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1	Preparation of immunostimulatory single-walled
2	carbon nanotube/CpG DNA complexes and
3	evaluation of their potential in cancer
4	immunotherapy
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1 Abstract

2 Carbon nanotubes (CNTs) have many interesting properties. In particular, their 3 photohyperthermic effect by near-infrared (NIR) irradiation could be used to kill cancer cells, 4 and could thus be applied in photohyperthermic therapy. However, the solubility of CNTs must 5 be improved before they can be used in biological applications. As DNA is reported to disperse 6 the CNTs in aqueous solution with π - π interactions, we hypothesis that immunostimulatory CpG 7 DNA may also disperse the CNTs in aqueous solution. In this study, we used CpG DNA to 8 disperse single-walled CNTs (SWCNTs) in aqueous solution, in order to combine 9 photohyperthermic effect and immunoactivation together to achieve a more effective cancer 10 therapy. As expected, CpG DNA effectively dispersed the SWCNTs in aqueous solution via the 11 formation of SWCNT/CpG DNA complexes. Moreover, the immunoreactivity of the 12 SWCNT/CpG DNA complexes was investigated. The results showed that intratumoral 13 administration of the SWCNT/CpG DNA complexes in mice enhanced the production level of inflammatory cytokines in tumor tissues. Finally, we evaluated the antitumor effects of the 14 15 SWCNT/CpG DNA complexes in tumor-bearing mice. The result indicated that intratumoral administration of the SWCNT/CpG DNA complexes combined with NIR irradiation was a more 16 17 effective approach to prevent the proliferation of tumor growth.

18



Keywords: Carbon nanotubes, CpG DNA, immune activation, photohyperthermia, cancer

1 1. Introduction

2 Carbon nanotubes (CNTs) consist exclusively of carbon atoms arranged in condensed 3 aromatic rings, which in turn are organized in one (single-walled carbon nanotube: SWCNT) 4 (Iijima and Ichihashi, 1993) or more (multi-walled carbon nanotube) (Iijima, 1991) concentric 5 graphene sheets rolled-up into cylinders. The diameters of the CNTs range from a few 6 nanometers to hundreds of nanometers. Owing to their unique structural and electronic 7 properties, CNTs have various possibilities for application in nanomedicine as biocompatible 8 materials for developing versatile drugs (Allen and Cullis, 2004; Bianco et al., 2005a), proteins 9 (Kam and Dai, 2005; Kam et al., 2004), genes (Cai et al., 2005; Liu et al., 2005), and vaccine 10 delivery systems (Panhuis, 2003). Especially, CNTs have a strong optical absorption in the near-11 infrared (NIR) region and can release this energy within tissue to produce localized heating 12 (Chakravarty et al., 2008; Kam et al., 2005). Therefore, CNTs may be promising photohyperthermic therapy agents for cancer therapy, because the selective thermal ablation of 13 14 malignant tissue is an important objective in cancer research and offers a viable alternative 15 treatment option when surgical resection is not possible (DeNardo GL and DeNardo SJ, 2008).

However, CNTs are completely insoluble in most organic solvents and aqueous buffers. Indeed, since the time when CNTs were first produced, there has much interest, particularly in the domain of biological and biomedical research, in enhancing the dispersibility and/or solubility of CNTs in aqueous solution to overcome this important problem. It has been reported that many detergents, such as sodium dodecyl benzene sulfonate and Triton-X100, are able to

disperse CNTs, but these detergents are generally toxic to normal cells (Dong et al., 2009). To 1 2 resolve this problem, several approaches to functionalize CNTs, including defect-group 3 chemistry, covalent sidewall chemistry, non-covalent wrapping by polymers, biopolymers, 4 surfactants and other amphiphilic molecules, have been explored (Heister et al., 2010; 5 Mackiewicz et al., 2008; Shim and Ahn, 2012; Stranks et al., 2012; Tasis et al., 2006;). More 6 recently, Zheng et al. reported that DNA can disperse CNTs in aqueous solution, where the aromatic nucleotide bases in DNA are suggested to interact with the CNTs through π - π stacking 7 8 (Hughes et al., 2007; Zheng et al., 2003a). Therefore, we considered that immunostimulatory 9 CpG DNA, which is capable of activating the immune system (Krieg, 2002), may be expected to 10 interact with CNTs to improve the dispersibility of the CNTs. Moreover, the immune responses 11 elicited by CpG DNA have the potential to inhibit tumor growth. In our previous studies, we 12 formulated complexes of phosphorothioate-type CpG DNA with cationic liposomes, and the 13 complexes showed strong immune responses and were effective in preventing proliferation of 14 cancer cells (Kuramoto et al., 2009, 2008; Zhou et al., 2012, 2010). Therefore, the combination 15 of photohyperthermic therapy and immunotherapy could be an effective approach for cancer 16 therapy.

