

Abstract

 Carbon nanotubes (CNTs) have many interesting properties. In particular, their photohyperthermic effect by near-infrared (NIR) irradiation could be used to kill cancer cells, and could thus be applied in photohyperthermic therapy. However, the solubility of CNTs must be improved before they can be used in biological applications. As DNA is reported to disperse 6 the CNTs in aqueous solution with $\pi-\pi$ interactions, we hypothesis that immunostimulatory CpG DNA may also disperse the CNTs in aqueous solution. In this study, we used CpG DNA to disperse single-walled CNTs (SWCNTs) in aqueous solution, in order to combine photohyperthermic effect and immunoactivation together to achieve a more effective cancer therapy. As expected, CpG DNA effectively dispersed the SWCNTs in aqueous solution via the formation of SWCNT/CpG DNA complexes. Moreover, the immunoreactivity of the SWCNT/CpG DNA complexes was investigated. The results showed that intratumoral administration of the SWCNT/CpG DNA complexes in mice enhanced the production level of inflammatory cytokines in tumor tissues. Finally, we evaluated the antitumor effects of the SWCNT/CpG DNA complexes in tumor-bearing mice. The result indicated that intratumoral administration of the SWCNT/CpG DNA complexes combined with NIR irradiation was a more effective approach to prevent the proliferation of tumor growth.

Keywords: Carbon nanotubes, CpG DNA, immune activation, photohyperthermia, cancer

1. Introduction

 Carbon nanotubes (CNTs) consist exclusively of carbon atoms arranged in condensed aromatic rings, which in turn are organized in one (single-walled carbon nanotube: SWCNT) (Iijima and Ichihashi, 1993) or more (multi-walled carbon nanotube) (Iijima, 1991) concentric graphene sheets rolled-up into cylinders. The diameters of the CNTs range from a few nanometers to hundreds of nanometers. Owing to their unique structural and electronic properties, CNTs have various possibilities for application in nanomedicine as biocompatible materials for developing versatile drugs (Allen and Cullis, 2004; Bianco et al., 2005a), proteins (Kam and Dai, 2005; Kam et al., 2004), genes (Cai et al., 2005; Liu et al., 2005), and vaccine delivery systems (Panhuis, 2003). Especially, CNTs have a strong optical absorption in the near- infrared (NIR) region and can release this energy within tissue to produce localized heating (Chakravarty et al., 2008; Kam et al., 2005). Therefore, CNTs may be promising photohyperthermic therapy agents for cancer therapy, because the selective thermal ablation of malignant tissue is an important objective in cancer research and offers a viable alternative treatment option when surgical resection is not possible (DeNardo GL and DeNardo SJ, 2008).

 However, CNTs are completely insoluble in most organic solvents and aqueous buffers. Indeed, since the time when CNTs were first produced, there has much interest, particularly in the domain of biological and biomedical research, in enhancing the dispersibility and/or solubility of CNTs in aqueous solution to overcome this important problem. It has been reported that many detergents, such as sodium dodecyl benzene sulfonate and Triton-X100, are able to

 disperse CNTs, but these detergents are generally toxic to normal cells (Dong et al., 2009). To resolve this problem, several approaches to functionalize CNTs, including defect-group chemistry, covalent sidewall chemistry, non-covalent wrapping by polymers, biopolymers, surfactants and other amphiphilic molecules, have been explored (Heister et al., 2010; Mackiewicz et al., 2008; Shim and Ahn, 2012; Stranks et al., 2012; Tasis et al., 2006;). More recently, Zheng et al. reported that DNA can disperse CNTs in aqueous solution, where the 7 aromatic nucleotide bases in DNA are suggested to interact with the CNTs through π - π stacking (Hughes et al., 2007; Zheng et al., 2003a). Therefore, we considered that immunostimulatory CpG DNA, which is capable of activating the immune system (Krieg, 2002), may be expected to interact with CNTs to improve the dispersibility of the CNTs. Moreover, the immune responses elicited by CpG DNA have the potential to inhibit tumor growth. In our previous studies, we formulated complexes of phosphorothioate-type CpG DNA with cationic liposomes, and the complexes showed strong immune responses and were effective in preventing proliferation of cancer cells (Kuramoto et al., 2009, 2008; Zhou et al., 2012, 2010). Therefore, the combination of photohyperthermic therapy and immunotherapy could be an effective approach for cancer therapy.

