

apunts

MEDICINA DE L'ESPORT

www.apunts.org



REVISION ARTICLE

Physical activity and lipid oxidation

Andreu Arquera, Roberto Elosuab, and Jaume Marrugatb,*

- ^a Centro de Alto Rendimiento de San Cugat, Barcelona, Spain
- b Instituto Municipal de Investigación Médica, Barcelona, Spain
- ^c CIBER Epidemiología y Salud Pública, Barcelona, Spain

Received December 4, 2009; accepted December 9, 2009

KEYWORDS

Arteriosclerosis; Lipid oxidation; Cardiovascular diseases; Physical activity

Abstract

Regular physical activity (PA) is associated with lower cardiovascular mortality and morbidity. Part of these benefits is related to the effects over the classic cardiovascular risk factors. These effects, however, only explain part of the protection of PA from these types of diseases. The oxidation of LDL cholesterol particles, which is the aetiopathogenic mechanism of a great part of cardiovascular diseases, plays an important role in the arteriosclerotic process. This narrative review presents current knowledge on the relationship between carrying out physical activity and lipid oxidation.

© 2009 Consell Català de l'Esport. Generalitat de Catalunya. Published by Elsevier

PALABRAS CLAVE

Arteriosclerosis; Oxidación lipídica; Patologías cardiovasculares; Actividad física

Actividad física y estrés oxidativo

España, S.L. All rights reserved.

Resumen

La práctica regular de actividad física (AF) se asocia con una menor mortalidad y morbilidad cardiovascular. Parte de estos efectos beneficiosos están relacionados con los efectos favorables sobre los factores de riesgo cardiovascular clásicos. Sin embargo, estos efectos explican sólo una parte de la protección de la AF sobre este tipo de enfermedades. La oxidación de las partículas de LDL colesterol tiene un papel fundamental en el proceso de la arteriosclerosis que es el mecanismo etiopatogénico de gran parte de las enfermedades cardiovasculares. En esta revisión narrativa se presenta el conocimiento actual sobre la relación entre la práctica de AF y la oxidación lipídica.

 \odot 2009 Consell Català de l'Esport. Generalitat de Catalunya. Publicado por Elsevier España, S.L. Todos los derechos reservados.

E-mail: jmarrugat@imim.es (Jaume Marrugat)

^{*}Author for correspondence.

Introduction

Atherosclerosis is the process that explains the aetiopathogenesis of various chronic cardiovascular pathologies responsible for much morbi-mortality in most of the world. Among this group of diseases, coronary disease constitutes the highest single cause of death in the western world^{1,2}. In Spain, coronary disease was responsible for 10.2% of all deaths and 3% of hospital morbidity in 2007^{3,4}. Prospects for the future, according to various authors, include an increasing trend that will also reach developing countries⁵.

Regular practice of physical activity (PA) reduces the risk of all-cause premature death in young and middle-aged individuals⁶ and is also associated with better survival in older people⁷. Regular exercise diminishes the risk of presenting with a cerebrovascular accident⁸ and cuts the risk of an acute coronary event in half⁹⁻¹². As a consequence, sedentary lifestyle is an independent risk factor for coronary disease^{13,14}, and promoting the practice of PA is one of the most important elements of public health campaigns for cardiovascular prevention^{15,16}.

Regular practice of PA produces favourable effects on classic risk factors for cardiovascular diseases: improves the lipid profile¹⁷, control blood pressure¹⁸ and prevents the appearance of non-insulin-dependent diabetes¹⁹. Nonetheless, these effects explain only part of the protection PA offers against this type of diseases²⁰.

PA has beneficial effects on lipid oxidation, haemostasis and endothelial function, factors that are also directly involved in the development and progression of atherosclerosis. In this narrative review of the literature we present the current knowledge about the relationship between PA and lipid oxidation.

Lipid oxidation and atherosclerosis

Oxidative stress has been associated to the development of diverse diseases and chronic processes, including atherosclerosis. The oxidative state is controlled by the equilibrium between the formation of free radicals, which are pro-oxidants, and the action of antioxidant systems. Oxidation of the components of low-density lipoprotein (LDL) is one of the cornerstones of atherosclerosis 1,21,22 . Oxidized LDL (LDL $_{\rm ox}$) participates in various processes that favour the appearance and progression of atheromatous plaque: LDL particles a) cause lesions on endothelial cells that change the vascular tone and permeability of the endothelium^{1,22}; b) induce expression of monocyte adhesion molecules on the surface of the endothelial light^{23,24}; c) act as chemotactic factors to attract monocytes from the bloodstream to the subendothelial space²⁵; d) enter the macrophages by way of the scavenger receptor, which are converted into foam cells and then form the fatty streak that constitutes the first atherosclerotic lesion²²; and e) induce proliferation of smooth muscle cells and their migration from the media layer of the arterial wall to the subendothelial space²⁶.