In this study, we used CpG DNA to disperse SWCNTs via the formation of SWCNT/CpG
DNA complexes. It is suggested that these SWCNT/CpG DNA complexes may possess not only
a photohyperthermic effect, but also an immunostimulatory effect. First, we obtained the results

that CpG DNA dispersed the SWCNTs in aqueous solution individually. Then, the 1 2 photohyperthermic effects of the SWCNT/CpG DNA complexes were observed in vitro and in 3 vivo with NIR laser irradiation. In addition, the SWCNT/CpG DNA complexes induced high 4 levels of inflammatory cytokine production in serum and in tumor tissues when intratumorally 5 administrated to colon 26 tumor-bearing mice. Furthermore, the SWCNT/CpG DNA complexes when irradiated with NIR laser, significantly inhibited tumor growth in colon 26 tumor-bearing 6 7 mice. Thus, SWCNT/CpG DNA complexes with NIR laser irradiation would be an effective 8 approach in cancer immunotherapy.

1 **2. Material and Methods**

2 2.1 Oligonucleotides

3 Oligonucleotides with phosphorothioate backbones were purchased from Operon (Tokyo,

4 Japan). The sequences of the oligonucleotides 5'were 5 TCGACGTTTTGACGTTTTGACGTTTT-3' 5'-(CpG DNA) and 6 TGCAGCTTTTGAGCTTTTGAGCTTTT-3' (GpC DNA).

7 2.2 Cell lines

Murine adenocarcinoma colon 26 tumor cells (Zhou et al., 2010) were grown in 5% CO₂ in
humidified air at 37 °C with RPMI1640 medium (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan)
supplemented with 10% fetal bovine serum (MP Biomedicals, Inc., Irvine, CA, USA), 100 IU/ml
penicillin, 100 µg/ml streptomycin, and 2 mM L-glutamine.

12 **2.3 Animals**

Male BALB/c (5-week-old) mice were purchased from the Shizuoka Agricultural Cooperative Association for Laboratory Animals (Shizuoka, Japan). All the animals were housed with free access to food and water. The light (dark/light cycle was 12/12 h), temperature, and humidity were kept constant throughout the experiments. All animal experiments were approved by the Animal Experimentation Committee of the Graduate School of Pharmaceutical Sciences,
 Kyoto University.

3 2.4 Preparation of SWCNT/CpG DNA complex

One milligram of HiPco SWCNTs (containing 9 wt% of TGA Residue as Fe(iv) from the
manufacturer information, Carbon Nanotechnologies Incorporated) and 5 mg of CpG DNA
(Operon, Tokyo, Japan) were sonicated in 5 ml of 5% dextrose solution for 20 minutes in an icewater bath by the formation of complexes (SWCNT/CpG DNA complexes).

8 **2.5 Evaluation of the amount of CpG DNA that's bound to the SWCNTs.**

9 The amount of CpG DNA adsorbing to SWCNTs surface was determined by 10 spectrophotometically. The SWCNTs suspension complexed with CpG DNA was filtrated with 11 0.22 um centrifugal filter device (Ultrafree-MC, Merck Millipore, Darmstadt, Germany) to 12 remove free CpG DNA witch dose not bind to SWCNTs. Filtrated SWCNTs was resuspended 13 with distilled water, and optical absorbance spectrum was measured using Jasco V-670 14 spectrophotometer (Tokyo, Japan). The absorbance spectrum showed a peak at 260 nm and a 15 trough at 230 nm in UV region, which are characteristics of the absorbance of DNA. Then, the 16 amount of CpG DNA contained in SWCNTs suspension was calculated from the absorbance 17 difference between at 260 nm and at 230 nm.

