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1 **Decomposition behaviors of various crystalline**
2 **celluloses as treated by semi-flow hot-compressed**
3 **water**

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32 **Abstract**

33 Various types of crystalline celluloses consisted of group I (Cell I, III_I, IV_I) and group II (Cell II, III_{II}, IV_{II})
34 prepared from cotton linter were adjusted for their degree of polymerization (DP) as starting materials. These
35 celluloses were, then, treated by semi-flow hot-compressed water (HCW) at 230-270°C/10MPa/2-15min to
36 study their decomposition behaviors. The treatments performed resulted in the residues of celluloses and water-
37 soluble (WS) portions. Consequently, the crystallinity of the residues was found to remain the same, but the DP
38 was reduced as temperature increased. Additionally, the X-ray diffractometry (XRD) and Fourier transform-
39 infrared (FT-IR) analyses demonstrated that the crystallographic changes were occurred for residues of Cell III_I,
40 IV_I and III_{II}. Despite of these changes, the overall results of the residues showed that group I has higher
41 resistance to be decomposed than group II. As for the WS portions, the yields of the hydrolyzed and degraded
42 products were more in group II as compared with group I, indicating that group II are lesser in resistance for
43 their decomposition by HCW treatment. Both results on the residues and WS portions are in agreement with
44 each other, concluding that the degree of difficulty for decomposition was higher in group I compared with
45 group II. Therefore, the decomposition behaviors of the celluloses are due to the differences in crystalline forms.

46

47 **Keywords** *Cellulose • Cotton linter • Crystalline structure • Hydrolysis • Semi-flow hot-*
48 *compressed water*

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62 Introduction

63 Cellulose together with hemicelluloses is often encrusted with lignin in a complex composite known as
64 lignocellulose. Such lignocellulose is not easy to separate and dissolve in almost any solvents, making them
65 resistant to enzymatic hydrolysis (O'Sullivan 1997). Cellulose is a crystalline homopolysaccharide composed of
66 D-glucopyranose linked together by β -1,4-glucosidic bonds. The β -1,4 linkages in cellulose form linear chains
67 that are highly stable and resistant to chemical attack because of the high degree of intra- and inter-molecular
68 hydrogen bondings (Nishiyama et al. 2002).

69 There are six known polymorphs of celluloses (I, II, III_I, III_{II}, IV_I, and IV_{II}) identified by their
70 characteristic X-ray diffraction (XRD) patterns as well as ¹³C nuclear magnetic resonance (NMR) spectra. This
71 crystalline structural arrangement of one allomorph can be converted to the other by treating it with chemicals
72 through various processes such as mercerization, hydrothermal processes etc (Isogai 1994).

73 Cellulose I, known as native cellulose, is abundant in nature and can be further classified into two
74 polymorphs, celluloses I _{α} and I _{β} (Atalla et al. 1984; Sugiyama et al. 1991), whose detailed structures have been
75 established through synchrotron X-ray and neutron fiber diffraction studies (Nishiyama et al. 2002; 2003).
76 Cellulose I _{α} is metastable and converted irreversibly into cellulose I _{β} by hydrothermal treatment, without losing
77 its crystallinity (Yamamoto et al. 1993; Wada et al. 2004).

78 Cellulose II can be prepared by two distinct routes: mercerization by alkali treatment and regeneration
79 through solubilization and subsequent recrystallization. It is thermodynamically more stable in structure with an
80 anti-parallel arrangement of the strands and some inter-sheet hydrogen bondings. On the other hand, celluloses
81 III_I and III_{II} can be prepared from celluloses I and II, respectively, by treatment with liquid ammonia or some
82 amines. The swelling of cellulose in liquid ammonia or some amines is a simple and classical way to increase
83 the accessibility of crystalline cellulose (Da Silva Perez et al. 2003), while celluloses IV_I and IV_{II} can be
84 obtained by heating celluloses III_I and III_{II}, respectively, at 260°C in glycerol (O'Sullivan 1997; Zugenmaier
85 2008).

86 Acid/alkali treatment and acid/alkali pretreatment followed by enzymatic hydrolysis are conventional
87 methods to produce glucose from cellulose (Camacho et al. 1996; Karr et al. 2000). The obtained glucose can be
88 used to synthesize various specialty chemicals or fermented to produce bioethanol. However, these treatments
89 are associated with serious economic and environmental constraints due to the heavy use of chemicals (Hsu
90 1996). Furthermore, cellulose utilization is still seen to be restricted by its resistance against hydrolysis due to
91 the encrusted structure with lignin (Grous et al. 1986; Mes-Hartree et al. 1988).

92 For the past three decades, hot-compressed water (HCW) treatments have attracted much attention
93 because of their suitability as a non-toxic, environmentally benign, inexpensive medium for lignocellulose
94 hydrolysis without any catalysts (Bobleter et al. 1976; Bobleter et al. 1983; Yu et al. 2010). The HCW treatment
95 system can be categorized into batch-type or flow-type arrangements. Due to significant decomposition
96 reactions occurred in batch-type treatment, flow-type HCW treatment was preferred to hydrolyze and fractionate
97 lignocellulose (Ehara et al. 2002; Lu et al. 2009; Phaiboonsilpa et al. 2010; 2011).

98 About 4-22% of cellulose decomposed in HCW (200-230°C/34.5MPa/15min) by using flow-type
99 treatment (Mok et al. 1992). Another configuration of HCW system is a semi-flow HCW treatment. It resulted
100 in the better digestible cellulose (Xiao et al. 2011), greater removal of lignin (Tirtowidjojo et al. 1988), higher

101 sugar recovery from hemicelluloses (Bobleter et al. 1983) and less inhibitors in the hydrolyzed liquid as
102 compared with the conventional batch and co-current flow systems (Liu et al. 2003). Recently, the two-step
103 semi-flow HCW treatments were used to study the chemical conversion of lignocellulose biomass (Lu et al.
104 2009; Phaiboonsilpa et al. 2010; 2011) and it revealed that 230°C/10MPa/15min and 270°C/10MPa/15min were
105 optimum conditions for decomposition of hemicelluloses and cellulose, respectively.

106 In this present study, we aimed to evaluate quantitatively decomposition behaviors of various
107 crystalline celluloses as treated by semi-flow HCW. The obtained residues of celluloses and water-soluble (WS)
108 portions after their treatments were, therefore, investigated in order to understand how different crystalline
109 forms of celluloses are decomposed.

110 **Materials and Methods**

111 **Preparation of various types of crystalline cellulose samples**

112 Various types of crystalline celluloses were prepared from cotton linter (Buckeye 1AY-500). Cotton
113 linter in its native form is cellulose I (Cell I). Mercerized cellulose with crystalline form of cellulose II (Cell II)
114 was prepared from Cell I by soaking it into 20.0% of aqueous NaOH solution for 24h at ambient temperature,
115 followed by washing thoroughly with water and freeze-drying (Isogai et al. 1998). Samples with the crystalline
116 forms of celluloses III_I (Cell III_I) and III_{II} (Cell III_{II}) were prepared from Cell I and Cell II, respectively, by
117 soaking them in 100% ethylenediamine (EDA) for 24h at ambient temperature, washed with dried methanol and
118 kept under vacuum.

119 The prepared Cell III_I and Cell III_{II} were further used for the preparation of celluloses IV_I (Cell IV_I) and
120 IV_{II} (Cell IV_{II}). They were firstly soaked in glycerol for 3 days at ambient temperature and then heated in a
121 reaction vessel at 260°C/0.6MPa for 30 min (Isogai et al. 1989). After cooling down to ambient temperature, the
122 product was washed with water and acetone successively and dried in vacuum. To simplify, the cotton linter
123 (Cell I) was converted into group I (Cell I, III_I and IV_I) and group II (Cell II, III_{II} and IV_{II}) celluloses. The
124 chemical compositions of these celluloses were also analyzed and all the cellulose samples were found to
125 contain similar components of about 99.9wt% glucose and 0.1wt% xylose.

126 To evaluate decomposition behaviors for various types of crystalline celluloses by semi-flow HCW
127 treatment, cellulose samples with the similar degree of polymerization (DP) are appropriate for their comparison.
128 Consequently, these celluloses were adjusted by trial and error for their DPs by changing the treatment condition
129 mentioned above for converting Cell I to various forms of celluloses.

