University of Nebraska - Lincoln DigitalCommons@University of Nebraska - Lincoln

Agronomy & Horticulture -- Faculty Publications

Agronomy and Horticulture Department

2021

Responses of soil surface greenhouse gas emissions to nitrogen and sulfur fertilizer rates to Brassica carinata grown as a bio-jet fuel

Dwarika Bhattarai South Dakota State University, Brookings

Gandura O. Abagandura University of Nebraska - Lincoln, gabagandura2@unl.edu

Thandiwe Nleya South Dakota State University, Brookings

Sandeep Kumar South Dakota State University, Brookings

Follow this and additional works at: https://digitalcommons.unl.edu/agronomyfacpub

Part of the Agricultural Science Commons, Agriculture Commons, Agronomy and Crop Sciences Commons, Botany Commons, Horticulture Commons, Other Plant Sciences Commons, and the Plant Biology Commons

Bhattarai, Dwarika; Abagandura, Gandura O.; Nleya, Thandiwe; and Kumar, Sandeep, "Responses of soil surface greenhouse gas emissions to nitrogen and sulfur fertilizer rates to Brassica carinata grown as a bio-jet fuel" (2021). *Agronomy & Horticulture -- Faculty Publications*. 1434. https://digitalcommons.unl.edu/agronomyfacpub/1434

This Article is brought to you for free and open access by the Agronomy and Horticulture Department at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Agronomy & Horticulture -- Faculty Publications by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

DOI: 10.1111/gcbb.12784

ORIGINAL RESEARCH



Responses of soil surface greenhouse gas emissions to nitrogen and sulfur fertilizer rates to *Brassica carinata* grown as a bio-jet fuel

Dwarika Bhattarai¹ | Gandura O. Abagandura² | Thandiwe Nleya¹ | Sandeep Kumar¹

¹Department of Agronomy, Horticulture and Plant Science, South Dakota State University, Brookings, SD, USA

²Agronomy and Horticulture, University of Nebraska-Lincoln, Lincoln, NE, USA

Correspondence

Sandeep Kumar, Department of Agronomy, Horticulture and Plant Science, South Dakota State University, Brookings, SD, USA. Email: Sandeep.Kumar@sdstate.edu

Funding information

South Dakota Oilseeds Initiative; USDA-NIFA, Grant/Award Number: 2014-38502-22598

Abstract

Carinata (Brassica carinata A. Braun), a non-food oilseed crop and an alternative bio-jet fuel feedstock, has received attention for its potential as a low-input option for production in the semi-arid region of the Northern Great Plains of the United States. Research addressing the impacts of nitrogen (N) and sulfur (S) fertilizers on soils and greenhouse gas (GHG; CO₂, N₂O, and CH₄) emissions from carinata production are limited. Thus, objective of this study was to evaluate the impact of different rates of N and S fertilizers applied to carinata on soil properties and GHG emissions. Field experiments were conducted in 2017 and 2018 to assess the response of carinata to four N (56, 84, 112, and 140 kg N ha^{-1}) and three S (0, 22, and 45 kg S ha^{-1}) rates. Soil samples were collected at crop harvest to measure soil properties; however, soil surface GHG fluxes were measured during 2017 and 2018 growing seasons using static chamber method. Data showed that application of N fertilizer increased soil EC, soil organic carbon (SOC), stable C, and labile N. However, sulfur fertilizer decreased SOC, labile N, and soil inorganic N contents. Results from GHG fluxes showed that higher rates of N fertilizer application increased the soil CO₂ and N₂O emissions, whereas the S fertilizer did not impact these fluxes. This study concludes that S and N fertilizers application to carinata crop affected soil properties, and higher rates of N fertilizer increased the GHG emissions. Therefore, N fertilizer application rate needs to be optimized to mitigate GHG emission for carinata production.

KEYWORDS

biofuel crops, carinata, greenhouse gas, nitrogen rates, soil properties, sulfur fertilizer

1 | INTRODUCTION

Bioenergy has potential to reduce greenhouse gas (GHG) emissions through sustainable resource development and the use of efficient bioenergy systems (Chum et al., 2011). One of the important components for this system is liquid biofuel.

Most common biofuel includes bioethanol derived from corn (*Zea mays* L.), sugar beet (*Beta vulgaris* L.), and sugarcane (*Saccharum officinarum* L.), which is mixed with gasoline, whereas oilseed crops-based biofuel, soybean (*Glycine max* L. Merr.), consumes high energy due to short hydrocarbon chains, to produce fuel (Perlack, 2005). Carinata (*Brassica*

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

 $\ensuremath{\mathbb O}$ 2020 The Authors. GCB Bioenergy Published by John Wiley & Sons Ltd

-WILEY-GCB-BIOENERG

carinata A. Braun), a non-food biofuel crop, has been the focus of much research attention because it can be used as an alternative source of biofuel production (Cardone et al., 2003), which has the potential to reduce global warming through producing low GHG fuel (Agrisoma-USA, 2020). Carinata is originated from the Ethiopian Highlands and is commonly known as Ethiopian mustard (Warwick et al., 2009). Carinata has a mean oil content of 41.7% and a protein content of 30%, depending on the variety and environmental conditions (Hossain et al., 2019). The seed contains longchain unsaturated fatty acids suitable for the production of bio-jet fuel, lubricants, and bioplastics (Taylor et al., 2010). Carinata is heat and drought-tolerant (Agrisoma-USA, 2020), which makes it an ideal crop for production in semi-arid environments, including central and western South Dakota (SD). Moreover, carinata is also an excellent rotational crop for cereal-based rotations, enhancing overall crop productivity and soil health (Wright, 2017).

Similar to other crops, oilseed crops require N, P, K, and S fertilizers, among which N and S fertilizers are crucial for vegetative and reproductive growth (Abdallah et al., 2010; CFIA, 2017). The current research has less focus on P and K fertilizers as the soil has adequate amount of these nutrients available in the given area. Brassica crops have high sulfur demand for the synthesis of sulfur containing compound, glucosinolate, and amino acids, methionine and cysteine, that determines the oilseed quality (Walker & Booth, 2003). Oilseed crops require balanced plant nutrition for enhanced seed and oil production (Ma et al., 2019). The growth and seed yield of carinata were increased with N and S application in India (Verma et al., 2018). A recent study in SD reported that inorganic N fertilizer requirements for carinata are lower compared to the cereal crops (Osborne et al., 2019), which can reduce the greenhouse gas (GHG) emissions (Zhong et al., 2016).

The application of inorganic N fertilizers is important for plant and soil health; however, repeated application of high rates of N fertilizers can affect soil health (Singh, 2018). Growing carinata can help to sustain soil health through low input demand, improvement in soil aggregates, and better carbon sequestering (Agrisoma-USA, 2019). Nitrogen fertilizer can improve soil organic carbon (SOC) content through increased microbial activity (Palmer et al., 2017). A number of studies have found that the long-term application of N fertilizers has significantly improved SOC (Lugato et al., 2010; Zhou et al., 2013). Biomass yield and root growth were improved by the application of inorganic N fertilizers, resulting in improved root activity and accelerated organic C accumulation (Liu et al., 2013). However, a study conducted in SD by Li et al. (2019) reported that increasing N fertilization rates from 0 to 84 kg N ha⁻¹ in a carinata crop grown on fine-silt soil did not affect SOC compared to the control. A study conducted in a canola field reported an increase in SOC

with the application of N fertilizers due to increased crop residue (Kazemeini et al., 2010). However, a study of mustard (Brassica juncea L.) showed increased biomass yield with the application of N fertilizers but unaffected SOC (Prasad et al., 2018). Application of N fertilizer is necessary to replenish the depleted N from soil resulting from plant uptake, leaching or evaporation loss. A study in canola showed that the field receiving higher rates of N fertilizer significantly increased soil mineral N (Herath et al., 2017). Not many studies reported the impact of S fertilizers on SOC under oilseed crops; however, a study in Canada reported a 2-51% reduction in microbial biomass C with the application of elemental S fertilizer in grass field (Gupta et al., 1988). Studies have shown that the application of S fertilizer can increase soil acidity (Fageria et al., 2010; Kissel et al., 2020). However, in contrast, Wiedenfeld (2011) reported that an increase in salinity level was directly correlated with the increasing rate of S fertilizer. Application of ammonium sulfate can result in increased salinity because of higher salt index (3.25) of this fertilizer (Bunt, 1988).

Application of high rates of fertilizers can also cause accumulation and concentration of mineral salts that lead to soil compaction and resistance to root penetration (Massah & Azadegan, 2016). However, inorganic fertilizers (e.g., N, P, and K) were found to have no impact on soil bulk density and SOC content in long-term studies including corn and wheat (Zhou et al., 2013, 2017). Our recent study on the impact of N fertilizers on carinata and camelina (*Camelina sativa* L.) found that there was no significant influence of N fertilizers on SOC (Li et al., 2019). Long-term inorganic N fertilizer application in corn reduced the soil microbial activity and decreased SOC, whereas short-term application of inorganic N fertilizer had limited effects on soil microbial activities (Fauci & Dick, 1994).