18 **2.6 Evaluation of microscopic characterization of SWCNT/CpG DNA complex**

1	The SWCNT/CpG DNA complex was centrifuged for 155 minutes at 284,000 g to remove
2	any insoluble material. Then, the supernatant was filtered through a 30-kDa molecular weight
3	filter (Amicon), and extensively washed several times with 5 ml of 5% dextrose to remove any
4	unbound CpG DNA. The absorption spectra of the SWCNT/CpG DNA complex were analyzed
5	using a Perkin-Elmer Lambda 900 UV/Vis/NIR spectrometer. Luminescence spectra of the
6	SWCNT/CpG DNA complex were analyzed using a SHIMADZU NIR-PL System. AFM images
7	were analyzed using an Asylum Technology MFP-3D-SA.
8	2.7 Temperature increase of SWCNT/CpG DNA complex solutions by NIR laser
9	irradiation
10	The SWCNT/CpG DNA complex was suspended at concentrations of 10 μ g of
11	SWCNTs/ml or 20 µg of SWCNTs/ml in 48-well plates. Two hundred microliters of each
12	suspension was irradiated at 2 W/cm ² or 3 W/cm ² for 5 minutes by using an 808-nm NIR laser
13	(Chameleon-RF, Coherent Japan Inc., Tokyo, Japan). The temperature was measured using a
14	thermocouple.
15	2.8 Evaluation of cancer cell ablation induced by SWCNT/CpG DNA complex with NIR

17 The SWCNT/CpG DNA complex (1 μ g of SWCNTs) or naked CpG DNA (2 μ g) were 18 added to colon 26 cells (1×10⁴) and incubated for 24 hours. Then, the cells were washed three times with PBS, and exposed to 808 nm of NIR laser for 2 minutes at 5 W/cm². Cell death was
 assessed using a Live-Dead-Cell Staining Kit (BioVision Inc., California, USA).

3 2.9 Evaluation of temperature increase and HSP70 production induced by SWCNT/CpG 4 DNA complex with NIR laser irradiation in vivo

Colon 26 cells (3×10^5) were transplanted into the flanks of mice (n=5 for each group). Once 5 6 the tumors reached a mean diameter of 5 mm, 5% of dextrose, naked CpG DNA (10 µg), the 7 SWCNT/CpG DNA complex (1 µg of SWCNTs and 10 µg of CpG DNA) or the SWCNT/GpC 8 DNA complex (1 µg of SWCNTs and 10 µg of GpC DNA) were intratumorally injected into the 9 mice. Then, 5% of dextrose, naked CpG DNA (10 µg) and the SWCNT/CpG DNA complex 10 treated groups were divided into NIR laser irradiated and non-NIR laser irradiated groups. For 11 the NIR laser irradiated groups, 24 hours after administration, the tumors were irradiated with 12 NIR laser 3 times of 30 s-on and 30 s-off once daily for 3 days. Tumor tissues were collected at different time points and HSP70 production levels were measured using a HSP70 ELISA kit 13 14 (Enzo Life Sciences Inc., New York, USA). The temperature increase of the tumor tissues was 15 monitored using an InfReC Thermography R300 (NEC Avio Infrared Technologies Co., Ltd., Tokyo, Japan) after NIR laser irradiation for 30 s. 16

17 2.10 Evaluation of cytokine production in tumor tissues and serum after SWCNT/CpG 18 DNA complex intratumoral injection

1	Colon 26 cells (3×10^5) were transplanted into the flanks of mice (n=5 for each group). Once
2	the tumors reached a mean diameter of 5 mm, naked CpG DNA (10 μ g), the SWCNT/CpG DNA
3	complex (1 μg of SWCNTs and 10 μg of CpG DNA) or the SWCNT/GpC DNA complex (1 μg
4	of SWCNTs and 10 μg of GpC DNA) were intratumorally injected into the mice. Then, 2 hours
5	later, tumor tissues and the serum were collected. Then, inflammatory cytokines including TNF-
6	α , IL-12p70 and IL-6 productions were measured with a BD Cytometric Bead Array (CBA)
7	Mouse Inflammation Kit (Becton, Dickinson and Company, Tokyo, Japan) by using a BD
8	FACSCantoTM II Flow Cytometer (Becton, Dickinson and Company, Tokyo, Japan).
9	2.11 Evaluation of cytokine enhancement in tumor tissues induced by SWCNT/CpG DNA
10	complex after NIR laser irradiation
11	The SWCNT/CpG DNA complex (1 μ g of SWCNTs and 10 μ g of CpG DNA) was
12	intratumorally injected into mice. Then, the SWCNT/CpG DNA complex treated mice were
13	divided into NIR laser irradiated and non-NIR laser-irradiated groups. For the NIR laser
14	irradiated groups, 24 h after administration, tumors were irradiated with NIR laser three times at
15	30-s-on and 30-s-off bursts once daily for 3 days. The tumor tissues were collected at 6 hours
16	after the last NIR laser irradiation and cytokines including TNF- α , IL-12p70 and IL-6
17	productions were measured using a BD Cytometric Bead Array (CBA) Mouse Inflammation Kit.
18	2.12 Evaluation of antitumor effect by SWCNT/CpG DNA complex with NIR laser
19	irradiation