130 **Determination of the degree of polymerization (DP) and crystallinity of the** 131 **celluloses**

132 The degree of polymerization (DP) is an important parameter to be considered in order to study the
133 decomposition behaviors. The viscosities of the celluloses were, therefore, measured by using 0.5M
134 cupriethylenediamine (Cuen) (TAPPI, 1982) in a Cannon Fenske capillary viscometer. The viscosity average
135 DP of the cellulose samples was calculated from the intrinsic viscosity $[\eta]$ according to the Eq. (1) (Sihtola et al.
136 1963):

137
$$DP^{0.905} = 0.75[\eta] \quad \text{Eq. (1)}$$

138 Apart from the DP, the crystallinity of the celluloses is another essential parameter. The XRD patterns
139 of these celluloses were recorded by X-ray diffractometer Rigaku RINT 2200 equipped with monochromator.
140 About 20mg of cellulose was placed on the glass sample holder and flattened carefully, then mounted on the
141 sample holder. X-ray diffraction (XRD) was conducted on reflectance modes through a 2θ range between 7.5°
142 and 32.5° by Cu- K_α radiation ($\lambda=0.1542\text{nm}$), operated at 40kV and 30mA at ambient temperature.

143 Gaussian functions were used to deconvolute the XRD patterns of various crystalline celluloses, and
144 the crystallinity was calculated from the ratio of the area of all crystalline peaks with less background to the total
145 area (Park et al. 2010).

146 The diffraction angles of each XRD pattern and their assignment diffraction planes are summarized
147 based on a newly proposed conventional indexing for various unit cells of cellulose (French 2013). Here, the
148 unit cells were mainly referenced to the orientation along c-axis, whereby various data from literatures and
149 experimental approaches were used, thus some variations may be occurred (Gardiner et al. 1985; Isogai 1994;
150 Langan et al. 2001; Nishiyama et al. 2002; 2003; Wada et al. 2004).

151 **Fourier transform-infrared (FT-IR) analysis of residual celluloses**

152 The FT-IR analysis was carried out for the residues of the celluloses obtained after HCW treatments.
153 The spectra of the dried sample pellets in KBr were recorded using a Shimadzu IR-8000 spectrophotometer. All
154 the spectra were recorded with an accumulation of 64 scans, revolution of 4 cm^{-1} , in a range from 4000 to 400
155 cm^{-1} .

156 **Treatment of various crystalline celluloses by semi-flow HCW**

157 The prepared celluloses (about 0.4g) as starting materials were treated individually within a 5ml
158 reaction vessel in a semi-flow HCW system at temperatures of 230°C , 250°C and 270°C under 10MPa for 2min
159 to 15min. The semi-flow HCW conversion system and its operational procedures as explained elsewhere were
160 adapted for this study (Lu et al. 2009; Phaiboonsilpa et al. 2010; 2011). Briefly, the ambient distilled water from
161 a water tank was flown through the reaction vessel by a pump in order to pressurize the system at 10MPa
162 controlled by a back-pressure regulator. To raise the temperature, the preheating unit monitored by
163 thermocouples was used to reach at the designated temperature for about 20min under 10MPa. In addition,
164 another heating unit was installed at the reaction vessel to maintain the designated temperature in the reaction
165 vessel, into which the HCW was passed through at the flow-rate of 10ml/min.

166 These treatments yielded residues of celluloses and WS portions. After the HCW passing through
167 reaction vessel, the WS portions were cooled down immediately by the cooling system to terminate all reactions.
168 The WS portions collected every 5min were allowed to settle in ambient temperature and pressure for a
169 minimum of 12h, before filtering them by $0.45\mu\text{m}$ membranes prior to subsequent analysis. The residues of
170 celluloses left in the reaction vessel were, on the other hand, collected, dried unless otherwise mentioned and
171 evaluated for its DP and crystallinity, again (Ehara et al. 2002; Kumar et al. 2010). The experimental process for
172 the study is illustrated in Fig. 1.

173 The WS portions were, then, characterized and analyzed by using high-performance anion-exchange
174 chromatography (HPAEC), high-performance liquid chromatography (HPLC) and capillary electrophoresis
175 (CE) as described previously (Lu et al. 2009; Phaiboonsilpa et al. 2010; 2011). The product percentages,
176 presented on oven-dried weight basis of the initial material, are based on the chromatogram peak areas of the
177 HPAEC, HPLC and CE.

178 Results and Discussion

179 To evaluate decomposition behaviors for various types of crystalline celluloses by semi-flow HCW
180 treatment, the celluloses must be the same in their DPs as the starting materials. Therefore, as shown in Table 1,
181 the DP of those celluloses was adjusted by trial and error to be similar. Thus, they can be directly compared as
182 they are decomposed. The XRD patterns of these starting celluloses are shown in Fig. 2. The diffraction angles
183 of each XRD pattern and their assignment diffraction planes are summarized in Table 2. For the celluloses in
184 Table 1 and Fig. 2, the semi-flow HCW treatments were carried out at temperatures of 230°C, 250°C and 270°C
185 under 10MPa for 2 min to 15min (230-270°C/10MPa/2-15min).

186 Evaluation of the residues of celluloses

187 The XRD patterns recorded for the residues of celluloses in group I and group II after XRD analysis are
188 shown in Figs. 3 and 4, respectively. In Fig. 3, the XRD patterns of the residues from Cell I are observed to
189 remain the same as in the starting Cell I, in contrast with the XRD patterns of residues from Cell III_I and Cell
190 IV_I. Two equatorial reflections of Cell III_I at $2\theta \approx 11.7$ and 20.8° indexed as 010 and $1\bar{1}0$, respectively, can be
191 observed at the starting of Cell III_I. After the HCW treatments at 230-270°C/10MPa/15min, the peak at $2\theta \approx$
192 11.7° , for all residues from Cell III_I was totally disappeared, whereas a peak at $2\theta \approx 20.8^\circ$ was noticeably
193 becoming smaller at elevated temperatures. In addition, the XRD peaks at $2\theta \approx 14.4$, 16.3 and 22.5° were
194 intensively appeared. These peaks were similar to those of Cell I, indexed as $\bar{1}10$, 110 and 200, respectively.
195 Here, therefore, all residues from Cell III_I were observed to be totally converted to Cell I. A similar result is
196 reported for Cell III_I treated in ethylenediamine (EDA) to be reversed and converted into Cell I by treatment in
197 warm water, whereas Cell III_I treated with liquid ammonia (140°C, 12kPa) was found to be stable to boiling
198 water for a few hours (Sueoka et al. 1973; Roche et al. 1981; Yatsu et al. 1986; Wada 2001; Wada et al. 2008).

199 Whereas for Cell IV_I, the peak at $2\theta \approx 15.4^\circ$ (indexed as $1\bar{1}0$) was shown to emerge into two peaks as
200 it was treated in the elevated temperatures. Although the changes were not seen for residue from Cell IV_I at
201 230°C, this corresponding peak started to be transformed at 250°C and 270°C, which was found to be
202 corresponded to that of Cell I; indexed as $\bar{1}10$ and 110.

203 As for the residues from group II shown in Fig. 4, the XRD patterns of residues from Cell II and Cell
204 IV_{II} remained the same. In contrast, the residues from Cell III_{II} were observed to be transformed to other
205 crystalline form. The peak at $2\theta \approx 12.1^\circ$ (indexed as 010) for all residues from Cell III_{II} was shown to have no
206 changes compared with the control Cell III_{II}. However, a peak at $2\theta \approx 20.4^\circ$ was completely emerged into two
207 peaks at $2\theta \approx 19.7$ and 22.0° , as can be seen with residues from Cell III_{II}. These peaks corresponded,
208 respectively, to crystalline peaks at 110 and 020 of Cell II. Moreover, a small peak at $2\theta \approx 15.1^\circ$ was noticed and
209 became more prominent at 270°C, which also matched up with crystalline peak of Cell IV_{II} that indexed as $1\bar{1}0$.

210 Thus, these outcomes of residues from Cell III_{II} after HCW treatments comprised a mixture of Cell II and Cell
211 IV_{II}.

212 All XRD patterns on residues from celluloses in Figs. 3 and 4 were recorded and compared for the
213 dried residual samples. Therefore, they were also examined under the wet conditions by X-ray diffractometry
214 for the residues from celluloses of Cell III_I, Cell IV_I and Cell III_{II}. Figure 5 shows the obtained XRD patterns of
215 residues from Cell III_I as treated by semi-flow HCW at lower temperature (230°C/10MPa/2-15min). It seems
216 apparent that Cell III_I has started to be converted to Cell I at an early stage of the HCW treatment. The peaks for
217 1 $\bar{1}$ 0 of Cell III_I were disappearing even after 2 min treatment. Similar observations were recorded for Cell III_I
218 treated at the higher temperature (250-270°C/10MPa/2-10min). In the same way, the wet residues of celluloses
219 were also examined and found that they had already been converted as well.

