

## Effectiveness of Sirolimus in Combination with Cyclosporine against Chronic Rejection in a Pediatric Liver Transplant Patient

Haruka Shinke,<sup>a</sup> Sachiyo Hashi,<sup>a</sup> Risa Kinoshita,<sup>a</sup> Risa Taniguchi,<sup>a</sup> Mitsuhiro Sugimoto,<sup>a</sup> Kazuo Matsubara,<sup>a</sup> Eri Ogawa,<sup>b</sup> Mari Sonoda,<sup>b</sup> Narito Takada,<sup>b</sup> Atsushi Yoshizawa,<sup>b</sup> Kohei Ogawa,<sup>b</sup> Shinya Okamoto,<sup>b</sup> Shinji Uemoto,<sup>b</sup> and Satohiro Masuda<sup>\*,a</sup>

<sup>a</sup>Department of Pharmacy, Faculty of Medicine, Kyoto University Hospital; and <sup>b</sup>Divisions of Hepato-Pancreato-Biliary, Transplant and Pediatric Surgery, Department of Surgery, Kyoto University Hospital; Sakyo-ku, Kyoto 606–8507, Japan.

Received March 22, 2013; accepted May 10, 2013; advance publication released online May 15, 2013

The patient is a 3-year-old boy who received living-donor liver transplantation (LDLT) for hepatoblastoma, with his mother as the donor. Oral tacrolimus was started at a dose of 0.3 mg every 12 h from day 1, with the dosage adjusted on the basis of trough concentrations. The levels of aspartate aminotransferase (AST), alanine transferase (ALT), and total bilirubin (T-bil) were 110 U/L, 182 U/L, and 12.6 mg/dL, respectively, when chronic rejection (CR) was pathologically diagnosed. Then, sirolimus at a dose of 1.0 mg/d was added to the tacrolimus-based regimen. The T-bil level rapidly decreased to 5.4 mg/dL, without changes in AST and ALT. Because the intracellular receptor of sirolimus and tacrolimus is FK506-binding protein 12, we switched tacrolimus to cyclosporine at a dose of 60 mg/d to avoid competitive inhibition between these 2 drugs. The target trough concentration of sirolimus and cyclosporine was set to around 15 ng/mL and 180 ng/mL, respectively. The concentration/dose ratio of sirolimus was significantly correlated with the blood cyclosporine level ( $r=0.5293$ ,  $p<0.05$ ), suggesting the pharmacokinetic interaction between these 2 drugs. Thereafter, the levels of AST and ALT as well as the T-bil were successfully decreased to 73 U/L, 83 U/L, and 3.0 mg/dL, respectively. These results suggest that sirolimus therapy in combination with cyclosporine may be an effective treatment against CR after liver transplantation.

**Key words** liver transplantation; humoral rejection; immunosuppressant; tacrolimus; hepatoblastoma; chemotherapy

Living-donor liver transplantation (LDLT) with subsequent immunosuppressive therapy with tacrolimus is used to treat pediatric patients with end-stage liver disease. However, intensive therapeutic drug monitoring (TDM) is required to control the concentration of tacrolimus in the blood between the narrow therapeutic ranges, because of the large inter- and intra-individual variation of tacrolimus pharmacokinetics. In addition, several adverse effects of tacrolimus, such as neurotoxicity, kidney injury, and malignancy, have been raised as serious problems that need to be considered in pediatric patients.