LDL oxidation is a complex process that basically depends on three factors:

- a) Formation of free radicals (FR). FR are unstable and very reactive molecules that are produced in any process in which oxygen intervenes²⁷. They can react with all of the molecules in an organism (proteins, lipids, DNA), changing their structure and function²⁸. The reaction of FR with fatty acids is responsible for LDL oxidation in a process called lipid peroxidation^{29,30}.
- b) Activity of antioxidant substances. To protect itself from the action of FR, the organism has an antioxidant defense system³¹. This system consists of endogenous substances, synthesized by the organism itself, such as superoxide dismutase (SOD), glutathione reductase (GSR) or paraoxonase (PON), and others of exogenous origin, ingested with food, such as vitamins E and C, B-carotenes and polyphenols³¹⁻³³. In addition, antioxidants are a heterogeneous group of substances that act synergistically, some in liquid medium (GSR, SOD, vitamin C, polyphenols) and others in the lipid environment (PON, vitamin E, B-carotenes) (Figure 1).
- c) Intrinsic properties of LDL. The larger and less dense the LDL particle, the less susceptible it is to oxidation³⁴; a higher polyunsatured fatty acid content³⁵ and glycosylation of the particle result in greater susceptibility to oxidation³⁶; on the other hand, a greater presence antioxidant substances, primarily vitamin E and to a lesser extent the B-carotenes in the LDL particle itself protect against the action of free radicals³⁷.

Physical activity and lipid oxidation

Both acute and regular PA practice influence FR production, antioxidant activity and LDL susceptibility to oxidation.

Acute physical activity practice and production of free radicals

The average FR life span is very short, and therefore direct quantification is impossible. For this reason, indirect markers are used. For the lipid oxidation study, the markers most often used are malondialdehyde (MDA) and substances that react with thiobarbituric acid (TBARS)^{38,39}.

During exercise, oxygen consumption increases, which translates into higher FR production. This increase could be so great that it surpasses the protective capacity of the antioxidant systems, resulting in increased oxidation processes, among them lipid peroxidation⁴⁰.

Various studies have observed that after acute PA there is a higher plasma level of by-products of lipid oxidation⁴¹⁻⁴⁸ (Tables 1 and 2). Some researchers have observed that this increase is directly related to the intensity of the PA involved⁴³. In any case, this increased oxidative stress does not persistent and, at least in people accustomed to exercise, is normalized in a few hours^{44,48}. Another fact that emerges from these studies is that in animals or individuals who regularly train, the observed increase is less than in sedentary subjects^{45,46}, suggesting the

32 A. Arquer et al

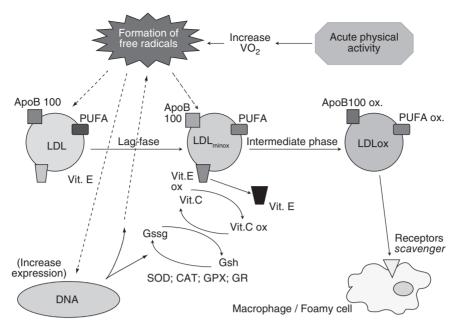


Figure 1 Simplified relationship between the practice of acute physical activity, regular physical activity practice and production of free radicals, lipid oxidation and antioxidant systems. [Adapted from Codina et al. Med Clín (Barc). 1999;112:508-15]. ApoB 100: apoprotein B-100; PUFA: polyunsaturated fatty acids; Vit. E: vitamin E; LDL: low density lipoproteins; Vit. Eox: vitamin E oxidized; LDL_{minox}: low density lipoproteins minimally oxidized; LDLox: low density lipoproteins oxidized; Apo B100ox: apoproteín B-100 oxidized; PUFAox: polyunsaturated fatty acids oxidized; Vit. C: vitamin C; Vit. Cox: vitamin C oxidized; Gssg: glutathione oxidized; Gsh: glutathione reduced.

Table 1 Basic description of studies that have assessed the relationship between the practice of physical activity and the production of free radicals and other oxidation products in animal models

Author	Physic	cal activity				Results
	n	Model	Level of training	Intervention	Variable	-
Davies ⁴⁰	6	Rats	Sedentary	Submaximal exercise until exhaustion	MDA	↑MDA in muscles and liver
Ji ⁴¹	6	Rats	Sedentary	Submaximal exercise until exhaustion	MDA	↑MDA in liver, without muscle changes
Alessio ⁵⁸	32	Rats	One group that trains	Submaximal exercise until exhaustion	MDA	↑MDA in sedentary subjects not found in those who trained
Jenkins ⁴⁸		Rats	One group that trains	Submaximal exercise until exhaustion	MDA	trinary excretion of MDA in both those who trained and those who did not

existence of a protective effect from the regular practice of PA.

Another of the mechanisms that explain lower free radicals production in those who train is explained by an improvement in metabolic efficiency mainly related to the metabolic substrate used to obtain energy. A person who weighs 70 kg has approximately 15 kg of fat in the form of triglycerides in the adipose tissue, which represents about 140,000 Kcal. With the availability of this great quantity of energy, the question is why triglycerides are not the organism's only source of energy, since obtaining energy

while exerting physical effort at maximum intensity requires the utilization of carbohydrates. The explanation for this limitation on using adipose tissue as the metabolic substrate to derive energy during the practice of maximum intensity exercise is not entirely clear, although it could be related to different factors:

1) The speed at which fat is released from the peripheral adipose tissue. Lipolysis in the adipose tissue is regulated by the nervous and hormonal systems. It has recently been demonstrated that 70% of fatty acids released from

Basic description of studies that have assessed the relationship between the practice of physical activity and production of free radicals other products of oxidation in humans

