Serveur Académique Lausannois SERVAL serval.unil.ch

# **Author Manuscript**

**Faculty of Biology and Medicine Publication** 

This paper has been peer-reviewed but does not include the final publisher proof-corrections or journal pagination.

Published in final edited form as:

Title: Lymphoma Classification. Authors: de Leval L, Jaffe ES Journal: Cancer journal (Sudbury, Mass.) Year: 2020 May/Jun Issue: 26 Volume: 3 Pages: 176-185 DOI: 10.1097/PPO.00000000000451

In the absence of a copyright statement, users should assume that standard copyright protection applies, unless the article contains an explicit statement to the contrary. In case of doubt, contact the journal publisher to verify the copyright status of an article.



UNIL | Université de Lausanne Faculty of Biology and Medicine

## The Cancer Journal: The Journal of Principles & Practice of Oncology Lymphoma classification --Manuscript Draft--

Manuscript Number:	
Full Title:	Lymphoma classification
Short Title:	Lymphoma classification
Article Type:	Review Article
Keywords:	Lymphoma; classification; Hodgkin; gray zone; indolent; cutaneous; digestive; WHO
Corresponding Author:	Laurence de Leval, MD pHD Lausanne University Hospital Lausanne, Vaud SWITZERLAND
Corresponding Author Secondary Information:	
Corresponding Author's Institution:	Lausanne University Hospital
Corresponding Author's Secondary Institution:	
First Author:	Laurence de Leval, MD pHD
First Author Secondary Information:	
Order of Authors:	Laurence de Leval, MD pHD
	Elaine S. Jaffe, MD
Order of Authors Secondary Information:	
Manuscript Region of Origin:	SWITZERLAND
Abstract:	Twenty-five years after the Revised European American Lymphoma (REAL) classification was published, its principle of an integrative approach to disease definition based on several parameters still prevails and has been adopted and expanded in the following World Health Organization (WHO) classifications of tumors of the haematopoietic organs. The latest WHO classification revised in 2017 comprises more than 80 entities of mature lymphoid neoplasms (B-cell, T-cell and Hodgkin lymphomas) which are defined according to their morphology, immunophenotype, genetic lesions and molecular profiles, clinical features and cellular derivation. The classification also recognizes both incipient and indolent lymphoid neoplasms with a low potential of proegression. In this review we highlight some of the new data and recent modifications introduced in the 2017 classification.

## Lymphoma classification

Laurence de Leval MD PhD (1) and Elaine S. Jaffe MD (2)

Institute of Pathology, Lausanne University Hospital (CHUV) and Lausanne University, Lausanne, Switzerland (1) and Hematopathology Section, Laboratory of Pathology, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, Maryland (2)

Corresponding author : Laurence de Leval

Professor Laurence de Leval, MD PhD, Institute of Pathology, University Hospital Lausanne (CHUV), 25 rue du Bugnon, CH- 1011 – Lausanne, Switzerland. Phone: +41 21 3147194. Fax: +41 21 3147205.

E-mail: Laurence.deLeval@chuv.ch

### Abstract

Twenty-five years after the Revised European American Lymphoma (REAL) classification was published, its principle of an integrative approach to disease definition based on several parameters still prevails and has been adopted and expanded in the following World Health Organization (WHO) classifications of tumors of the haematopoietic organs. The latest WHO classification revised in 2017 comprises more than 80 entities of mature lymphoid neoplasms (B-cell, T-cell and Hodgkin lymphomas) which are defined according to their morphology, immunophenotype, genetic lesions and molecular profiles, clinical features and cellular derivation. The classification also recognizes both incipient and indolent lymphoid neoplasms with a low potential of proegression. In this review we highlight some of the new data and recent modifications introduced in the 2017 classification.

## Keywords

Lymphoma - classification - Hodgkin - gray zone - indolent - cutaneous - digestive - WHO

#### A historical perspective of lymphoma classification

Few areas in pathology have evoked as much controversy and confusion as the classification of lymphoid neoplasms, dating back to the first attempts to organize the variety of described neoplasms into a comprehensive scheme. The difficulties resulted from several factors, notably the diversity of lymphoid tumors, and the relative insensitivity of routine histopathological techniques that are useful in other organ systems in recognizing defining features of lymphoid cells and their tumors. Later on, in parallel with advances in immunology and genetics, and with the development of novel technologies, the past decades have witnessed major advances in the knowledge of lymphoma pathology, biology and genetics, which translated to novel concepts for the classification of lymphoma (**Figure 1**)<sup>1,2</sup>.

Before 1970, several purely morphologically oriented descriptive classifications of lymphomas were proposed <sup>3-6</sup>. The **Rappaport proposal** (1966) was the first clinically relevant classification<sup>6</sup>. Broadly speaking, Rappaport divided lymphomas into those composed of small cells and of large cells, and further subdivided according to follicular (or nodular) *versus* diffuse growth patterns. The follicular and small-cell tumors were clinically less aggressive; a better survival could therefore be expected and, importantly, less potent and less toxic therapy was suitable for these cases. The converse applied to cases with a diffuse growth pattern, especially if composed of large cells.

Immunologic advances in the 1960's and 1970's revolutionized the understanding of the immune system and lymphomas. The existence of several lymphoid lineages was discovered, and it became possible to establish the phenotype of lymphoma cells using monoclonal antibodies and immunolabeling techniques. Soon it became evident that the lymphoma cells were closely related to normal lymph node cells and that the cells of the non-Hodgkin lymphomas recapitulated the cytology of normal lymphocytes, particularly the B cells of the follicle center. Several new schemes based on these new concepts appeared. The **Kiel Classification**<sup>7,8</sup> developed by Karl Lennert in Germany became widely used in Europe; in this approach primarily intended for classification of nodal diseases, lymphomas were classified according to a hypothetical scheme of B- and T-lymphocyte differentiation, and the nomenclature reflected the putative normal counterpart of the neoplastic cells. According to their cytological characteristics rather than their predetermined clinical behavior and in keeping with established schemes for other tumors, the lymphomas were designated low-grade and high-grade. The classification developed by Lukes and Collins (1974) in the United States was based on similar concepts<sup>9</sup>, and several other proposals appeared shortly after.

Because of the multiplicity of classifications causing much confusion, the compromise **« Working Formulation for Clinical Use »** (NHL Classification Project, 1982) was developed and designed to « translate » between the various classifications<sup>10</sup>. In this scheme, non-Hodgkin lymphomas were divided on clinical grounds into three prognostic groups: low, intermediate and high-grade. Within each group, classification was based purely on morphologic criteria, in particular the nodular versus diffuse growth pattern and the size of the tumor cells (small *versus* large *versus* mixed small and large). As a result, different clinico-pathological entities were lumped together and the Working Formulation did not incorporate immunophenotypic data. In the 1980's and early 1990's, the Working Formulation became the standard American<sup>11</sup> classification, while the Kiel classification was most widely used in Europe. This lack of consensus on lymphoma classification and terminology caused problems for both pathologists and clinicians, and caused difficulty in interpreting published studies. In addition, in the 1980's and 1990's, many new disease entities were described, for example mantle cell lymphoma<sup>12</sup> and

extranodal marginal zone lymphoma<sup>13</sup>, which were not included in either classification. At the same time there were tremendous advances in the understanding of the genetic basis of B-cell lymphomas with the successive discovery of several recurrent translocations and protooncogenes (*BCL2, MYC, CCND1*)<sup>14,15</sup>. The progresses made in immunogenetics with the descriptions of the process of gene rearrangement of immunoglobulin and T-cell receptor genes in normal lymphocytes, and of somatic hypermutations occurring in immunoglobulin genes during transit through the germinal center, were soon exploited to infer clonality and lineage derivation of lymphoid neoplasms, and differentiation stage of B-cell neoplasms <sup>11,16</sup>. Ultimately, the introduction of the new techniques of immunophenotyping and molecular genetic analysis led to confusion about what, if anything, should be the modern "gold standard" for defining disease entities.

In 1991, Peter Isaacson and Harald Stein founded the International Lymphoma Study Group (ILSG) to promote a better communication between hematopathologists from Europe and America. The group composed of 19 prominent hematopathologists from the United States, Europe and Asia at that time, aimed at defining "real" disease entities on which they could reach a consensus based on published data. This approach represented a new paradigm in lymphoma definition and classification. A consensus list of well-defined disease entities was published in 1994 as the REAL (Revised European-American Classification of Lymphoid Neoplasms) classification<sup>1</sup>. Some entities were listed as provisional due to insufficient knowledge or evidence for establishing them as definitive. In the REAL classification, distinct lymphomas defined by a constellation of five properties, namely morphology, immunophenotype, genetic features, clinical features, and if possible the normal cell counterpart from which they derive. The degree to which these properties contribute to the classification of each disease entity is variable. The integration of clinical features into lymphoma definition, which is particularly important for T-cell and NK-cell lymphomas, was a very innovative aspect of the REAL classification. Later on, the notion that the site of lymphoma presentation could represent a signpost for important underlying biologic distinctions was later reflected in the delineation of several "organ-specific" lymphoma entities, notably specific extranodal entities of diffuse large B-cell lymphomas (primary mediastinal, leg-type, of the central nervous system) or follicular lymphomas (cutaneous, duodenaltype, testicular). Although the initial publication of the REAL classification elicited considerable controversy<sup>17</sup>, experience over the following years showed that it could be reproducibly applied by most pathologists and that the entities described have distinctive clinical features, making it a useful and practical classification<sup>18,19</sup>.