Greenhouse gas emissions have an important role in regulating the earth's surface temperature (Zell, 2010); however, higher emissions can make the planet warmer. The United States Environmental Protection Agency (USEPA) (EPA, 2019) reported that agriculture is responsible for 9% of the total GHG emissions in the United States in 2017. Nitrogen fertilizer is one of the main sources of GHG emissions by influencing the processes of microbial decomposition and root respiration (Kim et al., 2014; Li, Watson, et al., 2013; Li et al., 2019; Mbonimpa et al., 2015; Ozlu & Kumar, 2018). There are few data examining the effect of S fertilizer on GHG emissions from oilseed crops. Studies in Canada (Gupta et al., 1988) and China (Wang et al., 2008) reported that the application of S fertilizer reduces microbial biomass and soil pH from grass fields, which can impact the soil GHG emissions. Our previous study by Li et al. (2019) found that increasing N fertilizer application (0-84 kg N ha⁻¹) in carinata resulted in higher N₂O emissions. This research further

extended the study of Li et al. (2019) by evaluating the impact of N (>84 kg N ha⁻¹) and S fertilizer applications to carinata on GHG emissions. The N rate to carinata did not impact SOC and total N (TN) in our previous study (Li et al., 2019) as changes in these parameters due to management practices occur very slowly (Purakayastha et al., 2008; Zhong et al., 2014). Therefore, the response of labile and stable soil C and N to different N and S fertilizer rates applied to carinata were evaluated in the current study. Specific objective of this study was to assess the impacts of different N and S fertilizer rates applied to carinata on soil properties including soil bulk density (BD), pH, electrical conductivity (EC), SOC, TN, stable and labile C and N, and surface GHG fluxes.

2 | MATERIALS AND METHODS

2.1 | Site description and treatment details

The study was conducted at Aurora Agricultural Experiment Station near Brookings, South Dakota (44°18'35"N, 96°40'15.9"W) during 2017 and 2018. The experiment was established on Brandt series soils characterized by fine silty, super active, frigid calcic hapludolls (Malo, 2003; Soil Survey Staff, 2017). Carinata fits well in rotation with wheat and other small grains. South Dakota has over 800,000 ha (USDA-NASS, 2012) dedicated to wheat production therefore, excellent potential land for carinata production in rotation with wheat. Therefore, the present study was established on winter wheat stubble each year but with different fields used in 2017 and 2018. The average soluble salts at the experimental site in 2017 was 100 µS cm⁻¹. The pH of the soil was found to be acidic (pH = 5.6), and P, K, and S levels were 10 mg kg⁻¹, 141 mg kg⁻¹, and 9.0 kg ha⁻¹, respectively, with 47 g kg⁻¹ organic matter. The pH of the soil in 2018 site was 5.7 with 53 g kg⁻¹ organic matter, and P and K levels were 21 and 220 mg kg⁻¹, respectively. The planting dates of carinata in 2017 and 2018 were 24 April and 8 May, respectively. Planting was done using a seven-row Hege 500[®] (Wintersteiger-Austria) at 22 cm row spacing. After crop emergence, Poast (Sethoxydim, BASF, Research Triangle, NC) herbicide was applied at the rate of 2.1 L ha⁻¹ 4 weeks after planting to control grassy weeds. Broadleaf weeds were managed by manually removing weeds from each plot as required. Carinata was harvested in August and September in 2017 and 2018, respectively. According to US Climate Data 2020, the 30 year (1980-2010) annual mean precipitation was 617 mm and mean high and low temperature were 12.2°C and 0°C, respectively, whereas the mean precipitation, high and low temperature during the cropping season (April–September) were 496 mm, 22.2°C, and 9.6°C, respectively.

-WILEY

The experimental plots were 1.62 m wide by 9.14 m long, arranged in a randomized complete block design (RCBD) with 12 treatments and three replications. Study treatments included four different rates of N fertilizer: 56, 84, 112, and 140 kg N ha⁻¹, and three different rates of S fertilizer: 0 (control), 22, and 45 kg S ha⁻¹ arranged in a factorial design to make 12 treatments within each replication for both years. A previous study by Osborne et al. (2019) has determined the optimum N rate for carinata is between 60 and 81 kg N ha⁻¹; thus, we used the lowest N rate of 56 kg ha^{-1} and increased the rate at the interval of 28 up to 140 kg N ha⁻¹. The optimum S fertilizer required to canola, crop similar to carinata, was around 20 kg S ha^{-1} (Jackson, 2000); thus, based on this study, we selected three different S fertilizer rates. Both urea (46-0-0) and ammonium sulfate (21-0-0-24) were broadcast in two-equal spilt doses: first half immediately after planting (24 April 2017 and 8 May 2018) and the next at bolting stage (13 June 2017 and 22 June 2018).

2.2 | Soil surface GHG monitoring

Soil GHG fluxes were monitored for the whole growing season of carinata in 2017 and 2018. Nine treatments (a factorial combination of 56, 112, and 140 kg N ha⁻¹ and 0, 22, and 45 kg S ha^{-1}) in three replications were selected for monitoring the GHG emissions. The whole experimental plots were not selected for gas sampling because of resources and time limitations and the study was focused on determining treatment differences; thus, we selected the lowest N rate and two highest N rate treatments. Polyvinyl chloride (PVC) static chambers (25 cm diameter and 15 cm height) were installed in each plot to monitor soil surface GHG fluxes, according to the method of Parkin and Venterea (2010). Gas samples were taken once or twice a week depending on weather conditions from June to August in 2017, and from May to August in 2018. During the GHG sampling, a lid (10 cm height with ports for gas sampling) was placed on top of the anchor. Gas samples were collected three times (0, 20, and 40 min) over an hour period by inserting a needle attached to a 10 ml syringe into the port and transferring the gas to pre-evacuated 10 ml vials. Gas chromatograph was used to measure the concentrations of CO₂, N₂O, and CH₄ for each sample. Soil GHG fluxes were calculated as the change in headspace gas concentration over time within the enclosed chamber volume (Hutchinson & Mosier, 1981; Ussiri & Lal, 2009). During each gas-sampling event, soil temperature at the 0-5 cm depth was measured with a thermometer (Taylor 14769 Digital 0.7" Lcd Folding Thermometer). The volumetric soil moisture content (%) at 0-5 cm depth was also measured at the time of gas sampling using a thetaprobe moisture sensor (Delta-T-Devices).

2.3 | Soil sampling

WILEY-GC

Soil samples were collected from the experimental site in August 2017 and November 2018 at 0-5, 5-15, 15-30, and 30–45 cm depths using a soil probe (3.2 cm diam.). For the analysis of pH and electric conductivity (EC), soil samples were air-dried and ground to pass through a 2 mm sieve, and pH and EC (1:2.5 soil/water) were determined as described by McLean (1982). Total C and N were measured by dry combustion using TruSpec carbon-hydrogen-nitrogen analyzer (LECO Corporation). Soil samples had low pH (≤ 6); therefore, measured TC was considered equal to SOC (Guo et al., 2016). Soil C and N fractions were analyzed using cold water extraction (for labile C and N) and hot water extraction (for stable C and N) methods (Ghani et al., 2003). A TOC-L analyzer (Shimadzu Corporation, model-TNM-L-ROHS) was used to determine the cold-water and hot-water C (CWC and HWC) and N (CWN and HWN) fractions. Soil bulk density was determined using the core method (Grossman & Reinsch, 2002).

2.4 | Statistical analysis

The impacts of N and S fertilizers applied to carinata on selected soil parameters in 2017 and 2018 were analyzed using PROC MIXED method. In this analysis, the year, N fertilizer, S fertilizer, and their interactions were considered fixed effects and replication as a random effect. No significant interactions were found; therefore, only main effects are discussed in this paper. There were no significant differences among soil properties between depths; hence, average values from 0 to 45 cm depth were compared. Statistical comparisons of soil BD, pH, EC, SOC, TN, C-fractions, and N-fractions between N and S fertilizers rates were obtained using pairwise differences method (adjusted by Tukey) in a mixed model approach using GLIMMIX procedure. The repeated measures analysis for comparing the soil CO_2 , CH_4 , and N_2O fluxes under different N and S rates was conducted using PROC MIXED, with N and S fertilizers and year as fixed effects, replication as a random effect, and date of gas sampling as a repeated measure variable. All statistical analyses were conducted using SAS 9.4 at a significance level of $\alpha = 0.05$.

3 | RESULTS

3.1 | Weather conditions, soil moisture, and temperature

Air temperature and rainfall data throughout the crop growing period at the site in 2017 and 2018 are shown in Figure 1. During the critical crop growth period (June-July), temperature often exceeded 25°C in both years. In 2017, the experimental site received a total rainfall of 377.2 mm over the crop-growing season (April-August); most of the precipitation was received in July and August (Figure 1). The early days of June 2017 did not receive significant rainfall but were followed by scattered rainfall mid-month. Unlike 2017, the 2018 year was a wet year where the total amount of rainfall during the growing period was 442.9 mm; on a single day in mid-July, the total rainfall was 150 mm (Figure 1). Soil temperature and moisture for different N and S rates during the crop growing period are presented in Figure 2. The soil temperature trend in the 2017 growing season was similar to that of 2018 under N and S fertilizer rates (Figure 2a,b). Soil moisture was lower in the 2017 (average 16.9%) season compared to the 2018 (average 18.4%) for N and S fertilizer rates ($p \le 0.05$, Figure 2c,d). Soil temperature and moisture (Figure 2) were not affected by N and S fertilizer rates in 2017 and 2018 (averaged across measurement dates in each year, p > 0.05).