220 The same was performed for Cell IV_I and Cell III_{II}, at 230-270°C/10MPa/2-10min and they were found
221 to behave in a similar manner. Cell IV_I has started to be converted to Cell I at 10 min treatments under 250-
222 270°C/10MPa, whereas for Cell III_{II}, the conversion has occurred even at 2 min treatment under 230°C/10MPa
223 to Cell II mainly and to Cell IV_{II} to some extent. All these results showed that they have already been converted
224 at the early stage of the HCW treatment at 230-270°C/10MPa. Since Cell III_{II} was observed to be converted to
225 Cell II and Cell IV_{II}, further treatments for Cell II were also performed at 270°C/10MPa/2-10min in order to
226 check whether it would be converted to Cell IV_{II}. However, no significant crystallographic changes was
227 observed, thus it remained as Cell II at 270°C/10MPa/2-10min.

228 In relation to the residues from Cell III_I, Cell IV_I and Cell III_{II} in Figs. 3-5, the FT-IR measurements
229 were performed to compare with the XRD results. For the FT-IR spectra, they were focused only on the OH
230 stretching regions before and after the HCW treatments as shown in Fig. 6. The extensively discussed bands in
231 literatures at 3,480, 3,300 and 3150 cm⁻¹ were found in the spectra of Cell III_I, whereas the residues of celluloses
232 were found to be similar as those of halocynthia and ramie which are Cell I (Cell I β type) (Wada, 2001; Kokot et
233 al. 2002; Zugenmaier, 2008). The bands at 3,720 cm⁻¹ was clearly observed showing that Cell III_I has been
234 converted to Cell I.

235 In contrast, neither Cell IV_I nor Cell III_{II} was regularly discussed in literatures. The residues from Cell
236 IV_I that consisted of a mixture of Cell I and Cell IV_I, usually has higher proportion of Cell IV_I (Marrinan et al.
237 1956; Zugenmaier, 2008). Despite the presence of a high proportion of Cell IV_I, there seems to be no
238 distinctive absorption which could be attributed to Cell IV_I. It is, therefore, assumed that the spectra of the
239 residues from Cell IV_I resemble that from Cell I.

240 The Cell III_{II} spectrum has close resemblance with that of Cell II as suggested by Marrinan et al. (1956)
241 that the distance of the hydrogen bonding must be very similar in the two forms. Similarly, the general
242 appearance of Cell IV_{II} spectrum resembles that of Cell II. The FT-IR spectra of the residues from Cell III_{II} at
243 230-270°C agreed with the corresponding XRD patterns in Fig. 4 that consisted of a mixture of Cell II and Cell
244 IV_{II}.

245 All the observations above showed that the interconversion of various crystalline celluloses of Cell III_I,
246 Cell IV_I and Cell III_{II} has occurred at either early or later stage of the HCW treatments. From these results, a
247 possible interconversion for the preparation process and transformation of celluloses by HCW treatments can be
248 summarized as in Fig. 7.

249 In spite of the crystallographic changes occurred for the residues from Cell III_I, IV_I and III_{II}, the
250 changes in crystallinity and DP for residues of celluloses after HCW treatments were demonstrated in Fig. 8.
251 The crystallinity for residues from Cell III_I, IV_I and III_{II} was evaluated from the obtained XRD patterns as
252 treated by the corresponding temperatures. The overall results were found out to be increased slightly and
253 remained almost constant approximately at 90%. The slight increase could be due to the combined effects of an
254 aqueous environment that removed the paracrystalline portions of celluloses during hydrolysis (Weimer et al.
255 1995) and annealing that could take place on the cellulose at temperature between 220°C and 280°C (Yamamoto
256 et al. 1993). This observation was also in agreement with previous studies, that even at a severe treatment
257 temperature 270°C, the crystallinity was observed to remain almost unchanged (Sasaki et al. 2004; Jollet et al.
258 2009; Kumar et al. 2010, Tolonen et al. 2011). One study emphasized that (i) crystallinity and (ii) changes in the
259 crystalline form of a cellulose; as the two of the main determining factors efficacy of hydrolysis of biomass
260 (Cetinkol et al. 2010). However, it is generally thought that drying could lead to the formation of new hydrogen
261 bonds and recrystallization, which would increase the crystallinity rather than reducing it (Tolonen et al. 2011).

262 Contrary to the crystallinity results, the DP for all residues of celluloses was seen to decrease slightly
263 with increase in temperature (Tolonen et al. 2011). The expectation that the lower DP celluloses such as Cell
264 IV_{II} would be easier to decompose due to their shorter average lengths of molecules, however, was not observed
265 in Fig. 8. The DP was decreased and leveling off to about 70 even after the treatment at 270°C/10MPa/15min.
266 The present interpretations suggested that the crystalline structure of various types of celluloses were still rigid,
267 but only broken down into shorter-chain of cellulose molecules. As temperature increases, the DP decreases
268 which would raise the solubility of cellulose in water and conversion of cellulose to the hydrolyzed and
269 degraded products (Sasaki et al. 2004; Yu et al. 2009). In Fig. 8, the DP of group I celluloses was observed to be
270 generally higher than that of group II. This shows that group II is easier to be decomposed as compared with
271 group I. Also, based on visual examination, the fibers as the starting materials have been changed into powder-
272 liked residues after the HCW treatment.

273 Figure 9 illustrates the relationship of DP and residues of celluloses after HCW treatments. Both are
274 seen to decrease as the treatment temperatures were increased. It has also been reported in previous studies that
275 the DP of residues gradually decreased with increasing cellulose decomposition (Sasaki et al. 2004; Tolonen et
276 al. 2011).

277 The overall yield of the residues after semi-flow HCW treatment for various types of celluloses is
278 shown in Fig. 10. These residues notably decreased with the increase in temperatures. The residues of various
279 types of celluloses obtained particularly at treatment condition of 230°C/10MPa/15min were still quite high in
280 yield, in the range of 70-90wt%. This is agreeable as the condition was known not to hydrolyze the crystalline
281 cellulose in wood (Lu et al. 2009; Phaiboonsilpa et al. 2010; 2011).

282 Since in the present study, the starting DP was adjusted to be the same for all celluloses, a direct
283 comparison can be made for decomposition behaviors of celluloses. As DP in the residues of celluloses is a
284 function of cellulose decomposition, the observed changes on residues were only due to the different crystalline
285 structures of celluloses as starting materials. The various crystalline celluloses prepared as the starting materials
286 have assisted the decomposition process in HCW treatments even though the interconversion of some celluloses
287 have occurred.

288 Cell I yielded the highest residue among various types of celluloses used in this study followed by Cell
289 III_I and IV_I, as in Fig. 10. At 230 and 250°C, both Cell III_I and IV_I behaved very similarly, however, Cell IV_I
290 was decomposed more at 270°C as compared with Cell III_I. This trend was later followed by Cell II, III_{II} and IV_{II}.
291 Both Cell III_{II} and IV_{II} behaved in a similar way, nevertheless, Cell IV_{II} was decomposed more at 270°C as
292 compared with Cell III_{II}. Cell II was observed to have no residues left at 270°C. Therefore, no crystallinity and
293 DP values were recorded in Fig.8.

294 As a conclusion, Cell I in native crystalline form was found to be the most resistant against
295 decomposition, while Cell II, III_{II} and IV_{II} were decomposed the most as compared with the other types of
296 celluloses such as Cell III_I and IV_I. This could be an indication of the easiness for decomposition of the various
297 crystalline cellulose samples and it seemed likely that group I celluloses (Cell I, III_I and IV_I) had higher
298 resistance to be decomposed as compared with group II celluloses (Cell II, III_{II} and IV_{II}).

299 **Evaluation of the water-soluble (WS) portions**

300 For the temperature ranges under high pressure, ionic products of water increases, thus, hydrolysis
301 reaction would be proceeded without any catalysts (Franck 1987; Dinjus et al. 2004; Kruse et al. 2007; Lu et al.
302 2009, Phaiboonsilpa et al. 2010). If such a reaction is prolonged, hydrolyzed products would be degraded due to
303 high temperature treatment. A reaction scheme showing how cellulose I is decomposed as treated by semi-flow
304 HCW is shown in Fig. 11 (Phaiboonsilpa et al. 2010). The products resulted from cellulose decomposition in the
305 treatment can be categorized into hydrolyzed and degraded products.