Shortly after the publication of the REAL classification, the World Health Organization (WHO) commissioned the European and American Societies of Hematopathology, to provide an update of the classification of hematopoietic and lymphoid neoplasms for its "blue book" series. In addition to many hematopathologists, the joined project which took several years also involved a large clinical advisory committee to address questions of clinical relevance. The principles of disease definition and consensus developed in the REAL classification were essentially maintained. The **WHO classification** of lymphomas published at the end of 2001 <sup>2</sup> represented the first true international consensus on the classification of hematologic malignancies, and gained acceptance worldwide as the reference for pathologists, clinicians, scientists and researchers. Significantly, the monograph belonged to a new series of blue books entitled "pathology and genetics of cancer" emphasizing the contribution of genetics to tumor classification. Since 2001, the WHO classification of tumors of hematopoietic and lymphoid tissues was updated in 2008<sup>20</sup> and the latter edition was revised in 2017<sup>21</sup>. While the original principles still prevail the successive editions have been expanded to incorporate new knowledge and emphasize

novel concepts resulting in an increasing number of entities and amount of information. In the last edition, the "genetic profile" sections are in general markedly enlarged and comprise a lot of new data derived from the many next-generation sequencing studies performed over the past few years, including molecular signatures, (epi)genetic profiles, mutational landscapes and new fusion genes, which mechanistically may correspond to translocations, intrachromosomal deletions or inversions. The genomic landscapes of the most common non-Hodgkin lymphoma entities show wide heterogeneity within and across subtypes, but there are also similar altered pathways and mechanisms of oncogenesis overlapping across different entities.<sup>22</sup>. Various genes belonging to multiple functional pathways are mutated, and the pattern of mutations for each gene is variable: some genes like *BRAF* or *MYD88* are characterized by one or a few hotspot alterations<sup>23,24</sup>, or the mutations may be more heterogeneous. Sometimes distinct mutations in the same gene may be either activating or loss-of-function, and mutations with apparently discordant functional impact may be encountered in different cases of the same disease<sup>25</sup>. Some disease entities are associated to a very characteristic mutational profile, involving one or a few genes, while other diseases are genetically more heterogeneous <sup>26</sup>.

#### The WHO classification of lymphoid neoplasms

In the 2017 classification, more than 80 mature lymphoma entities are recognized, grouped into three major categories: B-cell neoplasms, T-cell and NK-cell neoplasms, and Hodgkin lymphomas (**Table 1**). Both lymphomas and lymphoid leukemias are included in this classification. The disease entities are listed according to predominant clinical presentation (predominantly disseminated diseases that often involve bone marrow and may be leukemic, primary extranodal lymphomas, and predominantly nodal diseases - which are often disseminated and may also involve extranodal sites -). There is an additional section on immunodeficiency-associated lymphoproliferative disorders addressing the spectrum of lymphoproliferations and lymphomas associated with primary immune deficiencies, HIV infection, in the post-transplant setting or in other iatrogenic conditions.

Following the principles of lymphoma classification, lymphoma diagnosis is based on the appreciation of morphologic and immunophenotypic features, genetic characteristics, in the context of clinical presentation. Assignment to a disease category requires the integration of these pieces of data for a final interpretation. In practice, morphologic interpretation combined to immunophenotyping represent the essential elements and are sufficient to establish the diagnosis in many cases. Technical factors in the preparation of the lymph node specimens are of critical importance for obtaining highquality conventional histologic preparations, which are the essential starting point for accurate diagnosis<sup>27</sup>. In principle the immunologic or molecular studies should always be interpreted in the light of the clinical and histopathogical features. Immunophenotype is an essential defining characteristic of lymphomas. In the assessment of malignant lymphoproliferations, immunophenotyping is used (1) to determine cell lineage and differentiation stage, (2) to infer clonality from immunoglobulin light chain expression in the case of B-cell lymphoproliferations, and (3) to identify markers of specific genetic alterations. Immunoglobulin and T-cell receptor gene rearrangement studies are useful to characterize the clonality status of lymphoproliferations, results should again always be interpreted with caution as there are many examples of clonal proliferations that may remain subclinical or indolent. Moreover, pseudoclonal results may be generated from small biopsies with limited infiltrates and PCR amplification of a limited repertoire. Molecular or genetic testing is mandatory to assign a diagnostic category in some instances, for example FISH studies searching for MYC, BCL2 or BCL6 gene rearrangements are necessary for the categorization of high-grade B-cell neoplasms<sup>28</sup>. Next generation sequencing technology has also entered the diagnostic arena, and can provide complementary information to support diagnoses<sup>29</sup>.

#### **B-cell lymphomas**

Mature B-cell neoplasms comprise over 80% of NHL. The most common types are follicular lymphoma and diffuse large B-cell lymphoma, which together account for more than half of all NHL. B-NHL entities comprise a range of morphologically, phenotypically, genetically and clinically distinct malignancies. The majority of B-NHL harbor somatically mutated IG genes and therefore are derived from germinal center (GC) or post-GC B cells. The correspondence to normal B-cell subpopulations initially assessed by a combination of morphology and immunophenotype, has been refined by the comparison of gene expression signatures of lymphomas and normal B cell subpopulations across the full range of normal B-cell differentiation. **Figure 2** illustrates the model of histogenesis and pathogenesis of B-cell neoplasms.

Several entities of B-cell neoplasms represent early or indolent clonal lymphoid lesions. Monoclonal B-cell lymphocytosis and monoclonal gammopathy of unknow significance are the subclinical precursor lesions to B-chronic lymphocytic leukemia and plasma cell myeloma, respectively <sup>30</sup>. *In situ* follicular neoplasia (not uncommon) and *in situ* mantle cell lymphoma (distinctively rare) denote tissue accumulation restricted to germinal centers of mantle zones, of B cells harboring the *BCL2* or *CCND1* hallmark translocations of follicular or mantle cell lymphoma<sup>31</sup>. *In situ* follicular neoplasia is considered the tissue equivalent of circulating blood cells harboring the t(14;18) translocation found in many healthy individuals; compared to clinically overt follicular lymphoma the load of additional genetic alterations is low in *in situ* follicular neoplasia<sup>32</sup>, and the risk of progression to clinical disease is low<sup>33</sup>. Duodenal-type follicular-type is somewhat assimilated to *in situ* follicular neoplasia, because this t(14;18)+ neoplasm which arises in the mucosa of the proximal duodenum is often discovered incidentally and has an indolent course with a very low risk of local progression or dissemination<sup>34</sup>.

Major changes have been introduced in the classification and diagnosis of aggressive B-cell lymphomas, which fall into three main categories: diffuse large B-cell lymphomas (DLBCLs), Burkitt lymphoma (BL), and two novel entities of high-grade B-cell lymphomas<sup>35</sup>. DLBCLs constitutes the most prevalent group and comprises many entities unified by a simple definition, i.e. a proliferation of large transformed B cells. Specific entities are related to occurrence in peculiar anatomic sites (primary mediastinal, primary cutaneous leg-type, primary DLBCL of the central nervous system, intravascular), some are classified based on a plasmablastic phenotype (plasmablastic lymphoma and ALK-positive DLBCL) or an abundant microenvironment (T-cell/histiocyte-tich large B-cell lymphoma), others are defined by their association to viral infections +/- a peculiar clinical context (EBV+ DLBCL, EBV+ muco-cutaneous ulcer, DLBCL with chronic inflammation, lymphomatoid granulomatosis, primary effusion lymphoma, HHV8+ DLBCL-NOS). For the usual form of DLBCL ("NOS") it is recommended to identify the two molecular subtypes defined by gene expression profiling, germinal center-like and activated B-cell like, because these two subtypes correspond to distinct pathogenetic pathways. In a routine setting, this is achieved in most centers by the use of immunohistochemical algorithms or simplified RNA profiling methods<sup>36-39</sup>. Besides Burkitt lymphoma which is a very homogeneous category, a new provisional entity named Burkitt-like lymphoma with 11q aberration has been introduced, referring to a limited number of cases resembling Burkitt lymphoma, but lacking a MYC translocation, having more complex karyotypes and a chromosome 11q aberration<sup>40</sup>. The categories of high-grade B-cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements, and high-grade B-cell lymphoma, not otherwise specified, were newly introduced. These new categories partly substitute for the former group of "unclassifiable B-cell lymphoma with features intermediate between diffuse large B-cell lymphoma and Burkitt lymphoma", which is no longer recognized. High-grade B-cell lymphomas may morphologically resemble DLBCL, have features intermediate between DLBCL and Burkitt lymphoma (high proliferation fraction and a starry sky appearance with cells larger or more pleomorphic than the spectrum of classical Burkitt lymphoma, or a morphology resembling Burkitt lymphoma with a discordant immunophenotype, typically strong BCL2 expression), or have a blastoid morphology. A sizeable fraction of high-grade B-cell lymphomas with MYC and BCL2 and/or BCL6 rearrangements, also simply termed «double hit» or «triple hit» lymphomas, represent transformed follicular lymphomas. MYC plus BCL2 double hit lymphomas are usually GCB and those with MYC and BCL6 rearrangement are usually ABC. High-grade B-cell lymphomas NOS by definition do not have double or triple hits, but a significant proportion carries a MYC rearrangement (alone). An important consequence of this new classification is that assignement of a large B-cell neoplasm to the DLBCL category requires genetic testing at least for MYC +/- BCL2 and BCL6 rearrangements, in order to exclude the possibility of a "double hit" lymphoma.

