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RESEARCH ARTICLE

Staphylococcus aureus secreted lipases do not inhibit innate immune killing mechanisms [version 1; peer review: 2 approved, 1 approved with reservations]

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Abstract

Background: *Staphylococcus aureus* causes an array of diseases in both humans and livestock. Pathogenesis is mediated by a plethora of proteins secreted by *S. aureus*, many of which remain incompletely characterised. For example, *S. aureus* abundantly secretes two isoforms of the enzyme lipase into the extracellular milieu, where they scavenge upon polymeric triglycerides. It has previously been suggested that lipases may interfere with the function of innate immune cells, such as macrophages and neutrophils, but the impact of lipases on phagocytic killing mechanisms remains unknown.

Methods: We employed the epidemic *S. aureus* clone USA300 strain LAC and its lipase deficient isogenic mutant, along with recombinant lipase proteins, in *in vitro* experimental infection assays. To determine if lipases can inhibit innate immune killing mechanisms, the bactericidal activity of whole blood, human neutrophils, and macrophages was analysed. In addition, gentamycin protection assays were carried out to examine the influence of lipases on *S. aureus* innate immune cell escape.

Results: There were no differences in the survival of *S. aureus* USA300 LAC wild type and its lipase-deficient isogenic mutant after incubation with human whole blood or neutrophils. Furthermore, there was no detectable lipase-dependent effect on phagocytosis, intracellular survival, or escape from both human primary and immortalised cell line macrophages, even upon supplementation with exogenous recombinant lipases.

Conclusions: *S. aureus* lipases do not inhibit bacterial killing mechanisms of human macrophages, neutrophils, or whole blood. These findings broaden our understanding of the interaction of *S. aureus* with the innate immune system.

Keywords

Staphylococcus, lipase, neutrophils, macrophages, innate immune cells

Open Peer Review

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Introduction

The Gram-positive bacterium *Staphylococcus aureus* is the cause of an array of nosocomial and community-acquired infections. To be a successful pathogen, *S. aureus* must evade killing by the innate immune system which it does via a large number of secreted factors. Within the *Staphylococcus* genus, a lipase-encoding gene (*lip1*) is present in at least 12 species, and a second lipase gene is present in *S. aureus* (*lip2*) and *S. epidermidis* (*gehD*)^{1,2}. *S. aureus* lipases are glycerol-ester hydrolases that cleave triglyceride lipids, resulting in the release of glycerol derivatives and free fatty acids¹. Lipase 1 has an affinity for short-chain fatty acids, whereas lipase 2 has no bias towards chain length¹. Transcription of lipase genes is regulated by the accessory gene regulator (*agr*) two component system, leading to the expression of a pre-pro-lipase precursor that is secreted into the extracellular milieu^{1,3}. The catalytic activity of lipases is regulated through downstream processing by the secreted zinc metalloprotease, Aur, which proteolytically cleaves the pre-pro precursor enzyme resulting in the mature, active form of the enzyme⁴. The activity of the mature lipase is governed by a catalytic triad, which cleaves glycerol-ester bonds through a serine hydrolase mechanism^{1,2}. Lipases have been reported to account for approximately 20% of the total *S. aureus* secretome, but our understanding of the role of lipases in host-pathogen interactions is limited⁵. It has been shown that 80% of clinical isolates from both systemic and localised *S. aureus* infections exhibit lipolytic activity, and patients typically test positive for anti-lipase IgG in serum^{6,7}. Lipases have further been attributed to the formation of biofilm, which subsequently confers resistance to toxic polyamines thus promoting bacterial persistence^{3,8,9}. Previous studies have further demonstrated that lipases can produce free-fatty acids from host lipid metabolites, such as low-density lipoproteins, which subsequently are incorporated into the lipid moieties of *S. aureus*¹⁰. The incorporation of lipoprotein particles has been shown to render the bacterium resistant to the antimicrobial drug triclosan, which is commonly used in the treatment of *S. aureus* infection¹⁰.

In a previous study, human granulocytes were treated with *S. aureus* lipases resulting in the loss of microvilli, projections, and pseudopodia on their surface suggesting a potential impact on phagocytosis or neutrophil extracellular trap (NET) formation^{11,12}. More recently, it was demonstrated that lipase 2 interferes with macrophage signalling, which subsequently diminishes the downstream pro-inflammatory response¹³. Specifically, lipase 2 inactivates *S. aureus* secreted lipoproteins, which are a major pattern-associated molecular pattern recognised by Toll-like receptor 2 (TLR2) in response to *S. aureus* infection¹³.