Author	С	Level of training	Intervention	Variables	Results
Maughan⁴²	16	Sedentary	45 min with incline of 12% to 75% TMHR	MDA	↑MDA. (Gradient: 6 hours post-exercise > 24 hours > 48 hours)
Kanter⁴⁵	20	Training, with VO ₂ max >50 ml·kg ⁻¹ · min ⁻¹	30 min running at 60% VO ₂ max. + 5 min of gradual increase to 2.5 min at 90% VO ₂ max.	MDA	↑MDA in serum
Viinikka ⁴⁶	10	Cyclists	10-14 min. exercise on stationary bicycles	MDA	No change, MDA in serum
Laaksonen⁴³	22	Sedentary	40 min. exercise on stationary bicycles at 60% VO, máx.	TBARS	↑TBARS
Treuth⁴	∞	Sedentary	PA (intense): 60 min bicycling at 50% VO ₂ max. PA (very intense): intervals bicycling at 100% VO ₂ max.		↑Oxidation, greater with higher PA intensity
Neubauer ⁴⁷	42	Triathletes	Triatĥlon	Various products	Increase in products of oxidation after competition, normalized by day 5 post-competition

MDA: malondialdehyde; TBARS: thiobarbituric acid reactive substances; TMHR: theoretical maximum heart rate; Vo, max: maximum oxygen volume.

- the adipose tissue are reesterified; this value drops to 25% at the start of submaximal exercise (40% of the maximum oxygen consumption). Therefore, a mechanism that increases fat utilization could be related to a reduction in reesterification.
- 2) The carrying capacity of fatty acids in the fatty tissue surrounding muscles and the muscles' captation capacity. A correlation has been found between increased plasma concentration of free fatty acids (FFA) and captation of these FFA by the muscle during exercise. Increased muscle capacity for FFA captation is directly related to an increase in muscle lipoprotein lipase (LPL) activitiv⁴⁹. Muscle captation of FFAs increases linearly with the availability of FFAs circulating in the trained muscle, while the untrained muscle reaches a maximum FFA captation capacity in time. This difference in behaviour between the trained and untrained muscle partly explains the greater utilization of fats in the trained muscle during PA and suggests that local adaptations secondary to training, as for example the increase in LPL activity, are important⁵⁰. On the other hand, muscle captation of glucose increases during prolonged exercise, both in those who train and those who don't, although this captation is much higher in those who do not train.
- 3) Muscle deposits of fats. Training increases the intramyocellular deposits of fat, parallel to the muscle's capacity for oxidation of fats⁵¹. Circulating FFAs from the peripheral adipose tissue and intramuscular deposits are the primary sources of fats. Catecholamines are the main stimulants for lipolysis in the adipose tissue, although the low plasma concentration of insulin also has a relevant role. On the other hand, lipolysis of intramuscular fat deposits is mediated only by beta-adrenergic stimulation. Training induces a progressive increase in the utilization of the fat in intramuscular deposits and a reduction in the utilization of carbohydrates⁵². These changes have now been observed at 5 days after the start of training, even before any increase in enzymatic mitochondrial muscle activity⁵³.

Lipolitic capacity in response to exercise diminishes with higher adiposity. The slightest increase in lipolysis capacity in overweight or obese individuals limits the availability of FFA as a metabolic substrate for energy production, compared with individuals of normal weight⁵⁴. This is important for the treatment of obesity; it has been demonstrated that training improves fat catabolism in people who have been sedentary and obese, while diet alone does not55. Some authors have suggested that decreased fatty mass and not age or maximum oxygen volume is the best individual predictor of a decline in fat oxidation capacity at rest. These results support the theory that a reduced fat oxidation capacity that occurs with age is associated with higher adiposity and fewer FFAs. Interventions that increase the quantity of FFAs, such as physical training, might increase oxidation capacity using fats as a metabolic substrate and limit the increase in peripheral adiposity that occurs with aging⁵⁶. Finally, children are better adapted to aerobic metabolism because their energy expenditure depends fundamentally on oxidative metabolism using fat as a metabolic substrate⁵⁷.

34 A. Arguer et al

The currently available data support the hypothesis that one of the protective effects of training for PA could be based on the production of fewer FRs due to higher metabolic efficiency, since less oxygen is consumed to obtain the same amount of energy.

Acute and regular physical activity practice and antioxidant systems

With the greater FR production secondary to PA practice, the organism can adapt itself by increasing endogenous antioxidant capacity. Experimental animal studies (Table 3) have demonstrated that regular practice of PA increases the activity of endogenous antioxidant enzymes⁵⁸⁻⁶². In many of these studies, a direct relationship has been observed between the intensity of the PA during the training programme⁶³⁻⁶⁶ and increased capacity of the antioxidants systems.

In cross-sectional studies (Table 4), greater antioxidant enzyme activity has been observed in trained human subjects

compared to those inactive^{65,66}. The few experimental studies present conflicting results: in one study, no changes were observed in antioxidant enzyme activity after a training period⁶⁷; in other findings, activity increased^{68,69}. These differences could be explained by differences in the duration and intensity of the training programme.

FRs can directly influence the expression of DNA, producing higher expression of the genes that codify these enzymes⁷⁰⁻⁷² and explain the antioxidant enzyme activity increase observed after a training period.

Physical activity and susceptibility of LDL to oxidation

Another mechanism by which PA might protect LDL from oxidation could be the reduction of LDL susceptibility to the oxidation process.

