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Sustained heterozygosity across a self-incompatibility locus in an inbred ascidian

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ABSTRACT

Because self-incompatibility loci are maintained heterozygous and recombination within self-incompatibility loci would be disadvantageous, self-incompatibility loci are thought to contribute to structural and functional differentiation of chromosomes. Although the hermaphrodite chordate, *Ciona intestinalis*, has two self-incompatibility genes, this incompatibility system is incomplete and self-fertilization occurs under laboratory conditions. Here, we established an inbred strain of *C. intestinalis* by repeated self-fertilization. Decoding genome sequences of sibling animals of this strain identified a 2.4 mega-base (Mb) heterozygous region on chromosome 7. A self-incompatibility gene, *Themis-B*, was encoded within this region. This observation implied that this self-incompatibility locus and the linkage disequilibrium of its flanking region contribute to the formation of the 2.4-Mb heterozygous region, probably through recombination suppression. We showed that different individuals in natural populations had different numbers and different combinations of *Themis-B* variants, and that the rate of self-fertilization varied among these animals. Our result explains why self-fertilization occurs under laboratory conditions. It also supports the concept that the *Themis-B* locus is preferentially retained heterozygous in the inbred line and contributes to the formation of the 2.4-Mb heterozygous region. High structural variations might suppress recombination, and this long heterozygous region might represent a preliminary stage of structural differentiation of chromosomes.

Introduction

Balancing selection maintains diversity of self-incompatibility loci (Wright 1939), and self-incompatibility loci are maintained heterozygous, because they prevent self-fertilization. Hence, self-incompatibility loci could cause linkage disequilibrium of flanking regions, and recombination suppression (Uyenoyama 1997), and might eventually cause structural differentiation of chromosomes, as a sex determining gene does in sex chromosomes (Eichler, Sankoff 2003; Fraser, Heitman 2004; Charlesworth et al. 2005; Uyenoyama 2005; Bachtrog 2013). While this prediction has partially been tested in plants (Kamau, Charlesworth 2005; Llaurens et al. 2009), it could also be tested by inbreeding more directly. First, genomic regions near self-incompatibility loci will be retained heterozygous by linkage disequilibrium. Second, if recombination is suppressed, large regions flanking to self-incompatibility loci will be retained heterozygous.

The genome of *Ciona intestinalis*, which is a hermaphrodite chordate belonging to a sister group of vertebrates (Delsuc et al. 2006), provides a unique opportunity to address this problem. It encodes two self-incompatibility loci, *Themis-A* and *Themis-B* (Harada et al. 2008). These loci are encoded on different chromosomes and are expected to be preferentially maintained heterozygous, because fertilization rarely occurs between sperm and eggs bearing self-incompatibility proteins that are encoded by the same alleles in both loci. Therefore, theoretically, either of the self-incompatibility loci is always heterozygous, and out-crossing is the natural reproductive mode of *Ciona intestinalis* (Morgan 1944). However, the self-incompatibility system of this animal is however incomplete, and self-fertilization occurs in laboratory

conditions.

Each of these incompatibility loci encodes two genes (Harada et al. 2008; Saito et al. 2012); one is expressed in sperm and the other is expressed in eggs. The *Themis-B* locus encodes the genes, *s-Themis-B* and *v-Themis-B*. *s-Themis-B* is expressed from a haploid genome of sperm, and *v-Themis-B* is expressed from a diploid genome of oocytes, and their protein products interact with each other to reject fertilization. These two genes contain a hyper-variable region, which enables a specific interaction between *s-Themis-B* and *v-Themis-B* proteins encoded by the same allele. The *Themis-A* locus has a structure similar to the *Themis-B* locus. An incompatible reaction occurs, only if both *Themis-A* and *Themis-B* alleles are matched between sperm and eggs.

To maintain the incompatibility system of *Themis-A* and *Themis-B*, recombination needs to be suppressed within these loci, because such recombination would destroy their function by rendering gametes carrying different alleles incompatible. However, there are no indication that structural differentiation occurs around the *Themis-A* and *Themis-B* loci, because the heterozygosity of regions close to these two loci is not prominently high compared to other regions (Satou et al. 2012). Hence, if these two self-incompatibility loci do evoke structural differentiation of chromosomes, the process remains at a preliminary stage.

In the present study, we found that a long heterozygous region in the genome of an inbred line of *C. intestinalis* contained the self-incompatibility *Themis-B* locus. We examined a possibility that this locus contributes to the formation of this long

heterozygous region.

Results

Sequencing of genomes of two sibling animals of an inbred line

We established an inbred strain of the tunicate, *C. intestinalis*, from an individual that was caught in Onagawa-Bay, Miyagi Prefecture, Japan. Taking advantage of the incomplete self-incompatibility of this animal (Harada et al. 2008), we repeated self-fertilization. Using sperm, we analyzed genomes of two mature siblings (specimens A and B) obtained after 11 self-fertilization (F11 generation). Using illumina and 454 sequencers, we obtained approximately 1.6×10^8 sequence tags from each of these two sibling genomes (Table 1). Over 1×10^{10} bases were mapped onto the non-repeated region of the reference genome (Dehal et al. 2002; Satou et al. 2008), which corresponded to over 100x sequence coverage. Then, we called genotypes of the 89,770,299 and 89,531,196 nucleotide positions, which correspond to approximately 80% of the reference sequence (Table 1).

We found that 0.04% of nucleotide positions were heterozygous in each sibling genome (Table 1). Genotypes could not be determined for 20 and 19 positions in the genomes of specimens A and B, respectively, which could be due to misalignments of sequence tags to the reference genome. The observed frequencies of heterozygous sites of specimens A and B were much lower than heterozygosity rates in natural populations (1.1~1.2%) (Dehal et al. 2002; Satou et al. 2012), but higher than the rate expected after 11 generations of self-fertilization assuming neutrality ($1.1 \sim 1.2\% \times (1/2)^{11} = 0.00054 \sim 0.00059\%$).

Among homozygous sites, 1.1% of genomic positions of the inbred animals were different from the reference (Table 1). Thus, each of the four haplotypes of the two siblings we analyzed differed from the reference by approximately 1.12% ($=1.1\%+0.04\%/2$). This estimate showed great agreement with the mean nucleotide diversity of *C. intestinalis* (Dehal et al. 2002; Satou et al. 2012), confirming the accuracy of genotype calling.

There are few neutral heterozygous sites that would become homozygous by further inbreeding

To test whether any neutral heterozygous sites or regions were remaining in the genome of the inbred strain, we compared genome sequences between the two sibling specimens. If neutral heterozygous sites or regions remained in the genome of the inbred strain, a quarter of these sites would become homozygous in both specimens, another quarter would remain heterozygous in both specimens, and the rest would become heterozygous in one specimen and homozygous in the other (Hom/Het sites or regions). At 89,068,420 positions where genotypes were determined commonly in both siblings, 2,962 positions were homozygous in specimen A and heterozygous in specimen B, and another 2,820 positions were heterozygous in specimen A and homozygous in specimen B (5,782 Hom/Het sites in total; Table 2).

Deep inspection of raw data revealed that at 5,675 of these 5,782 Hom/Het sites, two heterozygous bases were found in aligned tags in both specimens, although in one animal, frequencies of the secondarily common bases were below the threshold (see Materials and Methods). Thus, these 5,675 (2,894 and 2,781 in specimens A and B,

respectively) sites could have been miscalled in either specimen (not *bona fide* Hom/Het sites). Even if all of these sites were indeed heterozygous, the above estimated frequencies of heterozygous sites are not significantly increased.

The remaining 107 (=5782-5675) Hom/Het sites were considered candidates for *bona fide* Hom/Het sites. Because of genetic linkage, most Hom/Het sites are expected to make clusters on the genome. Of the 107 Hom/Het sites, 56 heterozygous sites were indeed found in a 5,224-bp region of Chromosome 5 of specimen B (Figure 1A–C), and therefore these sites likely represent *bona fide* Hom/Het sites, or multiple copies of this region in one haplotype of the genome of specimen B. In the case of multiple copies, the above 56 sites might not be actual Hom/Het sites, but the corresponding region should be considered as a Hom/Het ‘region’. Indeed, the mean sequence coverage of this region in specimen B (160x) was 1.6-times as deep as that in specimen A (101x). An additional 31 potential Hom/Het sites were found in seven small genomic regions (length = 2, 7, 11, 29, 461, 1,662, and 2,612 bp), although the remaining 20 (=107-56-31) sites were dispersed over the genome. Statistically, half of the neutral heterozygous sites/regions are expected to become homozygous in one sibling and heterozygous in the other. Hence, even if the above 107 sites or 87 (=107-20) sites in eight regions actually represented neutral Hom/Het sites, most heterozygous regions in the parental F10 animal could hardly become homozygous by further inbreeding.

As described earlier, a quarter of neutral heterozygous regions in the parental genome are expected to become homozygous in both siblings, and half of them are expected to be different from each other. There was only one such site at nucleotide position 5,250 of scaffold KHL81 (‘G’ in specimen A and ‘C’ in specimen B). Direct sequencing of

the amplicon of this region of the ancestral F9 genome revealed a homozygous 'C', but showed no indication of 'G' (Figure 2). Therefore, the 'G' nucleotide most likely occurred by mutation between the F9 and F11 generations. This observation again supports the concept that there are few neutral heterozygous sites that could become homozygous by further inbreeding, and that most heterozygous sites found in the inbred strain will be maintained by further inbreeding.

A 2.4-Mb region containing a self-incompatibility locus is retained heterozygous

Although the above analysis showed that most neutral sites became homozygous in the F11 genome, sequence coverage and distribution of heterozygous sites of specimens A and B in non-overlapping 100-kb chromosomal windows showed that an approximately 2.4-Mb region on chromosome 7 is highly heterozygous (Figure 3A and Supplementary Figure 1A). This region contained over 40% of the heterozygous sites we identified, and encoded more than 300 genes. Because the result in the preceding section predicts that this region will not easily become homozygous by further inbreeding, and because the recombination rate in *C. intestinalis* is estimated to be 25 to 49 kb/cM (Kano et al. 2006), this heterozygous region likely lacks recombination.

We confirmed the high heterogeneity of this 2.4-Mb region on chromosome 7 by sequencing an amplicon of a 783-bp long genomic region of the F4 to F9 genomes. These genomes all contained 19 heterozygous sites (Supplementary Table 1).

The detailed view of the 2.4-Mb region of specimen A in non-overlapping 10-kb windows showed that there were two peaks of heterozygosity at the

1,840,001~1,850,000-th and 1,920,001~1,930,000-th positions (Figure 3B). This and a still more detailed view of the regions containing these two peaks in non-overlapping 1-kb windows (Figure 3C) showed that the peak regions encoded a self-incompatibility locus, *Themis-B* (in the reference sequence, there are three *Themis-B* genes). Figure 3C also shows that the observed frequency of heterozygous sites of regions around the *Themis-B* locus, particularly the intervening region, is higher than the mean heterozygosity rate in natural populations (1.1~1.2%; Dehal et al. 2002; Satou et al. 2012) and in more distant regions.

A hyper-variable region is contained in the N-terminal end of s-*Themis-B* (Harada et al. 2008). While no sequence tags were mapped onto the hyper-variable regions of the second and third copies of *Themis-B* genes (middle and right genes in Figure 3C), a considerable number of sequence tags were mapped onto the hyper-variable region of the first copy (left gene in Figure 3C). This *Themis-B* allele appears to be encoded in either haplotype of the inbred strain, because no heterozygous nucleotides were found and the sequence coverage was almost half the average (Figure 3C). Careful manual inspection of Roche 454 tags succeeded in identifying four different alleles of *Themis-B*, which we will further confirm in the following section. In contrast, a substantial number of sequence tags were mapped onto the constant regions of the three *Themis-B* genes. Therefore, the inbred strain likely had multiple copies of *Themis-B*, and sequence tags for their hyper-variable regions were not mapped because of their variability.

In contrast, the genomic region containing the other self-incompatibility locus, *Themis-A*, did not show high heterogeneity (Supplementary Figure 1B). In addition, deep

inspection of Roche 454 tags succeeded in identifying one allele but not multiple alleles. To confirm this observation, we amplified a genomic fragment containing the hyper-variable region of *Themis-A* using a primer for a constant region of the *s-Themis-A* gene with a primer for the upstream region of *Themis-A* from the F9 genome. We obtained only one allele, which was the same as that identified by deep inspection of Roche 454 tags (Supplementary Figure 2). Although we could not determine whether the *Themis-A* locus in the F0 genome was homozygous, the above results strongly indicate that the *Themis-A* locus is homozygous in the inbred strain.

Structural variations of the self-incompatibility locus

The above observation raised the possibility that the *Themis-B* locus, which is thought to be maintained heterozygous, is responsible for the long heterozygous region.

Although a previous study speculated that multiple copies of *Themis-B* are artifacts caused by mis-assembly of the genome (Harada et al. 2008), the above observation implied that multiple copies were indeed encoded in each haplotype of the inbred line.

To confirm that the inbred line has multiple copies of the *Themis-B* locus, we amplified a genomic fragment containing the hyper-variable regions between alleles of *s-Themis-B* and *v-Themis-B* using a primer for a constant region of the *s-Themis-B* gene with a primer for the *v-Themis-B* gene. We obtained four different genomic fragments from the F9 genome and determined the nucleotide sequences of them individually (alleles A, B, C and D in Supplementary Figures 3, 4 and 5). Allele B was identical to the allele encoded in the first copy of the reference genome sequence. Thus, the inbred animals had multiple copies of the *Themis-B* locus per haplotype.

To further confirm this result, we determined the copy number of *s-Themis-B* in the genome of the F9 animal using quantitative PCR (qPCR). Under the assumption that two copies of a control *Macho-1* gene are encoded per diploid, the qPCR assay indicated that six copies of *s-Themis-B* and two copies of another control gene, *FoxA-a*, are encoded in the F9 diploid genome (Figure 4A). Thus, our assay found six copies of four different *Themis-B* alleles in the F9 genome.

