we are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

5,300 Open access books available 130,000

International authors and editors

155M

154 Countries delivered to Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Therapeutic Potential of IL-9 in Allergic and Autoimmune Diseases

Ahmed Ummey Khalecha Bintha, Amani Souwelimatou Amadou, Mursalin Md Huzzatul and Muhammad Fauziyya

Abstract

Interleukin-9 (IL-9) is a pleiotropic cytokine produced by several immune and epithelial cells. Recently, many studies have eluded the physiological and pathological roles of IL-9 and its lineage-specific helper T cell subset (Th9). In this chapter, we will focus on the immunological role of Interleukin 9 (IL-9) in allergy and autoimmunity. We will introduce the basics of IL-9 and describe the cells involved in the secretion, signaling, and regulation of IL-9. After establishing the background, we will discuss the pathogenesis and regulation of IL-9 in allergic and autoimmune diseases. We will conclude the chapter by providing an updated therapeutics that target IL-9 and their potential uses in autoimmune and allergic diseases.

Keywords: IL-9, Th9, multiple sclerosis, Th17, IBD, uveitis, mast cells, asthma, atopic dermatitis, food allergy, diabetes, TGF- β , ILC2

1. Introduction

Interleukin-9 (IL-9) is a pleotropic cytokine that regulates diverse immunological functions (**Figure 1**). This cytokine was first identified in the late 1980s as a T cell growth factor [1]. Because of the molecular weight of IL-9, it was initially known as P40 [2]. Later studies revealed that the observed molecular weight was due to N-link glycosylation, and actual molecular weight for this discovered molecule is 14 kDa [3]. A similar factor was also identified from Th2 cells and mast cells where it was initially named as T-cell growth Factor III (TCGF III) and mast cell growth-enhancing activity (MEA), respectively [2, 4]. Further studies revealed that both TCGF III and MEA actually represent the P40 factor [4]. In later years, considering its pleotropic roles and the redundant nomenclature the P40 factor was renamed as IL-9 [5].

The locus encoding IL9 in mouse is about 11 kb in size, and located on chromosome 13 [6]. The II9 locus is comprised of 5 exons and 4 introns [3]. The II9 locus encode for a precursor peptide of 144 amino acids, first 18 amino acids of which is signal sequence peptide. The mature IL-9 peptide, a single-chain glycoprotein of 126 amino acids, and similar to other cytokines of IL-2 family folds into a fouralpha-helix bundles [7]. Human IL-9 locus is present on chromosome 5 in the region q31–35 [6]. Homology between mouse and human IL-9 is about 55%, and both of them contain a conserved 10 cysteine residue to form a disulfide bond that is critical for a mature IL-9 peptide. Interestingly, three conserved non-coding sequences,

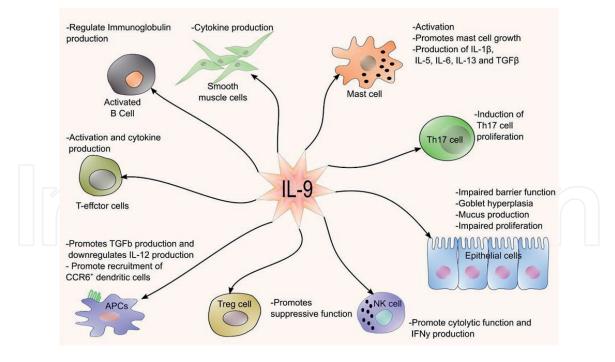


Figure 1.

Functions of IL-9. IL-9 contributes to different immunopathology and physiology through activation of multiple cell types. Illustration by MHuzzatul.

CNS0, CNS1, and CNS2 are present on both mouse and human *il9* locus sequence similarity of which is 63% [3, 7]. CNS0 is positioned in the upstream (-6 kb) of transcription start site (TSS), CNS1 is the promoter region, and CNS2 is located at the downstream of TSS (+5.4 kb) [8]. CNS1 provide binding site to numerous transcription factors that includes PU.1, STAT5, STAT6, GATA1, GATA3, IRF1, IRF4, NF-kb, BATF, AP-1, Smads 2/3/4, Gcn5, Notch [9]. Etv5 can bind to both CNS0 and CNS2, and recruit histone acetyltransferase p300 to mediate chromatin remodeling [8–11]. Regulation of IL-9 expression by this multiple numbers of transcription factors explain the necessity of a delicate cytokine milieu that requires to stimulate IL-9 producing cells. The miscellaneous origin of IL-9 and the complexity of its regulation underscore the need for a comprehensive assessment of IL-9 function. Therefore, in this chapter, we will elucidate the basis of IL-9 function in health and diseases and its therapeutic potentials in autoimmune and allergic diseases.

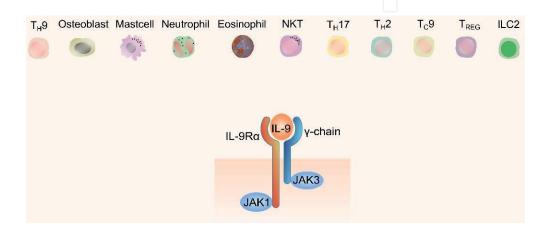
2. IL-9, a lineage specific Th9 cytokine

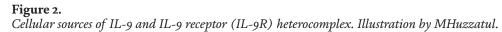
T cells were originally thought to be the main source of IL-9 [12–14]. IL-9 was defined as a Th2 cytokine. The reason for this Th2 designation by many research findings included IL-9 genome. The *il-9* gene is positioned within a Th2 cytokine clusters. Also, increased expression of IL-9 was observed in a Th2-predominate BALB/c mouse model of cutaneous leishmaniasis (BALB/c mice) but not in Th1-predominate model (using C57BL/6 mice). This finding suggested IL-9 as a Th2 signature cytokine [12]. In addition, Th2-like responses such as airway epithelial hyperplasia, proliferation of mast cells, mucin-producing cells, and eosinophils were found in the lungs of IL-9 transgenic mice [15]. More recently, the designation of IL-9 as a Th2 cytokine loses credence, due to the identification of PU.1, an ETS family transcription factor that induces IL-9 secretion. Mice with T-cell-specific deletion of PU.1 did not develop IL-9 dependent inflammation of the lungs [16]. However, the mice had similar frequencies of Th2 cells [16]. In another experiment that utilized siRNA-mediated disruption of PU.1 resulted in impaired IL-9

production in human T-cells. Recently, a distinct helper T cell subset, Th9 was identified as IL-9 lineage-specific cells. Studies observed increased PU.1 expression under Th9 polarizing conditions but not Th2 conditions [16]. The finding of another helper T cell subset suggested that Th2 is not the main source for IL-9, and PU.1 as a unique transcription factor necessary for IL-9 production emphasized the identity of Th-9. Later, *in vitro* studies identified IL-4 and TGF- β as cytokines that facilitate the differentiation of naïve T cells to Th9 cells [17, 18]. Though IL-4 is a known Th2 cytokine, TGF- β exhibit pleotropic functions and regulates the development of other helper T cells including Th17 and Treg cells [19]. Presence of IL-4 with TGF- β facilitates the differentiation of naive T cells into IL-9-secreting Th9 but not Tregs or Th17. Also, IL-4 can directly block the expression of FoxP3 in T cells thus reprogramming Treg cells into Th9 cells [17]. And, addition of TGF- β in culture medium reprograms Th2 cells to Th9 cells [18]. IL-4 and TGF- β -mediated induction of IL-9-producing cells are dependent on both activated STAT6 and GATA3, suggesting the initial identification of IL-9 as a Th2 cytokine. And Th2 including other helper T cells secrete small amounts of IL-9 [20].

3. Sources of IL-9

In addition to Th9 and Th2, other immune cells have been identified as potential sources of IL-9 (Figure 2). Prominent among these immune cells is Th17 cells. Th17 cells are involved in mounting immune responses against extracellular bacteria and fungi and are implicated in autoimmunity [21]. Activation of a Th17-associated transcription factor, retinoic acid receptor-related orphan receptor-yt (RORyt) with phorbol 12-myristate 13-acetate and ionomycin (PMA) leads to IL-9 secretion [22]. Tregs have also be shown to secrete IL-9 both in vivo and in vitro, however, the role is IL-9-secreting Tregs is conflicting [23, 24]. Another recently identified source of IL-9 is V δ 2 T cells in human peripheral blood. This $\gamma\delta$ T cell subset population can be stimulated with antigens, TGF- β , and IL-15 to produce IL-9 [24]. Mast cells, natural killer T cells (NKT) have also been found to produce IL-9. Mast cells cross-linked with IgE and inflammatory mediators like histamine produce IL-9 in the presence of IL-1 β and LPS [25–29]. Stimulation of NKT cells with IL-2 leads to secretion of IL-9 [30]. A large number of infiltrating IL-9 producing NKT has been found in histological section from patient with nasal NKT cell lymphomas [31]. Decreased expression of IL-9 was observed in CD1d-restricted NKT deficient mouse model of allergic inflammation suggesting NKT cell can also promote IL-9 production in vivo [32]. In addition, innate lymphoid cells such as ILC2s, eosinophils, neutrophils, and osteoblasts also have been found to produce IL-9 [33-35].





