

1 **Graphene: a disruptive opportunity for COVID-19 and future pandemics?**

2
3 Giacomo Reina,[#] Daniel Iglesias,[#] Paolo Samorì,^{*} Alberto Bianco^{*}

4
5 Dr. G. Reina, Dr. A. Bianco

6 CNRS, Immunology, Immunopathology and Therapeutic Chemistry, UPR 3572, University of
7 Strasbourg, ISIS, 67000 Strasbourg, France

8 E-mail: a.bianco@ibmc-cnrs.unistra.fr

9 Dr. D Iglesias, Prof. P. Samorì, Dr. A. Bianco

10 University of Strasbourg, CNRS, ISIS, 67000 Strasbourg, France

11 E-mail: samori@unistra.fr

12
13 [#]These authors contributed equally

14 15 **Abstract**

16 The graphene revolution, which has taken place during the last 15 years, has represented a
17 paradigm shift for science. The extraordinary properties possessed by this unique material have
18 paved the road to a number of applications in materials science, opto-electronics, energy and
19 sensing. Graphene related materials (GRMs) are now produced in large scale and found niche
20 applications also in the biomedical technologies, defining new standards for drug delivery and
21 biosensing. Such advances positioned GRMs as novel tools to fight against the current COVID-
22 19 and future pandemics. In this regard, GRMs can play a major role in sensing, as an active
23 component in antiviral surfaces or in virucidal formulations. This essay showcases the most
24 promising strategies reported in the literature on the interface of GRMs with COVID-19 and other
25 types of viruses, with a strong focus on the impact of functionalization, deposition techniques and
26 integration into devices and surface coatings.

1 The spread of COVID-19 worldwide has dramatically changed our daily lives resulting in the most
2 unstable and uncertain period our society is facing after the two world wars. The infections are
3 coming as waves with increasing contagion and deaths.^[1] Many governments are grappling with
4 the competing demands of protecting the economy and protecting the healthcare systems. At the
5 same time, researchers worldwide are devoting a significant effort in the search for strategies
6 capable to contain the pandemic, diminish the hospitalization and develop as fast as possible a
7 reliable vaccine.^[2] So far we have learnt that our society will have to coexist at least for the next
8 one to two years, with the virus forcing the mankind to adopt new habits, use protective equipment
9 and take the precaution of sanitising commonly touched surfaces. In this context, research of
10 new antiviral solutions is highly sought after.

11 Over the past 15 years, graphene and graphene related materials (GRMs) have attracted a
12 great attention because of their unique physical and chemical properties, which rendered them
13 powerful components for applications in opto-electronics, energy storage and generation,
14 (bio)chemical sensing, reinforcement for aeronautics and constructions, membranes for
15 purification of water, etc. GRMs are composed by various carbon 2D materials each of them with
16 a specific structure and nomenclature.^[3,4] Among the different GRMs, the most promising
17 materials are graphene (G), graphene oxide (GO) and reduced graphene oxide (rGO) (Figure 1).
18 G displays a typical honeycomb sp^2 carbon structure that is responsible for its flexibility and
19 strength, exceptional electrical conductivity and high lipophilicity. Conversely, GO, the oxidized
20 form of G, contains various oxygenated groups and some sp^3 carbons that decrease the
21 mechanical and electrical performances but renders it highly hydrophilic and water dispersible.
22 Additionally, GO can be reduced to partially restore the electrical conductivity but retaining, to a
23 great extent, the water dispersibility, yielding rGO. GRMs are already produced at the industrial
24 scale and their application has been included in different device prototypes.

25 This Essay will showcase and rationalize the most relevant applications based on graphene and
26 related materials that can be foreseen to combat viral pandemics. In particular, GRMs
27 applications as antiviral material for drug delivery, smart surfaces and (bio)sensing will be critically
28 discussed and analysed. Timely applications, opportunities and future challenges will be
29 addressed. With this contribution, we would like to stimulate scientists to think out of the box to
30 find new efficient solutions against COVID-19 and the next viral pandemics.