Macrophages are equipped with an array of pathogen recognition receptors and, alongside a role in modulation of cellular signalling, are professional phagocytes that aid in the clearance *S. aureus*^{14,15}. However, studies have shown that once entrapped within the macrophage phagolysosome, *S. aureus* can subvert killing mechanisms and persist for several days^{16,17}. The subsequent death of the macrophage through membrane blebbing and caspase-3 activation results in the release of viable bacteria, promoting intra-host dissemination in a Trojan horse-like system^{14,17,18}.

Here, we tested the hypothesis that lipases can interfere with the antibacterial activity of whole blood, neutrophils and macrophages. We report that, despite their abundant secretion, lipases have no effect on killing, phagocytosis, intracellular survival or escape of *S. aureus* USA300 LAC.

Methods

Bacterial growth conditions

40% (v/v) glycerol stocks of both a wild type (*S. aureus* USA300 WT) and an isogenic mutant (*S. aureus* USA300 Δ *lip1*/ Δ *lip2*) of the CC8 epidemic clone *S. aureus* USA300 LAC generated in a previous study¹⁹ were stored at -80°C. When required, stocks were sub-cultured onto tryptone soy agar (TSA, Oxoid CM131B) or cultured into tryptone soy broth (TSB, Oxoid CM129B) overnight at 37°C with agitation (200 rpm). The culture was diluted 1 in 100 in TSB and incubated, until exponential phase (OD_{600} =0.6–0.8), as measured using an Amersham Biosciences Ultrospec 2100 pro spectrophotometer. For infection protocols, bacteria were washed in cell culture media and suspended at the required OD_{600} .

Purification of recombinant lipase1 (rLip1) and 2 (rLip2)

Expression plasmid constructs pET156:*lip1* or pET156:*lip2*¹⁹ were transformed into ClearColi® BL21 (DE3) electrocompetent cells (Lucigen, 60810-1) by electroporation, according to the manufacturer's instructions. Cells were grown in LB Miller broth (Sigma, L3522-250G) to an OD_{600} of 0.6 and protein expression was induced with 1 mM isopropyl β -D-1-thiogalactopyranoside (IPTG, Formedium Ltd, IPTG025) for 4 h at 37°C, with agitation (200 rpm), before centrifugation and storage at -20°C.

Hexa-histidine tagged proteins were purified by immobilised metal affinity chromatography as described previously¹⁹. Western blot analysis confirmed the presence of hexa-histidine tagged proteins at 76 kDa (1 in 10,000 monoclonal anti-poly His, α -diagnostics HISP12-HRP, in 8% (w/v) skimmed milk (Sigma, 70166-500G) in sterile phosphate buffered saline (PBS)). Primary antibody binding was detected using enhanced luminol-based chemiluminescent (ECL) western blotting substrate (GE Healthcare, RPN2232). For lipopolysaccharide (LPS) removal, 1 ml of Pierce high capacity endotoxin removal resin (Thermo Fisher Scientific, 88271) was used according to the manufacturer's instructions and proteins were quantified using a bicinchoninic acid (BCA) assay (Merck millipore, 71285-3). To analyse recombinant protein lipolysis, a turbidimetric assay was used following the methodology outlined previously²⁰. For each of the following experiments, 200 nM of recombinant lipase 1 (rLip1) or 2 (rLip2) was used, according to previous estimates of lipase secretion levels by *S. aureus*²¹.

Ethics statement

Human blood was obtained from healthy volunteers in syringes treated with anticoagulant citrate dextrose. Ethical approval for the collection of blood from anonymous donors was granted by the University of Edinburgh Research Ethics Committee. This study was reviewed by the University Of Edinburgh College Of Medicine Ethics Committee (2009/01) and subsequently renewed by the Lothian Research Ethics Committee

(11/AL/0168). Written informed consent was received from all volunteers participating in the study.

