (III) demonstrating shallower transitions reflecting the additional protein interactions of the multimeric state. Without atomic resolution structural data, it is impossible to entirely rule out protein misfolding as the reason for the appearance of the less active monomeric species. However, insight from related systems together with the data presented here support the hypothesis that Spa47 oligomerization is involved in activation and regulation of the *Shigella* T3SS.

Recent transmission electron microscopy electron density maps provided new insight into the architecture of the *Shigella* T3SA sorting platform and confirmed the relevance of Spa47 oligomer formation by identifying a symmetric homo-hexameric Spa47 ring at the base of the intact T3SA. The Spa47 hexamer is positioned directly below the MxiA export gate[35,46] suggesting that secreted proteins interact with Spa47 prior to entering the apparatus needle for export. The structure further suggests that Spa47 is positioned via symmetric hexameric linkage to six individual Spa33 units of the C ring through direct interaction with MxiN, confirming yeast two-hybrid and co-immunoprecipitation assays that first described these interactions.[21,36,37] The observed interaction between Spa47 and MxiN is particularly interesting as MxiN is homologous to the regulatory protein FliH in the flagellar export apparatus which appears to prevent formation of the active FliI homo-hexamer, preventing the wasteful hydrolysis of ATP before the flagellar export apparatus is prepared for productive transport.[43,44] A similar mechanism would be beneficial to the *Shigella* T3SS although the association of MxiN and Spa47 appears to be necessary for

proper formation of the sorting platform[35] and a *mxiN* null *Shigella* strain less efficiently secretes MxiH and MxiI, preventing proper T3SA needle formation.[21]

We suspect that the role of Spa47 in the *Shigella* T3SS is multifaceted, serving as both a structural component within the sorting platform and as an ATPase that provides the energy necessary for secretion, perhaps through recognition, chaperone release, and partial unfolding of transported proteins as has been demonstrated for the *Salmonella enterica* ATPase InvC.[34] In *Shigella*, it has been shown that a *spa47* null *Shigella* strain is unable to form the external needle structure of the T3SA and as a result is secretion and invasion deficient.[21,47] We tested the invasion phenotype of a *Shigella spa47* null strain complemented with the engineered ATPase inactive Spa47^{K165A} mutant finding that it resulted in the same non-invasive phenotype as the null strain even though invasion was restored when complemented with native *spa47*. Together with recent structural findings[35] and the enzymatic activity characterization provided in this study, it appears that Spa47 may play critical structural roles in the T3SA and is in fact a functional ATPase whose activity may be tied directly to *Shigella* virulence. The findings presented here are the result of overcoming the long-standing hurdle of recombinant expression and purification of the *Shigella* ATPase Spa47 and together have uniquely positioned us to begin answering a number of key questions regarding Spa47 activation, T3SS secretion regulation, and potential structural/organizational roles of Spa47 within the sorting platform of the T3SA. Ultimately, answering these questions will require a direct comparison of protein interactions, phenotype assays, and further characterization of enzyme activity and structure. Together, the methods developed in this study to produce active recombinant Spa47, the generation of a key ATPase inactive Spa47 point mutant, and the link identified between Spa47 oligomeric state and ATPase activity will provide a strong platform for this additional work dissecting the mechanism of Spa47 catalyzed ATP hydrolysis and its role in the *Shigella* T3SS.

REFLECTIVE WRITING (1187 words)

The most daunting thing about an Honors capstone project is getting started. As a freshman, even thinking about what I would do for a capstone project was overwhelming. However, the Honors program is perfectly set up to make the capstone experience a seamless transition from the classroom to outside experience.

As a biochemistry major, I needed to complete several contracts in chemistry courses. One of the courses I selected to complete a contract in was CHEM 5700: Biochemistry I. I was interested in how research was conducted in the biochemistry department, so for my contract, I completed a literature review of a new topic of interest in the Dickenson lab. The topic was on the current state of knowledge (before the above research) of Spa47 purification.

By the end of the semester, I had complied a literature review not only on the very little knowledge in the field about Spa47, but also on current ideas and possibilities for purifying Spa47 recombinantly. I found myself surprised at how intrigued I was of the topic, and luckily I had impressed Dr. Dickenson enough with my dedication to the literature review that I was invited to try my hand at research in his lab.

Entering a research lab was so different from a teaching lab. Each lab group has their own variations of biochemical techniques. The nuances of what order to do things, what recipes to use, what method is best to use, all needed to be learned. As a result, I am still learning new techniques! The great thing about being a part of a research lab group though is that you aren't alone. You can ask for help from not only your PI, but also senior members of the lab about new techniques and ideas.