31

32 **Medical applications blocking the viral replication machinery**

33 Since their discovery, GRMs have become powerful allies to fight against different diseases.^[5] By
34 taking full advantage of their tunable size, high biocompatibility and tailored surface functionalities,

1 GRMs have become useful components for application in different biomedical fields including
2 drug delivery, bioimaging and tissue engineering. Current progress of GRMs for antiviral
3 applications is still at the early stage. Yet, the encouraging results achieved in other biomedical
4 applications, including antimicrobial and immune activation, make G and GO potentially
5 promising tools to fight against viral pandemics. G, GO and rGO have been used in biomedical
6 research against different kinds of viruses. They combine remarkable figures of merit including a
7 high surface area offering good contact with the viral protein, a well-established surface chemistry
8 enabling the preparation of multifunctional platforms and an efficient photothermal activity under
9 near infrared laser irradiation allowing the local temperature enhancement. Thus, GRMs can be
10 ad hoc designed targeting common viral replication machinery (broad spectrum antiviral) or a
11 specific viral antigen (specific antiviral) making them useful allies against SARS-CoV-2 and other
12 viral infections. G, GO and rGO have been already explored for their virustatic activity, and so
13 designed to reversibly bind viral proteins or host cell proteins responsible for the attachment of
14 the virus, thus blocking the infection at the early stage. The most popular targeting agent reported
15 has been the mimicking cell heparan sulphate (HS), a polysaccharide present in a variety of
16 mammalian cells and broadly used by several viruses (e.g., HIV, Herpes simplex virus, Zika and
17 Hepatitis B virus) to penetrate host cell membrane.^[6] SARS-CoV-2 seems also to require HS as
18 an assisting cofactor to enter host cells.^[7] Thus, first clinical data have shown that the
19 administration of HS in COVID-19 patients with severe coronavirus disease is beneficial and that
20 one possible antiviral mechanism is that HS molecules bind spike protein to block viral
21 attachment or entry.^[7] Due to the strong electrostatic interaction between the virus and HS,^[6] the
22 mimicking of this linear sulphated polysaccharide has been achieved using ligands containing
23 sulphate or sulfonate pendent groups.^[8] These findings render HS particularly interesting for viral
24 treatment and the combination with GRMs offers two additional advantages: 1) when using
25 surface functionalized GRMs, the local concentration of the targeting moieties is higher, resulting
26 in an enhanced binding constant (multivalent effect),^[9] that, as for other carbon nanoparticles,
27 may reduce the therapeutically relevant concentration by different orders of magnitude;^[10] 2)
28 chemistry on GRMs allows multifunctionalization permitting the incorporation of different
29 desirable groups (Figure 1). For these reasons, functionalized GRMs are now studied as antiviral
30 drugs for medical therapies. For example, sulfonate groups have been introduced onto G, GO
31 and rGO through different chemical strategies. The most common one is through sp² carbon
32 derivatization. For instance, rGO functionalized with sulfonic acids (sulfanilic acid as precursor)
33 have shown interesting antiviral activity against *Herpes simplex Virus-1* (HSV-1).^[11] To enhance
34 the functionalization degree, more complex precursors have been grafted onto the GRMs surface.
35 Dendritic polyglycerol sulfate functionalized rGO has been prepared through epoxide ring
36 opening polymerization of glycidol and sulfonation of the hydroxyl groups. These conjugate has
37 shown similar efficacy to heparin in inhibiting Orthopoxviruses.^[12] Due to similarity with the