Bacterial killing by neutrophils

Neutrophils were purified from human blood using a ficoll gradient. Briefly, 10 ml of 1.077 g/mol ficoll paque plus (Fisher, 11778538) was gently layered onto 12 ml of 1.119 g/mol Histopaque plus (Sigma, 11191). Fresh human blood was diluted at a 1:1 ratio in Ca^{2+} and Mg^{2+} free PBS (Lonza, BE17-515F), then slowly pipetted onto the ficoll gradient prior to centrifugation for 20 min at $400 \times g$ (without a brake). The neutrophil layer was collected, cells were centrifuged and erythrocytes lysed by osmotic shock. Cells were suspended in RPMI-1640 (Sigma, R5886), 0.05% (v/v) human serum albumin (Sigma, A9080-10ML) and 1% (v/v) GlutaMAX (Gibco, 35050-061) prior to use. $50 \mu\text{l}$ of 1.5×10^5 colony forming units (CFU) of *S. aureus* USA300 WT or *S. aureus* USA300 *Alip1/Alip2* bacterial cells were opsonised in $50 \mu\text{l}$ of 10% autologous human plasma for 15 min in a 96 well Cellstar U bottomed plate (Greiner Bio-One Inc, 650101) (37°C). Subsequently, 1.5×10^4 neutrophils (MOI=10) were incubated with the bacteria in the presence or absence of 200 nM rLip1 or rLip2. The plate was shaken at 600 rpm for 30 min at 37°C followed by cell lysis in 0.1% Triton X-100 (Sigma, P6416-100ML) and plated onto TSA using a modified Miles-Misra technique²², whereby $10 \mu\text{l}$ of each 10-fold bacterial dilution was plated, followed by incubation overnight at 37°C and counting of colonies.

Bacterial killing by whole blood

$75 \mu\text{l}$ of whole blood was infected with $25 \mu\text{l}$ of 1.5×10^5 CFU of *S. aureus* USA300 WT and *S. aureus* USA300 *Alip1/Alip2* in the presence or absence of 200 nM rLip1, rLip2 or both in a 96 well Cellstar U bottomed plate for 1, 2 and 4 h at 37°C , with shaking at 200 rpm. Blood was lysed in 0.1% (v/v) TritonX-100 (Sigma), viable bacteria counts were determined with $10 \mu\text{l}$ of ten-fold bacterial dilutions in PBS onto TSA using a modified Miles-Misra technique²² and incubated overnight at 37°C .

Isolation of CD14⁺ monocytes

Monocytes were isolated from human whole blood following centrifugation at $1200 \times g$ (no break) for 20 min. Buffy coats were combined and diluted with PBS and subsequently slowly pipetted over 15 ml of 1.199 g/mol ficoll paque plus (Sigma). A gradient was generated by centrifugation for 45 min at $200 \times g$ (no break), in which the mononuclear cell layer was subsequently removed. Ficoll was removed by centrifugation for 10 min with $300 \times g$, and resuspension in PBS. CD14⁺ monocytes were collected using a MAC-LS column as per the manufacturer's instructions (Miltenyi Biotec, 130-042-401).

Macrophage differentiation

For THP1 differentiation into macrophages, 5×10^5 THP1 cells were seeded in a 96-well Nunc flat bottomed plate in RPMI-1640 (Sigma), 10% (v/v) heat-inactivated foetal bovine serum (Gibco, 10270-106) and 1% (v/v) GlutaMAX (Gibco) in the presence of 200 nM phorbol 12-myristate 13-acetate (PMA, VWR P1585-1MG) for 3 d, before being left to rest

for 1 d in media without PMA. For blood monocyte-derived macrophages, 5×10^5 of purified human blood CD14⁺ cells were incubated for 5 d in 1:100 dilution of 10^4 U/ml recombinant human colony-stimulating factor-1 (hCSF-1, provided by Prof. D. Hume) in media. On the 5th day, cells were topped up with 25% complete medium containing $3 \times$ the target concentration of hCSF-1 and cells were used at day 7.

Gentamycin-protection assay

THP1 macrophages and blood-monocyte derived macrophages were infected at an MOI of 1 with bacteria suspended in fresh media (RPMI-1640 (Sigma), 10% (v/v) heat-inactivated foetal bovine serum (Gibco) and 1% (v/v) GlutaMAX (Gibco). Cells were centrifuged at $400 \times g$ for 5 min and incubated for 1 h at 37°C , 5% CO_2 . For analysing internalised bacteria (phagocytosis), cells were subsequently incubated with $100 \mu\text{g/ml}$ gentamycin (Sigma, G1397-10ML) in cRPMI for 30 min. To analyse intracellular survival, cells were subsequently left in $20 \mu\text{g/ml}$ gentamycin in media and were incubated for a further 24 h at 37°C , 5% CO_2 . Finally, to analyse the escape of intracellular bacteria, cells were incubated for 24 h in antibiotic-free media at 37°C , 5% CO_2 . At each time point, corresponding to the degree of phagocytosis, bacterial intracellular survival, and bacterial escape from the macrophage, cells were lysed in 0.1% Triton X-100 in PBS for 5 min at room temperature, and viable cell counting by plating onto TSA as described above.

