

The Therapeutic Potential of Epigenome-modifying drugs in Cardiometabolic disease

Marwa Berjawi¹ & Christopher G Bell^{1*}

1. William Harvey Research Institute, Barts and The London School of Medicine & Dentistry, Queen Mary University of London, London EC1M 6BQ, U.K.

*Corresponding author: c.bell@qmul.ac.uk

Abstract

Purpose of review:

Cardiometabolic disorders, including cardiovascular disease, obesity and type 2 diabetes are an extreme burden on the health of individuals and societies worldwide. The combination of genetic susceptibility, environmental, and lifestyle factors drive disease development. Therefore, understanding the exact molecular changes occurring in their pathogenesis brings not only insights into mechanism, but also potential novel therapeutic angles. Epigenetic changes - the mitotically heritable chemical marks or packaging of DNA that influence gene expression without changing the genetic code itself - have been identified as highly compelling targets. Therapeutic success modulating the epigenome has initially come in cancer, but is now expanding into non-malignant diseases. Here, we discuss the different epigenetic mechanisms implicated in cardiometabolic disease and the advancing science of epigenetic therapeutics for these disorders.

Recent findings:

Disruption of epigenetic networks can be causative in cardiometabolic diseases, including for example, endothelial dysfunction and vascular ageing. Epigenomic changes may also influence or indicate known risk factors, including hypertension, inflammation and dyslipidemia. The dynamic plasticity that characterizes the epigenome lends itself to therapeutic manipulation with many epigenetic drugs now under clinical trials and others have already been approved for medical use. Major epigenetic inhibitor classes include: DNA Methyltransferase inhibitors (DNMTi); Histone Deacetylase inhibitors (HDACi); Isocitrate Dehydrogenase inhibitors (IDHi); Bromodomain and the extra-terminal motif proteins inhibitors

35 (BETi); & EZH2 inhibitors (EZH2i); with natural and synthetic examples of these
36 possessing potential cardiometabolic effects. For example, there is accumulating
37 evidence for the BETi Apabetalone to reduce vascular inflammation, thereby
38 decreasing cardiovascular events, and the HDACi Sodium Butyrate to suppress
39 experimentally inflammatory cytokines in atherosclerosis.

40 **Summary:**

41 Despite challenges in the development and application of epigenetic drugs, the
42 achievements in oncology and increasing experimental evidence indicate the
43 clinical utility of targeting the readers, writers and erasers of the epigenome.
44 Therefore, precise modifying of epigenetic information is likely to be a key player
45 in the future of personalized therapeutic approaches in individuals with
46 cardiometabolic disorders.

47

48

49 **Keywords:**

50 Epigenomics, Epigenetics, Cardiovascular Disease, Type 2 Diabetes, Obesity,
51 Hypertension, Hyperlipidaemia, Inflammation, Atherosclerosis, DNA
52 methylation, Histone post-translational modifications.

53 Introduction

54 The burden of cardiometabolic disease is growing globally [1], and further
55 significant improvements are needed in early prevention, prediction and
56 detection to avoid the disease-related mortality and morbidity [2]. During the past
57 two decades, the study of genomic variation has been highly successful in
58 establishing the field of personalised medicine. International efforts have
59 extensively described the human genome and analysed its variation in both
60 healthy and diseased individuals. It is undoubtable that the knowledge gained
61 from these genetic studies has paved the way to our understanding of the
62 molecular basis of different human disorders. Nevertheless, whilst studies of
63 genotype-phenotype relationships have brought many insights to researchers and
64 clinicians with robust benefits to patients [3], this work has not been a complete
65 panacea. Some observations do not follow the classic genetic heritability [4] and,
66 furthermore, the integration of tissue-specific regulatory information brings an
67 additional dimension to the intricacies of pathogenesis [5].

68 Epigenetics is a molecular explanation that bridges the gap between the static
69 genome, developmental trajectories [6], and specialised cell-specific activity [7],
70 as well as the complexity of environmental signals [8]. Epigenetic marks, in
71 contrast to genetic mutations, do not alter the DNA sequence but rather alter the
72 way cells use these DNA instructions [9]. Epigenetic signatures can be associated
73 with lifestyle and environmental factors, during intrauterine, postnatal
74 development, or across the entire lifecourse. Being modifiable, the epigenome
75 provides an enticing opportunity to comprehend disease-related variation, and
76 when causative, its possible reversion [4]. Many initiatives have sought to
77 determine the changes in the epigenome of human disorders, including
78 cardiometabolic disease, cancer, and neurodegenerative diseases. Whilst, many of
79 these observed changes are not causally involved in the pathogenesis, robust
80 associations are still potential disease biomarkers with clinical utility. Those
81 epigenetic modifications that do display evidence of driving these conditions are
82 targets for novel drugs to revert these defects. Therefore, epigenetics is emerging
83 as a key player in the risk prediction, diagnosis, prognosis, as well as personalized
84 treatment of cardiovascular diseases and cardiometabolic syndrome. This review
85 highlights those epigenetic modifications discovered in relation to

86 cardiometabolic disease and explores the current state of the potential
87 therapeutic use of epigenome-modifying drugs in these conditions. It also
88 presents some of the challenges associated with pharmacological targeting of
89 epigenetic networks.

90

91 **1. The epigenome**

92 In 1942, embryologist Conrad Waddington coined the term 'epigenetics', defining
93 it as being the sum of developmental processes that lie between the genotype and
94 the phenotype and dynamically connect them [10]. Although much conjecture has
95 ensued regarding definitions following this [11], at its most straightforward, this
96 term refers to the mitotically-heritable chemical modifications and packaging of
97 genomic DNA leading to a change in the gene expression without altering the
98 genetic sequence itself [12]. Fundamental to the packing mechanism is chromatin,
99 a nucleoprotein-complex composed of DNA and histone proteins that make up the
100 chromosome in eukaryotic cells [13]. It is the scaffold for packaging the whole
101 genome and is also a dynamically adjusted structure that mirrors the regulatory
102 signals responsible for various cell processes, including DNA replication,
103 transcription, DNA repair and cell division [14]. Therefore, the epigenome is the
104 sum of changes involved in chromatin modelling and post-translational histone
105 tail modifications, which can at times be guided by non-coding RNA molecules, as
106 well as direct modifications to the DNA itself [15].

107

108 **1.1 DNA Modifications**

109 The most common direct chemical addition to DNA is the classical epigenetic
110 mark of DNA methylation. In eukaryotes, DNA methylation only targets cytosine
111 residues (C-5 position of the pyrimidine ring in DNA). It is functionally involved
112 in a range of regulatory processes [16], including different stable epigenetic
113 suppression mechanisms, such as parent-of-origin imprinting, silencing of
114 repetitive DNA and inactivation of the X chromosome [16]. The DNA methylome,
115 or the genomic state of methylation, is strongly encoded by CpG density as well as
116 the interaction of transcription factors and their sequence motifs [16]. Cytosine
117 methylation occurs mainly at CpG dinucleotides (C followed by G in the 5' to 3'
118 direction of the DNA strand) that are sparsely located or at repetitive elements,
119 and it is usually not found in normal somatic cells where CpGs are densely packed

120 together (termed CpG islands). Methylated cytosine is susceptible to spontaneous
121 deamination to thymine. This 5mC mutation causes a guanine-thymine mismatch,
122 which is repaired in an error-prone manner [17]. This hypermutability means that
123 CpG sequence context is reduced to ~20% of the expected level in the human
124 genome [17] and ~30% of common human SNPs reside at disrupted CpG
125 dinucleotides [18]. Recent analysis of vital loss-of-function intolerant genes
126 identified that purifying selection acts to suppress germline methylation at their
127 promoters to reduce the CpG mutation rate in these critical regulatory regions
128 [19].

129 One group of enzymes that play an essential role in regulating DNA methylation
130 is the DNA methyltransferase family (DNMT), which includes the maintenance
131 DNMT1, and *de novo* DNMT3A and DNMT3B, as well as the regulatory factor
132 DNMT3L [20]. DNMTs are the writers of DNA methylation (see Figure 1). They
133 are highly conserved proteins that catalyze the addition of a methyl group from
134 the methyl donor S-adenosylmethionine (SAM) to cytosine to form 5-
135 methylcytosine [20,21]. DNA methylation removal can occur in an active or
136 passive way. Active demethylation is mediated by the ten-eleven translocation
137 (TET) enzymes [22]. The rare DNA modifications are steps in the active DNA
138 demethylation process. TETs add a hydroxyl group to the methyl group of 5mC to
139 convert it into its oxidized derivative 5-hydroxymethylcytosine (5hmC) and
140 sequentially further to 5-formyl cytosine (5fC) and then 5- carboxylcytosine
141 (5caC). A final step of base excision repair leads to the restoration of an
142 unmodified cytosine [23]. On the other hand, the passive loss of a methyl group
143 takes place due to a reduction of DNMT1 activity through replication [24].
144 Therefore, mutations within these epigenetic modifiers (DNMTs and TETs) can
145 lead to a change in genome-wide methylation profile, resulting in global changes
146 in the activation and repression pattern of downstream genes [25]. The extensive
147 cancer sequencing projects over the last decade have identified an enrichment of
148 oncogenic driver mutations in epigenomic modifier genes [13,26].

149

150 1.2 Post Translational Histone Tail Modifications

151 The N-terminal tails that project out from the core histones are exposed to
152 different potential post-translational modifications [27]. Currently, more than ten
153 varieties of histone modifications have been identified at different histone sites,
154 and these modifications include acetylation, methylation, ubiquitination, and

155 phosphorylation [28]. The latest fascinating addition being serotonylation and its
156 involvement in permissive expression [29]. Post-translational modifications of
157 histones regulate the shift from a tighter chromatin (heterochromatin) to a looser
158 chromatin (euchromatin) state, or vice versa [30]. The heterochromatin state
159 encapsulates either constitutive heterochromatin, with associated H3K9me3 and
160 high levels of DNA methylation found in pericentromeric regions and telomeres;
161 or facultative heterochromatin, with high H3K27me3 in nonrepetitive regions and
162 genes silenced via histone deacetylation [31]. Histone tail modifications thus
163 result in either transcriptional activation or repression through the action of
164 different modification writing and erasing enzymes (see Figure 1) [30]. For
165 instance, the mechanism of acetylation is regulated by the action of histone
166 acetyltransferases (HATs), which catalyze the addition of acetyl groups using
167 acetyl-coA as a coenzyme, and histone deacetylases (HDAC) that catalyze their
168 removal. Lysine acetyltransferases (KATs) and lysine deacetylases (KDACs) are
169 critical and well-studied enzymes with acknowledged roles in pathology [32]. In
170 general, histone acetylation influences or is correlated with transcription,
171 whereas deacetylation is associated with an inactive chromatin state [33]. At both
172 promoters and enhancers, classically delineated by H3K4me3 and H3K4me1
173 modifications, respectively, the co-occurrence of acetylation in the form of
174 H3K27ac indicates activity at these regulatory regions, compared to only a likely
175 permissive/poised state without [34].

176

177 1.3 Non-coding RNA

178 An extremely high proportion (>~98%) of the transcribed human genome does
179 not encode proteins but instead numerous non-coding RNAs (ncRNAs) [35], which
180 are divided into two classes: regulatory and structural. According to their length,
181 regulatory ncRNAs can be further categorized into two major groups: (i) small
182 ncRNAs (<200 nucleotides) that include microRNA (miRNA) and PIWI-interacting
183 RNAs (piRNA), and (ii) long ncRNAs, which are longer than 0.2 kb [36]. ncRNAs
184 can influence the expression of other genes (e.g. CHD2 by the lncRNA *Chaserr* [37])
185 and are involved in chromatin regulation and remodelling through the interaction
186 with transcription factors and epigenetic regulators to adjust their retention or
187 catalytic activity [38]. They act with both methyl- or acetyl- writing and erasing

188 enzymes, resulting in a change in chromatin conformation and gene activity [39].
189 lncRNAs can function as chromatin remodellers and can act as scaffolds for
190 additional proteins or protein complexes (e.g. lncRNA *KHPS1* activating a poised
191 enhancer)[36,40–42]. Moreover, miRNAs are found to impact epigenetic
192 processes by directly inhibiting the role of some enzymes related to DNMTs,
193 histone modifications and chromatin remodelling or by changing the availability
194 of substrates required for these enzymatic reactions. Hence, a disruption in the
195 miRNA-epigenetic regulatory circuitry may result in the impairment of normal
196 chromatin function, leading to various diseases [33].