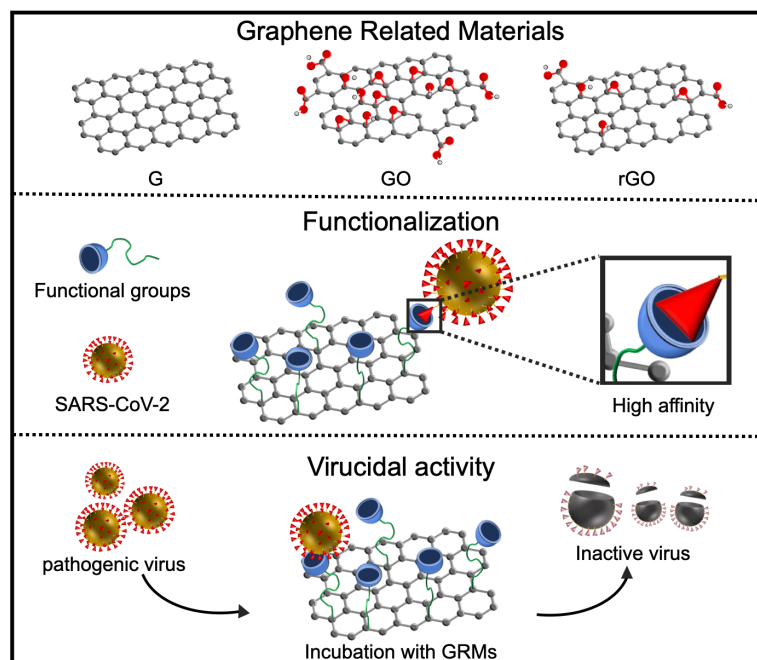
1 structure of HS, the inhibition effect could be enhanced by using a linear polyglycerol sulfonated
2 rGO, instead of the dendritic polymer.^[13] Another functionalization pathway well reported for the
3 functionalization of G is the triazine derivatization proposed by the group of Haag.^[14] This strategy
4 allows an easy and elegant surface multifunctionalization of G flakes through a controlled
5 derivatization of the triazine ring. This approach was used to introduce both sulfonated
6 polyglycerols and long fatty chains onto G surface. *In vitro* results on HSV-1 showed that a
7 synergistic effect between electrostatic and hydrophobic interactions resulted in a high inhibition
8 of infectivity.^[14] These *in vitro* examples show how G, GO and rGO can be *ad hoc* functionalized
9 in order to reversibly bind viruses and boost the antiviral activity. The use sulfonic groups may be
10 a key strategy to target *in vivo* different classes of viruses including SARS-CoV-2. However, due
11 to the reversible nature of the binding mechanism to viral capsid, this approach can be used to
12 slow down the infection (as for HS)^[7] but not to eradicate the pathology. Indeed, following the
13 binding thermodynamics, at low concentration the viral particles would be released from the
14 nanomaterial surface causing the recurrence of the infection. Thus, the design of a system
15 capable to irreversibly deactivate the viral machinery (virucidal activity) is timely and highly
16 desirable (Figure 1). Pioneering works have demonstrated that G and GO may possess intrinsic
17 antiviral activity resulting from their mechanical interaction with the viral capsid or the
18 envelope.^[15,16] However, this effect has not yet been confirmed neither *in vitro* nor *in vivo* where
19 the antiviral activity of non-functionalized materials was found being negligible. This *per se*
20 virucidal activity is present only when viruses are incubated with the materials prior to contact
21 with host cells, making pristine G and GO interesting for surface disinfection or at the early stages
22 of infection. The mechanism of virucidal effect is associated to the unique physical and chemical
23 properties of GRMs. In particular, this virucidal activity has been attributed to the combination of
24 three main factors: the physical insertion into the viral capsid, the negative surface charge, and
25 the amphiphilic nature of the materials.^[8] Therefore, by exploiting these characteristics it should
26 be possible to prepare a wide spectrum antiviral agents. For instance, *in silico* studies have
27 demonstrated that the exposure of Ebola virus to G leads to the poration of the protein protective
28 shell, which induces the leakage of genomic RNA leading to virus inhibition.^[15] Such virucidal
29 activity was associated to the sharp, rigid structure together with the hydrophobic surface allowing
30 G sheets to recognize and break VP40 protein-protein interactions. However, these studies have
31 never been confirmed by further *in vitro* or *in vivo* assays. In this context, it is worth to note that,
32 due to its highly hydrophobic nature, G is not colloidally stable in water and biological media, thus
33 limiting its application. One possible solution to prepare water stable G dispersions is by using
34 amphiphilic molecules during the graphite exfoliation process. Stable G water dispersions have
35 been obtained using different classes of surfactants including biocompatible molecules such as
36 proteins or sugars.^[17] The surfactants used during the exfoliation process play an important role
37 in the interaction of G with the viral surface, thus screening through modelling and *in vitro* tests

1 would be very helpful to select the most efficient G-based antiviral system. Conversely, GO
2 exposes several oxygen containing groups onto its surface that renders it highly hydrophilic to
3 form colloiddally stable aqueous dispersions. The GO structure has been theorized as composed
4 of oxygenated C sp³ islands surrounded by C sp² hydrophobic areas.^[5] This unique patchy
5 amphipathic structure is associated with a high capacity to adsorb lipophilic and hydrophilic
6 molecules onto its surface. GO is able to disrupt the cellular lipid bilayers, and interfere with
7 protein-protein assemblies leading to unfolding.^[18] For these reasons, GO is considered being
8 extremely promising as antiviral material. A study demonstrated the ability of GO to denature and
9 induce agglomeration of fragments of the viral protein R, which has a fundamental role on HIV1
10 host cell binding and uptake.^[16] The interaction of GO with HIV1 fragments were associated to
11 the hydrophobic interaction between the protein R and the nanomaterial that causes the
12 conversion of the active α -helix segments into β -sheets promoting the formation of biologically
13 inactive fibrillar agglomerates. GO were also used to treat enteric EV71 (e.g., pathogenic agents
14 of hand, foot, and mouth disease) and H9N2 (e.g., gastrointestinal avian influenza A)
15 demonstrating to have a strong capacity to absorb both viruses onto its surface and showing
16 good virucidal activity.^[19] In a comparative investigation using GO, rGO, graphite and graphite
17 oxide, it has been shown that GO and rGO display the highest virucidal activity against
18 Pseudorabies virus, indicating that the nanosized dimension of the material plays a crucial role
19 on their activity.^[20] Additionally, the covering of GO with cationic polymer causes a loss of antiviral
20 activity, thereby highlighting the importance of negative charges present on GO surfaces.