#### **T-cell lymphomas**

Neoplasms of mature NK or T cells (PTCLs) are overall rare. They comprise approximately 10% of all lymphomas in Western world, while in Asia where EBV-associated extranodal NK/T-cell lymphoma (and adult T-cell leukemia/lymphoma) are more frequent, they represent up to 20% of lymphomas. The delineation of mature NK/T-cell neoplasms into entities along the WHO scheme is not as sharp as for the B-cell tumors, in regard to the numerous T-cell subsets and their functional plasticity. Indeed many T/NK-cell neoplasms have a broad range of cellular composition, and immunophenotypic profiles may be heterogeneous within a disease category while overlapping across different entities. Recent advances in characterizing the molecular profiles and mutational landscapes of these diseases have translated into significant changes in the classification.

With regard to nodal PTCLs (**Figure 3**), a major change was the grouping of angioimmunoblastic T-cell lymphoma and two related entities, namely follicular T-cell lymphoma and nodal T-cell lymphoma with a T follicular helper phenotype, under a common heading named "nodal lymphomas derived from follicular helper T cells (TFH cells). The rationale is that these three entities have in common not only an immunophenotype and gene expression signature similar to those normal TFH cells, but also a similar genetic landscape, characterized by recurrent mutations in epigenetic modifier genes (*TET2, DNMT3A, IDH2*), *RHOA* and other T-cell receptor signaling genes <sup>41-48</sup>. Nodal T-cell lymphoma with a T follicular helper phenotype is defined as a PTCL lacking the peculiar morphological features characteristic of AITL, but showing expression of several (at least two, optimally at least three) TFH markers, and this entitiy includes many cases categorized in the past as "Lennert lymphomas" (rich in epithelioid cells) or nodal T-zone lymphomas. Anaplastic large cell lymphoma (ALCL), ALK-negative, is now viewed as a genetically heterogeneous entity comprising at least two subsets defined by recurrent translocations affecting *DUSP22* (about 30% of the cases, and associated to a good prognosis similar to that of ALK-positive ALCL) or *TP63* (10% of the cases, associated with a poor outcome) <sup>49,50</sup>. Since *DUSP22*-rearranged cases may not fulfill all the phenotypic attributes of

ALK-negative ALCL, their identification is important to clarify the sometimes difficult differential diagnosis with CD30-positive PTCL-NOS. In addition, a proportion of ALK- ALCL also carry mutations impacting on the JAK/STAT pathway<sup>51</sup>, a feature also reported in primary cutaneous ALCL<sup>51</sup> and breast implant-associated ALCL <sup>52-54</sup>. The latter was introduced as a provisional entity in the classification due to its peculiar clinical presentation usually as an effusion and less commonly as a tumor mass around a breast implant, and in the light of its peculiar epidemiology and usually indolent clinical course<sup>55,56</sup>. PTCL-NOS remains a diagnosis of exclusion - notably implying the exclusion of a TFH expression - and a heterogeneous category. Two molecular subtypes based on gene expression profiles related to Th1 or TH2 subsets of helper T cells, have been defined and can be identified by a surrogate immunohistochemistry-based algorithm<sup>57,58</sup>. A small subset of primary nodal PTCL-NOS are associated to EBV infection of the neoplastic cells; these cases differ from extranodal NK/T-cell lymphoma, nasal type as they tend to be more monomorphic, usually lack angioinvasion and necrosis, and do not express CD56<sup>59</sup>.

#### Hodgkin lymphomas and gray zone lymphomas

Hodgkin lymphomas (HLs)differ from most other lymphomas in their unique cellular composition comprising a minority of large atypical neoplastic cells (Hodgkin and Reed-Sternberg (HRS) cells and their variants) outnumbered by non-neoplastic reactive cells<sup>60</sup>. After decades of uncertainties and speculations regarding the nature of this malignancy, genetic molecular studies conducted on microdissected HRS cells established that HLs are neoplasms of large B cells and represent clonal populations of transformed germinal center B cells <sup>61-66</sup>. Remarkably, however, the historical distinction between Hodgkin's disease and non-Hodgkin lymphomas survived the successive classifications of lymphomas over time - reflecting not only the peculiar morphological range of HL but also its distinctive clinical presenting features and distinctive responses to certain chemotherapeutic combinations and radiation therapy.

Classic HLs (cHLs) and nodular lymphocyte predominant HL (NLPHL) represent two distinctive groups of diseases (**Table 2**). In cHL subclassified into four subtypes according to the cellular composition of the background infiltrate, the neoplastic Hodgkin and Reed-Sternberg (HRS) which may harbor the Epstein-Barr virus (EBV) cells are typically positive for CD30 (an activation marker) and for CD15 (an antigen associated with the granulocyte and monocyte lineages), while they usually lack expression of CD45 and of molecules characteristic of the B and T-cell lineages<sup>67</sup>. In contrast, in NLPHL, the neoplastic LP "popcorn" cells, have a B-cell immunophenotype <sup>68</sup>, and in its classical form develop within large follicular structures<sup>69</sup>. Six distinct histological patterns of NLPHL (A to F) have been recognized<sup>70</sup> and should be described in the diagnosis since the "non-classical" patterns (C to F) tend to correlate with higher disease stage and a higher risk of relapse<sup>71</sup>. There are clinical similarities and some morphologic overlap between the lymphocyte-rich subtype of cHL and NLPHL, which explains a high rate of misdiagnoses in the past<sup>72</sup>. However, despite the facts that the neoplastic cells in lymphocyte-rich cHL tend to have preserved expression of B-cell markers and develop in a follicular microenvironment rich in PD1-positive cells, similar to NLPHL <sup>73</sup>the two entities are separated by other key distinctive features.

The term "gray-zone lymphomas" was coined to denote lymphoproliferations rich in large cells and difficult to classify due to morphological or immunophenotypic overlapping features between Hodgkin and non-Hodgkin lymphomas, suggesting the possibility of a biological overlap or continuum<sup>74</sup>. Some of the differential diagnoses considered under the "gray-zone" umbrella, in particular anaplastic large cell lymphoma versus cHL, have been resolved with the refinement of diagnostic criteria and the use of expanded immunohistochemistry and molecular genetic testing. Currently, there remains essentially two gray zone areas, one each around cHL and NLPHL.

The category of B-cell lymphoma, unclassifiable, with features intermediate between diffuse large B-cell lymphoma and cHL, which was a provisional entity in the 2008 classification, was promoted to a definitive status in 2017. This entity mostly applies mostly to mediastinal cases showing both overlapping features between nodular sclerosis HL and primary mediastinal large B-cell lymphoma, and significant discordances between morphology and immunophenotype preventing from allocating the case to one of each of the well-defined entities<sup>75</sup>. The patients who are often young males, tend to experience a clinical course more aggressive than those with either Hodgkin or large B-cell lymphoma<sup>76</sup>. The notion of a biological continuum is supported by the partial overlap in the molecular and genetic features of mediastinal cHL, large B-cell lymphoma and gray-zone cases<sup>77-79</sup>. Recent studies have highlighted considerable variability among pathologists towards the diagnosis of mediastinal gray zone lymphomas, and the lack of consensus criteria<sup>79,80</sup>. EBV-positive cases should be excluded from the gray zone category as it turns out that these cases are either syncytial forms of EBV-positive nodular sclerosis HL, or part of the spectrum of EBV-positive diffuse large B-cell lymphomas.