4. IL-9 receptor signaling

IL-9 exerts its biological effect on its target cells through IL-9R receptor. The IL-9R is a heterocomplex of the alpha chain (IL-9R α) and the common gamma chain [36]. IL-9R α is specific only to IL-9, whereas the gamma chain is present in the receptor complexes of several other cytokines such as IL-2, IL-4, IL-7, IL-13, IL-15, and IL-21 [37–39]. About 25% of the IL-9R α exist in complex with the gamma chain outside IL-9 heterocomplex. IL-9R α is of 522 amino acids in human, and 468 amino acids in mouse, and contains 11 exons [40]. This 64 kDa glycoprotein is a member of type I hematopoietin receptor super family due to the presence of the Box1 and Box2 motifs in the intracellular domain, and WSXWS motif in the extracellular domain [41]. Formation of a heterocomplex with the γ-chain is enhanced as IL-9 binds to IL-9R α (**Figure 2**) [42]. The binding of IL-9 to IL-9R α results in a conformational change in IL-9R. This conformational change recruit JAK molecules to Box1 motif which results in the phosphorylation of tyrosine residues of IL-9R α -associated JAK1 and γ-chain associated JAK3 [41]. BOX1 motif is very critical in IL-9 mediated signaling as disruption of Box1 results in loss of

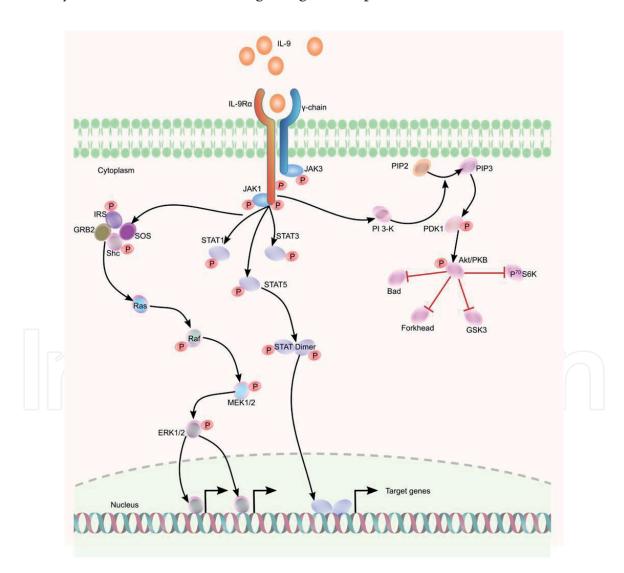


Figure 3.

Schematic representation of IL-9 signaling pathway. IL-9 cytokine binds to IL-9R complex. This leads to phosphorylation of JAKs. The phosphorylated JAKs activate STATs, PI3 kinase, and the MAP kinase pathway. IL-9R, interleukin-9 receptor; JAK, Januse kinase; STAT, signal transducer and activator of transcription; PI3K, phosphatidylinositol-3 kinase; PIP, phosphoinositide; PDK1, pyruvate dehydrogenase kinase 1; bad, GSK3, glycogen synthase kinase 3; PS6K, IRS, insulin receptor substrate; SOS, suppressors of cytokine signaling; GRB2, ERK, extracellular signal regulated kinase; Shc; Ras/Raf/MEK, mitogen-activated protein kinases; illustration by MHuzzatul.

phosphorylation of JAK1 and JAK3 [43]. Activated JAK molecules then phosphorylate a tyrosine residue (Tyr407) in the IL-9R α , which results in the phosphorylation of intermediate molecules, STAT molecules (STAT1, STAT3, and STAT5), MAPK, and IRS-PI3 pathways (**Figure 3**) [44–46]. Activation of these pathways contribute to the upregulation of IL-9, as well as important in the growth, differentiation, and development of the IL-9 targeted cells [47, 48].

5. IL-9 and allergic diseases

Allergic diseases including respiratory, food, and skin allergies are mainly mediated by Th2 cells through the expression of various cytokines such as IL-4, IL-5, and IL-13 (reviewed in [49]). The cytokine IL-9, which was initially studied in the context of Th2-mediated immune response and later associated with T-helper 9 (Th9) cells, has been shown to play an important role in allergic inflammation [50, 51]. IL-9 and its receptor IL-9R α regulate antibody synthesis, specifically IgE, in both murine and human B cells [52, 53]. To contribute to allergic disease pathogenesis, IL-9 also promotes activation and recruitment of inflammatory cells [54–57].

6. Asthma including airway allergies

Various studies have shown that IL-9 and its receptor contribute to airway allergic diseases and asthma. Sputum, serum, and lungs of patients with asthma were shown to have increased concentrations of the cytokine [58–60]. IL-9 levels were also increased in the airways of murine asthma models [61]. IL-9Rα is expressed on human tonsillar germinal center and memory B cells, and smooth muscles in the airways. IL-9/ IL-9R α signaling in B cells induces STAT3 and STAT5 pathways to potentiate IgE production [52, 53, 55, 62, 63]. Overexpression of IL-9 in transgenic mice or treatment with recombinant cytokine induces expansion of B-1 cells, and accumulation of mast cells in the tissues [64, 65]. IL-9 induces the release of proteases and pro-inflammatory cytokines by the mast cells to promote survival of eosinophils and increase airway permeability [66, 67]. IL-9/IL-9Rα signaling also stimulates human airway smooth muscle to secrete eotaxin1/CCL1 and induces production of IL-13 in airway epithelial cells. Eotaxin1/CCL11 and IL-13 significantly increase eosinophil recruitment and cause lung epithelial cell hypertrophy. These effects result in asthma-like symptoms, including lung inflammation, bronchial hyper-responsiveness, and mucus accumulation. Moreover, IL-9 worsens lung injury in a murine model of chronic obstructive pulmonary disease (COPD) [63, 68, 69]. The cytokine also appears to be a critical player in allergic rhinitis. Serum IL-9 in patients strongly correlates with irritative nasal symptoms including rhinorrhea [70]. In mice, Th9 cells are significantly upregulated during allergic rhinitis and neutralization of IL-9 alleviates symptoms. Blocking IL-9 decreases the level of inflammatory cytokines (IFN-γ, IL-4, and IL-17) and eosinophils infiltration in the nasal mucosa. This causes a decrease in the frequency of sneezing and nasal rubs in experimental models of allergic rhinitis [71].

7. Food allergies

Studies in patients with food allergy and experimental oral hypersensitivity have shown that allergic reactions in the gastrointestinal tract are mediated by various players, including Th2-secreted cytokines, such as IL-4 and IL-9 [72–74]. Various

studies have shown that IL-9 drives intestinal inflammation and plays a critical role in food allergies [75, 76]. In patients with food allergies, the severity of clinical symptoms strongly correlates with increased intestinal permeability [77]. In vitro experiments have shown that patients with peanut allergy have increased levels of IL-9. The memory T helper cell response specific to peanuts in allergic children is dominated by IL-9. Thus, cytokine levels can be used as a biomarker to determine individuals with peanut allergy [78, 79]. In mice, overexpression of intestinal IL-9 or induction of IL-9-producing mucosal mast cells (MMC9s) also increases susceptibility to food allergy [80]. Migration of mast cell progenitors and their development into MMC9s is regulated by basic leucine zipper transcription factor ATF-like (BATF) and Th2-secreted IL-4 [81]. The large amount of MCC9s-derived IL-9 and other mast cell mediators cause intestinal mastocytosis and increased intestinal permeability, which is central to the induction of experimental oral hypersensitivity [82]. The actions of the IL-9-stimulated mast cells cause allergic diarrhea and hypothermia [75]. IL-9 can additionally be secreted by the group 2 innate lymphoid cells (ILC2) and Th9 cells to amplify the intestinal allergic inflammatory response, which may lead to anaphylaxis [83–88].