197

198 **2. Epigenetics as a biomarker of cardiometabolic disease**

199 Epigenetic biomarkers, specifically DNA modifications, are highly attractive for
200 clinical practice because of their long-term stability [43]. They are molecular
201 markers with the ability to accurately quantitate environmental, lifestyle and
202 endogenous contributors to disease (*e.g.* pollution, smoking and inflammation
203 *etc.*) [4,44–46]. Cardiometabolic disease is associated with many risk factors that
204 can be detected by epigenetic biomarkers, as well as concurrently observable
205 epigenetic network modulations driving its pathogenesis.

206

207 **2.1 Ageing-associated changes**

208 Ageing is an obvious but major risk factor for cardiometabolic-related phenotypes
209 [47] due to many interrelated pathophysiological mechanisms [48]. It constitutes
210 progressive damage of tissue and reduced organ function over a period of time.
211 Oxidative stress and cellular senescence are involved in ageing and many acute
212 and chronic pathological processes, including cardiovascular, cardiometabolic,
213 acute and chronic kidney diseases [49]. Epigenetic alterations are one of the
214 hallmarks of ageing [47]. As cells age, we observe widespread changes in
215 transcription factor binding, heterochromatin formation, DNA methylation and
216 the histone modification landscape [50]. Several specific CpG sites that undergo
217 consistent directional age-related methylation changes have been determined and
218 enabled the construct of age predictors or DNA methylation ‘clocks’ [51]. These
219 clocks have been shown to capture ‘biological’ aspects of ageing, which are good

220 predictors of cardiovascular disease (CVD) and all-cause mortality [52–54]. One
221 gene involved in ageing and is modulated through epigenetic changes is *SIRT1*. It
222 is an important gatekeeper against inflammation, oxidative stress as well as
223 deterioration of cardiovascular structure and function and is found to be
224 downregulated in ageing [55]. Due to recent technological advances there is
225 significant interest in the analysis of the ageing-related epigenomic changes, as
226 these may bring novel functional insights to ageing-related disease mechanisms,
227 including cardiometabolic disorders [54,56,57].

228 Another fascinating insight into ageing, with ramifications for both
229 epigenomics and CVD, is age-related clonal haematopoiesis (ARCH) [58]. With age,
230 somatic mutations accumulate in the DNA of hematopoietic stem cells, and those
231 conferring a competitive advantage to the mutated cell result in its clonal
232 expansion [59]. Clonal haematopoiesis has been associated with haematological
233 malignancy, but additionally recent findings have connected this process with
234 increased cardiovascular risk, with a potential role as a driver of atherosclerosis
235 [59]. Therefore, clonal hematopoiesis is identified as being a possible novel
236 biomarker of vascular ageing [47,59] and is also associated with epigenetic
237 measures of biological age [60]. More than 70% of cases of ARCH are a result of
238 mutations in the genes coding for *DNMT3A*, *TET2* and *ASXL1*, which modulate DNA
239 and histone methylation and thus lead to distinct clonal epigenome-wide changes
240 [61]. Causal evidence has also come from a mouse model, where TET2-deficient
241 macrophages increased NLRP3-mediated inflammasome IL-1beta secretion and,
242 thus, accelerated atherosclerotic plaque growth [62].

243

244 2.2 Hypertension

245 Modifications in the epigenome are identified in essential hypertension [63,64].
246 Many genes related to pathways and biological processes involved in
247 hypertension are regulated by epigenetic changes. One example is *HSD11B2*, a
248 gene responsible for catalysing the conversion of cortisol to cortisone, with
249 promoter DNA hypermethylation resulting in abnormal levels of cortisol- and
250 cortisone-active metabolites and consequently in the onset of essential
251 hypertension [63,65]. Moreover, there is evidence that endothelial nitric oxide
252 synthase (eNOS) activity is downregulated in CVD by cell-specific histone

253 modifications [66]. This enzyme is responsible for NO production in the vascular
254 endothelium and plays an essential role in the regulation of vascular tone [67,68].
255 A recent epigenome-wide study (EWAS) in peripheral blood for hypertension
256 identified 7 significant CpGs in a European ancestry cohort for Diastolic blood
257 pressure but only 1 (cg07598370 near *OR5AP2*) in a trans-ancestry analysis
258 including a South Asian ancestry group [69]. However, differentially methylated
259 region (DMR) analysis in the trans-ancestry cohort identified numerous Systolic
260 (n=387) and Diastolic (n=237) DMRs. Of these DMRs, 12 were in common
261 between both traits, including overlap with *TFAP2D* and *HLX* previously
262 genetically implicated with hypertension [70,71].

263

264 2.3 Obesity and Type 2 Diabetes

265 Obesity and type 2 diabetes (T2D) are also detectable in changes to the
266 epigenome. Costantino *et al.* determined that the mitochondrial adaptor p66^{Shc} is
267 upregulated in visceral fat arteries of obese patients and is associated with
268 endothelial dysfunction, oxidative stress and insulin resistance [72]. A group of
269 chromatin remodelers were found through unbiased gene profiling and chromatin
270 immunoprecipitation to regulate p66^{Shc} transcription by inducing demethylation
271 and H3K9 acetylation [72]. Interestingly, intensive glycemic control failed to
272 revert p66^{Shc} -related epigenetic modifications in peripheral blood monocytes of
273 T2D individuals [73]. T2D is a heterogenous disorder associated with obesity,
274 inflammation and insulin resistance [74]. Furthermore, in T2D, pancreatic β -cells,
275 whose dysfunction is a dominant contribution to this disease's development [75],
276 undergo modifications in gene expression. DNA methylation has a significant role
277 in the development of the β -cell [76] but also in the age-related progression of T2D
278 [77]. The importance of the regulation of islet epigenetics by DNA methylation is
279 demonstrated in that combined knock-out of *Dnmt1* and the homeobox gene
280 *Aristaless-related homeobox (Arx)* convert adult islets α -cells into functional β -
281 cells [78]. Hyperglycaemia has been implicated in the β -cell dysfunction and
282 altered cellular identity that occurs in T2D [79]. Intergrated β -cell epigenomic
283 analysis also revealed weakening of β -cell specific Polycomb Repressive Complex
284 2 (PRC2) bivalent domains leads to ectopic gene activation in T2D patients [74].
285 Loss of transcription integrity of lineage-defining genes was also observed. These

286 polycomb-dependent changes were proposed as amenable epigenomic
287 therapeutic targets to stabilise β -cell identity with also the potential for directing
288 α - to β -cell conversion [80]. *PRMT1* methylates arginine-3 of histone H4
289 (H4R3me2a), enables p300 to acetylate further H4 residues, and is also found to
290 be required to maintain mature β -cell state [81]. Recent evidence also points to
291 the critical role of the lnc-RNA, lncRNA-*PAX6-AS1*, with high glucose states driving
292 increased expression of this repressor of the pancreatic β -cell identity and
293 functional transcription factor Pax6 [82].

294 Volkov et al., using whole-genome bisulfite sequencing, performed the first
295 comprehensive analysis of the DNA methylome of a T2D case-control cohort in
296 pancreatic islets [83]. A very large number of DMRs (n=25,820) were found in
297 islets from T2D patients compared to controls. However, the key islet
298 transcription factor, *PDX1*, was an outlier overlapping with seven DMRs covering
299 >2.5kb, which included 105 CpGs, and this result was also independently
300 replicated. The total DMR set showed enriched overlap with binding sites for islet-
301 specific transcription factors. When integrating the DMR analysis with pancreatic
302 islet RNA-seq data, *SOCS2*, *PARK2*, *PID1* and *NR4A3* showed changes in both DNA
303 methylation and expression. Functional studies of these genes in rat β -cells
304 revealed that the expression changes impaired glucose-stimulated insulin
305 secretion [84]. Furthermore, islet PDX-1, which has a reduced expression in T2D
306 patients, showed increased methylation when exposed to glucose [85,86]. PDX-1
307 is also regulated through histone methylation by the action of Set7/9, a lysine
308 methyltransferase (the mouse ortholog of human SETD7; KMT7) [87].

309 An epigenetic role in the complications of diabetes, such as nephropathy has also
310 been explored [88,89]. Khamis *et al.* recently identified an intriguing epigenetic
311 connection between lipid-lowering statins and the insulin sensitivity role of
312 mature adipocytes [90]. Statins are known to increase T2D risk [91] and their
313 experimental administration led to promoter hypomethylation and increased
314 expression of the epigenomic-erasor *HDAC9*, which in turn reduced the activity of
315 crucial adipogenic genes, such as *ABCG1* [90]. The impact on adipocyte maturation
316 subsequently increased insulin resistance. This mechanistic insight will need to be
317 carefully appraised, as both *HDAC9* and *ABCG1* are considered potential
318 therapeutic targets for metabolic diseases.

319 From three meta-analysis EWAS performed in peripheral blood for obesity
320 [92,93], ten significant CpGs were consistently identified [94]. None have any
321 biological evidence of being causative in obesity, but instead are consequential to
322 the obese state, such as changes driven by hyperlipidaemia, hyperglycaemia and
323 inflammation. In line with this, a meta-analysis of EWAS in only children and
324 adolescents (n=4,133 from 23 studies) found minimal DNA methylation
325 association with childhood and adolescent BMI [95]. However, with advancing age
326 there was an increasing overlap with known associations observed with adult
327 BMI. Such findings further support that differences in blood DNA methylation are
328 a consequence rather than a cause of obesity.

329

330 2.4 Dyslipidemia

331 Epigenomic modifications have been implicated with a role in inter-individual
332 postprandial lipemia (PPL) variability. An epigenome-wide association study for
333 PPL in isolated CD4+ T cells, from 979 adults from the GOLDN study challenged
334 with a high-fat meal, identified eight significant DNA methylation sites involving
335 five genes (*CPT1A*, *LPP*, *APOA5*, *SREBF1* and *ABCG1*) [96]. The increased
336 methylation at *LPP*, *APOA5*, *ABCG1* and *SREBF1* and the decreased methylation at
337 *CPT1A* correlated with the increase of triglyceride-PPL response [96]. With the
338 same cohort, another study revealed *CPT1A* methylation being associated with
339 fasting very-low LDL-cholesterol and triglyceride [97]. *CPT1A* has been previously
340 pinpointed in multiple cardiometabolic-related EWAS [94]. A landmark study
341 from Dekkers *et al.* [98] determined, using multivariable Mendelian
342 randomization, that DNA methylation change is induced by triglycerides at three
343 CpGs, by HDL cholesterol at two CpGs, and by LDL cholesterol at one CpG. These
344 six CpGs were found to be associated with the expression of key regulators of lipid
345 metabolism, which include *CPT1A* and *SREBF1*, *ABCG1*, and *DHCR24* for
346 triglycerides, HDL cholesterol, and LDL cholesterol, respectively.

347 A recent EWAS of metabolic syndrome and its components was conducted on
348 1,887 individuals of European ancestry [99]. The study implicated cg08309687, a
349 locus that was previously associated with T2D, with lipid metabolism. Another
350 DNA methylation locus, cg17901584, was found to be connected with HDL levels.

351 Also, the study concluded that cg19693031, within the 3'UTR of *TXNIP*, possesses
352 multiple intersecting lines of evidence for it to be a potential central epigenetic
353 hub locus linking separate metabolic syndrome components. A further EWAS in
354 peripheral leukocyte DNA of 226 metabolic measurements determined by nuclear
355 magnetic resonance (NMR) spectroscopy identified 161 robustly replicated
356 associations for 57 unique metabolic measures at 16 CpG sites (discovery n= 1662,
357 replication n = 3752) [100]. All these CpGs had been previously associated via
358 biochemical and clinical measures with metabolic components.