Hepatoblastoma develops in children, until the age of 4 year, with highly elevated alpha-fetoprotein (AFP) levels of  $>100$  ng/mL.<sup>1,2)</sup> Several chemotherapeutic drugs such as anthracyclines, cisplatin, taxanes, or irinotecan have been evaluated as treatments against hepatoblastoma for patients who are contraindicated for surgical treatment.<sup>3,4)</sup> Liver transplantation has been considered an attractive optional treatment for patients with hepatoblastoma.<sup>5,6)</sup>

Although acute cellular rejection (ACR) after LDLT is curable with high-dose steroid pulse therapy, there is no effective treatment against chronic rejection (CR) mediated by humoral immunity. Tacrolimus and cyclosporine, common immunosuppressive agents, inhibit calcium-dependent T-cell activation.<sup>7)</sup> On the other hand, a mammalian target of rapamycin (mTOR) inhibitor acts on B-cells independently of its effects on helper T-cells, causing an inhibition of antigen- and cytokine-driven B-cell proliferation.<sup>8)</sup> Maintenance immunosuppression with mTOR inhibitors is reported to be associated with a significantly reduced risk of developing any posttransplant malig-

nancy.<sup>9)</sup> mTOR is acknowledged as a major player in cell proliferation. Recently, the mTOR inhibitors sirolimus (rapamycin) and everolimus have been used for cancer treatment.<sup>10,11)</sup> Elsharkawi *et al.*<sup>12)</sup> reported that sirolimus administration diminished the lung metastasis of hepatocellular carcinoma after liver transplantation. On the basis of these findings, mTOR inhibitors such as sirolimus and everolimus are considered to act as both anticancer agents and immunosuppressants.

In the present study, we examined the effectiveness of sirolimus against CR and the relapse of hepatoblastoma in a boy who received LDLT. CR was successfully controlled, without a relapse of hepatoblastoma, using sirolimus therapy in combination with cyclosporine rather than with tacrolimus.

### MATERIALS AND METHODS

**Case Report** The patient is a 3-year-old boy admitted to Kyoto University Hospital (Kyoto, Japan) with poorly differentiated hepatoblastoma and placed on the waiting list for liver transplantation. He was affected with chronic heart failure, chronic pulmonary disorder, bronchial asthma, chronic colon pseudo-obstruction and inguinal hernia. Before the transplantation, he was admitted in another hospital. Because of heart failure, the first-line therapy for hepatoblastoma, anthracycline antitumor drugs, could not be given to this patient. Therefore, he was treated with cisplatin (80 mg/sqm) biweekly. The tumor marker, alpha-fetoprotein (AFP), was effectively decreased from 602000 to 14900 ng/mL. However, because cisplatin-induced nephropathy occurred, the regimen was changed to weekly carboplatin (120 mg/sqm) 4 times. AFP was decreased further to 3000 ng/mL; however, the portal vein tumor throm-

The authors declare no conflict of interest.