As I gained (and am still gaining) experience in lab research, I found a passion for my project. Now I had a topic and data for my honors capstone that I didn't mind putting in hundreds of hours towards. What had seemed like a daunting task as a freshman has become a relatively simple task because of the interest I had in the project. I think overall, just getting the writing polished to its final draft has been the most difficult part of the entire capstone experience; I already knew my topic, had my research data, and my discussion laid out. In fact, my capstone project here is an adaption of my first author published manuscript found in the journal of *Protein Science.* So getting my paper to peer-reviewed publishing quality required MANY

drafts, revisions, and edits. As a result of my journey through finding a research lab and completing my capstone project, I would like to specifically point out some problems, challenges, triumphs, and some advice I have for future Honors students.

Problems:

Science. While science is an amazing subject, sometimes getting your research to work properly doesn't always work out. One of the biggest hurdles I had to overcome in my research was being able to obtain recombinant protein to study. The most common technique biochemists use to purify recombinant proteins is to use a type of affinity tag. When first working with Spa47, several affinity tags were found to not be compatible with soluble protein expression (including maltose binding protein tag, poly-histidine nickel tag, etc.).

The way I and my lab members were able to overcome this hurdle was by talking to other members of the department. The research lab just across the hall from us suggested trying a chitin binding domain tag, a large protein tag that seems to help with solubility. Amazingly, it worked! The chitin binding tag was able to solubilize Spa47 and allow for recombinant expression. It was a humbling lesson on the importance of intra-departmental communication. The answer you may be looking for could be found in the insight/experience of your peers.

Challenges:

The biggest challenge for my capstone experience is learning lab techniques. Even after one and a half years of experience in the Dickenson research lab with over 1,290 hours logged, I have found that I still don't know half of the techniques performed in the lab. If I had known even just the techniques I know now, my research progress would have been ten times faster, but learning is all a part of the process. It taught me patience and how to take good notes so I can repeat what I have been shown. It has taught me that there is more to biochemistry than you learn in the classroom or even teaching labs. I also learned that you get what you put in. In other words, I learned lots of skills by putting in my hundreds of hours into the lab. I had to sacrifice time for my learning but I believe the benefits from my developed lab skills far outweigh the cost.

Triumphs:

By far, the biggest triumph of my capstone experience was getting a first-author publication in *Protein Science* as an undergraduate. Being able to participate in research is satisfying, but being able to have YOUR work published in a journal felt very rewarding. All the hours spent not only on research but also on writing and editing finally paid off. Not only was my research was published, but it was peer-review approved. Peer-review is the gold-standard in the research field for publications. In order to get to that point though, a lot of time and effort has to go into the writing process. I learned a lot about scientific writing skills and how to present research to the scientific community in a meaningful way.

Advice:

My biggest advice to upcoming Honors students is to get started early! Explore disciplines, beyond the classroom. Find something that intrigues you and that you would be happy to put your time and effort into. It makes the whole process so much easier if you can find a passion for your work. Regardless of if you like the subject, you will HAVE to put in a lot of hours into your capstone. The writing process itself takes a lot of time, let alone the hours to collect the research content.

My other piece of advice is to find a good mentor that fits your style. For me, I needed a mentor who was willing to explain the nitty gritty details to me about everything…at first. But once I learned the required techniques, I had free reigns on moving my project forward. I found this with Dr. Dickenson. Some people like more guidance or would rather have a "hands-off" mentor. Regardless of which, find the right mentor for you because it made a world of difference for me. I don't think I would of progressed as far as I have without the proper mentorship.

Above all else, have fun! The Honors capstone experience is really just that: an amazing, above the average experience. It really brought together my extracurricular and academic experience at USU and I hope all future Honors students will find it to be as rewarding as I did.

WORKS CITED

1. Kotloff, K. L., Nataro, J. P., Blackwelder, W. C., Nasrin, D., Farag, T. H., Panchalingam, S., Wu, Y., Sow, S. O., Sur, D., Breiman, R. F., Faruque, A. S., Zaidi, A. K., Saha, D., Alonso, P. L., Tamboura, B., Sanogo, D., Onwuchekwa, U., Manna, B., Ramamurthy, T., Kanungo, S., Ochieng, J. B., Omore, R., Oundo, J. O., Hossain, A., Das, S. K., Ahmed, S., Qureshi, S., Quadri, F., Adegbola, R. A., Antonio, M., Hossain, M. J., Akinsola, A., Mandomando, I., Nhampossa, T., Acacio, S., Biswas, K., O'Reilly, C. E., Mintz, E. D., Berkeley, L. Y., Muhsen, K., Sommerfelt, H., Robins-Browne, R. M., and Levine, M. M. (2013) Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study, Lancet 382, 209-222.