306 Under these treatment conditions, the glucosidic linkages of cellulose are cleaved and cellulose starts
307 to be hydrolyzed into cello-oligosaccharides, subsequently, hydrolyzed to monosaccharide such as glucose
308 (Antal et al. 1990a; Ehara et al. 2002; Kruse et al. 2003; Phaiboonsilpa et al. 2010). Fructose and mannose are
309 also obtained during the process as a result of isomerization of glucose. Monosaccharides are unstable at high
310 temperature and thus some parts of them are further converted into their degraded products such as furfural, 5-
311 hydroxymethyl furfural (5HMF), levoglucosan through dehydration, and erythrose, glycolaldehyde,
312 methylglyoxal through fragmentation (Antal et al. 1990b). A prolonged treatment allows further degradation to
313 take place, generating other products such as organic acids. It is important to know the decomposition pathway
314 of cellulose as the degraded products could inhibit the fermentation process for ethanol production (Palmqvist et
315 al. 2000). The WS portions for various crystalline celluloses obtained from each treatment were found to follow
316 similar decomposition pathway as in Fig. 11.

317 To observe the decomposition behavior of various crystalline celluloses, the yields of the hydrolyzed
318 and degraded products were investigated. However, the latter is a result of the degradation of the former. Thus,
319 the total yields reflect the easiness of the hydrolysis decomposition behavior. Figure 12 shows one example of
320 the results on such yields. It can be seen that for both groups, more than 50wt% of the products consisted of
321 hydrolyzed products and that higher portion of them are obtained as compared with degraded products.
322 According to Fig.11, the hydrolyzed yields produced were still at the upper paths of cellulose decomposition
323 pathway. This could be an indication that the cellulose samples have resistance against decomposition. From Fig.
324 12, it is apparent that the group II is easier than group I for hydrolysis reactions. These results from WS portions
325 are in good agreement with those from residues of celluloses.

326

327 **Concluding Remarks**

328 The decomposition behaviors on various types of crystalline celluloses were investigated at 230-
329 270°C/10MPa/2-15min. To compare directly the effect of the treatment, the DP of the celluloses was adjusted
330 by trial and error to be similar prior to the treatment. Based on the results of the residues, crystallographic
331 changes were found to be occurred during the HCW treatment for Cell III_I, Cell IV_I and Cell III_{II}. In general,
332 group I celluloses (Cell I, III_I, Cell IV_I) have been converted to Cell I and group II (Cell II, III_{II}, IV_{II}) to Cell II.
333 Despite of these changes, the overall results of residues showed that group I has higher resistance to be
334 decomposed than group II. In the meantime, for the WS portions, celluloses in group II were decomposed more
335 to be the hydrolyzed and degraded products as compared with those in group I. The results on the WS portion
336 revealed that the degree of difficulty for decomposition is greater in group I than group II. Based on these
337 findings, it can be concluded that group I has more resistance to be decomposed than group II.

338 Based on the obtained results, it was clear that the decomposition behaviors were due to their different
339 crystalline forms of celluloses. For that reason, it is recommended to transform one type crystalline allomorph of
340 cellulose to the other for a better hydrolysis reaction. All these data are, therefore, useful in order to understand
341 the behaviors of different types of crystalline celluloses that provide information for efficient use of
342 lignocellulose.

343

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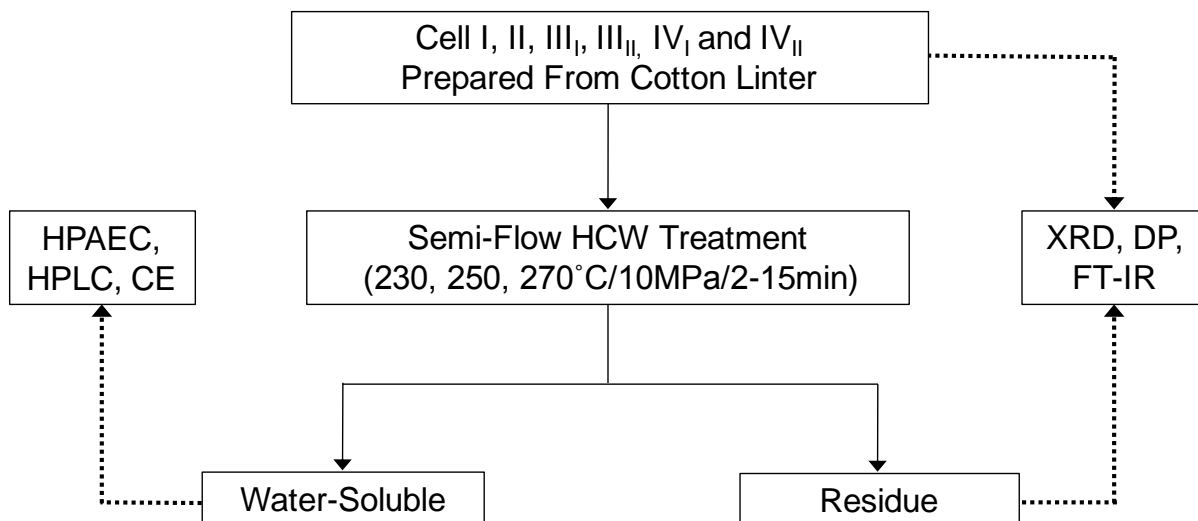


Fig.1 The experimental process to study the decomposition behaviors for various types of crystalline celluloses prepared from cotton linter

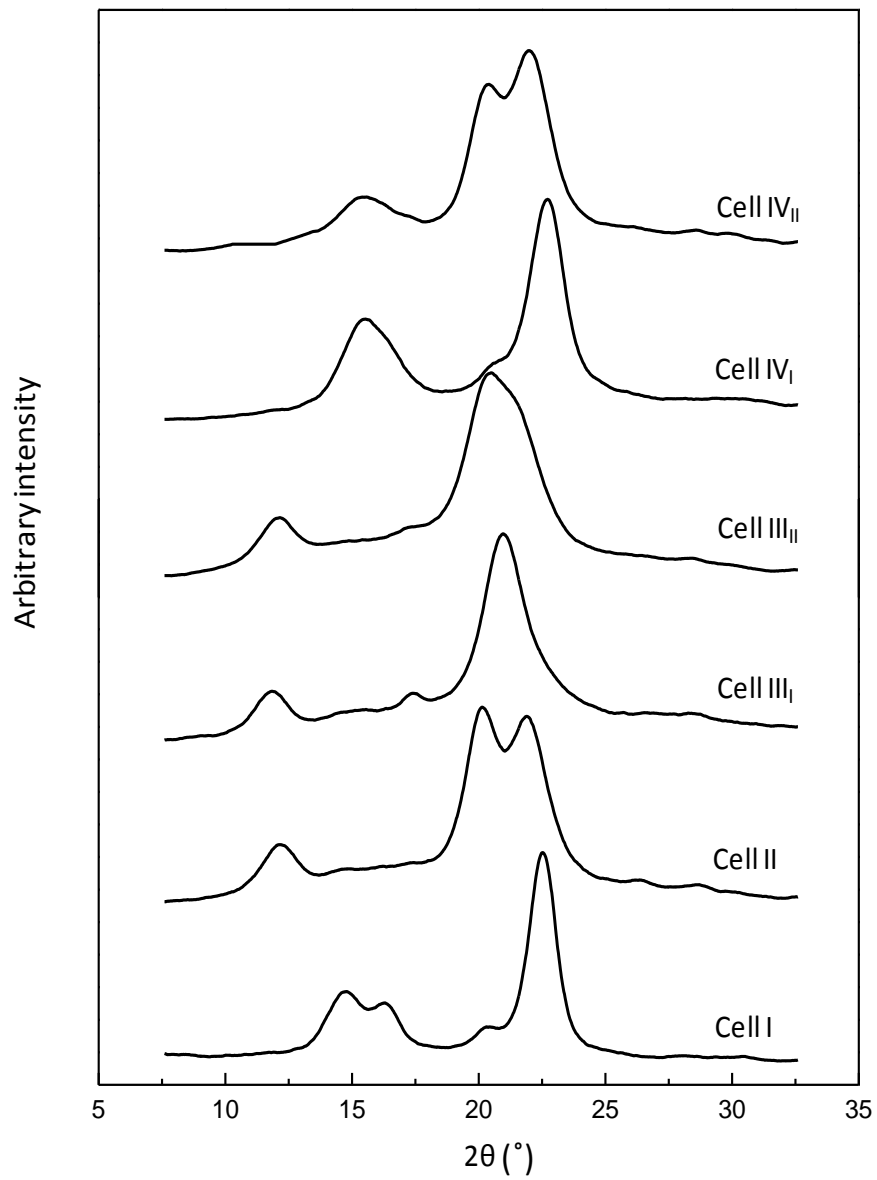


Fig.2 The XRD patterns for various types of crystalline celluloses prepared in this study