21 Understanding the interaction of GRMs with viruses may help to conceive more efficient broad-
22 spectrum antiviral agents. Hitherto, there are no reports proving the virucidal activity of GRMs
23 against SARS-CoV-2. It is worth noting that the viricidal activity is associated to different factors
24 including surface polarity, size and edges. Most of the performed tests did not consider the
25 biological media and/or the presence of protein corona. Coronation is a process that takes place
26 between serum proteins and GRMs flakes, which may irreversibly alter the materials surface
27 chemistry causing an attenuation or a drop of the virucidal activity. Additionally, the impact of
28 surface functionalization has not been exhaustively explored yet. The grafting of targeting
29 molecules (e.g., ACE2, 3CLpro, S Protein)^[21] may enhance the binding capacity of the
30 nanomaterial and hence improve the specificity and the virucidal activity against SARS-CoV-2.
31 In this context, another strategy is to vehicle antiviral drugs through GRMs for the selective
32 delivery to targeted tissues and organs. GRMs combine good biocompatibility, longer blood
33 circulation capacity and good permeability through biological barriers and membranes thus
34 offering outstanding drug delivery capacity. For instance, GO can efficiently release hypericin
35 (e.g., an antiviral drug) into duck reovirus infected ducklings preventing pathological lesions and
36 decreasing the viral load in the organs with a consequent prolonged animal survival.^[22]

1 Multifunctionalization strategies have been also developed for the preparation of virucidal GRMs.
2 GO modified with sulfanilic acid and β -cyclodextrin displayed high binding efficacy to respiratory
3 syncytial virus and incorporation of curcumin, a powerful lipophilic antiviral drug capable to induce
4 oxidation of the capsids.^[23]

5 Another interesting but less developed approach regards the virucidal activity of G, GO and rGO
6 through phototherapy. The inherent photothermal activity of G, GO and rGO was successfully
7 proved in cancer therapy.^[5] In this context, G, GO and rGO are able to efficiently convert near
8 infrared (NIR) light into heat generating a local and rapid temperature increase that induces cell
9 ablation. The same strategy can be applied for virucidal purposes where the virus upon
10 adsorption onto the nanomaterial surface can be irreversibly damaged by heat.^[24] The possibility
11 to prepare antiviral films that can be easily regenerated upon harmless NIR light might be an
12 easy, versatile and cost-effective solution for the next pandemics.



13

14 **Figure 1.** Sketch describing the preparation of antiviral GRMs. Top panel: chemical structures
15 of graphene, graphene oxide and reduced graphene oxide. GRMs can enhance their affinity
16 for Sars-Cov-2 through chemical functionalization with specific functional groups (middle
17 panel). Following incubation, GRMs can irreversibly damage the virus, dropping down their
18 infectivity (bottom panel).