1	Colon 26 cells (3×10^5) were transplanted into the flanks of mice. Once the tumors reached a
2	mean diameter of 5 mm, naked CpG DNA (10 μ g), the SWCNT/CpG DNA complex (1 μ g of
3	CNT and 10 μ g of CpG DNA), or the SWCNT/GpC DNA complex (1 μ g of CNT and 10 μ g of
4	GpC DNA) were intratumorally injected into the mice. Then, 24 hours later, NIR laser was
5	irradiated (30 s on and 30 s off for 3 times) for 3 days. The tumor sizes were measured up to 62
6	days after the transplantation of the colon 26 cells.
7	

8 **3. Results and Discussion**

9 3.1 Characterization of SWCNT/CpG DNA complexes

10 CNTs have various possible applications in nanomedicine. However, such potential 11 biological applications of CNTs will require improvements to be made to the dispersibility 12 and/or solubility of CNTs in aqueous solution. Several papers have reported that wrapping of the 13 CNTs by DNA was found to be sequence-dependent (Zheng et al., 2003b), especially in the case 14 of Poly (T), which has the highest dispersion efficiency among fixed length homopolymers (Zheng et al., 2003a). Other combinations of DNA sequence motifs such as $d(GT)_n$ or $d(AC)_n$ 15 were also investigated (Vogel et al., 2007). However, the interaction of CNTs with CpG DNA is 16 17 less well studied. In this study, 1 mg of HiPco SWCNTs and 5 mg of CpG DNA were sonicated in 5 ml of 5% dextrose solution for 20 minutes in an ice-water bath to form SWCNT/CpG DNA 18 19 complexes. Microscopic characterizations of the resultant SWCNT/CpG DNA complexes were

analyzed by UV/Visible-NIR absorption spectroscopy (Fig. 1a), photoluminescence 1 2 spectroscopy (Fig. 1b), and atomic force microscopy (AFM) (Fig. 1c). The SWCNT/CpG DNA 3 complexes were dispersed individually without the formation of remarkable aggregates, as 4 evidenced by the appearance of a high peak in the absorption spectra (Fig. 1a,). As the 5 photoluminescence isn't observed for aggregates but only for individualized SWCNTs, the strong 6 fluorescence in the photoluminescence spectra (Fig. 1b) indicated that well dispersed SWCNTs 7 were present. However, this result could not rule out the presence of some bundles. Meanwhile, 8 AFM images clearly showed that the SWCNT/CpG DNA complexes were well dispersed and 9 had a mean diameter of about 1–4 nm (Fig. 1c) and hight of about 0.5-2 nm (Fig. 1d). In addition, 10 removal of free DNA by a 30-kDa molecular weight filter did not cause SWCNT flocculation to 11 occur, and the resultant SWCNT/CpG DNA complexes were stable for months at 4 °C. The 12 SWCNT/CpG DNA complexes were also redispersed in 5% dextrose solution once freeze-dried, 13 indicating that the CpG DNA binding to the SWCNTs was very strong. Furthermore, the amount 14 of CpG DNA contained in SWCNTs suspension was calculated from the absorbance difference 15 between at 260 nm and at 230 nm, and determined to be 1.27 ± 0.24 ug/1 ug SWCNTs. Taegar 16 and collaborators have reported that the best dispersion efficiency is achieved at DNA: CNT 17 ratios of 1:1 or 1:2 (Taegar et al., 2005). Our result was comparable with their result. The 18 fragmentation of CpG DNA was not achieved by sonication for 20 minutes (Fig. S1a). The 19 fragmentation of CpG DNA by sonication in the presence of SWCNTs was also checked. The 20 results were showed in Fig. S2b. The concentration of CpG DNA which by the sonication with

1	SWCNTs were slightly decreased compared with CpG DNA that were not sonicated. However,
2	Large fragment of CpG DNA in SWCNT/CpG DNA complex were not detected. We considered
3	that during the preparation of SWCNT/CpG DNA complex, SWCNTs may have litter effect on
4	CpG DNA fragmentation. Moreover, in vitro studies showed that the sonicated CpG DNA
5	maintained an equivalent immunostimulatory potential to that of CpG DNA prior to sonication
6	(Fig. S1c). These results suggested that CpG DNA may still exhibit immunostimulatory activity
7	after sonication.