Statistical methods

Statistical analysis was performed with GraphPad Prism 8 software (GraphPad, USA).

Results

Lipases do not inhibit *S. aureus* survival in human whole blood

Peripheral whole blood contains an array of innate immune components involved in the direct killing of *S. aureus*^{23–27}. To evaluate if lipases can promote *S. aureus* survival in blood, human whole blood was incubated with *S. aureus* USA300 LAC (*S. aureus* USA300 WT) or its isogenic mutant deficient in both lipase 1 and lipase 2 production (*S. aureus* USA300 *Alip1/Alip2*) for 1, 2, and 4 h at 37°C . Concurrently, *S. aureus* USA300 *Alip1/Alip2* was also incubated with 200 nM of functionally active rLip1 and rLip2 (Extended Figure 1²⁸). There was a 10-fold reduction in the number of recoverable bacteria in the first hour post-infection, followed by a stabilisation of the number of viable bacteria recovered up to 4 h, but there was no difference between the *S. aureus* USA300 WT and the lipase-deficient mutant or strains supplemented with recombinant lipase (Figure 1²⁸). Overall, these data indicate that lipases do not inhibit killing of *S. aureus* USA300 LAC in human whole blood.

S. aureus lipases do not inhibit neutrophil bactericidal activity

It was previously demonstrated that purified *S. aureus* lipases alter the phenotype of granulocytes, suggesting a possible impact on their function^{11,12}. To establish if lipases can inhibit neutrophil killing of *S. aureus*, human neutrophils were isolated from

fresh whole blood and incubated with opsonised *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$ for 30 min. As with whole blood, there was a 10-fold reduction in the number of viable bacteria after incubation with neutrophils, but viability between the *S. aureus* USA300 WT and the lipases-deficient strain did not differ (Figure 2²⁸). In addition, neutrophils were incubated with *S. aureus* USA300 $\Delta lip1/\Delta lip2$ in the presence of exogenous recombinant lipases and there were no differences in the number of recovered viable bacteria between the tested conditions (Figure 2²⁸). Taken together, these data indicate that lipases do not inhibit neutrophil-mediated killing of *S. aureus* USA300 LAC.

Lipases do not influence phagocytosis, intracellular survival or escape of *S. aureus* from macrophages

Recently, it was demonstrated that lipolysis of *S. aureus* lipoproteins by lipase 2 facilitated the survival of *S. aureus* through the

manipulation of macrophage cellular signalling¹³. In addition, *S. aureus* can interfere with macrophage phagolysosomal killing, enabling intracellular persistence¹⁶. To examine the capacity for *S. aureus* lipases to influence phagocytosis, intracellular survival, and escape from macrophages, primary human monocyte-derived macrophages were incubated with *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$ in the presence or absence of rLip1 or rLip2 (Figure 3a²⁸). Considerable variation in the number of recovered bacteria was observed between technical replicates due to donor variability, but no significant lipase-dependent differences were observed (Figure 3b²⁸). To further explore the effect of lipases on macrophage function, an immortalised cell line derived from human peripheral blood monocytes (THP1) cells was employed²⁹. PMA induces THP1 monocyte differentiation into adherent macrophages which represent a model of human monocyte-derived macrophages³⁰. *S. aureus* USA300 LAC infection of THP1 macrophages exhibited less variation between