3.2 | Soil properties



Data showed that none of the studied treatments significantly influenced soil bulk density in either year (Tables 1 and 2). The mean soil pH was acidic, but was not influenced by

FIGURE 1 Daily maximum and minimum air temperature and precipitation in 2017 and 2018 for the study site. Orange line represents the maximum air temperature, dotted line represents the minimum air temperature, and blue bar represents the precipitation

FIGURE 2 Average soil temperature of plots applied with different (a) N rates, and (b) S rates and average soil moisture of plots applied with different (c) N rates and (d) S rates over the observed days in 2017 and 2018



TABLE 1 Means of soil bulk density (BD), pH, electrical conductivity (EC), soil organic carbon (SOC), total nitrogen (TN), and labile and stable C and N as influenced by different N and S fertilizers rates to carinata for the 0–45 cm depth in 2017

	BD (g cm ⁻³)	рН	EC (µS cm ⁻¹)	SOC (g kg ⁻¹)	TN (g kg ⁻¹)	Labile C (mg kg ⁻¹)	Stable C (mg kg ⁻¹)	Labile N (mg kg ⁻¹)	Stable N (mg kg ⁻¹)
$N \ rates \ (kg \ ha^{-1})$									
56	1.4^{\dagger}	5.4	322.4 ^b	24.5 ^b	2.7	288.6	936.3 ^b	50.0 ^b	30.5
84	1.4	5.4	319.2 ^b	25.1 ^{ab}	2.7	307.9	992.5 ^{ab}	54.4 ^b	33.1
112	1.4	5.4	368.6 ^a	26.0 ^a	2.7	311.1	1071.9 ^a	90.0 ^a	35.8
140	1.4	5.3	370.0 ^a	26.0 ^a	2.9	297.1	1020.4 ^a	67.5 ^{ab}	34.1
	Analysis of variance $(p > F)$								
	0.306	0.137	0.009	0.041	0.058	0.573	0.012	0.013	0.114
S rates	$(kg ha^{-1})$								
0	1.4^{\dagger}	5.3 ^b	369.2 ^a	25.2	2.7	286.1	1016.2	63.1	33.4
22	1.4	5.5 ^a	329.9 ^b	25.7	2.7	306.7	967.6	68.5	32.6
45	1.4	5.5 ^a	336.1 ^b	25.3	2.8	308.2	1028.2	62.2	33.9
	Analysis of variance $(p > F)$								
	0.363	0.016	0.007	0.634	0.241	0.535	0.103	0.991	0.487

[†]Mean values followed by different lowercase letters within the column represent significant differences due to the treatments at $p \le 0.05$. Values are the average values for 0–5, 5–15, 15–30, and 30–45 cm depth soil samples as there was no significant difference between the depths.

different N fertilizer treatments in either year (Tables 1 and 2). However, soils with S fertilizer applied at the rate of 22 and 45 kg S ha⁻¹ in 2017 had significantly higher pH values than 0 kg S ha⁻¹ (Table 1). Application of S fertilizer in 2018 did not affect pH (Table 2). Mean EC at 112 and $140 \text{ kg N} \text{ ha}^{-1}$ in 2017 were significantly higher than the 56 and 84 kg N ha⁻¹ (Table 1). While the application of 22 and 45 kg S ha⁻¹ reduced the EC by 12% and 10%, respectively, compared to S control soils in 2017 (Table 1), and S control plot had significantly lower EC compared to higher rates in 2018 (Table 2). For SOC, 140 and 112 kg N ha⁻¹ increased SOC more than the 56 kg N ha⁻¹; however, 84 kg N ha⁻¹ had a similar concentration compared to all other rates in 2017 (Table 1). As seen in Table 2, N fertilizer rates did not show any significant impact on SOC in 2018. Sulfur fertilizer rates showed a significant impact only in 2018, when

the application of 22 and 45 kg S ha⁻¹ had 3.2% and 1.8% lower SOC than the S control soils, respectively (Table 2). Nitrogen and S fertilizers did not influence total soil N in either year (Tables 1 and 2). Thus, total N results are not discussed here. Data on C and N pools showed significant impact of N fertilizers only on stable C and labile N in 2017, whereas these parameters were not influenced in 2018 (Tables 1 and 2). Stable C in 2017 was higher under 140 and 112 kg N ha⁻¹ than 56 kg N ha⁻¹ but similar to 84 kg N ha⁻¹ (Table 1). Labile N in 2017 was higher under 112 kg N ha⁻¹ compared to 84 and 56 kg N ha⁻¹ (Table 1). Data on C and N pool were non-significant to S fertilizers in 2017 (Table 1). In 2018, however, soil in the 45 kg S ha⁻¹ had 17% lower concentration of labile N compared to S control soil, which was similar to 22 kg S ha⁻¹ (Table 2).

Y- GCB-BIOENERGY

TABLE 2 Means of soil bulk density (BD), pH, electrical conductivity (EC), soil organic carbon (SOC), total nitrogen (TN), and labile and stable C and N as influenced by different N and S fertilizers rates to carinata for the 0–45 cm depth in 2018

	BD (g cm ⁻³)	рН	EC (µS cm ⁻¹)	SOC (g kg ⁻¹)	TN (g kg ⁻¹)	Labile C (mg kg ⁻¹)	Stable C (mg kg ⁻¹)	Labile N (mg kg ⁻¹)	Stable N (mg kg ⁻¹)
N rates (kg ha ⁻¹)									
56	1.4	5.6	123.2	28.6	3.1	105.5	420.7	21.9	43.2
84	1.4	5.7	120.3	29.0	3.1	110.8	436.2	23.8	46.6
112	1.5	5.6	126.5	28.3	3.0	106.5	418.8	23.3	43.7
140	1.5	5.7	125.9	28.4	3.1	100.8	394.6	23.9	41.9
	Analysis of variance $(p > F)$								
	0.804	0.401	0.656	0.266	0.478	0.167	0.180	0.427	0.173
S rates	$(kg ha^{-1})$								
0	1.5	5.6	116.0 ^{b†}	29.0 ^a	3.1	108.6	424.4	24.2 ^a	45.4
22	1.4	5.7	129.7 ^a	28.1 ^b	3.1	108.9	418.6	25.5 ^a	44.6
45	1.5	5.6	126.6 ^a	28.5 ^b	3.0	100.7	411.6	20.1 ^b	41.9
	Analysis of variance $(p > F)$								
	0.908	0.281	0.027	0.011	0.254	0.091	0.531	0.001	0.106

[†]Mean values followed by different lowercase letters within the column for N and S rates represent significant differences due to the treatments at $p \le 0.05$. Values are the average values for 0–5, 5–15, 15–30, and 30–45 cm depth soil samples as there was no significant difference between the depths.

3.3 | Soil GHG fluxes

Data on daily mean soil GHG fluxes from carinata crop managed with different N and S fertilizers rates over the observed days in 2017 and 2018 are presented in Figure 3. For N fertilizer, CO₂ peaks were observed on 16 June and 13 July in 2017 (Figure 3). No significant differences on CO2 fluxes were observed among N fertilizer rates during these two peaks in this year (p > 0.05,analysis not shown in the figure). The largest difference among N fertilizer rates in CO₂ fluxes was observed on 6 July 2017, with 56 kg N ha⁻¹ recording lower CO_2 flux (16.28 kg CO₂-C ha⁻¹ day⁻¹) than 140 kg N ha^{- $\overline{1}$} $(24.04 \text{ kg CO}_2\text{-C ha}^{-1} \text{ day}^{-1})$ and 112 kg N ha^{-1} $(21.12 \text{ kg CO}_2\text{-C ha}^{-1} \text{ day}^{-1}; p < 0.001, \text{ Figure 3}).$ In 2018, the highest CO_2 fluxes were observed on 24 June and 6 July, with 140 and 112 kg N ha⁻¹ recording higher CO_2 fluxes than 56 kg N ha⁻¹ (p < 0.001, Figure 3). The peaks of CO₂ flux under S fertilizer rates were observed on 16 June and 13 July in 2017; however, no significant differences on CO2 fluxes were observed among S fertilizer rates during these two peaks (p > 0.05). In 2018, soil CO₂ fluxes increased from May to the beginning of July and decreased thereafter under all S fertilizer rates (Figure 3). The fluxes of CO_2 were similar among S fertilizer rates at all sampling dates in 2018 (p > 0.05).