8. Skin allergies

IL-9 has been identified as a potential mediator of cutaneous allergies, including atopic dermatitis (AD) and allergic contact dermatitis (ACD). Patients with atopic dermatitis have a significantly higher level of IL-9 in the serum and skin lesions [89]. The concentration of the cytokines also positively correlates with the severity of the disease and serum IgE levels [90]. These observations were made in both adult and pediatric patients [91, 92]. A study in a Korean population also linked IL-9 and IL-9R gene polymorphisms to AD [93]. IL-9 induces IL-5 and IL-13 by ILC2. ILC2 and the cytokines are associated with AD pathogenesis. IL-5 and IL-13 contribute to the defective skin barrier in AD patients by downregulating tight junctions genes [94, 95]. IL-9 also promotes the secretion of the vascular endothelial growth factor (VEGF) by keratinocytes and mast cells [92, 96]. An increased level of VEGF contributes to the dilatation of capillaries, erythema, and inflammatory edema characteristics of AD [97, 98]. Moreover, IL-9 has been shown to regulate Th1-mediated allergic contact dermatitis. Patients with positive patch tests to nickel have a higher level of allergen-specific IL-9 expression in skin, peripheral blood mononuclear cells (PBMCs). Also, IL-9 potentially mediates infiltration of eosinophils in the skins as its levels strongly correlate with the cell infiltration in the tissues. This demonstrates a potential pathogenic role of the cytokine IL-9 in ACD [99, 100].

9. IL-9 and autoimmunity

The etiology or trigger of autoimmune diseases is not well understood [101, 102]. However, there is a consensus that many factors, including genetic, environmental, and cytokine dysregulation are implicated in causing aberrant immune responses that drive tissue damage [102–104]. Many studies on divergent immune responses in autoimmunity have shown dysfunction of helper T cell subsets, which include Th1, Th17, and/or Treg cells [104, 105]. Studies in the last decade have identified IL-9-secreting Th9 cells as another T helper cell subset involved in immune responses [23, 106]. The IL-9 cytokine has become the focus of many autoimmune studies [107, 108]. Initial studies showed IL-9

to be a growth factor and a Th2 cytokine [13, 108]. More recently, IL-9 has been characterized as a lineage-specific cytokine for Th9 cells [109]. Thereafter, many immune cells involved in autoimmunity, such as Th17 and Treg cells, have demonstrated secretion of IL-9 [16, 110]. In EAE, a rodent model of MS, researchers identified Th9 and its signature cytokine, IL-9, in driving the disease process [111]. Its close association with Th17 and TGF- β has renewed interest in the role of IL-9 in the pathogenesis of autoimmune diseases [23]. In this section, we will examine the role of IL-9 in some autoimmune diseases such as multiple sclerosis (MS), systemic lupus erythematosus (SLE), inflammatory bowel diseases (IBD), rheumatoid arthritis (RA), and uveitis.

10. IL-9 and IL-17 dynamics in autoimmunity

The role of IL-9 in autoimmunity was illuminated when many studies reported that IL-9 and IL-17 are intricately related in driving the pathogenesis of diseases [111]. Human and animal studies revealed that Th17 cells secrete some amount of IL-9, in addition to other proinflammatory cytokines [112]. During the differentiation of naive T cells, TGF- β , a key driver of Th17 polarization, plays an important role in the differentiation of Th9 cells [23]. This was well elaborated in a study by Nowak *et al* in which *in vitro* polarization of MOG-specific Th17 cells was shown to generate IL-9-secreting Th9 [22, 113]. Secretion of IL-9 was further enhanced by the addition of IL-1 β or IL-21 to the culture [113]. In addition, TGF- β and IL-6 induce Th17 cells that co-express IL-9 and IL-17 [22]. Studies have shown an increased frequency of memory CD4 cells that co-express IL-9 and IL-17 in patients with Type 1 diabetes [23].

On the other hand, IL-9 potentiates Th17 functions in an autocrine manner on Th17 cells [22, 110]. Th17 is a predominant helper T-cell subset that expresses IL-9 receptors (IL-9R) [22]. Through this receptor, IL-9 acts as an activator of Th17 cells [22]. IL-9 also synergizes with TGF- β to differentiate naive T cells into Th17 cells [110]. The presence of IL-9 in T cell cultures leads to the expansion of Th17 cells [110]. The importance of IL-9 in Th17 cell function is emphasized in IL-9R-deficient experimental autoimmune encephalomyelitis (EAE) model. Mice that lack IL-9 signaling showed decreased Th17 cells and defective migration of Th17 cells into the CNS [22, 114]. Neutralization of IL-9 led to attenuation of disease in EAE [22]. This unique relationship between IL-9 and Th17 provides the premise to examine the role of IL-9 in Th17-mediated autoimmune diseases.

11. Multiple sclerosis (MS)

Most autoimmune diseases like MS occur due to alteration of immune responses, which leads to tissue damage. The importance of IL-9 in MS has been enhanced through our understanding of the roles of IL-9-secreting T cells in EAE, an animal model of MS orchestrated by helper T cells [115]. Most studies revealed IL-9 plays a pathogenic role in EAE [22]. Th9 cells and Th17 cells were observed in the central nervous system (CNS) during EAE [115]. Blockade of IL-9 signaling in EAE resulted in contradictory conclusions. One study reported increased severity of disease in IL9Ra KO mice on a C57BL/6 background through a loss of Treg function and increased secretion of GM-CSF [116]. Other studies showed attenuation of disease and decreased Th17 cell infiltration into the CNS of SJL mice treated with IL-9 blocking antibody [22, 117]. This opposing view in disease outcome may be due to differences in the helper T cell composition and dysfunction driving the

pathogenesis in the mouse strains. Also, IL-9 has been shown to increase chemokine CCL20, which enhances migration of Th17 into the CNS [22]. Accumulation and activation of mast cells during the Th17-IL9 immune response could explain the feedback loop [113]. Adoptive transfer of IL-9⁺ Th9 into recipient mice resulted in EAE [118]. Th9-EAE model manifested a unique disease profile independent of Th1 and Th17 EAE models [118].

The role of IL-9 in MS patients is complex. A study by Roucco *et al* showed that IL-9 activates STAT1 and STAT 5, which are inhibitors of Th17 function [119]. IL-9 directly interfered with IL-17 expression in Th17 cells. Levels of IL-9 in the cerebrospinal fluid (CSF) of relapsing and remitting MS patients were inversely correlated with the disease pathogenesis and the disability indices [119]. These findings suggested the immunoregulatory role of IL-9 in MS. In another study, CSF of MS patients showed increased amounts of IL-9, and levels of IL-9 correlated with IL-17 [120]. Therefore, more studies are needed to understand the functional role of IL-9 in MS.

12. Uveitis

Unlike other autoimmune diseases, uveitis is a heterogeneous disorder that results in inflammation of the eye [121]. In animal models of uveitis, adoptive transfer of *in vitro* polarized Th9 cells induced ocular inflammation [122, 123]. However, IL-9 was not detected in the eyes or lymph nodes of these mice [123]. Analysis of inflammatory cytokines in the vitreous humor of patients with uveitis detected increased levels of IL-9, among other proinflammatory cytokines [124]. However, the biological relevance of increased IL-9 in the study was not elaborated.

Another study examined the role of IL-9 in patients with Vogt-Koyanagi-Harada (VKH) disease. VKH is a systemic autoimmunity that manifests with bilateral panuveitis [125]. Patients with active disease had significantly higher levels of IL-9 in culture supernatants and higher IL-9 mRNA in PBMCs than did healthy controls and inactive patients [126]. The synergy of IL-9 and IL-17 was demonstrated in the study. The secretion of IL-17 by IL-9-treated PBMCs of active patients was significantly higher compared to the controls or inactive patients [126]. In a study that evaluated the serum of patients with Behcet's disease, another complex autoimmune disease with uveitis, serum IL-9 was neither elevated in disease state nor correlated with disease index [127]. More studies are needed to understand whether IL-9 signaling plays any immunological role in the eye.

13. Rheumatoid arthritis (RA)

The study of IL-9 in RA highlights its functional relationship with Tregs. In an antigen-induced animal model of arthritis, mice that lacked IL-9 had a chronic disease [128]. Treatment with rIL-9 resolved the joint inflammation, swelling, and tissue damage. The absence of IL-9 led to impaired suppressive functions of Treg cells [128]. Type 2 innate lymphoid cells (IL-C2) are documented to express IL-9 and have an anti-inflammatory function [128, 129]. These studies highlight the role of IL-9 in the resolution of inflammation in arthritis [130]. In human studies, IL-9-producing IL-C2 cells were also identified in the PBMCs of RA patients [130, 131]. In a study of treatment-induced remission of RA, synovial fluid of patients showed high levels of IL-9 [128].