359 In regard to non-coding RNA, two lncRNAs have been determined as powerful
360 regulators of lipid levels as well as atherosclerosis: LeXis, which is expressed in
361 the liver and controls genes involved in cholesterol biosynthesis, and MeXis, which
362 is expressed in macrophages and controls genes involved in cholesterol efflux
363 [101,102].

364

365 **2.5 Atherosclerosis**

366 The pathogenesis of atherosclerosis involves variations in epigenetic
367 modifications and gene expression in a cell type- and stage- specific manner [103].
368 Significant changes in DNA methylation occur in the atherosclerotic plaque [104] with
369 early studies on humans and mice noting global and focal DNA hypermethylation
370 [105,106]. Using bisulfite sequencing of healthy and atherosclerotic human
371 aortas, a positive relationship between DNA methylation and atherosclerotic
372 lesion grade was determined [107]. A DNA methylation array analysis comparing
373 aortic healthy versus normal identified >1,800 plaque-associated CpGs and
374 validated 16 by pyrosequencing, including promoter CpGs with inversely
375 associated transcription levels for *HOXA6* and *MIR23b* [105]. A more recent 850k
376 DNA methylation array analysis identified 2,679 CpGs reaching genome-wide
377 significance between paired atheromatous plaque lesions and corresponding plaque-
378 free aortic intima tissue in 128 post-mortem specimens from 64 Japanese patients [108].

379 *In vitro* studies have suggested that inflammatory signaling pathways can regulate
380 DNA methylation [103]. For instance, proinflammatory stimuli, such as oxidized
381 LDL (oxLDL), resulted in DNMT1 upregulation and caused the DNA methylation
382 of the promoter of the gene that encodes Krüppel-like factor 2 (*KLF2*) [109]. KLFs

383 belong to a group of anti-inflammatory transcription factors, thus *KLF2*
384 suppression increases proatherogenic endothelial inflammation [109].

385 Post-translation histone tail methylation and acetylation modifications occur in the
386 development of atherosclerosis as well as its progression to atheroma involving a
387 number of different cell types, including monocytes, macrophages, vascular smooth
388 muscle cells and endothelial cells [110]. A distinct subset of CD4⁺ T cells with
389 expression markers of T cell exhaustion are observed in the atherosclerotic lesions
390 [111]. Macrophages become activated and with uptake of lipid particles differentiate
391 into foam cells, becoming further dysregulated in the plaque [112].

392 Several lncRNAs have been identified as key players in vascular diseases [103].
393 One example is *ANRIL*, which is NF- κ B inducible [113] and promotes human
394 vascular smooth muscle cell (vSMC) proliferation [114,115]. It also upregulates
395 the expression of IL6 and IL8 through interaction and recruitment of the
396 transcription factor YY1 [113]. Additionally, long intergenic non-coding RNA p21
397 (lincRNA-p21) is implicated in atherosclerosis [36]. Its expression is
398 downregulated in atherosclerotic plaques of *ApoE*^{-/-} mice and human coronary
399 arteries [116]. Moreover, the knockdown of this lncRNA stimulated the expansion
400 of the neointima in a mouse model of carotid artery vascular injury [116].

401

402 **3. Potential use of epigenome-modifying drugs in cardiometabolic disease**

403

404 3.1 Success with therapeutic manipulation of the epigenome in cancer

405 Since the clear recognition of a global loss of DNA methylation in cancer cells in
406 1983 [117], epigenetic drug discovery efforts have investigated oncogenic
407 pathways [118]. Epigenome modulation is a feature of almost all human cancers;
408 different mutations in chromatin-controlling genes are common, some of which
409 are drivers for tumour initiation, while others may influence immune invasion,
410 metastasis or even drug resistance [119]. Therefore, epi-drugs in cancer are
411 targeted at reversing these changes and reactivating tumour suppressor genes
412 [120]. These therapies tested in clinical trials can be divided into two major
413 groups: i) broad 'reprogrammers' and ii) targeted therapies. Broad

414 reprogrammers cause genome-wide modifications in the epigenome, and include
415 DNA methyltransferase inhibitors (DNMTi), histone deacetylase inhibitors
416 (HDACi), and inhibitors of the bromodomain and the extra-terminal motif
417 proteins (BETi). These were the first-generation modulators to be permitted for
418 clinic use. Targeted therapies, on the other hand, treat specific genetic mutations
419 in the epigenetic pathways [121–123], or in the future, epigenetically modify a
420 specific regulatory locus.

421

422 Although many drugs are in development, presently nine epigenetic agents have
423 been approved by the FDA in the US for the standard treatment of haematological
424 cancers and solid tumours [13]. These are currently from the writer and eraser
425 category of epigenomic regulators and include DNMTi, HDACi, IDHi (Isocitrate
426 dehydrogenase inhibitors that block IDH mutant inhibition of TET2 and lysine
427 demethylation) and EZH2i (inhibition of the EZH2 involved in H3K27 methylation
428 and the enzymatic component of the Polycomb Repressive Complex 2) [13].
429 Interestingly, most of the work on multidrug combinations is currently focused on
430 the synergistic role of epi-drugs as potentiators to increase sensitivity to other
431 anticancer therapies, such as immunotherapy in patients with cancer [124–126].
432 Both HDACi and DNMTi have shown an effective immunomodulatory activity on
433 host immune cells and tumour cells.

434

435 3.2 Toward epigenetic therapies for cardiometabolic disease

436 Altered epigenetic regulation has been related to the development and
437 progression of cardiometabolic disease and the modulation of the biological
438 processes underlying it. The six currently approved epigenomic eraser inhibitors,
439 which are all HDACis, are: Vorinostat, Belinostat, Panobinostat and Romidepsin,
440 plus repurposed compounds now recognised to affect this pathway; the diet
441 supplement Sodium Butyrate [127] and the anticonvulsant and affective disorder
442 treatment, Valproic acid (see Table 1). Therefore, after their success as
443 epigenomic modulators in cancer therapy, HDACis experimental benefits have
444 been explored across a range of diseases, including cardiovascular,
445 neurodegenerative, and inflammatory diseases (e.g. Atherosclerosis [32], long-
446 term memory [128], Systemic Lupus Erythematosus (SLE) [129]). This has

447 subsequently led to several clinical trials now taking place to study the use of
448 epigenetic drugs in targeting conditions related to cardiometabolic disease.
449 Supportive evidence for HDACis for the treatment of metabolic and cardiovascular
450 diseases [130], also includes improved endothelial dysfunction and diabetic
451 nephropathy, and reduced ischemia/reperfusion myocardial injury [131,132].
452 Humans possess 18 HDACs, which can be categorized into four major classes
453 according to their homology to yeast (class I, II, & IV possessing conserved
454 deacetylase domains differing by specific cofactor dependence, and class III sirtuin
455 proteins) [133]. Whilst there is significant experimental evidence for
456 cardiovascular benefits of HDACis [134], the inhibition of the distinctive classes of
457 HDAC differ in their action. Inhibitors of Class II and class III HDACs have
458 protective roles, not only in heart injury but also in vessel injury, whereas those
459 acting on class I HDACs protect against vessel damage [135], but can led to cardiac
460 arrhythmias, atherosclerosis and vessel calcification [32].

461 Administration of Trichostatin A (TSA), a selective class I & II HDAC inhibitor, in a
462 mouse model of acute myocardial infarction reduced inflammation by
463 downregulating TNF- α expression and stimulated angiogenesis through AKT-1
464 phosphorylation, leading to ventricular remodeling reduction [136]. Vorinostat
465 (SAHA; suberanilohydroxamic acid), which is approved for the treatment of
466 Cutaneous T-Cell Lymphoma (CTCL), has also gained interest due to its ability to
467 reduce inflammation and oxidative stress [137,138]. Oral administration for 18
468 weeks in diabetic mice, resulted in the reduction of albuminuria, collagen
469 deposition and oxidative-nitrosative stress by inhibiting eNOS coupling [139]. In
470 addition, in a rabbit model of myocardial infarction provoked by ischemia
471 followed by reperfusion, SAHA was shown to reduce myocardial infarct size and
472 partially improve systolic function [132]. This was by inducing cell death and
473 cardiomyocyte autophagy, a mechanism implicated across a range of disorders
474 [140]. However, the molecular mechanisms underlying autophagy in cardiac
475 muscle cells remain unknown [132]. Similarly, the HDACi Sodium Butyrate was
476 found to lessen the inflammatory response in atherosclerosis and myocardial
477 infarction by suppressing the inflammatory molecules TNF- α , IL-6, ICAM-1 and
478 VCAM-1 and driving cardiomyocyte autophagy [132,141]. Also, this compound
479 has shown therapeutic effects in diet-induced obesity by increasing mitochondrial

480 oxidative phosphorylation and activating adiponectin-mediated pathways as well
481 as the ability to reduce low-grade inflammation, vascular disease, and protect
482 against cardiac dysfunction in T2D and obesity [127,141,142].

483 Resveratrol, a polyphenol that is present in red wine, is recognized to activate the
484 histone deacetylase SIRT1, and has been implicated in improving endothelial
485 function, insulin sensitivity and myocardial dysfunction in obese and T2D patients
486 [143]. Interestingly, resveratrol and its derivatives may also play a role as DNMT3
487 inhibitors [138,144]. However, its rapid metabolism and poor bioavailability are
488 acknowledged weaknesses [145]. Therefore, the benefit of this compound
489 requires further clinical studies to be robustly assessed; it exhibits a broad range
490 of cardioprotective, vasculoprotective and atheroprotective-related activities,
491 suggesting its therapeutic potential for the treatment, or prevention, of
492 cardiometabolic disease [138]. The anti-diabetic medications metformin and
493 glucagon-like peptide (GLP-1) analogues are also modulators of SIRT1 activity,
494 thus modifying histone acetylation and transcription of genes involved in insulin
495 signaling and pancreatic β -cell homeostasis [39].

496 A promising future for DNA methylation modifier clinical therapies is suggested
497 as aberrations in DNA methylation patterns, as well as being biomarkers of
498 disease, can also potentially be drivers of cardiovascular and metabolic disorders.
499 Increased DNA methylation was observed in some atheroprotective genes
500 including those encoding estrogen receptors (ER)- α and - β (ESR1 and ESR2,
501 respectively) in human coronary atherosclerotic tissues and plaque regions of
502 ascending aorta [146]. Hypermethylation of ESR1 and ESR2 was also detected in
503 *in vitro* senescing smooth muscle cells and endothelial cells [147]. Such findings
504 imply that epigenetic modifications in both ESR1 and 2 can drive vascular ageing
505 and atherosclerosis [147]. The DNMTi 5-aza-2-deoxycytidine (5-AZA), which is
506 now a well-established anti-cancer therapy, administered in addition to TSA,
507 restored the expression of ER genes in normal smooth muscle cells and
508 endothelial cells, showing low toxicity in these cells [138,148]. Therefore, the
509 combination of epigenetic therapy with hormone replacement therapy has been
510 proposed as potentially beneficial for the prevention and/or treatment of CVD
511 [149]. Experimental intraperitoneal injections of 5-AZA in *Ldlr*^{-/-} and *Apoe*^{-/-}

512 mouse models was observed to cause a reduction in the atherosclerotic lesion
513 burden [150,151].

514 Different natural compounds can also affect relevant epigenetic networks and
515 consequently prevent cardiometabolic features. For instance, Curcumin, a
516 naturally occurring molecule that gives turmeric its yellow color, modulates the
517 activity of DNMT1, HATs (including p300) and HDACs, and has the potential to
518 prevent several diabetes-related outcomes as well as recognized cardiovascular
519 protective effects [137,138,152]. Administration of Curcumin in rats averted the
520 development of hypertension-induced heart failure by inhibiting GATA4
521 transcription, which is a hypertrophy-responsive transcription factor [153]. The
522 atheroprotective action includes enhancing cholesterol efflux in human and
523 mouse macrophages *in vitro*, with an anti-inflammatory effect acting through the
524 downregulation of NF- κ B activity [154–156].

