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1 **Cellulase Activity in Meiobenthos in Wetlands**

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12

13 **Abstract**

14 To validate the involvement of meiobenthos in cellulose breakdown in wetlands,  
15 meiobenthos were collected from the sediments of Lake Furen and the Biwase River in  
16 Hokkaido prefecture, the Kako River in Hyogo prefecture, and the Chinai River in  
17 Shiga prefecture. Cellulase activities of the meiobenthos were measured by cellulose  
18 zymographic analysis using SDS-PAGE gels containing 0.5% carboxymethyl cellulose.  
19 The results showed that most of the Turbellaria, Nematoda, Harpacticoida and  
20 Oligochaeta species exhibited cellulase activity. The molecular sizes of the  
21 cellulase-active bands of the sediments in Lake Furen, the Biwase River, and the Chinai  
22 River coincided with those of meiobenthos. The findings suggest that meiobenthos  
23 might play a major function in cellulose breakdown in these wetlands. This paper is the  
24 first to report cellulase activity in meiobenthos and that they are possibly involved in the  
25 breakdown of cellulose in wetlands.

26 **KEY WORDS:** cellulase • cellulose • meiobenthos • sediment • wetland

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28

29 **Introduction**

30

31 Cellulose, the most abundant organic matter on earth, is a high molecular weight  
32 substance consisting of glucose residues bound by  $\beta$ -1,4 linkages, unlike starch, another  
33 glucan consisting of a  $\alpha$ -1,4 linked glucose residues. Cellulose is resistant to enzyme  
34 degradation [1, 2]. However, cellulose can be degraded by specific enzymes collectively  
35 named cellulases [3].

36 Fungi and bacteria as well as symbiotic protozoa in herbivorous animals and  
37 termites have been studied as known consumers of cellulose [4]. Recently, an intrinsic  
38 cellulase gene was isolated from termite [5], and a variety of intrinsic cellulase genes  
39 were identified from various animals, including beetle [6], nematode [7, 8], abalone [9],  
40 mussel [10], sea urchin [11], and brackish-water clam [12].

41 Recent stable isotopic analysis showed that a brackish water clam *Corbicula*  
42 *japonica* consumes land-derived organic materials mainly composed of cellulose [13].  
43 Identification of the intrinsic cellulase gene and immunological detection of the enzyme  
44 protein in *C. japonica* strongly suggest that *C. japonica* plays an important role in the  
45 process of degradation of cellulose in rivers [14].

46 Besides macrobenthos such as *C. japonica*, a group of small animals called  
47 meiobenthos also inhabits the sediments of aquatic areas. Meiobenthos are defined as

48 animals that pass through a 1-mm mesh filter and are known to be composed of a  
49 variety of fauna corresponding to 22 phyla [15].

50 In the present study, we attempted to validate the role of meiobenthos in the  
51 process of breakdown of cellulose in wetlands. We chose Lake Furen and the Biwase  
52 River located in the subarctic area, since they were reported to be the wetlands  
53 demonstrating the highest cellulase activities in Japan [16]. On the other hand, we chose  
54 the Kako River and the Chinai River as typical rivers in temperate area. We also  
55 expected the difference in distribution of meiobenthos between the Kako River and the  
56 Chinai River, because the Chinai River is a fresh water river. We report that  
57 meiobenthos have cellulase activity and possibly play substantially important roles in  
58 the cellulose degradation process in the sediments of some wetlands.

59

## 60 **Materials and Methods**

61

### 62 **Materials**

63

64 We collected sediments from Lake Furen and the Biwase River in Hokkaido prefecture,  
65 the Kako River in Hyogo prefecture, and the Chinai River in Shiga prefecture. River

66 sediments from all rivers were collected in the wetland within 50 m from the river  
67 mouth. From each site, we collected sediment samples from a 5-cm depth at low tide  
68 from May 2006 to October 2007. The sediment samples were transported to the  
69 laboratory in Kyoto University at 4°C, and meiobenthos were recovered in the fraction  
70 that included material small enough to pass through a 1mm-mesh filter but too large to  
71 pass through a 40 µm-mesh filter. These meiobenthos were classified under microscopic  
72 observation according to Higgins and Thiel [15]. About 500 g of the sediments were  
73 filtered through 1 mm-mesh. Sediment samples that were not filtered were designated  
74 “total sediment fraction,” while sediments less than 1 mm and larger than 40 µm were  
75 collected and designated “meiobenthos fraction.”