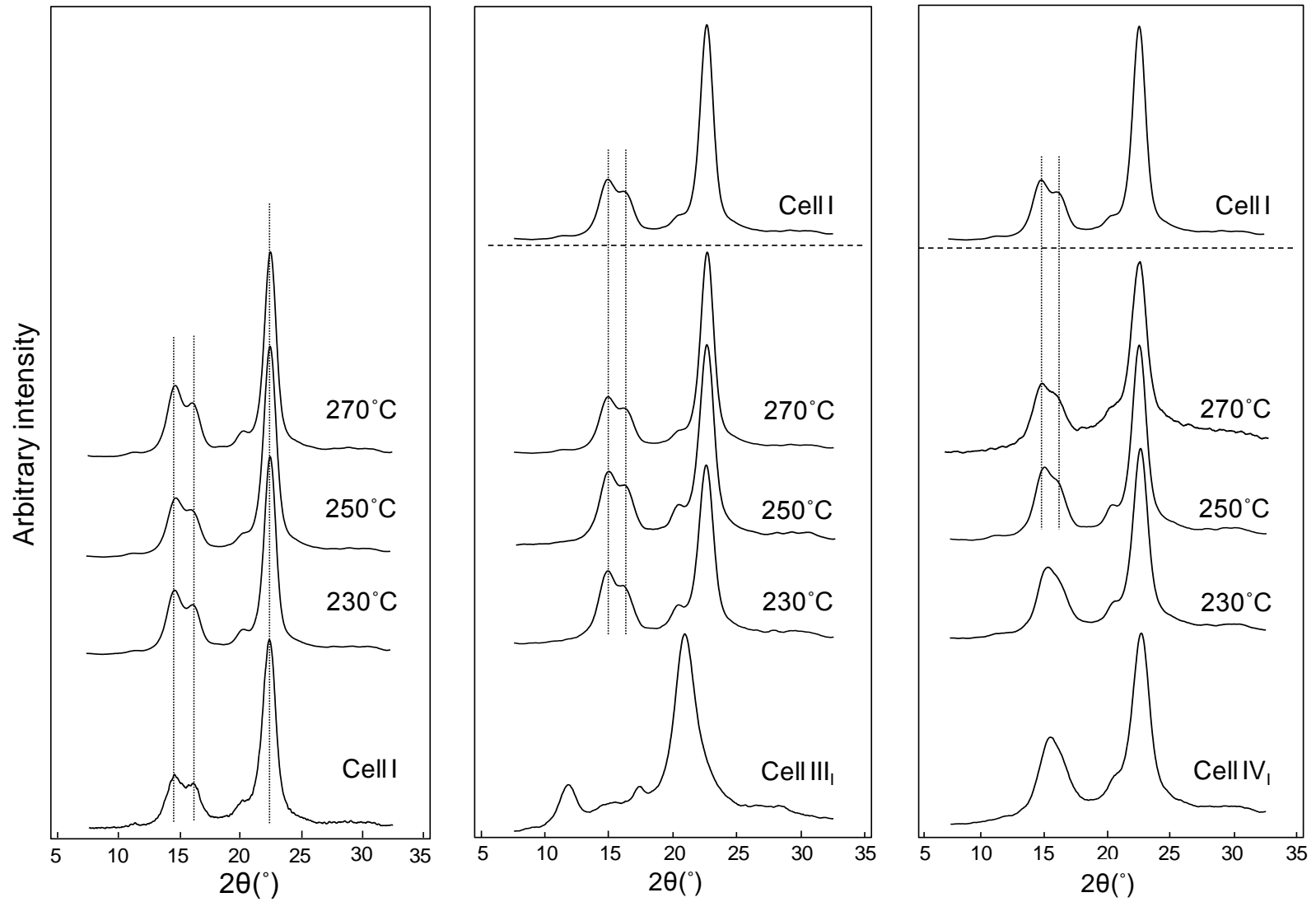


Fig. 3 The XRD patterns for the residues from Cell I, Cell III₁ and Cell IV₁ (from left to right) as treated by semi-flow HCW at 230-270°C/10MPa/15min

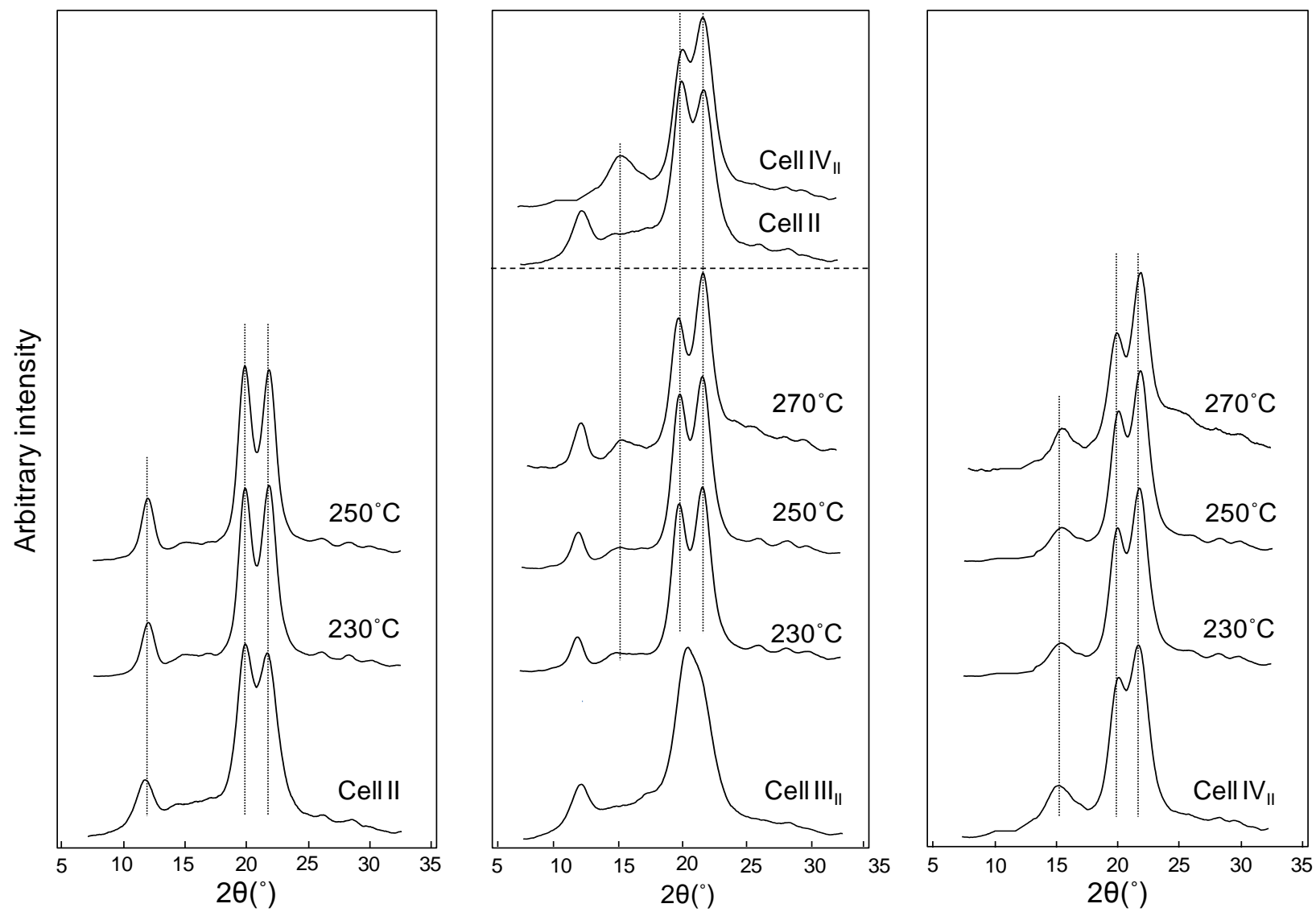


Fig. 4 The XRD patterns for the residues from Cell II, Cell IIIII and Cell IVII (from left to right) as treated by semi-flow HCW at 230-270°C/10MPa/15min