The relationship and borderline between NLPHL and T-cell/histiocyte-rich large B-cell lymphoma has been addressed abundantly in the literature and remains a matter of debate<sup>81</sup>. The diffuse pattern of NLPHL which is often T-cell-rich, closely resembles de novo T-cell/histiocyte-rich large B-cell lymphoma not only on morphological grounds, but also at the transcriptional and genomic levels, with common recurrent mutations in *DUSP2*, *JUNB*, *SGK1* and *SOCS1*, and tends to follow a somewhat aggressive course<sup>82,83</sup>. However, it is recommended in the WHO classification to restrict the diagnosis of T-cell/histiocyte-rich large B-cell lymphoma to de novo cases, and to employ the term "T-cell/histiocyte-rich large B-cell lymphoma-like transformation" in cases progressing from NLPHL or arising in association with the classical nodular pattern of NLPHL <sup>21</sup>.

#### Lymphomas of the digestive system

Lymphomas involving the gastrointestinal (GI) tract account for about 30 to 40% of extranodal lymphomas <sup>84,85</sup>. Most cases correspond to entities that are not specific to the GI tract, and a smaller fraction correspond to few specific tumor entities primarily arising in the gastrointestinal tract, including duodenal-type follicular lymphoma and intestinal T-cell lymphomas. The latest classification introduced significant changes in the intestinal T-cell lymphoma comprises four categories: enteropathy-associated T-cell lymphoma (EATL), formerly type I EATL and linked to celiac disease and frequent in Northern Europe; monomorphic epitheliotropic intestinal T-cell lymphoma (MEITL), formerly type II EATL, not associated to celiac disease and relatively more frequent in Asia; intestinal T-cell lymphoma NOS, and indolent T-cell lymphoproliferative disorder of the GI tract<sup>21</sup>. The two subtypes of EATL previously considered as variants of the same disease, have been reclassified as two distinct diseases (**Figure 4**) due to differences in their epidemiology and clinical, features, morphology and phenotype, and genetic features as highlighted in recent works<sup>56,86,87</sup>. Intestinal T-cell lymphoma NOS is a category created to comprise the cases of primary intestinal T-cell lymphomas not meeting the criteria for either EATL or MEITL. Indolent T-cell lymphoproliferative disorder of the GI tract represent a novel category of clonal

monomorphic proliferations of small T cells showing a superficial/mucosal distribution in the colon or anywhere along the GI tract, usually characterized by an indolent behavior and a chronic relapsing course<sup>88,89</sup>. Recent works have demonstrated an association with recurrent gene fusions or point mutations<sup>90,91</sup>. All the recent updates of the 2017 lymphoma classification are also included in a specific section of the blue book on digestive system tumours dedicated to hematolymphoid malignancies<sup>92</sup>.

#### **Cutaneous lymphomas**

Primary cutaneous lymphomas represent the second most common form of extranodal lymphomas. They comprise a heterogenous group of lymphomas that present in the skin without extracutaneous disease at the time of diagnosis. Primary cutaneous T-cell lymphomas represent the vast majority of the cases while B-cell lymphomas account for 20-25% primary cutaneous lymphomas. Several primary cutaneous lymphoma entities like mycosis fungoides or lymphomatoid papulosis, have highly characteristic clinical and histological features; conversely some lymphomas histologically similar to systemic lymphomas like follicular lymphoma or diffuse large B-cell lymphomas, often have a completely different clinical behavior and prognosis when occurring primarily in the skin. Because of these specificities, primary cutaneous lymphomas have been considered separately from systemic lymphomas in different classification systems. The consensus classification of the WHO-EORTC (World health Organization -European Organization for Research and Treatment of Cancer) published the first time in 2005 and recently revised, serves as the reference for the classification and diagnosis of primary cutaneous lymphomas<sup>93,94</sup>. The approach and principles for disease definition and diagnosis are similar to those used for systemic lymphomas, with increased contribution of genetic and molecular markers in the last edition, but perhaps more importantly than for systemic lymphomas, it must be stressed that clinical data remain essential for a correct diagnosis. For the most part, the terminology and defining criteria of the different primary cutaneous lymphoma entities of the updated WHO-EORTC classification were already introduced in the 2017 WHO classification of tumors of haematopoietic tissues<sup>21</sup>. The latest edition of the WHO blue book on skin tumors published in 2018<sup>95</sup>, also incorporates the revised WHO-EORTC classification more completely with regards to the distinction of two different subtypes of cutaneous marginal zone lymphomas, one class-switched and comprising a small proportion of neoplastic B cells that are CXCR3-negative, and the other characterized by large nodules of neoplastic cells that are IgM- and CXCR3-positive<sup>96-98</sup>.

#### Table 1

2017 WHO classification of mature lymphomas (adapted from <sup>21</sup>)

#### **B-cell Neoplasms**

#### Predominantly disseminated

- Chronic lymphocytic leukemia/ B-cell small lymphocytic lymphoma
- Monoclonal B-cell lymphocytosis,
- B-cell prolymphocytic leukemia
- Splenic marginal zone B-cell lymphoma
- Hairy cell leukemia
- Splenic B-cell lymphoma/leukemia, unclassifiable\*
- Splenic diffuse red pulp small B-cell lymphoma\*
- Hairy cell leukemia-variant\*
- Lymphoplasmacytic lymphoma
- Waldenström macroglobulinemia
- Monoclonal gammopathy of unknown significance (MGUS) IgM
- Mu heavy-chain disease
- Gamma heavy-chain disease
- Alpha heavy-chain disease
- Monoclonal gammopathy of unknown significance (MGUS) IgG/A+
- Plasma cell myeloma
- Solitary plasmacytoma of bone
- Extraosseous plasmacytoma
- Monoclonal immunoglobulin deposition disease

#### Primary extranodal or nodal

- Extranodal marginal zone B-cell lymphoma of mucosa-associated lymphoid tissue
- Nodal marginal zone B-cell lymphoma
  - Pediatric nodal marginal zone lymphoma\*
- Follicular lymphoma
  - In situ follicular neoplasia
  - Duodenal-type follicular lymphoma
- Pediatric-type follicular lymphoma
- Large B-cell lymphoma with IRF4 rearrangement\*
- Primary cutaneous follicle center lymphoma
- Mantle cell lymphoma
  - In situ mantle cell neoplasia
- Diffuse large B-cell lymphoma (DLBCL)-NOS Germinal center B-cell type
  - Activated B-cell type
- T-cell/histiocyte-rich large B-cell lymphoma
- Primary DLBCL of the central nervous system
- Primary cutaneous DLBCL, leg-type
- EBV+ DLBCL-NOS
- EBV+ muco-cutaneous ulcer

- DLBCL associated with chronic inflammation
- Lymphomatoid granulomatosis
- Mediastinal (thymic) large B-cell lymphoma
- Intravascular large B-cell lymphoma
- ALK+ large B-cell lymphoma
- Plasmablastic lymphoma
- Primary effusion lymphoma
- HHV8+ DLBCL-NOS\*
- Burkitt lymphoma
- Burkitt-like lymphoma with 11q aberration\*
- Lymphomatoid granulomatosis
- High-grade B-cell lymphoma with MYC and BCL2 and/or BCL6 gene rearrangements\*
- High-grade B-cell lymphoma NOS\*
- B-cell lymphoma unclassifiable with features intermediate between DLBCL and classic Hodgkin lymphoma

#### T and NK-cell Neoplasms

#### Predominantly disseminated

- T-cell prolymphocytic leukemia
- T-cell large granular lymphocytic leukemia
- Aggressive NK-cell leukemia
- Chronic lymphoproliferative disorder of NK cells\*
- Systemic EBV-positive T-cell lymphoma of childhood\*
- Chronic active EBV infection of T- and NK-cell type, systemic form
- Adult T-cell lymphoma/leukemia (HTLV-1+)

#### Primary extranodal

- Extranodal NK/T-cell lymphoma, nasal type
- Enteropathy-associated T-cell lymphoma
- Monomorphic epitheliotropic intestinal T-cell lymphoma
- Indolent T-cell lymphoproliferative disorder of the gastro-intestinal tract\*
- Hepatosplenic T-cell lymphoma
- Subcutaneous panniculitis-like T-cell lymphoma
- Breast implant-associated anaplastic large cell lymphoma\*
- Subcutaneous panniculitis-like T-cell lymphoma

#### Primary cutaneous

- Mycosis fungoides
- Sezary syndrome
- Primary cutaneous CD30 + lymphoproliferative disorders Lymphomatoid papulosis
  - Primary cutaneous anaplastic large cell lymphoma
- Primary cutaneous  $\gamma\delta$  T-cell lymphoma
- Provisional CD4+ or CD8+ entities\*
- Hydroa vacciniforme-like lymphoproliferative disorder
- Severe mosquito bite allergy

#### Predominantly nodal

- Peripheral T-cell lymphoma, not otherwise specified
- Angioimmunoblastic T-cell lymphoma
- Follicular T-cell lymphoma
- Nodal peripheral T-cell lymphoma with T follicular helper phenotype
- Anaplastic large cell lymphoma, ALK-positive
- Anaplastic large cell lymphoma, ALK-negative