19

20

21

22

1 **Antiviral surfaces and coatings**

2 Many studies were devoted to evaluate the resistance of viruses under different environmental
3 conditions. For instance, the exhaled viral particles remain infective even 3 h after
4 exhalation.^[25,26] Beside aerosols and direct contact, one the most plausible mechanisms of
5 transmission for SARS-CoV-2 and other pathogens is through fomites. Fomites are inanimate
6 items that can transfer the pathogens to a new host through unexpected transfer by contact
7 with virus on inanimate objects. Common examples of fomites are door handles, lift buttons,
8 ATM machine touchscreens, shared equipment in workplaces, etc. In a recent study, Riddell
9 *et al.* have analyzed the survival rates of the SARS-CoV-2 in materials present in our every-
10 day life including stainless steel, polymer and paper banknotes, cotton, and vinyl.^[27]
11 Remarkably, SARS-CoV-2 remains active in non-porous substrates up to 28 days when it is
12 incubated in dark, at 20 °C, at 50% relative humidity. The persistence in cotton (i.e., a highly
13 porous material) was markedly shorter. Despite the variations due to the experimental
14 conditions (e.g., viral titer, relative humidity, tested materials), these results are in line with
15 other reports on the topic,^[28] which underlined the need for constant sanitization and hand-
16 washing, calling for investigations to develop new functional coatings that help to shorten the
17 persistence of pathogens. A recent perspective article highlighted the importance of the
18 development of efficient antimicrobial surfaces, with particular emphasis on polymeric
19 materials due to their constant presence in our every-day life.^[29] Common strategies to boost
20 the antimicrobial properties rely on the use of metal nanoparticles (e.g., CuO, Ag or Au
21 nanoparticles) or the exploitation of surface chemistry to tune the surface-pathogen interface.
22 With sufficient adjustment, the strategies developed against bacteria could be translated to
23 viruses. Chemical approaches leading to modify surfaces with pendent cationic, anionic heads
24 or hydrophobic chains have been reported to inactivate viruses via different mechanisms. For
25 example, polyethyleneimines (PEI) were exploited to prepare glass slides with different
26 surface chemistry and to evaluate their antiviral activity against influenza viruses.^[30] Significant
27 differences were found related to the length of the alkyl chains and the quaternization of
28 amines. We would like to underline that the growing chemistry of 2D materials could find
29 application in the development of antiviral coatings with *ad hoc* functional groups. Recently,
30 we reported the preparation of GO with branched PEI to study the complexation of siRNA,^[31]
31 or for water remediation.^[32] These studies might inspire the use of PEI-modified GO as antiviral
32 surface coatings.

33 A key aspect for the realization of smart coatings based on GRMs concerns its integration into
34 the actual surfaces (Figure 2). Extensive research effort has been dedicated to the
35 development of GRMs inks or dispersions with high processability. Application of these inks
36 to surfaces can be done by a number of techniques such as dip-coating, (ink-jet) printing,

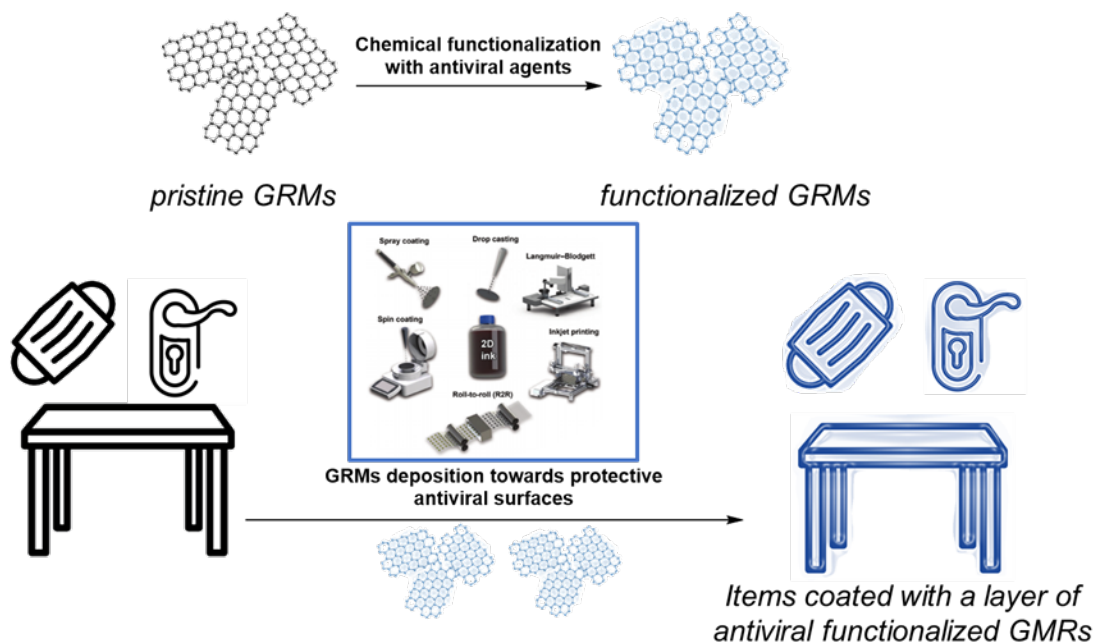
1 screen printing, spray-coating, roll-to-roll processing, etc.^[33] The chemical nature of the
2 surface and the ink to be deposited determine which technique suits best. For instance, spray-
3 coating might be the preferred option when transparency is required,^[34] while pad-dry process
4 works better for industrial scale coating of textiles as reported in a work describing the
5 development of e-textiles.^[35] In this study, GO was simultaneously reduced and non-covalently
6 functionalized with a hydrophilic polymer to avoid the re-stacking of the flakes. The adequate
7 control of the chemical functionalization of 2D sheets open the door to their use in virtually any
8 media. This could enable their dispersion in precursor matrices of certain polymeric substrates.
9 For example, the covalent functionalization of rGO with silsesquioxane moieties allowed their
10 integration into electrospun fibers.^[36] The electrospinning process in the preparation of textiles,
11 filters, wound dressing materials, etc. has shown promising results in the last years.^[37,38]
12 Integration of GRMs into electrospun mats was beneficial in many applications as shown in
13 the case of poly(lactide-co-glycolide/chitosan) fibers decorated with GO functionalized with Ag
14 nanoparticles in a post-modification step with strong antibacterial activity against several
15 tested microorganisms.^[39]