76

77 Measurement of cellulase activity

78

79 Meiobenthos were separated from the sediments by using a pair of tweezers under the  
80 microscope (Olympus, S2X12, Tokyo), and each single meiobenthos was homogenized  
81 with 20 µl of phosphate-buffered saline (PBS) containing 140 mM NaCl, 2.7 mM KCl,  
82 8 mM Na<sub>2</sub>HPO<sub>4</sub>, and 1.5 mM KH<sub>2</sub>PO<sub>4</sub> (pH 7.4) to prepare a meiobenthos extract. The  
83 total sediment and meiobenthos fractions were homogenized with 1.5-fold volume of

84 PBS, and the supernatants were obtained by centrifugation at  $10,000 \times g$  for 10 min.  
85 Ten microgram of the supernatant was applied on cellulase zymographic analysis.  
86 Cellulase zymographic analysis was performed as described previously by using 7.5%  
87 or 10% SDS-PAGE gel containing 0.5% carboxymethyl cellulose (CMC, Sigma, St  
88 Louis, MO, US). After electrophoresis, the gels were soaked in 10 mM acetate buffer  
89 (pH 5.5) containing 0.1% TritonX-100 for 30 min to remove SDS from the gels. The  
90 gels were transferred to 10 mM acetate buffer (pH 5.5), incubated at 37°C overnight,  
91 and then stained with 0.1% Congo Red. The gels were destained using 1 M NaCl. The  
92 active bands were detected as non-stained bands. Unless otherwise specified, special  
93 grades of reagents were commercially obtained from Nacalai Tesque (Kyoto). Protein  
94 concentration was measured according to the method of Bradford (17).

95

## 96 **Results**

97

### 98 Distribution of meiobenthos

99

100 Oligochaeta species were the dominant meiobenthos in Lake Furen, where the  
101 sediments are mainly composed of sand. In addition, a variety of Turbellaria, Nematoda,

102 Harpacticoida species were also observed. Oligochaeta species were also dominant in  
103 the Chinai River, where the sediments were, like in Lake Furen, mainly composed of  
104 sand.

105 In the Biwase and Kako Rivers, where the sediments were mainly composed of  
106 clay, Nematoda and Harpacticoida species were dominantly observed.

107

108 Cellulase activity of sediments and meiobenthos

109

110 Cellulase activity in meiobenthos extracts, total sediment fractions, and meiobenthos  
111 fractions were measured by cellulose zymography.

112 Cellulase activity bands of the total sediment fraction were detected between  
113 32.5 kDa and 47.5 kDa in samples from Lake Furen, which coincided with those of the  
114 meiobenthos fraction (lanes 1 and 2 in Fig. 1). Oligochaeta species assayed by using a  
115 single animal showed an active band corresponding to 32.5 kDa, which is the band size  
116 observed for the total sediment fraction and the meiobenthos fraction (lane 3 in Fig. 1).

117 Different species of Nematoda showed active bands of different molecular sizes, as  
118 shown in lanes 4–7 of Fig. 1. Interestingly, common active bands at 25 kDa were  
119 detected for all the Nematoda species. Various active bands were also detected for

Fig.1



120 Turbellaria species (lane 8 in Fig. 1), but their sizes differed from those of Nematoda  
121 species.

122 Figure 2 shows the cellulase activity in sediment samples corresponding to  
123 approximately 5 mg per one lane from the Biwase River. Intensive active bands were  
124 observed, especially above 47.3 kDa in the total sediment fraction and the meiobenthos  
125 fraction (lanes 1 and 2 in Fig. 2), and the active band patterns were nearly identical.  
126 Harpacticoida species, the dominant organisms of meiobenthos in the Biwase River,  
127 also exhibited remarkably intensive activity bands of above 47.3 kDa (lane 3 in Fig. 2).