Next we examined the genomes of seven wild animals (wt1 to wt7) and found that these animals carried two to ten copies of the *s-Themis-B* gene (Figure 4A). By genotyping of the *Themis-B* locus in the wt1 and wt2 genomes, which have five and seven copies of *Themis-B*, respectively, we identified three (alleles *D-F*) and two (*A* and *C*) alleles of *Themis-B* (Figure 4B and Supplementary Figures 3, 4 and 5). Therefore, multiple copies of multiple *Themis-B* variants are likely encoded in these genomes, and are not specific to the inbred strain.

The self-fertilization rate in association with structural variations of the self-incompatibility locus

Multiple copies of *Themis-B* imply that the self-incompatibility system is not as simple as previously proposed. Indeed, 79% of eggs obtained from the F9 animal (884 of 1117) were self-fertilized. We reasoned that structural variations of this locus should be related to the self-fertilization rate, if the *Themis-B* locus is responsible for formation of the 2.4-Mb heterozygous region. For this purpose, we determined the rate of self-fertilization in association with copy numbers and nucleotide sequences of alleles of *Themis-B* in nine different wild type animals (wt8 to wt16) (Figure 4B, C and

Supplementary Figures 3, 4 and 5). We identified only one allele (allele A) in wt16, the eggs of which were almost completely self-fertilized. This observation indicates that the pair of s-Themis-B and v-Themis-B encoded in this allele does not effectively block self-fertilization. On the other hand, s-Themis-B and v-Themis-B proteins encoded in allele H, which was the only allele found in wt10, apparently reject self-fertilization efficiently (Figure 4B and C). This allele H is most similar to allele F; s-Themis-B and v-Themis-B proteins encoded in these two alleles were 90% and 81% identical, respectively (Supplementary Figures 4 and 5). Two individuals with allele F (wt8 and wt9) showed the lowest self-fertilization rates (Figure 4B and C). Thus, different variants likely have different reactivity, and self-fertility and genotypes of this locus appear correlated.

The high self-fertilization rate of wt15 indicates that alleles *D*, *I*, and *J* do not react very efficiently (Figure 4B and C); if either of these three alleles reacted efficiently, the self-incompatibility reaction would take place (Saito et al. 2012), and no self-fertilization would occur. Wt11 had only alleles *I* and *J*, and showed a lower self-fertilization rate than wt15 (Figure 4B and C). Wt11 had a smaller number of *Themis-B* copies and a smaller number of variants, which might increase the likelihood that a specific type of s-Themis-B expressed in a sperm cell would encounter its counterpart (v-Themis-B) on the egg's vitelline coat. This hypothesis is also supported by the following observation. The self-fertilization rate of wt12, which had three variants including alleles *I* and *J*, was higher than that of wt11, but lower than that of wt15 (Figure 4B and C), while the copy number was larger than that of wt11 and smaller than that of wt15. Although we cannot rule out the possibility that some alleles of the *Themis-A* self-

incompatibility locus did also not react efficiently in wt8–wt16, the above observations strongly indicate that the self-fertilization efficiency is affected by different reactivity among variants, copy number and allele number of *Themis-B*.

Discussion

Establishment of inbred lines

Because of the simplicity and compactness of the genome, as well as simple development, *Ciona* is widely used for experimental biology including developmental and evolutionary studies. Nevertheless no inbred *Ciona* strain has yet been made available, and animals from natural populations are typically used for experiments. Because this animal has a large gene pool (Satou et al. 2012), genetic variations might often have affected reproduction of results obtained from different animals. Deep genomic sequencing of the inbred strain established here has shown that this strain possesses only 0.04% of heterozygous nucleotides, which is much smaller than the nucleotide diversity rate in natural populations (1.1~1.2%). In addition, decoding of the genomes of two siblings predicted that there are few neutral sites that could potentially become homozygous by further inbreeding. Thus, this strain can now be used as a new biological resource.

The self-incompatibility locus *Themis-B*

We identified twelve *Themis-B* alleles in the genomes of the F9 and eleven wild animals, which were derived from animals caught in Onagawa Bay. Eight of these

alleles were shared among multiple individuals. In addition, the *B* allele is found in the reference genome, which is derived from a specimen caught on West Coast of the United States (Dehal et al. 2002). These observations imply that this animal has a limited number of variants of the self-incompatibility gene in natural populations.

The number of variants appears much smaller than the theoretical expectation based on population sizes (Wright 1939; Kimura, Crow 1964); 60 to 90 variants are expected for even a small population of 10,000 individuals. However, our results strongly indicated that different reactivity among variants, copy number variation, and variant number variation all influence self-fertilization efficiency, and a combination of these factors probably increases the functional complexity of this self-incompatibility locus. At the same time, this system makes self-incompatibility between eggs and sperm of *Ciona* incomplete, such that outcrossing occurs more frequently under natural conditions where self-sperm and non-self-sperm are available and self-fertilization occurs in laboratory conditions.

A half of offspring of our inbred line are expected to be homozygous for *Themis-B*.

The observation that this locus is maintained heterozygous indicates that the fertility of animals homozygous for *Themis-B* is lower than the fertility of heterozygous animals, and animals homozygous for *Themis-B* rarely generate offspring. In this sense, this incomplete self-incompatibility locus has strong heterozygous advantage in the inbred strain, and likely causes inbreeding depression, which is widely observed in offspring of related individuals (Charlesworth, Willis 2009).

The large heterozygous region might represent a preliminary stage of structural

differentiation of chromosomes

The recombination rate in *C. intestinalis* is estimated to be 25 to 49 kb/cM (Kano, Satoh, Sordino 2006), and three *Themis-B* genes in the reference genome are encoded within approximately 100-kb. Therefore, the 2.4-Mb heterozygous region in inbred animals strongly suggests that recombination rarely occurs in this region, and it seems likely that this recombination suppression is related to the *Themis-B* locus. First, *Themis-B* is retained heterozygous. Previous studies have shown that large genomic regions segregate together with self-incompatibility or mating type loci in other organisms (Boyes et al. 1997; Lahn, Page 1999; Ferris et al. 2002; Lengeler et al. 2002; Liu et al. 2004). Second, any recombination within the hyper-variable region of *Themis-B* across alleles would probably impair their function, because v-*Themis* and s-*Themis* encoded in each allele specifically react with each other and do not react with those encoded in other alleles. Third, the high degree of nucleotide variability and copy number variations should physically suppress recombination at this locus. Fourth, it is possible that different haplotypes have different arrangements of multiple *Themis-B* copies, including inversions. Such variation may be directly related to recombination suppression in the large regions flanking the *Themis-B* locus, although this hypothesis remains to be tested. Finally, the observation that the frequency of heterozygous sites in the genomic region between the second and third *Themis-B* genes was higher may suggest that this region has started to diverge.

Intervening homozygous regions within the 2.4-Mb heterozygous region (Figure 3B), however, indicate that recombination is not completely suppressed. There may be several loci with heterozygous advantage in the 2.4-Mb heterozygous region, although

we could not find candidate genes with heterozygous advantage other than *Themis-B*, and linked deleterious mutations might also confer heterozygous advantage. Because the observed frequency of heterozygous sites of regions close to the *Themis-B* locus is higher than those of more distant regions within the 2.4-Mb heterozygous region, genes or genomic regions with heterozygous advantage in distant regions and modifiers of recombination rates would have been acquired under the linkage disequilibrium caused by the *Themis-B* locus. Thus, the 2.4-Mb heterozygous region of chromosome 7 might represent a preliminary stage of structural differentiation of chromosomes.

Materials and Methods

Inbreeding and sequence data

A single individual originating from a natural population in Onagawa, Miyagi Prefecture, Japan was chosen as the F0 animal. Although *Ciona intestinalis* is a hermaphroditic and out-crossing is its normal reproductive mode, self-fertilization occurs under laboratory conditions. Each generation, offspring were obtained by self-crossing. DNA extracted from sperm cells of two F11 animals were individually sequenced with illumina GA2 and Roche 454 sequencers.

Mapping and genotype calling

We first trimmed low quality regions (QV<25) from sequencing tags with a 'TrimmingReads.pl' script (Patel, Jain 2012). Obtained tags were mapped onto the reference genome sequence using ssaha2 (Ning et al. 2001) with default parameters for illumina tags and Roche 454 tags. We used nucleotide positions in non-repeat regions that were covered by 20 or more tags because of the error-prone nature of sequence tags. Because nucleotide positions that are too deeply covered might represent repeat sequences, we also excluded nucleotide positions that were covered by more than 203 and 212 tags, which were twice as large as the averaged sequence depths of specimens A and B, respectively (Table 1).

Before genotype calling, we screened out repeats in the reference genome sequence (KH version) (Satou et al. 2008). We first took 100-bp sliding windows with 50-bp steps and aligned them onto the reference genome using blat (Kent 2002) with the

'fastMap' option. Sequence fragments that were more than 50% identical with multiple regions were regarded as repeats. As a result, 15,119,916 nucleotides were found in repeats and were excluded from the subsequent analyses.

Using these mapped sequence tags, genotypes were called using previously published criteria (Harismendy et al. 2009; Nielsen et al. 2011). We called positions with common allele read frequencies over 80% as homozygous, and positions with secondarily common allele read frequencies over 20% as heterozygous, unless ternary common allele read frequencies were over 20%.

PCR genotyping

We amplified a genomic fragment shown in Supplementary Table 1 from the F4 to F9 genomes by polymerase-chain reactions with a set of primers; 5'-ACTGCAAACAAGTTTCCGTTGT-3' and 5'-AGTGTTACTGACTTGAGATTACTA-3'. Amplified sequence fragments were directly sequenced using an ABI3130xl sequencer. Genomic DNA was extracted from sperm cells of the F4 to F9 animals. The genomes of F0 to F3 and F10 were accidentally lost and were not analyzed.

We also amplified a genomic fragment of the scaffold KHL81 from the F9 genome with the following primers. 5'-AACGAATTGTAAAGTACGCTCACGA-3', and 5'-AGAAGCTCTCAGCCAATGAGCGT-3'.

The *Themis-B* locus of the genomes of the F9 animal and wild-caught animals was amplified with the following primers: 5'-CTGGAAAGTTATCCAACCAGTT-3' and 5'-TTGTTGGGATATTT(A/G)TTGATTCT-3'. Amplified fragments were cloned and

sequenced. For each genome, we sequenced at least 18 clones.

The *Themis-A* locus of the F9 genome was amplified with the following primers: 5'-ATTTTCGATCTCAATAGACACCAA-3' and 5'-TGTTACATTCAATGTGCCAAGT-3'.

Measurement of copy number variations

DNA extracted from sperm cells of the F9 animal and wild-caught animals were used to determine copy number variation. We performed two distinct multiplex-quantitative-PCRs for each genomic DNA. We amplified a constant region of the *s-Themis-B* and *Macho-1* genes in the first reaction, and *FoxA-a* and *Macho-1* genes in the second reaction. We adopted a TaqMan method using a TaqMan Universal PCR Master Mix (Invitrogen). Probes for *s-Themis-B*, *FoxA-a*, and *Macho-1* are as follows: *s-Themis-B*, 5'-(FAM)-CAGCGCTATCATTAGAT-(MGB)-3'; *FoxA-a*, 5'-(FAM)-TCTGCCGTTGAAGTTAGTTCGCCATCC-(TAMARA)-3'; *Macho-1*, 5'-(VIC)-ACGGTCACTTTAGCACCTCCACCA-(TAMARA)-3'. As calibrators, we made two DNA constructs, which contained amplicon sequences of *s-Themis-B* and *Macho-1*, and of *FoxA-a* and *Macho-1*. Sequences of these calibrators are shown in Supplementary Figure 6. After calibrating the different efficiencies between probe/primer sets with these calibrators, we calculated expected numbers of *s-Themis-B* and *FoxA-a* per diploid, assuming that *Macho-1* exists in a single copy gene per haploid (two copies per diploid). Standard errors were calculated among triplicates. PCR primers were as follows: *s-Themis-B*, 5'-TGATGAATGTAAATTGGTTCAAGTCAA-3' and 5'-TGAGGAACGGTTTCAAACACTTG-3'; *FoxA-a*, 5'-

TTCAACACCACCACACTCAACAG-3' and 5'-CGTGTTCAATGCCATGTTC-3';
Macho-1, 5'-CCCAGTATGCACCAAATTCAGA-3', and 5'-
TGGTGAGAAAACGGGTGAAAC-3'. The fertilization ratio was determined as
previously described (Harada et al. 2008).

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Figure Legends

Figure 1. The number of sequence tags aligned to the region from the 303,101-th to 317,100-th positions on chromosome 5. At each position, the number of the nucleotide that was most frequently found is shown by black dots, and the number of secondarily most frequently identified nucleotide is shown by red dots. Regions where no sequence tags are aligned have no dots. Two siblings are shown in **(A)** and **(B)**, respectively. Two genes encoded in this region are indicated in **(C)**. Exons are shown by black boxes and introns are shown by thin lines.

Figure 2. A chromatogram showing homozygosity of the 5,250-th nucleotide position of scaffold KHL81 of the F9 animal. While the deep sequencing data indicated that this position is homozygous ‘G’ in specimen A and homozygous ‘C’ in specimen B, direct sequencing of a genomic fragments amplified from the ancestral F9 genome revealed that this position is actually ‘C’ (an arrow).

Figure 3. A chromosomal distribution for heterozygosity in specimen A. **(A–C)** Sequence coverage (blue lines) and frequencies of heterozygous sites (red lines) **(A)** in non-overlapping 100kb chromosomal windows across the 14 chromosomes, which comprise 68% of the current assembly, **(B)** in non-overlapping 10kb windows across the highly heterozygous region on chromosome 7, and **(C)** in non-overlapping 1kb windows across the region encoding the *Themis-B* locus. Positions of *Themis-A* and *Themis-B* are shown by arrows in **(A)** and **(B)**. In **(C)**, the constant regions of three copies of *Themis-B* are indicated by with black arrows. Arrows shows directions of s-*Themis-B* on the genome. The hyper-variable region of the first (left) copy of *Themis-*

B is shown with a red line.