The experimental studies that have analyzed the effect of acute PA on LDL susceptibility obtained inconclusive results. Sanchez-Quesada et al⁷³ observed, in trained

Table 3 Basic description of studies that have assessed the relationship between regular practice of physical activity and the antioxidants system in animal models

Author	Sample		Physical activity		Results
	Animal	Tissue	Intervention	Variables	_
Powers ⁶¹	Rats	Diaphragm	Training (continuous running) in diverse groups, by intensity (high, moderate, light) and duration (30, 60, 90 min/day) 4 days/week for 10 weeks		↑OD, GRX, CS at all intensities and durations of activity. SOD increase was greater at higher and moderate intensities and at 60 min/day or more
Powers ⁶²	Rats	Myocardium	Training (continuous running) in diverse groups, by intensity (high, moderate, light) and duration (30, 60, 90 min/day) 4 days/week for 10 weeks	SOD, GRS, CAT	↑SOD activity at high intensity, all durations, and at moderate intensity at 90 min. No changes found in other enzymes
Criswell ⁶³	Rats	Muscle	Two types of training (12 weeks): intervals (6 series of 5 min al 80-90% VO ₂ max) and continuous (45 min at 70% VO ₂ max)	SOD,GRS	↑GRS activity in interval training group. ↑SOD activity in both groups
Sen ⁵⁹	Dogs	Liver, muscle	Training (continuous running): 40 km/day at 5.5-6.8 km/hour with 15% incline, 5 days/week for 55 weeks	GRS	↑GRS (quantity)
Sen ⁵⁹	Rats	Liver, muscle	Training (continuous running): 2 hours/day at 2.1 km/hour, 5 days/week for 8 weeks	GRS	↑GRS (quantity)
Marin ⁶⁰	Dogs	Muscle, liver	Training (continuous running): 5 days/week with 15% incline for 30 weeks	GRS	↑GRS activity in muscles exercised. No changes fund in liver
Vani ⁶⁴	Rats	Liver	Training (continuous running): 3 groups at same intensity, different duration: 1 day, 10 days and 60 days.	MDA, SOD, GRS, XO	↑SOD, XO activity as training period increased. No changes found in GRS

GRS: glutathione reductase; SOD: superoxide dismutase; CS: citrate synthase; CAT: catalase; XO: xanthine oxidase; VO_2 max: maximum oxygen volume.

Author	Sample				Physical activity		Results
	_ _	Groups	Tissue	Design	Intervention	Variables	
Mena ⁶⁵		Sedentary	Erythrocytes	Transversal		SOD, CAT, GPX	Basal: SOD activity in cyclists (professionals and amateurs) higher than in codentary subjects
		Cyclist (amateur) Cyclist					CAT and GPX activity higher in professionals than in amateurs and
Covas ⁶⁶	488	Women		Transversal		SOD, GPX	Practice of physical activity directly associates with SOD and GPX activity
Tiidus ⁶⁷	7 9	Men	Muscle	Experimental	Training (cycling) 35 min 3 days/week SOD, CAT, for 8 weeks	SOD, CAT,	No changes in SOD, GPX and CAT activity after training period
Elosua ⁶⁸	7 0	Men		Experimental	Training (aerobic), 45-60 min 3-5 days/ SOD,GPX, GR ↑GPX, ↑GR activity after the training week for 16 weeks	SOD, GPX, GR	↑GPX, ↑GR activity after the training
Evelo ⁶⁹	18 73 9	Women	Erythro-cytes	Erythro-cytes Experimental	Training in two 20 weeks periods. Running 15 km after 20 weeks, half-marathon after 40 weeks	X	GDX after 20 wks of training. Levels maintained in succeeding 20 weeks. After the two races (15 km after 20 weeks of training; 21 km after 40 weeks of training) GPX activity decreased sharply, with full normalization at 5 days

GPX: glutathione peroxidase; SOD: superoxide dismutase; CAT: catalase; GR: glutation reductase.

36 A. Arquer et al

individuals, an increase in LDL susceptibility to oxidation immediately after completing a 4-hour run. Other studies that have analysed changes more than 8 hours after PA did not observe an increase⁷⁴⁻⁷⁵. From these studies we could hypothesize that trained individuals have increased susceptibility to LDL oxidation during a short period of time after engaging in intense, prolonged, acute PA.

No studies have addressed the effect of acute PA on oxidation susceptibility in sedentary subjects, although it would be reasonable to expect an increase of greater magnitude and duration than in the athletes studied.

Another key point is the effect of a training period on LDL susceptibility to oxidation. The crossectional studies that compared physically active and sedentary subjects have established that those actives have less LDL susceptibility to oxidation 76,77. It has been reported that after a training period LDL susceptibility to oxidation decreases, reducing the oxidized LDL in circulation 68. The reason for this greater resistance to oxidation is not entirely clear, although some evidence suggests that it is related to a qualitative change in LDL subclasses with a reduction of the dense fractions and an increase in the mean diameter of the LDL particles 78-80.

The global effect of all these mechanisms that adapt to training is reduced levels of lipid oxidation that has been reported in healthy individuals^{68,81} and also in patients with previous ischemic heart disease⁸².

Physical activity and inflammation

One of the most important consequences of this decrease in lipid oxidation is the simultaneous reduction in systemic markers of inflammation. Various transversal studies have described an inverse association between the regular practice of physical activity and diverse inflammation markers, especially high sensitivity C-reactive protein (CRPhs)⁸³⁻⁹¹. Although a recent meta-analysis of experimental studies that considered this association concluded that aerobic exercise does not reduce CRPhs levels in adults⁹², that meta-analysis included only 5 studies with a total of 323 participants. Therefore, the findings should be interpreted with caution.