#### Hodgkin lymphomas

• Nodular lymphocyte predominant Hodgkin lymphoma

Classic Hodgkin lymphoma
 Nodular sclerosis classic Hodgkin lymphoma
 Mixed cellularity classic Hodgkin lymphoma
 Lymphocyte-rich classic Hodgkin lymphoma
 Lymphocyte-depleted classic Hodgkin lymphoma

#### Immunodeficiency-associated lymphoproliferative disorders

Post-transplantation lymphoproliferative disorders (PTLD) Non-destructive PTLD plasmacytic hyperplasia infectious mononucleosis PTLD florid follicular hyperplasia PTLD Polymorphic PTLD Monomorphic PTLD Classic Hodgkin Lymphoma PTLD Other iatrogenic immunodeficiency-associated lymphoproliferative disorders

\* provisional entities

Figure legends

#### Figure 1

#### Evolution of lymphoma classifications (adapted from <sup>99</sup>)

#### Figure 2

#### Histogenetic and pathogenetic model of mature B-cell neoplasms

The follicular (T-cell dependent) and extrafollicular pathways of normal B-cell differentiation are represented. Rearrangement of the IG heavy chain gene is symbolized with a horizontal bar in the nucleus, and somatic mutations in the IGV region are symbolized with vertical lines. The most common types of mature B-cell neoplasms are linked to their putative normal counterpart. The genetic lesion most frequently associated with each lymphoma category is also indicated.

#### Figure 3

#### Overview of the main peripheral T-cell lymphoma entities presenting as nodal diseases

The three groups of nodal lymphomas depicted on the scheme account for the majority of cases presenting with nodal involvement. The category of peripheral T-cell lymphoma not otherwise specified (PTCL-NOS) is a diagnosis of exclusion, especially after having discarded the possibility of a TFH lymphoma (expressing at least two TFH markers), and after having excluded the possibility of nodal involvement by a disseminated T-cell neoplasm (T-cell prolymphocytic leukemia, adult T-cell leukemia/lymphoma) or an extranodal or cutaneous lymphoma. PTCL-NOS is a heterogenous category comprising several subsets defined by gene expression signatures resembling those of T Helper 1 or 2 cells (T-BET and GATA-3, respectively), EBV-positive tumors and neoplasms with a cytotoxic phenotype.

AITL: angioimmunoblastic T-cell lymphoma; ALCL: anaplastic large cell lymphoma; ALK: anaplastic lymphoma kinase; F-TCL: follicular T-cell lymphoma; PTCL-TFH PTCL with follicular helper T-cell phenotype.

#### Figure 4

#### Evolution of the categorization of intestinal T-cell lymphomas

Primary intestinal T-cell lymphomas were collectively considered in the 2008 classification as one disease entity, enteropathy-associated T-cell lymphoma, of which two variants, type I (classical), and type II were recognized. In the 2017 classification, these two variants are considered as separate diseases, and designated enteropathy-associated T-cell lymphoma (EATL) (former type I) and monomorphic epitheliotropic intestinal T-cell lymphoma (MEITL) (former type II). EATL is typically pleomorphic, often associated with an inflammatory component and necrosis. MEITL as its name implies, is composed of a monomophic lymphoid cell proliferation and infiltrates the adjacent epithelium of intestinal villi and crypts. In addition to EATL and MEITL, the current classification also recognizes a subset of cases not fulfilling the criteria for either EATL or MEITL, designated "intestinal T-cell lymphoma, not otherwise specified" as well as a category of indolent clonal T-cell

lymphoproliferative disorders. Moreover, other lymphoma types may present with intestinal involvement (for example extrandoal NK/T-cell lymphoma, ALK+ anaplastic T-cell lymphoma).

## References

1. Harris NL, Jaffe ES, Stein H, et al. A revised European-American classification of lymphoid neoplasms: a proposal from the International Lymphoma Study Group. *Blood.* 1994;84(5):1361-1392.

2. Jaffe E, Harris N, Stein H, Vardiman J. *Pathology and genetics. Tumours of haematopoietic and lymphoid tissues.* Lyon: IARC Press; 2001.

3. Robb-Smith AHT. Reticulosis and reticulosarcoma. A histological classification. *J Path Bact.* 1938;47:457-480.

4. Gall EA, Mallory TB. Malignant lymphoma. A clinicopathologic survey of 618 cases. *Am J Pathol.* 1942;18:381-395.

5. Jackson H, Jr., Parker F, Jr. Hodgkin's disease II: pathology. N Engl J Med. 1944;231:35-44.

6. Rappaport H. Tumors of the hematopoietic system. In. *Atlas of Tumor Pathology*. Vol Section III. Series I ed. Washington, DC: Armed Forces Institute of Pathology; 1966:.

7. Lennert K, Mohri N, Stein H, Kaiserling E. The histopathology of malignant lymphoma. *Br J Haematol.* 1975;31 [suppl]:193-203.

8. Stansfeld AG, Diebold J, Noel H, et al. Updated Kiel classification for lymphomas [letter] [published erratum appears in Lancet 1988 Feb 13;1(8581):372]. *Lancet*. 1988;1(8580):292-293.

9. Lukes R, Collins R. Immunologic characterization of human malignant lymphomas. *Cancer*. 1974;34:1488-1503.

10. National Cancer Institute sponsored study of classifications of non-Hodgkin's lymphomas: summary and description of a working formulation for clinical usage. The Non-Hodgkin's Lymphoma Pathologic Classification Project. *Cancer.* 1982;49(10):2112-2135.

11. Arnold A, Cossman J, Bakhshi A, al e. Immunoglobulin gene rearrangements as unique clonal markers in human lymphoid neoplasms. *N Engl J Med.* 1983;309:1594-1599.

12. Banks P, Chan J, Cleary M, et al. Mantle cell lymphoma: a proposal for unification of morphologic, immunologic, and molecular data. *Am J Surg Pathol.* 1992;16:637-640.

13. Isaacson P, Wright D. Malignant lymphoma of mucosa associated lymphoid tissue. A distinctive B cell lymphoma. *Cancer*. 1983;52:1410-1416.

14. Dalla-Favera R, Bregni M, Erikson J, Patterson D, Gallo RC, Croce CM. Human c-myc onc gene is located on the region of chromosome 8 that is translocated in Burkitt lymphoma cells. *Proc Natl Acad Sci U S A*. 1982;79(24):7824-7827.

15. Yunis JJ, Oken MM, Kaplan ME, Ensrud KM, Howe RR, Theologides A. Distinctive chromosomal abnormalities in histologic subtypes of non-Hodgkin's lymphoma. *N Engl J Med*. 1982;307(20):1231-1236.

16. Kuppers R, Klein U, Hansmann ML, Rajewsky K. Cellular origin of human B-cell lymphomas. *N Engl J Med.* 1999;341(20):1520-1529.

17. Rosenberg SA. Classification of lymphoid neoplasms. *Blood*. 1994;84(5):1359-1360.

18. A clinical evaluation of the International Lymphoma Study Group classification of non-Hodgkin's lymphoma. The Non-Hodgkin's Lymphoma Classification Project. *Blood.* 1997;89(11):3909-3918.

19. Armitage JO, Weisenburger DD. New approach to classifying non-Hodgkin's lymphomas: clinical features of the major histologic subtypes. Non-Hodgkin's Lymphoma Classification Project. *J Clin Oncol.* 1998;16(8):2780-2795.

20. Swerdlow S, Campo E, Harris N, et al. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues, Fourth Edition.* Lyon: IARC Press; 2008.

21. Swerdlow SH, Campo E, Harris NL, et al. *WHO classification of tumours of haematopoietic and lymphoid tissues*. Lyon: International Agency for Research on Cancer; 2017.

22. Moffitt AB, Dave SS. Clinical Applications of the Genomic Landscape of Aggressive Non-Hodgkin Lymphoma. *J Clin Oncol.* 2017;35(9):955-962.

23. Tiacci E, Trifonov V, Schiavoni G, et al. BRAF mutations in hairy-cell leukemia. *N Engl J Med.* 2011;364(24):2305-2315.

24. Treon SP, Xu L, Yang G, et al. MYD88 L265P somatic mutation in Waldenstrom's macroglobulinemia. *N Engl J Med.* 2012;367(9):826-833.

25. Nagata Y, Kontani K, Enami T, et al. Variegated RHOA mutations in adult T-cell leukemia/lymphoma. *Blood.* 2016;127(5):596-604.

26. Pasqualucci L, Trifonov V, Fabbri G, et al. Analysis of the coding genome of diffuse large B-cell lymphoma. *Nature genetics*. 2011;43(9):830-837.

27. Banks P. Technical factors in the preparation and evaluation of lymph node biopsies. In: Knowles D, ed. *Neoplastic hematopathology*. Philadelphia: Lipincott Williams & Wilkins; 2001:467-482.