\* To whom correspondence should be addressed. e-mail: masuda@kuhp.kyoto-u.ac.jp

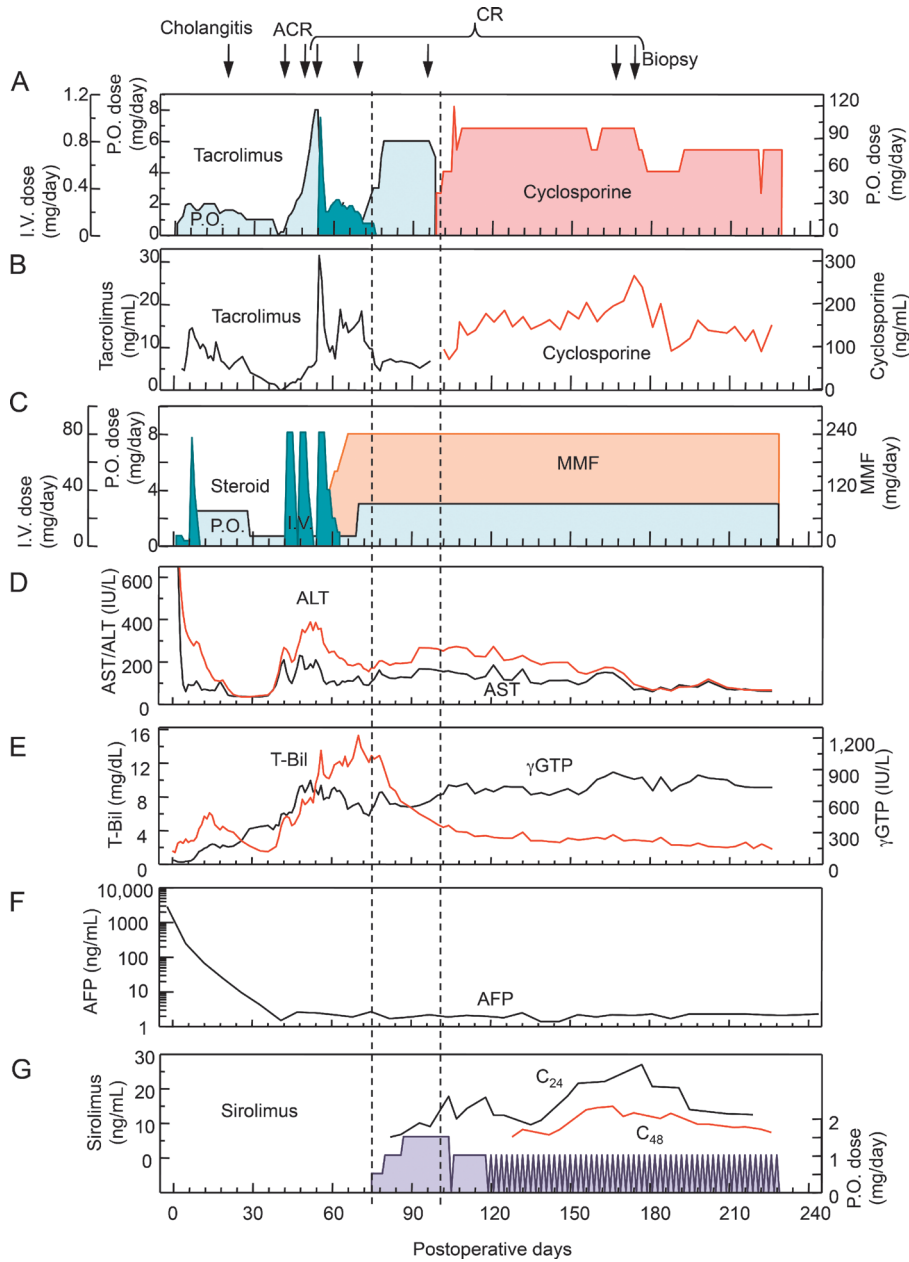


Fig. 1. Dosage (A) and Blood Concentration (B) of Calcineurin Inhibitors; Dosage of Steroid or Mycophenolate Mofetil (MMF) (C); Monitoring of Transaminases (D),  $\gamma$ -GTP or Total Billirubin (E), and  $\alpha$ -Fetoprotein (F); and Dosage and Blood Concentration of Sirolimus (G)

The trough levels of calcineurin inhibitors (B) were determined approximately 12h after the evening dose every day. The blood concentrations of tacrolimus and cyclosporine were measured with the CLIA and ACMIA methods, respectively. The blood concentration of sirolimus was measured by CLIA method. The dotted line shows the time points of the start and end of sirolimus therapy in combination with tacrolimus. ALT, alanine aminotransferase; AST, aspartate aminotransferase; T-bil, total bilirubin;  $\gamma$ -GTP, gamma-glutamyl transpeptidase; CR, chronic rejection; ACR, acute cellular rejection; P.O., per oral administration; I.V., intravenous administration; AFP, alpha-fetoprotein.