16 While waiting for an effective medical treatment or the herd immunity achieved with the new
17 and upcoming vaccines against SARS-CoV-2, the use of face-masks is among the most
18 effective precautions to be exploited in order to avoid the spread of viral particles when the
19 social distancing is not possible. As a consequence, millions of surgical masks are daily
20 produced, used and disposed around the globe causing a strong environmental impact.^[40]
21 Research activities are being directed towards the reutilization of masks after decontamination
22 by microwave irradiation or dry heating.^[41] The multifaceted GRMs can play an important role
23 to mitigate this massive use of resources. For instance, the superhydrophobicity of laser-
24 induced graphene (LIG) was exploited to develop self-cleaning recyclable masks.^[42] LIG is
25 produced after the irradiation of commercial polymers (i.e., polyimides) usually with a CO₂
26 laser. This is a photothermal process driven by the high pressure and temperatures around
27 the laser spot.^[43,44] In this case, LIG synthesis was adapted to preserve the integrity of the
28 masks. The process provided superhydrophobic masks and can be easily brought to large-
29 scale manufacturing. Besides, the strong light absorption and superhydrophobicity of LIG
30 would permit the deactivation of most pathogens by only exposing it to the sun light. The strong
31 light response would allow the re-cycling of the masks in solar steam generators. In another
32 work, the health protection capacity of commercial melt-blown fabrics (MBF) and commercial
33 masks with activated carbon fiber (ACF) was compared to LIG.^[45] *Escherichia coli* was used
34 as a model microorganism with an inactivation temperature over 60 °C. Interestingly, LIG
35 showed 81.57 % intrinsic antibacterial activity in contrast to 9.13 and 2.0 % of MBF and ACF,
36 respectively. After 10 min of light irradiation, LIG films reached 62 °C, which resulted in an

1 enhanced antibacterial activity of 99.998%, while ACF reached only 52 °C and 67.24 % activity.
2 On the other hand, MBF fibres reached only 36 °C, yet its activity raised up to 85.3%,
3 suggesting that temperature is not the only important factor.

4 The use of graphene in masks was not limited to LIG. For instance, a prototype filter was
5 designed as a disposable part of reusable respiratory masks.^[46] The structural part of the filter
6 was composed of multilayers of electrospun polylactic acid and cellulose acetate. By adding
7 CuO nanoparticles and GO in a post-processing step using ultrasonication, the mechanical,
8 antiviral and antibacterial properties were enhanced. The photothermal activity of LIG is
9 expected also for other GRMs. Remarkably, LIG grows tightly on the substrates, which
10 envisions certain stability. The stability of GRMs masks thereof will depend on the kind of
11 GRMs and the deposition technique, being key factors in view of the incorporation of GRMs-
12 containing masks into the market.

13



14

15 **Figure 2.** Chemical functionalization of GRMs to generate antiviral coatings on fomites. The
16 inset box illustrate several common techniques used for GRMs deposition. Adapted with
17 permission.^[33] Copyright 2019, John Wiley & Sons, Inc.

18

19 (Bio)sensing

20 A recent mathematical study has suggested the benefits of proper testing, tracing, isolation
21 and quarantine of population to control the spread of viral infections.^[47] Unfortunately,

1 scenarios with confirmed cases around 20000 per day would require to quarantine over
2 500000 people ideally for two weeks. Such circumstances impose to carry out a vast number
3 of tests every single day. Alongside, the silent state of the infection leads to fast propagation
4 of the virus, which is in contrast with the slow available testing methodologies. Current testing
5 relies mainly on the use of reverse transcription polymerase chain reaction (RT-PCR). Testing
6 by means of this method requires few hours and the intervention of trained personnel at every
7 step of the analysis including sampling, sample treatment and data interpretation.
8 Consequently, fast and reliable methods without the use of sophisticated equipment or trained
9 staff at a reasonable cost are needed for proper triage of SARS-CoV-2 in the current pandemic.