28. Swerdlow SH, Campo E, Pileri SA, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood.* 2016;127(20):2375-2390.

29. Sujobert P, Le Bris Y, de Leval L, et al. The Need for a Consensus Next-generation Sequencing Panel for Mature Lymphoid Malignancies. *Hemasphere*. 2019;3(1):e169.

30. Karube K, Scarfo L, Campo E, Ghia P. Monoclonal B cell lymphocytosis and "in situ" lymphoma. *Semin Cancer Biol.* 2014;24:3-14.

31. Jegalian AG, Eberle FC, Pack SD, et al. Follicular lymphoma in situ: clinical implications and comparisons with partial involvement by follicular lymphoma. *Blood*. 2011;118(11):2976-2984.

32. Schmidt J, Salaverria I, Haake A, et al. Increasing genomic and epigenomic complexity in the clonal evolution from in situ to manifest t(14;18)-positive follicular lymphoma. *Leukemia*. 2014;28(5):1103-1112.

33. Pillai RK, Surti U, Swerdlow SH. Follicular lymphoma-like B cells of uncertain significance (in situ follicular lymphoma) may infrequently progress, but precedes follicular lymphoma, is associated with other overt lymphomas and mimics follicular lymphoma in flow cytometric studies. *Haematologica*. 2013;98(10):1571-1580.

34. Mamessier E, Song JY, Eberle FC, et al. Early lesions of follicular lymphoma: a genetic perspective. *Haematologica*. 2014;99(3):481-488.

35. Rimsza L, Pittaluga S, Dirnhofer S, et al. The clinicopathologic spectrum of mature aggressive B cell lymphomas. *Virchows Arch.* 2017;471(4):453-466.

36. Hans CP, Weisenburger DD, Greiner TC, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a tissue microarray. *Blood*. 2004;103(1):275-282.

37. Coutinho R, Clear AJ, Owen A, et al. Poor concordance among nine immunohistochemistry classifiers of cell-of-origin for diffuse large B-cell lymphoma: implications for therapeutic strategies. *Clin Cancer Res.* 2013;19(24):6686-6695.

38. Scott DW, Wright GW, Williams PM, et al. Determining cell-of-origin subtypes of diffuse large B-cell lymphoma using gene expression in formalin-fixed paraffin-embedded tissue. *Blood.* 2014;123(8):1214-1217.

39. Mareschal S, Ruminy P, Bagacean C, et al. Accurate Classification of Germinal Center B-Cell-Like/Activated B-Cell-Like Diffuse Large B-Cell Lymphoma Using a Simple and Rapid Reverse Transcriptase-Multiplex Ligation-Dependent Probe Amplification Assay: A CALYM Study. *J Mol Diagn.* 2015.

40. Salaverria I, Martin-Guerrero I, Wagener R, et al. A recurrent 11q aberration pattern characterizes a subset of MYC-negative high-grade B-cell lymphomas resembling Burkitt lymphoma. *Blood.* 2014;123(8):1187-1198.

41. de Leval L, Rickman DS, Thielen C, et al. The gene expression profile of nodal peripheral T-cell lymphoma demonstrates a molecular link between angioimmunoblastic T-cell lymphoma (AITL) and follicular helper T (TFH) cells. *Blood*. 2007;109(11):4952-4963.

42. de Leval L, Savilo E, Longtine J, Ferry JA, Harris NL. Peripheral T-cell lymphoma with follicular involvement and a CD4+/bcl-6+ phenotype. *Am J Surg Pathol.* 2001;25(3):395-400.

43. Lemonnier F, Couronne L, Parrens M, et al. Recurrent TET2 mutations in peripheral T-cell lymphomas correlate with TFH-like features and adverse clinical parameters. *Blood*. 2012;120(7):1466-1469.

44. Palomero T, Couronne L, Khiabanian H, et al. Recurrent mutations in epigenetic regulators, RHOA and FYN kinase in peripheral T cell lymphomas. *Nature genetics*. 2014;46(2):166-170.

45. Sakata-Yanagimoto M, Enami T, Yoshida K, et al. Somatic RHOA mutation in angioimmunoblastic T cell lymphoma. *Nature genetics*. 2014;46(2):171-175.

46. Vallois D, Dobay MP, Morin RD, et al. Activating mutations in genes related to TCR signaling in angioimmunoblastic and other follicular helper T-cell-derived lymphomas. *Blood*. 2016;128(11):1490-1502.

47. Vallois D, Dupuy A, Lemonnier F, et al. RNA fusions involving CD28 are rare in peripheral T-Cell lymphomas and concentrate mainly in those derived from follicular helper T cells. *Haematologica*. 2018.

48. Dobay MP, Lemonnier F, Missiaglia E, et al. Integrative clinicopathological and molecular analyses of angioimmunoblastic T-cell lymphoma and other nodal lymphomas of follicular helper T-cell origin. *Haematologica*. 2017;102(4):e148-e151.

49. Parilla Castellar ER, Jaffe ES, Said JW, et al. ALK-negative anaplastic large cell lymphoma is a genetically heterogeneous disease with widely disparate clinical outcomes. *Blood.* 2014.

50. Pedersen MB, Hamilton-Dutoit SJ, Bendix K, et al. DUSP22 and TP63 rearrangements predict outcome of ALK-negative anaplastic large cell lymphoma: a Danish cohort study. *Blood*. 2017;130(4):554-557.

 Crescenzo R, Abate F, Lasorsa E, et al. Convergent mutations and kinase fusions lead to oncogenic STAT3 activation in anaplastic large cell lymphoma. *Cancer cell*. 2015;27(4):516-532.
 Oishi N, Brody GS, Ketterling RP, et al. Genetic subtyping of breast implant-associated anaplastic large cell lymphoma. *Blood*. 2018.

53. Letourneau A, Maerevoet M, Milowich D, et al. Dual JAK1 and STAT3 mutations in a breast implant-associated anaplastic large cell lymphoma. *Virchows Arch.* 2018.

54. Laurent C, Nicolae A, Laurent C, et al. Gene alterations in epigenetic modifiers and JAK-

STAT signaling are frequent in breast implant-associated ALCL. *Blood.* 2020;135(5):360-370.
Laurent C, Delas A, Gaulard P, et al. Breast implant-associated anaplastic large cell lymphoma: two distinct clinicopathological variants with different outcomes. *Ann Oncol.*

2016;27(2):306-314.

56. de Leval L. Breast implant-associated anaplastic large cell lymphoma and other rare T-cell lymphomas. *Hematol Oncol.* 2019;37 Suppl 1:24-29.

57. Iqbal J, Wright G, Wang C, et al. Gene expression signatures delineate biological and prognostic subgroups in peripheral T-cell lymphoma. *Blood*. 2014;123(19):2915-2923.

58. Amador C, Greiner TC, Heavican TB, et al. Reproducing the Molecular Subclassification of Peripheral T-cell Lymphoma-NOS by Immunohistochemistry. *Blood*. 2019.

59. Ng SB, Chung TH, Kato S, et al. Epstein-Barr virus-associated primary nodal T/NK-cell lymphoma shows a distinct molecular signature and copy number changes. *Haematologica*. 2018;103(2):278-287.

60. Reed D. On the pathological changes in Hodgkin's disease, with especial reference to its relation to tuberculosis. *Johns Hopkins Hosp Rep.* 1902;10:133-196.

61. Kuppers R, Schwering I, Brauninger A, Rajewsky K, Hansmann ML. Biology of Hodgkin's lymphoma. *Ann Oncol.* 2002;13 Suppl 1:11-18.

62. Kuppers R, Rajewsky K, Zhao M, et al. Hodgkin disease: Hodgkin and Reed-Sternberg cells picked from histological sections show clonal immunoglobulin gene rearrangements and appear to be derived from B cells at various stages of development. *Proc Natl Acad Sci U S A*. 1994;91(23):10962-10966.

63. Kanzler H, Kuppers R, Hansmann ML, Rajewsky K. Hodgkin and Reed-Sternberg cells in Hodgkin's disease represent the outgrowth of a dominant tumor clone derived from (crippled) germinal center B cells. *J Exp Med.* 1996;184(4):1495-1505.

64. Braeuninger A, Kuppers R, Strickler JG, Wacker HH, Rajewsky K, Hansmann ML. Hodgkin and Reed-Sternberg cells in lymphocyte predominant Hodgkin disease represent clonal populations of germinal center-derived tumor B cells. *Proc Natl Acad Sci U S A*. 1997;94(17):9337-9342.

65. Marafioti T, Hummel, M.D., Anagnostopoulos I, et al. Origin of nodular lymphocytepredominant Hodgkin's disease from a clonal expansion of highly mutated germinal-center B cells. *The New England Journal of Medicine*. 1997;337(7):453-458.