bus was not diminished. His physicians changed the regimen again to irinotecan (100mg/sqm) and etoposide (100mg/sqm) for 3d. Although the patient developed adverse effects of chemotherapy, such as myelosuppression, diarrhea, and sepsis, he was treated with a reduced dose of irinotecan (3.3mg/kg) and etoposide (3.3mg/kg) and/or carboplatin (2.6mg/kg) for 3d. Then, he was referred to our hospital in August 2009. His AFP level was 5110ng/mL; however, portal vein tumor thrombus persisted and metastasis to the lymph nodes was suspected. Therefore, he was not approved for liver transplantation. He was then treated with irinotecan for 6 months from August 2009 to February 2010 in the other hospital, and his AFP decreased to 700ng/mL. He was eventually admitted to

our hospital for LDLT. In June 2010, he underwent successful LDLT, with his 41-year-old mother (ABO blood type identical) as donor. He weighed 7665g and had a height of 74.5cm. The graft size was 190g, and the graft-to-recipient weight ratio (GRWR) was 2.53%. Oral administration of immunosuppressive agent tacrolimus (FK506) was started at a dose of 0.3mg every 12h with the target window around 10ng/mL, and its dose was adjusted on the basis of trough concentrations measured 12h after the evening dose (Figs. 1A,B). However, at postoperative day (POD) 43, ACR was observed on biopsy (Fig. 1A). He received steroid pulse therapy to control ACR, 3 times at POD 43, 48, and 53 (Fig. 1C). Intravenous injection of tacrolimus was also started at the time. The high

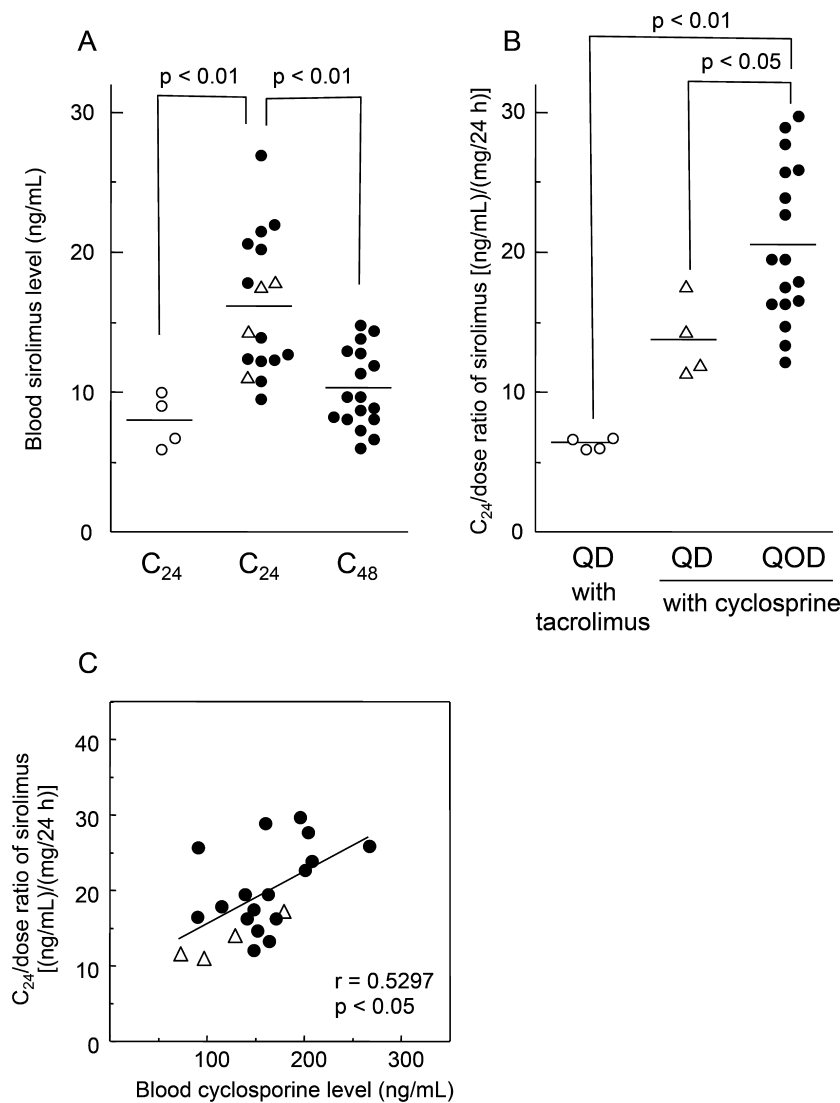


Fig. 2. The Blood Concentration of Sirolimus Was Elevated When Given in Combination with Cyclosporine