A decrease in levels of interleukin 6 (IL-6) have been observed in some studies^{91,93} but not in others⁹⁴. In this context, we must take into account the role of myokines in muscle function. During contractile activity, the muscle produces a series of myokines, among them IL-6, that can exert beneficial effects at the systemic level. Some studies have observed that muscle IL-6 induces systemic expression of anti-inflammatory molecules, such as IL-10 and the IL-1 receptor antagonist, and reduces the presence of pro-inflammatory molecules such as tumour necrosis factoralpha. In addition, muscle IL-6 induces lipolysis, which stimulates the utilization of fatty acids as a metabolic substrate to obtain energy.

Conclusions

Regular practice of PA produces a series of beneficial effects on oxidative metabolism that translate into less oxidative stress. The mechanisms involved are related to an increase in energy efficiency and the utilization of fatty acids as a metabolic substrate for energy production, a greater capacity to defend against oxidative stimulants due to an increase in endogenous antioxidant enzymes activity, and an increase in LDL resistance to oxidation. All of this translates into a reduction in the levels of oxidized LDL and systemic markers of inflammation.

Conflict of interest

The authors declare no conflict and no financial interests.

Funding sources

This work was supported by the Ministerio de Ciencia e Innovación, Instituto Carlos III/FEDER (Red HERACLES RD06/0009), and Agencia de Gestió d'Ajuts Universitaris y d'Investigació de la Generalitat de Catalunya (2009 SGR 1195).

References

- 1. Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990s. Nature. 1993;362:801-9.
- Marrugat J, Elosua R, Martí E. Epidemiología de la cardiopatía isquémica en España: estimación del número de casos y de las tendencias entre 1997 y 2005. Rev Esp Cardiol. 2002;55:337-46
- Instituto Nacional de Estadística. Tablas de mortalidad de la población de España por comunidades autónomas, sexo, edades, años y funciones. [Accessed January 30, 2009]. In: http:// www.ine.es/jaxi/menu.do?type=pcaxis&path=%2Ft20%2Fp 319a%2F1992-2005&file=pcaxis&N=&L=0
- Centro Nacional de Epidemiología. Área de Epidemiología Aplicada del Servicio de Epidemiología de Enfermedades Cardiovasculares. [Accessed January 30, 2009]. In: http://www.isciii. es/htdocs/centros/epidemiologia/epi_cardio_tabla3.jsp
- Murray CJ, López AD. The global burden of disease: a comprehensive assessment of mortality and disability from diseases, injuries and risk factors in 1990 and projected to 2020. Cambridge: Harvard University Press; 1996.
- Nocon M, Hiemann T, Müller-Riemenschneider F, Thalau F, Roll S, Willich SN. Association of physical activity with all-cause and cardiovascular mortality: a systematic review and meta-analysis. Eur J Cardiovasc Prev Rehabil. 2008;15:239-46.
- 7. Hakim AA, Petrovitch H, Burchfiel M, Ross WG, Rodriguez BL, White LR, et al. Effects of walking on mortality among non-smoking retired men. N Engl J Med. 1998;338:94-9.
- Guillum RF, Mussolino ME, Ingram DD. Physical activity and stroke incidence in women and men. The NHANES I epidemiologic follow-up study. Am J Epidemiol. 1996;143:860-9.
- Powell KE, Thompson PD, Caspersen CJ, Kendrich LS. Physical activity and the incidence of coronary heart disease. Ann Rev Publ Health. 1987;8:253-87.
- Morris JN, Everit MG, Pollard R, Chave SP, Semmence AM. Vigorous exercise in leisure time. Protection against coronary heart disease. Lancet. 1980;2:1207-10.
- Paffenbarger RS, Wing AL, Hyde RT. Physical activity as and index of heart attack risk in college alumni. Am J Epidemiol. 1978;108:161-75.