8 3.2 In vitro and in vivo photohyperthermic effects of SWCNT/CpG DNA complexes with 9 NIR irradiation

10 Supratim et al. recently demonstrated that DNA-encased CNTs are efficient for thermal 11 ablation (Ghosh et al., 2009). To investigate whether the SWCNT/CpG DNA complexes also 12 keep the photohyperthermic effect by NIR laser irradiation, the temperature increases achieved 13 following NIR laser irradiation of the SWCNT/CpG DNA complexes were evaluated. In Fig. 2, when a SWCNT/CpG DNA complex solution was irradiated with a NIR laser at 3 W/cm² (Fig. 14 2b), the temperature increased more rapidly than irradiation at 2 W/cm² (Fig. 2a). Additionally, 15 the increases in temperature of the SWCNT/CpG DNA complex solution were dependent on the 16 17 concentration of SWCNTs in solution and the NIR laser irradiation time. Moreover, the 18 photohyperthermic effect of the SWCNT/CpG DNA complexes was equal to the photohyperthermic effect of SWCNT/GpC DNA complexes (in which the SWCNTs dispersed 19

with GpC DNA exhibited no immunostimulatory effects), and SWCNT/Pluronic F-127 (in which
the SWCNTs were dispersed with the surfactant, Pluronic F127) (Fig. 2c). These results suggest
that the SWCNT/CpG DNA complexes also exhibit a thermal effect when irradiated with an NIR
laser, thereby highlighting their potential use for photohyperthermic therapy.

5 As the SWCNT/CpG DNA complexes released substantial vibrational energy after exposure 6 to NIR light, the use of NIR irradiation for the induction of hyperthermia is particularly attractive 7 because living tissues do not strongly absorb in this range (Weissleder, 2001). Therefore, a 8 number of biomedical investigations of CNTs have focused on their application for the treatment 9 of cancer (Liu et al., 2008a; Meng et al., 2008; Singh et al., 2005). In this study, the results of an 10 in vitro photohyperthermic ablation of the murine colorectal cancer cell line, colon 26, treated with the SWCNT/CpG DNA complex, using NIR irradiation, were evaluated with a Live-Dead-11 12 Cell Kit. It was shown that most of the colon 26 cells treated with the SWCNT/CpG DNA 13 complex (1 µg of SWCNTs) were dead after NIR laser irradiation (Fig. 3f), as compared with the 14 same cells treated with the SWCNT/CpG DNA complex without NIR laser irradiation (Fig. 3e). 15 Meanwhile, the untreated (NT) cells (Fig. 3b) and naked CpG DNA treated cells (Fig. 3d) 16 survived even after receiving NIR laser irradiation. These results suggested that the 17 SWCNT/CpG DNA complex induced colon 26 cell cytotoxicity after NIR laser irradiation.

18 Moreover, we also investigated the photohyperthermic effects of the SWCNT/CpG DNA 19 complex in vivo by evaluating the temperature increases and the production of HSP70 in tumor

1	tissue after NIR laser irradiation. HSPs are induced by elevated temperatures (typically in excess
2	of 43 °C) and serve as endogenous cellular markers of thermal stress (Burke et al., 2009). As
3	seen in Fig. 4a, after 30 s of NIR laser irradiation, the temperature of the tumor tissues treated
4	with the SWCNT/CpG DNA complex or the SWCNT/GpC DNA (not immunostimulatory)
5	complex were increased approximately up to 43 °C. On the other hand, the temperature of the
6	tumor tissues treated with 5% dextrose or naked CpG DNA remained the same as the
7	temperature before NIR laser irradiation. Then, we evaluated the production of HSP70 in tumor
8	tissue after NIR laser irradiation. As shown in Fig. 4b, HSP70 production in the tumor tissues of
9	mice treated with the SWCNT/CpG DNA complex + NIR were significantly higher than that in
10	mice treated only with the SWCNT/CpG DNA complex at 6 hours after NIR laser irradiation (P
11	< 0.01). In addition, in the tumor treated with the SWCNT/GpC DNA complex + NIR laser, the
12	HSP70 production was induced to a high level (Fig. 4c). In contrast, the HSP70 production in
13	mice treated with 5% dextrose or naked CpG DNA was of a comparable level to that in the mice
14	that did not receive NIR laser irradiation (Fig. 4c). These results suggest that both SWCNT
15	complexes when stimulated with NIR laser irradiation increased the temperature in the tumor
16	tissue and induced the enhancement of HSP70 production.