The dosage regimen of sirolimus was changed from QD to QOD because the calcineurin inhibitor was switched from tacrolimus to cyclosporine. The blood concentration of sirolimus (A) and the 24h concentration of sirolimus (ng/mL) per dosage (mg/d) ratio (B) were significantly elevated when given in combination with cyclosporine. (C) The blood cyclosporine level significantly correlated with the C<sub>24</sub>/dose ratio of sirolimus. Data are derived from Figs. 1B and 2B. White circle, QD sirolimus with tacrolimus; black circle, QOD sirolimus with cyclosporine; white triangle, QD sirolimus with cyclosporine.

levels of ALT and AST were decreased, while T-bil level was increased (Figs. 1D,E). Although the pathological findings related to ACR were diminished, CR was observed at POD 52. The levels of AST, ALT and T-bil were 110 U/L, 182 U/L, and 12.6 mg/dL, respectively. At POD 72, the findings related to CR were still positive. We started to administer the mTOR inhibitor sirolimus (rapamycin) orally with intravenous tacrolimus from postoperative day 75. The T-bil level rapidly decreased to 5.4 mg/dL without changes in AST and ALT. Therefore, treatment with sirolimus seems to be effective against CR. However, the gamma-glutamyl transpeptidase ( $\gamma$ -GTP) was slightly increased. At POD 96, he underwent biopsy and late-phase CR was diagnosed. Because the intracellular receptor of sirolimus and tacrolimus is FK506-binding protein 12 (FKBP1A), we switched tacrolimus to cyclosporine at a dose of 60 mg/day at postoperative day 100 to avoid competitive inhibition between sirolimus and tacrolimus.<sup>13)</sup> We thought cyclosporine was more effective against CR than tacrolimus in combination with sirolimus. The target trough concentration

of sirolimus and cyclosporine was set to around 15 ng/mL and 180 ng/mL, respectively. The blood concentration of sirolimus was elevated in combination with cyclosporine, but not with tacrolimus. At POD 177, when the final biopsy was performed, the levels of AST and ALT as well as T-bil were found to be successfully decreased to 73 U/L, 83 U/L, and 3.0 mg/dL, respectively. Thereafter, the liver function did not improve but remained stable. Finally, the patient was discharged without deterioration of liver function and relapse of the hepatoblastoma at POD 247.

**Measurement of the Blood Concentration of Immunosuppressant Drugs** The dose of tacrolimus and sirolimus was adjusted on the basis of trough concentrations measured 12h after the evening dose by means of the chemiluminescent enzyme immunoassay (CLIA) method (ARCHITECT™; Abbott Japan, Tokyo, Japan). The lower limit of this system is 0.5 ng/mL for tacrolimus or 1.0 ng/mL for sirolimus with a whole blood sample. The blood concentration of cyclosporine was measured by the antibody-conjugated magnetic immuno-

assay (ACMIA) method (Dimension™; Siemens Japan, Tokyo, Japan). The lower limit for cyclosporine is 25 ng/mL.