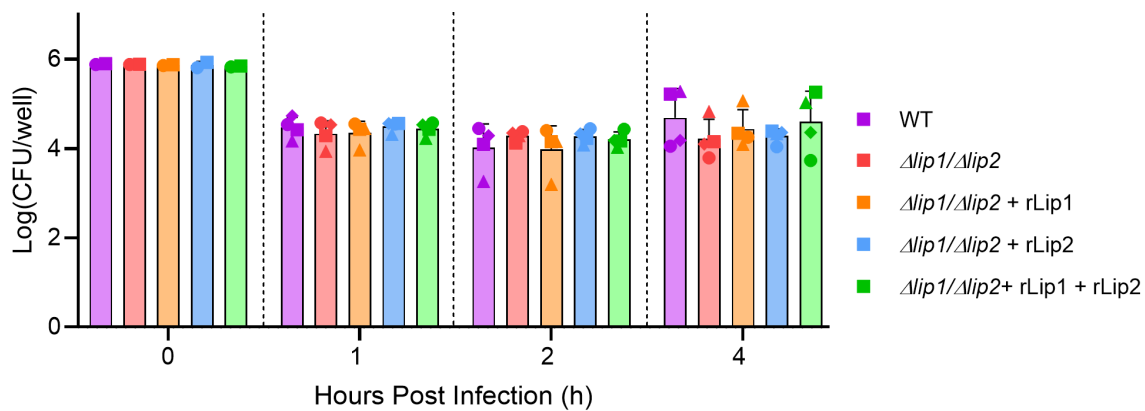


Figure 1. Lipases do not promote survival of *S. aureus* in human whole blood. Human whole blood was collected from healthy donors (each donor represented by a different symbol) and incubated with *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$, alongside *S. aureus* USA300 $\Delta lip1/\Delta lip2$ supplemented with 200 nM rLip1 and/or rLip2 for 0, 1, 2 and 4 h, at 37°C (with agitation). Each symbol represents the average of technical triplicates. Two-way ANOVA with Tukey's multiple comparisons. Bars show mean + SD, n=4.

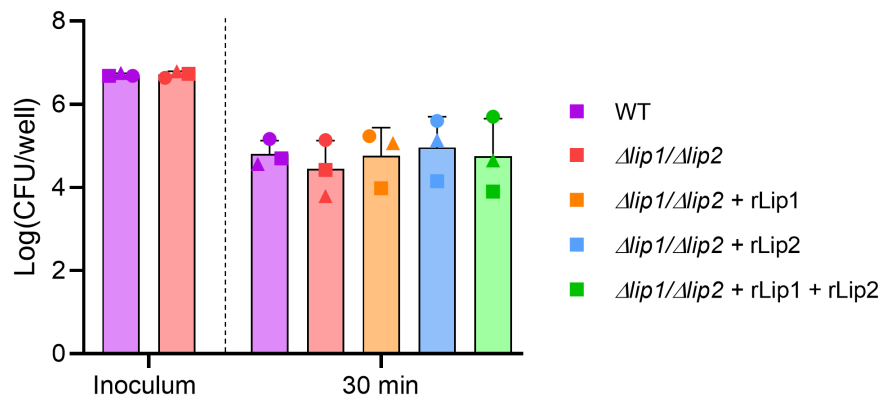


Figure 2. Lipases do not inhibit human neutrophil bactericidal activity. Human neutrophils were isolated from healthy donors (each donor represented by a different symbol) and incubated with plasma opsonised *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$ at an MOI of 10, in the presence or absence of 200 nM rLip1 and/or rLip2 for 30 min at 37°C (with vigorous agitation). Each symbol represents the mean of 5 technical replicates for an individual donor. One-way ANOVA with Tukey's multiple comparisons. Bars show mean + SD, n=3.