14. Systemic lupus erythematosus (SLE)

Proinflammatory cytokines are generally believed to be involved in the pathogenesis of SLE. High levels of IL-9 mRNA and Th17 cells were seen in SLE patients compared with healthy controls (HC) [132, 133]. Dantas *et al*, evaluated the level of IL-9 in SLE and observed that patients with SLE had elevated IL-9 compared with levels in healthy individuals [134]. Further, IL-9⁺ CD4 cells were more abundant in patients with SLE [132]. Serum IL-9 and mRNA of IL-9 were significantly elevated in SLE patients [132]. Also the elevated serum IL-9 and mRNA correlated with the SLE severity index [132, 135]. Animal studies corroborated these findings. Spleens and kidneys of lupus-prone mice showed high expression of IL-9 [136]. Neutralizing antibodies of IL-9 decreased kidney manifestation of SLE (lupus nephritis) and decreased anti-dsDNA antibody titers in these animal models [136].

15. Inflammatory bowel disease (IBD)

Aberrant adaptive immune response to the gut epithelial cells involving both CD4 and CD8 is implicated in the IBD [137]. These T cells are shown to express $\alpha 4/\beta 7$ integrin, which binds to MAdcam1 on the gut epithelium [138, 139]. Gut T cells including cells that secrete IL-9 have been shown to express high levels of this integrin, and they propagate inflammation in the gut [140]. Gene expression studies have highlighted IR4 and GATA3 expression on immune cells that reside in the epithelial lining of the gut [141]. IRF4 is a transcription factor that drives the induction of Th9 immune responses in the gut [141]. Animal models of colitis confirms this finding of an abundance of the IL-9-producing T cells in the gut. These T-cells-producing IL-9 are involved in breaking the intestinal barrier [142]. In a DSS colitis model, anti-IL-9 blocking antibodies suppressed mucosal inflammation, and attenuation of disease was observed [142]. Adoptive transfer of IL-9-producing T cells into Rag2 knockout (Rag $2^{-/-}$ KO) mice also induced colitis [143]. Furthermore, IL-9 was found to directly modulate the expression of tight junction proteins, claudin and occludin in the animal model of colitis [144]. This indicates that IL-9 directly inhibited membrane integrity.

Immunological assessment of patients with inflammatory bowel disease (IBD) revealed high expression of IL-9 in the lamina propria [145]. In addition to other gut-residing T cells in IBD, CD4 cells had increased production of proinflammatory cytokines, including IL-9, which drive gut inflammation [145, 146]. Elevated levels of IL-1 β and IL-9 were observed in the serum of IBD patients, and these correlated with disease prognosis [147]. Epithelial cells of UC also showed high expression of IL-9 receptor (IL-9R) [147, 148]. This receptor expression is most pronounced in patients with active disease [147]. *Ex vivo* IL-9 treatment of intestinal epithelial cells from UC patients showed increased proliferation of epithelial cells and pSTAT 5 expression [110].

Together, these findings highlight the role of IL-9 in IBD and colitis models. IL-9 could serve as a therapeutic target for IBD. Mice treated with GATA 3 DNAzyme showed it directly reduced IL-9 production and some Th2 cytokines to attenuate disease [149].

16. Type I diabetes

Studies by Vasanthakumar *et al* examined the role of IL-9 in patients with diabetes mellitus (DM) [150]. They observed that memory T cells from patients

stimulated with Th17 polarizing conditions led to IL-9 production [150]. This shows that Th17 cells from DM patients have an increased ability to secrete IL-9 [23]. The study also identified TGF- β as the critical activator of IL-9 secretion [23]. TGF- β activity links Th17 and IL-9 secretion.

IL-9 appears to play both anti- and pro-inflammatory functions in autoimmunity. The functional heterogeneity of IL-9 may result from the unique cells or the microenvironment producing it. In RA, IL-9 exhibits anti-inflammatory function [128]. Studies have elaborated the anti-inflammatory function of IL-9 as it potentiates Treg-dependent immune tolerance to allografts [151]. In the gut, it is regarded as proinflammatory [142]. Some studies have shown that the expression of the activation marker CD96 on Th9 cells may explain the immunological status of the secreted IL-9 [152]. Researchers have reported that Th9 with high expression of CD96 showed a reduced ability to cause colitis compared with Th9 with low expression of CD96, which is associated with severe intestinal inflammation [152]. More studies must be done to identify the immunological heterogeneity of IL-9.

17. IL-9 as a therapeutic target

One principle of treatment of autoimmune diseases involves inhibition of mediators of inflammation. Drugs that target proinflammatory cytokines are extensively used in the treatment of autoimmune diseases [153]. Here we explore the use of IL-9 blockade as a therapeutic target in different disease conditions.

Medimmune LLC developed a humanized anti-IL-9 monoclonal antibody, MEDI-528 [154]. This humanized anti-IL-9 monoclonal antibody was indicated for use in allergen-induced asthma in adults [154]. Results from the clinical trial of Medimmune MEDI-528 showed no increased efficacy in improving respiratory functions and control of asthma compared to placebo [155]. Preclinical studies in mice showed the efficacy of blocking IL-9 in maintaining the airway [156]. Questions remain regarding why therapy directed at IL-9 failed to produce the desired response in humans. Heterogeneity of IL-9 sources and functions could explain the differences in airway response observed in this clinic trial.

18. Other potential IL-9 treatments

IL-9R inhibitor (rhIL-9-ETA) is a chimeric toxin targeting IL9 receptor [157]. These IL-9R inhibitors have efficacy in targeting malignant cells in non-hodgkin's lymphoma (NHL) and acute myeloid leukemia (AML) expressing IL9 and IL-9R [157]. However, the efficacy of this drug has not been tested in autoimmunity. Pfizer Inc. developed a JAK/STAT pathway inhibitor, CP-690550 [158]. It specifically targets and inhibits the activation of JAK 3 [158]. This treatment effectively prevents transplant rejection [158]. This drug could be beneficial in inhibiting IL-9 signaling, which depends on the JAK/STAT pathway. JAK inhibitors have been used in the treatment of RA and psoriasis [159]. UC patients that were treated with JAK inhibitors showed decreased Th9 cells [160].

BNZ 132-1-40 peptide, an antagonist of IL-2, IL-9, and IL-15 from Bioniz Therapeutics is undergoing safety and tolerability testing in patients with moderate to severe alopecia areata, an autoimmune disease of the skin that leads to hair loss [161]. However, no results from the clinical trial were available at the time of this review. Recently, FDA approved the use of BNZ-1 for the treatment of cutaneous T cell lymphoma (CTCL) [162]. These studies suggest BNZ-1 could be used to target IL-9 in diseases [163].

Other potential drug options include RDP58, which targets IRF4, a transcription factor involved in Th9 induction [164]. Interferon gamma (IFN- γ) has the ability to inhibit Th9 polarization through IL-27-dependent mechanisms [165]. Actimmune, an IFN- γ -based therapy by Horizon Therapeutics, is FDA-approved for the treatment of chronic granulomatous disease (CGD) [166]. The efficacy of inhibiting IL-9 by this drug could be tested in IL-9-related disorders.

The immune modulatory roles of IL-9 in health and diseases are important and provides a basis for exploring IL-9 as a therapeutic target. However, the divergent roles of IL-9 in promoting and inhibiting inflammation complicate definitive drug development. Some studies have highlighted the function of IL-9 in promoting immune tolerance. Future studies to understand cell-specific IL-9 regulation and function may resolve the conundrum of therapy development targeting IL-9. More studies in disease will broaden our knowledge about IL-9 function.

19. Conclusion

Significant progress has been made in our understanding of the functions of IL9 in health and diseases. For a long time, IL-9 was considered as a T cell growth factor, however, the identification of Th9 helper T cells has expanded our understanding on the roles IL-9 play in diseases. The pathogenic functions of IL-9 in autoimmunity and allergy suggest that IL-9 signaling can be targeted for therapy development. In this chapter, we focused on the function of IL-9 in different autoimmune diseases that include MS, SLE, RA, uveitis, and allergic conditions. We also highlighted IL-9-Th17 paradigm and its complexity in autoimmune diseases. Animal models of autoimmune diseases revealed contrasting roles of IL-9 and human studies are limited. Therefore, extensive animal and human research are necessary to elucidate the divergent immunological roles of IL-9. Such studies will be required for effective drug development that targets IL-9 signaling.

Acknowledgements

We will like to thank Kathy Kyler of the Office of the Vice President for Research and Mary Carter Ph.D. of the Writing center, University of Oklahoma Health Sciences center. Also, we will like to extend our gratitude to Dr. Jimmy Ballard and the staff of Microbiology and Immunology, OUHSC.

Conflict of interest

The authors declare no conflict of interests.