## DISCUSSION

Sirolimus treatment rapidly decreased the high level of T-bil. However, the levels of AST and ALT tended to increase when sirolimus was added to the tacrolimus-based regimen. Recently, Nielsen *et al.*<sup>14)</sup> reported that 4 and 6 of 12 pediatric patients with chronic graft dysfunction after liver transplantation developed completely normal liver function and showed partial response, respectively. However, there was no precise analysis in drug interaction between tacrolimus and everolimus. In the present study, both AST and ALT became well controlled after switching from tacrolimus to cyclosporine. Therefore, part of the patients with partial response of the past report<sup>14)</sup> might develop completely normal liver function when tacrolimus was switched to cyclosporine. These results suggested that sirolimus in combination with cyclosporine might be an effective treatment against CR after liver transplantation. That is to say that sirolimus is efficacious against CR, and cyclosporine is better calcineurin inhibitor compared to tacrolimus to obtain pharmacological effects of both mTOR inhibitor and calcineurin inhibitor for concomitant administration.

Several groups have shown that the mTOR pathway is required for the development and maturation of B-cells.<sup>15,16)</sup> It is also reported that mTOR inhibitors block B-cell development and antibody production.<sup>17)</sup> On the basis of these findings and the present experience, sirolimus treatment might be effective against CR mediated by humoral immunity.

The target sirolimus trough level of between 10 and 15 ng/mL was recommended for pediatric renal transplantation.<sup>18)</sup> However, the frequency of leukopenia was increased with the elevation of the sirolimus trough concentration to as high as 15 ng/mL.<sup>19)</sup> Nevertheless, we have reported the initial target trough concentration of sirolimus at 15 ng/mL in patients receiving islet transplantation.<sup>20,21)</sup> Based on these reports, we have set the initial target trough concentration of sirolimus at 15 ng/mL in this case.

The blood concentration of the immunosuppressive drugs used is shown in Fig. 2. The mean blood concentration of sirolimus, at 24 h after the treatment in combination with tacrolimus, was 8.0 ng/mL (range, 6–10.1 ng/mL). However, the blood concentration of sirolimus when given in combination with cyclosporine was significantly increased to 16.2 ng/mL (range, 9.6–27.0 ng/mL;  $p < 0.01$ ). The 24 h concentration of sirolimus (ng/mL) per dosage (mg/d) ratio was also significantly elevated in combination with cyclosporine ( $p < 0.01$ ). It is well acknowledged that sirolimus is mainly metabolized in the intestine and liver by cytochrome P450 3A4 (CYP3A4).<sup>22)</sup> In addition, the apparent  $IC_{50}$  values of tacrolimus and cyclosporine on for CYP3A4 inhibition were determined to be 53  $\mu$ M and 90  $\mu$ M, respectively.<sup>23)</sup> However, the target trough concentration of tacrolimus was approximately 10  $\mu$ g/mL and that of cyclosporine was approximately 180  $\mu$ g/mL in this case. Therefore, the inhibitory effect of cyclosporine on CYP3A4-mediated sirolimus metabolism would be stronger than that of tacrolimus in clinical use. Taking together these findings, the blood concentration of sirolimus could be expected to be elevated in combination with cyclosporine, but not with tacro-

limus. In addition, as Fig. 2C clearly indicates, the  $C_{24}$ /dose ratio of sirolimus correlates with blood cyclosporine concentration. These results clearly demonstrate the pharmacokinetic interaction between cyclosporine and sirolimus for the first time in a clinical liver transplant case.

Previously, it has been reported that sirolimus effectively inhibits hepatoblastoma growth both *in vitro* and *in vivo*.<sup>24)</sup> Therefore, in the present case, it is indicated that sirolimus treatment may not only be effective against CR but also contributes to the avoidance of hepatoblastoma relapse. In fact, since the treatment with sirolimus was started, the AFP levels remained markedly decreased to around 2.0 ng/mL (Fig. 1F), showing that the hepatoblastoma did not spread.

In summary, sirolimus therapy in combination with cyclosporine may be an effective treatment against CR after liver transplantation. Because of the competitive inhibition between tacrolimus and sirolimus, cyclosporine would be better than tacrolimus from use in combination with sirolimus. However, further investigation with more cases is required to confirm whether sirolimus treatment in combination with cyclosporine is effective against humoral immunity and hepatoblastoma.