66. Ohno T, Stribley JA, Wu G, Hinrichs SH, Weisenburger DD, Chan WC. Clonality in nodular lymphocyte-predominant Hodgkin's disease. *N Engl J Med.* 1997;337(7):459-465.

67. von Vasielewski R, Mengel M, Fischer R, et al. Classical Hodgkin's disease: clinical impact of the immunophenotype. *Am J Pathol.* 1997;151:1123-1130.

68. Falini B, Bigerna B, Pasqualucci L, et al. Distinctive expression pattern of the BCL-6 protein in nodular lymphocyte predominance Hodgkin's disease. *Blood.* 1996;87(2):465-471.

69. Hansmann ML, Stein H, Dallenbach F, Fellbaum C. Diffuse lymphocyte-predominant Hodgkin's disease (diffuse paragranuloma). A variant of the B-cell-derived nodular type. *Am J Pathol.* 1991;138(1):29-36.

70. Fan Z, Natkunam Y, Bair E, Tibshirani R, Warnke RA. Characterization of variant patterns of nodular lymphocyte predominant hodgkin lymphoma with immunohistologic and clinical correlation. *Am J Surg Pathol.* 2003;27(10):1346-1356.

71. Hartmann S, Eichenauer DA, Plutschow A, et al. The prognostic impact of variant histology in nodular lymphocyte-predominant Hodgkin lymphoma: a report from the German Hodgkin Study Group (GHSG). *Blood.* 2013;122(26):4246-4252; quiz 4292.

72. Diehl V, Sextro M, Franklin J, et al. Clinical presentation, course, and prognostic factors in lymphocyte- predominant Hodgkin's disease and lymphocyte-rich classical Hodgkin's disease: report from the European Task Force on Lymphoma Project on Lymphocyte-Predominant Hodgkin's Disease [see comments]. *J Clin Oncol.* 1999;17(3):776-783.

73. Nam-Cha SH, Montes-Moreno S, Salcedo MT, Sanjuan J, Garcia JF, Piris MA. Lymphocyterich classical Hodgkin's lymphoma: distinctive tumor and microenvironment markers. *Mod Pathol.* 2009;22(8):1006-1015.

74. Rudiger T, Jaffe ES, Delsol G, et al. Workshop report on Hodgkin's disease and related diseases ('grey zone' lymphoma). *Ann Oncol.* 1998;9 Suppl 5:S31-38.

75. Traverse-Glehen A, Pittaluga S, Gaulard P, et al. Mediastinal Gray Zone Lymphoma: The Missing Link Between Classic Hodgkin's Lymphoma and Mediastinal Large B-Cell Lymphoma. *Am J Surg Pathol.* 2005;29(11):1411-1421.

76. Wilson WH, Pittaluga S, Nicolae A, et al. A prospective study of mediastinal gray zone lymphoma. *Blood.* 2014.

77. Eberle FC, Rodriguez-Canales J, Wei L, et al. Methylation profiling of mediastinal gray zone lymphoma reveals a distinctive signature with elements shared by classical Hodgkin's lymphoma and primary mediastinal large B-cell lymphoma. *Haematologica*. 2011;96(4):558-566.

78. Eberle FC, Salaverria I, Steidl C, et al. Gray zone lymphoma: chromosomal aberrations with immunophenotypic and clinical correlations. *Mod Pathol.* 2011;24(12):1586-1597.

79. Sarkozy C, Molina T, Ghesquieres H, et al. Mediastinal gray zone lymphoma: clinicopathological characteristics and outcomes of 99 patients from the Lymphoma Study Association. *Haematologica*. 2017;102(1):150-159.

80. Pilichowska M, Pittaluga S, Ferry JA, et al. Clinicopathologic consensus study of gray zone lymphoma with features intermediate between DLBCL and classical HL. *Blood Adv*. 2017;1(26):2600-2609.

81. Harris NL. Shades of gray between large B-cell lymphomas and Hodgkin lymphomas: differential diagnosis and biological implications. *Mod Pathol.* 2013;26 Suppl 1:S57-70.

82. Hartmann S, Doring C, Jakobus C, et al. Nodular lymphocyte predominant hodgkin lymphoma and T cell/histiocyte rich large B cell lymphoma--endpoints of a spectrum of one disease? *PLoS One.* 2013;8(11):e78812.

83. Hartmann S, Schuhmacher B, Rausch T, et al. Highly recurrent mutations of SGK1, DUSP2 and JUNB in nodular lymphocyte predominant Hodgkin lymphoma. *Leukemia*. 2016;30(4):844-853.

84. Howell JM, Auer-Grzesiak I, Zhang J, Andrews CN, Stewart D, Urbanski SJ. Increasing incidence rates, distribution and histological characteristics of primary gastrointestinal non-Hodgkin lymphoma in a North American population. *Can J Gastroenterol.* 2012;26(7):452-456.

85. Andrews CN, John Gill M, Urbanski SJ, Stewart D, Perini R, Beck P. Changing epidemiology and risk factors for gastrointestinal non-Hodgkin's lymphoma in a North American population: population-based study. *Am J Gastroenterol.* 2008;103(7):1762-1769.

86. Roberti A, Dobay MP, Bisig B, et al. Type II enteropathy-associated T-cell lymphoma features a unique genomic profile with highly recurrent SETD2 alterations. *Nat Commun.* 2016;7:12602.

87. Moffitt AB, Ondrejka SL, McKinney M, et al. Enteropathy-associated T cell lymphoma subtypes are characterized by loss of function of SETD2. *J Exp Med.* 2017;214(5):1371-1386.

88. Perry AM, Warnke RA, Hu Q, et al. Indolent T-cell lymphoproliferative disease of the gastrointestinal tract. *Blood*. 2013;122(22):3599-3606.

89. Matnani R, Ganapathi KA, Lewis SK, Green PH, Alobeid B, Bhagat G. Indolent T- and NK-cell lymphoproliferative disorders of the gastrointestinal tract: a review and update. *Hematol Oncol.* 2017;35(1):3-16.

90. Sharma A, Oishi N, Boddicker RL, et al. Recurrent STAT3-JAK2 fusions in indolent T-cell lymphoproliferative disorder of the gastrointestinal tract. *Blood.* 2018;131(20):2262-2266.

91. Soderquist CR, Patel N, Murty VV, et al. Genetic and phenotypic characterization of indolent T-cell lymphoproliferative disorders of the gastrointestinal tract. *Haematologica*. 2019.

92. Cree IA, Centre international de recherche sur le cancer, Organisation mondiale de la santé. *WHO classification of tumours of the digestive system.* 5th ed.

93. Willemze R, Jaffe ES, Burg G, et al. WHO-EORTC classification for cutaneous lymphomas. *Blood.* 2005;105(10):3768-3785.

94. Willemze R, Cerroni L, Kempf W, et al. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. *Blood*. 2019;133(16):1703-1714.

95. Elder DE, Organisation mondiale de la santé, Centre international de recherche sur le cancer. *WHO classification of skin tumours.* 4th ed.

96. van Maldegem F, van Dijk R, Wormhoudt TA, et al. The majority of cutaneous marginal zone B-cell lymphomas expresses class-switched immunoglobulins and develops in a T-helper type 2 inflammatory environment. *Blood.* 2008;112(8):3355-3361.

97. Edinger JT, Kant JA, Swerdlow SH. Cutaneous marginal zone lymphomas have distinctive features and include 2 subsets. *Am J Surg Pathol.* 2010;34(12):1830-1841.

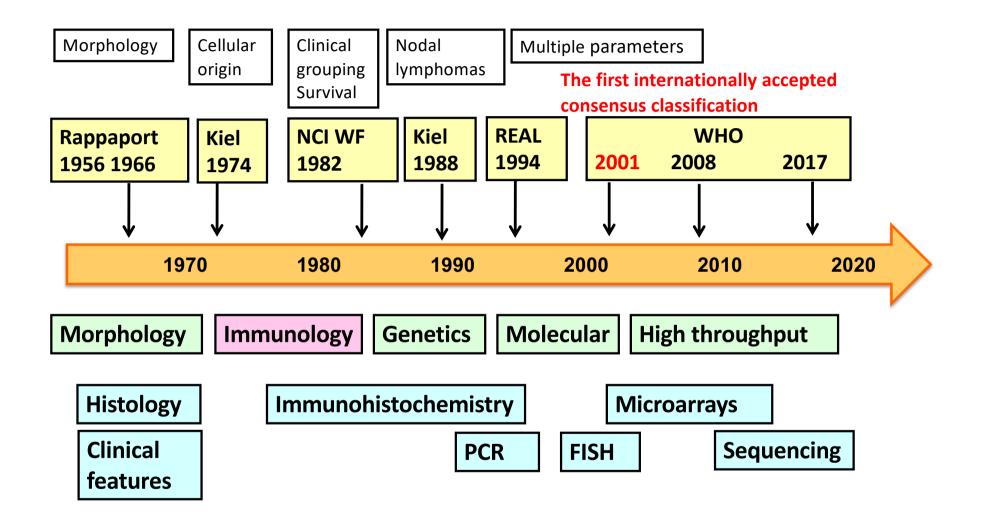
98. Swerdlow SH. Cutaneous marginal zone lymphomas. *Semin Diagn Pathol.* 2017;34(1):76-84.
99. Jaffe E, Arber D, Campo E, Harris N, Quintanilla-Martinez L. *Hematopathology*.
Philadelphia: Elsevier; 2017.