Figure 4. Allele variants, copy number variation, and variant number variation of the *Themis-B* locus may affect the rate of self-fertilization. (A) Copy numbers of *FoxA-a* (gray bars) and *s-Themis-B* (red bars) in eight animals including the F9 inbred animal. Error bars indicate standard errors among triplicates. (B) Alleles found in the present study are indicated by red boxes. (C) In addition to the copy numbers of *FoxA-a* (gray bars) and *s-Themis-B* (red bars) in nine animals, numbers of *Themis-B* variants identified by PCR cloning (blue bars) and the rate of self-fertilization (a black line) are shown. Error bars on gray and red bars indicate standard errors among triplicates.

Table 1. Statistics of sequencing data and mapping data.

	Specimen A	Specimen B
Number of Illumina sequence tags	158,996,466	162,332,248
Total length of Illumina sequence tags	15,693,529,667	16,034,418,242
Number of 454 sequence tags	953,649	860,633
Total length of 454 sequence tags	130,066,159	102,565,614
Total nucleotide length	15,823,595,826	16,136,983,856
Total number of nucleotides mapped onto the reference genome sequence (112,162,187 bases)	11,393,757,651	11,895,122,867
Mean sequence depth	101.5x	106.1x
Total number of nucleotides mapped onto the repeat-masked genome sequence (97,042,271 bases)	10,032,892,695	10,576,941,089
Nucleotide positions where genotypes are called	89,770,299	89,531,196
Heterozygous positions	38,072	39,177
Homozygous positions	89,732,207	82,492,000
Homozygous positions that are different from the reference sequence	991,115	992,810
Ambiguous positions	20	19

Table 2. Comparisons of heterozygous and homozygous sites between the two sibling specimens.

	Number of sites
Genomic positions whose genotypes were called in both siblings	89,068,420
Homozygous sites that are identical between the siblings	89,028,708
Homozygous sites that are not identical between the siblings	1
Heterozygous sites that are identical between the siblings	33,903
Heterozygous sites that are not identical between the siblings	0
Sites that are homozygous in the specimen A and heterozygous in the specimen B	2,962 (68*)
Sites that are homozygous in the specimen B and heterozygous in the specimen A	2,820 (39*)
Ambiguous sites	26

* Candidates for *bona fide* Hom/Het sites (see Materials and Methods).

Fig.1

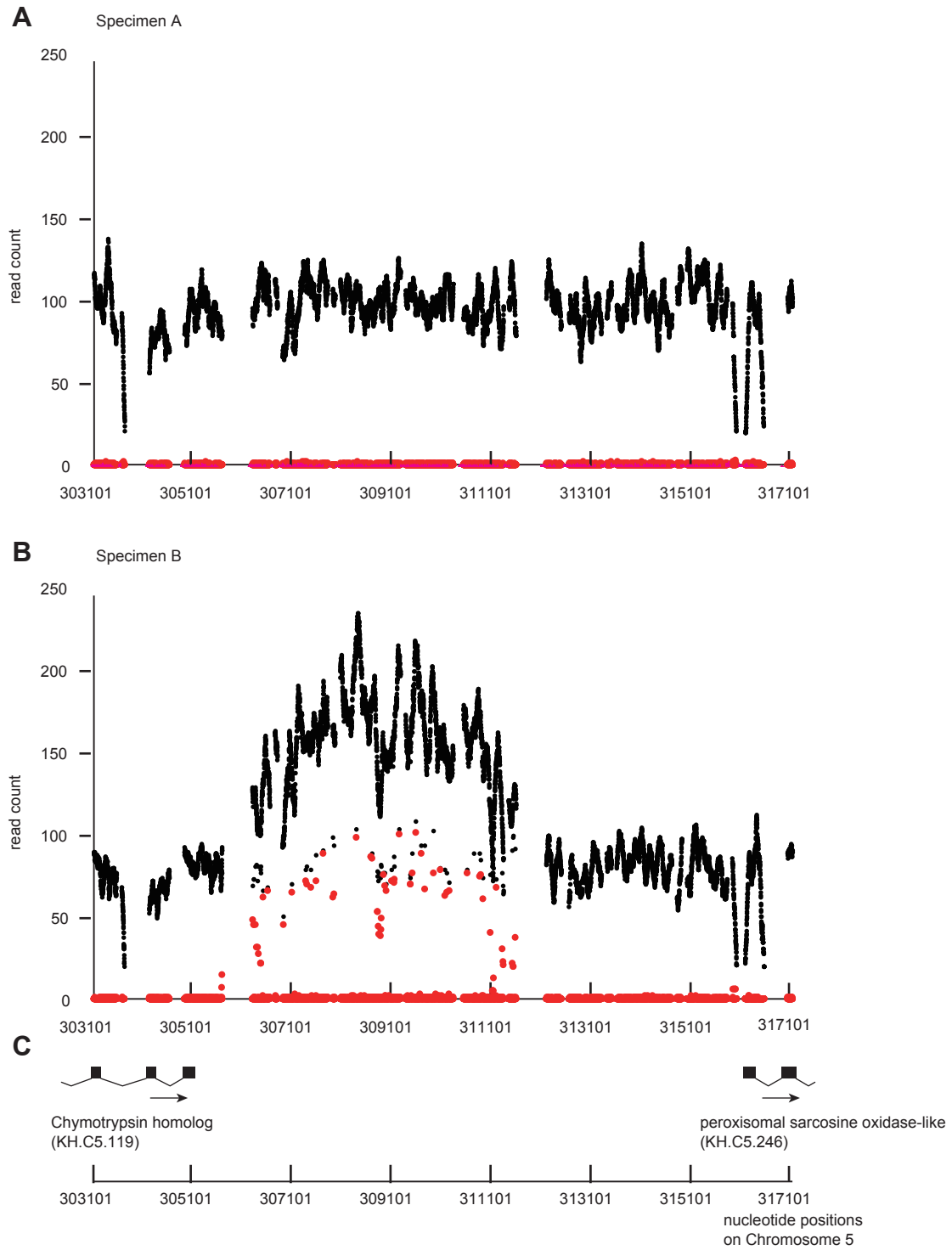


Fig.2

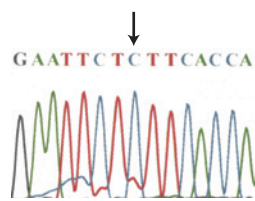


Fig.3

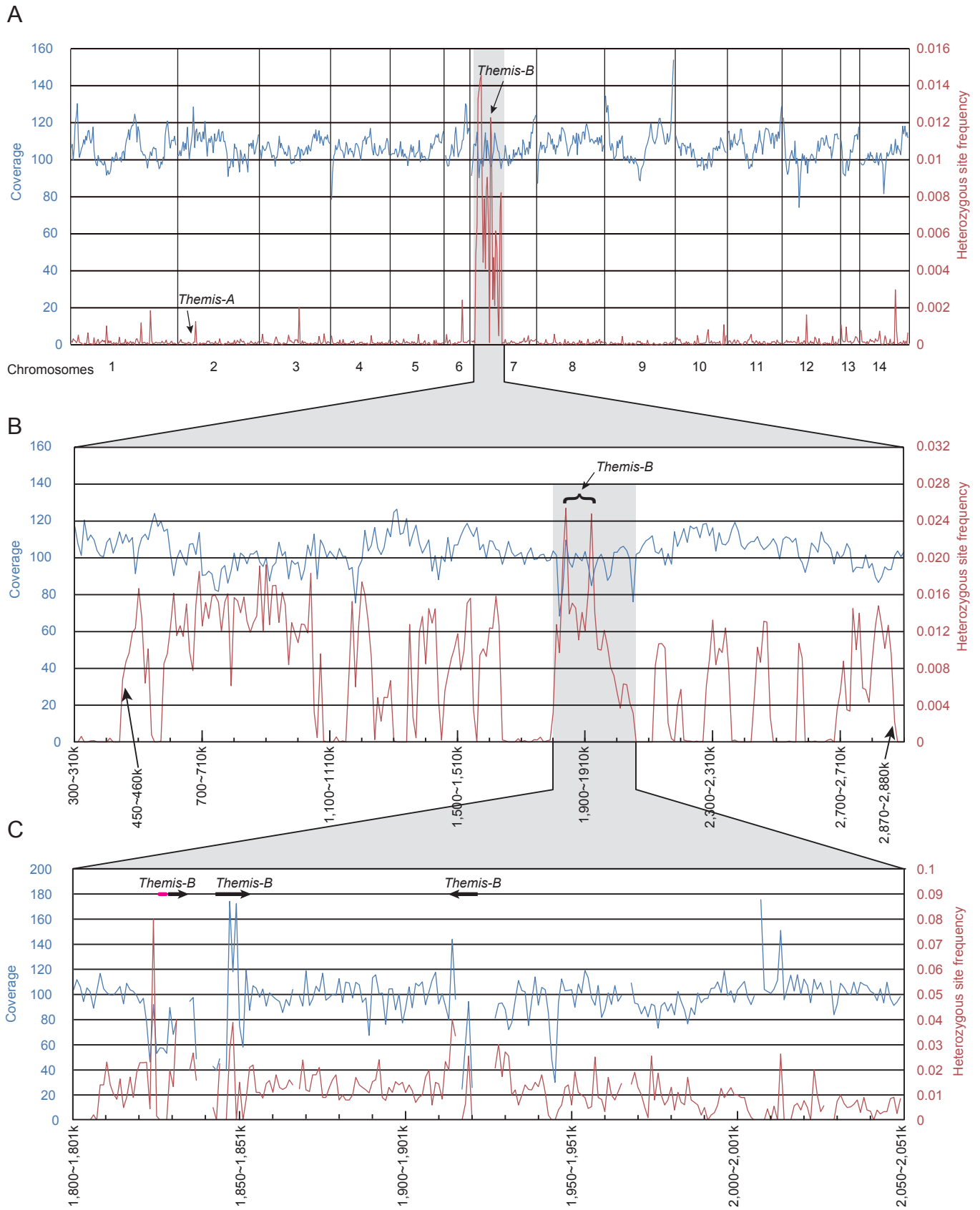
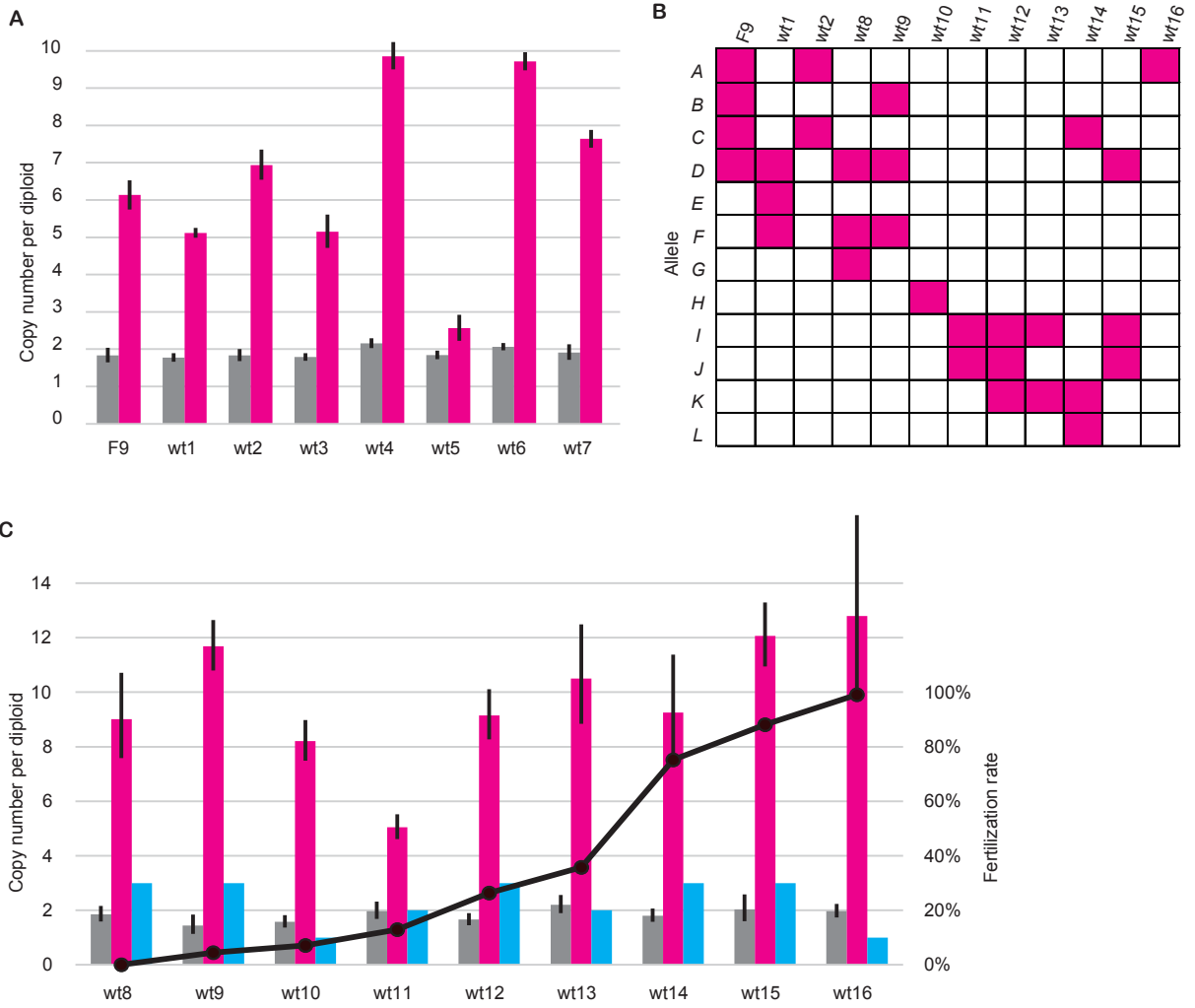
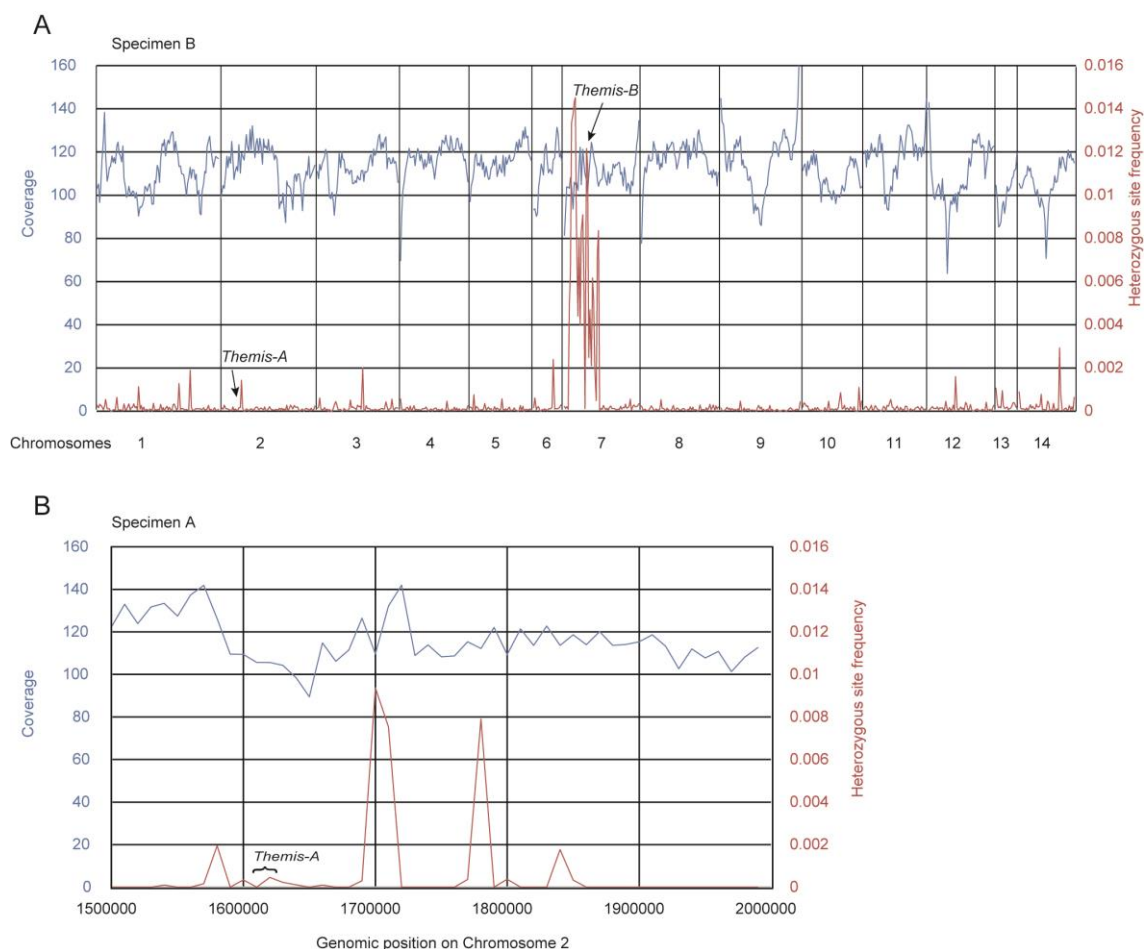