525 Similarly, dietary supplementation of folate, a crucial vitamin responsible for the
526 production of SAM, has the ability to revert aberrations in chromatin in different
527 cardiometabolic states. Folate-enriched diet reduced endothelial dysfunction and
528 hypertension as well as increasing the bioavailability of nitric oxide [157]. In
529 addition, a study demonstrated that folic acid supplementation prevents
530 atherosclerosis by investigating its atheroprotective role in high-fat diet-fed *Apoe*
531 knockout mice and in oxidized low-density lipoprotein-treated human umbilical
532 vein endothelial cells [158]. An increase in folic acid levels elevates DNA
533 methyltransferase activity and expression, and thus modifies the expression of
534 atherosclerosis-related genes [158]. Several other compounds like apicidin,
535 valproic acid and peroxisome proliferator-activated receptor gamma agonists
536 (PPAR γ -agonists) also have the ability to revert chromatin alterations in
537 cardiometabolic conditions (see Table 1) [39,137].

538 The BET protein family belongs to the class of epigenomic readers [13]. They
539 modulate gene transcription through their interaction with acetylated histone
540 residues, and they have been implicated in different pathological conditions,
541 including inflammation [137]. The BET inhibitor Apabetalone (RVX 208) is able to
542 regulate reverse cholesterol transport, coagulation, vascular inflammation and
543 complement activation in cultured primary human hepatocytes [159]. Moreover,

544 a pooled analysis of phase II clinical trials for this BETi demonstrated a significant
545 reduction of major adverse cardiovascular events (MACE) [160]. A phase III
546 clinical trial on T2D individuals with recent acute coronary syndrome revealed
547 promising outcomes related to specific secondary endpoints like cardiovascular
548 death or nonfatal myocardial infarction and heart failure. However, this data could
549 not clearly establish a reduction in MACE [161]. Although, a short-term analysis of
550 established coronary artery disease patients showed benefit with respect to
551 cardiovascular events compared to placebo [162]. Additionally, a recent study
552 assessed the effect of Apabetalone on *ex vivo* inflammatory responses of
553 monocytes from patients with T2D and CVD [163]. They demonstrated that
554 Apabetalone treatment suppresses this pro-inflammatory phenotype, which
555 further supports the potential role of BETi as a therapeutic agent for high risk T2D
556 and CVD patients.

557

558 3.3 Challenges and future directions

559 Whilst our understanding of the complexity of the epigenome is still limited,
560 pharmacological targeting of epigenetic networks will remain challenging [4]. The
561 variability in cellular composition can confound or dilute biomarker studies due
562 to the cell-type specificity of epigenetic factors, particularly for rare cell subtype
563 signatures. Moreover, in most cases, detailed functional evaluation is required to
564 distinguish whether the epigenetic modification is a consequence of disease and
565 possibly helpful for diagnosis, patient stratification or response prediction, or a
566 cause that could be a suitable therapeutic target [4]. Another issue to be addressed
567 when considering implementing epigenetic therapy is the lack of locus specificity.
568 Genome-wide epi-drugs can disrupt epigenetic marks indiscriminately, which
569 may modulate the expression of off-target genome sequences. This can be
570 accompanied with substantial side effects [164,165]. The CRISPR-Cas9 editing
571 system has been modified to perform locus-specific epigenetic editing [166],
572 including DNA methylases [167], demethylases, and histone modifiers, including
573 acetylators, deacetylators and demethylators [168]. Using dCas9 fused to different
574 epigenetic modifiers and transcriptional regulatory domains, or using scaffold
575 RNAs recruited with effector proteins, CRISPR-Cas9 system-mediated site-specific

576 transcriptional and epigenetic modulations can be achieved [168–170]. Although
577 targeting efficiency and intracellular delivery are still considerations [171],
578 CRISPR-Cas9 is a very promising tool for the precise modulation of the
579 cardiometabolic disease epigenome [172]. Illustrating the capability of
580 epigenomic editing, a study succeeded in stimulating human β -cell proliferation
581 even using the older TALE targeting methodology coupled with the TET1 enzyme
582 [173]. In the imprinting disorder Beckwith-Wiedemann syndrome (BWS), β -cell
583 hyperproliferation is driven by hypomethylation of the maternal imprinting control
584 region 2 (ICR2) of the *CDKN1C* gene (expressing the cell-cycle inhibitor p57). In
585 this study, the epigenomic targeting of the TET1 demethylating enzyme to this
586 *CDKN1C* ICR2, produced a similar increase in β -cell mass. The CRISPR/CAS9-based
587 genetic deletion of enhancers induced by the common hypertensive therapeutic
588 target angiotensin II (AngII), which also promotes atherosclerosis by growth and
589 pro-inflammatory pathways in vSMCs, blocks this AngII-induced pathology [174].
590 These knocked-out enhancers are clear cardiovascular targets for more nuanced
591 future therapeutic epigenomic modulation.

592

593 **Conclusions**

594 Clinical epigenetics is still in a developmental phase. Despite its infancy, a few epi-
595 drugs have been approved and implemented in clinical use, and a variety of
596 epigenetic biomarkers for disease diagnosis, prognosis or response to therapy are
597 now available or are in development. Currently, oncology is the main focus for epi-
598 drugs, but an increasing number of trials suggest their potential for use in many
599 other non-malignant conditions in the very near future. In this review, we have
600 shed light on the strong prospects of these epigenetic modulators in alleviating
601 the progression of cardiometabolic disease. Furthermore, a deeper understanding
602 of the epigenetic landscape in disease enabled by advancing technologies,
603 including single cell analyses, may reveal additional novel targets for the
604 prevention and treatment of cardiometabolic-related disorders. Epigenetic drugs
605 represent true genomic therapeutics, and the development of a new era of
606 personalized genomic medicine using predictive biomarkers or personalized
607 therapy will undoubtedly incorporate the epigenome as a vital player.

608

609 **Conflicts of Interest**

610 The authors have no conflicts of interest to declare.

611 **Table 1**

612

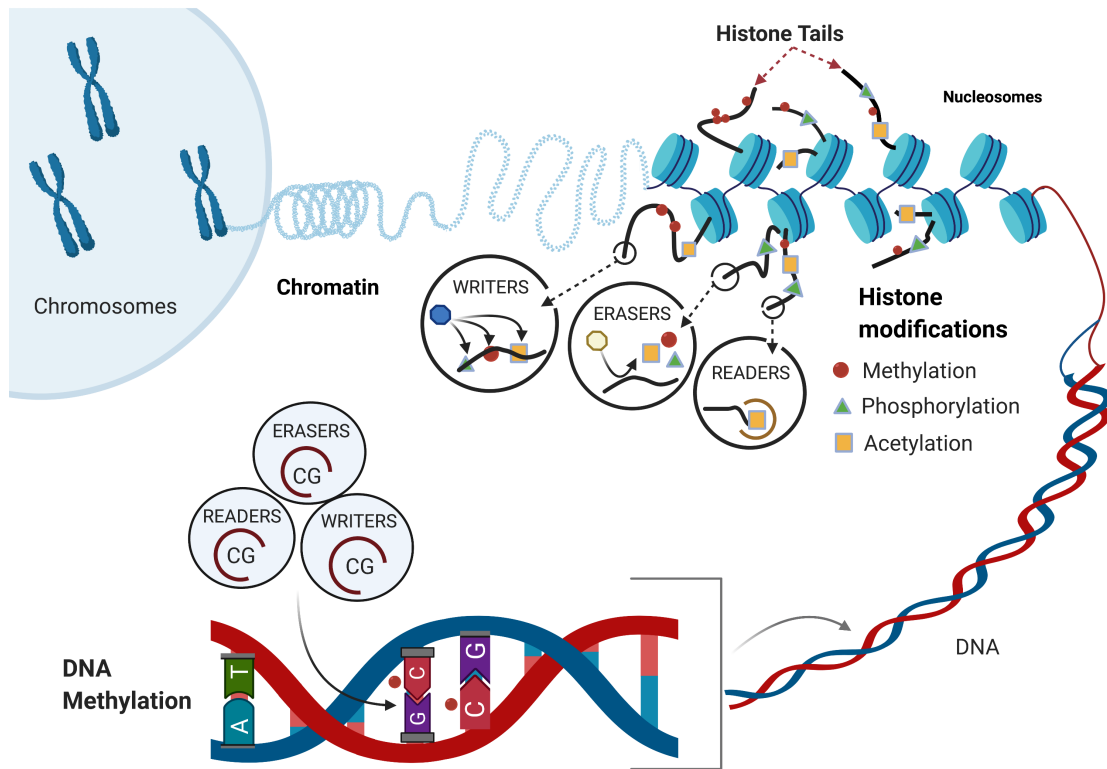
Compound	Epigenetic Mechanism	Models Tested	Effect
Apabetalone	BET inhibition	Human, Mouse	Modulates reverse cholesterol transport, coagulation, vascular inflammation and complement activation [159] and reduces cardiovascular events in patients with coronary artery disease [175].
Apicidin	HDAC inhibition	Human cells, Mouse	Reduces myocardial hypertrophy after 1-week pressure overload prompted by thoracic aortic constriction [176].
Curcumin	HAT inhibition	Rat	Inhibits cardiac hypertrophy and preserves cardiac/endothelial function [177].
Folate	DNA and histone methylation	Human cells, Mouse	Influences SAM production, modifies the expression of atherosclerosis-related genes and prevents atherosclerosis [158].
GLP1 analogue	SIRT1 activation	Mouse	Reduces blood pressure, vascular inflammation, and oxidative stress and prevents uncoupling of eNOS [178].
Metformin	SIRT1 activation	Human	Protects vascular endothelial cells from cellular stress, downgrades signs of cellular ageing and production of reactive oxygen species [179].
PPAR γ agonists	HAT/HDAC recruitment	Human	Improve vascular function in T2D patients [180].

Resveratrol	SIRT1 activation	Human, Mouse	Improves endothelial function, cardiac dysfunction and insulin sensitivity in obesity and T2D [143].
Sodium Butyrate	HDAC inhibition	Human cells, Rat	Suppresses inflammatory cytokines in experimental models of myocardial infarction and atherosclerosis [141].
Trichostatin A	HDAC inhibition	Human cells, Mouse	Promotes angiogenic response and cardiomyocyte survival and prevents ischemia-induced left ventricular remodelling [136].
Valproic Acid	HDAC inhibition	Rat	Attenuates hypertrophic and hypertensive responses through modulation of ROS-generating and pro-inflammatory pathways [181].
Vorinostat	HDAC inhibition	Mouse, Rabbit	Promotes the autophagic flux, prevents uncoupling of eNOS and activation of NF- κ B and reduces oxidative stress [132,139,182].

613

Table 1 Epigenetic drugs assessed for treatment of cardiometabolic diseases⁶¹⁴ in alphabetic order [39,137]. BET: Bromodomain and the Extra-Terminal motif; eNOS: Endothelial Nitric Oxide Synthase; GLP1: Glucagon-Like Peptide 1; HAT: Histone acetyltransferase; HDAC: Histone Deacetylase; NF- κ B: Nuclear Factor kappa-light-chain-enhancer of activated B cells; PPAR γ : Peroxisome Proliferator-Activated Receptor Gamma; ROS: Reactive Oxygen Species; SAM: S-adenosylmethionine; SIRT1: Sirtuin 1, a NAD⁺-dependent Histone Deacetylase; & T2D: Type 2 Diabetes.

615 **Figure 1**



616

617 **Figure Legend**

618 Figure 1. The Epigenome

619 The epigenome comprises of chromatin, a nucleoprotein-complex composed of
 620 DNA and histone proteins, which make up the chromosome in eukaryotic cells. It
 621 interacts with DNA modifications, post-translational histone tail modifications,
 622 and non-coding RNA molecules. Figure adapted from Bates [13] and Keating *et al.*
 623 [89].