2. DuPont, H. L., Levine, M. M., Hornick, R. B., and Formal, S. B. (1989) Inoculum size in shigellosis and implications for expected mode of transmission, J Infect Dis 159, 1126-1128.

3. Kotloff, K. L., Winickoff, J. P., Ivanoff, B., Clemens, J. D., Swerdlow, D. L., Sansonetti, P. J., Adak, G. K., and Levine, M. M. (1999) Global burden of Shigella infections: implications for vaccine development and implementation of control strategies, Bull World Health Organ 77, 651-666.

4. Dutta, S., Ghosh, A., Ghosh, K., Dutta, D., Bhattacharya, S. K., Nair, G. B., and Yoshida, S. (2003) Newly emerged multiple-antibiotic-resistant Shigella dysenteriae type 1 strains in and around Kolkata, India, are clonal, J Clin Microbiol 41, 5833-5834.

5. Carayol, N., and Nhieu, G. T. V. (2013) The Inside Story of Shigella Invasion of Intestinal Epithelial Cells, Csh Perspect Med 3,

6. Schroeder, G. N., Jann, N. J., and Hilbi, H. (2007) Intracellular type III secretion by cytoplasmic Shigella flexneri promotes caspase-1-dependent macrophage cell death, Microbiology 153, 2862-2876.

7. Chatterjee, S., Chaudhury, S., McShan, A. C., Kaur, K., and De Guzman, R. N. (2013) Structure and Biophysics of Type III Secretion in Bacteria, Biochemistry 52, 2508-2517.

8. Ashida, H., Mimuro, H., and Sasakawa, C. (2015) Shigella manipulates host immune responses by delivering effector proteins with specific roles, Front Immunol 6, 219.

9. Enninga, J., Mounier, J., Sansonetti, P., and Tran Van Nhieu, G. (2005) Secretion of type III effectors into host cells in real time, Nat Methods 2, 959-965.

10. Cordes, F. S., Komoriya, K., Larquet, E., Yang, S., Egelman, E. H., Blocker, A., and Lea, S. M. (2003) Helical structure of the needle of the type III secretion system of Shigella flexneri, J Biol Chem 278, 17103-17107.

11. Epler, C. R., Dickenson, N. E., Olive, A. J., Picking, W. L., and Picking, W. D. (2009) Liposomes recruit IpaC to the Shigella flexneri type III secretion apparatus needle as a final step in secretion induction, Infection and immunity 77, 2754-2761.

12. Deane, J. E., Abrusci, P., Johnson, S., and Lea, S. M. (2010) Timing is everything: the regulation of type III secretion, Cell Mol Life Sci 67, 1065-1075.

13. Marteyn, B., Gazi, A., and Sansonetti, P. (2012) Shigella: a model of virulence regulation in vivo, Gut Microbes 3, 104-120.

14. Pallen, M. J., Bailey, C. M., and Beatson, S. A. (2006) Evolutionary links between FliH/YscL-like proteins from bacterial type III secretion systems and second-stalk components of the FoF1 and vacuolar ATPases, Protein Science 15, 935-940.

15. Blocker, A., Komoriya, K., and Aizawa, S. (2003) Type III secretion systems and bacterial flagella: insights into their function from structural similarities, Proc Natl Acad Sci U S A 100, 3027-3030.

16. Eichelberg, K., Ginocchio, C. C., and Galan, J. E. (1994) Molecular and functional characterization of the Salmonella typhimurium invasion genes invB and invC: homology of InvC to the F0F1 ATPase family of proteins, J Bacteriol 176, 4501-4510.

17. Akeda, Y., and Galan, J. E. (2004) Genetic analysis of the Salmonella enterica type III secretion-associated ATPase InvC defines discrete functional domains, Journal of bacteriology 186, 2402-2412.

18. Woestyn, S., Allaoui, A., Wattiau, P., and Cornelis, G. R. (1994) YscN, the putative energizer of the Yersinia Yop secretion machinery, J Bacteriol 176, 1561-1569.

19. Lee, P. C., and Rietsch, A. (2015) Fueling type III secretion, Trends Microbiol 23, 296-300.

20. Burkinshaw, B. J., and Strynadka, N. C. (2014) Assembly and structure of the T3SS, Biochim Biophys Acta 1843, 1649-1663.