Fig.4





Supplementary Figure 1. Distributions for heterozygosity in specimen B across the 14 chromosomes and in specimen A across the region encoding *Themis-A*. Sequence coverage (blue lines) and frequencies of heterozygous sites (red lines) (A**) in non-overlapping 100kb windows and (**B**) in non-overlapping 10kb windows are shown.**

[Supplementary Figure 2 (1/2)]

A

at t t c g a t c t c a a t a g a c a c c a a a g a t a a c a g c a a a t g a a g a a a t t a t t a c a t t t t t a g c t a t g g g t g
 t g a a a t t t g g t t a c t c t g t t t c a c g t g g a c g a a g t c a c a c a g t a a a g t a c g c c g t g g a a t c c a g c t t
 c a g t t g t g t a c c a g t g c t t t a t a a a a t a a a a c t c c a a c a a a t t g c t g a t g t t a t a t a a t t a t t t t t a
 c t g t c t t g a a c a a t t g c a a t t a g t g a t g t c a a a c t t t t c t t t g t t t a t a t c t t t t a t a t t t a c a g g
 a g a a g t t t t t c c t t a a g a a c a c a t t a t c c a g c a a c a t c a c a c a a a a a g c a a t t a g c a a g t a a a g t c

tgcttttggaa **ATGAAGTGGATTTTTTCATACGCTGTTAATAGTTTTATTACAATTTTAACGTCGTTAACC**
AAGgtaagtacgatattggcttaaattccttaagaaaaacttcattaaaaataactaaacatgtctattat
 caccatactaactattgcgattgaaagtattaacatcgtatttgggtgtgtataccacagcagcagcagta
 aatggtaatTTTTgttctatttcccgttttatacttaac **TTAATACTGAAATTTTCATGGAGACGGACTT**
AAATGCTGTATTATTTTTCGTTTACCCTGTACCAGTCATACCAGTAAATATTGGAAGGGGAGTTTTTATC
TCCTATTTGAGGATAAGGACCGTTTAAATTAGATTCGCCGCAAGCTAGAAACCACCAGCCACTCTGAAA
TGTTCCGGGCATCCTGAAATGCATTATTATTGTGCAATGTGCTAAACCAGTGATTGAAATCGGAGT
AGGAACCCCGGGGGGTATAGCTGTTGCTTGATCAAGATAGGCAGCAATCAAGCGATACTCATCTTTTGC
CCCCGCTACTTTAAACAGCTCATACTCAGCGTACGCAGTAACTCCGTCCTGATCAACAAAATCTATTTCG
CATACGTGGTGGAGTAACATAACTTCCAATCCAATCAATAGTTGTATTTGGTTGCTTAAAGCATGGAT
ATGCTCCAACCCAATCCAATATTCTTCACGAACATTTCCAATCCGTC AACATAATTTTGCCAACCTCT
GTCAAATTAATAGCCCCATTTACTCTCTTTTGTATAGTAATCCATCCCGTTTTATTACTTAATGCTTG
AACATCCATGTCGAAAATACTTTGGTAAACTTGTACAATTGATTTATCCAAATTCATGAACACCCT
TCTTACATAACCAAGCATCATATATTTTCGTTGCAATctaaaatagttgtaagtgttaaggctgtctttaa
 aatgtatttaaacatgtttgcatgatgtacgttagtactagaaaataacatac **CTTGAAATAAAGAATT**
AATAGTATTTTTTTGTGCGATTGTAGCCTGTTACATTATCTTCTGGCTGTTTCAATTCATTGGCGGTAAT
AATCGAGCAATCCTTTCTTTGCTGTTGTGCTAGATTTTTAATGTTTGTAACCTTCCTTGCGATTTTTGT
TATCCTCAGTTGTTCTTTAAAATTATTCATTTTAAAGTTTGTCTCATTATTGTATGCTAAAATCAGATT
CACTACAAGTAATACCAACACTATGCAGCACATATAGTCGGACATatcacctccttttgtatctatgc

v-Themis-A ←

ttacattcagtttgcacttcacag **CAATCGCGAACATTTATTGAAATAACAGAAGCAAAGGTGAAATTA**
CCTCTCCTTCTTCATCGGAATGTAATGAAGGCTTTTTATACAAAGTCTTGGAAATTCAGTTTTTCAACCG
ACTCCGTTTTGTTTGTAACTTTACCAACTTTAATATAACACGgtgagtgcgaaatggataaatatatag
 tatagtgggggaagatggaacatcttagcacgtattgccaaatatttccaaaatcaaaaaatggcgac
 tttgaggtaataccacgaattagtagtactatcatatttctttattaattgacaacaattaaaaaataata
 ataaaatagtcgatcagttagttagcgtatcgcacaacaggtgtaatggacgttatattacgtaaaact
 cggtgaaatttactagtttgcagtttgaatcaccgatgtatagtagtttaataagtaaaatagctgaaat
 atttattatacagtttgatttaaggtaaaatgacacatgtctttttttgtgaatatttatacagatagg
 gtgggaaaagacggcacctaataacatatacgtataattgaactatatag **ATGTAAGGAGGCCACTCTT**
GTAGTTGGCAGCGTTTCACGAGAGACTTACTGTGAGGATCAAATACCTTCAACCTTTTTTGCCTCGGAA
ACCATCACAATTACATTTACATCATATAGACCGCGTGTCTGACTTGTTTCATTTGATTTATAAGATC
ATACgtaagtacattcgtaatttttaaaattgtgggcactattatttacccttataaccagtgacgcat
 atttatttgatatag **GTATTACAGTGACGGTATTAACCCCTCTTTTAATTACTTTGTTGGAAAGAAAA**
TAAGTTTTCTCGTTGACGTTACATATGTTCCCAAGCATGAGCTTGAGTTGTTGTGTTTCATTACCTGTA
ATCGAAGAAGACACGAGCTTGCCAATACTACAGAAAATAGTGTGTCATGTGATTTCAAATTCCTGGAAGAG
CTCCATTTCTCGGTGAATGTGAATCGATCTTACCGAGATTAAGAATTAATAAATAAGATAAATTAATTT
TGAAAATAAACTGTTTCACAAAAACTTGCAGCTGCGACACATTCAAACGAAAAGTTCAAAGAATGGTA
ACGCTGACCCAAACGGCATAACGATATCGCAGCTATTTATATATCCTATTTTCATACACGCAAGTGTG

s-Themis-A hyper-variable region ← *s-Themis-A* constant region

GAGATTCCTCGGTAACAATATCTTTTACTCCAAAACCATTTGACCTATCGAAATACACAAGTCAATCAA
GAGAAGAAATTAAGTTTCAATTACAACCACAACAAGTTACCTCACTTGGACCAGGCCAGCAGCATTTTC
ACCTAGTCTTACAAAACAACGTTTCTTACGTTACATACACTTTTCCAATATATATTAACGAAAACTTG
GCACATTGAATGTAACA

[Supplementary Figure 2 (2/2)]

B

F9 genome **MSDY**CC**IVLVLLV**NL**L**LAY-----**NNE**TNFK**NN**KE**QLRITK**IA**RK**VT**TN**IK**NLR**Q
 Reference **MYRI**Y**C**IL**LLLLLE**QNR**I**ETL**DSK**SQR**I**NT**TN**LT**V**DD**YK**QL**K**L**I**N**V**RK**I**P**N**IK**N**F**Q**K****

F9 genome **QRKDC**SI**I**T**A**N**E**L**-KQ**P**E**D**N**V**T**G**Y**N**RK**N**T**I**N**S**L**F**Q**D**C**N**E**I**Y**D**A**G**V**V**R**S**G**V**H**A**I**W**I**N**Q**L**Y**
 Reference **QQT**I**C**S**G**I**C**T**T**N**Q**V**N**L**Q**R**D**N**A**T**-R**L**RQ**N**A**R**K**I**I**VY**Q**D**C**K**AM**Y**D**A**G**V**V**R**S**G**V**Y**P**I**W**I**Y**Q****L****Y****************************

F9 genome **KFT**V**V**C**D**M**D**V**Q****A**S**N**K**T**G**WI**T**I**Q****K**R**V**N**G**A**I**N**F**D**R**G**WQ****NY**V**D**G**F**G**NV**R**E**E**Y**W**I**G**L**E**H**I**H**A**
 Reference **KFT**V**V**C**D**M**D**V**Q****A**S**G**D**K**N**G**WI**T**I**Q****K**R**V**N**G**A**I**N**F**D**R**G**WQ****NY**V**D**G**F**G**NV**Y**C**E**Y**W**I**G**L**E**H**I**H**A************************************

F9 genome **L**S**NQ**NT**T**I**D**W**I**G**SV**T**P**P**RM**RI**D**F**V**D**Q****D**G**V**T**A**Y**A**EY**E**L**E**K**VA**G**A**K**D**E**Y**R**L**I**A**A**Y**L**D**Q**A**T**A**
 Reference **L**T**YQ**NT**T**I**D**Q**FG**SV**T**P**P**RM**RI**D**F**V**D**Q****D**G**V**T**A**Y**A**EY**RV**VS**G**A**K**Q**K**Y**R**L**I**A**A**HL**D**Q**A**T**A**

F9 genome **I**P**P**G**V**P**I**P**S**N**H**W**F**S**T**E**D**N**N**A**F**P**G**C**P**E**T**F**Q****S**G**W**F**L**A**C**G**E**S**N**L**N**G**P**Y**Q**I**G**D**K**N**S**P**S**N**I
 Reference **I**P**P**N**G**S**P**I**Y**R**W****F**S**I**V**D**N**N**Y**V****F**S**N**C**P****E**K**L****H**G**G**W****F**S**CP**R**S**N**L**N**G**L**Y**P**T**Q**G**E**K****N**S**P**S**N**I********************************

F9 genome **Y**W**Y**D**W**Y**T**V**N**E**N**N**T**A****F**K**S**V**S**M**K**F**Q**Y**
 Reference **Y**W**Y**D**W**Y**T**V**N**E**N**N**T**A****F**K**S**V**S**M**K**F**Q**Y******************

C

F9 genome **M**K**W**I**E**T**L**L**I**V**Y**N**F**N**V**N**Q**A**I**A**N**I**E**T**T**E**A**K**G**E**I**T**S**P**S**S**E**C**N**E**G**F**T**K**S**W**K****F**S**F**S**T**D**S**
 Reference **M**K**W**I**E**T**L**L**M**V**F**Y**N**L**I**F**V**N**P**A**N**P**K**V**V**E**T**E**A**E**G**E**I**T**S**P**S**E**S**I**CY**N**G**F**M**Q**I**W**K****F**S**F**S**A****D****S**