Fig.2

128 Figure 3 shows the active bands of the samples from the Chinai River. Faint  
129 active bands were observed at 25 kDa and between 47.5 kDa and 62 kDa in the total  
130 sediment fraction and the meiobenthos fraction (lanes 1 and 2 in Fig. 3). Bands with  
131 nearly the same activity were observed for Oligochaeta species, as shown in lane 3.

Fig.3

132 Figure 4 shows the active bands in the Kako River samples. In the total  
133 sediment fraction and the meiobenthos fraction, faint bands of less than 25 kDa were  
134 observed (lanes 1 and 2 in Fig. 4). In lane 3, active bands of Harpacticoida species were  
135 observed at approximately 47.5 kDa, which did not coincide with the active bands in the  
136 total sediment fraction and the meiobenthos fraction.

Fig.4

137

138 **Discussion**

139 Cellulase activities were detected in the extracts of the meiobenthos collected from all  
140 the sampling sites examined, suggesting that meiobenthos may be involved in the  
141 breakdown of cellulose in the sediments. Interestingly, 25 kDa active bands were  
142 commonly detected in the extracts of morphologically distinct Nematoda species  
143 collected from Lake Furen (lanes 4–7 in Fig. 1), suggesting that Nematoda species  
144 possibly share a related cellulase gene. On the other hand, Oligochaeta species collected  
145 from Lake Furen and the Chinai River were morphologically distinct and the band sizes  
146 of the cellulase were also different (compare lane 3 in Fig. 1 and lane 3 in Fig. 3).

147           The International Collaborative Research on the Management of Wetland  
148 Ecosystem of the National Institute for Environmental Studies [16] reported the  
149 outstanding strong cellulase activities of sediments collected from Lake Furen and the  
150 Biwase River in Hokkaido among many Japanese wetlands tested. The report attributed  
151 the strong cellulase activity in the sediments of Lake Furen and the Biwase River to  
152 microorganisms including bacteria and fungus. However, the active bands in cellulose  
153 zymographic analyses showed that the position of the active bands coincided with the  
154 sediment fractions and the extracts of meiobenthos (Figs. 1 and 2), which supported the

155 hypothesis that meiobenthos might be involved in the breakdown of cellulose in Lake  
156 Furen and the Biwase River.

157           Because recent molecular biological studies suggest the endogenous origin of  
158 the cellulase genes in aquatic invertebrates [9, 11, 12], cellulase genes of meiobenthos  
159 could be encoded in the DNA of meiobenthos themselves. We are now trying to clone  
160 the cellulase genes of meiobenthos to validate the possibility of its endogenous origin.

161           As shown in Fig. 3, active bands of the total sediment fraction and the  
162 meiobenthos fraction coincided with those of Oligochaeta species from the Chinai River.  
163 On the other hand, active bands of the total sediment fraction and the meiobenthos  
164 fraction did not coincide with those of the Harpacticoida species from the Kako River,  
165 as shown in Fig. 4. Thus, the origin of cellulase could not be concluded to be the  
166 meiobenthos in the case of Kako River sediment. Further studies are required to  
167 evaluate the contribution of meiobenthos to the breakdown of cellulose in wetlands in  
168 the temperate area.

169           The contribution of termites to the breakdown of cellulose in the forests of  
170 tropical zones is assumed to correspond to 80% the total cellulose breakdown in this  
171 area [18]. Like termites, meiobenthos could be major consumers of cellulose, especially  
172 in some wetlands in Hokkaido, because cellulase activity of meiobenthos in Lake Furen

173 and the Biwase River were detected at 4°C (data not shown), which is a temperature at  
174 which the growth of bacteria and fungi would be suppressed. Therefore, it seems  
175 probable that meiobenthos would play important roles in cellulose degradation  
176 especially in low temperature environments like wetlands in Hokkaido.

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178

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- 231

232 Figure legends

233

234 **Fig. 1.** Cellulase activities in Lake Furen. Activities were detected in 10% SDS-PAGE  
235 gel containing 0.5% carboxymethyl cellulose. The positions for molecular mass marker  
236 proteins are shown by arrows. Lanes: 1, total sediment fraction; 2, meiobenthos  
237 fraction; 3, Oligochaeta species; 4–7, morphologically distinct species of Nematode; 8,  
238 Turbellaria species.