 In this study, we used CpG DNA to disperse SWCNTs via the formation of SWCNT/CpG DNA complexes. It is suggested that these SWCNT/CpG DNA complexes may possess not only a photohyperthermic effect, but also an immunostimulatory effect. First, we obtained the results

 that CpG DNA dispersed the SWCNTs in aqueous solution individually. Then, the photohyperthermic effects of the SWCNT/CpG DNA complexes were observed in vitro and in vivo with NIR laser irradiation. In addition, the SWCNT/CpG DNA complexes induced high levels of inflammatory cytokine production in serum and in tumor tissues when intratumorally administrated to colon 26 tumor-bearing mice. Furthermore, the SWCNT/CpG DNA complexes when irradiated with NIR laser, significantly inhibited tumor growth in colon 26 tumor-bearing mice. Thus, SWCNT/CpG DNA complexes with NIR laser irradiation would be an effective approach in cancer immunotherapy.

2. Material and Methods

2.1 Oligonucleotides

Oligonucleotides with phosphorothioate backbones were purchased from Operon (Tokyo,

 Japan). The sequences of the oligonucleotides were 5'- TCGACGTTTTGACGTTTTGACGTTTT-3' (CpG DNA) and 5'- TGCAGCTTTTGAGCTTTTGAGCTTTT-3' (GpC DNA).

2.2 Cell lines

8 Murine adenocarcinoma colon 26 tumor cells (Zhou et al., 2010) were grown in 5% CO₂ in humidified air at 37 °C with RPMI1640 medium (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan) supplemented with 10% fetal bovine serum (MP Biomedicals, Inc., Irvine, CA, USA), 100 IU/ml 11 penicillin, 100 μ g/ml streptomycin, and 2 mM L-glutamine.

2.3 Animals

 Male BALB/c (5-week-old) mice were purchased from the Shizuoka Agricultural Cooperative Association for Laboratory Animals (Shizuoka, Japan). All the animals were housed with free access to food and water. The light (dark/light cycle was 12/12 h), temperature, and humidity were kept constant throughout the experiments. All animal experiments were approved

 by the Animal Experimentation Committee of the Graduate School of Pharmaceutical Sciences, Kyoto University.

2.4 Preparation of SWCNT/CpG DNA complex

 One milligram of HiPco SWCNTs (containing 9 wt% of TGA Residue as Fe(iv) from the manufacturer information, Carbon Nanotechnologies Incorporated) and 5 mg of CpG DNA (Operon, Tokyo, Japan) were sonicated in 5 ml of 5% dextrose solution for 20 minutes in an ice-water bath by the formation of complexes (SWCNT/CpG DNA complexes).

2.5 Evaluation of the amount of CpG DNA that's bound to the SWCNTs.

 The amount of CpG DNA adsorbing to SWCNTs surface was determined by spectrophotometically. The SWCNTs suspension complexed with CpG DNA was filtrated with 0.22 um centrifugal filter device (Ultrafree-MC, Merck Millipore, Darmstadt, Germany) to remove free CpG DNA witch dose not bind to SWCNTs. Filtrated SWCNTs was resuspended with distilled water, and optical absorbance spectrum was measured using Jasco V-670 spectrophotometer (Tokyo, Japan). The absorbance spectrum showed a peak at 260 nm and a trough at 230 nm in UV region, which are characteristics of the absorbance of DNA. Then, the amount of CpG DNA contained in SWCNTs suspension was calculated from the absorbance difference between at 260 nm and at 230 nm.

2.6 Evaluation of microscopic characterization of SWCNT/CpG DNA complex

17 The SWCNT/CpG DNA complex (1 µg of SWCNTs) or naked CpG DNA (2 µg) were 18 added to colon 26 cells (1×10^4) and incubated for 24 hours. Then, the cells were washed three

1 times with PBS, and exposed to 808 nm of NIR laser for 2 minutes at 5 W/cm². Cell death was assessed using a Live-Dead-Cell Staining Kit (BioVision Inc., California, USA).

2.9 Evaluation of temperature increase and HSP70 production induced by SWCNT/CpG DNA complex with NIR laser irradiation in vivo

5 Colon 26 cells (3×10^5) were transplanted into the flanks of mice (n=5 for each group). Once 6 the tumors reached a mean diameter of 5 mm, 5% of dextrose, naked CpG DNA (10 μ g), the 7 SWCNT/CpG DNA complex (1 µg of SWCNTs and 10 µg of CpG DNA) or the SWCNT/GpC 8 DNA complex (1 µg of SWCNTs and 10 µg of GpC DNA) were intratumorally injected into the 9 mice. Then, 5% of dextrose, naked CpG DNA (10 µg) and the SWCNT/CpG DNA complex treated groups were divided into NIR laser irradiated and non-NIR laser irradiated groups. For the NIR laser irradiated groups, 24 hours after administration, the tumors were irradiated with NIR laser 3 times of 30 s-on and 30 s-off once daily for 3 days. Tumor tissues were collected at different time points and HSP70 production levels were measured using a HSP70 ELISA kit (Enzo Life Sciences Inc., New York, USA). The temperature increase of the tumor tissues was monitored using an InfReC Thermography R300 (NEC Avio Infrared Technologies Co., Ltd, Tokyo, Japan) after NIR laser irradiation for 30 s.