Intechopen

Author details

Ahmed Ummey Khalecha Bintha¹, Amani Souwelimatou Amadou¹, Mursalin Md Huzzatul² and Muhammad Fauziyya^{3*}

1 Department of Microbiology and Immunology, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA

2 Department of Ophthalmology, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA

3 Department of Neurosurgery, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA

*Address all correspondence to: fauziyya-muhammad@ouhsc.edu

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Goswami, R. and M.H. Kaplan, *A brief history of IL-9*. J Immunol, 2011. 186(6): p. 3283-3288.

[2] Uyttenhove, C., R.J. Simpson, and J. Van Snick, *Functional and structural characterization of P40, a mouse glycoprotein with T-cell growth factor activity.* Proc Natl Acad Sci U S A, 1988. **85**(18): p. 6934-6938.

[3] Van Snick, J., et al., *Cloning and characterization of a cDNA for a new mouse T cell growth factor (P40).* J Exp Med, 1989. **169**(1): p. 363-368.

[4] Hültner, L., et al., *Mast cell growthenhancing activity (MEA) is structurally related and functionally identical to the novel mouse T cell growth factor P40/ TCGFIII (interleukin 9).* Eur J Immunol, 1990. **20**(6): p. 1413-6.

[5] Renauld, J.C., et al., *Human P40/IL-9. Expression in activated CD4+ T cells, genomic organization, and comparison with the mouse gene.* J Immunol, 1990. **144**(11): p. 4235-4241.

[6] Mock, B.A., et al., *IL9 maps to mouse chromosome 13 and human chromosome 5.* Immunogenetics, 1990. **31**(4): p. 265-70.

[7] Simpson, R.J., et al., *Complete amino acid sequence of a new murine T-cell growth factor P40*. Eur J Biochem, 1989. **183**(3): p. 715-722.

[8] Koh, B., et al., *A conserved enhancer regulates II9 expression in multiple lineages.* Nat Commun, 2018. **9**(1): p. 4803.

[9] Kaplan, M.H., *The transcription factor network in Th9 cells*. Semin Immunopathol, 2017. **39**(1): p. 11-20.

[10] Staudt, V., et al., *Interferon-regulatory factor 4 is essential for the developmental program of T helper 9 cells*. Immunity, 2010. **33**(2): p. 192-202. [11] Jash, A., et al., Nuclear factor of activated T cells 1 (NFAT1)-induced permissive chromatin modification facilitates nuclear factor- κB (NF- κB)mediated interleukin-9 (IL-9) transactivation. J Biol Chem, 2012. **287**(19): p. 15445-15457.

[12] Gessner, A., H. Blum, and M. Röllinghoff, *Differential regulation* of IL-9-expression after infection with Leishmania major in susceptible and resistant mice. Immunobiology, 1993. **189**(5): p. 419-435.

[13] Else, K.J., L. Hültner, and R.K. Grencis, *Cellular immune responses to the murine nematode parasite Trichuris muris. II. Differential induction of TH-cell subsets in resistant versus susceptible mice.* Immunology, 1992. **75**(2): p. 232-237.

[14] Schmitt, E., et al., *TCGF III/P40 is produced by naive murine CD4+ T cells but is not a general T cell growth factor.* Eur J Immunol, 1989. **19**(11): p. 2167-2170.

[15] Temann, U.A., et al., *Expression of interleukin 9 in the lungs of transgenic mice causes airway inflammation, mast cell hyperplasia, and bronchial hyperresponsiveness.* J Exp Med, 1998. **188**(7): p. 1307-1320.

[16] Chang, H.C., et al., *The transcription factor PU.1 is required for the development of IL-9-producing T cells and allergic inflammation.* Nat Immunol, 2010. **11**(6): p. 527-534.

[17] Dardalhon, V., et al., *IL-4 inhibits TGF-beta-induced Foxp3+ T cells and*, *together with TGF-beta, generates IL-9+ IL-10+ Foxp3(-) effector T cells*. Nat Immunol, 2008. **9**(12): p. 1347-1355.

[18] Veldhoen, M., et al., *Transforming* growth factor-beta 'reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9-producing *subset*. Nat Immunol, 2008. **9**(12): p. 1341-1346.

[19] Li, M.O., Y.Y. Wan, and R.A. Flavell, *T cell-produced transforming growth factor-beta1 controls T cell tolerance and regulates Th1- and Th17-cell differentiation.* Immunity, 2007. **26**(5): p. 579-591.

[20] Schmitt, E., et al., *IL-9 production* of naive CD4+ T cells depends on *IL-2*, is synergistically enhanced by a combination of TGF-beta and *IL-4*, and is inhibited by *IFN-gamma*. J Immunol, 1994. **153**(9): p. 3989-3996.

[21] Tesmer, L.A., et al., *Th17 cells in human disease*. Immunol Rev, 2008. **223**: p. 87-113.

[22] Nowak, E.C., et al., *IL-9 as a mediator of Th17-driven inflammatory disease.* J Exp Med, 2009. **206**(8): p. 1653-1660.

[23] Beriou, G., et al., *TGF-beta induces IL-9 production from human Th17 cells.* J Immunol, 2010. **185**(1): p. 46-54.

[24] Putheti, P., et al., *Human CD4 memory T cells can become CD4+IL-9+ T cells.* PLoS One, 2010. **5**(1): p. e8706.

[25] Hültner, L., et al., *In activated mast cells*, *IL-1 up-regulates the production of several Th2-related cytokines including IL-9*. J Immunol, 2000. **164**(11): p. 5556-63.

[26] Stassen, M., et al., *IL-9 and IL-13 production by activated mast cells is strongly enhanced in the presence of lipopolysaccharide: NF-kappa B is decisively involved in the expression of IL-9.* J Immunol, 2001. **166**(7): p. 4391-8.

[27] Stassen, M., et al., *p38 MAP kinase drives the expression of mast cell-derived IL-9 via activation of the transcription factor GATA-1*. Mol Immunol, 2007. **44**(5): p. 926-33. [28] Stassen, M., et al., *Murine bone marrow-derived mast cells as potent producers of IL-9: costimulatory function of IL-10 and kit ligand in the presence of IL-1. J Immunol, 2000.* **164**(11): p. 5549-55.

[29] Wiener, Z., A. Falus, and S. Toth, IL-9 increases the expression of several cytokines in activated mast cells, while the IL-9-induced IL-9 production is inhibited in mast cells of histamine-free transgenic mice. Cytokine, 2004. **26**(3): p. 122-130.

[30] Lauwerys, B.R., et al., *Cytokine* production and killer activity of NK/T-NK cells derived with IL-2, IL-15, or the combination of IL-12 and IL-18. J Immunol, 2000. **165**(4): p. 1847-53.

[31] Nagato, T., et al., *Expression of interleukin-9 in nasal natural killer/Tcell lymphoma cell lines and patients.* Clin Cancer Res, 2005. **11**(23): p. 8250-8257.

[32] Jones, T.G., et al., *Antigen-induced increases in pulmonary mast cell progenitor numbers depend on IL-9 and CD1d-restricted NKT cells.* J Immunol, 2009. **183**(8): p. 5251-5260.

[33] Gounni, A.S., et al., *IL-9 expression by human eosinophils: regulation by IL-1beta and TNF-alpha*. J Allergy Clin Immunol, 2000. **106**(3): p. 460-466.

[34] Sun, B., et al., *Characterization and allergic role of IL-33-induced neutrophil polarization*. Cell Mol Immunol, 2018. **15**(8): p. 782-793.

[35] Xiao, M., et al., Osteoblasts support megakaryopoiesis through production of interleukin-9. Blood, 2017. **129**(24): p. 3196-3209.

[36] Russell, S.M., et al., *Interaction of IL-2R beta and gamma c chains with Jak1 and Jak3: implications for XSCID and XCID.* Science, 1994. **266**(5187): p. 1042-1045.

[37] Demoulin, J.B. and J.C. Renauld, *Signalling by cytokines interacting with the interleukin-2 receptor gamma chain.* Cytokines Cell Mol Ther, 1998. **4**(4): p. 243-256.

[38] Renauld, J.C., et al., *Expression cloning of the murine and human interleukin 9 receptor cDNAs*. Proc Natl Acad Sci U S A, 1992. **89**(12): p. 5690-5694.

[39] Kimura, Y., et al., *Sharing of the IL-2 receptor gamma chain with the functional IL-9 receptor complex.* Int Immunol, 1995. 7(1): p. 115-120.

[40] Bauer, J.H., et al., *Heteromerization* of the gammac chain with the interleukin-9 receptor alpha subunit leads to STAT activation and prevention of apoptosis. J Biol Chem, 1998. **273**(15): p. 9255-9260.