F9 genome **V**L**F**V**N**E**T**N**F**N**I**T**R**C**K**E**A**T**L**V**V**G**S**V**S**R**E**T**Y**C**E**D**Q**I**P**S**T**E**F**A**S**E**T**I**T**T**F**T**S**Y**R**P**-R**C**S**D**L**F
 Reference **A**L**F**L**Q****E**I**N**F**N**I**T**R**C**D**E**A**T**L**V**V**D**S**F**S**R**S**V**Y**C**E**D**E**S**P**P**T**I**E**V**I**D**T**T**E**V**T**F**T**S**Q**T**P**Q****H**C****S**G**V****F**

F9 genome **H**L**I**Y**K**I**R**L**T**V**T**V**L**T****P**S**E**N**F**V**G**K**K**I**S**F**L**V**D**V**T**Y**V**P****H**E**L**E**L**L**C**S**F**T**C**N**R**R**R**H**E**L**A**N**T**T****E**
 Reference **H**L**I**Y**K**I**R**L**T**V**T**V**L**T**P**S**E**H**R**R**V**G**S**E**I**S**F**L**V**D**V**K****H**L**P**H**Y**N**L**Q****L**K****C**S**L**I**C**N**K**M**V**K**L**A****N**T**T****E**

F9 genome **I**V**S**C**D**E**F**K**I**P**G**R**A**P**F**S**G**E**C**E**S**I**L**P**R**L**I**K**K**I**D**K**L**N**L**E**N**K**L**F**H**K**N**L**Q**L**R**H**T**Q**T****K**S**S**K**N**G**N**A**D
 Reference **I**V**R**F**M**S**V**P**G**S**V**Q****F**I**G**E**C**E**T**A****L**S**R**L**K**I**E**K**L**G**E**L**H**L**E**N**Q**L**S**Q****N**D**L**E**V**K**Y**S**Q**N**K**---**M**N**Y****G**********

s-Themis-A hyper-variable region ← → s-Themis-A constant region

F9 genome **P**N**G**I**T**I**S**H**V****I**Y**L****P**I**S****Y****N**A**S****V**G**D**S**S****V**T**I****S**F**T****P**K**P****F**D**L****S**K**Y****T****S**Q**S**R**E****E**I**K****F**Q**L****Q****P**Q**O****V****T**S**L****G**
 Reference **S**N**D**V**I****S**H**I****Y****L****P**I**S****Y****S**V**S**I**G****N**S**S**V**A****M**S**L****P****P**K**P****F**D**L****S**K**Y****T****S**H**S**K**E**E**A****I****F**Q**L****Q****P**Q**O****V****T**S**L****G**

F9 genome **P**G**Q****H**D**F****H**L**V****L**Q**N****N**V**S**Y**V****T**Y**T****F**P**I****Y**I**N****E**K**L****G**T**L****N**V**T**
 Reference **P**G**Q****H**D**F****H**L**V****L**Q**N****N**V**S**Y**V****T**Y**N****F**P**I****Y**I**N****E**K**L****G**T**L****N**V**T**

Supplementary Figure 2. A *Themis-A* allele found in the F9 genome. (A) A genomic fragment encoding *v-Themis-A* and a hyper-variable region of *s-Themis-A* that were amplified using PCR from the F9 genome. Putative exons of *v-Themis-A* and *s-Themis-A* are shown in red and green capital letters, respectively. Introns are shown in black lower case. PCR primers are indicated by underscores. (B, C) Alignments of (B) *v-Themis-A* and (C) a hyper-variable region of *s-Themis-A* variants encoded in the F9 genome and the reference genome. Identical residues are highlighted in black, and similar residues are highlighted in gray.

[Supplementary Figure 3 (1/5)]

A CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTCCCATGTGCGAATAAATGTGAATAAACCGATATCCAACCTTTAACCAGATT
B CTGGAAGTTATCCAACCAAGTTTGTGGTTATGCGGCTACCAGTTTCCATGTGCAAAATGAATAAAAAAACCAGATATCCAACCTTTCAACCAATA
C CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAAATGTGATGAAGCGATATCCAACCTTTAACCAGATT
D CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAAATGTGATGAAGCGATATCCAACCTTTAACCAGATT
E CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAAATGTGAACAAACCGATATCCAACCTTTCAACCAATT
F CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAGATGTGAACAAACCGATATCCAACCTTTCAACCAATT
G CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAAATGTGAATAAACCGATATCCAACCTTTAACCAGATT
H CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAGATGTGAATAAACCGATATCCAACCTTTCAACCAATT
I CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAATAAATGTGAATAAACCGATATCCAACCTTTCAACCAATA
J CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAATAAATGTGAATAAACCGATATCCAACCTTTCAACCAATA
K CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAGTAAATGTGAATAAACCGATATCCAACCTTTAACCAGATT
L CTGGAAGTTATCCAACCAAGTTTGTGGTTATGTGGCTTCCACTTCCATGTGCGAATAAATGTGAATAAACCGATATCCAACCTTTCAACCAATA

A GGATAAACATCGTTGACATGTTTCCAGATTGAAGTATACCATTGCAGTct-----
B GGATAAACGTCGCTGCTGTATCTTCTAACCCTGAAGATTGTACAATct-----
C GGATAAACATTAAGTCTGCTTTTCCAAATGAACAATGCTTTACAATct-----
D GGATAAACATCACTGACGTAATCCAGATAGATGAATATAGTGCAGTct-----
E GGATAAACGTCGCTGCCATATCTTCCAGATTGAAGATACCGCTGCAGTct-----
F GGATAAACGCCCTGTCATATATCCAGATATATAATATCGTTGCAGTct-----
G GGATAAACGTCGGTGACGTTTCCATGTTGAAGAATACCATTGCAGTctgtgaaaaataataactagtttatttatatttacgactgagagtttt
H GGATAAACGTCGCTGTCATATATCCAGATTGAATAATACCGCTGCAGTct-----
I GGATAAACATCGCTGACATATCTCCAGATGGAACAACCTTTTACATTct-----
J GGATAATATTTCACTTCTCCGTTTATAACGTTGAATCCAATGACAATct-----
K GGATAAAA---TGCTGCCATTTTATCGGACTGAATAACACTTTTACAATct-----
L GGATAACGTCGCTAACATGTTTCCAAATGAATAATATCATTGCAATct-----

A -----gca
B -----g
C -----attgta
D -----gt
E -----
F -----atga
G agactgtcttacttaagaaaaacagatcgaccatatttcttagtaataaaaaacacaacgagttttgacttgctctttgaaacaataacctgcagttga
H -----
I -----
J -----aacgcatatacaaaaggact
K -----
L -----gaaag

A tgtaaaagtaaaacttagttagctgtaaacagcattcgcggttttaagagccactacCGAAATACAGCCTTCCAATTGCCAAAATCCGTTTAATTT
B ctttgcaaaaaatagtttaatttaaatgatctagactaaaatgtagaccatcctacCTGTGTTAAACTTTGAATTGCTGACGAGTTCATTAATAT
C aaaaaaatagcaaatattttaatacaatataatattttgagcatgctgataacCTGTATATAAGCTTTGAATTGCCGCCAAAGACAAACATTT
D tggaaaatcatacgcggtttattttgacaactataaacgaagagttttgactctcgtaacCTGTATTTAAGTTTGGAGCAGCGTCAAAGCATTCCTTTT
E gaaaataatgtaattagaataataagcttagtttaaaaaatttgacagcctgtacCTGTATAAAATCTCTTAATGGCCATTATTTTTTCAATGT
F aaaaaagttatttaaatataataagcttagtttcgaggttttaacagcctgtacCTGTATAAAATTTCTTAACAGCCATCTTTTTTTCAAAAGT
G gcttgcgatgtttttgccttagcttactaaaagctatatgcaactggtgctgacCTGTGTTCAAGTTTGGAGCTGCCGCCAAAGCATTCGCTTT
H -gaaaataagtaattagaataataagcttagtttttagattttaaacaacctgtacCTGTATAAAATTTCTTAATGGACATTATTTTTTCAATGT
I ---ggtataaaaaataacttttactatttaacaacagcctaataatagattatcgtaacCTGTGTTTACATTTTGAATTGCCCTTTACAATCTTAGTGT
J ttagtaaatctgtaagatccgttgtaacactttcttttagatatattttttagcttacCTGTAAAAATGCTTTTGATACGGATAAAGCGTTGATATT
K ---ggttcatggagaacagtaagttgtcatatcatggttattttcgacagcttacCTGCGTAAAAGTTTGAATTGCCAACGAGTTGAACAGTTC
L taaagtcaaaaacagttcaaatatttaaaaaaaccttttatctatttaaacataacCTGTGAATAAATTTCTTAATGGCGAGAAATATCCAATTT

A TGGGTTCTCTT-----AGAAACTAAACCATGTTTAACTGGGTAGTCTGTTGCTAAAAAATAGTGGTAACTTCACATCTGTTCTGTGTAAG
B CTTTTCTCTT-----GGAAAATGAACCTGCTTATATTTAAATGATCTTTACAAAACAGGTCACCCGAAACCTTGCATATTTTACACGTATG
C AAAATCGTCTT-----CAGAAATAGGACGAATGTAATCGTTGGTAACCTTTCACTAAT---CCGCTGATAATTTGCATCTGTCCTATGCTCA
D TCGCCAACTTC-----AAATAACTAACTTTAAGCTTTCGTATTTCTTTTCCAAT-----ATATCTTACATTTCTCATTGCGTAAA
E TGCATCATCTCT---GTGAGGATCTAAACGAACCATGAACCTGGGTATTCTTTTACTAATAGATCGCCGGTAAACTGACATTTGTTCTTACGTTCC
F TGCATCTCTGTAAGCATTAACCTCAAACGATCCATGAACCTTAGGATTTCTTTTACCAATAGATCGCCGATAAACTTACATTTGTTCTTACGTTCC
G TGAGTCTTTCTT-----AGAAAACCACTAACTTTGAGGTTGCGGGATTTCGTTTACTAAT-----TTTTTCTCACAGTACTCATTGCGAAC
H TGCATCTCTGTG-----AGGATGTAACGATCTATGAACCTGGGTATTCTTTTACTAATAGATCGCCGATAAGCTTACATTTGTTCTTACGTTCC
I ATCCCTTTCTT-----TGAAAATAAATTAAGTTAAGCAATTCAGTCTGTAATAATGATTAAGCTTAACTTTACATCTGTACTTGCCTAGAC
J CTGAAACTTTT-----GTTTGAATAAAGTGGAGTTTATGATACCGCTCTCT-----AAAATACCACCTTCGACTTGCCTGAAA
K GAACTGTTTCTTGAAGGTAAGCCACTTTAAGTTTGAACGATTTCTTAAATAACAACATATCCATATAGACCTTGCACCTTTTCTGCGTCTG
L TGATCCGTTT-----TTTTGATTTGCGTAATTTGTCACCAGTGAATTATG-----TCATACCTGCATCTTACTCTGCTGCG

A TTTATTAACCTCGCAATGTAGCAGCTGTCTGATAATCTAAGTACTCTTGTGAAATTTGCTTTTGGTCTGTTTTGGTATAGCGCTTCTAGCAG
B CTTAATACTCCTCGCAAAGTTGAATTTGACTTACCAAGTTACTAATTTTACTTGGATTTTTTACTAGGCTTTTCTGTCAGAAAAGTTTTATTGA
C TPCAATAATATCTCAACAATGCAATTTGTTTCGAAAACTTAATACTTTCAATTTGAACACTCCTT---GCTGTTATGACACTAAGGTTGCGTTC
D CCAATTAGCTCGCGACAGAGTAAAAGTTGCTGGACAAATTTATTACTTTTATTGAAAACC-----TGTTGCTG-----TTATATGGA
E TTTATTAACCTTGGCAATGTAGTAGCTGCTTGTATAATTTAAGTACTTTGTTTGAATATTTCTTCTTCCGCTATTGGTGTGCGGTTACAACAA
F TTTATTAACCTTGTACAATATAGTAGCTGCTTGTATAATTTAAGTACTTTGTTTGAATATTTCTTTCTTTGTTATTGGTGTGCGGTTACAACAA
G CTAATTAGCTCGCGGCAAAATAAAAGTTGCTTGTACAAGTTTATTACTTTTATTGAAAACCT---GTGGTCTGTTATTGGCAATTCGGCTGTATTGA
H TTTATTAACCTTGTACAATGTAGTAGCTGCTTGTATAATTTAAGTACTTTGTTTGAATATTTCTTTTCCGCTATTGGTGTGCGGTTACAACAA
I CTAATAATTTCCGACAAACGAAACACTTGTCTGACAGGTTCAATAATTTTGTGGTAACTTCTTT---CTGTTAACG---TTATGTTTCACTGC
J TATTGTAATTTGTGACATAGGATTTGTTGCTTGTGTAATTTGAAAAGTTGTAATGAACTGCGGTGCTATTTTTAGAGAATGAGCAGAAAATATCC
K TTTAACACTCATGGCAAAGTTGTAATTTGACGTGACAAGTTCAATAACTTTAATTTGACTTTCGGTACTTCAATATTTGGAAGAAAAGTTTCACTGA
L CTTATTAGTTTGGCAAAAGTTGAATCCGTTGGATAAGTTTAACTGCTGCTGTAATTTCTTCCAGATGCTGGAGGAATAGTTTCATTAAGT

[Supplementary Figure 3 (2/5)]