- Sofi F, Capalbo A, Cesari F, Abbate R, Gensini GF. Physical activity during leisure time and primary prevention of coronary heart disease: an updated meta-analysis of cohort studies. Eur J Cardiovasc Prev Rehabil. 2008;15:247-57.
- 13. Fletcher GF, Blair SN, Blumenthal J, Caspersen C, Chaitmain B, Epstein S, et al. Statement on exercise. Benefits and recommendations for physical activity programs for all Americans. A statement for health professionals by the Committee on exercise and cardiac rehabilitation of the Council on clinical cardiology, American Heart Association. Circulation. 1992;86:340-4.
- Bijnen FC, Caspersen DJ, Mosterd WL. Physical inactivity as a risk factor for coronary heart disease: a WHO and International Society and Federation of Cardiology. Position Statement. Bull World Health Organ. 1994;72:1-4.
- 15. Giannuzzi P, Mezzani A, Saner H, Björnstad H, Fioretti P, Mendes M, et al; Working Group on Cardiac Rehabilitation and Exercise Physiology. European Society of Cardiology. Physical activity for primary and secondary prevention. Position paper of the Working Group on Cardiac Rehabilitation and Exercise Physiology of the European Society of Cardiology. Eur J Cardiovasc Prev Rehabil. 2003;10:319-27.
- Haskell WL, Lee IM, Pate RR, Powell KE, Blair SN, Franklin BA, et al. Physical activity and public health: updated recommendation for adults from the American College of Sports Medicine and the American Heart Association. Circulation. 2007;116:1081-93
- Marrugat J, Elosua R, Covas MI, Molina L, Rubies-Prat J, and the MARATHOM investigators. Amount and intensity of physical activity, physical fitness and serum lipid in men. Am J Epidemiol. 1996;143:562-9.
- 18. American College of Sports Medicine. Physical activity, physical fitness and hypertension. Med Sci Sports Exerc. 1993;25:I-X.
- Helmrich SP, Ragland DR, Leung RW, Paffenbarger RS. Physical activity and reduced occurrence of non-insulin-dependent diabetes mellitus. N Engl J Med. 1991;325:874-7.
- Ekelund LG, Haskell WL, Johnson JL, Whaley FS, Criqui MH, Sheps DS. Physical fitness as a predictor of cardiovascular mortality in asymptomatic North American Men. The Lipid Research Clinics Mortality Follow-up Study. N Engl J Med. 1988;319:1379-84.
- 21. Witzum JL. The oxidation hypothesis of atherosclerosis. Lancet. 1994;344:793-5.
- 22. Chison GM, Penn MS. Oxidized lipoproteins and Atherosclerosis. In: Fuster V, Ross R, Topol EJ, editors. Atherosclerosis and coronary artery disease. Philadelphia-Nework: Lipincott-Raven Publishers; 1996. p. 129-49.
- Ross R. The pathogenesis of atherosclerosis: An update. N Eng J Med. 1986;314:488-500.
- 24. Maier JA, Barenghi L, Pagani F, Bradamante S, Comi P, Ragnotti G. The protective role of high density lipoprotein on oxidized-low density-lipoprotein-induced U937 endothelial cell interactions. Eur J Biochem. 1994;221:35-41.
- Terkeltaub R, Banka CL, Solan J, Santoro D, Braud K, Curtiss LK. Oxidized LDL induces monocytic cell expression of interleukin-8, a chemokine with T-lymphocyte chemotactic activity. Arterioscler Thromb. 1994;14:47-53.
- Kume N, Gimbrone MA Jr. Lysophosphatidylcholine transcriptionally induces growth factor gene expressions in cultured human endothelial cells. J Clin Invest. 1994;93:907-11.
- Ballester M. Antioxidantes, radicales libres y salud. Un enfoque químico-orgánico-físico. Med Clin (Barc). 1996;107:509-15.
- 28. Davies KJ. Oxidative stress. The paradox of aerobic life. Bioch Soc Sym. 1995;6:1-31.
- Esterbauer H, Jürgens G. Mechanistic and genetic aspects of susceptibility of LDL to oxidation. Curr Opin Lipidol. 1993;4:114-24.
- Halliwell B. Reactive oxygen species in living systems: Source, biochemistry and role in human disease. Am J Med. 1991; 91(3C):14S-21S.

- 31. Sen CK. Oxidants and antioxidants in exercise. J Appl Physiol. 1995;61:1-31.
- Romero D, Villalba MP, Mur M, Cabeza F, Guerrero L, Simal E. Importancia de los antioxidantes en la alimentación humana. Med Clin (Barc). 1990;94:69-75.
- 33. Gutteridge JMC. Lipid peroxidation and antioxidants as biomarkers of tissue damage. Clin Chem. 1995;41:1819-28.
- Beard CM, Barnard J, Robbins D, Ordovas JM, Shaefer EJ. Effects of diet and exercise on qualitative and quantitative measures of LDL an its susceptibility to oxidation. Arterioscler Thromb Vasc Biol. 1996;16:201-7.
- 35. Lyons TJ. Glycation and oxidation: a role in the pathogenesis of atherosclerosis. Am J Cardiol. 1993;71:26B-31B.
- Esterbauer H, Puhl H, Dieber-Rothender M. Effect on oxidative modification of LDL. Ann Med. 1991;23:573-81.
- 37. Armstrong D, Browne R. The analysis of free radicals, lipid peroxides, antioxidant enzymes and compounds related to oxidative stress as applied to the clinical chemistry laboratory in Free Radical in Diagnostic Medicine. New York: Plenum Press; 1994. p. 43-58.
- 38. Vasankari T, Kujala U, Heinonen O, Kapanen J, Ahotupa M. Mesurement of serum lipid peroxidation during exercise using three different methods: diene conjugation, thiobarbituric acid reactive material and fluorescent chromolipids. Clin Chim Acta. 1995;234:63-9.
- 39. Jenkins RR. Exercise, oxidative stress, and antioxidants:a review. Int J Sport Nutr. 1993;3:356-75.
- Davies KJA, Quintanilha AT, Brooks GA, Packer L. Free radicals and tissue damage produced by exercise. Biochem Biophys Res Commun. 1982;107:1198-205.
- 41. Ji LL, Fu R. Responses of glutathione system and antioxidant enzymes to exhaustive exercise and hydroperoxide. J Appl Physiol. 1992;72:549-54.
- Maughan RJ, Donnelly AE, Gleeson M, Whiting DH, Walker KA, Clough PJ. Delayed onset muscle damage and lipid peroxidation in man after a downhill run. Muscle Nerve. 1989;12:332-6
- Laaksonen DE, Atalay M, Niskanen L, Uusitupa M, Hanninen O, Sen CK. Increased resting and exercise-induced oxidative stress in young IDDM men. Diabetes Care. 1996;19:569-74.
- Treuth MS, Hunter GR, Williams M. Effects of exercise intensity on 24-h energy expenditure and substrate oxidation. Med Sci Sports Med. 1996;28:1138-43.
- 45. Kanter MM, Nolte LA, Holloszy JO. Effects of an antioxidant vitamine mixture on lipid peroxidation at rest and post-exercise. J Appl Physiol. 1993;74:965-9.
- Viinikka L, Vaoni J, Ylikokala O. Lipid peroxides prostacyclin, and thromboxane A2 in runners during acute exercise. Med Sci Sports Exerc. 1984;16:275-7.
- 47. Neubauer O, Konig D, Kern N, Nics L, Wagner KH. No indications of persistent oxidative stress in response to an ironman triathlon. Med Sci Sports Exerc. 2008;40:2119-28.
- 48. Jenkins RR, Goldfarb. Introduction: oxidant stress, aging, and exercise. Med Sci Sports Exerc. 1993;25:210-2.
- 49. Guezennec CY. Role of lipids on endurance capacity in man. Int J Sports Med. 1992;13:S114-8.
- Turcotte LP, Richter EA, Kiens B. Increased plasma FFA uptake and oxidation during prolonged exercise in trained vs. untrained humans. Am J Physiol. 1992;262(6 Pt 1):E791-9.
- 51. Pruchnic R, Katsiaras A, He J, Kelley DE, Winters C, Goodpaster BH. Exercise training increases intramyocellular lipid and oxidative capacity in older adults. Am J Physiol Endocrinol Metab. 2004;287:E857-62.
- 52. Martin WH 3rd. Effects of acute and chronic exercise on fat metabolism. Exerc Sport Sci Rev. 1996;24:203-31.
- 53. Phillips SM, Green HJ, Tarnopolsky MA, Heigenhauser GF, Hill RE, Grant SM. Effects of training duration on substrate turno-