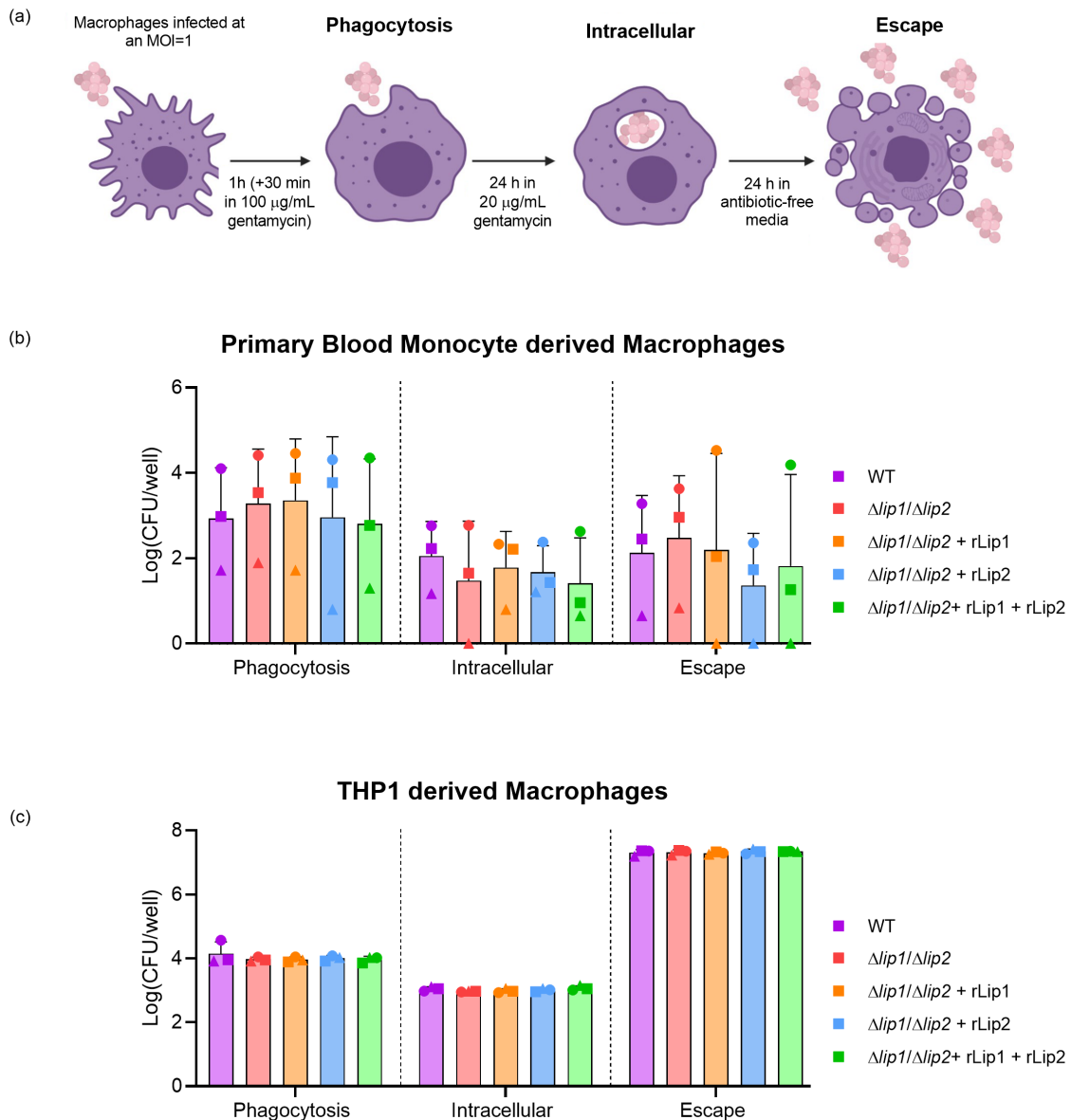


Figure 3. Lipases do not affect bacterial survival in human blood monocyte or THP1 derived macrophages. (a) Schematic of the assay used to analyse the phagocytosis, intracellular proliferation and escape of *S. aureus* from within macrophages. (b) Primary macrophages were differentiated from human blood monocytes isolated from 3 different healthy donors (represented by different symbols) and were incubated with *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$, alongside *S. aureus* USA300 $\Delta lip1/\Delta lip2$ supplemented with 200 nM rLip1 and/or rLip2 as per the schematic in Fig 4a, followed by plating and viable counting. (c) THP1 macrophages were incubated with *S. aureus* USA300 WT or *S. aureus* USA300 $\Delta lip1/\Delta lip2$, alongside *S. aureus* USA300 $\Delta lip1/\Delta lip2$ supplemented with 200 nM rLip1 and/or rLip2 following the schematic in Fig 4a. CFU analysis of phagocytosis, 24 h intracellular survival and 24 h escape was quantified for 3 technical replicates. Paired data was analysed using a two-way ANOVA, Tukey's multiple comparisons. Bars show mean + SD, n=3.

replicates when compared to primary cultures but no lipase-dependent differences in the number of bacteria recovered was observed (Figure 3c²⁸). Together, these data indicate that lipases do not affect phagocytosis, survival or escape of *S. aureus* from human macrophages.

Discussion

The importance of neutrophils in the initial response to *S. aureus* infection is well established^{24,31}. Previously, Roloff *et al.*, demonstrated, using scanning electron microscopy, that supernatant-purified *S. aureus* lipases altered granulocyte

morphology by denuding surface projections¹¹. As neutrophil phagocytosis is reliant on pseudopod extensions for ingesting bacteria, it was hypothesised that this phenotype could inhibit bactericidal activity²⁴. Furthermore, the release of extracellular DNA into the environment, through NETosis, could be impacted by lipase-mediated changes to the cellular membrane which could influence bacterial killing.