[41] Zhu, Y.X., et al., *Critical cytoplasmic* domains of human interleukin-9 receptor alpha chain in interleukin-9-mediated cell proliferation and signal transduction. J Biol Chem, 1997. **272**(34): p. 21334-21340.

[42] Malka, Y., et al., Ligand-independent homomeric and heteromeric complexes between interleukin-2 or -9 receptor subunits and the gamma chain. J Biol Chem, 2008. **283**(48): p. 33569-33577.

[43] Fujiwara, H., et al., Homodimerization of the human interleukin 4 receptor alpha chain induces Cepsilon germline transcripts in B cells in the absence of the interleukin 2 receptor gamma chain. Proc Natl Acad Sci U S A, 1997. **94**(11): p. 5866-5871.

[44] Ihle, J.N. and I.M. Kerr, *Jaks and Stats in signaling by the cytokine receptor superfamily.* Trends Genet, 1995. **11**(2): p. 69-74.

[45] Demoulin, J.B., et al., *A single tyrosine of the interleukin-9 (IL-9) receptor is required for STAT activation*, *antiapoptotic activity, and growth regulation by IL-9.* Mol Cell Biol, 1996. **16**(9): p. 4710-6.

[46] Levy, D.E. and J.E. Darnell, Jr., *Stats: transcriptional control and biological impact*. Nat Rev Mol Cell Biol, 2002. **3**(9): p. 651-662.

[47] Malik, S. and A. Awasthi, *Transcriptional Control of Th9 Cells: Role of Foxo1 in Interleukin-9 Induction*. Frontiers in Immunology, 2018.
9: p. 995.

[48] Humblin, E., et al., *IRF8-dependent molecular complexes control the Th9 transcriptional program.* Nature Communications, 2017. **8**(1): p. 2085.

[49] Ngoc, L.P., et al., *Cytokines, allergy, and asthma.* Current opinion in allergy and clinical immunology, 2005. 5(2): p. 161-166.

[50] Veldhoen, M., et al., *Transforming* growth factor- β 'reprograms' the differentiation of T helper 2 cells and promotes an interleukin 9–producing subset. Nature immunology, 2008. **9**(12): p. 1341-1346.

[51] Gessner, A., H. Blum, and M. Röllinghoff, *Differential regulation* of IL-9-expression after infection with Leishmania major in susceptible and resistant mice. Immunobiology, 1993. **189**(5): p. 419-435.

[52] Dugas, B., et al., Interleukin-9 potentiates the interleukin-4-induced immunoglobulin (IgG, IgM and IgE) production by normal human B lymphocytes. European journal of immunology, 1993. **23**(7): p. 1687-1692.

[53] Takatsuka, S., et al., *IL-9 receptor* signaling in memory *B cells regulates* humoral recall responses. Nature immunology, 2018. **19**(9): p. 1025-1034.

[54] MURPHY, K. and C. WEAVER, *JANEWAY'S 9TH EDITION.*

[55] Petit-Frere, C., et al., *Interleukin-9* potentiates the interleukin-4-induced IgE and IgG1 release from murine B lymphocytes. Immunology, 1993. **79**(1): p. 146.

[56] Dong, Q., et al., *IL-9 induces* chemokine expression in lung epithelial cells and baseline airway eosinophilia in transgenic mice. European journal of immunology, 1999. **29**(7): p. 2130-2139.

[57] Longphre, M., et al., *Allergen-induced IL-9 directly stimulates mucin transcription in respiratory epithelial cells.* The Journal of clinical investigation, 1999. **104**(10): p. 1375-1382.

[58] Sherkat, R., et al., *Innate lymphoid cells and cytokines of the novel subtypes of helper T cells in asthma*. Asia Pacific Allergy, 2014. **4**(4): p. 212-221.

[59] Hoppenot, D., et al., *Peripheral blood Th9 cells and eosinophil apoptosis in asthma patients*. Medicina, 2015. **51**(1): p. 10-17.

[60] Erpenbeck, V.J., et al., *Increased* expression of interleukin-9 messenger RNA after segmental allergen challenge in allergic asthmatics. Chest, 2003. **123**(3): p. 370S.

[61] Kim, M.S., et al., *Effects of interleukin-9 blockade on chronic airway inflammation in murine asthma models.* Allergy, asthma & immunology research, 2013. 5(4): p. 197-206.

[62] Fawaz, L.M., et al., *Expression* of *IL-9* receptor α chain on human germinal center B cells modulates *IgE secretion*. Journal of allergy and clinical immunology, 2007. **120**(5): p. 1208-1215.

[63] Gounni, A.S., et al., *IL-9-mediated induction of eotaxin1/CCL11 in human airway smooth muscle cells*. The Journal of Immunology, 2004. **173**(4): p. 2771-2779. [64] Levitt, R.C., et al., *IL-9 pathway in asthma: new therapeutic targets for allergic inflammatory disorders.* Journal of Allergy and Clinical Immunology, 1999. **103**(5): p. S485-S491.

[65] Temann, U.-A., et al., *Expression* of interleukin 9 in the lungs of transgenic mice causes airway inflammation, mast cell hyperplasia, and bronchial hyperresponsiveness. The Journal of experimental medicine, 1998. **188**(7): p. 1307-1320.

[66] Wiener, Z., A. Falus, and S. Toth, *IL-9 increases the expression of several cytokines in activated mast cells, while the IL-9-induced IL-9 production is inhibited in mast cells of histamine-free transgenic mice.* Cytokine, 2004. **26**(3): p. 122-130.

[67] Matsuzawa, S., et al., *IL-9 enhances the growth of human mast cell progenitors under stimulation with stem cell factor*. The Journal of Immunology, 2003. **170**(7): p. 3461-3467.

[68] Temann, U.-A., et al., *IL9 leads to airway inflammation by inducing IL13 expression in airway epithelial cells.* International immunology, 2007. **19**(1): p. 1-10.

[69] McLane, M.P., et al., Interleukin-9 promotes allergen-induced eosinophilic inflammation and airway hyperresponsiveness in transgenic mice. American journal of respiratory cell and molecular biology, 1998. **19**(5): p. 713-720.

[70] Ciprandi, G., *Serum interleukin 9 in allergic rhinitis.* Annals of Allergy, Asthma & Immunology, 2010. **104**(2): p. 180-181.

[71] Gu, Z.W., Y.X. Wang, and Z.W. Cao, *Neutralization of interleukin-9 ameliorates symptoms of allergic rhinitis by reducing Th2, Th9, and Th17 responses and increasing the Treg response in a murine model.* Oncotarget, 2017. **8**(9): p. 14314.

[72] Osterfeld, H., et al., *Differential roles* for the IL-9/IL-9 receptor alpha-chain pathway in systemic and oral antigeninduced anaphylaxis. J Allergy Clin Immunol, 2010. **125**(2): p. 469-476.e2.

[73] Nakajima-Adachi, H., et al., *Critical* role of intestinal interleukin-4 modulating regulatory T cells for desensitization, tolerance, and inflammation of food allergy. PLoS One, 2017. **12**(2): p. e0172795.

[74] Burton, O.T., et al., *Direct effects of IL-4 on mast cells drive their intestinal expansion and increase susceptibility to anaphylaxis in a murine model of food allergy*. Mucosal Immunol, 2013. **6**(4): p. 740-750.

[75] Shik, D., et al., *IL-9-producing cells in the development of IgE-mediated food allergy*. Semin Immunopathol, 2017. **39**(1): p. 69-77.

[76] El Ansari, Y.S., C. Kanagaratham, and H.C. Oettgen, *Mast Cells as Regulators of Adaptive Immune Responses in Food Allergy*. Yale J Biol Med, 2020. **93**(5): p. 711-718.

[77] Ventura, M., et al., *Intestinal permeability in patients with adverse reactions to food.* Digestive and liver disease, 2006. **38**(10): p. 732-736.

[78] Xie, J., et al., *Elevated antigen-driven IL-9 responses are prominent in peanut allergic humans.* PloS one, 2012. 7(10): p. e45377.

[79] Brough, H.A., et al., *IL-9 is a key component of memory TH cell peanut-specific responses from children with peanut allergy*. Journal of allergy and clinical immunology, 2014. **134**(6): p. 1329-1338. e10.

[80] Chen, C.Y., et al., *Induction of Interleukin-9-Producing Mucosal Mast Cells Promotes Susceptibility to IgE-Mediated Experimental Food Allergy.* Immunity, 2015. **43**(4): p. 788-802. [81] Tomar, S., et al., *IL-4–BATF* signaling directly modulates *IL-9* producing mucosal mast cell (MMC9) function in experimental food allergy. Journal of Allergy and Clinical Immunology, 2021. **147**(1): p. 280-295.