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A CATCGTGAATAAGAGGCACGCAACT---AAAATAA---CGTAACATAATTGAACC---ATTTCAT-----tttgcatttgcaggagtgtgt
B CTGCGTTGTTTTGTGATAAAGCAGCTTTTAAAACAAAAGACAAAATAATATAAGCCAATAATTCAT-----ctttagcatttccacttctagta
C TTTTGTGGTTTAGAGTTAGGGCAACTTGGGATGCAAAAACAGCATAATGTAAGTCC---ATTTCAT---ttttgttaactatttacagtacttgtt
D CACCTTGGTTCAAGGGTATAGCCATATATAAAGAAAATACCACCTGAAAGTAAATCC---ATTTCATttttctatccctttatccacagttattgtt
E GACCGTTAATAACAGGCACGACAACAAAAACAACATTT-----TACAATTAACATTTTCAT-----tttgcgttggcaggactgggt
F GACTGTTAATAATATGCACGACAACAAAAAACAACCAACATTTCAAATTAAGCATTTCAT-----tttgcatttggcaggactgggt
G CTGCATGATTTATGGGAATTACCACATTTAAAGAAA---TACCACCTGAAAGCAAATCAATCTCATttttttatcctattatccacagttattgat
H GACTGTTAATAGTAGGCACGACAACAAAAAACAACCAACATTTTACAATTAAGCATTTCAT-----tttgcgttggcaggactgggt
I TCGTGGATTCCGAGATGCTACGATTTTAAAACAATAACCAAGTATAAAGCAAAGCT---GTTTCAT-----ctttgattaccagtgtttgtt
J TTTGCAAATAAACAGAACAAACACAACATTAC-----TCTCAT-----acttgcctttttatttttagactggct
K CGCCGTTATTCAATGTTATTGCAGTTTTTAAAATAAG---AGCCAACGATGTACAAGCAAATACAT--acttgcctttttatttttagactggct
L --TATAACTCCGGATATAGCAAATTTCTCCCGTTAAA---GTTAACATAACGTAAACC-CATTTCAT-----

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v-Themis-B ←

→ *s-Themis-B* (2nd exon)

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A ccaaaagcatatgcaacctcgatagAGA-----TTCGTGCTCCCTTGAGTGAAGAATGCAATCAA--ACTG
B ctggtttgcaactggaagcgccgtagAAA-----TCCATGCTCCAGTAGCCGAAGAATGCAACCAA--GCAA
C cgtgatacctctgcaagcgtaaaagAAA-----TTCGTAGTCCCGTACACGAAGAATGCAATCAA--ACAA
D tctgatacctcaacgaacgtttagAAG-----TTTACGCTCCAGTGGTTGAAGAATGCAACCAA--ACAA
E tctaatacatatgcgaaactcggtagAGA-----TTCGTGCTCCTTTAACTGAAGAATGCAACCAA--CCTA
F tctaatacctatgctggcctcggtagAGA-----TTCGTGCGCCTTTGACTGAAGAATGCAACCAA--CCTA
G tctgatacctcaacgaacgtttagAAG-----TTCATGCTCCAGTGGCTAAAGAATGCAACCAA--GCCA
H tctaatacatatgcgaaactcggtagAGA-----TTCGTGCTCCTTTAAGCAAAGAATGCAACCAA--CCTA
I ctcgatgtcactgcaaatgtggttagAGA-----TTCGTGCTCCCGTACAGAAAAATGCAACCAAGCCATG
J -----attttcccttcaagTTCTGACTGGTAATGTTATCGCAATCTCTTTGGAAAGCACTGGCACGCTGCAAAATCCGCTTGGTGAAAAGT
K tgaatagtagtactgcaagcgtagtagAAA-----TAAATGCTCCGGTGGACGAAGAATGCAACCGA--GCGA
L -----tttatcgtcttggtttcaagTTATCGCTTCTGACAC---TTTACTCGGGTAATTCATGCTCTGTGCACGAAGAATGCAAAATTA--ACAA

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A ACGTTTCAAACAGGTTGCTCTCGAATTCGGGTT-AAAAAGTACACAGATGCTGGT-----TTTACG-TTTACAAAATACGAGgt-----
B ACGTTCAAACCTCCATTTCTCTCGAATTCGATTT-AAATGCGAAAGAGATGCTTGT-----TCTTCG-TTTGGAAAACGCAAGgt-----
C AATACCCAAGTGAGATTACACACGAGTTTGATTT-AAAGGAAACGCAAGTTCTGGT-----TTTTCGGTTTAAAGAAC-CAAGgt-----
D ATATTCAAACGAGATTTCTCTCGAATTTGATTT-AAAAGATAAACAGATGTTGGT-----CATTCA-TTTTCAAATGCCAGgt-----
E ATGTTTCAATCCAGATTGATCTCCAATTTGATTT-AACGCAAGCCCAGGTTTGGT-----TCTTCG-TTTACAAAATACAAGgt-----
F ACGTTTCAAACGAGATTGATCTCCAATTTGATTT-GACGCAATCACAGATGTTGGT-----TCTTCG-TTTACAAAATACAAGgtactaaat
G ATATTCAAACGAGATTTCTCTCGAATTTGATTT-AAAAGAAAACACAGATGCTGGT-----CATTCA-TTTTCAAATGCCAGgt-----
H ATGTTTCAATCCAGATTGATCTCCAATTTGATTT-AACGCATTCACAGGTGTTGAT-----TCTTCG-TTTACAAAAACAAGgt-----
I AACGTTTCAAACGAGATTACACTTGCATTTAATTTAAAGGAAAACCCAGATCTGGT-----TCTTCG-TATTCAAGAAGACAGgt-----
J GCAACAAAAGCAGTGGTTCGCCACAATTTGATTTGAAATTCAGCTTGTGAGTAGTCAAATCCTTATCATCCATTTCCAGAACAAAAGgt-----
K ACGTTTCTAATCAAATTTCTCTCGAATTTAATTT-AAAGGAAAACAGATGCTTGT-----TCTTCG-TTGGAAAATGGAAGgt-----
L ATATACCGAACGAGATTTCTCTAGAATTTGATTT-AAAAGAAAACGAGATGTTGAT-----CCTGCA-GTTAAAAAATACAAGgt-----

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A -----
B -----
C -----
D -----
E -----
F gcacattattatatatagtaaagtgcgggaagatgggacaccttagctcgaatatccaacatcctcatcgggtttaaacatttaacaacggctct
G -----
H -----
I -----
J -----
K -----
L -----

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A -----
B -----
C -----
D -----
E -----
F atggaagtcgtaggttaaggtttcacaattcttcgattgtttattttgcttactactaaattgaaaataaaaatagaatgaaagggtgtccaatct
G -----
H -----
I -----
J -----
K -----
L -----

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A -----
B -----
C -----
D -----
E -----
F tttcctaccctactctataacaataagaacagatttaatgaacttattttactatatttaagaaagtcgtatagtatataaaactggtatcgcac
G -----
H -----
I -----
J -----
K -----
L -----

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[Supplementary Figure 3 (4/5)]

A -----
B -----
C -----
D acacaattagtcataataatctaaacctccttaacgtattttaacaatatacaatggctctatgggagtcacaaggatgcatctttctaaatctttg
E -----
F -----
G -----
H -----
I -----
J -----
K -----
L -----

A -----atgatttttattgcttatttttctattattggaagcaaaactaaaaactgttttcttctgctta
B -----gattactatatttttataaggtattttaaaatggtaacgtaaattccgactaaatttgataataattcc
C -----agtataatgatgaaactttcaggcgctgttaaacaaataggtatttaactacatcgtatttttccctatgct
D aatgtttactacgaaatggaacaagaaaatataaaatgccccacctttccccacactcaccatactcgttataaaacattattttcc
E -----agtatttttattgcttaactaaatagctttttatggttttcttaatagtaaaacataatggtttgtttacc
F -----agtatttttattgcttaactaggtttttatcgttttctgaaatgtaaaacacaaactgtttgttttcc
G -----agttatggtcgtgtagttgtttatccttataagataaactcctttttttat
H -----agtatttttattgcttaactaaatgctttttatcgttttcttaaatgtaaaacataatggtttgttttcc
I -----aactacttttatacagcttgcatccaagctataataactcttactatttacttttttggcttaaatccgcttccattca
J -----aagtgattacttagaagagaactattttctgtcaaaactaatattgtatacaaca
K -----gatgttataatatttaaatgcatgataatatttattgtttacc
L -----ttgatttcattattttatgtttatataaccaaactgaaactgtaattttgtctgct

A tagAATGGTGGTGGTGTGACCCTTACAAAAGTTTCTGTCGGTGAAAACGTTACAGTTTATAGTAAAATCTAAAACCTCAAACACTACGTTGAAC
B cagAAATGATACGTGATTTTACCAGTGCATTCATTACTGCCGAGAAAACGTAACAATTTATTGCTGAAATCGATAACGTTAAGCAATACAGTGAAT
C cagAAATGTCATTTGATATTACCACCACATCCAAATCCCGTTGATAAGAAATGTCACAGTTTACGCCAAAATAACGAAACAACAGGAAATACCTCAGATGC
D tagAAACGATACTGATTTTACCAACACAACCGCTCTCCACTCGGGAAAATGTAACATTTTATGCTGGAATTAACGACATCGGCCAATACCGTGAATC
E tagAAGTGGCAGTGTATTTACCAAAAACAAATGGTCTGTCGGGAGAAAACGTCACAATTTACAGCAAAAATCTCAACCTCCGACAATATTTGGAGTT
F tagAAGTGGCAGTGGTCTACCAAAAACAAATGGTCTGTCGGGAGAAAACGTCACAATTTACAGCAAAAATCTCAACCTCCGACAATATTTGGAGTT
G tagAAACGATGCTGTGTTTTACCAACACAACCGCTCTCTCGGAAAATGTAACATTTTATGCTGGATACACTGACATCAGCCAAATACGCGCAAT
H tagAAGTGGCAGTGGTCTACCAAAAACAAATGGTCTGTCGGGAGAAAACGTCACAATTTACAGCAAAAATCTCAACCTCCGACAATATTTGGAGTT
I cagAAATGGCGTTGATTTGATACCATTTACACCAGATTTTACAGCAAAAATGTTTACAATCTACGCTGAAATTTATTAATGCTAAAACAAATACACAGAGTC
J tagAGATGGCAGCTGTATCTACC-----GAAGCAACCGATTCCTATTGGCAAGAGAGTTCCTGATGTAAGTGGCAAGAACGCTCAATTTTAAACAAAATCAAAAATTTGAAAAGTTAG
K cagAAATGGCTCTCACTTTACCAATCAAGTCGGTTTCTACTAGAGAAAATATAACAATCTATGCTGATATTAGTAACTACAGACAATACACTGAAAA
L cagAAGTATATGTAATTTACCAGTGCACCGGTGCCAGTTGGGAAAACATCACAAATTCACGCTGGAATTTACAATATACGCGCAATCCATTCAATC

A ATTTCTGTGCACAATCACCTGTTCTATTTCTTTGAAAATAAATCTCCCATGTCAATGAAATTCACCGAACAGACCAATTTTACCATCAGTTTTTTG
B GTTCTTGTGCACCATCAGCTGTTCTGATTTCCCGTCGGAGATAGTTCCTGTAACATTCGAAATCCACCAAAA---ACAAAATTTCCCGAGCGTGGTTGTA
C TTTCTTTGTTCCATAGCTGTTCTATTTCTTTGCGGACGATTTCAATGTTATGATGATATCTACGAAA---ATGCATTTTCTCATCGTTTTCTG
D ATTTGCCATAAACCTCGAATGTTCTTTAACCTTTGGTGATAATTTCTCTAAAACAGTTGGAATTCACGACA---GTAANAATGTATCATATATTTGGA
E TTTTATGTTACTGCCAGTGTGTTCTATTACTGTCCGAAAGAGTTCCTCCAGTATCAGTTGAAACCCAAACGAAAGGCCCCATTTTACCATCAGTTTTTCA
F TTTTATGTTACTGTCAGCTGTTCTATTTCTTTGCGGAAAGAGTTCCTCCAGTATCAGTTGAAACCTTTACGAAAGGCCCCATTTTACCATCAGTTTTTCA
G ATTTGTGCATAACTCTCAGCTGTTTTTAACTTTTCCGCGAAGCTCTGCGGTGCTTTTTGGAATCCACCGTA---GGGTACTTTATCATATATTTGGA
H TTTTATGTTACTGTCAGCTGTTCTATTTCTTTGCGGAAAGAGTTCCTCCAGTATCAGTTGAAACCTTTACGAAAGGCCCCATTTTACCATCAGTTTTTCA
I ATTTCTTTGTCACCATCAGCTGTTCTATTTCTTTGCGGAAAGAGTTCCTCCAGTATCAGTTGAAACCTTTACGAAAGGCCCCATTTTACCATCAGTTTTTCA
J TTTAGAACGGGCCGAATGTACAATTTCTGACGAAAGGATTACAACCTCAATTCAGTTTCCAGTATCAGTGAACCTTA---GACTATTTTATCATATGCTTTTTT
K ATTTCTTACAACCATTAGCTGTTCTATTTGCGTTTGGTAAACGTTGGAATTCGTCAAA---ATGTAATTTATCATATGATTTTTTCT
L TTTTGGTGTACTTTTAACTGTTTTATTTCTTTTGGAAAAGGTTCTCCTTTATCGGTTGACACTCAGCAAAA---ACACATTTTACCATCGTTTTTCTG