For N_2O fluxes, the peaks in 2017 were recorded on 16 June, 19 and 29 July, with 112 and 140 kg N ha⁻¹ recording higher N_2O than 56 kg N ha⁻¹ in all peaks

(p < 0.001, Figure 3). In 2018, the highest daily peak was seen on 24 June (Figure 3). Again, 112 and 140 kg N ha⁻¹ emitted higher N₂O than 56 kg N ha⁻¹ during this peak (p < 0.001, Figure 3). The peaks of N₂O fluxes were also observed under S fertilizer treatment on 16 June, 19 and 29 July 2017, and 24 June, 6 and 24 July 2018, but no significant differences on N₂O flux among treatments were recorded in either year at any sampling dates (p > 0.05, Figure 3). The trend of the CH₄ fluxes under N and S fertilizers varied on the sampling dates over the 2 years (Figure 3). There were no clear seasonal patterns over the 2 years under all treatments (Figure 3). However, no significant effects of N and S fertilizer rates on the daily CH₄ fluxes were observed during the sampling dates for both years (p > 0.05).

Data on the seasonal means of CO₂, N₂O and CH₄ fluxes as influenced by different N and S fertilizer treatments are presented in Table 3. In 2017, application of N fertilizer rates significantly influenced CO₂ flux, with 140 kg N ha⁻¹ recording higher CO₂ flux than 56 kg N ha⁻¹; however, the flux at 112 kg N ha⁻¹ was similar with that of both 56 and 140 kg N ha⁻¹ (Table 3). No significant differences on CO₂ fluxes due to N fertilizer rates were observed in 2018 (Table 3). The fluxes of N₂O were lower in the plots with 56 kg N ha⁻¹ than 112 and 140 kg N ha⁻¹ in 2017 (Table 3). Like CO₂ flux, no significant differences in N₂O fluxes due to N fertilizer rates were observed in 2018 (Table 3). Methane fluxes were unaffected by N and fertilizer rates in both years (Table 3).





633



FIGURE 3 Daily mean soil CO_2 , N_2O , and CH_4 fluxes from carinata crop managed with different N fertilizer rates (a, b, c, respectively) and S fertilizer rates (d, e, f, respectively) over the observed days in 2017 and 2018

TABLE 3	Means of CO ₂ , N ₂ O, and CH	fluxes as influenced by different N as	and S fertilizer rates to carinata in 2017 a	and 2018
---------	---	--	--	----------

	2017			2018			
	$ \frac{\text{CO}_2}{(\text{kg C ha}^{-1} \text{day}^{-1})} $	N_2O (g N ha ⁻¹ day ⁻¹)	CH ₄ (g C ha ⁻¹ day ⁻¹)	CO ₂ (kg C ha ⁻¹ day ⁻¹)	N ₂ O (g N ha ⁻¹ day ⁻¹)	CH ₄ (g C ha ⁻¹ day ⁻¹)	
$N \ rates \ (kg \ ha^{-1})$							
56	24.8 (±5.2) ^{b†}	$6.5 (\pm 1.2)^{b}$	9.6 (±3.4)	16.8 (±3.9)	6.3 (±1.1)	8.9 (±3.4)	
112	$26.4 (\pm 5.1)^{ab}$	$9.4(\pm 2.4)^{a}$	10.4 (±3.1)	17.3 (±5.7)	7.1 (±1.9)	9.9 (±2.9)	
140	27.2 $(\pm 5.1)^{a}$	$8.5 (\pm 2.8)^{a}$	12.5 (±2.7)	18.1 (±7.2)	7.6 (±2.0)	9.1 (±3.1)	
	Analysis of variance $(p > F)$						
	0.015	<0.001	0.118	0.954	0.109	0.623	
$S \text{ rates } (kg ha^{-1})$							
0	26.4 (±4.2)	8.3 (±2.4)	11.3 (±2.6)	17.7 (±6.1)	7.9 (±2.9)	10.9 (±3.7)	
22	26.3 (±5.7)	8.0 (±1.6)	11.0 (±2.5)	16.9 (±5.1)	6.8 (±1.5)	8.1 (±3.1)	
45	25.7 (±5.1)	8.0 (±1.9)	10.1 (±2.6)	16.9 (±4.5)	6.5 (±1.5)	9.3 (±3.2)	
	Analysis of variance $(p > F)$						
	0.487	0.893	0.803	0.708	0.054	0.060	

Note: Standard error values (\pm) are shown in the parentheses.

[†]Mean values followed by different lowercase letters within the column for N and S rates represent significant differences due to the treatments at $p \le 0.05$.

4 | DISCUSSION

4.1 | Soil moisture and temperature

Increasing the N rate from 56 to $140 \text{ kg N} \text{ ha}^{-1}$ in this study did not affect soil moisture and temperature. In line with the results

of current study, our previous study reported no impact of N fertilizer applications $(0-84 \text{ kg N ha}^{-1})$ to carinata did not affect soil temperature and moisture for 3 years which was attributed to the short growing season of the carinata crop (Li et al., 2019). Similarly, nitrogen fertilizer application $(0-112 \text{ kg ha}^{-1})$ to switchgrass (*Panicum virgatum* L.) did not affect soil moisture

and temperature in South Dakota (Mbonimpa et al., 2015). Conversely, Sainju, Caesar-TonThat, et al. (2012) reported that the N fertilization to malt barley (Hordeum vulgaris L.) decreased soil temperature compared to the control (no fertilizer). This difference in the effect of N fertilization rates on soil temperature between our study and Sainju, Caesar-TonThat, et al. (2012) can be attributed to the differences in crops used in each study. No differences on soil moisture and temperature were observed among S fertilizer treatments in this study. The impact of S fertilizer rates on soil moisture and temperature has not been reported in the literature yet. Bulk density can impact the soil pores and hence the soil moisture. In general, a higher bulk density can reduce the soil pores and the moisture. In this study, bulk density was similar under all N and S fertilizer rates in both years (Tables 1 and 2), perhaps explaining why similar soil moisture were recorded among the treatments in this study.

4.2 Soil properties

Data showed that there was no significant impact of N and S fertilizer rates on soil bulk density, which can be attributed to the short duration of the study (2 years). Longterm application of fertilizers may affect soil bulk density (Blanco-Canqui et al., 2015). Singh et al. (2019) reported that a 10-year application of 112 kg N ha⁻¹ to switchgrass decreased soil bulk density compared to the control (no fertilizer). The increased biomass production and residue retention on the soil under high N fertilization can increase SOC and reduce soil bulk density (Wagner et al., 1994). Similar to our findings, a study of maize crop in China Zhong et al. (2014) reported non-significant impacts of different rates of NPK fertilizers including urea on soil porosity and bulk density. Similarly, even a 17-year study on chemical fertilization (N and P fertilizers) in China did not show any significant impact on soil bulk density (Li, Xu, et al., 2013).

Nitrogen fertilizer rate did not affect soil pH in either year. Excess N fertilizer in soil is associated with H⁺ ion formation (Barak et al., 1997; Zhang et al., 2017), which was not observed in this study. Bryla et al. (2008) reported ammonium-releasing fertilizers lower the pH gradually, and so their effect might not be detectable in a short period. Similar results for the soil pH in response to different N rates were reported by a study on switchgrass (Mbonimpa et al., 2015). However, the change in pH with S fertilizer application might be due to H⁺ ions being released after microbial conversion of ammonium to nitrate (nitrification; Barak et al., 1997). One molecule of ammonium sulfate can release four H⁺ ions (IPNI, 2019) in the soil, which lowers the soil pH; however, our results are contradictory to this, which might be due to the release of hydroxide ions in soil during the nutrients uptake by plants. When plants

uptake NO₃-N, it equals or exceeds the uptake of potassium, calcium, and magnesium, and leading to proton consumption by plants and thus, hydroxide is released in the soil (Jaillard et al., 2003). Our results were similar to the findings from Wiedenfeld (2011) where the application of S fertilizer increased soil salinity. High soil water content might have limited the nitrification process, which would prevent the release of H⁺ ions (Tu et al., 2000), thus exhibiting no effect on soil pH from N and S fertilizer rates.

Increasing N fertilizer rates increased the EC of the soil in 2017, supporting the results reported by Liu et al. (2014). Similarly, our previous study reported that increasing N rates increased the soil EC, which was associated with the increase in the salts due to the fertilizer application (Li et al., 2019). Both fertilizers, urea and ammonium sulfate, serve as the source of N in the soil; higher salt index of ammonium sulfate (3.25) might have increased the salt concentration in the soil. Our result was supported by the findings by Han et al. (2015) and Gandois et al. (2011). The increase in soil EC with higher S fertilizer rate in 2018 may be due to the higher concentration of ammonium sulfate (Bunt, 1988). The addition of ammonium sulfate linearly increased the saturated EC (Bryla et al., 2008). Soil EC increases with the application of S fertilizer (SO_4^{-}) , as reported by other researchers (Hashemimajd et al., 2012; Turan et al., 2013). Soil EC decreased with increasing S fertilizer rates in 2017, which is not common and thus, additional measurements are needed to determine this impact.