[82] Forbes, E.E., et al., *IL-9- and mast cell-mediated intestinal permeability predisposes to oral antigen hypersensitivity.* J Exp Med, 2008. **205**(4): p. 897-913.

[83] Osterfeld, H., et al., *Differential* roles for the IL-9/IL-9 receptor α-chain pathway in systemic and oral antigen—induced anaphylaxis. Journal of allergy and clinical immunology, 2010. **125**(2): p. 469-476. e2.

[84] Ahrens, R., et al., *Intestinal mast cell levels control severity of oral antigen-induced anaphylaxis in mice*. The American journal of pathology, 2012. **180**(4): p. 1535-1546.

[85] Chen, C.-Y., et al., *Induction of interleukin-9-producing mucosal mast cells promotes susceptibility to IgEmediated experimental food allergy.* Immunity, 2015. **43**(4): p. 788-802.

[86] Tomar, S., et al., *IL-4-BATF* signaling directly modulates *IL-9* producing mucosal mast cell (MMC9) function in experimental food allergy. Journal of Allergy and Clinical Immunology, 2020.

[87] Steenwinckel, V., et al., *IL-9* promotes *IL-13-dependent paneth cell* hyperplasia and up-regulation of innate immunity mediators in intestinal mucosa. The journal of immunology, 2009. **182**(8): p. 4737-4743.

[88] Forbes, E.E., et al., *IL-9–and mast cell–mediated intestinal permeability predisposes to oral antigen hypersensitivity.* The Journal of experimental medicine, 2008. **205**(4): p. 897-913.

[89] Ma, L., et al., Possible pathogenic role of T helper type 9 cells and interleukin *(IL)-9 in atopic dermatitis*. Clin Exp Immunol, 2014. **175**(1): p. 25-31.

[90] Klonowska, J., et al., *New Cytokines in the Pathogenesis of Atopic Dermatitis-New Therapeutic Targets*. Int J Mol Sci, 2018. **19**(10).

[91] Ciprandi, G., et al., Serum interleukin-9 levels are associated with clinical severity in children with atopic dermatitis. Pediatric dermatology, 2013. **30**(2): p. 222-225.

[92] Ma, L., et al., Possible pathogenic role of T helper type 9 cells and interleukin (IL)-9 in atopic dermatitis. Clinical & Experimental Immunology, 2014.
175(1): p. 25-31.

[93] Namkung, J.-H., et al., *An association between IL-9 and IL-9 receptor gene polymorphisms and atopic dermatitis in a Korean population.* Journal of dermatological science, 2011. **62**(1): p. 16-21.

[94] Stockinger, B.B., et al., *Interleukin* 9 fate reporter reveals induction of innate *IL-9 response in lung* inflammation. 2011.

[95] Brunner, P.M., E. Guttman-Yassky, and D.Y. Leung, *The immunology of atopic dermatitis and its reversibility with broad-spectrum and targeted therapies*. Journal of Allergy and Clinical Immunology, 2017. **139**(4): p. S65-S76.

[96] Sismanopoulos, N., et al., *IL-9 induces VEGF secretion from human mast cells and IL-9/IL-9 receptor genes are overexpressed in atopic dermatitis.* PLoS One, 2012. 7(3): p. e33271.

[97] Zhang, Y., H. Matsuo, and E. Morita, *Increased production of vascular endothelial growth factor in the lesions of atopic dermatitis*. Archives of dermatological research, 2006. **297**(9): p. 425.

[98] Chen, L., et al., *The progression* of inflammation parallels the dermal

angiogenesis in a keratin 14 IL-4transgenic model of atopic dermatitis. Microcirculation, 2008. **15**(1): p. 49-64.

[99] Liu, J., et al., *IL-9 regulates allergen-specific Th1 responses in allergic contact dermatitis.* Journal of Investigative Dermatology, 2014. **134**(7): p. 1903-1911.

[100] Louahed, J., et al., *Interleukin 9* promotes influx and local maturation of eosinophils. Blood, The Journal of the American Society of Hematology, 2001. **97**(4): p. 1035-1042.

[101] Sarvetnick, N., *Etiology of autoimmunity*. Immunol Res, 2000. **21**(2-3): p. 357-362.

[102] Jörg, S., et al., *Environmental factors in autoimmune diseases and their role in multiple sclerosis*. Cell Mol Life Sci, 2016. **73**(24): p. 4611-4622.

[103] Smith, D.A. and D.R. Germolec, *Introduction to immunology and autoimmunity*. Environ Health Perspect, 1999. **107 Suppl 5**(Suppl 5): p. 661-5.

[104] Rosenblum, M.D., K.A. Remedios, and A.K. Abbas, *Mechanisms of human autoimmunity*. J Clin Invest, 2015. **125**(6): p. 2228-2233.

[105] Kaur, G., K. Mohindra, and S. Singla, *Autoimmunity-Basics and link with periodontal disease*. Autoimmun Rev, 2017. **16**(1): p. 64-71.

[106] Li, J., et al., *IL-9 and Th9 cells in health and diseases-From tolerance to immunopathology*. Cytokine Growth Factor Rev, 2017. **37**: p. 47-55.

[107] Pan, L.L., et al., *IL-9-producing Th9 cells may participate in pathogenesis of Takayasu's arteritis.* Clin Rheumatol, 2016. **35**(12): p. 3031-3036.

[108] Deng, Y., et al., *Th9 cells and IL-9 in autoimmune disorders: Pathogenesis and*

therapeutic potentials. Hum Immunol, 2017. **78**(2): p. 120-128.

[109] Weigmann, B. and M.F. Neurath, *Th9 cells in inflammatory bowel diseases.* Semin Immunopathol, 2017. **39**(1): p. 89-95.

[110] Elyaman, W., et al., *IL-9 induces* differentiation of TH17 cells and enhances function of FoxP3+ natural regulatory T cells. Proc Natl Acad Sci U S A, 2009. **106**(31): p. 12885-12890.

[111] Malik, S., V. Dardalhon, and A. Awasthi, *Characterization of Th9 Cells in the Development of EAE and IBD*. Methods Mol Biol, 2017. **1585**: p. 201-216.

[112] Noelle, R.J. and E.C. Nowak, *Cellular sources and immune functions of interleukin-9*. Nat Rev Immunol, 2010. **10**(10): p. 683-687.

[113] Nowak, E.C. and R.J. Noelle, *Interleukin-9 as a T helper type 17 cytokine*. Immunology, 2010. **131**(2): p. 169-173.

[114] Zhou, Y., et al., *IL-9 promotes Th17 cell migration into the central nervous system via CC chemokine ligand-20 produced by astrocytes.* J Immunol, 2011. **186**(7): p. 4415-4421.

[115] Elyaman, W. and S.J. Khoury, *Th9 cells in the pathogenesis of EAE and multiple sclerosis*. Semin Immunopathol, 2017. **39**(1): p. 79-87.

[116] Yoshimura, S., et al., *IL-9 Controls Central Nervous System Autoimmunity by Suppressing GM-CSF Production.* J Immunol, 2020. **204**(3): p. 531-539.

[117] Li, H., et al., *IL-9 is important for T-cell activation and differentiation in autoimmune inflammation of the central nervous system*. Eur J Immunol, 2011. **41**(8): p. 2197-2206.

[118] Jäger, A., et al., *Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with* *different pathological phenotypes.* J Immunol, 2009. **183**(11): p. 7169-7177.

[119] Ruocco, G., et al., *T helper 9 cells induced by plasmacytoid dendritic cells regulate interleukin-17 in multiple sclerosis*. Clin Sci (Lond), 2015. **129**(4): p. 291-303.

[120] Khaibullin, T., et al., *Elevated Levels of Proinflammatory Cytokines in Cerebrospinal Fluid of Multiple Sclerosis Patients.* Front Immunol, 2017. 8: p. 531.

[121] Squires, H., et al., A systematic review and economic evaluation of adalimumab and dexamethasone for treating non-infectious intermediate uveitis, posterior uveitis or panuveitis in adults. Health Technol Assess, 2017. **21**(68): p. 1-170.

[122] Kaplan, M.H., *Th9 cells:differentiation and disease*. Immunol Rev, 2013. 252(1): p. 104-115.

[123] Horai, R. and R.R. Caspi, *Cytokines in autoimmune uveitis*. J Interferon Cytokine Res, 2011. **31**(10): p. 733-744.

[124] Fukunaga, H., et al., *Analysis of inflammatory mediators in the vitreous humor of eyes with pan-uveitis according to aetiological classification*. Sci Rep, 2020. **10**(1): p. 2783.

[125] Baltmr, A., S. Lightman, and O. Tomkins-Netzer, *Vogt-Koyanagi-Harada syndrome - current perspectives*. Clin Ophthalmol, 2016. **10**: p. 2345-2361.