239

240 **Fig. 2.** Cellulase activities in the Biwase River. Ten percent SDS-PAGE gel containing  
241 0.5% carboxymethyl cellulose was used for the detection of cellulase bands of the  
242 sediments, while 7.5% gel was used for Harpacticoida species. The positions for  
243 molecular mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction;  
244 2, meiobenthos fraction; 3, Harpacticoida species.

245

246 **Fig. 3.** Cellulase activities in the Chinai River. Activities were detected in 10%  
247 SDS-PAGE gel containing 0.5% carboxymethyl cellulose. The positions for molecular  
248 mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction; 2,  
249 meiobenthos fraction; 3, Oligochaeta species.

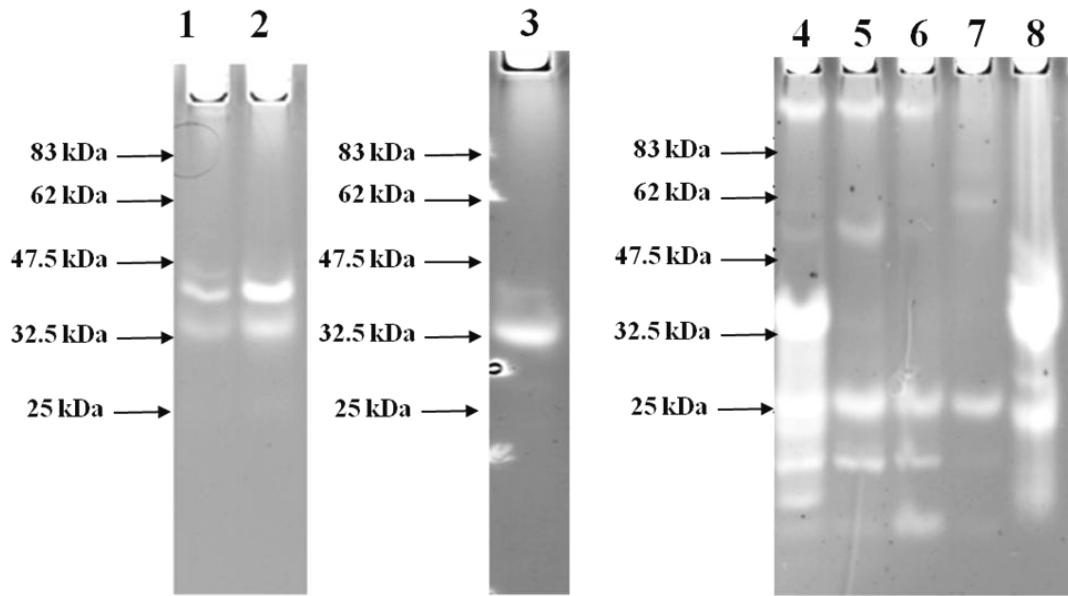


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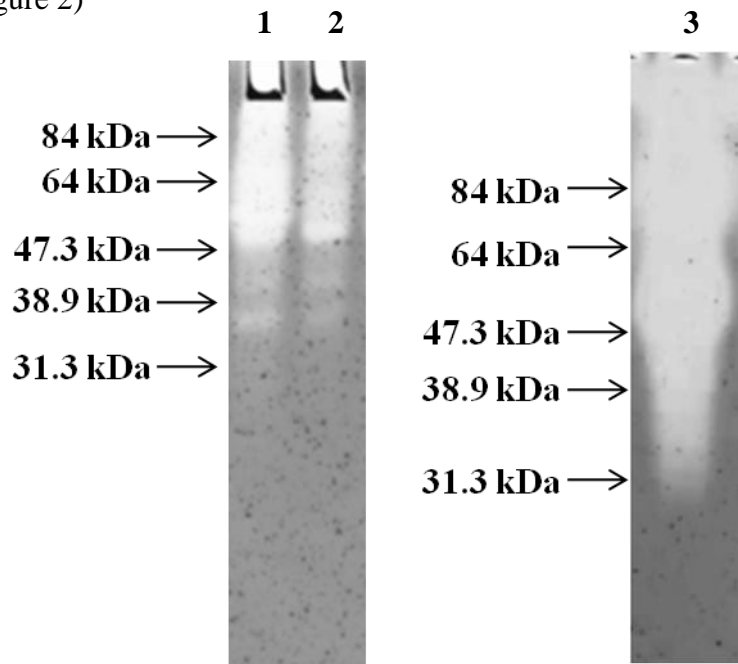
251 **Fig. 4.** Cellulase activities in the Kako River. Activities were detected in 10%  
252 SDS-PAGE gel containing 0.5% carboxymethyl cellulose. The positions for molecular  
253 mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction; 2,  
254 meiobenthos fraction; 3, Harpacticoida species.