10 The electronic properties, high surface area-to-volume ratio and the ultra-high response to
11 changes in the environment make GRMs ideal candidates for the development of novel and
12 highly sensitive sensors associated with SARS-CoV-2 and other viruses. Sensing using GRMs
13 and other 2D materials is a growing field that have provided very promising results in the last
14 years.^[48,49] The available know-how is serving as a solid starting point for the development of
15 new sensors. The process for the development of GRMs-based sensors is illustrated in Figure
16 3. To the best of our knowledge, Seo *et al.* reported for the first time the use of G for the
17 detection of SARS-CoV-2 in nasopharyngeal swab specimens.^[50] In this work, CVD G was
18 transferred onto SiO₂/Si substrates and patterned into linear shapes with photolithography
19 before growing top Au/Cr electrodes. SARS-CoV-2 spike antibody was then immobilized onto
20 G using a bifunctional linker containing pyrene that binds to G via π - π stacking interaction,
21 and an activated carboxylic acid to form a stable amide bond with the antibody. The use of the
22 antibody enabled the specific detection of the spike protein, a transmembrane protein that
23 differs from one coronavirus to another. This device architecture permitted to evaluate the
24 field-effect transistor biosensing response to three different samples including the SARS-CoV-
25 2 antigen protein, the cultured SARS-CoV-2 and real clinical samples. Interestingly, the
26 authors reported a limit of detection (LOD) of 1 or 100 fg/ml of protein in PBS or universal
27 transport medium, respectively, 1.6×10^1 pfu/ml of the cultured virus with a linear response up
28 to 1.6×10^4 pfu/ml. Besides, Mers-CoV spike protein gave no response, suggesting high
29 specificity of the biosensor. Although the high complexity of the clinical samples provided a
30 signal with high noise background, it was possible to distinguish between positive and negative
31 patients. The LOD in nasopharyngeal swabs was 242 copies/ml, which is close to the \sim 100
32 copies/ml of the most performing RT-PCR.^[51]

33 Alternatively, G-based electrochemical biosensors are attracting a lot of attention for SARS-
34 CoV 2 detection. The integration of this biosensors into electrochemical devices has evolved
35 rapidly in the last years allowing the fabrication of point of care testing of relevant biomolecules.
36 ^[52,53] A supersandwich-type electrochemical device able to detect RNA of SARS-CoV-2

1 without nucleic acid amplification or reverse transcription was recently developed.^[54] In this
2 type of sensors, the target oligonucleotide sequence hybridizes with the capture probe and
3 the signal probe (i.e., sandwich), which hybridizes in turn with another extreme of the target
4 strand forming long concatamers (i.e., supersandwich) that amplify the electrochemical signal.
5 A sophisticated synthetic approach was applied to prepare the active material, which is
6 composed of Au@Fe₃O₄ nanoparticles decorated with a short oligonucleotide capture probe
7 for the viral RNA, and rGO functionalized with Au nanoparticles binding the labelled signal
8 probe. In addition, the basal plane of rGO was functionalized with p-sulfocalix[8]arene hosting
9 toluidine B to improve the electrochemical signal. The electrochemical response to the SARS-
10 CoV-2 RNA was measured by differential pulse voltammetry using an electrochemical
11 workstation operated by a smartphone. Remarkably, a LOD of 200 copies/ml was achieved
12 with this sensor, which outperforms the LOD for viral RNA reported in literature, and was
13 suitable for different clinical samples including throat swabs, saliva, urine, sputum, etc.
14 Moreover, the sensor displayed no significant response when it was exposed to samples from
15 other viruses (e.g., Mers-CoV, influenza A, rhinoviruses, etc.), which confirmed its high
16 specificity and selectivity.