624 **Highlighted References**

- 625 • 81. Kim H, Yoon BH, Oh CM, Lee J, Lee K, Song H, et al. PRMT1 is required
626 for the maintenance of mature β -cell identity. *Diabetes*. 2020;69:355–68
627 ○ This study highlighted the importance of histone H4 arginine 3
628 asymmetric dimethylation to maintain pancreatic β -cell identity.
629 Identifying critical epigenomic marks pinpoints targets for future
630 epi-drugs, in this case for β -cell dysfunction.
- 631 • 111. Fernandez DM, Rahman AH, Fernandez NF, Chudnovskiy A, Amir E ad
632 D, Amadori L, et al. Single-cell immune landscape of human atherosclerotic
633 plaques. *Nat Med*. 2019;25:1576–88.
634 ○ This single cell proteomic and transcriptomic analysis identified the
635 specific innate and adaptive immune cells present in atherosclerotic
636 plaques. Precise pathogenic cell type identification reveals targets
637 for novel epi-drug action.
- 638 • 126. Morel D, Jeffery D, Aspeslagh S, Almouzni G, Postel-Vinay S. Combining
639 epigenetic drugs with other therapies for solid tumours — past lessons and
640 future promise. *Nat. Rev. Clin. Oncol*. 2020. p. 91–107.
641 ○ This review details the potentiating effect of combining epi-drugs
642 with standard therapies in oncology. This approach may also be
643 successful in cardiometabolic diseases.
- 644 • 132. Xie M, Kong Y, Tan W, May H, Battiprolu PK, Pedrozo Z, et al. Histone
645 deacetylase inhibition blunts ischemia/reperfusion injury by inducing
646 cardiomyocyte autophagy. *Circulation*. 2014;129:1139–51.
647 ○ The use of an anticancer histone deacetylase inhibitor (SAHA) was
648 tested in an animal model and shown to have a cardioprotective
649 effect in myocardial ischemia by blunting reperfusion injury and
650 reducing infarct size.
- 651 • 163 - Wasiak S, Dzobo KE, Rakai BD, Kaiser Y, Versloot M, Bahjat M, et al.
652 BET protein inhibitor apabetalone (RVX-208) suppresses pro-
653 inflammatory hyper-activation of monocytes from patients with
654 cardiovascular disease and type 2 diabetes. *Clin Epigenetics*. 2020;12.

- 655 ○ This study indicates the therapeutic potential in CVD & T2D of
656 inhibiting the reading of acetylated lysine histone residues by the
657 BETi Apabetalone.
- 658 • 173. Ou K, Yu M, Moss NG, Wang YJ, Wang AW, Nguyen SC, et al. Targeted
659 demethylation at the CDKN1C/p57 locus induces human β cell replication.
660 J Clin Invest. 2019;129:209–14.
- 661 ○ The targeted DNA demethylation by the epigenomic modifier TET
662 enzyme is shown to induce β -cell proliferation and indicates a
663 possible future epi-drug treatment for diabetes.

664 **References**

- 665 1. Mancia G, Carugo S, Grassi G. Primary Prevention of Cardiovascular Disease.
666 *Cardiol.* 1997;88:32–7.
- 667 2. Sattar N, Gill JMR, Alazawi W. Improving prevention strategies for
668 cardiometabolic disease. *Nat Med.* 2020;26:320–5.
- 669 3. Claussnitzer M, Cho JH, Collins R, Cox NJ, Dermitzakis ET, Hurles ME, et al. A brief
670 history of human disease genetics. *Nature.* 2020;577:179–89.
- 671 4. Berdasco M, Esteller M. Clinical epigenetics: seizing opportunities for
672 translation. *Nat. Rev. Genet.* 2019;20:109–27.
- 673 5. Roadmap Epigenomics Consortium, Kundaje A, Meuleman W, Ernst J, Bilenky M,
674 Yen A, et al. Integrative analysis of 111 reference human epigenomes. *Nature.*
675 2015;518:317–29.
- 676 6. Eckersley-Maslin MA, Alda-Catalinas C, Reik W. Dynamics of the epigenetic
677 landscape during the maternal-to-zygotic transition. *Nat. Rev. Mol. Cell Biol.*
678 2018;19:436–50.
- 679 7. Kelsey G, Stegle O, Reik W. Single-cell epigenomics: Recording the past and
680 predicting the future. *Science (80-.).* 2017;358:69–75.
- 681 8. Feil R, Fraga MF. Epigenetics and the environment: Emerging patterns and
682 implications. *Nat. Rev. Genet.* 2012;13:97–109.
- 683 9. Cavalli G, Heard E. Advances in epigenetics link genetics to the environment and
684 disease. *Nature.* 2019;571:489–99.
- 685 10. Waddington CH. Canalization of development and the inheritance of acquired
686 characters. *Nature.* 1942;150:563–5.
- 687 11. Greally JM. A user’s guide to the ambiguous word “epigenetics.” *Nat. Rev. Mol.*
688 *Cell Biol.* 2018;19:207–8.
- 689 12. Alexanian M, Padmanabhan A, McKinsey TA, Haldar SM. Epigenetic therapies
690 in heart failure. *J Mol Cell Cardiol.* 2019;130:197–204.
- 691 13. Bates SE. Epigenetic Therapies for Cancer. *N. Engl. J. Med.* 2020;383:650–63.
- 692 14. Luger K, Dechassa ML, Tremethick DJ. New insights into nucleosome and
693 chromatin structure: An ordered state or a disordered affair? *Nat Rev Mol Cell Biol.*
694 2012;13:436–47.
- 695 15. Allis CD, Jenuwein T. The molecular hallmarks of epigenetic control. *Nat. Rev.*
696 *Genet.* 2016;17:487–500.
- 697 16. Schübeler D. Function and information content of DNA methylation. *Nature.*
698 2015;517:321–6.
- 699 17. Gujar H, Weisenberger DJ, Liang G. The roles of human DNA
700 methyltransferases and their isoforms in shaping the epigenome. *Genes (Basel).*
701 2019;10:172.
- 702 18. Bell CG, Gao F, Yuan W, Roos L, Acton RJ, Xia Y, et al. Obligatory and facilitative
703 allelic variation in the DNA methylome within common disease-associated loci.
704 *Nat Commun.* 2018;9(1):8
- 705 19. Boukas L, Bjornsson H, Hansen K. Purifying selection acts on germline
706 methylation to modify the CpG mutation rate at promoters. *BioRxiv* 2020;
707 2020.07.04.187880
- 708 20. Ambrosi C, Manzo M, Baubec T. Dynamics and Context-Dependent Roles of
709 DNA Methylation. *J. Mol. Biol.* 2017;429:1459–75.
- 710 21. Lyko F. The DNA methyltransferase family: A versatile toolkit for epigenetic
711 regulation. *Nat. Rev. Genet.* 2018;19:81–92.
- 712 22. Wu X, Zhang Y. TET-mediated active DNA demethylation: Mechanism, function

- 713 and beyond. *Nat. Rev. Genet.* 2017;18:517–34.
- 714 23. Bochtler M, Kolano A, Xu GL. DNA demethylation pathways: Additional players
715 and regulators. *BioEssays.* 2017;39:1–13.
- 716 24. Masi S, Ambrosini S, Mohammed SA, Sciarretta S, Lüscher TF, Paneni F, et al.
717 Epigenetic Remodeling in Obesity-Related Vascular Disease. *Antioxid Redox*
718 *Signal.* 2020; doi: 10.1089/ars.2020.8040
- 719 25. Lee CJ, Ahn H, Jeong D, Pak M, Moon JH, Kim S. Impact of mutations in DNA
720 methylation modification genes on genome-wide methylation landscapes and
721 downstream gene activations in pan-cancer. *BMC Med Genomics.* 2020;13; 27
- 722 26. Plass C, Pfister SM, Lindroth AM, Bogatyrova O, Claus R, Lichter P. Mutations
723 in regulators of the epigenome and their connections to global chromatin patterns
724 in cancer. *Nat. Rev. Genet.* 2013;14:765–80.
- 725 27. Biswas S, Rao CM. Epigenetic tools (The Writers, The Readers and The Erasers)
726 and their implications in cancer therapy. *Eur. J. Pharmacol.* 2018;837:8–24.
- 727 28. Ling C, Rönn T. Epigenetics in Human Obesity and Type 2 Diabetes. *Cell Metab.*
728 2019;29:1028–44.
- 729 29. Farrelly LA, Thompson RE, Zhao S, Lepack AE, Lyu Y, Bhanu N V., et al. Histone
730 serotonylation is a permissive modification that enhances TFIID binding to
731 H3K4me3. *Nature.* 2019;567:535–9.
- 732 30. Costantino S, Libby P, Kishore R, Tardif JC, El-Osta A, Paneni F. Epigenetics and
733 precision medicine in cardiovascular patients: From basic concepts to the clinical
734 arena. *Eur. Heart J.* 2018;39:4150–8.
- 735 31. Saksouk N, Barth TK, Ziegler-Birling C, Olova N, Nowak A, Rey E, et al.
736 Redundant Mechanisms to Form Silent Chromatin at Pericentromeric Regions
737 Rely on BEND3 and DNA Methylation. *Mol Cell.* 2014;56:580–94.
- 738 32. Li P, Ge J, Li H. Lysine acetyltransferases and lysine deacetylases as targets for
739 cardiovascular disease. *Nat. Rev. Cardiol.* 2020;17:96–115.
- 740 33. Sadakierska-Chudy A, Filip M. A Comprehensive View of the Epigenetic
741 Landscape. Part II: Histone Post-translational Modification, Nucleosome Level,
742 and Chromatin Regulation by ncRNAs. *Neurotox. Res.* 2014;27:172–97.
- 743 34. Ernst J, Kheradpour P, Mikkelsen TS, Shores N, Ward LD, Epstein CB, et al.
744 Mapping and analysis of chromatin state dynamics in nine human cell types.
745 *Nature.* 2011;473:43–9.
- 746 35. Hon CC, Ramilowski JA, Harshbarger J, Bertin N, Rackham OJL, Gough J, et al.
747 An atlas of human long non-coding RNAs with accurate 5' ends. *Nature.*
748 2017;543:199–204.
- 749 36. Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs
750 and its biological functions. *Nat. Rev. Mol. Cell Biol.* 2020;22:96–118.
- 751 37. Rom A, Melamed L, Gil N, Goldrich MJ, Kadir R, Golan M, et al. Regulation of
752 CHD2 expression by the Chaserr long noncoding RNA gene is essential for
753 viability. *Nat Commun.* 2019; 10(1):5092
- 754 38. Kaikkonen MU, Adelman K. Emerging Roles of Non-Coding RNA Transcription.
755 *Trends Biochem. Sci.* 2018;43:654–67.
- 756 39. Costantino S, Mohammed SA, Ambrosini S, Paneni F. Epigenetic processing in
757 cardiometabolic disease. *Atherosclerosis.* 2019;281:150–8.
- 758 40. Blank-Giwojna A, Postepska-Igielska A, Grummt I. lncRNA KHPS1 Activates a
759 Poised Enhancer by Triplex-Dependent Recruitment of Epigenomic Regulators.
760 *Cell Rep.* 2019;26:2904-2915.e4.
- 761 41. Squillaro T, Peluso G, Galderisi U, Di Bernardo G. Long non-coding RNAs in