3.3 Immune effect of SWCNT/CpG DNA complexes in tumor-bearing mice

CpG DNA can be taken up by macrophages and dendritic cells, leading to the production of
Th1-type cytokines such as IFN-γ (Krieg, 2002). In our previous studies, we have demonstrated

1	that CpG DNA Lipoplex showed strong immune responses and were effective in preventing
2	proliferation of cancer cells (Kuramoto et al., 2009, 2008; Zhou et al., 2012, 2010). Therefore,
3	using CpG DNA to disperse the SWCNTs may lead to complexes having an immunostimulatory
4	effect. In this study, the immunoreactivity of the SWCNT/CpG DNA complex was evaluated by
5	measuring the production of inflammatory cytokines in serum and in the tumor tissues in colon
6	26 bearing mice at different time points (Fig. S3-S4). After intratumoral injection of the
7	SWCNT/CpG DNA complex, the production of TNF- α and IL-6 in the serum and the tumor
8	tissues showed a peak value at 2 hours after administration (Fig. S3a, S3b, S4a and S4b). The
9	production of IL-12p70 showed a peak value in the serum at 8 hours after administration (Fig.
10	S3c) and in the tumor tissues at 2 hours after administration (Fig. S4c).

11 In addition, compared with untreated mice or mice injected with naked CpG DNA or the 12 SWCNT/GpC DNA complex, mice injected with the SWCNT/CpG DNA complex significantly 13 induced higher levels of TNF- α and IL-6 production both in serum (Fig. 5a and 5b) and in tumor tissue (Fig.6a and 6b) at 2 hours after injection. The SWCNT/CpG DNA complex also induced 14 15 higher levels of IL-12p70 production in tumor tissues than compared with either naked CpG 16 DNA or the SWCNT/GpC DNA complex (Fig. 5c). These results demonstrated that the 17 SWCNT/CpG DNA complex induced TNF- α and IL-6 cytokine production in tumor tissues and 18 in serum, and may have the potential to inhibit tumor growth. As several papers have already 19 demonstrated that CNTs are able to enter immune cells such as macrophages actively and

1 passively via incomplete phagocytosis or by impaling the membrane (Cheng et al., 2009; Kam et 2 al., 2005; Kraszewski et al., 2012), we considered that the SWCNT/CpG DNA complex may be 3 more effectively recognized by the immune cells than naked CpG DNA. Moreover, our results 4 suggest that the SWCNT/CpG DNA complex induced higher levels of cytokine production than 5 naked CpG DNA, thereby confirming the reports that functionalized CNTs could enhance CpG 6 DNA uptake and their immunostimulatory properties (Bianco et al., 2005b; Zhao et al., 2011). 7 Furthermore, Fan et al. also has reproted that functionalized carbon nanotubes that conjugated 8 with CpG DNA enhanced activity of CpG DNA and was effective for both brain and systemic 9 melanomas (Fan et al., 2012).

10 As some reports have shown that the photohyperthermic effect could enhance immune 11 responses in vivo (Zhang et al., 2008; Gastpar et al., 2005), next, we investigated the immune 12 effects of the SWCNT/CpG DNA complex after NIR laser irradiation. The results showed that 13 there was no significant difference between the cytokine production levels of the SWCNT/CpG 14 DNA complex and the SWCNT/CpG DNA complex + NIR laser irradiation (data not shown). 15 This result may be because the photohyperthermic effects induced by the SWCNT/CpG DNA 16 complex + NIR were either not enough to enhance the immune response, or the enhanced 17 immune response was too low to be detected. We will investigate these details in the future.

3.4 Enhancement of anti-tumor effect by SWCNT/CpG DNA complexes after NIR laser
 irradiation

1	Then, the antitumor effect of the SWCNT/CpG DNA complex was also evaluated in tumor-
2	bearing mice. For this study, 3×10^5 colon 26 cells were subcutaneously injected into the flanks of
3	the tumor-bearing mice. Once the tumor volumes reached approximately 200 mm ³ , the mice
4	were injected intratumorally with 50 μ l of treatment solution, which contained 10 μ g of either
5	naked CpG DNA, the SWCNT/CpG DNA complex (1 μ g of SWCNTs and 10 μ g of CpG DNA)
6	or the SWCNT/GpC DNA complex, which are not immunostimulatory (1 μ g of SWCNTs and 10
7	μ g of GpC DNA). (n = 6 per group). It was found that there was no statistical differences among
8	the group mean tumor volumes of the three solutions tested at the onset of treatment. After
9	treatment, compared with the untreated (NT), the NIR laser-only, and the SWCNT/GpC DNA
10	complex + NIR studies, the tumor volumes of the SWCNT/CpG DNA complex + NIR treated
11	mice had decreased significantly by day 24 (P < 0.001 compared with the NT and NIR laser –
12	only; P < 0.01 compared with the SWCNT/GpC DNA complex + NIR) and by day 26 (P < 0.001
13	compared with NT; $P < 0.01$ compared with NIR laser – only; $P < 0.05$ compared with the
14	SWCNT/GpC DNA complex + NIR) after tumor cell implantation (Fig. 7). Moreover, the
15	SWCNT/CpG DNA complex + NIR treated mice were even significantly decreased compared
16	with naked CpG DNA or naked CpG DNA + NIR treated mice ($P < 0.05$) (Fig. 7). The tumor
17	volumes of the SWCNT/CpG DNA complex + NIR treated mice were smaller than that of the
18	SWCNT/CpG DNA complex treated mice, albeit a minor difference. These results demonstrated
19	that the combination of SWCNT/CpG DNA complex administration and NIR laser irradiation