**Acknowledgments** This work was supported in part by a Grant-in-Aid for Scientific Research (KAKENHI) from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan and by a Funding Program for Next Generation World-Leading Researchers (NEXT Program: LS073) initiated by the Council for Science and Technology Policy of the Japan Society for the Promotion of Science of Japan.

## REFERENCES

- 1) Darbari A, Sabin KM, Shapiro CN, Schwarz KB. Epidemiology of primary hepatic malignancies in U.S. children. *Hepatology*, **38**, 560–566 (2003).
- 2) Van Tornout JM, Buckley JD, Quinn JJ, Feusner JH, Krailo MD, King DR, Hammond GD, Ortega JA. Timing and magnitude of decline in alpha-fetoprotein levels in treated children with unresectable or metastatic hepatoblastoma are predictors of outcome: a report from the Children's Cancer Group. *J. Clin. Oncol.*, **15**, 1190–1197 (1997).
- 3) Hishiki T, Matsunaga T, Sasaki F, Yano M, Ida K, Horie H, Kondo S, Watanabe K, Oue T, Tajiri T, Kamimatsuse A, Ohnuma N, Hi-yama E. Outcome of hepatoblastomas treated using the Japanese Study Group for Pediatric Liver Tumor (JPLT) protocol-2: report from the JPLT. *Pediatr. Surg. Int.*, **27**, 1–8 (2011).
- 4) Lieber J, Ellerkamp V, Wenz J, Kirchner B, Warmann SW, Fuchs J, Armeanu-Ebinger S. Apoptosis sensitizers enhance cytotoxicity in hepatoblastoma cells. *Pediatr. Surg. Int.*, **28**, 149–159 (2012).
- 5) Koneru B, Flye MW, Busuttil RW, Shaw BW, Lorber MI, Emond JC, Kalayoglu M, Freese DK, Starzl TE. Liver transplantation for hepatoblastoma. The American experience. *Ann. Surg.*, **213**, 118–121 (1991).
- 6) Otte JB, Pritchard J, Aronson DC, Brown J, Czauderna P, Maibach R, Perilongo G, Shafford E, Plaschkes J, International Society of Pediatric Oncology (SIOP). Liver transplantation for hepatoblastoma: results from the International Society of Pediatric Oncology (SIOP) study SIOPEL-1 and review of the world experience. *Pediatr. Blood Cancer*, **42**, 74–83 (2004).
- 7) Flanagan WM, Corthésy B, Bram RJ, Crabtree GR. Nuclear association of a T-cell transcription factor blocked by FK-506 and cyclosporin A. *Nature*, **352**, 803–807 (1991).