- 762 regulation of adipogenesis and adipose tissue function. *Elife*. 2020;9:1–15.
- 763 42. Baker M. Long noncoding RNAs: The search for function. *Nat Methods*.
- 764 2011;8:379–83.
- 765 43. García-Giménez JL, Seco-Cervera M, Tollefsbol TO, Romá-Mateo C, Peiró-Chova
- 766 L, Lapunzina P, et al. Epigenetic biomarkers: Current strategies and future
- 767 challenges for their use in the clinical laboratory. *Crit. Rev. Clin. Lab. Sci*.
- 768 2017;54:529–50.
- 769 44. Lee MK, Xu CJ, Carnes MU, Nichols CE, Ward JM, Kwon SO, et al. Genome-wide
- 770 DNA methylation and long-term ambient air pollution exposure in Korean adults.
- 771 *Clin Epigenetics*. 2019; 11; 37
- 772 45. Joehanes R, Just AC, Marioni RE, Pilling LC, Reynolds LM, Mandaviya PR, et al.
- 773 Epigenetic Signatures of Cigarette Smoking. *Circ Cardiovasc Genet*. 2016;9:436–
- 774 47.
- 775 46. Ligthart S, Marzi C, Aslibekyan S, Mendelson MM, Conneely KN, Tanaka T, et al.
- 776 DNA methylation signatures of chronic low-grade inflammation are associated
- 777 with complex diseases. *Genome Biol*. 2016; 17(1):255
- 778 47. Hamczyk MR, Nevado RM, Barettino A, Fuster V, Andrés V. Biological Versus
- 779 Chronological Aging: JACC Focus Seminar. *J. Am. Coll. Cardiol*. 2020;75:919–30.
- 780 48. Pietri P, Stefanadis C. Cardiovascular Aging and Longevity: JACC State-of-the-
- 781 Art Review. *J. Am. Coll. Cardiol*. 2021;77:189–204.
- 782 49. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, et al. Oxidative
- 783 stress, aging, and diseases. *Clin. Interv. Aging*. 2018;13:757–72.
- 784 50. Booth LN, Brunet A. The Aging Epigenome. *Mol. Cell*. 2016;62:728–44.
- 785 51. Horvath S, Raj K. DNA methylation-based biomarkers and the epigenetic clock
- 786 theory of ageing. *Nat. Rev. Genet*. 2018;19:371–84.
- 787 52. Marioni RE, Shah S, McRae AF, Chen BH, Colicino E, Harris SE, et al. DNA
- 788 methylation age of blood predicts all-cause mortality in later life. *Genome Biol*.
- 789 2015; 16(1):25
- 790 53. Field AE, Robertson NA, Wang T, Havas A, Ideker T, Adams PD. DNA
- 791 Methylation Clocks in Aging: Categories, Causes, and Consequences. *Mol. Cell*.
- 792 2018;71:882–95.
- 793 54. Bell CG, Lowe R, Adams PD, Baccarelli AA, Beck S, Bell JT, et al. DNA
- 794 methylation aging clocks: Challenges and recommendations. *Genome Biol*. 2019;
- 795 20(1):249
- 796 55. Costantino S, Paneni F, Cosentino F. Ageing, metabolism and cardiovascular
- 797 disease. *J. Physiol*. 2016;594:2061–73.
- 798 56. Bell CG, Xia Y, Yuan W, Gao F, Ward K, Roos L, et al. Novel regional age-
- 799 associated DNA methylation changes within human common disease-associated
- 800 loci. *Genome Biol*. 2016;17(193)
- 801 57. Acton RJ, Yuan W, Gao F, Xia Y, Bourne E, Wozniak E, et al. The genomic loci of
- 802 specific human tRNA genes exhibit ageing-related DNA hypermethylation.
- 803 *bioRxiv*. 2019. 10.1101/870352.
- 804 58. Steensma DP, Bejar R, Jaiswal S, Lindsley RC, Sekeres MA, Hasserjian RP, et al.
- 805 Clonal hematopoiesis of indeterminate potential and its distinction from
- 806 myelodysplastic syndromes. *Blood*. 2015;126:9–16.
- 807 59. Fuster JJ, Walsh K. Somatic mutations and clonal hematopoiesis: Unexpected
- 808 potential new drivers of age-related cardiovascular disease. *Circ. Res*.
- 809 2018;122:523–32.
- 810 60. Robertson NA, Hillary RF, McCartney DL, Terradas-Terradas M, Higham J,

- 811 Sproul D, et al. Age-related clonal haemopoiesis is associated with increased
812 epigenetic age. *Curr. Biol.* 2019;29:R786–7.
- 813 61. Michalak EM, Burr ML, Bannister AJ, Dawson MA. The roles of DNA, RNA and
814 histone methylation in ageing and cancer. *Nat Rev Mol Cell Biol.* 2019;20:573–89.
- 815 62. Fuster JJ, MacLauchlan S, Zuriaga MA, Polackal MN, Ostriker AC, Chakraborty
816 R, et al. Clonal hematopoiesis associated with TET2 deficiency accelerates
817 atherosclerosis development in mice. *Science (80-)*. 2017;355:842–7.
- 818 63. Raftopoulos L, Katsi V, Makris T, Tousoulis D, Stefanadis C, Kallikazaros I.
819 Epigenetics, the missing link in hypertension. *Life Sci.* 2015;129:22–6.
- 820 64. Arif M, Sadayappan S, Becker RC, Martin LJ, Urbina EM. Epigenetic
821 modification: a regulatory mechanism in essential hypertension. *Hypertens. Res.*
822 2019;42:1099–113.
- 823 65. Udali S, Guarini P, Moruzzi S, Choi SW, Friso S. Cardiovascular epigenetics:
824 From DNA methylation to microRNAs. *Mol. Aspects Med.* 2013;34:883–901.
- 825 66. Daiber A, Xia N, Steven S, Oelze M, Hanf A, Kröller-Schön S, et al. New
826 therapeutic implications of endothelial nitric oxide synthase (eNOS)
827 function/dysfunction in cardiovascular disease. *Int. J. Mol. Sci.* 2019;20:187.
- 828 67. Han S, Uludag MO, Usanmaz SE, Ayaloglu-Butun F, Akcali KC, Demirel-Yilmaz
829 E. Resveratrol affects histone 3 lysine 27 methylation of vessels and blood
830 biomarkers in DOCA salt-induced hypertension. *Mol Biol Rep.* 2015;42:35–42.
- 831 68. Wise IA, Charchar FJ. Epigenetic modifications in essential hypertension. *Int. J.*
832 *Mol. Sci.* 2016;17:451.
- 833 69. Kazmi N, Elliott HR, Burrows K, Tillin T, Hughes AD, Chaturvedi N, et al.
834 Associations between high blood pressure and DNA methylation. *PLoS One.*
835 2020;15(1):e0227728.
- 836 70. Levy D, Larson MG, Benjamin EJ, Newton-Cheh C, Wang TJ, Hwang SJ, et al.
837 Framingham Heart Study 100K Project: Genome-wide associations for blood
838 pressure and arterial stiffness. *BMC Med Genet.* 2007;8:S3.
- 839 71. Wakil SM, Ram R, Muiya NP, Mehta M, Andres E, Mazhar N, et al. A genome-
840 wide association study reveals susceptibility loci for myocardial
841 infarction/coronary artery disease in Saudi Arabs. *Atherosclerosis.* 2016;245:62–
842 70.
- 843 72. Costantino S, Paneni F, Viridis A, Hussain S, Mohammed SA, Capretti G, et al.
844 Interplay among H3K9-editing enzymes SUV39H1, JMJD2C and SRC-1 drives p66
845 Shc transcription and vascular oxidative stress in obesity. *Eur Heart J.*
846 2019;40:383–91.
- 847 73. Costantino S, Paneni F, Battista R, Castello L, Capretti G, Chiandotto S, et al.
848 Impact of glycemic variability on chromatin remodeling, oxidative stress, and
849 endothelial dysfunction in patients with type 2 diabetes and with target HbA1c
850 levels. *Diabetes.* 2017;66:2472–82.
- 851 74. Lu TTH, Heyne S, Dror E, Casas E, Leonhardt L, Boenke T, et al. The Polycomb-
852 Dependent Epigenome Controls β Cell Dysfunction, Dedifferentiation, and
853 Diabetes. *Cell Metab.* 2018;27:1294-1308.e7.
- 854 75. Thurner M, van de Bunt M, Torres JM, Mahajan A, Nylander V, Bennett AJ, et al.
855 Integration of human pancreatic islet genomic data refines regulatory
856 mechanisms at type 2 diabetes susceptibility loci. *Elife.* 2018;7:e31977
- 857 76. Dhawan S, Georgia S, Tschening S, Fan G, Bhushan A. Pancreatic β Cell Identity
858 Is Maintained by DNA Methylation-Mediated Repression of Arx. *Dev Cell.*
859 2011;20:419–29.

- 860 77. Bacos K, Gillberg L, Volkov P, Olsson AH, Hansen T, Pedersen O, et al. Blood-
861 based biomarkers of age-associated epigenetic changes in human islets associate
862 with insulin secretion and diabetes. *Nat Commun.* 2016;7:11089
- 863 78. Chakravarthy H, Gu X, Enge M, Dai X, Wang Y, Damond N, et al. Converting Adult
864 Pancreatic Islet α Cells into β Cells by Targeting Both Dnmt1 and Arx. *Cell Metab.*
865 2017;25:622–34.
- 866 79. Wang Z, York NW, Nichols CG, Remedi MS. Pancreatic β cell dedifferentiation
867 in diabetes and redifferentiation following insulin therapy. *Cell Metab.*
868 2014;19:872–82.
- 869 80. Remedi MS. Should I Stay or Should I Go: A Clash of α -Cell Identity. *Cell Metab.*
870 2017;25:488–90.
- 871 81. Kim H, Yoon BH, Oh CM, Lee J, Lee K, Song H, et al. PRMT1 is required for the
872 maintenance of mature β -cell identity. *Diabetes.* 2020;69:355–68.
- 873 82. Lopez-Noriega L, Callingham R, Martinez-Sánchez A, Pizza G, Haberman N,
874 Cveticic N, et al. The long non-coding RNA Pax6os1/PAX6-AS1 modulates
875 pancreatic β -cell identity and function. *bioRxiv.* 2020. 2020.07.17.209015
- 876 83. Volkov P, Bacos K, Ofori JK, Esguerra JLS, Eliasson L, Rönn T, et al. Whole-
877 Genome bisulfite sequencing of human pancreatic islets reveals novel
878 differentially methylated regions in type 2 diabetes pathogenesis. *Diabetes.*
879 2017;66:1074–85.
- 880 84. Zhang H, Pollin TI. Epigenetics Variation and Pathogenesis in Diabetes. *Curr.*
881 *Diab. Rep.* 2018; 18(11):121
- 882 85. Al-Haddad R, Karnib N, Assaad RA, Bilen Y, Emmanuel N, Ghanem A, et al.
883 Epigenetic changes in diabetes. *Neurosci. Lett.* 2016;625:64–9.
- 884 86. Hall E, Dekker Nitert M, Volkov P, Malmgren S, Mulder H, Bacos K, et al. The
885 effects of high glucose exposure on global gene expression and DNA methylation
886 in human pancreatic islets. *Mol Cell Endocrinol.* 2018;472:57–67.
- 887 87. Maganti A V., Maier B, Tersey SA, Sampley ML, Mosley AL, Özcan S, et al.
888 Transcriptional activity of the islet β cell factor Pdx1 Is augmented by lysine
889 methylation catalyzed by the methyltransferase Set7/9. *J Biol Chem.*
890 2015;290:9812–22.
- 891 88. Bell CG, Teschendorff AE, Rakyan VK, Maxwell AP, Beck S, Savage DA. Genome-
892 wide DNA methylation analysis for diabetic nephropathy in type 1 diabetes
893 mellitus. *BMC Med Genomics.* 2010;3:33
- 894 89. Keating ST, van Diepen JA, Riksen NP, El-Osta A. Epigenetics in diabetic
895 nephropathy, immunity and metabolism. *Diabetologia.* 2018;61:6–20.
- 896 90. Khamis A, Boutry R, Canouil M, Mathew S, Lobbens S, Crouch H, et al. Histone
897 deacetylase 9 promoter hypomethylation associated with adipocyte dysfunction
898 is a statin-related metabolic effect. *Clin Epigenetics.* 2020;12(68)
- 899 91. Galicia-Garcia U, Jebari S, Larrea-Sebal A, Uribe KB, Siddiqi H, Ostolaza H, et al.
900 Statin treatment-induced development of type 2 diabetes: From clinical evidence
901 to mechanistic insights. *Int. J. Mol. Sci.* 2020;21:1–25.
- 902 92. Demerath EW, Guan W, Grove ML, Aslibekyan S, Mendelson M, Zhou YH, et al.
903 Epigenome-wide association study (EWAS) of BMI, BMI change and waist
904 circumference in African American adults identifies multiple replicated loci. *Hum*
905 *Mol Genet.* 2015;24:4464–79.
- 906 93. Mendelson MM, Marioni RE, Joehanes R, Liu C, Hedman ÅK, Aslibekyan S, et al.
907 Association of Body Mass Index with DNA Methylation and Gene Expression in
908 Blood Cells and Relations to Cardiometabolic Disease: A Mendelian