38 A. Arquer et al

ver and oxidation during exercise. J Appl Physiol. 1996;81:2182-91

- 54. Mittendorfer B, Fields DA, Klein S. Excess body fat in men decreases plasma fatty acid availability and oxidation during endurance exercise. Am J Physiol Endocrinol Metab. 2004;286: E354-62.
- 55. Amati F, Dubé JJ, Shay C, Goodpaster BH. Separate and combined effects of exercise training and weight loss on exercise efficiency and substrate oxidation. JAppl Physiol. 2008;105:825-31
- Calles-Escandón J, Arciero PJ, Gardner AW, Bauman C, Poehlman ET. Basal fat oxidation decreases with aging in women. J Appl Physiol. 1995;78:266-71.
- 57. Boisseau N, Delamarche P. Metabolic and hormonal responses to exercise in children and adolescents. Sports Med. 2000;30: 405-22.
- Alessio HM, Goldfarb A. Lipid peroxidation and scavenger enzymes during exercise. Adaptative response to training. J Appl Physiol. 1988;64:1333-6.
- Sen CK, Marin E, Kretzschmar M, Hanninen O. Skeletal muscle and liver glutathione homeostasis in response to training exercise and immobilization. J Appl Physiol. 1992;73:1265-72.
- Marin E, Kretzschmar M, Arokoski J, Hanninen O, Klinger W. Enzymes of glutathione synthesis in dog skeletal muscles and their response to training. Acta Physiol Scand. 1993;147:369-73
- Powers SK, Criswell D, Lawler J, Martin D, Ji LL, Herb RA, et al. Regional training-induced alterations in diaphragmatic oxidative and antioxidant enzymes. Respir Physiol. 1994;95:227-37.
- Powers SK, Criswell D, Lawler J, Martin D, Lien FK, Ji LL, et al. Rigorous exercise training increases superoxide dismutase activity in ventricular myocardium. Am J Physiol. 1993;265: 2094-8.
- 63. Criswell D, Powers S, Dodd S, Lawler J, Edwuards W, Reushler K, et al. High intensity training-induced changes in skeletal muscle antioxidant enzyme activity. Med Sci Sports Exerc. 1993;25:1140-53.
- 64. Vani M, Reddy GP, Reddy GR, Thyagasaju K, Reddauna P. Glutathione-S-transferase, superoxide dismutase, xanthine oxidase, catalase, glutathione peroxidase and lipid peroxidation in the liver of exercised rats. Biochem Int. 1990;21:17-26.
- 65. Mena P, Maynar M, Gutierrez JM. Erythrocyte free radical scavenger enzymes in bicycle professional racers. Adaptation to training. Int J Sports Med. 1991;12:563-6.
- 66. Covas MI, Elosua R, Fitó M, Alcántara M, Coca L, Marrugat J. Relationship between physical activity and oxidative stress biomarkers in women. Med Sci Sports Exerc. 2002;34:814-9.
- Tiidus PM, Pushkarenko J, Houston ME. Lack of antioxidant adaptation to short-term aerobic training in human muscle. Am J Physiol. 1996;271:832-6.
- 68. Elosua R, Molina L, Fito M, Arquer A, Sanchez-Quesada JL, Covas MI, et al. Response of oxidative stress biomarkers to a 16-week aerobic physical activity program, and to acute physical activity in healthy young men and women. Atherosclerosis. 2003;167:327-34.
- 69. Evelo CT, Palmen NG, Artur Y, Janssen GM. Changes in blood glutathione concentrations and in erythrocyte glutathione reductase and glutathione s-transferase activity after running training and after participation in contests. Eur J Appl Physiol. 1992;64:354-8.
- 70. Sen CK, Packer L. Antioxidant and redox regulation of gene transcription. FASEB J. 1996;10:709-20.
- Sun Y, Oberley LW. Redox regulation of transcriptional activators. Free Radical Biol Med. 1996;21:335-48.
- 72. Roche E, Romero-Alvira D. Papel del estrés oxidativo en la expresión de genes: isquemia miocárdica, cerebral, cáncer y otras enfermedades. Med Clin (Barc). 1995;104:468-76.