2.10 Evaluation of cytokine production in tumor tissues and serum after SWCNT/CpG DNA complex intratumoral injection

3. Results and Discussion

3.1 Characterization of SWCNT/CpG DNA complexes

 CNTs have various possible applications in nanomedicine. However, such potential biological applications of CNTs will require improvements to be made to the dispersibility and/or solubility of CNTs in aqueous solution. Several papers have reported that wrapping of the CNTs by DNA was found to be sequence-dependent (Zheng et al., 2003b), especially in the case of Poly (T), which has the highest dispersion efficiency among fixed length homopolymers 15 (Zheng et al., 2003a). Other combinations of DNA sequence motifs such as $d(GT)_n$ or $d(AC)_n$ were also investigated (Vogel et al., 2007). However, the interaction of CNTs with CpG DNA is less well studied. In this study, 1 mg of HiPco SWCNTs and 5 mg of CpG DNA were sonicated in 5 ml of 5% dextrose solution for 20 minutes in an ice-water bath to form SWCNT/CpG DNA complexes. Microscopic characterizations of the resultant SWCNT/CpG DNA complexes were

 analyzed by UV/Visible-NIR absorption spectroscopy (Fig. 1a), photoluminescence spectroscopy (Fig. 1b), and atomic force microscopy (AFM) (Fig. 1c). The SWCNT/CpG DNA complexes were dispersed individually without the formation of remarkable aggregates, as evidenced by the appearance of a high peak in the absorption spectra (Fig. 1a,). As the photoluminescence isn't observed for aggregates but only for individualized SWCNTs, the strong fluorescence in the photoluminescence spectra (Fig. 1b) indicated that well dispersed SWCNTs were present. However, this result could not rule out the presence of some bundles. Meanwhile, AFM images clearly showed that the SWCNT/CpG DNA complexes were well dispersed and had a mean diameter of about 1–4 nm (Fig. 1c) and hight of about 0.5-2 nm (Fig. 1d). In addition, removal of free DNA by a 30-kDa molecular weight filter did not cause SWCNT flocculation to occur, and the resultant SWCNT/CpG DNA complexes were stable for months at 4 °C. The SWCNT/CpG DNA complexes were also redispersed in 5% dextrose solution once freeze-dried, indicating that the CpG DNA binding to the SWCNTs was very strong. Furthermore, the amount of CpG DNA contained in SWCNTs suspension was calculated from the absorbance difference 15 between at 260 nm and at 230 nm, and determined to be 1.27 ± 0.24 ug/1 ug SWCNTs. Taegar and collaborators have reported that the best dispersion efficiency is achieved at DNA: CNT ratios of 1:1 or 1:2 (Taegar et al., 2005). Our result was comparable with their result. The fragmentation of CpG DNA was not achieved by sonication for 20 minutes (Fig. S1a). The fragmentation of CpG DNA by sonication in the presence of SWCNTs was also checked. The results were showed in Fig. S2b. The concentration of CpG DNA which by the sonication with

3.2 In vitro and in vivo photohyperthermic effects of SWCNT/CpG DNA complexes with NIR irradiation

 Supratim *et al*. recently demonstrated that DNA-encased CNTs are efficient for thermal ablation (Ghosh et al., 2009). To investigate whether the SWCNT/CpG DNA complexes also keep the photohyperthermic effect by NIR laser irradiation, the temperature increases achieved following NIR laser irradiation of the SWCNT/CpG DNA complexes were evaluated. In Fig. 2, 14 when a SWCNT/CpG DNA complex solution was irradiated with a NIR laser at 3 W/cm² (Fig. 15 2b), the temperature increased more rapidly than irradiation at 2 W/cm² (Fig. 2a). Additionally, the increases in temperature of the SWCNT/CpG DNA complex solution were dependent on the concentration of SWCNTs in solution and the NIR laser irradiation time. Moreover, the photohyperthermic effect of the SWCNT/CpG DNA complexes was equal to the photohyperthermic effect of SWCNT/GpC DNA complexes (in which the SWCNTs dispersed

 with GpC DNA exhibited no immunostimulatory effects), and SWCNT/Pluronic F-127 (in which the SWCNTs were dispersed with the surfactant, Pluronic F127) (Fig. 2c). These results suggest that the SWCNT/CpG DNA complexes also exhibit a thermal effect when irradiated with an NIR laser, thereby highlighting their potential use for photohyperthermic therapy.