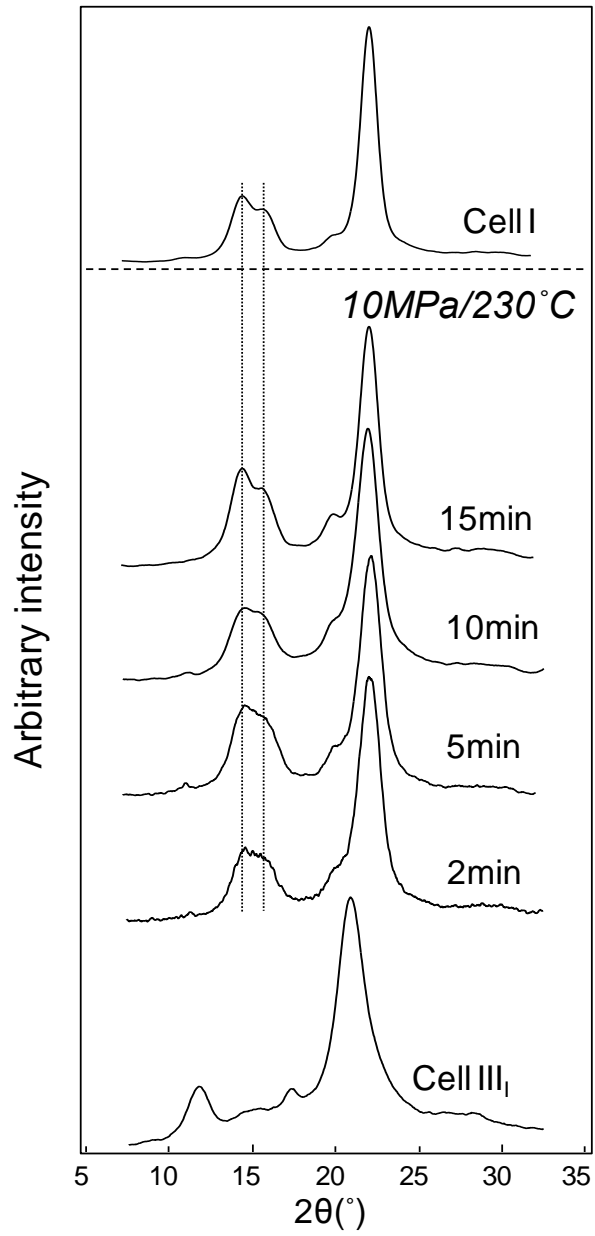


Fig. 5 The XRD patterns for the residues from Cell III₁ as treated by semi-flow HCW at 230°C/10MPa for different treatment times

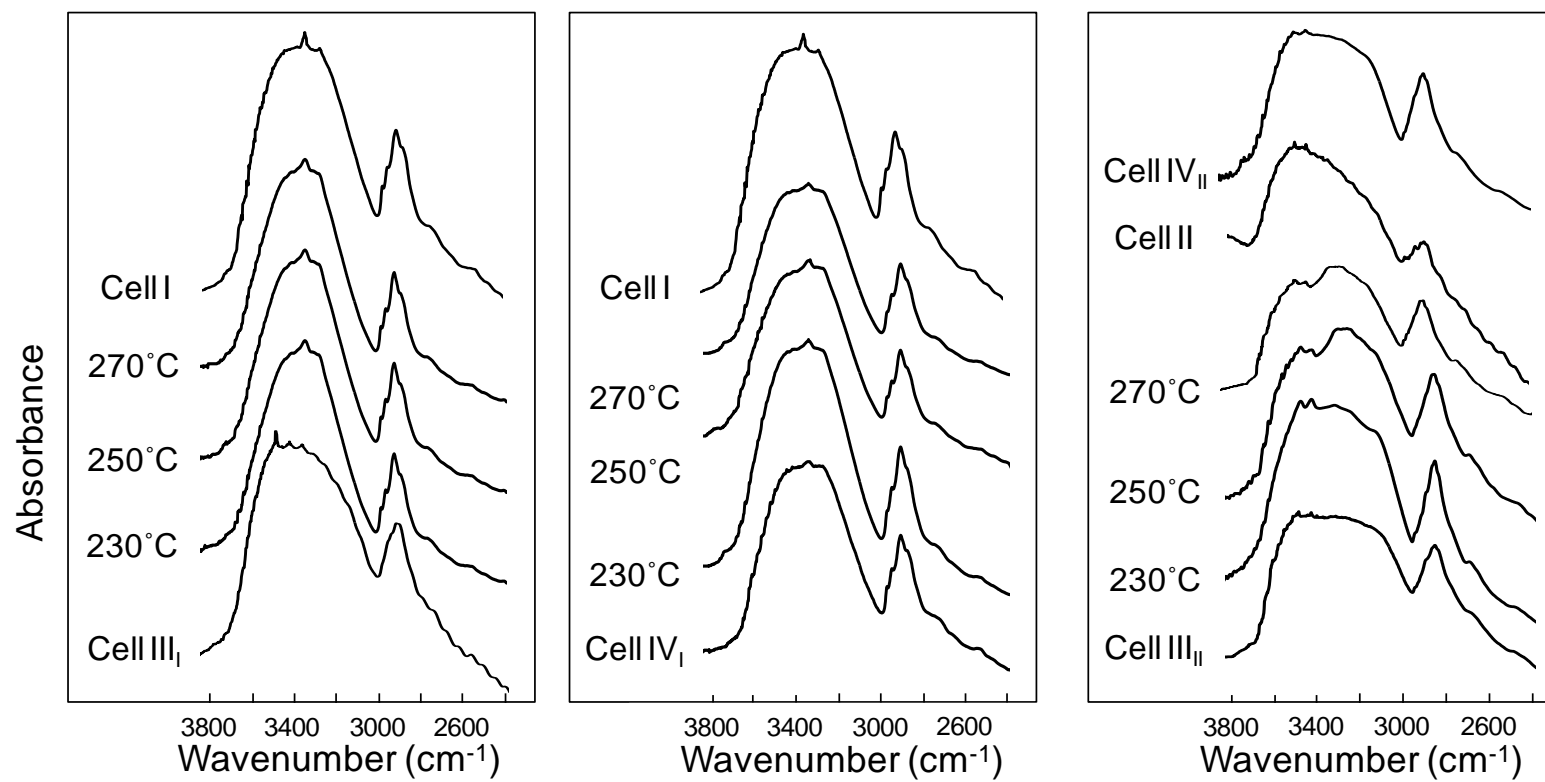


Fig. 6 The FT-IR spectra for the residues from Cell III_I, Cell IV_I and Cell III_{II} (from left to right) as treated by semi-flow HCW at 230-270°C/10MPa/15min

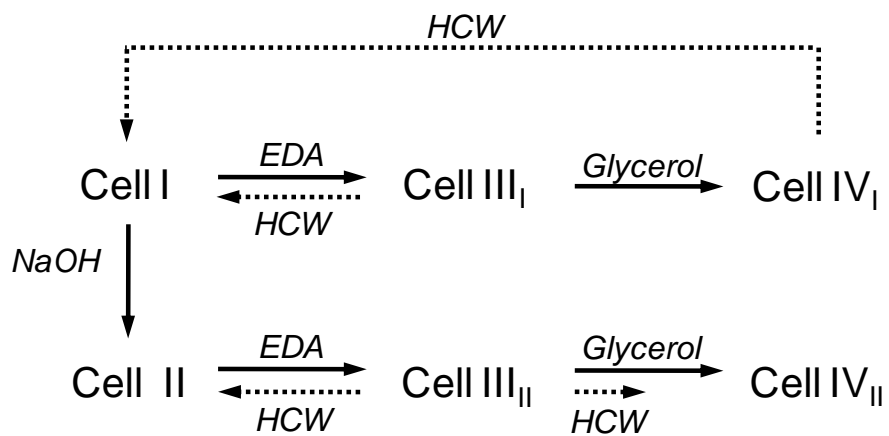


Fig. 7 The possible interconversion pathways for various types of crystalline celluloses