The current study found increased SOC with increasing use of N fertilizer. When N is a limiting factor, N fertilizer can impact SOC mineralization through influencing microbial activity and plant biomass (Chen et al., 2014). Sekaran et al. (2019) reported that increasing N rates in soil seeded with switchgrass increased the activity of urease and β -glucosidase enzymes. Other studies reported an increase in SOC with increased N fertilizer rates (Ghimire et al., 2017; Li et al., 2015). Increase in SOC might also have resulted from higher crop biomass and C sequestration (Giacometti et al., 2013). Carinata produced higher biomass under higher N rate (data not shown), which may have added organic carbon to the soil through microbial activity (Seepaul et al., 2016). A study in continuous corn and corn-soybean rotation (Poffenbarger et al., 2017) stated that application of N fertilizer increased SOC until an optimum N rate, above which SOC storage was not affected. Analogous to this study, SOC increased until the application of 112 kg N ha⁻¹ and remained constant with further increase in N fertilizer rate in our present study. The application of S fertilizer in 2018 reduced the SOC compared to the S control plots, which might be due to the acidic effect of sulfate in 2017 causing reduced enzyme activity (Lv et al., 2014). The presence of high H⁺ ions can inhibit soil microbial activity (Chen et al., 2012). A study in Canada by Gupta et al. (1988) showed that the application of S fertilizer reduced microbial biomass.

A study reported that increase in stable C with higher rates of N fertilizer might be due to the residue decomposition accelerated by higher microbial activity (Alvarez, 2005). Our results of increased stable C with higher N fertilizer rates are in line with the previous study. Water extractable labile N is the source of energy for soil microbial activity (Zhang et al., 2016), which increased with increasing N fertilizer rate in 2017. However, labile N decreased in 2018 with the application of S fertilizer, which might be attributed to the reduced SOC. The non-significant influence on other C and N fractions is analogous to the data reported by Singh et al. (2019) in their study of switchgrass in South Dakota.

4.3 | Daily and seasonal GHG fluxes

The CO₂ fluxes varied with date of sampling, peaked after short precipitation and fertilizer application. Similar findings were also reported by Sainju, Stevens, et al. (2012). Our results showed that the CO_2 fluxes peaked on 16 June in 2017 might probably have resulted from the similar phenomenon as explained by previous studies (Sainju, Stevens, et al., 2012). There were similar activities including increased soil temperature, soil moisture (precipitation from 10 to 14 June), and fertilizers application (13 June) that might have increased the CO₂ peak. Peaks on 13 July, 2017 resulted from precipitation on 12 July and higher soil temperature (Figures 1 and 2). In 2018, the peaks in June and July might have resulted from fertilizers application (22 June) and continuous rainfall from 17 June to 21 June. A number of studies have reported significant effects of soil temperature, soil moisture, and fertilizer application on CO₂ fluxes (Abagandura, Şentürklü, et al., 2019; Davidson et al., 1998; Mbonimpa et al., 2015; Schaufler et al., 2010; Soosaar et al., 2011). Similar to our results, application of N and S increased CO₂ emissions in a study by Hu et al. (2017), which might be due to increased root respiration with the application of exogenous nutrients.

The N₂O peaks in June and July in 2017 were a response to N and S fertilizers, higher temperature, and soil moisture resulting from heavy rainfall (Figures 1 and 2), which may trigger N mineralization and the nitrification/denitrification process, leading to higher N₂O emissions. Similar findings were also reported by Chatskikh and Olesen (2007). Similar to CO₂ fluxes, the N₂O peaks in 2018 were observed on 24 June, probably due to fertilizer application (22 June) and continuous rainfall (17–21 June), resulting in higher soil moisture and temperature (Figures 1 and 2). These conditions may induce the nitrification/denitrification process, resulting in high N₂O emission. Many studies reported loss of applied N because of the increased stimulatory response of N₂O emissions 635

-WILEY-

(Drury et al., 2014; Omonode et al., 2011; Pelster et al., 2011). Application of ammonium sulfate releases NH_4^+ , which is the substrate for nitrification and the aerobic source of N₂O (Deppe et al., 2016); however, N₂O fluxes often peak under oxygen-limited denitrifying (high soil moisture, fertilization) conditions (Drury et al., 2006; Pelster et al., 2012).

Soil CH₄ fluxes were very low for both years in this study. Several studies have reported low soil CH₄ fluxes under aerobic soil conditions (Abagandura, Şentürklü, et al., 2019; Li et al., 2019). Methane flux was positive on most of the sampling dates in both years, perhaps due to heavy and continuous rainfall in both years (Figure 1), resulting in high soil moisture (Figure 2), which, in turn, may have produced an anaerobic condition. Release or uptake of CH₄ depends on the microbes present in the soil. The anaerobic microbes release CH₄ and increase these fluxes, while aerobic microbes uptake CH₄, and result in negative flux (Abagandura, Chintala, et al., 2019). Higher amounts of crop residues (senesced leaves of carinata) at the maturity stage, in combination with moisture and higher temperature, might have triggered the decomposition process, which is the substrate for CH₄ production (Ding et al., 2004).

The increase in mean seasonal CO₂ flux with increasing rate of N fertilizer in 2017 may be due to higher microbial activity, which increases C and N mineralization, as discussed earlier. The SOC, stable C, and labile N significantly increased with increasing N fertilizer rates, which may cause this increase in CO₂ flux with increasing N fertilizer rate. Similar to our results, Sainju et al. (2008) reported a 14% increase in CO₂ fluxes in N-applied soils compared to the N control soils in North Dakota. The present research results have concurred with many other studies (Hu et al., 2017; Jiang et al., 2010). In contrary, a study by Mbonimpa et al. (2015) reported a reduction in CO₂ fluxes in plots applied with N fertilizer due to the combination of lower SOC, lower porosity, and higher bulk density.

Nitrogen fertilizer increased mean seasonal N2O flux in 2017, which may be due to N mineralization and the nitrification/denitrification process, depending on the environmental conditions throughout the crop-growing period. Our results of seasonal N₂O fluxes agree with other studies (Drury et al., 2014; Dusenbury et al., 2008). In 2018, there was no significant influence of either fertilizer on CO2, and N2O fluxes; higher precipitation might have resulted in nitrate leaching, which could reduce the amount of volatilizing N and, in addition, higher precipitation might have reduced the soil air space by filling soil pores with water. In addition, soil properties were least affected by the fertilizer treatments in 2018. Sulfur fertilizer rates did not affect CO2 and N2O fluxes in both years in this study, suggesting that increasing S fertilizer rate from 0 to 45 kg S ha⁻¹ in carinata field for 2 years will not increase the flux of these gases under similar conditions.

The lack of effect of N and S fertilization rates on CH₄ fluxes in this study was attributed to the similar soil moisture

636

-WILEY-GCB-BIOENERGY

among treatments. Several studies reported no significant effect of N fertilizer rate on soil CH_4 fluxes compared with the control from bioenergy crops in the dryland cropping system (Abagandura, Chintala, et al., 2019; Li et al., 2019; Mbonimpa et al., 2015). Management practices usually have little effect on CH_4 flux in the upland agricultural soils, as reported in our previous studies (e.g., Abagandura, Şentürklü, et al., 2019; Ozlu & Kumar, 2018).

5 | CONCLUSIONS

This study demonstrated the impact different rates of N and S fertilizers applied to carinata on soil properties and greenhouse gas fluxes in Brookings, South Dakota in 2017 and 2018. The growing season of 2018 was a comparatively wet year with heavy precipitation. Data from this study showed mixed impacts of N and S fertilizers on soil parameters. In 2017, N fertilizer rates significantly increased soil EC, SOC, stable C, and labile N, whereas the S fertilizer rates decreased the soil acidity and EC. In 2018, application of S fertilizer increased soil EC but decreased the SOC. The application of N fertilizer at higher rates significantly increased CO₂ and N₂O emissions only in 2017. No significant influence of S fertilizer application was found on GHG emissions in either year. We can conclude from this study that, in general, sulfur fertilizer at a rate of $0-45 \text{ kg S} \text{ ha}^{-1}$ did not affect GHG fluxes but required for the carinata crop growth. Furthermore, application of N fertilizer did not impact soil properties, however, higher N fertilizer rate (140 kg ha⁻¹) increased soil surface CO₂ and N₂O fluxes. This study suggests that the application of N fertilizer beyond 112 kg N ha⁻¹ can increase soil surface GHG emissions, and this N rate with 22 kg S ha⁻¹ needed for the carinata growth with minimal GHG emissions.

ACKNOWLEDGEMENTS

The project was funded by the South Dakota Oilseeds Initiative, the North Central Regional Sun Grant Center at South Dakota State University through a grant provided by the US Department of Agriculture under award number 2014-38502-22598 and USDA-NIFA. We thank Nathan Braun, Christopher Owusu, and Reshma Thapa for their help in the fieldwork; Agrisoma USA for supplying carinata seed; and the South Dakota Agricultural Experiment Station for support.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

DATA AVAILABILITY STATEMENT

The data that support the findings of the present study are available from the corresponding author upon reasonable request.