 As the SWCNT/CpG DNA complexes released substantial vibrational energy after exposure to NIR light, the use of NIR irradiation for the induction of hyperthermia is particularly attractive because living tissues do not strongly absorb in this range (Weissleder, 2001). Therefore, a number of biomedical investigations of CNTs have focused on their application for the treatment of cancer (Liu et al., 2008a; Meng et al., 2008; Singh et al., 2005). In this study, the results of an in vitro photohyperthermic ablation of the murine colorectal cancer cell line, colon 26, treated with the SWCNT/CpG DNA complex, using NIR irradiation, were evaluated with a Live-Dead- Cell Kit. It was shown that most of the colon 26 cells treated with the SWCNT/CpG DNA 13 complex (1 μ g of SWCNTs) were dead after NIR laser irradiation (Fig. 3f), as compared with the 14 same cells treated with the SWCNT/CpG DNA complex without NIR laser irradiation (Fig. 3e). Meanwhile, the untreated (NT) cells (Fig. 3b) and naked CpG DNA treated cells (Fig. 3d) survived even after receiving NIR laser irradiation. These results suggested that the SWCNT/CpG DNA complex induced colon 26 cell cytotoxicity after NIR laser irradiation.

 Moreover, we also investigated the photohyperthermic effects of the SWCNT/CpG DNA complex in vivo by evaluating the temperature increases and the production of HSP70 in tumor

3.3 Immune effect of SWCNT/CpG DNA complexes in tumor-bearing mice

 CpG DNA can be taken up by macrophages and dendritic cells, leading to the production of Th1-type cytokines such as IFN-γ (Krieg, 2002). In our previous studies, we have demonstrated

11 In addition, compared with untreated mice or mice injected with naked CpG DNA or the SWCNT/GpC DNA complex, mice injected with the SWCNT/CpG DNA complex significantly 13 induced higher levels of TNF- α and IL-6 production both in serum (Fig. 5a and 5b) and in tumor tissue (Fig.6a and 6b) at 2 hours after injection. The SWCNT/CpG DNA complex also induced higher levels of IL-12p70 production in tumor tissues than compared with either naked CpG DNA or the SWCNT/GpC DNA complex (Fig. 5c). These results demonstrated that the 17 SWCNT/CpG DNA complex induced TNF- α and IL-6 cytokine production in tumor tissues and in serum, and may have the potential to inhibit tumor growth. As several papers have already demonstrated that CNTs are able to enter immune cells such as macrophages actively and

 passively via incomplete phagocytosis or by impaling the membrane (Cheng et al., 2009; Kam et al., 2005; Kraszewski et al., 2012), we considered that the SWCNT/CpG DNA complex may be more effectively recognized by the immune cells than naked CpG DNA. Moreover, our results suggest that the SWCNT/CpG DNA complex induced higher levels of cytokine production than naked CpG DNA, thereby confirming the reports that functionalized CNTs could enhance CpG DNA uptake and their immunostimulatory properties (Bianco et al., 2005b; Zhao et al., 2011). Furthermore, Fan et al. also has reproted that functionalized carbon nanotubes that conjugated with CpG DNA enhanced activity of CpG DNA and was effective for both brain and systemic melanomas (Fan et al., 2012).

 As some reports have shown that the photohyperthermic effect could enhance immune responses in vivo (Zhang et al., 2008; Gastpar et al., 2005), next, we investigated the immune effects of the SWCNT/CpG DNA complex after NIR laser irradiation. The results showed that there was no significant difference between the cytokine production levels of the SWCNT/CpG DNA complex and the SWCNT/CpG DNA complex + NIR laser irradiation (data not shown). This result may be because the photohyperthermic effects induced by the SWCNT/CpG DNA complex + NIR were either not enough to enhance the immune response, or the enhanced immune response was too low to be detected. We will investigate these details in the future.

 3.4 Enhancement of anti-tumor effect by SWCNT/CpG DNA complexes after NIR laser irradiation

 led to a more efficient antitumor effect than compared with that achieved by naked CpG DNA administration or SWCNT/GpC DNA complex administration.