A ATGCCTGGAGCACATGAAGTGTGGTTCATGTAATATACCTGATGCAGAACTATTAGAACATACGGTGTGCTGAAATGTGGAAGGCTTTTTAAACCA
B CTGCCCGGTACACACAGTCATATGTTCTCTTGCTTTATTCCAAAGCGGAAAAGTCAATTACACAACGAAAATCTTTACAAAGTGGAAAGTTGTTGTTCA
C TTACCGGGTCTTATGAAATATTTATTTTGTGCTCATACCAAGCGGCATTGACATTTACACAACAAGAACCTTTTACAAAGTGAAGTTGTTGACCC
D CTGCCTGGTAGCCATTATATACATTTTCTTGCCATATGCCGAGCGGAAGCGCGGTTACGGAACGAAAATTAATACAAAGCTGAAAGTTGGTTAAACG
E TTCCCAAAAACCTATGCTGTATCGTTTTTCAATGTGAAAATACCAGTGGGAGATTAATAGAGCATATGGTGCCGTAACAGTGGAAAGTAAATTTGAACA
F TTCCCAAAAAGCAGGATGTATGGTTTTTCAATGTGAAAATACCAGTGGGAGATCTTACAGCATACGGTGCCTGAAAGTGAAGTAAATTTAAACA
G CTGCCTGGTAGCCATTACATTTACATTTCTTGCCAATACAAAGCGGAAAAGTCAATTACAGAACGAAAATCTGTTACAAAGTGAAGTGGTTGAAAG
H TTCCCAAAAAGCAGGATGTATGGTTTTTCAATGTGAAAATACCAGTGGGAGATTAATAGAGCATACGGTGCCTGAAAGTGAAGTAAATTTAAACA
I CTACCTGGTACTCAGCTTATGTTATTTCTGTCGAATATTTCAAGCGGAAAAGACATTTATACACGAAAATTTTGCAAACGAAAGTTGCTGACCG
J CGGCCAACCGAGCATGCCATTTCAATATACGTGTAATGTTTTTAAACAATGTACGCATGTACACCAAAAAGACTCTGCATGTGAAAGGCTACTTGGAAA
K GTGCCCTGGTAAACAGGAAATATCCTTTTCTCTGTTATGTACCAAGCTCAAAAAGAAATAGAGAACAAAAGATATACAAAGTGGAAAGTGTGTTGAATG
L CTTCCTCGTGAGCACAAAATATCATTTAATTTGCCGTATACCAAGCGGAGAGTAAATTAAGCAAGAAAATGATACAAAGTGAAGTGGTTAAACG

A CCAACAAATTACAAAATAAGAAACACAG---ACAAACCGGTACAGTTTCTAACGAAAACAACTTACTTTTTCTGTCATGTGTATCAATACCCCATAAA
B GTAACAAATTAATAAATAAACATCAC---CAGCAAAATTCACATTTCTAACTGAGGCAGAAAATACTTTTCAGTACGTGTACCACATATCCCATATC
C ATGACAACCTTAAAAATAAAACCGAGGT---CGGCACCAGTACAGTACTTTACGGATGCAAGTATACCTATCAACACGTTTACCAATATCCTCTACA
D AGGACAGTCTAAAAATGATACCTGCAC---CGACAACAGTCAAGTTTTGACAGACATTAATGTTTCTGTACGTTACCAATATCCAAATATCAATTAAT
E TAGACAACCTTGAANAATTAATTCAGTGT---TAAAGCGGCAAGATTTCTAACAGATACTGCTGCTTTTTTCAGTTTATATATCAATATCCCATACA
F AAGACAATTTGAAAATTAATTCAGAT---CACAACCGACAAGTTTCTAACAGATATTAATTTGCTTTTTTCAGTACATTTATCAGTATCCTATACA
G AGGACAGTCTGAAAATGATACCTGCAC---CGACAAAAAGTCAAGTTTTGACAGACAATACTTAATGTTTCTGTATGCTTACCAATATCCAAATATCAAT
H TAGACAATTTGAAAATTAATTCAGAT---CACAACCGACAAGTTTCTAACGGATACTACTGCTTTTTTCAGTACATTTATCAGTATCCTATACA
I ATGACATTTTGAAGATTAACCTGGTTAAAACCTACACCAGTAAGATTTTCTGTTAGGTCAGTCTTACATTTCAACACGTTGATCAATATCCTATAAAA
J AAGAAAACCTGCAAGTACAAAATGACC---CCTTTGTTATACGATACCCGAAAGAGCAAGTACTGCAAAATTAACAATATTTGACAGATCCAAATCA
K ATAATACCTGAAAATAAATCGGCAT---CGGAACCGGTACAGTTTCTAACAGATACCAATCTGCTTCTCCAGCATGCTGACCAATATCCTGCAAT
L TCGACAACCTGAATATAAAACCTGAAT---CATTACCAGTACAGTTTCAACAACCTGCCGATTTAGTCTTCCATCACACCTATCAATATCCCATCA

[Supplementary Figure 3 (5/5)]

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A      GTATTGTTTTATTTTGAAAAATTATCAAAATTGTGACCAAACTGAGGATTTTTTACGAAACAAGTACACCAATAGAGAAGTTGTTGCAGCCAAGTTC
B      GTACTGCTTTTATTTTAAATGATTATCAAAGTTGTGACCGAACTAAAAGAAATTTTACAAAACCAATATAACGATAAAAGAAGTTGCTGAGATCAAGTTT
C      GTACTGCTTTCGTTTTTAAATAGATTATCAAAAATGTGACCAAAACAAAAGACTTTTTACAAAACAAGTACAACAATAAAGAATTTGGTTGAGGTTGAATTC
D      CTCTGCATCGTTTTTAAATGATAATCAAGTTTGTGACAGAACCGAATCCTTTTCAGTAGAAAAGTTTGAAGGAAAAGAAGTTGTTGTAATGCCATAT
E      ACATTGCTTTCGTTTTGAGAACATATCAAAGTTGTGACCGAATTTAAACTTTTTTCACAAAACCAATATGAAAAATAAGATCTTATTGAAGTTTCAGTTT
F      ACATTGCTTTCGTTTAAAGAACATATCAAAGTTGTGACCGAAGTACACTTTTTTCACAAAACCAATATGAAAAATAAGATCTTGTGATGTTTCAGTTT
G      GTACTGCATCATTTTTAAATGATTATCAAAGTTGTGGCAGAACTGAAACATTTTTAGTAGACAAGTTTGAAGGAAAAGAGCTTGTGAAATGCCATTT
H      ACATTGCTTTCGTTTTGAGAACATATCAAAGTTGTGACCGAAGTAAAAGTTTTCACAAAACCAATATGAAAAATAAGATCTTATTGATGTTTCAGTTT
I      GTATTGCTTTATTTATGAAGCAATATGAAAGTTGCGATCAAAGTTCGGAGTTTTTCAGAAAACAATAATAAAAAATAAGAAAGTTGTTGAAGTTTCAGTTT
J      ATACATTTTATTAATTTGGTACACATGTGATTACGCAGAACACGGGGTATTTGAATCTAGCAAATATGAAACTAAAAAAGTTGCTCCGTTTCAGTTT
K      ACTGTGCTTCCTTTTAAAAAGATTATAAAAGCTGTGACACAACATAAAGATTTCTTACCAACTAAATACGAAAAACAAGAGGTTGTTGAGGTTTCAGTTT
L      GFACTGCTAATTTTGCACACGTATAAAAAGTTGCGACCGAACTAAAATTTTTTCGAAAACAATAATGACAACAACGAGATAAATTGAGGTATCATT

A      AAACTAACAACAGAAATTCAAACTGAAACTGGGCCAGGAATACATTTTTGTACATTGTTTAAATGCAAAAACAATGTATCTGATGTAACATATAAGTCAA
B      AAACTAACAACAAGCATTCAACTCTACAATCGGACCGGATACATTCCTTCACACGTTTTATGCAAAAACAATGTGTCGCGATGTAATAATACAGGTTTC
C      CAACTAACACAACAACTTCAATCTAAAATTTGGACCCGGAATACACCCAATTACATTGTACTTTCAAATAATGTTTCTGTTGTAACAACTTCGTCAA
D      CCACCTAACAAAACAGTATACAATCTTCCACTGGCCCGGAAACACATCTCATTACAGTTTTCTTGCAGAAATAATGTGTGCGGAGTGAATACAATTTTA
E      CAACCTTACAACAGACATCCAGTCATCTATCGGACCCGGAATACATCCAGTCCACATTATTTATGCAAAAATAATATATCTGATGTAAGATATATGTC
F      CAACTTACAACAGATATTCAGTCATCAATCGGACCCGGAATACATCCAGTCCACATTGTTTATGCAAAAATAATATATCTGATGTAAGATATATGTC
G      AAACTAACAACAAGAAATATGCAATCTTCAACTGGCCCGGAAACACATCTCATTACAGTTTTCTTGCAAAATAATGTGTGCGGAGTGAATACAATTTTA
H      CAACCTTACAACAGATATTCAGTCATCAATCGGACCCGGAATACATCCAGTCCACATTGTTTATGCAAAAACAATAATATCTGATGTAAGATATATGTC
I      CAACTTGCACACAGACATTCAGTCATCAATCGGACCCGGAATACACCCAGTTACATTTATTTATGCAAAAATAATATATCTGATGTAAGATATATGTC
J      AAACGTGCTGCCAGATGCAACTGGAAGTTGGTCCCTGGAAGTTACGCAATTTGGTTTTATATTTACAAAACAATGTGTGCAACATTTGATTTTACATCAG
K      GAACCTAACACAACATCATTCAATCTACAATTTGACCTGGAATACATCCATTTGCATTTACTTCCAAAATAATGTTTCTGCAGTGCATATAGGTTCTG
L      GAAATTACAAGAGCATTCAATCTTCAATCCAACAGGAATGTGTACCGCAGTTCTCTTCTTGCAAAATAATGTATCTGCAGTCCGTTACAGTCTC

A      TAGTTTGGATTAACGAACAAGTTACAGGAGTTACGGTAACATGCAATAGCTTTTGTGGAAATACATCCAAATCAATTCACGATCAGCATCACACCTAA
B      TTTTTATCTTAACCAAGCTGGTTACAGGAATGAAAGTAATATGCCATCACTTTGTCGGAATACAACCAAAATTACTTTACAATCAACATTACGCTTGA
C      CAGTTTGGATTAACCAACAAGTAACAGGAATGAAAGTTACATGTAATTCATTTGTTGGAATTCACCCAAATTACTTTCACAATCAACATTACGCTTGA
D      CAGTTTGGATTAACCGACAATTACATCAATAAAAAGTGGCATGCGATTCCTTTGTTGGTATTCAACCAAAATTACTTTACAATCAACATCACACTTGA
E      CGGTATGGATTAACCAAGCAGTAAACAGGAATAAAAGTAACTTGTAAATCCCTTTGTCGGAATACAACCAAAATTACTTTGTAATTAACATCACACTTGA
F      CGGTCTGGATTAACAAGCAGGTAACCTGGAATAAAAGTACATGCGATCTCTTTGTCGGAATACAACCAAAATTACTTTACAATCAACATTACGCTTGA
G      CAGTTTGGATTAACCTACAATTACATCAATAAAAAGTAAATGTGATACTTTTGTTGGTATTCAACCAAAATTACTTTACAATCAACATCACACTTGA
H      CAGTCTGGATTAACAAGCAGGTAACCTGGAATAAAAGTACATGCGATCTCTTTGTCGGAATATATCCAAATCAGTTTACAATCAACATTACGCTTGA
I      CAGTCTGGATTAACAAGCAGGTAACCTGGAATAAAAGTACATGCGATCCCTTTGTCGGAATACAACCAAAATTACTTTGTAATTAACATCACACTTGA
J      AAATTTGGTTAAACCAACCGTTAGTTGAACTGAAAGCAACTGTCCACCATTTTGGTTGGAGTTTATCCACATGATGTTGTTGTAACATCACGCTCGC
K      TGATTTGGATTAACCAACAAGTAACTGGAATAAAAGTACATGCGATCCCTTTGTTGGAATACATCCAAATTACTTTACAATCAACATCACACTTAA
L      CAGTCTGGATTAACCGTCCACTTACAGATCTCAAAGTACATGATGCTCTTTGTCGGAATACATCCGAGTTACTTTACAATCAACATCACACTTGA