ORCID

Dwarika Bhattarai Dhttps://orcid. org/0000-0002-9241-693X Sandeep Kumar Dhttps://orcid.org/0000-0002-2717-5455

REFERENCES

- Abagandura, G. O., Chintala, R., Sandhu, S. S., Kumar, S., & Schumacher, T. E. (2019). Effects of biochar and manure applications on soil carbon dioxide, methane, and nitrous oxide fluxes from two different soils. *Journal of Environmental Quality*, 48, 1664–1674.
- Abagandura, G. O., Şentürklü, S., Singh, N., Kumar, S., Landblom, D. G., & Ringwall, K. (2019). Impacts of crop rotational diversity and grazing under integrated crop-livestock system on soil surface greenhouse gas fluxes. *PLoS One*, 14(5), e0217069.
- Abdallah, M., Dubousset, L., Meuriot, F., Etienne, P., Avice, J.-C., & Ourry, A. (2010). Effect of mineral sulphur availability on nitrogen and sulphur uptake and remobilization during the vegetative growth of *Brassica napus* L. *Journal of Experimental Botany*, *61*(10), 2635–2646. https://doi.org/10.1093/jxb/erq096
- Agrisoma-USA. (2019). Southeastern US 2017–18 carinata management handbook. https://agrisoma.com/wp-content/uploads/2018/ 10/2017_18_SE_Handbook.pdf
- Agrisoma-USA. (2020). Grow carinata. https://growcarinata.com/
- Alvarez, R. (2005). A review of nitrogen fertilizer and conservation tillage effects on soil organic carbon storage. Soil Use and Management, 21(1), 38–52.
- Barak, P., Jobe, B. O., Krueger, A. R., Peterson, L. A., & Laird, D. A. (1997). Effects of long-term soil acidification due to nitrogen fertilizer inputs in Wisconsin. *Plant and Soil*, 197(1), 61–69.
- Blanco-Canqui, H., Hergert, G. W., & Nielsen, R. A. (2015). Cattle manure application reduces soil compactibility and increases water retention after 71 years. *Soil Science Society of America Journal*, 79, 212–223.
- Bryla, D. R., Shireman, A. D., & Machado, R. M. (2008). Effects of method and level of nitrogen fertilizer application on soil pH, electrical conductivity, and availability of ammonium and nitrate in blueberry. VI International Symposium on Mineral Nutrition of Fruit Crops, 868, 95–102.
- Bunt, A. (1988). A manual on the preparation and use of growing pot plants. *Media and mixes for container grown plants*. Unwin Hyman Ltd.
- Cardone, M. et al. (2003). Brassica carinata as an alternative oil crop for the production of biodiesel in Italy: Agronomic evaluation, fuel production by transesterification and characterization. Biomass and Bioenergy, 25(6), 623–636.
- CFIA. (2017). The biology of Brassica carinata (A.) Braun (Abyssinian cabbage). Author. http://www.inspection.gc.ca/ plants/plants-with-novel-traits/applicants/directive-94-08/biolo gy-documents/brassica-carinata/eng/1501087371874/15010 87468251#a43
- Chatskikh, D., & Olesen, J. E. (2007). Soil tillage enhanced CO₂ and N₂O emissions from loamy sand soil under spring barley. *Soil and Tillage Research*, 97(1), 5–18.
- Chen, R., Senbayram, M., Blagodatsky, S., Myachina, O., Dittert, K., Lin, X., & Kuzyakov, Y. (2014). Soil C and N availability determine the priming effect: Microbial N mining and stoichiometric decomposition theories. *Global Change Biology*, 20(7), 2356–2367.

- Chen, S., Shen, X., Hu, Z., Chen, H., Shi, Y., & Liu, Y. (2012). Effects of simulated acid rain on soil CO₂ emission in a secondary forest in subtropical China. *Geoderma*, 189-190, 65–71. https://doi. org/10.1016/j.geoderma.2012.05.002
- Chum, H., Faaij, A., Moreira, J., Berndes, G., Dhamija, P., Dong, H., & Ribeiro, S. (2011). Bioenergy. In O. Edenhofer, R. Pichs-Madruga, Y. Sokona, K. Seyboth, S. Kadner, T. Zwickel, P. Eickemeier, G. Hansen, S. Schlömer, C. von Stechow, & P. Matschoss (Eds.), *Renewable energy sources and climate change mitigation: Special report of the Intergovernmental Panel on Climate Change* (pp. 209–332). Cambridge University Press. https://doi.org/10.1017/ CBO9781139151153.006
- Davidson, E., Belk, E., & Boone, R. D. (1998). Soil water content and temperature as independent or confounded factors controlling soil respiration in a temperate mixed hardwood forest. *Global Change Biology*, 4(2), 217–227.
- Deppe, M., Well, R., Kücke, M., Fuß, R., Giesemann, A., & Flessa, H. (2016). Impact of CULTAN fertilization with ammonium sulfate on field emissions of nitrous oxide. *Agriculture, Ecosystems & Environment*, 219, 138–151. https://doi.org/10.1016/j.agee.2015. 12.015
- Ding, W., Cai, Z., & Tsuruta, H. (2004). Cultivation, nitrogen fertilization, and set-aside effects on methane uptake in a drained marsh soil in Northeast China. *Global Change Biology*, 10(10), 1801–1809.
- Drury, C. F., Reynolds, W. D., Tan, C. S., Welacky, T. W., Calder, W., & McLaughlin, N. B. (2006). Emissions of nitrous oxide and carbon dioxide. *Soil Science Society of America Journal*, 70(2), 570–581. https://doi.org/10.2136/sssaj2005.0042
- Drury, C. F., Reynolds, W. D., Tan, C. S., McLaughlin, N. B., Yang, X. M., Calder, W., Oloya, T. O., & Yang, J. Y. (2014). Impacts of 49–51 years of fertilization and crop rotation on growing season nitrous oxide emissions, nitrogen uptake and corn yields. *Canadian Journal of Soil Science*, 94(3), 421–433. https://doi.org/10.4141/ cjss2013-101
- Dusenbury, M., Engel, R., Miller, P., Lemke, R., & Wallander, R. (2008). Nitrous oxide emissions from a northern great plains soil as influenced by nitrogen management and cropping systems. *Journal of Environmental Quality*, 37(2), 542–550.
- EPA. (2019). Sources of greenhouse gas emissions. https://www.epa. gov/ghgemissions/sources-greenhouse-gas-emissions
- Fageria, N. K., Dos Santos, A. B., & Moraes, M. F. (2010). Influence of urea and ammonium sulfate on soil acidity indices in lowland rice production. *Communications in Soil Science and Plant Analysis*, 41(13), 1565–1575.
- Fauci, M. F., & Dick, R. P. (1994). Soil microbial dynamics: Short- and long-term effects of inorganic and organic nitrogen. *Soil Science Society of America Journal*, 58, 801–806.
- Gandois, L., Perrin, A. S., & Probst, A. (2011). Impact of nitrogenous fertiliser-induced proton release on cultivated soils with contrasting carbonate contents: A column experiment. *Geochimica et Cosmochimica Acta*, 75(5), 1185–1198.
- Ghani, A., Dexter, M., & Perrott, K. (2003). Hot-water extractable carbon in soils: A sensitive measurement for determining impacts of fertilisation, grazing and cultivation. *Soil Biology and Biochemistry*, 35(9), 1231–1243.
- Ghimire, R., Lamichhane, S., Acharya, B. S., Bista, P., & Sainju, U. M. (2017). Tillage, crop residue, and nutrient management effects on soil organic carbon in rice-based cropping systems: A review. *Journal of Integrative Agriculture*, 16(1), 1–15.