Solid line: Preparation process; Dotted line: Transformation by HCW

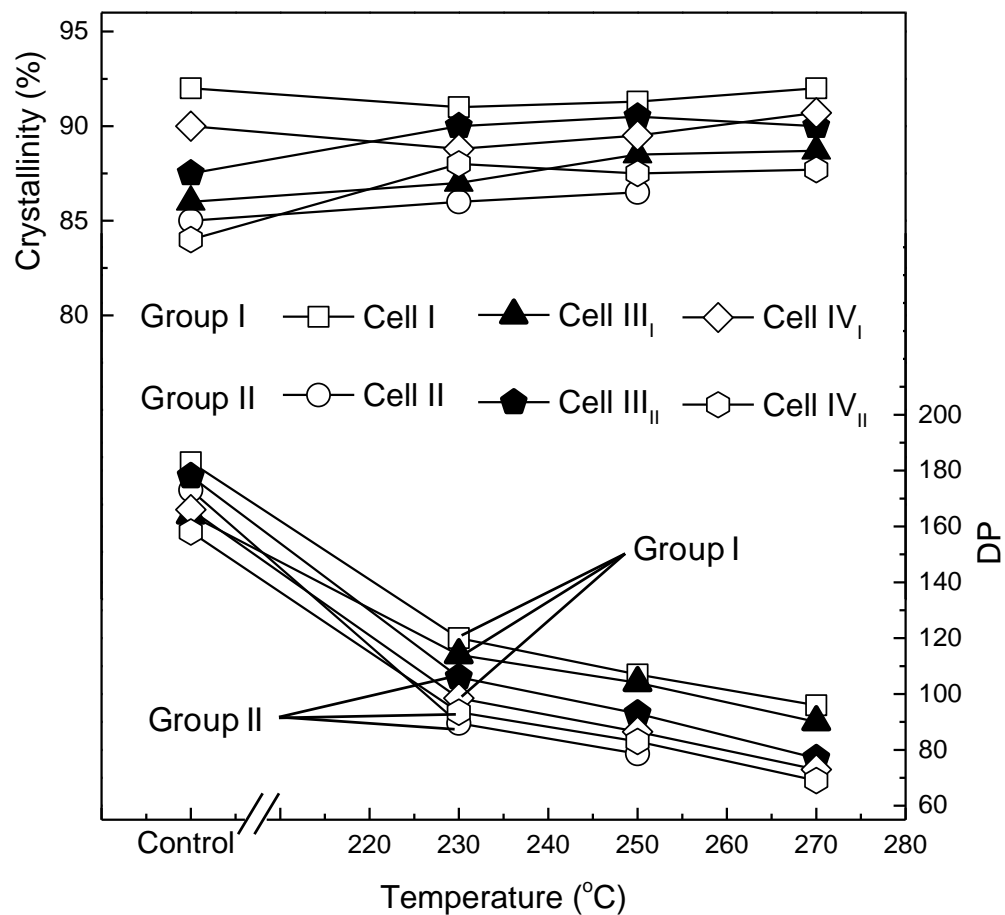


Fig. 8 The changes on crystallinity and DP for the residues from various crystalline celluloses as treated at different temperatures under 10MPa for 15 min (230-270°C/10MPa/15min) by semi-flow HCW

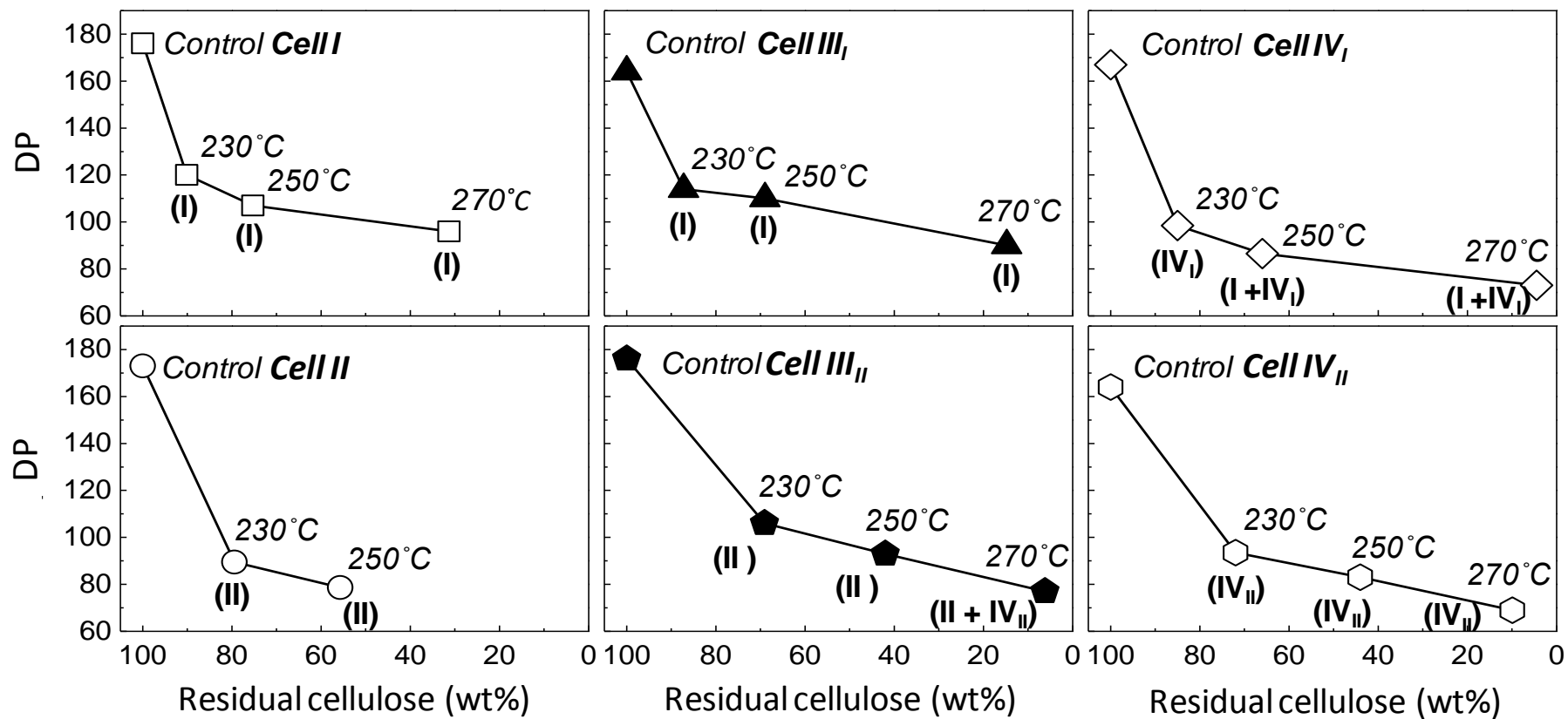


Fig. 9 The relationship between DP and residues from various crystalline celluloses as treated by semi-flow HCW at 230-270°C/10MPa/15min

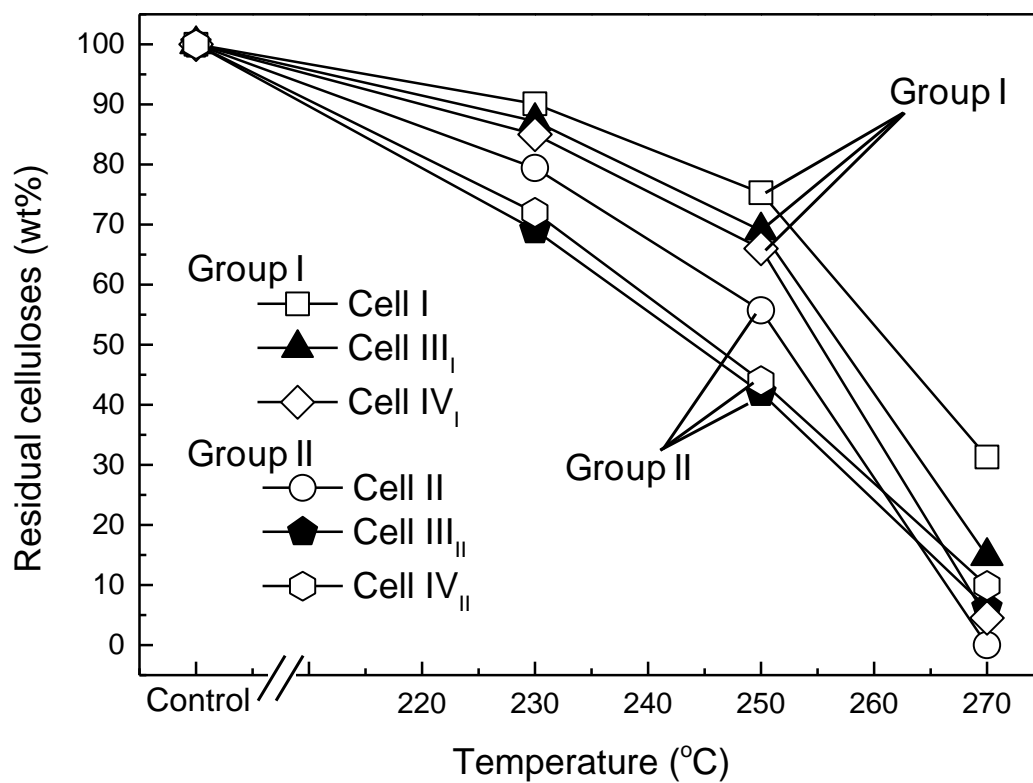


Fig. 10 The changes on the residues from various types of crystalline celluloses as treated at different temperatures under 10MPa for 15min (230-270°C/10MPa/15min) by semi-flow HCW