Table

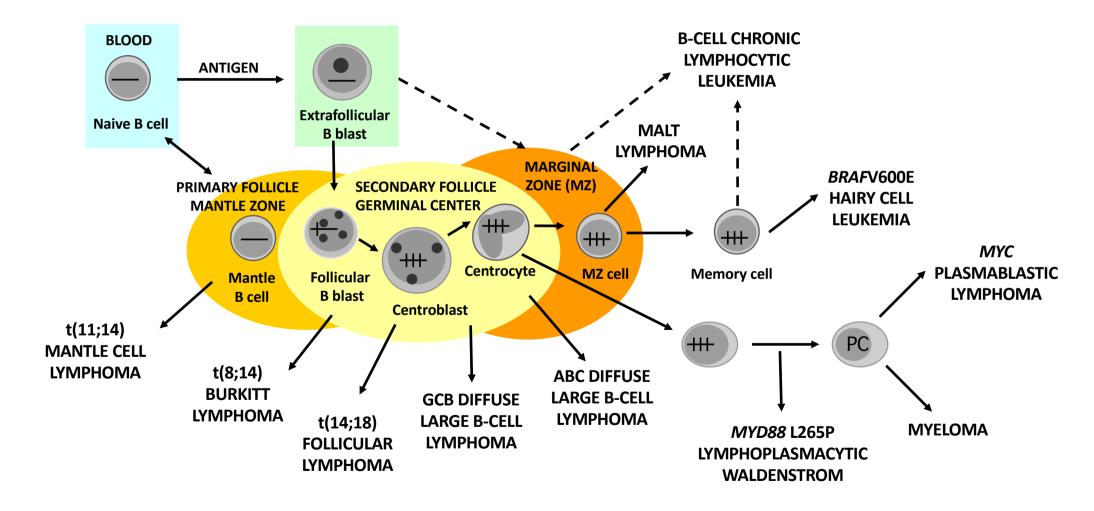
## Table 2

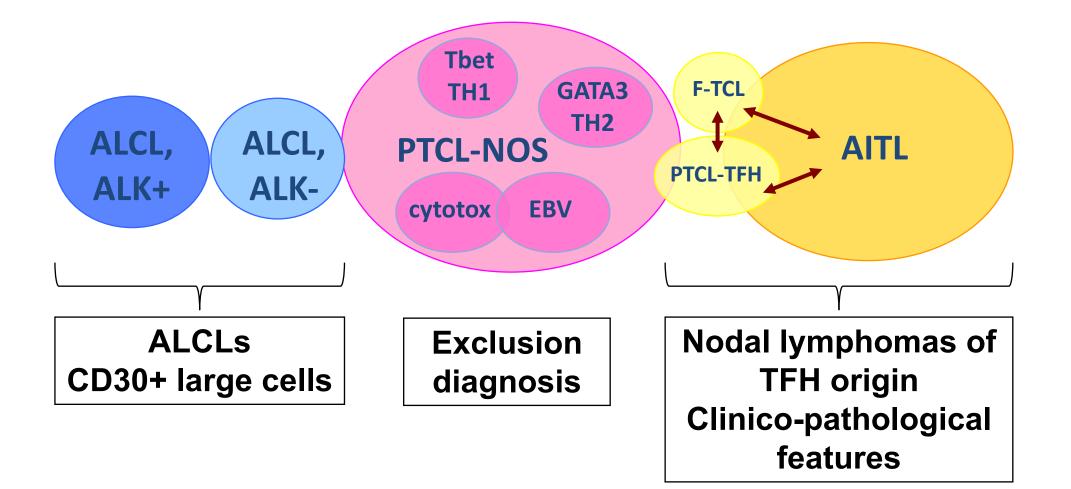
Summary of distinguishing featrures of classic and nodular lymphocyte predomionant Hodgkin lymphomas

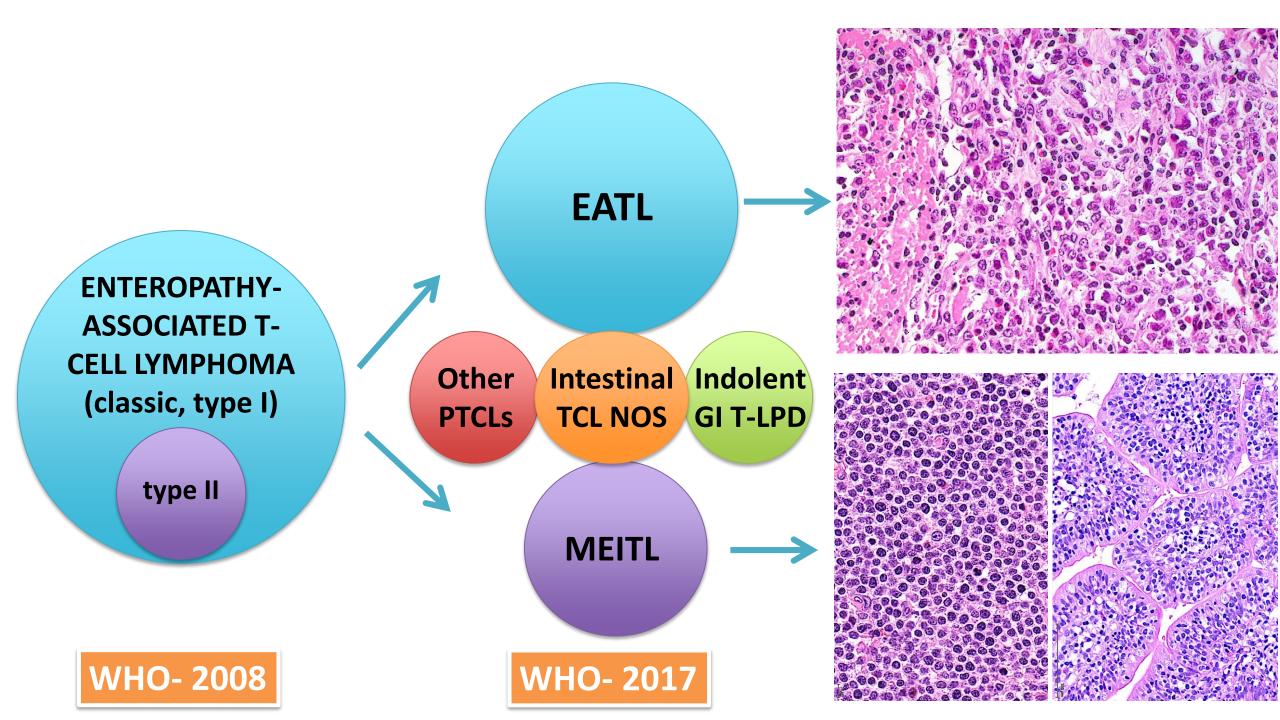
Classic Hodgkin Lymphoma (cHL) > 90% of HL	Nodular Lymphocyte Predominant Hodgkin Lymphoma (NLPHL)
HRS cells: CD20-/+ PAX5+ CD30+ CD15+/- BCL6- MUM1+ BOB1- OCT2- EBV+ (40%)	LP cells CD20+ PAX5+ CD30- CD15- BCL6+ MUM1+ BOB1+ OCT2+ EBV-
Four types: Nodular Sclerosis and Mixed Cellularity most common; Lymphocyte-rich, or Lymphocyte–depleted less common	Six patterns: classical B-cell rich nodular (A), serpiginous/interconnexted (B); with promiennt extranodular LP cells (C); T-cell rich nodular (D); diffuse (T-cell/histiocyte-rich large B-cell lymphoma like) (E); diffuse B-cell-rich (F).
IG: clonal, mutated, crippled mutations Mutations in <i>STAT6, SOCS1</i> ; <i>PDL1/PDL2</i> alterations, <i>CIITA</i> rearrangements	IG: clonal, mutated BCL6 rearrangements, mutations in DUSP2, JUN, SGK1, SOCS1
Gray zone: unclassifiable B-cell lymphoma with features intermediate between diffuse large B-cell lymphoma and cHL	Gray zone: diffuse NLPHL versus T-cell/histiocyte-rich large B-cell lymphoma



Manuscript (All Manuscript Text Pages in MS Word format, including References and Figure Legends)







Manuscript (All Manuscript Text Pages in MS Word format, including References and Figure Legends)

Click here to access/download **Copyright Transfer and Disclosure Form** NIH Publishers Cover Sheet de leval and Jaffe.doc