led to a more efficient antitumor effect than compared with that achieved by naked CpG DNA
 administration or SWCNT/GpC DNA complex administration.

3 Several papers have reported that CNTs are effective in photohyperthermic therapy (Burke et al., 2009; Ghosh et al., 2009; Xiao et al., 2009). An advantage of NIR is that biological 4 5 systems largely lack chromophores that absorb in this region, and as such, the NIR light should 6 effectively and safely penetrate normal tissue and ablate any cells to which the CNTs are 7 attached. Moreover, hyperthermic therapy has been clinically used in the management of solid 8 tumors because it can synergistically enhance tumor cytotoxicity when combined with 9 chemotherapy or radiotherapy (Falk and Issels, 2001; Wust et al., 2002). However, the 10 combination of immunotherapy with CNTs is less well studied. In the present study, we 11 successfully combined the photohyperthermic effects and immune effects together using the 12 SWCNT/CpG DNA complex, and observed the resultant antitumor effect in vivo (Fig. 7). In 13 practice, the toxicology of SWCNTs also needs to be addressed prior to using them in clinical 14 situations. It has been found that well-functionalized CNTs, which are stable in physiological 15 environments, are non-toxic to cells in vitro and non-toxic to mice in vivo (Liu et al., 2009; Liu 16 et al., 2008; Schipper et al., 2008; Yang et al., 2008). Furthermore, Kagan et al. recently showed 17 that hypochlorite and reactive radical intermediates of the human neutrophil enzyme, 18 myeloperoxidase, enhance the biodegradation of SWCNTs in vitro, in neutrophils, and to a lesser

1	degree, in macrophages (Kagan et al., 2010). Therefore, the SWCNT/CpG DNA complex may
2	be applied for effective cancer immunotherapy without significant toxicity.

3 4. Conclusions

We successfully prepared the SWCNTs / CpG DNA complexes, which were dispersed well 4 5 and possessed both the photohyperthemic effects and the immune effects. The SWCNTs / CpG 6 DNA complexes enhanced the production level of inflammatory cytokines in tumor tissues when intratumoral administration. We also have shown an effective combination cancer therapy using 7 8 SWCNTs / CpG DNA complexes in mice. Intratumoral administration of SWCNTs / CpG DNA 9 complexes with NIR irradiation prevented the proliferation of tumor growth in mice and prolonged the survival time of mice. These were suggested that combination of 10 11 photohyperthermic therapy and immunotherapy would be an effective approach in cancer 12 therapy.

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2 FIGURES

3	Figure 1. Characterization of the SWCNT/CpG DNA complex. (a) UV-Vis-NIR absorption		
4	spectra of the SWCNT/CpG DNA complex. (b) Photoluminescence maps of the SWCNT/CpG		
5	DNA complex. (c) AFM images of the SWCNT/CpG DNA complex. (d) Height of		
6	SWCNT/CpG DNA complex in AFM images. Results are representative of three experiments.		
7	Figure 2. Temperature increases of the SWCNT/CpG DNA complex through NIR laser		
8	irradiation. SWCNT/CpG DNA complex solutions were prepared at two concentrations (10 μg		
9	CNT/ml and 20 μg CNT/ml) and placed in 48-well plates. Each sample was then irradiated with		
10	an 808-nm NIR laser at a different power level—(a) power level 2W/cm ² , (b) power level		
11	3W/cm ² —for 5 minutes. The temperatures were measured every 20 s. (c) Temperature increases		
12	of the SWCNT/CpG DNA complex, the SWCNT/GpC DNA complex, and the SWCNT /F-127		
13	after NIR laser irradiation. Results are representative of three experiments.		
14	Figure 3. In vitro photohyperthermic effects of the SWCNT/CpG DNA complex. The viabilities		
15	of colon 26 cells treated with naked CpG DNA or the SWCNT/CpG DNA complex following		
16	NIR laser irradiation were examined using a Live-Dead-Cell Kit. The representative images from		
17	each sample are shown. Cells with green fluorescence were considered alive, whereas those with		
18	red fluorescence were dead. (a) Untreated colon 26 cells. (b) Untreated colon 26 cells + NIR		
19	laser. (c) Colon 26 cells treated with CpG DNA alone. (d) Colon 26 cells treated with CpG DNA		