Here, we demonstrate that lipases do not inhibit direct killing of *S. aureus* mediated by human neutrophils, macrophages or whole blood *in vitro*. The findings are consistent with the findings of Nguyen *et al.*, who did not observe any differences in bacterial burden in the heart and liver in an *in vivo* murine sepsis model 24 h after infection with *S. aureus* USA300 WT LAC or an isogenic lipase-deficient mutant³. These data suggest that lipases do not interfere with the initial clearance of *S. aureus* from the blood.

A recent study by Chen *et al.*, reported that lipases have no direct effect on initial bacterial clearance in the early stages of infection. However they demonstrated that after 48 h, there was an indirect effect of lipase 2 resulting in reduced pro-inflammatory cytokine release by macrophages¹³. The authors found that *S. aureus* lipase 2 mediates cleavage of *S. aureus* lipoproteins, which are well characterised TLR2 ligands, resulting in increased bacterial burden by thwarting macrophage responses.

Previously it has been shown that *S. aureus* virulence factors regulated by the *agr* quorum-sensing system are required for survival and escape of *S. aureus* from macrophages, including the zinc metalloprotease Aur which is responsible for the downstream activation of the catalytically active lipases^{16,32,33}. Here, we report that the *agr*-regulated lipases do not influence the survival of *S. aureus* in human monocyte-derived macrophages, although considerable donor specific variation was observed with primary cells. Data obtained using the THP1 macrophage cell line further support the finding that *S. aureus* lipases do not affect phagocytosis, intracellular survival or escape of *S. aureus* from human macrophages. The lack of an observable effect of lipases may reflect the fact that bacterial capture by macrophages is dependent on dynamic actin-rich protrusions, with negligible involvement of triglyceride lipids in the process³⁴.

Conclusion

Overall, we report that *S. aureus* lipases do not directly impact on the killing mechanisms of neutrophils and macrophages. These data add to our understanding of *S. aureus* interactions with the innate immune system and the role of lipases in the pathogenesis of *S. aureus* disease.

Data availability

Underlying data

Edinburgh Datashare: *Staphylococcus aureus* secreted lipases do not inhibit innate immune killing mechanisms: Extended Figure 1. <https://doi.org/10.7488/ds/2881>²⁸

This project contains the following underlying data:

- Validation of rLip1 and rLip2.xlsx (ClarioSTAR (BMG Labtech) readouts of both rLip1 and rLip2, alongside 400 nM BSA. Absorbance was measured at OD₄₉₅ every 5 min for 20 h. Each experiment contained three technical repeats, n=3)
- Recombinant lipases Western blot, raw-unedited image. jpg (Raw gel image for Western presented in Extended Figure 1)

Extended data

Edinburgh Datashare: *Staphylococcus aureus* secreted lipases do not inhibit innate immune killing mechanisms: Extended Figure 1. <https://doi.org/10.7488/ds/2881>²⁸

- Extended Figure 1.docx

Extended Figure 1: Functional characterisation of purified recombinant *S. aureus* lipase 1 and 2.

(a) Purification of recombinant lipase 1 and 2 was analysed using western blot, in which bands present at 76 kDa indicated the correct protein elution (detected by hexa-his tag, α -diagnostics HISP12-HRP). Page-Ruler ladder (furthest left well) shows the visible protein marker at 75 kDa. Measurement of lipolytic activity of recombinant protein 1 (rLip1) (b) and 2 (rLip2) (c). It was observed that both lipase 1 and 2 were functionally active enzymes which were able to cleave Tween-20 over a broad scope of concentrations. Indeed, it was also observed that lipase 2 was much more kinetically active in comparison to lipase 1, which could be attributed to its broader substrate range. Two-way ANOVA, Dunnett's Multiple Comparisons against the BSA negative control, $\alpha=0.05$, **** $p<0.001$. Each point shows mean + SD (Data represent a representative experiment, from three independent experiments).

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/) (CC-BY 4.0).

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Meera Unnikrishnan 

Division of Biomedical Sciences, University of Warwick, Coventry, UK

This is a clearly written manuscript, investigating the role of the *S. aureus* lipase 1 and 2 in bacterial killing in whole blood, neutrophils and macrophages. This work is important as lipases are generally considered to impact innate cell functions, although effects on bacterial killing have not been directly assessed yet.