- Sanchez-Quesada JL, Homs-Serradesanferm R, Serrat-Serrat J, Serra-Grima JR, González-Sastre F, Ordoñez-Llanos J. Increase susceptibility to oxidation occurring after intense, long duration aerobic exercise. Atherosclerosis. 1995;118:297-305
- 74. Baumstark MW, Frey I, Berg A. Acute and delayed effects of prolonged exercise on serum lipoproteins II. Concentration and composition of low density lipoproteins subfractions and very low density lipoproteins. Eur J Appl Physiol. 1993;66:526-30.
- 75. Lamon-Fava S, McNamara JR, Farber HW, Hill NS, Schaefer EJ. Acute changes in lipid, lipoprotein, apolipoprotein and low-density lipoprotein particle size after and endurance triathlon. Metabolism. 1989:38:921-5.
- 76. Williams PT, Krauss RM, Wood PD, Lindgren FT, Giotas C, Vranizan KM. Lipoproteins subfractions of runners and sedentary men. Metabolism. 1986;35:45-52.
- 77. Sánchez-Quesada JL, Ortega H, Payes-Romero A, Serrat-Serrat J, González-Sastre F, Lasunción MA, et al. LDL from aerobically-trained subjects shows higher resistence to oxidative modification than LDL from sedentary subjects. Atherosclerosis. 1997;132:207-13.
- Williams PT, Krauss RM, Vranizan KM, Wood PDS. Changes in lipoprotein subfractions during diet-induced and exercise-induced weight loss in moderately overweight men. Circulation. 1990;81:1293-304.
- 79. Lofgren I, Zern T, Herron K, West K, Sharman MJ, Volek JS, et al. Weight loss associated with reduced intake of carbohydrate reduces the atherogenicity of LDL in premenopausal women. Metabolism. 2005;54:1133-41.
- 80. Pihl E, Zilmer K, Kullisaar T, Kairane C, Pulges A, Zilmer M. High-sensitivity C-reactive protein and oxidative stress-related status in former athletes in relation to traditional cardiovascular risk factors. Atherosclerosis. 203;171:321-6.
- Vasankari TJ, Kujala UM, Vasankari TM, Ahotupa M. Reduced oxidized LDL levels alter a 10-month exercise program. Med Sci Sports Exerc. 1998;30:1496-501.
- 82. Mosca L, Ruberfire M, Tarshis T, Tsai T, Pearson T. Clinical predictors of oxidized low-density lipoproteins in patients with coronary heart disease. Am J Cardiol. 1997;80:825-30.
- 83. Pihl E, Zilmer K, Kullisaar T, Kairane C, Pulges A, Zilmer M. High-sensitivity C-reactive protein and oxidative stress-related status in former athletes in relation to traditional cardiovascular risk factors. Atherosclerosis. 2003;171:321-6.
- 84. Geffken DF, Cushman M, Burke GL, Polak JF, Sakkinen PA, Tracy RP. Association between physical activity and markers of inflammation in a healthy elderly population. Am J Epidemiol. 2001;153:242-50.
- Ford ES. Does exercise reduce inflammation? Physical activity and C-reactive protein among U.S. adults. Epidemiology. 2002; 13:561-8.
- Abramson JL, Vaccarino V. Relationship between physical activity and inflammation among apparently healthy middle-aged and older US adults. Arch Intern Med. 2002;162:1286-92.
- 87. Wannamethee SG, Lowe GD, Whincup PH, Rumley A, Walker M, Lennon L. Physical activity and hemostatic and inflammatory variables in elderly men. Circulation. 2002;105:1785-90.
- 88. King DE, Carek P, Mainous AG 3rd, Pearson WS. Inflammatory markers and exercise: differences related to exercise type. Med Sci Sports Exerc. 2003;35:575-81.
- 89. Pitsavos C, Chrysohoou C, Panagiotakos DB, et al. Association of leisure-time physical activity on inflammation markers (Creactive protein, white cell blood count, serum amyloid A, and fibrinogen) in healthy subjects (from the ATTICA Study). Am J Cardiol. 2003;91:368-70.
- 90. Mora S, Lee IM, Buring JE, Ridker PM. Association of physical activity and body mass index with novel and traditional cardio-vascular biomarkers in women. JAMA. 2006;295:1412-9.

- 91. Elosua R, Bartali B, Ordovas JM, Corsi AM, Lauretani F, Ferrucci L, InChianti Investigators. Association between physical activity, physical performance and inflammatory biomarkers in an elderly population: the InCHIANTI Study. J Gerontol A Biol Sci Med Sci. 2005;60:760-7.
- 92. Kelley GA, Kelley KS. Effects of aerobic exercise on C-reactive proteins, body composition, and maximum oxygen consumption in adults: a meta-analysis of randomized controlled trials. Metabolism. 2006;55:1500-7.
- 93. Kullo IJ, Khaleghi M, Hensrud DD. Markers of inflammation are inversely associated with ${\rm VO_2}$ max in asymptomatic men. J Appl Physiol. 2007;102:1374-9.
- 94. Olson TP, Dengel DR, Leon AS, Schmitz KH. Changes in inflammatory biomarkers following one-year of moderate resistance training in overweight women. Int J Obes. 2007;31:996-1003.