[126] Peng, Z., et al., *Expression and role of interleukin-9 in Vogt-Koyanagi-Harada disease*. Mol Vis, 2017. **23**: p. 538-547.

[127] Nouri-Vaskeh, M., et al., *Lack of association between serum IL-9 levels and Behçet's disease*. Immunol Lett, 2019. **211**: p. 23-27.

[128] Rauber, S., et al., *Resolution of inflammation by interleukin-9-producing type 2 innate lymphoid cells*. Nat Med, 2017. **23**(8): p. 938-944. [129] Karagiannis, F. and C. Wilhelm, *More Is Less: IL-9 in the Resolution of Inflammation*. Immunity, 2017. **47**(3): p. 403-405.

[130] Wu, X., *Innate Lymphocytes in Inflammatory Arthritis*. Front Immunol, 2020. **11**: p. 565275.

[131] Hughes-Austin, J.M., et al., Multiple cytokines and chemokines are associated with rheumatoid arthritis-related autoimmunity in first-degree relatives without rheumatoid arthritis: Studies of the Aetiology of Rheumatoid Arthritis (SERA). Ann Rheum Dis, 2013. **72**(6): p. 901-907.

[132] Ouyang, H., et al., *Increased interleukin-9 and CD4+IL-9+ T cells in patients with systemic lupus erythematosus.* Mol Med Rep, 2013. 7(3): p. 1031-1037.

[133] Leng, R.X., et al., *Potential roles of IL-9 in the pathogenesis of systemic lupus erythematosus*. Am J Clin Exp Immunol, 2012. **1**(1): p. 28-32.

[134] Dantas, A.T., et al., Increased
Serum Interleukin-9 Levels in
Rheumatoid Arthritis and Systemic Lupus
Erythematosus: Pathogenic Role or Just
an Epiphenomenon? Dis Markers, 2015.
2015: p. 519638.

[135] Ouyang, H., et al., [Abnormality and significance of interleukin-9 and CD4(+)interleukin-9(+) T-cells in peripheral blood of patients with systemic lupus erythematosus]. Zhonghua Yi Xue Za Zhi, 2013. **93**(2): p. 99-103.

[136] Yang, J., et al., Interleukin-9 Is Associated with Elevated Anti-Double-Stranded DNA Antibodies in Lupus-Prone Mice. Mol Med, 2015. **21**(1): p. 364-370.

[137] Funderburg, N.T., et al., *Circulating CD4(+)* and *CD8(+) T* cells are activated *in inflammatory bowel disease and* *are associated with plasma markers of inflammation*. Immunology, 2013. **140**(1): p. 87-97.

[138] Wittner, M., et al., Comparison of the integrin $\alpha 4\beta 7$ expression pattern of memory T cell subsets in HIV infection and ulcerative colitis. PLoS One, 2019. **14**(7): p. e0220008.

[139] Kurmaeva, E., et al., *T cell*associated $\alpha 4\beta 7$ but not $\alpha 4\beta l$ integrin is required for the induction and perpetuation of chronic colitis. Mucosal Immunol, 2014. 7(6): p. 1354-1365.

[140] Jovani, M. and S. Danese, Vedolizumab for the treatment of IBD: a selective therapeutic approach targeting pathogenic a4b7 cells. Curr Drug Targets, 2013. **14**(12): p. 1433-1443.

[141] Malik, S. and A. Awasthi, *Transcriptional Control of Th9 Cells: Role of Foxo1 in Interleukin-9 Induction*. Front Immunol, 2018. **9**: p. 995.

[142] Yuan, A., et al., *IL-9 antibody injection suppresses the inflammation in colitis mice*. Biochem Biophys Res Commun, 2015. **468**(4): p. 921-926.

[143] de Heusch, M., et al., *IL-9* exerts biological function on antigenexperienced murine T cells and exacerbates colitis induced by adoptive transfer. Eur J Immunol, 2020. **50**(7): p. 1034-1043.

[144] Gerlach, K., et al., *IL-9 regulates intestinal barrier function in experimental T cell-mediated colitis.* Tissue Barriers, 2015. **3**(1-2): p. e983777.

[145] Hufford, M.M. and M.H. Kaplan, *A gut reaction to IL-9*. Nat Immunol, 2014. **15**(7): p. 599-600.

[146] Matusiewicz, M., et al., Systemic interleukin-9 in inflammatory bowel disease: Association with mucosal healing in ulcerative colitis. World

J Gastroenterol, 2017. **23**(22): p. 4039-4046.

[147] Gerlach, K., et al., *TH9 cells that express the transcription factor PU.1 drive T cell-mediated colitis via IL-9 receptor signaling in intestinal epithelial cells.* Nat Immunol, 2014. **15**(7): p. 676-686.

[148] Vyas, S.P. and R. Goswami, *A* Decade of Th9 Cells: Role of Th9 Cells in Inflammatory Bowel Disease. Front Immunol, 2018. **9**: p. 1139.

[149] Popp, V., et al., *Rectal Delivery of a DNAzyme That Specifically Blocks the Transcription Factor GATA3 and Reduces Colitis in Mice.* Gastroenterology, 2017. **152**(1): p. 176-192.e5.

[150] Vasanthakumar, R., et al., Serum IL-9, IL-17, and TGF- β levels in subjects with diabetic kidney disease (CURES-134). Cytokine, 2015. 72(1): p. 109-112.

[151] Burrell, B.E., et al., *Regulatory T cell induction, migration, and function in transplantation.* J Immunol, 2012. **189**(10): p. 4705-4711.

[152] Stanko, K., et al., *CD96 expression determines the inflammatory potential of IL-9-producing Th9 cells.* Proc Natl Acad Sci U S A, 2018. **115**(13): p. E2940-e2949.

[153] Willrich, M.A., D.L. Murray, and M.R. Snyder, *Tumor necrosis factor inhibitors: clinical utility in autoimmune diseases*. Transl Res, 2015. **165**(2): p. 270-282.

[154] Antoniu, S.A., *MEDI-528, an anti-IL-9 humanized antibody for the treatment of asthma.* Curr Opin Mol Ther, 2010. **12**(2): p. 233-239.

[155] White, B., et al., Two firstin-human, open-label, phase I dose-escalation safety trials of MEDI-528, a monoclonal antibody against *interleukin-9, in healthy adult volunteers.* Clin Ther, 2009. **31**(4): p. 728-740.

[156] Kim, M.S., et al., *Effects of interleukin-9 blockade on chronic airway inflammation in murine asthma models*. Allergy Asthma Immunol Res, 2013. 5(4): p. 197-206.

[157] Klimka, A., et al., A deletion mutant of Pseudomonas exotoxin-A fused to recombinant human interleukin-9 (rhIL-9-ETA') shows specific cytotoxicity against IL-9-receptor-expressing cell lines. Cytokines Mol Ther, 1996. 2(3): p. 139-146.

[158] Flanagan, M.E., et al., *Discovery of CP-690,550: a potent and selective Janus kinase (JAK) inhibitor for the treatment of autoimmune diseases and organ transplant rejection.* J Med Chem, 2010. **53**(24): p. 8468-8484.

[159] Kvist-Hansen, A., P.R. Hansen, and L. Skov, *Systemic Treatment of Psoriasis with JAK Inhibitors: A Review.* Dermatol Ther (Heidelb), 2020. **10**(1): p. 29-42.

[160] Imam, T., et al., Effector T Helper Cell Subsets in Inflammatory Bowel Diseases. Front Immunol, 2018. 9: p. 1212.

[161] NCT03532958, P.T.o.B.i.P.W.M.t.S.A.A., 2019.

[162] A Dose-Ranging Study of IV BNZ-1 in LGL Leukemia or Refractory CTCL-NCT03239392.

[163] Wang, T.T., et al., *IL-2 and IL-15* blockade by BNZ-1, an inhibitor of selective *y*-chain cytokines, decreases leukemic T-cell viability. Leukemia, 2019. **33**(5): p. 1243-1255.

[164] Li, J., et al., *Toll-like receptors as therapeutic targets for autoimmune connective tissue diseases*. Pharmacology & Therapeutics, 2013. **138**(3): p. 441-451. [165] Murugaiyan, G., et al., *IFN-γ limits Th9-mediated autoimmune inflammation through dendritic cell modulation of IL-27.* J Immunol, 2012. **189**(11): p. 5277-5283.

[166] Green, D.S., et al., Production of a cellular product consisting of monocytes stimulated with Sylatron® (Peginterferon alfa-2b) and Actimmune® (Interferon gamma-1b) for human use. Journal of Translational Medicine, 2019. 17(1): p. 82.