<u>GCB-BIOENERGY</u>

- Giacometti, C., Demyan, M. S., Cavani, L., Marzadori, C., Ciavatta, C., & Kandeler, E. (2013). Chemical and microbiological soil quality indicators and their potential to differentiate fertilization regimes in temperate agroecosystems. *Applied Soil Ecology*, 64, 32–48. https://doi.org/10.1016/j.apsoil.2012.10.002
- Grossman, R. B., & Reinsch, T. G. (2002). 2.1 Bulk density and linear extensibility. In J. H. Dane & G. Clarke Topp (Eds.), *Methods of soil analysis*. Wiley. https://doi.org/10.2136/sssabookser5.4.c9
- Guo, Y., Wang, X., Li, X., Wang, J., Xu, M., & Li, D. (2016). Dynamics of soil organic and inorganic carbon in the cropland of upper Yellow River Delta, China. *Scientific Reports*, 6(1), https://doi. org/10.1038/srep36105
- Gupta, V., Lawrence, J., & Germida, J. (1988). Impact of elemental sulfur fertilization on agricultural soils. I. Effects on microbial biomass and enzyme activities. *Canadian Journal of Soil Science*, 68(3), 463–473.
- Han, J., Shi, J., Zeng, L., Xu, J., & Wu, L. (2015). Effects of nitrogen fertilization on the acidity and salinity of greenhouse soils. *Environmental Science and Pollution Research*, 22(4), 2976–2986.
- Hashemimajd, K., Farani, T., & Jamaati-e-Somarin, S. (2012). Effect of elemental sulphur and compost on pH, electrical conductivity and phosphorus availability of one clay soil. *African Journal of Biotechnology*, 11(6), 1425–1432.
- Herath, A., Ma, B., Shang, J., Liu, J., Dong, T., Jiao, X., Kovacs, J., & Walters, D. (2017). On-farm spatial characterization of soil mineral nitrogen, crop growth, and yield of canola as affected by different rates of nitrogen application. *Canadian Journal of Soil Science*. https://doi.org/10.1139/cjss-2017-0024
- Hossain, Z., Johnson, E. N., Wang, L., Blackshaw, R. E., & Gan, Y. (2019). Comparative analysis of oil and protein content and seed yield of five Brassicaceae oilseeds on the Canadian prairie. *Industrial Crops and Products*, 136, 77–86.
- Hu, M., Wilson, B. J., Sun, Z., Ren, P., & Tong, C. (2017). Effects of the addition of nitrogen and sulfate on CH₄ and CO₂ emissions, soil, and pore water chemistry in a high marsh of the Min River estuary in southeastern China. *Science of The Total Environment*, 579, 292–304.
- Hutchinson, G., & Mosier, A. (1981). Improved soil cover method for field measurement of nitrous oxide fluxes 1. Soil Science Society of America Journal, 45(2), 311–316.
- IPNI. (2019). Nutrient source specifics. Author. https://www.ipni.net/ specifics-en
- Jackson, G. D. (2000). Effects of nitrogen and sulfur on canola yield and nutrient uptake. Agronomy Journal, 92(4), 644–649.
- Jaillard, B., Plassard, C., & Hinsinger, P. (2003). Measurements of H+ fluxes and concentrations in the rhizosphere. In Z. Rengel (Ed.), *Handbook of soil acidity* (pp. 231–266). Marcel Dekker, Inc.
- Jiang, C., Yu, G., Fang, H., Cao, G., & Li, Y. (2010). Short-term effect of increasing nitrogen deposition on CO₂, CH₄ and N₂O fluxes in an alpine meadow on the Qinghai-Tibetan Plateau, China. *Atmospheric Environment*, 44(24), 2920–2926.
- Kazemeini, S. A., Hamzehzarghani, H., & Edalat, M. (2010). The impact of nitrogen and organic matter on winter canola seed yield and yield components. *Australian Journal of Crop Science*, 4(5), 335–342.
- Kim, D.-G., Rafique, R., Leahy, P., Cochrane, M., & Kiely, G. (2014). Estimating the impact of changing fertilizer application rate, land use, and climate on nitrous oxide emissions in Irish grasslands. *Plant and Soil*, 374(1–2), 55–71.

-WILEY

WILEY-CCB-BIOENERGY

- Kissel, D. E., Bock, B. R., & Ogles, C. Z. (2020). Thoughts on acidification of soils by nitrogen and sulfur fertilizers. *Agrosystems, Geosciences & Environment*, 3(1), e20060.
- Li, D., Watson, C. J., Yan, M. Jia, Lalor, S., Rafique, R., Hyde, B., Lanigan, G., Richards, K. G., Holden, N. M., & Humphreys, J. (2013). A review of nitrous oxide mitigation by farm nitrogen management in temperate grassland-based agriculture. *Journal of Environmental Management*, 128, 893–903.
- Li, J., Cooper, J. M., Lin, Z., Li, Y., Yang, X., & Zhao, B. (2015). Soil microbial community structure and function are significantly affected by long-term organic and mineral fertilization regimes in the North China Plain. *Applied Soil Ecology*, 96, 75–87. https:// doi.org/10.1016/j.apsoil.2015.07.001
- Li, N., Kumar, P., Lai, L., Abagandura, G. O., Kumar, S., Nleya, T., Sieverding, H. L., Stone, J. J., & Gibbons, W. (2019). Response of soil greenhouse gas fluxes and soil properties to nitrogen fertilizer rates under camelina and carinata nonfood oilseed crops. *BioEnergy Research*, 12(3), 524–535. https://doi.org/10.1007/s12155-019-09987-4
- Li, Q., Xu, M., Liu, G., Zhao, Y., & Tuo, D. (2013). Cumulative effects of a 17-year chemical fertilization on the soil quality of cropping system in the Loess Hilly Region, China. *Journal of Plant Nutrition and Soil Science*, 176(2), 249–259.
- Liu, C.-W., Sung, Y., Chen, B.-C., & Lai, H.-Y. (2014). Effects of nitrogen fertilizers on the growth and nitrate content of lettuce (*Lactuca sativa* L.). *International Journal of Environmental Research and Public Health*, 11(4), 4427–4440.
- Liu, E., Yan, C., Mei, X., Zhang, Y., & Fan, T. (2013). Long-term effect of manure and fertilizer on soil organic carbon pools in dryland farming in northwest China. *PLoS One*, 8(2), e56536.
- Lugato, E., Simonetti, G., Morari, F., Nardi, S., Berti, A., & Giardini, L. (2010). Distribution of organic and humic carbon in wet-sieved aggregates of different soils under long-term fertilization experiment. *Geoderma*, 157(3-4), 80–85. https://doi.org/10.1016/j.geode rma.2010.03.017
- Lv, Y., Wang, C., Jia, Y., Wang, W., Ma, X., Du, J., Pu, G., & Tian, X. (2014). Effects of sulfuric, nitric, and mixed acid rain on litter decomposition, soil microbial biomass, and enzyme activities in subtropical forests of China. *Applied Soil Ecology*, 79, 1–9. https:// doi.org/10.1016/j.apsoil.2013.12.002
- Ma, B.-L., Zheng, Z., Whalen, J. K., Caldwell, C., Vanasse, A., Pageau, D., Scott, P., Earl, H., & Smith, D. L. (2019). Uptake and nutrient balance of nitrogen, sulfur, and boron for optimal canola production in eastern Canada. *Journal of Plant Nutrition* and Soil Science, 182(2), 252–264. https://doi.org/10.1002/ jpln.201700615
- Malo, D. D. (2003). South Dakota soil classification key. Technical Bulletins. Paper 9. http://openprairie.sdstate.edu/agexperimentsta_ tb/9. SDSU Agricultural Experiment Station, pp. 33–35.
- Massah, J., & Azadegan, B. (2016). Effect of chemical fertilizers on soil compaction and degradation. *Ama, Agricultural Mechanization in Asia Africa & Latin America*, 47(1), 44–50.
- Mbonimpa, E. G., Hong, C. O., Owens, V. N., Lehman, R. M., Osborne, S. L., Schumacher, T. E., Clay, D. E., & Kumar, S. (2015). Nitrogen fertilizer and landscape position impacts on CO₂ and CH₄ fluxes from a landscape seeded to switchgrass. *GCB Bioenergy*, 7(4), 836–849. https://doi.org/10.1111/gcbb.12187
- Mclean, E. O. (1982). Soil pH and lime requirement. In A. L. Page (Ed.), *Methods of soil analysis. Part 2. Chemical and microbiological properties* (pp. 199–224). American Society of Agronomy, Soil Science Society of America.