- 909 Randomization Approach. *PLoS Med.* 2017;14(1):e1002215
- 910 94. Bell CG. The Epigenomic Analysis of Human Obesity. *Obesity.* 2017;25:1471–
- 911 81.
- 912 95. Vehmeijer FOL, Küpers LK, Sharp GC, Salas LA, Lent S, Jima DD, et al. DNA
- 913 methylation and body mass index from birth to adolescence: meta-analyses of
- 914 epigenome-wide association studies. *Genome Med.* 2020;12(1):105
- 915 96. Lai CQ, Wojczynski MK, Parnell LD, Hidalgo BA, Irvin MR, Aslibekyan S, et al.
- 916 Epigenome-wide association study of triglyceride postprandial responses to a
- 917 high-fat dietary challenge. *J Lipid Res.* 2016;57:2200–7.
- 918 97. Irvin MR, Zhi D, Joehanes R, Mendelson M, Aslibekyan S, Claas SA, et al.
- 919 Epigenome-wide association study of fasting blood lipids in the genetics of lipid-
- 920 lowering drugs and diet network study. *Circulation.* 2014;130:565–72.
- 921 98. Dekkers KF, van Iterson M, Sliker RC, Moed MH, Bonder MJ, van Galen M, et
- 922 al. Blood lipids influence DNA methylation in circulating cells. *Genome Biol.*
- 923 2016;17(1):138
- 924 99. Nuotio ML, Pervjakova N, Joensuu A, Karhunen V, Hiekkalinna T, Milani L, et al.
- 925 An epigenome-wide association study of metabolic syndrome and its components.
- 926 *Sci Rep.* 2020;10(1):20567
- 927 100. Gomez-Alonso M del C, Kretschmer A, Wilson R, Pfeiffer L, Karhunen V,
- 928 Seppälä I, et al. DNA methylation and lipid metabolism: an EWAS of 226 metabolic
- 929 measures. *Clin Epigenetics.* 2021;13(1):7
- 930 101. Sallam T, Jones MC, Gilliland T, Zhang L, Wu X, Eskin A, et al. Feedback
- 931 modulation of cholesterol metabolism by the lipid-responsive non-coding RNA
- 932 LeXis. *Nature.* 2016;534:124–8.
- 933 102. Sallam T, Jones M, Thomas BJ, Wu X, Gilliland T, Qian K, et al. Transcriptional
- 934 regulation of macrophage cholesterol efflux and atherogenesis by a long
- 935 noncoding RNA. *Nat Med.* 2018;24:304–12.
- 936 103. Khyzha N, Alizada A, Wilson MD, Fish JE. Epigenetics of Atherosclerosis:
- 937 Emerging Mechanisms and Methods. *Trends Mol. Med.* 2017;23:332–47.
- 938 104. Aavik E, Babu M, Ylä-Herttua S. DNA methylation processes in
- 939 atherosclerotic plaque. *Atherosclerosis.* 2019;281:168–79.
- 940 105. Zaina S, Heyn H, Carmona FJ, Varol N, Sayols S, Condom E, et al. DNA
- 941 methylation map of human atherosclerosis. *Circ Cardiovasc Genet.* 2014;7:692–
- 942 700.
- 943 106. Yoo T, Yoon YS, Ryu SH, Ahn JY, Kim S, Ha TY, et al. Hypermethylation of
- 944 repetitive DNA elements in livers of mice fed an atherogenic diet. *Nutrition.*
- 945 2012;28:127–30.
- 946 107. Del Pilar Valencia-Morales M, Zaina S, Heyn H, Carmona FJ, Varol N, Sayols S,
- 947 et al. The DNA methylation drift of the atherosclerotic aorta increases with lesion
- 948 progression. *BMC Med Genomics.* 2015;8:7
- 949 108. Yamada Y, Horibe H, Oguri M, Sakuma J, Takeuchi I, Yasukochi Y, et al.
- 950 Identification of novel hyper- or hypomethylated CpG sites and genes associated
- 951 with atherosclerotic plaque using an epigenome-wide association study. *Int J Mol*
- 952 *Med.* 2018;41:2724–32.
- 953 109. Kumar A, Kumar S, Vikram A, Hoffman TA, Naqvi A, Lewarchik CM, et al.
- 954 Histone and DNA methylation-mediated epigenetic downregulation of endothelial
- 955 kruppel-like factor 2 by low-density lipoprotein cholesterol. *Arterioscler Thromb*
- 956 *Vasc Biol.* 2013;33:1936–42.
- 957 110. Jiang W, Agrawal DK, Boosani CS. Cell-specific histone modifications in

- 958 atherosclerosis (Review). *Mol. Med. Rep.* 2018;18:1215–24.
- 959 111. Fernandez DM, Rahman AH, Fernandez NF, Chudnovskiy A, Amir E ad D,
960 Amadori L, et al. Single-cell immune landscape of human atherosclerotic plaques.
961 *Nat Med.* 2019;25:1576–88.
- 962 112. Kuznetsova T, Prange KHM, Glass CK, de Winther MPJ. Transcriptional and
963 epigenetic regulation of macrophages in atherosclerosis. *Nat. Rev. Cardiol.*
964 2020;17:216–28.
- 965 113. Zhou X, Han X, Wittfeldt A, Sun J, Liu C, Wang X, et al. Long non-coding RNA
966 ANRIL regulates inflammatory responses as a novel component of NF- κ B pathway.
967 *RNA Biol.* 2016;13:98–108.
- 968 114. Yap KL, Li S, Muñoz-Cabello AM, Raguz S, Zeng L, Mujtaba S, et al. Molecular
969 Interplay of the Noncoding RNA ANRIL and Methylated Histone H3 Lysine 27 by
970 Polycomb CBX7 in Transcriptional Silencing of INK4a. *Mol Cell.* 2010;38:662–74.
- 971 115. Holdt LM, Hoffmann S, Sass K, Langenberger D, Scholz M, Krohn K, et al. Alu
972 Elements in ANRIL Non-Coding RNA at Chromosome 9p21 Modulate Atherogenic
973 Cell Functions through Trans-Regulation of Gene Networks. *PLoS Genet.*
974 2013;9(7)
- 975 116. Wu G, Cai J, Han Y, Chen J, Huang ZP, Chen C, et al. LincRNA-p21 regulates
976 neointima formation, vascular smooth muscle cell proliferation, apoptosis, and
977 atherosclerosis by enhancing p53 activity. *Circulation.* 2014;130:1452–65.
- 978 117. Feinberg AP, Vogelstein B. Hypomethylation distinguishes genes of some
979 human cancers from their normal counterparts. *Nature.* 1983;301:89–92.
- 980 118. Hunter P. The second coming of epigenetic drugs. *EMBO Rep.* 2015;16:276–
981 9.
- 982 119. Pfister SX, Ashworth A. Marked for death: Targeting epigenetic changes in
983 cancer. *Nat. Rev. Drug Discov.* 2017;16:241–63.
- 984 120. Ghasemi S. Cancer's epigenetic drugs: where are they in the cancer
985 medicines? *Pharmacogenomics J.* 2020;20:367–79.
- 986 121. Jones PA, Ohtani H, Chakravarthy A, De Carvalho DD. Epigenetic therapy in
987 immune-oncology. *Nat. Rev. Cancer.* 2019;19:151–61.
- 988 122. Mohammad HP, Barbash O, Creasy CL. Targeting epigenetic modifications in
989 cancer therapy: erasing the roadmap to cancer. *Nat. Med.* 2019;25:403–18.
- 990 123. Jones PA, Issa JPJ, Baylin S. Targeting the cancer epigenome for therapy. *Nat.*
991 *Rev. Genet.* 2016;17:630–41.
- 992 124. Mazzone R, Zwergel C, Mai A, Valente S. Epi-drugs in combination with
993 immunotherapy: a new avenue to improve anticancer efficacy. *Clin. Epigenetics.*
994 2017;9:59
- 995 125. Chiappinelli KB, Zahnow CA, Ahuja N, Bylin SB. Combining epigenetic and
996 immunotherapy to combat cancer. *Cancer Res.* 2016;76:1683–9.
- 997 126. Morel D, Jeffery D, Aspeslagh S, Almouzni G, Postel-Vinay S. Combining
998 epigenetic drugs with other therapies for solid tumours — past lessons and future
999 promise. *Nat. Rev. Clin. Oncol.* 2020;17:91–107.
- 1000 127. Zhang L, Du J, Yano N, Wang H, Zhao YT, Dubielecka PM, et al. Sodium Butyrate
1001 Protects -Against High Fat Diet-Induced Cardiac Dysfunction and Metabolic
1002 Disorders in Type II Diabetic Mice. *J Cell Biochem.* 2017;118:2395–408.
- 1003 128. Ziemka-Nalecz M, Jaworska J, Sypecka J, Zalewska T. Histone deacetylase
1004 inhibitors: A therapeutic key in neurological disorders? *J. Neuropathol. Exp.*
1005 *Neurol.* 2018;77:855–70.
- 1006 129. Reilly CM, Regna N, Mishra N. HDAC inhibition in lupus models. *Mol Med.*

- 1007 2011;17:417–25.
- 1008 130. Wang Z, Zhao YT, Zhao TC. Histone deacetylases in modulating cardiac
1009 disease and their clinical translational and therapeutic implications. *Exp. Biol.*
1010 *Med.* 2020;246:213–25.
- 1011 131. Costantino S, Ambrosini S, Paneni F. The epigenetic landscape in the
1012 cardiovascular complications of diabetes. *J. Endocrinol. Invest.* 2019;42:505–11.
- 1013 132. Xie M, Kong Y, Tan W, May H, Battiprolu PK, Pedrozo Z, et al. Histone
1014 deacetylase inhibition blunts ischemia/reperfusion injury by inducing
1015 cardiomyocyte autophagy. *Circulation.* 2014;129:1139–51.
- 1016 133. Park SY, Kim JS. A short guide to histone deacetylases including recent
1017 progress on class II enzymes. *Exp. Mol. Med.* 2020;52:204–12.
- 1018 134. Yoon S, Eom GH. HDAC and HDAC Inhibitor: From Cancer to Cardiovascular
1019 Diseases. *Chonnam Med J.* 2016;52:1.
- 1020 135. Aune SE, Herr DJ, Mani SK, Menick DR. Selective inhibition of class I but not
1021 class IIb histone deacetylases exerts cardiac protection from ischemia
1022 reperfusion. *J Mol Cell Cardiol.* 2014;72:138–45.
- 1023 136. Zhang L, Qin X, Zhao Y, Fast L, Zhuang S, Liu P, et al. Inhibition of histone
1024 deacetylases preserves myocardial performance and prevents cardiac remodeling
1025 through stimulation of endogenous angiomyogenesis. *J Pharmacol Exp Ther.*
1026 2012;341:285–93.
- 1027 137. Ambrosini S, Mohammed SA, Lüscher TF, Costantino S, Paneni F. New
1028 Mechanisms of Vascular Dysfunction in Cardiometabolic Patients: Focus on
1029 Epigenetics. *High Blood Press. Cardiovasc. Prev.* 2020;27:363–71.
- 1030 138. Chistiakov DA, Orekhov AN, Bobryshev Y V. Treatment of cardiovascular
1031 pathology with epigenetically active agents: Focus on natural and synthetic
1032 inhibitors of DNA methylation and histone deacetylation. *Int. J. Cardiol.*
1033 2017;227:66–82.
- 1034 139. Advani A, Huang Q, Thai K, Advani SL, White KE, Kelly DJ, et al. Long-term
1035 administration of the histone deacetylase inhibitor vorinostat attenuates renal
1036 injury in experimental diabetes through an endothelial nitric oxide synthase-
1037 dependent mechanism. *Am J Pathol.* 2011;178:2205–14.
- 1038 140. Rubinsztein DC, Codogno P, Levine B. Autophagy modulation as a potential
1039 therapeutic target for diverse diseases. *Nat. Rev. Drug Discov.* 2012;11:709–30.
- 1040 141. Hu XR, Zhang K, Xu CW, Chen ZQ, Jiang H. Anti-inflammatory effect of sodium
1041 butyrate preconditioning during myocardial ischemia/reperfusion. *Exp Ther Med.*
1042 2014;8:229–32.
- 1043 142. Jian H, Yimin J, Shifng P, Longfei J, Huifang L, Zhenqiang H, et al. Butyrate
1044 alleviates high fat diet-induced obesity through activation of adiponectin-
1045 mediated pathway and stimulation of mitochondrial function in the skeletal
1046 muscle of mice. *Oncotarget.* 2016;7:56071–82.
- 1047 143. Pollack RM, Crandall JP. Resveratrol: therapeutic potential for improving
1048 cardiometabolic health. *Am. J. Hypertens.* 2013;26:1260–8.
- 1049 144. Aldawsari FS, Aguayo-Ortiz R, Kapilashrami K, Yoo J, Luo M, Medina-Franco
1050 JL, et al. Resveratrol-salicylate derivatives as selective DNMT3 inhibitors and
1051 anticancer agents. *J Enzyme Inhib Med Chem.* 2016;31:695–703.
- 1052 145. Berman AY, Motechin RA, Wiesenfeld MY, Holz MK. The therapeutic potential
1053 of resveratrol: a review of clinical trials. *NPJ Precis Oncol.* 2017;1:35
- 1054 146. Kim GH, Ryan JJ, Archer SL. The role of redox signaling in epigenetics and
1055 cardiovascular disease. *Antioxidants Redox Signal.* 2013;18:1920–36.