- 8) Aagaard-Tillery KM, Jelinek DF. Inhibition of human B lymphocyte cell cycle progression and differentiation by rapamycin. *Cell Immunol.*, **156**, 493–507 (1994).
- 9) Kauffman HM, Cheriakh WS, Cheng Y, Hanto DW, Kahan BD. Maintenance immunosuppression with target-of-rapamycin inhibitors is associated with a reduced incidence of *de novo* malignancies. *Transplantation*, **80**, 883–889 (2005).
- 10) Atkins MB, Hidalgo M, Stadler WM, Logan TF, Dutcher JP, Hudes GR, Park Y, Liou SH, Marshall B, Boni JP, Dukart G, Sherman ML. Randomized phase II study of multiple dose levels of CCI-779, a novel mammalian target of rapamycin kinase inhibitor, in patients with advanced refractory renal cell carcinoma. *J. Clin. Oncol.*, **22**, 909–918 (2004).
- 11) Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, Grünwald V, Thompson JA, Figlin RA, Hollaender N, Urbanowitz G, Berg WJ, Kay A, Lebowitz D, Ravaud A, RECORD-1 Study Group. Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. *Lancet*, **372**, 449–456 (2008).
- 12) Elsharkawi M, Staib L, Henne-Bruns D, Mayer J. Complete remission of posttransplant lung metastases from hepatocellular carcinoma under therapy with sirolimus and mycophenolate mofetil. *Transplantation*, **79**, 855–857 (2005).
- 13) Bierer BE, Mattila PS, Standaert RF, Herzenberg LA, Burakoff SJ, Crabtree G, Schreiber SL. Two distinct signal transmission pathways in T lymphocytes are inhibited by complexes formed between an immunophilin and either FK506 or rapamycin. *Proc. Natl. Acad. Sci. U.S.A.*, **87**, 9231–9235 (1990).
- 14) Nielsen D, Briem-Richter A, Sornsakrin M, Fischer L, Nashan B, Ganschow R. The use of everolimus in pediatric liver transplant recipients: first experience in a single center. *Pediatr. Transplant.*, **15**, 510–514 (2011).
- 15) Lazorchak AS, Liu D, Facchinetti V, Di Lorenzo A, Sessa WC, Schatz DG, Su B. Sin1-mTORC2 suppresses rag and il7r gene expression through Akt2 in B cells. *Mol. Cell*, **39**, 433–443 (2010).
- 16) Benhamron S, Tirosh B. Direct activation of mTOR in B lymphocytes confers impairment in B-cell maturation and loss of marginal zone B cells. *Eur. J. Immunol.*, **41**, 2390–2396 (2011).
- 17) Zhang S, Readinger JA, DuBois W, Janka-Junttila M, Robinson R, Pruitt M, Bliskovsky V, Wu JZ, Sakakibara K, Patel J, Parent CA, Tessarollo L, Schwartzberg PL, Mock BA. Constitutive reductions in mTOR alter cell size, immune cell development, and antibody production. *Blood*, **117**, 1228–1238 (2011).
- 18) Kahan BD. The potential role of rapamycin in pediatric transplantation as observed from adult studies. *Pediatr. Transplant.*, **3**, 175–180 (1999).
- 19) Hong JC, Kahan BD. Sirolimus-induced thrombocytopenia and leukopenia in renal transplant recipients: risk factors, incidence, progression, and management. *Transplantation*, **69**, 2085–2090 (2000).
- 20) Matsumoto S, Okitsu T, Iwanaga Y, Noguchi H, Nagata H, Yonekawa Y, Yamada Y, Fukuda K, Tsukiyama K, Suzuki H, Kawasaki Y, Shimodaira M, Matsuoka K, Shibata T, Kasai Y, Maekawa T, Shapiro J, Tanaka K. Insulin independence after living-donor distal pancreatectomy and islet allotransplantation. *Lancet*, **365**, 1642–1644 (2005).
- 21) Sato E, Shimomura M, Masuda S, Yano I, Katsura T, Matsumoto S, Okitsu T, Iwanaga Y, Noguchi H, Nagata H, Yonekawa Y, Inui K. Temporal decline in sirolimus elimination immediately after pancreatic islet transplantation. *Drug Metab. Pharmacokinet.*, **21**, 492–500 (2006).
- 22) Gallant-Haidner HL, Trepanier DJ, Freitag DG, Yatscoff RW. Pharmacokinetics and metabolism of sirolimus. *Ther. Drug Monit.*, **22**, 31–35 (2000).
- 23) Haehner T, Refaie MO, Müller-Enoch D. Drug–drug interactions evaluated by a highly active reconstituted native human cytochrome P4503A4 and human NADPH-cytochrome P450 reductase system. *Arzneimittelforschung*, **54**, 78–83 (2004).
- 24) Wagner F, Henningsen B, Lederer C, Eichenmüller M, Gödeke J, Müller-Höcker J, von Schweinitz D, Kappler R. Rapamycin blocks hepatoblastoma growth *in vitro* and *in vivo* implicating new treatment options in high-risk patients. *Eur. J. Cancer*, **48**, 2442–2450 (2012).