+ NIR. (e) Colon 26 cells treated with the SWCNT/CpG DNA complex. (f) Colon 26 cells
 treated with the SWCNT/CpG DNA complex + NIR. Results are representative of more than
 three experiments.

4	Figure 4. Temperature increases and HSP70 production induced by NIR laser irradiation of the
5	SWCNT/CpG DNA complex in vivo. (a) Temperature increases in tumor tissues after NIR laser
6	irradiation. (b) HSP70 production induced by NIR laser irradiation of the SWCNT/CpG DNA
7	complex in tumor tissues at different time points. * $P < 0.05$ compared with SWCNT/CpG DNA
8	complex + NIR at 2 hours after NIR laser irradiation. ** $P < 0.01$ compared with SWCNT/CpG
9	DNA complex at 6 hours, and SWCNT/CpG DNA complex + NIR at 24 hours after NIR laser
10	irradiation. (c) HSP70 production induced by NIR laser irradiation of 5% dextrose, naked CpG
11	DNA, the SWCNT/CpG DNA complex or the SWCNT/GpC DNA complex in tumor tissue at 6
12	hours after NIR laser irradiation. * $P < 0.05$ compared with 5% dextrose and the SWCNT/CpG
13	DNA complex; ** P < 0.01 compared with 5% dextrose + NIR and naked CpG DNA + NIR. # P
14	< 0.05 compared with 5% dextrose + NIR, naked CpG DNA + NIR, and the SWCNT/CpG DNA
15	complex. Each value represents the mean + S. D. ($n = 5$). Results are representative of three
16	experiments.

Figure 5. Cytokine productions induced by the SWCNT/CpG DNA complex in tumor tissue. (a)
TNF-α production levels in tumor tissues after naked CpG DNA, SWCNT/CpG DNA complex
or SWCNT/GpC DNA complex administration. * P < 0.05 compared with NT and SWCNT/GpC

1	DNA complex. (b) IL-6 production levels in tumor tissues after naked CpG DNA, SWCNT/CpC		
2	DNA complex or SWCNT/GpC DNA complex administration. * $P < 0.05$ compared with naked		
3	CpG DNA; ** P < 0.01 compared with NT and the SWCNT/GpC DNA complex. (c) IL-12p70		
4	production levels in tumor tissues after naked CpG DNA, SWCNT/CpG DNA complex or		
5	SWCNT/GpC DNA complex administration. Each value represents the mean + S. D. ($n = 5-6$).		
6	Results are representative of more than three experiments.		
7			
8	Figure 6. Cytokine productions induced by the SWCNT/CpG DNA complex in serum. (a) TNF-		
9	α production levels in serum after naked CpG DNA, SWCNT/CpG DNA complex or		
10	SWCNT/GpC DNA complex administration. * $P < 0.05$ compared with NT and the		
11	SWCNT/GpC DNA complex. (b) IL-6 production levels in serum after naked CpG DNA,		
12	SWCNT/CpG DNA complex or SWCNT/GpC DNA complex administration. ** P < 0.01		
13	compared with NT, naked CpG DNA and the SWCNT/GpC DNA complex. Each value		
14	represents the mean + S. D. ($n = 5-6$). Results are representative of more than three experiments.		
15			
16	Figure 7. Tumor volume of mice after combination of SWCNT/CpG DNA complex intratumoral		

- 17 administration and NIR laser irradiation. # P < 0.05 compared with naked CpG DNA and naked
- 18 CpG DNA + NIR; ## P < 0.01 compared with SWCNT/GpC DNA +NIR; ### P < 0.001

1	compared with NT and NIR 1	aser - only; * $P < 0.05$ compared	ared with SWCNT/GpC DNA +NIR;
		27	

- 2 ** P < 0.01 compared with NIR laser only; *** P < 0.001 compared with NT. Each value
- 3 represents the mean + S. D. (n = 6). Results are representative of more than three experiments.





























Days after Tumor Implantation (Day)