- 1056 147. Schiano C, Vietri MT, Grimaldi V, Picascia A, Pascale MR De, Napoli C.
 1057 Epigenetic-related therapeutic challenges in cardiovascular disease. *Trends*
 1058 *Pharmacol. Sci.* 2015;36:226–35.
- 1059 148. Kim J, Kim JY, Song KS, Lee YH, Seo JS, Jelinek J, et al. Epigenetic changes in
 1060 estrogen receptor β gene in atherosclerotic cardiovascular tissues and in-vitro
 1061 vascular senescence. *Biochim Biophys Acta - Mol Basis Dis.* 2007;1772:72–80.
- 1062 149. Voelter-Mahlknecht S. Epigenetic associations in relation to cardiovascular
 1063 prevention and therapeutics. *Clin. Epigenetics.* 2016;8:1–17.
- 1064 150. Cao Q, Wang X, Jia L, Mondal AK, Diallo A, Hawkins GA, et al. Inhibiting DNA
 1065 methylation by 5-Aza-2'-deoxycytidine ameliorates atherosclerosis through
 1066 suppressing macrophage inflammation. *Endocrinology.* 2014;155:4925–38.
- 1067 151. Dunn J, Qiu H, Kim S, Jjingo D, Hoffman R, Kim CW, et al. Flow-dependent
 1068 epigenetic DNA methylation regulates endothelial gene expression and
 1069 atherosclerosis. *J Clin Invest.* 2014;124:3187–99.
- 1070 152. Li H, Sureda A, Devkota HP, Pittalà V, Barreca D, Silva AS, et al. Curcumin, the
 1071 golden spice in treating cardiovascular diseases. *Biotechnol. Adv.*
 1072 2020;38:107343.
- 1073 153. Morimoto T, Sunagawa Y, Kawamura T, Takaya T, Wada H, Nagasawa A, et al.
 1074 The dietary compound curcumin inhibits p300 histone acetyltransferase activity
 1075 and prevents heart failure in rats. *J Clin Invest.* 2008;118:868–78.
- 1076 154. Lin XL, Liu MH, Hu HJ, Feng HR, Fan XJ, Zou WW, et al. Curcumin Enhanced
 1077 Cholesterol Efflux by Upregulating ABCA1 Expression Through AMPK-SIRT1-
 1078 LXR α Signaling in THP-1 Macrophage-Derived Foam Cells. *DNA Cell Biol.*
 1079 2015;34:561–72.
- 1080 155. Aggarwal BB, Sung B. Pharmacological basis for the role of curcumin in
 1081 chronic diseases: an age-old spice with modern targets. *Trends Pharmacol. Sci.*
 1082 2009;30:85–94.
- 1083 156. Liu T, Li C, Sun H, Luo T, Tan Y, Tian D, et al. Curcumin inhibits monocyte
 1084 chemoattractant protein-1 expression and enhances cholesterol efflux by
 1085 suppressing the c-Jun N-terminal kinase pathway in macrophage. *Inflamm. Res.*
 1086 2014;63:841–50.
- 1087 157. Costantino S, Paneni F, Cosentino F. Targeting Chromatin Remodeling to
 1088 Prevent Cardiovascular Disease in Diabetes. *Curr Pharm Biotechnol.*
 1089 2015;16:531–43.
- 1090 158. Cui S, Li W, Lv X, Wang P, Gao Y, Huang G. Folic acid supplementation delays
 1091 atherosclerotic lesion development by modulating MCP1 and VEGF DNA
 1092 methylation levels in vivo and in vitro. *Int J Mol Sci.* 2017;18:990.
- 1093 159. Gilham D, Wasiak S, Tsujikawa LM, Halliday C, Norek K, Patel RG, et al. R VX-
 1094 208, a BET-inhibitor for treating atherosclerotic cardiovascular disease, raises
 1095 ApoA-I/HDL and represses pathways that contribute to cardiovascular disease.
 1096 *Atherosclerosis.* 2016;247:48–57.
- 1097 160. Ghosh GC, Bhadra R, Ghosh RK, Banerjee K, Gupta A. R VX 208: A novel BET
 1098 protein inhibitor, role as an inducer of apo A-I/HDL and beyond. *Cardiovasc. Ther.*
 1099 2017;35:e12265.
- 1100 161. Ray KK, Nicholls SJ, Buhr KA, Ginsberg HN, Johansson JO, Kalantar-Zadeh K,
 1101 et al. Effect of Apabetalone Added to Standard Therapy on Major Adverse
 1102 Cardiovascular Events in Patients with Recent Acute Coronary Syndrome and
 1103 Type 2 Diabetes: A Randomized Clinical Trial. *JAMA - J Am Med Assoc.*
 1104 2020;323:1565–73.

- 1105 162. Nicholls SJ, Ray KK, Johansson JO, Gordon A, Sweeney M, Halliday C, et al.
 1106 Selective BET Protein Inhibition with Apabetalone and Cardiovascular Events: A
 1107 Pooled Analysis of Trials in Patients with Coronary Artery Disease. *Am J*
 1108 *Cardiovasc Drugs*. 2018;18:109–15.
- 1109 163. Wasiak S, Dzobo KE, Rakai BD, Kaiser Y, Versloot M, Bahjat M, et al. BET
 1110 protein inhibitor apabetalone (RVX-208) suppresses pro-inflammatory hyper-
 1111 activation of monocytes from patients with cardiovascular disease and type 2
 1112 diabetes. *Clin Epigenetics*. 2020;12(166)
- 1113 164. Cheishvili D, Boureau L, Szyf M. DNA demethylation and invasive cancer:
 1114 Implications for therapeutics. *Br. J. Pharmacol*. 2015;172:2705–15.
- 1115 165. Hannan AJ. Epimimetics: Novel Therapeutics Targeting Epigenetic Mediators
 1116 and Modulators. *Trends Pharmacol. Sci*. 2020;41:232–5.
- 1117 166. Keung AJ, Joung JK, Khalil AS, Collins JJ. Chromatin regulation at the frontier
 1118 of synthetic biology. *Nat. Rev. Genet*. 2015;16:159–71.
- 1119 167. Holtzman L, Gersbach CA. Editing the epigenome: Reshaping the genomic
 1120 landscape. *Annu. Rev. Genomics Hum. Genet*. 2018;19:43–71.
- 1121 168. Stricker SH, Köferle A, Beck S. From profiles to function in epigenomics. *Nat.*
 1122 *Rev. Genet*. 2016;18:51–66.
- 1123 169. Konermann S, Brigham MD, Trevino AE, Joung J, Abudayyeh OO, Barcena C,
 1124 et al. Genome-scale transcriptional activation by an engineered CRISPR-Cas9
 1125 complex. *Nature*. 2015;517:583–8.
- 1126 170. Zalatan JG, Lee ME, Almeida R, Gilbert LA, Whitehead EH, La Russa M, et al.
 1127 Engineering complex synthetic transcriptional programs with CRISPR RNA
 1128 scaffolds. *Cell*. 2015;160:339–50.
- 1129 171. Sgro A, Blancafort P. Epigenome engineering: new technologies for precision
 1130 medicine. *Nucleic Acids Res*. 2020;48:12453–82.
- 1131 172. Bell CG. The emerging potential for epigenetic therapeutics in noncancer
 1132 disorders. *Drug Discov Cancer Epigenetics*. 2015. p. 437–56.
 1133 doi.org/10.1016/B978-0-12-802208-5.00017-5
- 1134 173. Ou K, Yu M, Moss NG, Wang YJ, Wang AW, Nguyen SC, et al. Targeted
 1135 demethylation at the CDKN1C/p57 locus induces human β cell replication. *J Clin*
 1136 *Invest*. 2019;129:209–14.
- 1137 174. Das S, Senapati P, Chen Z, Reddy MA, Ganguly R, Lanting L, et al. Regulation of
 1138 angiotensin II actions by enhancers and super-enhancers in vascular smooth
 1139 muscle cells. *Nat Commun*. 2017;8(1):1467
- 1140 175. Wasiak S, Gilham D, Tsujikawa LM, Halliday C, Calosing C, Jahagirdar R, et al.
 1141 Downregulation of the Complement Cascade In Vitro, in Mice and in Patients with
 1142 Cardiovascular Disease by the BET Protein Inhibitor Apabetalone (RVX-208). *J*
 1143 *Cardiovasc Transl Res*. 2017;10:337–47.
- 1144 176. Gallo P, Latronico MVG, Gallo P, Grimaldi S, Borgia F, Todaro M, et al.
 1145 Inhibition of class I histone deacetylase with an apicidin derivative prevents
 1146 cardiac hypertrophy and failure. *Cardiovasc Res*. 2008;80:416–24.
- 1147 177. Bai XJ, Hao JT, Wang J, Zhang WF, Yan CP, Zhao JH, et al. Curcumin inhibits
 1148 cardiac hypertrophy and improves cardiovascular function via enhanced
 1149 Na⁺/Ca²⁺ exchanger expression after transverse abdominal aortic constriction in
 1150 rats. *Pharmacol Reports*. 2018;70:60–8.
- 1151 178. Helmstädter J, Frenis K, Filippou K, Grill A, Dib M, Kalinovic S, et al.
 1152 Endothelial GLP-1 (Glucagon-Like Peptide-1) Receptor Mediates Cardiovascular
 1153 Protection by Liraglutide In Mice With Experimental Arterial Hypertension.

- 1154 Arterioscler Thromb Vasc Biol. 2020;40:145–58.
1155 179. Zhang E, Guo Q, Gao H, Xu R, Teng S, Wu Y. Metformin and resveratrol
1156 inhibited high glucose-induced metabolic memory of endothelial senescence
1157 through SIRT1/p300/p53/p21 pathway. PLoS One. 2015;10:e0143814.
1158 180. Plutzky J. The PPAR-RXR transcriptional complex in the vasculature: Energy
1159 in the balance. Circ. Res. 2011;108:1002–16.
1160 181. Cardinale JP, Sriramula S, Pariaut R, Guggilam A, Mariappan N, Elks CM, et al.
1161 HDAC inhibition attenuates inflammatory, hypertrophic, and hypertensive
1162 responses in spontaneously hypertensive rats. Hypertension. 2010;56:437–44.
1163 182. Sciarretta S, Boppana VS, Umapathi M, Frati G, Sadoshima J. Boosting
1164 autophagy in the diabetic heart: a translational perspective. Cardiovasc Diagn
1165 Ther. 2015;5:394–402.
1166