- Omonode, R. A., Smith, D. R., Gál, A., & Vyn, T. J. (2011). Soil nitrous oxide emissions in corn following three decades of tillage and rotation treatments. *Soil Science Society of America Journal*, 75(1), 152–163.
- Osborne, S., Mathew, F., Ali, S., Sieverding, H., Kumar, S., & Nleya, T. (2019). Nitrogen requirements of ethiopian mustard for biofuel feedstock in South Dakota. *Agronomy Journal*, 111(3), 1304– 1311. https://doi.org/10.2134/agronj2018.06.0419
- Ozlu, E., & Kumar, S. (2018). Response of surface GHG fluxes to long-term manure and inorganic fertilizer application in corn and soybean rotation. *Science of The Total Environment*, 626, 817–825.
- Palmer, J., Thorburn, P. J., Biggs, J. S., Dominati, E. J., Probert, M. E., Meier, E. A., Huth, N. I., Dodd, M., Snow, V., Larsen, J. R., & Parton, W. J. (2017). Nitrogen cycling from increased soil organic carbon contributes both positively and negatively to ecosystem services in wheat agro-ecosystems. *Frontiers in Plant Science*, 8. https://doi.org/10.3389/fpls.2017.00731
- Parkin, T. B., & Venterea, R. T. (2010). Sampling protocols. Chapter 3. Chamber-based trace gas flux measurements. In R. F. Follett (Ed.), *Sampling protocols* (pp. 3-1–3-39). USDA. www.ars.usda. gov/research/GRACEnet
- Pelster, D. E., Larouche, F., Rochette, P., Chantigny, M. H., Allaire, S., & Angers, D. A. (2011). Nitrogen fertilization but not soil tillage affects nitrous oxide emissions from a clay loam soil under a maize–soybean rotation. *Soil and Tillage Research*, 115, 16–26.
- Pelster, D. E., Chantigny, M. H., Rochette, P., Angers, D. A., Rieux, C., & Vanasse, A. (2012). Nitrous oxide emissions respond differently to mineral and organic nitrogen sources in contrasting soil types. *Journal of Environmental Quality*, 41(2), 427–435. https://doi. org/10.2134/jeq2011.0261
- Perlack, R. D. (2005). Biomass as feedstock for a bioenergy and bioproducts industry: The technical feasibility of a billion-ton annual supply. Oak Ridge National Laboratory.
- Poffenbarger, H. J., Barker, D. W., Helmers, M. J., Miguez, F. E., Olk, D. C., Sawyer, J. E., Six, J., & Castellano, M. J. (2017). Maximum soil organic carbon storage in Midwest U.S. cropping systems when crops are optimally nitrogen-fertilized. *PLoS One*, *12*(3), e0172293. https://doi.org/10.1371/journal.pone.0172293
- Prasad, A., Swaroop, N., Thomas, T., & Rao, P. S. (2018). Effect of different levels of phosphorus and sulphur on physico-chemical properties of soil, growth and yield of mustard (*Brassica juncea* L.) Cv. Varuna. *International Journal of Chemical Studies*, 6(3), 2109–2111.
- Purakayastha, T., Rudrappa, L., Singh, D., Swarup, A., & Bhadraray, S. (2008). Long-term impact of fertilizers on soil organic carbon pools and sequestration rates in maize-wheat-cowpea cropping system. *Geoderma*, 144, 370–378.
- Sainju, U. M., Caesar-TonThat, T., Lenssen, A. W., & Barsotti, J. L. (2012). Dryland soil greenhouse gas emissions affected by cropping sequence and nitrogen fertilization. *Soil Science Society of America Journal*, 76(5), 1741–1757.
- Sainju, U. M., Jabro, J. D., & Stevens, W. B. (2008). Soil carbon dioxide emission and carbon content as affected by irrigation, tillage, cropping system, and nitrogen fertilization. *Journal of Environmental Quality*, 37, 98–106.
- Sainju, U. M., Stevens, W. B., Caesar-TonThat, T., & Liebig, M. A. (2012). Soil greenhouse gas emissions affected by irrigation, tillage, crop rotation, and nitrogen fertilization. *Journal of Environmental Quality*, 41, 1774–1786.

- Schaufler, G., Kitzler, B., Schindlbacher, A., Skiba, U., Sutton, M. A., & Zechmeister-Boltenstern, S. (2010). Greenhouse gas emissions from European soils under different land use: effects of soil moisture and temperature. *European Journal of Soil Science*, 61(5), 683–696. https://doi.org/10.1111/j.1365-2389.2010.01277.x
- Seepaul, R., George, S., & Wright, D. L. (2016). Comparative response of *Brassica carinata* and *B. napus* vegetative growth, development and photosynthesis to nitrogen nutrition. *Industrial Crops and Products*, 94, 872–883.
- Sekaran, U., McCoy, C., Kumar, S., & Subramanian, S. (2019). Soil microbial community structure and enzymatic activity responses to nitrogen management and landscape positions in switchgrass (*Panicum virgatum* L.). *GCB Bioenergy*, 11(7), 836–851.
- Singh, B. (2018). Are nitrogen fertilizers deleterious to soil health? Agronomy, 8(4), 48.
- Singh, N., Dhaliwal, J. K., Sekaran, U., & Kumar, S. (2019). Soil hydrological properties as influenced by long-term nitrogen application and landscape positions under switchgrass seeded to a marginal cropland. *GCB Bioenergy*, 11, 1026–1040.
- Soil Survey Staff. (2017). *Web soil survey*. United States Department of Agriculture. http://websoilsurvey.sc.egov.usda.gov/
- Soosaar, K., Mander, Ü., Maddison, M., Kanal, A., Kull, A., Lõhmus, K., Truu, J., & Augustin, J. (2011). Dynamics of gaseous nitrogen and carbon fluxes in riparian alder forests. *Ecological Engineering*, 37(1), 40–53. https://doi.org/10.1016/j.ecoleng.2010.07.025
- Taylor, D. C., Falk, K. C., Palmer, C. D., Hammerlindl, J., Babic, V., Mietkiewska, E., Jadhav, A., Marillia, E.-F., Francis, T., Hoffman, T., Giblin, E. M., Katavic, V., & Keller, W. A. (2010). Brassica carinata - a new molecular farming platform for delivering bioindustrial oil feedstocks: case studies of genetic modifications to improve very long-chain fatty acid and oil content in seeds. *Biofuels, Bioproducts and Biorefining*, 4(5), 538–561. https://doi.org/10.1002/bbb.231
- Tu, C., Zheng, C., & Chen, H. (2000). Effect of applying chemical fertilizers on forms of lead and cadmium in red soil. *Chemosphere*, 41(1–2), 133–138.
- Turan, M. A., Taban, S., Katkat, A. V., & Kucukyumuk, Z. (2013). The evaluation of the elemental sulfur and gypsum effect on soil pH, EC, SO. *Journal of Food, Agriculture & Environment*, 11(1), 572–575.
- USDA-NASS. (2012). Census of agriculture. https://www.nass.usda. gov/Publications/AgCensus/2012/
- Ussiri, D. A., & Lal, R. (2009). Long-term tillage effects on soil carbon storage and carbon dioxide emissions in continuous corn cropping system from an alfisol in Ohio. *Soil and Tillage Research*, 104(1), 39–47.
- Verma, O., Singh, S., Pradhan, S., Kar, G., & Rautaray, S. (2018). Irrigation, nitrogen and sulphur fertilization response on productivity, water use efficiency and quality of Ethiopian mustard (*Brassica carinata*) in a semi-arid environment. *Journal of Applied* and Natural Science, 10(2), 593–600.
- Wagner, L., Ambe, N., & Ding, D. (1994). Estimating a proctor density curve from intrinsic soil properties. *Transactions of the ASAE*, 37(4), 1121–1125.

<u>GCB-BIOENERGY</u>

- Walker, K., & Booth, E. (2003). Sulphur nutrition and oilseed quality, sulphur in plants (pp. 323–339). Springer.
- Wang, Y., Li, Q., Hui, W., Shi, J., Lin, Q., Chen, X., & Chen, Y. (2008). Effect of sulphur on soil Cu/Zn availability and microbial community composition. *Journal of Hazardous Materials*, 159(2-3), 385–389. https://doi.org/10.1016/j.jhazmat.2008.02.029
- Warwick, S., Francis, A., & Gugel, R. (2009). Guide to wild germplasm of Brassica and allied crops (tribe Brassiceae, Brassicaceae). Agriculture and Agri-Food Canada.
- Wiedenfeld, B. (2011). Sulfur application effects on soil properties in a calcareous soil and on sugarcane growth and yield. *Journal of Plant Nutrition*, 34(7), 1003–1013.
- Wright, D. (2017). Carinata fit into SE cropping systems. Presented at: Brassica carinata Summit, Quincy, Florida. 30 March.
- Zell, H. (2010). Carbon dioxide controls earth's temperature. https://www.nasa.gov/topics/earth/features/co2-temperature. html
- Zhang, H., Zhang, Y., Yan, C., Liu, E., & Chen, B. (2016). Soil nitrogen and its fractions between long-term conventional and no-tillage systems with straw retention in dryland farming in northern China. *Geoderma*, 269, 138–144.
- Zhang, Y., de Vries, W., Thomas, B. W., Hao, X., & Shi, X. (2017). Impacts of long-term nitrogen fertilization on acid buffering rates and mechanisms of a slightly calcareous clay soil. *Geoderma*, 305, 92–99.
- Zhong, H., Wang, Q., Zhao, X., Du, Q., Zhao, Y., Wang, X., Jiang, C., Zhao, S., Cao, M., Yu, H. & Wang, D. (2014). Effects of Different Nitrogen Applications on Soil Physical, Chemical Properties and Yield in Maize (*Zea mays L.*). Agricultural Sciences, 5(14), 1440– 1447. https://doi.org/10.4236/as.2014.514155
- Zhong, Y., Wang, X., Yang, J., Zhao, X., & Ye, X. (2016). Exploring a suitable nitrogen fertilizer rate to reduce greenhouse gas emissions and ensure rice yields in paddy fields. *Science of The Total Environment*, 565, 420–426.
- Zhou, H., Fang, H., Hu, C., Mooney, S. J., Dong, W., & Peng, X. (2017). Inorganic fertilization effects on the structure of a calcareous silt loam soil. Agronomy Journal, 109(6), 2871–2880. https://doi. org/10.2134/agronj2016.10.0590
- Zhou, H., Peng, X., Perfect, E., Xiao, T., & Peng, G. (2013). Effects of organic and inorganic fertilization on soil aggregation in an Ultisol as characterized by synchrotron based X-ray micro-computed tomography. *Geoderma*, 195–196, 23–30.

How to cite this article: Bhattarai D, Abagandura GO, Nleya T, Kumar S. Responses of soil surface greenhouse gas emissions to nitrogen and sulfur fertilizer rates to *Brassica carinata* grown as a bio-jet fuel. *GCB Bioenergy*. 2021;13:627–639. <u>https://doi.org/10.1111/gcbb.12784</u>

WILE'