 Several papers have reported that CNTs are effective in photohyperthermic therapy (Burke et al., 2009; Ghosh et al., 2009; Xiao et al., 2009). An advantage of NIR is that biological systems largely lack chromophores that absorb in this region, and as such, the NIR light should effectively and safely penetrate normal tissue and ablate any cells to which the CNTs are attached. Moreover, hyperthermic therapy has been clinically used in the management of solid tumors because it can synergistically enhance tumor cytotoxicity when combined with chemotherapy or radiotherapy (Falk and Issels, 2001; Wust et al., 2002). However, the combination of immunotherapy with CNTs is less well studied. In the present study, we successfully combined the photohyperthermic effects and immune effects together using the SWCNT/CpG DNA complex, and observed the resultant antitumor effect in vivo (Fig. 7). In practice, the toxicology of SWCNTs also needs to be addressed prior to using them in clinical situations. It has been found that well-functionalized CNTs, which are stable in physiological environments, are non-toxic to cells in vitro and non-toxic to mice in vivo (Liu et al., 2009; Liu et al., 2008; Schipper et al., 2008; Yang et al., 2008). Furthermore, Kagan et al. recently showed that hypochlorite and reactive radical intermediates of the human neutrophil enzyme, myeloperoxidase, enhance the biodegradation of SWCNTs in vitro, in neutrophils, and to a lesser

4. Conclusions

 We successfully prepared the SWCNTs / CpG DNA complexes, which were dispersed well and possessed both the photohyperthemic effects and the immune effects. The SWCNTs / CpG DNA complexes enhanced the production level of inflammatory cytokines in tumor tissues when intratumoral administration. We also have shown an effective combination cancer therapy using SWCNTs / CpG DNA complexes in mice. Intratumoral administration of SWCNTs / CpG DNA complexes with NIR irradiation prevented the proliferation of tumor growth in mice and prolonged the survival time of mice. These were suggested that combination of photohyperthermic therapy and immunotherapy would be an effective approach in cancer therapy.

Acknowledgments

- This work was supported in part by a Grant-in-Aid for Scientific Research (A) (23240072) and
- 3 by a fellowship grant from the Japan Society for the Promotion of Sciences (23•2743).

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FIGURES

 $1 + NIR$. (e) Colon 26 cells treated with the SWCNT/CpG DNA complex. (f) Colon 26 cells 2 treated with the SWCNT/CpG DNA complex + NIR. Results are representative of more than three experiments.

$\overline{4}$	Figure 4. Temperature increases and HSP70 production induced by NIR laser irradiation of the
5	SWCNT/CpG DNA complex in vivo. (a) Temperature increases in tumor tissues after NIR laser
6	irradiation. (b) HSP70 production induced by NIR laser irradiation of the SWCNT/CpG DNA
τ	complex in tumor tissues at different time points. * $P \le 0.05$ compared with SWCNT/CpG DNA
8	complex + NIR at 2 hours after NIR laser irradiation. ** $P < 0.01$ compared with SWCNT/CpG
9	DNA complex at 6 hours, and SWCNT/CpG DNA complex + NIR at 24 hours after NIR laser
10	irradiation. (c) HSP70 production induced by NIR laser irradiation of 5% dextrose, naked CpG
11	DNA, the SWCNT/CpG DNA complex or the SWCNT/GpC DNA complex in tumor tissue at 6
12	hours after NIR laser irradiation. * $P < 0.05$ compared with 5% dextrose and the SWCNT/CpG
13	DNA complex; ** $P < 0.01$ compared with 5% dextrose + NIR and naked CpG DNA + NIR. # P
14	< 0.05 compared with 5% dextrose + NIR, naked CpG DNA + NIR, and the SWCNT/CpG DNA
15	complex. Each value represents the mean $+ S$. D. (n = 5). Results are representative of three
16	experiments.

 Figure 5. Cytokine productions induced by the SWCNT/CpG DNA complex in tumor tissue. (a) 18 TNF- α production levels in tumor tissues after naked CpG DNA, SWCNT/CpG DNA complex or SWCNT/GpC DNA complex administration. * P < 0.05 compared with NT and SWCNT/GpC

18 CpG DNA + NIR; ## $P < 0.01$ compared with SWCNT/GpC DNA +NIR; ### $P < 0.001$

administration and NIR laser irradiation. # P < 0.05 compared with naked CpG DNA and naked

- ** P < 0.01 compared with NIR laser only; *** P< 0.001 compared with NT. Each value
- 3 represents the mean $+ S$. D. (n = 6). Results are representative of more than three experiments.

Tumor

Tumor

Days after Tumor Implantation (Day)