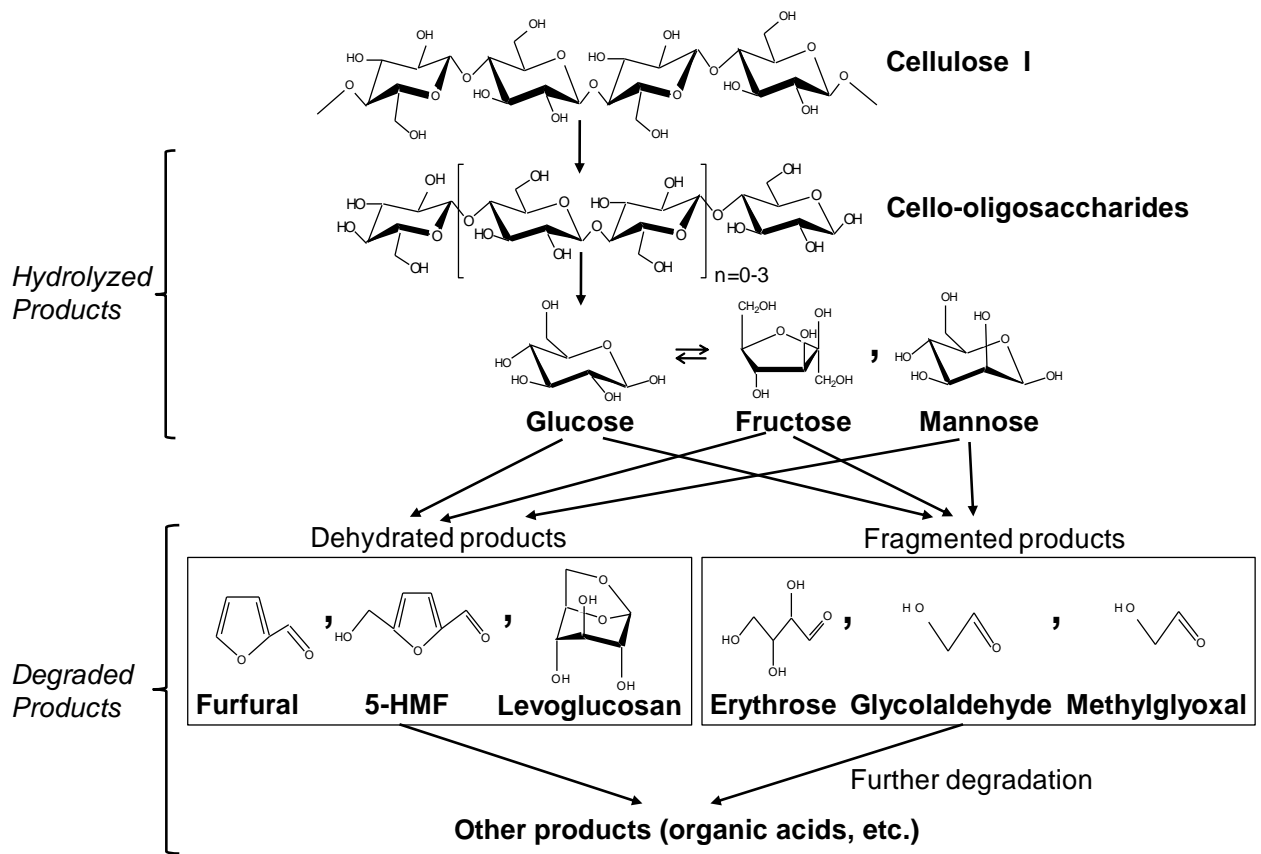


Fig. 11 Decomposition pathway of crystalline cellulose as treated by semi-flow HCW (adapted from Phaiboonsilpa et al. 2010)

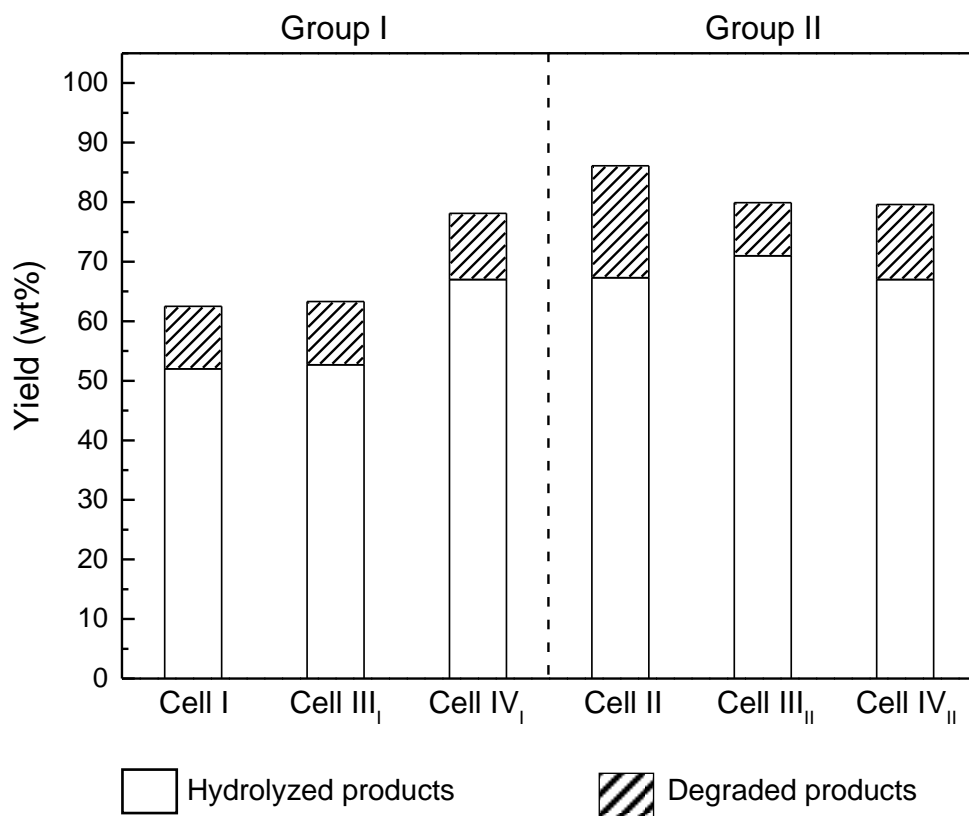


Fig. 12 The yields on the water-soluble portions from various types of crystalline celluloses as treated by semi-flow HCW at 270°C/10MPa/15min.

Table 1 The DP and crystallinity for various types of crystalline celluloses prepared in this study

Cell	DP	Crystallinity (%)
I	176	92
II	173	85
III _I	164	86
III _{II}	176	87
IV _I	167	90
IV _{II}	164	85

Table 2 The diffraction planes and angles in XRD patterns of various crystalline celluloses

Cell	Diffraction planes / Diffraction angles, 2θ (°)		
I	$\bar{1}10$ / 14.4	110 / 16.3	200 / 22.5
II	$\bar{1}10$ / 12.1	110 / 19.7	020 / 22.0
III _I	010 / 11.7	100 / 20.8	$1\bar{1}0$ / 20.8
III _{II}	010 / 12.1	100 / 20.4	$1\bar{1}0$ / 20.4
IV _I	$1\bar{1}0$ / 15.4	020 / 21.8	200 / 22.2
IV _{II}	$1\bar{1}0$ / 15.1	012 / 20.6	200 / 22.5

I, II and III_I according to French (2013)