In general experiments have been done well, and the main conclusions are valid. However a few technical clarifications are required.

For the macrophage internalisation assays, when following bacterial replication by CFU, usually it is good to look at an earlier time point as by 24h there can be significant cell lysis induced by the WT. Have the authors checked the morphologies or the states of the macrophages during their experiments?

The bacterial 'escape' measurements from macrophages need some clarification. It is not clear if after the 1h infection the macrophages were treated with gentamicin to kill all the extracellular bacteria first prior to adding the antibiotic-free medium. This is essential to do in order to measure the escape of intracellular bacteria. It is also not clear if escape was quantified from the culture supernatants or from the the cell lysates. Quantifying bacterial escape is quite tricky, and should be preferably done at multiple time points after infection to get a clear picture. Counting from supernatants are not accurate as most bacteria settle down to the well by 24h forming microcolonies, and by 24h there is substantial cell lysis, so hard to deduce the 'escaped' population by measuring intracellular counts (by cell lysis). Finally, the authors may want to comment on why the 'escape' numbers are so high between the THP1 vs the primary cells.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Partly

Are sufficient details of methods and analysis provided to allow replication by others?

Partly

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

No source data required

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Host pathogen interactions: Intracellular *S. aureus* infections, Type VII secretion systems, *C. difficile* colonisation

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 08 February 2021

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Julie A. Morrissey

Department of Genetics and Genome Biology, University of Leicester, Leicester, UK

This is a very clearly written interesting and relevant article on the role of two *Staphylococcus aureus* lipases that are secreted into the extracellular milieu. Previous studies have suggested that these lipases may interfere with the function of innate immune cells but no previous articles have investigated the direct role of the lipases. This article describes the phenotypic analysis of a *S. aureus* double lipase mutant and recombinant lipase proteins. There were no differences in the survival between the wild type and mutant strains with human whole blood or neutrophils and no effect on phagocytosis, intracellular survival, or escape from both human primary and immortalised cell line macrophages. Therefore, it was concluded that *S. aureus* lipases do not inhibit bacterial killing mechanisms of human macrophages, neutrophils, or whole blood. The article gives a very clear background of the literature of *S. aureus* lipases in innate immunity. The study design is logical and uses a series of well-established phenotypic assays that are well described and use appropriate controls and are shown to be statistically reproducible. There are

no major issues to address.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

Yes

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Staphylococcus aureus, pathogenesis, host-microbe interactions

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Reviewer Report 23 December 2020

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Francis Alonzo III

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In this Research Article, Sargison *et al.* test if the *S. aureus* secreted lipases, Lip1 and Lip2, interfere with the phagocytic killing mechanisms of innate immune cells. Prior work had suggested there might be potential effects on immune cell phagocytosis/killing on account of morphological changes that occurred after the treatment of granulocytes with lipases. Additional work has also demonstrated a role for lipases in interference with immune signaling on account of lipolysis of the lipopeptide ligands of TLR2. No studies to date have directly tested the role of lipases in phagocytic killing or escape from innate immune cells. The authors use a series of well-executed in

vitro survival assays with whole blood, primary human neutrophils, monocyte-derived macrophages, and THP-1 differentiated macrophages to monitor the survival of WT *S. aureus* and an isogenic *lip1/lip2* deletion mutant +/- recombinant Lip1/Lip2 over time. They find that lipases do not have a discernable effect on phagocytic killing or bacterial escape in the assays used. The methods contain sufficient detail, and the results are clear and unambiguous. Incidentally, the work also closely resembles unpublished observations made in our lab with murine neutrophils and macrophages, thus the data are further bolstered by similar outcomes among multiple groups. Overall, this work provides important information related to the roles of lipases in host-microbe interactions. I have no substantive criticism of this solid study.

Is the work clearly and accurately presented and does it cite the current literature?

Yes

Is the study design appropriate and is the work technically sound?

Yes

Are sufficient details of methods and analysis provided to allow replication by others?

Yes

If applicable, is the statistical analysis and its interpretation appropriate?

Yes

Are all the source data underlying the results available to ensure full reproducibility?

No source data required

Are the conclusions drawn adequately supported by the results?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: *S. aureus* pathogenesis, host-microbe interactions, immune response to infection.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