255

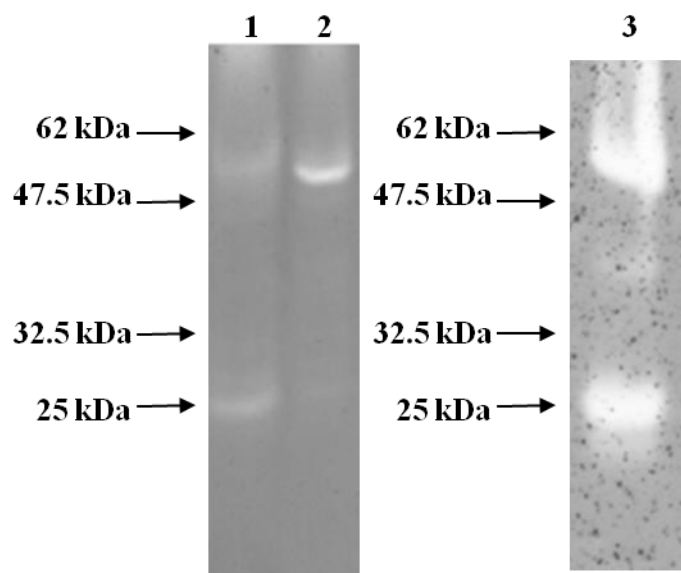
(Figure 1)



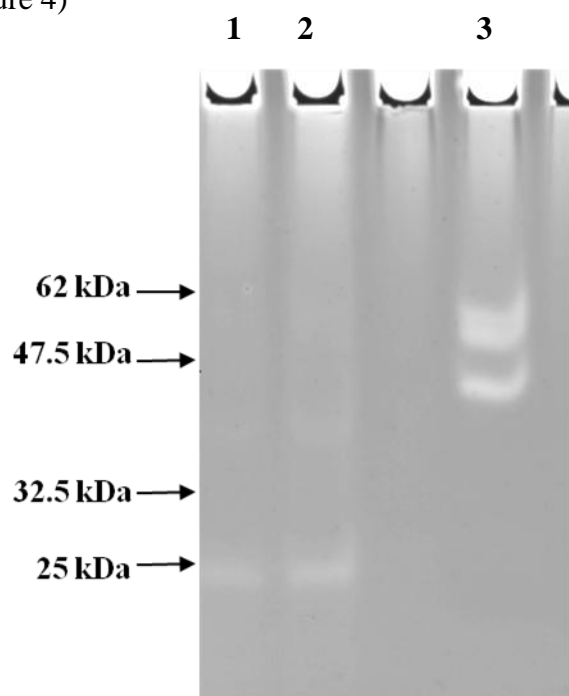
(Figure 2)



(Figure 3)



(Figure 4)



## 和文要旨

湿地帯に生息するメイオベントスのセルラーゼ活性

豊原治彦，朴 煥華，土屋佳奈子，劉 文（京大院農）

湿地帯におけるセルロース分解に関わるメイオベントスの役割を明らかにする目的で，琵琶瀬川と風連湖（北海道），知内川（滋賀県），加古川（兵庫県）の底泥に生息するメイオベントスのセルラーゼ活性を測定し，ほとんどのメイオベントスに活性を認めた。とくに風蓮湖，琵琶瀬川および知内川ではザイモグラフィー分析により，底泥とメイオベントスの活性バンドのサイズが一致したことから，これらの湿地帯においてはセルロース分解にメイオベントスが主要な役割を果たしていると考えられた。