A      AAAAGGTTGTCCAGTCAACATTACAGTAGAAATCAATAAATATCCCAACAA
B      AGAAGGTTGTCCGGCCAACATTACAATAGAAATCAATAAATATCCCAACAA
C      AGAAGGTTGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
D      AGAAGGTTGTCCAGCCAACATTACAATAGAAATCAATAAATATCCCAACAA
E      AGAAGGATGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
F      AGAAGGATGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
G      AGAAGGTTGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
H      AGAAGGTTGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
I      AGAAGGTTGTCCAGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
J      AGAAGGTTGTCTGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA
K      AGAAGGTTGTCCAGCCAACATCACAGTAGAAATCAACAATAATATCCCAACAA
L      AGAAGGTTGTCCGGCCAACATTACAATAGAAATCAACAATAATATCCCAACAA

```

Supplementary Figure 3. An alignment of genomic fragments encoding *v-Themis-B* and a hyper-variable region of *s-Themis-B* that were amplified using PCR. Putative exons of *v-Themis-B* and *s-Themis-B* are shown in red and green capital letters, respectively. Introns are shown in black lower case. Only exons were aligned. PCR primers are indicated by underscores.


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A MKWFNVTLVL--VVVPIHDAARAAEKTAKSKICTVLRLSROLLHCNGLINHKNNMNG--LVSK--NPKINGLAIGRLYEDCNGILOSQKHVND
B MNYWLVIIISFVAKAALSONNAVANKIFLTERPSKIQKLVNLSQQLCEELISRVKIKAG--SFSK--K-RILMNSSAIQSLENTDCTNHRIGVSSD
C MKW-TYIILVFA SQVAITINHKRNAIIV-SIARSTOMKVISFSQQLCEELINEHHRVYIR--PILKND-FRLSLAAIQSLEYTDCKDIQFGKASN
D MKW-TYFOVFSIYMAIPINOGVH-----IATGFOIKVINLSROLLCEELIGRNEIKVS--TF-EVG-AKRNAITAAQNLNTDCTNIHLSGKYVSD
E MKW-FNCKVLEF---VVVPIINGLVVATPIAEERNIQTKVILKLSROLLHCCELINERKNMVR--LDPHDDATLKKLMAIKREYTDGSGILOSQGYVSD
F MKWLNIEVVLFFVHVHINSVVAATPIITKEPNIQTKVILKLSROLLHCCELINERKNMDRLELDAYR-DATLKKKMAIKREYTDGNDIISGLYVDSG
G MKW-LCFQVFSINVVVPIINHAHTAEPI-IITTSFOIKVINLSROLLCEELIRRNEYKVS--WFSK--D-SKRNAIPAAQNLNTDCTNIHLSGKYVSD
H MKWLNCKLVVFFVVPPTNSAVITPIAKRRNIQTKVILKLSROLLHCCELINERKNIDR--LHPH--DATLKKMSIKREYTDGSGILOSQGYVSD
I MKW-LCFVIVIKVASRNPA-S-SENIT-LIPRSYQKLVNLSRQFRCKLIGRKYN---YFPK--R-DTSLVKAIQNLNTCKEIVFSGHYVSD
J MKW-LVVFVIFIQRFSA-----HSLKIDTQIQQLSNLQKQLCQOYRKS---LI-PNK--FQNLNASSIKSLFTDGHVQRYKRRSE
K MYL-LCTSAALVKTAITINNGVSEIFPILEVPKQKLVNLSRQQLCEELINRRFKGKGG--PSK--NSFELFNSLAIQNEVADCKSIQSLKNGS
L MKW-VVVLVPLTGEFALSQSYTNETIIPASARNIQTAVLNLSRQQLCKLISDQIG-----KKG-SKLDSEFSPKNLNTDCTNIHLSGKYVSD

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A VYPIWLKVGYRFIHIYCDMGSGSHITNKTGWITFQ
B VYPIWLKVGYRFIHIYCDMEIGSRITNKTGWITFQ
C VYPIWLKVGYRFIDIYCDMBSGSHITNKTGWITFQ
D VYPIWLKVGYRFIDIYCDMBSGSVITNKTGWITFQ
E VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ
F VYPIWLKVGYRFIHIYCDMEIGSHITNKTGWITFQ
G VYPIWLKVGYRFIHIYCDMEIGSHITNKTGWITFQ
H VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ
I VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ
J VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ
K VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ
L VYPIWLKVGYRFIHIYCDMBSGSHITNKTGWITFQ

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Supplementary Figure 4. An alignment of an N-terminal portion of v-Themis-B

variants. Identical residues are highlighted in black, and similar residues are highlighted in gray.

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A -----IRAPLSEECNQT-DVSNQVALEFGLKSIQMLVLRLLQNSCS-FASIDVQSKSE-ITFFCNKIPKKNQFYLFGRCSVTISPTPKQ
B -----IHAPVAEECNQA-NASNSILEFDLNAKEMLVLRLENASCR-FVSINVOLEWE-TLLEFCNOMPDK-KOFFELFRQFSLISPPQPN
C -----IRSPVHEECNQT-KYPSEIHEFDLKEIQMLVFRFKEFSCK-HVSNINQDRRE-TAFECNLSSTT-KQYLYLIGRETVYISPHKIN
D -----VYAPVVEECNQT-NIPNEISLEFDLKDQMLVHFQNASCO-NVFINVQLTRG-TOELCKQAPK-K-TTSEFNDRETVLISPPQPN
E -----IRAPLSEECNQP-NVSDIQDLOFDLTOAQMLVLRLLQNSCS-YAFINARSKSK-TIFFCNOSPQMNQYLFGRCSVSISSVPQPN
F -----IRAPLSEECNQP-NVSNQIDLOFDLTOQMLVLRLLQNSCS-HASITARSKSK-TVFCGGSPLPLNQYLFRRQSVSISPPQPN
G -----VHAPVAEECNQA-DIPNEISLEFDLKKQMLVHFQNASCO-NVFINVQLKRE-TOELCNQAPPK-K-TPVSEFNDRETVLISPPQPN
H -----IRAPLSEECNQP-NVSDIQDLOFDLTHSQMLVLRLLQNSCS-YAFINGVSESE-TVFCGGSPLPLNQYLFGRCSVSISSVPQPN
I -----IRAPVQKKNQAMNVSNEIILAFNLKEIQMLVLRLLQNSCS-IYVILINQVSIKK-TLSICMQSSPK-KQHSYLGRETVLISPPQPN
J LTGNVIAISLESTGTLQIPLEGKCNKS-SGSDQFDLKESSISSEQLLHFQNKSCSEKLVSHVQSDST-TTFCPETPLDLKKEYAFNGETALWFDPEPKN
K -----INAPVDEECNRA-NVSNQIDLEFNLKEIQMLVLRLLQNSCS-VVSLVVRLEQETTVFCNNSPK-K-EQYLYLTHGTVSISPPQPN
L IA-----SDTFTTRVTHAPVHEECKLT-NIPNEISLEFDLKEIQMLVLRLLQNSCS-TVILINVOIKRE-TIFFCNQSPFQ-TQNNLVNQCENVLISPPQPN

A CKTM-VIARYVILELVVVVPLQKQVSVGENVTIYSKILNFRHYVELEFVTLTSCSISE-ENNSPMSIETRTDQFYHQFLPGAREVSVSCNIPDAESITRY
B CKTH-VIARYVILELILPMHSITAGENVTIIEADNVKQYSELEFVTLTSCSISV-GDSSPVTFIIEQ-NKFSQRVYLPGEVILFSCFIPSGKAITQR
C CKDD-VIARYVILELISLPPHPFWDKQVTHAKITNKRKYSFAFVSIKCSISE-GHDSNVMIIIEYENASHRFLFLPGSEYELFLCLIPSGIDITTO
D CKNI-VIASYVILELITLIPQPVSTRENTVIYAGITDQYAESEFANLECSLIE-GDNSPKVGTID-SKMYHIFGLPGSHYIIFSCFIPSGSVAITER
E CKSI-VIARYVILEVAVLPKQMVSVGENVTIYSKILNROYLEBEFVTFASCSILVGRSSPVSVETQRKAPFYHOFSEBKUYAVSFSCFIPGCEITRAY
F CKSI-VIARYVILEVAVLPKQMVSVGENVTIYSKILNFRHYSEFATVSCSISE-GRSSPVSVETFRKAPFYHOFLEFQKODWFSCFIPGCEITRAY
G GENP-VIASYVILEMMLLPPQPVSSRENTVIYAGYTDQYAELEFVTLSCSIE-GESSAVSEGTIR-RVLYHIFGLPGSHYIILSQIQGCKAITER
H CKSI-VIARYVILEVAVLPKQMVSVGENVTIYSKILNROYLEBEFVTFASCSILVGRSSPVSVETQRKAPFYHOFLEFQKODWFSCFIPGCEITRAY
I QDNV-VIARYVILEALVPLHQSVRKQVNTIYAEIINAKQYSEFVTLTSCSISE-GFDSNVEVALIQ-NKPYRFRFLFLGSHVIVFSCNIPSGKDIITR
J QDSEHAAFYVILEALVLPKQPVPLGKNVETINKTKNEKVS---LERAECILISEGLQSLPVSVEL-RLHHVFLRTEHAIQMLCNVLNNVRYTK
K CKPN-VIASYVILEALPLPKSVSTRENTVIYAGITDQYAESEFANLECSLIE-GNGLSESLGIRQ-NVLYHRRFIPGKQEIIFSCYIPSSKETREQ
L CKNV-EVIASVILEVYVNLVRFVFGKNITTHACIYNROSQSISGVTFNCFISF-GRSSPVSVETQO-NTFYHRRFLLEREKIFENCRIPSGEILQIE

A GVVVNESLITTNKLOIRN-TDKPVTFLETTLLFRHVYOYPIKYCFILNYONCDQTEDEFLRNKYTN-EVVAAKFKLTTEIQLEETGPGIHFVTLFMQNNV
B KSLQVESLIFSKOLKIIT-SPAKFTFLTAEITFCYVYHYPISEYCFILIDYQSCDRTRFLOLKYNDKEMAEIKFRLTTSIQSIIIGPGIHSFTLFMQNNV
C ELLQVESLITHDNLKIKP-RAPVRLITDASILVQHVYOYPIQYCFVILIDYQKCDQTKDFLQNKYNNKELVEVEEOLTNNIOSKIGPGIHPVTLFMQNNV
D KLLQVESMLNEDSLKIP-APPTVFLTDIILFLYVYOYPIIYCFILNDNQVCDRTEFSFVEKERKEVVPVPLTNSIQSSTGPGIHLITFLFMQNNV
E GAVVNESLNDNLKITS-VLKAAREFLTDALLFOHYQYPIQHCFVILTYQSCDRTRFSONKYENKILLVVOFOLTDDIQSSIGPGIHPVTLFMQNNV
F GAVVNESLNDNLKITS-RSQETRFLETDIILFLYVYOYPIQHCFVILTYQSCDRTRFSONKYENKILLVVOFOLTDDIQSSIGPGIHPVTLFMQNNV
G KLLQVESMLNEDSLKIP-APTKVRFLETDIILFLYVYOYPIIYCFILNDYQSCDRTEFLVDEKERKELVEEERKLTKNQSSSTGPGIHLITFLFMQNNV
H GAVVNESLNDNLKITS-RSQETRFLETDIILFLYVYOYPIQHCFVILTYQSCDRTRFSONKYENKILLVVOFOLTDDIQSSIGPGIHPVTLFMQNNV
I KLLQVESLITDILKIKLVKTPVRFVSRSSLTFQHVYOYPIKYCFILMVOYESCQTPSFSFNKYENKELVEVFOFOLADIQSSIGPGIHPVTLFMQNNV
J RTHVESYLEKELQIQI-DPFVRLPEQVLOIKHYQHPYOYILLGTHVITQCTGVFESSKYETKELVSVQFRLQOLEGPGSYAVLVYVQNNV
K KSLQVESVLDNNTLTKS-ASEPVRFLETDIILFOHYQYPIIYCFILIDYQSCDRTRKDFLPSKYENKELVEVFOFOLTDDIQSIIIGPGIHPVTLFMQNNV
L KMLQVESLITVLENLHKP-EELVPRFQITADLVEHHTYOYPIQYCFILQTYRSCDRTRKFSQNKYNNKELVEVSEETKSIHSSIIEGCTAVLFLFMQNNV

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Supplementary Figure 5. An alignment of a hyper-variable region of s-Themis-B

variants. Identical residues are highlighted in black, and similar residues are highlighted in gray.

A

CTGCAGGCGGCCGCACTAGTGATT

TGGTGTGAAAACGGGTGAAACGAACTGGTGGAGGTGCTAAAGTGACCGTAAGTGTCTGAATTTGGTGCATACTGGG

AATCCCGCGGCCATGGCCGCGGGATT

TGATGAATGTAAATTGGTTCAAGTCAAACATCTTTGAAGAAAGCTGGAGAGAAATACAGAGTCACTGTGTTGCAAATAACCAAAATCTAATGATAGCGCTG

TATCCAAGTGTGAAACCGTTCCTCA

AATCACTAGTGCGGCCGCTGCAGGTCGAG

B

CTGCAGGCGGCCGCACTAGTGATT

TGGTGTGAAAACGGGTGAAACGAACTGGTGGAGGTGCTAAAGTGACCGTAAGTGTCTGAATTTGGTGCATACTGGG

AATCCCGCGGCCATGGCCGCGGGATT

TTCAACACCACCACACTCAACAGCAATCTGCCGTTGAAGTTAGTTCGCCATCCAACACCCCAACCTCAGAGAACATGGCATTGAACACG

AATCACTAGTGCGGCCGCTGCAGGTCGAG

Supplementary Figure 6. Nucleotide sequences of calibrators used for measuring copy numbers of the *s-Themis-B* locus. Each of the fragments shown in (A) and (B) includes portions of *Macho-1* (blue)/*s-Themis-B* (green) and *Macho-1* (blue)/*FoxA-a* (red), respectively. Linker sequences are shown in black letters. Primer sequences are underlined and probe sequences are enclosed with black lines.

Supplementary Table 1. Genotypes of a genomic region on Chromosome 7 of the F4 to F9 and F11 generations

Chr	Position	Reference	Genotypes *							
			F4	F5	F6	F7	F8	F9	F11A	F11B
7	459721	G	G/A	G/A	G/A	G/A	G/A	G/A	G/A	G/A
7	459725	C	A	A	A	A	A	A	A	A
7	459782	A	A/-	A/-	A/-	A/-	A/-	A/-	A/-	A/-
7	459813	T	T/C	T/C	T/C	T/C	T/C	T/C	T/C	T/C
7	459823	G	C	C	C	C	C	C	C	C
7	460003	T	T/G	T/G	T/G	T/G	T/G	T/G	T/G	T/G
7	460006	T	T/A	T/A	T/A	T/A	T/A	T/A	T/A	T/A
7	460073	C	C/T	C/T	C/T	C/T	C/T	C/T	C/T	C/T
7	460085	G	G/A	G/A	G/A	G/A	G/A	G/A	G/A	G/A
7	460144	C	C/T	C/T	C/T	C/T	C/T	C/T	C/T	C/T
7	460188	T	T/-	T/-	T/-	T/-	T/-	T/-	T/-	T/-
7	460189	G	G/A	G/A	G/A	G/A	G/A	G/A	G/A	G/A
7	460200	C	C/A	C/A	C/A	C/A	C/A	C/A	C/A	C/A
7	460246	C	T	T	T	T	T	T	T	T
7	460268	C	C/A	C/A	C/A	C/A	C/A	C/A	C/A	C/A
7	460270	T	T/A	T/A	T/A	T/A	T/A	T/A	T/A	T/A
7	460359	C	C/A	C/A	C/A	C/A	C/A	C/A	C/A	C/A
7	460401	C	C/T	C/T	C/T	C/T	C/T	C/T	C/T	C/T
7	460466	T	T/TA	T/TA	T/TA	T/TA	T/TA	T/TA	T/TA	T/TA
7	460467	G	A/T	A/T	A/T	A/T	A/T	A/T	A/T	A/T
7	460469	A	A/C	A/C	A/C	A/C	A/C	A/C	A/C	A/C
7	460482	A	G	G	G	G	G	G	G	G
7	460503	T	T/A	T/A	T/A	T/A	T/A	T/A	T/A	T/A

* Nucleotides identical and not identical to the reference are represented by black and red letters.

Nucleotides now shown in this table were the same as those in the reference genome.