21. Jouihri, N., Sory, M. P., Page, A. L., Gounon, P., Parsot, C., and Allaoui, A. (2003) MxiK and MxiN interact with the Spa47 ATPase and are required for transit of the needle components MxiH and MxiI, but not of Ipa proteins, through the type III secretion apparatus of Shigella flexneri, Molecular microbiology 49, 755-767.

22. Chatterjee, R., Halder, P. K., and Datta, S. (2013) Identification and molecular characterization of YsaL (Ye3555): a novel negative regulator of YsaN ATPase in type three secretion system of enteropathogenic bacteria Yersinia enterocolitica, PLoS One 8, e75028.

23. Andrade, A., Pardo, J. P., Espinosa, N., Perez-Hernandez, G., and Gonzalez-Pedrajo, B. (2007) Enzymatic characterization of the enteropathogenic Escherichia coli type III secretion ATPase EscN, Archives of biochemistry and biophysics 468, 121-127.

24. Stone, C. B., Johnson, D. L., Bulir, D. C., Gilchrist, J. D., and Mahony, J. B. (2008) Characterization of the putative type III secretion ATPase CdsN (Cpn0707) of Chlamydophila pneumoniae, Journal of Bacteriology 190, 6580-6588.

25. Pozidis, C., Chalkiadaki, A., Gomez-Serrano, A., Stahlberg, H., Brown, I., Tampakaki, A. P., Lustig, A., Sianidis, G., Politou, A. S., Engel, A., Panopoulos, N. J., Mansfield, J., Pugsley, A. P., Karamanou, S., and Economou, A. (2003) Type III protein translocase: HrcN is a peripheral ATPase that is activated by oligomerization, J Biol Chem 278, 25816-25824.

26. Zarivach, R., Vuckovic, M., Deng, W., Finlay, B. B., and Strynadka, N. C. (2007) Structural analysis of a prototypical ATPase from the type III secretion system, Nature structural & molecular biology 14, 131-137.

27. Habersetzer, J., Ziani, W., Larrieu, I., Stines-Chaumeil, C., Giraud, M. F., Brethes, D., Dautant, A., and Paumard, P. (2013) ATP synthase oligomerization: from the enzyme models to the mitochondrial morphology, Int J Biochem Cell Biol 45, 99-105.

28. Claret, L., Calder, S. R., Higgins, M., and Hughes, C. (2003) Oligomerization and activation of the FliI ATPase central to bacterial flagellum assembly, Mol Microbiol 48, 1349-1355.

29. Fan, F., and Macnab, R. M. (1996) Enzymatic characterization of FliI. An ATPase involved in flagellar assembly in Salmonella typhimurium, J Biol Chem 271, 31981-31988.

30. Walker, J. E., Saraste, M., Runswick, M. J., and Gay, N. J. (1982) Distantly related sequences in the alpha- and beta-subunits of ATP synthase, myosin, kinases and other ATP-requiring enzymes and a common nucleotide binding fold, EMBO J 1, 945-951.

31. Garcia de la Torre, J. G., and Bloomfield, V. A. (1981) Hydrodynamic properties of complex, rigid, biological macromolecules: theory and applications, Quarterly reviews of biophysics 14, 81-139.

32. Stone, C. B., Bulir, D. C., Gilchrist, J. D., Toor, R. K., and Mahony, J. B. (2010) Interactions between flagellar and type III secretion proteins in Chlamydia pneumoniae, Bmc Microbiology 10,

33. Allison, S. E., Tuinema, B. R., Everson, E. S., Sugiman-Marangos, S., Zhang, K., Junop, M. S., and Coombes, B. K. (2014) Identification of the docking site between a type III secretion system ATPase and a chaperone for effector cargo, J Biol Chem 289, 23734-23744.

34. Akeda, Y., and Galan, J. E. (2005) Chaperone release and unfolding of substrates in type III secretion, Nature 437, 911-915.

35. Hu, B., Morado, D. R., Margolin, W., Rohde, J. R., Arizmendi, O., Picking, W. L., Picking, W. D., and Liu, J. (2015) Visualization of the type III secretion sorting platform of Shigella flexneri, Proceedings of the National Academy of Sciences of the United States of America 112, 1047-1052.

36. Johnson, S., and Blocker, A. (2008) Characterization of soluble complexes of the Shigella flexneri type III secretion system ATPase, FEMS Microbiol Lett 286, 274-278.

37. Morita-Ishihara, T., Ogawa, M., Sagara, H., Yoshida, M., Katayama, E., and Sasakawa, C. (2006) Shigella Spa33 is an essential C-ring component of type III secretion machinery, Journal of Biological Chemistry 281, 599-607.

38. Olive, A. J., Kenjale, R., Espina, M., Moore, D. S., Picking, W. L., and Picking, W. D. (2007) Bile salts stimulate recruitment of IpaB to the Shigella flexneri surface, where it colocalizes with IpaD at the tip of the type III secretion needle, Infect Immun 75, 2626-2629.

39. Niebuhr, K., and Sansonetti, P. J. (2000) Invasion of epithelial cells by bacterial pathogens the paradigm of Shigella, Subcell Biochem 33, 251-287.

40. Dickenson, N. E., Zhang, L., Epler, C. R., Adam, P. R., Picking, W. L., and Picking, W. D. (2011) Conformational changes in IpaD from Shigella flexneri upon binding bile salts provide insight into the second step of type III secretion, Biochemistry 50, 172-180.

41. Dickenson, N. E., Arizmendi, O., Patil, M. K., Toth, R. T. t., Middaugh, C. R., Picking, W. D., and Picking, W. L. (2013) N-terminus of IpaB provides a potential anchor to the Shigella type III secretion system tip complex protein IpaD, Biochemistry 52, 8790-8799.

42. White, S. R., and Lauring, B. (2007) AAA+ ATPases: achieving diversity of function with conserved machinery, Traffic 8, 1657-1667.

43. Minamino, T., Kazetani, K., Tahara, A., Suzuki, H., Furukawa, Y., Kihara, M., and Namba, K. (2006) Oligomerization of the bacterial flagellar ATPase FliI is controlled by its extreme N-terminal region, Journal of molecular biology 360, 510-519.

44. Minamino, T., and MacNab, R. M. (2000) FliH, a soluble component of the type III flagellar export apparatus of Salmonella, forms a complex with FliI and inhibits its ATPase activity, Molecular microbiology 37, 1494-1503.

45. Imada, K., Minamino, T., Tahara, A., and Namba, K. (2007) Structural similarity between the flagellar type III ATPase FliI and F-1-ATPase subunits, P Natl Acad Sci USA 104, 485-490.

46. Abrusci, P., Vergara-Irigaray, M., Johnson, S., Beeby, M. D., Hendrixson, D. R., Roversi, P., Friede, M. E., Deane, J. E., Jensen, G. J., Tang, C. M., and Lea, S. M. (2013) Architecture of the major component of the type III secretion system export apparatus, Nature structural & molecular biology 20, 99-104.

47. Tamano, K., Aizawa, S., Katayama, E., Nonaka, T., Imajoh-Ohmi, S., Kuwae, A., Nagai, S., and Sasakawa, C. (2000) Supramolecular structure of the Shigella type III secretion machinery: the needle part is changeable in length and essential for delivery of effectors, Embo J 19, 3876-3887.

48. Formal, S. B., Dammin, G. J., Labrec, E. H., and Schneider, H. (1958) Experimental Shigella infections: characteristics of a fatal infection produced in guinea pigs, J Bacteriol 75, 604-610.

49. Lebowitz, J., Lewis, M. S., and Schuck, P. (2002) Modern analytical ultracentrifugation in protein science: a tutorial review, Protein Sci 11, 2067-2079.

50. Laue, T., Shah, B., Ridgeway, T., and Pelletier, S. (1992) Analytical Ultracentrifugation in Biochemistry and Polymer Science. in Analytical Ultracentrifugation in Biochemistry and Polymer Science (Harding, S., Rowe, A., and Horton, J. C. eds.), Royal Society of Chemistry, United Kingdom.

51. Niesel, D. W., Chambers, C. E., and Stockman, S. L. (1985) Quantitation of HeLa cell monolayer invasion by Shigella and Salmonella species, J Clin Microbiol 22, 897-902.

AUTHOR BIBLIOGRAPHY

Jamie Kingsford is a recently married USU undergraduate student also known as Jamie Burgess. She will be graduating with Departmental Honors in Biochemistry. She has been fully involved in research since January 2015 and had published her research as a first-author paper in the journal of *Protein Science.* She has presented at several conferences including Utah Research on Capitol Hill and the Utah Conference for Undergraduate Research. As a graduating senior, she has been awarded the Maeser-Bauer Outstanding Graduating Senior in Biochemistry and the College of Science Undergraduate Teaching Fellow of the Year award. Jamie plans on continuing her research as a Masters student in Biochemistry at USU followed by medical school.