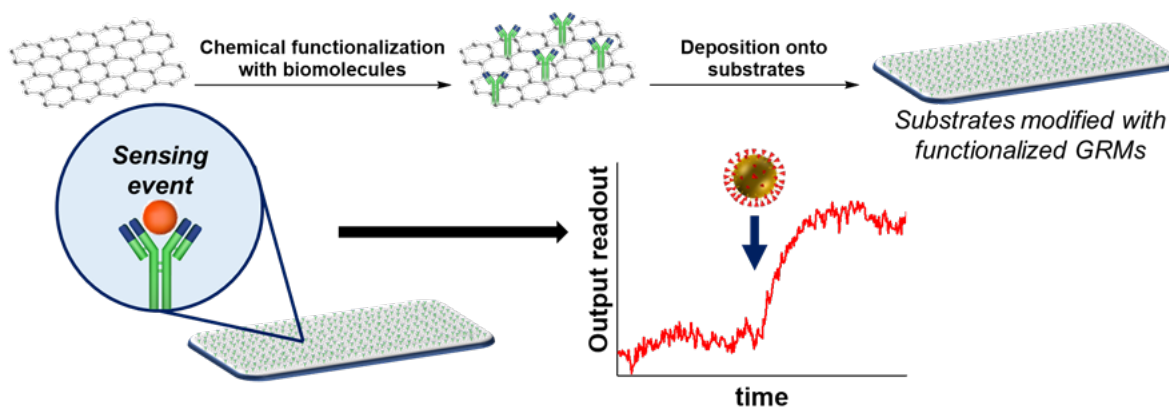
17 Beside the virus itself, the detection of other biomarkers associated to the disease is of vital
18 importance to control a pandemic. The identification and early detection of biomarkers can
19 anticipate dramatically the diagnosis with the consequent increase of survival rate in many
20 diseases. In this regard, high levels of C-reactive protein have been found in patients with
21 severe symptoms of COVID-19.^[55] Moreover, the presence or the absence of the specific
22 immunoglobulins IgM and IgG provides relevant information on the immune response of the
23 tested individuals. A smart-solution to detect the spike protein of SARS-CoV-2, the associated
24 immunoglobulins and the C-reactive protein in either blood or saliva using a multiplexed device
25 based on functionalized G was reported.^[56] To construct the device, a four channel LIG
26 electrode was engraved onto polyimide with a CO₂ laser. Pyrenebutyric acid was used as a
27 linker for the modification of each working electrode with the specific reagent for the
28 corresponding biomarker. Once assembled, the performance of the multiplexed device was
29 measured by amperometry against Ag/AgCl reference electrode fabricated by
30 electrodeposition. Incubation time as short as 10 min was enough to obtain a linear response
31 within a physiologically relevant range. Furthermore, the sensor was integrated in a device
32 powered by a small lithium-ion polymer battery. The device was able to measure and send
33 the data simultaneously *via* Bluetooth technology. The applicability of the device in real serum
34 and saliva samples was proven both for positive and negative cases. The potential use of non-
35 invasive sampling (i.e., saliva), together with the fast response and good performance without

1 sophisticated laboratory equipment or trained personnel at a reasonable price make this
2 pioneer device a strong candidate for fast patient triage to block the spread of the virus.

3 In most of the described works, a strong focus was given to achieve high specificity and
4 selectivity for SARS-CoV-2, which was assessed by exposing the sensors to different viruses.
5 A completely different approach was proposed by Hashemi *et al.*^[57] The authors put the focus
6 on the electrochemical sensing of glycoproteins by differential pulse voltammetry. The active
7 material consisted of functionalized GO decorated with Au nanostars deposited on glassy
8 carbon electrodes. This biosensor provided low LOD for SARS-Cov-2, infectious bronchitis
9 virus, avian influenza and two strains of Newcastle disease virus. The high performance was
10 attributed to the high adsorption capacity of functionalized GO and the electron transfer
11 mediated by the Au nanostars. The test could be done quickly using saliva. Interestingly, the
12 differential pulse voltammetry displayed a different pattern for all the tested viruses. This is of
13 high importance since distinguishing between the SARS-CoV-2 and other viruses is frequently
14 a challenging task because of the similarity of the symptoms. The concept developed here
15 could be of extreme importance to discriminate people infected with concomitant viruses (e.g.,
16 SARS-CoV-2 and influenza in winter) and stop the spread of dangerous pathogens.

17 Yet, novel sensors should not be limited to biomarkers. For instance, a high detection rate of
18 SARS-CoV-2 was achieved through the analysis of forced cough recording with artificial
19 intelligent.^[58] GRMs-based architectures responsive to sound^[59] or pressure^[60] could find
20 application here. In addition, integration of 2D material-based thermometers into wearables
21 could constantly measure the temperature of an individual. Sensors based on 2D materials
22 could find application to assess the state of personal protective equipment like masks. Playing
23 with the experimental conditions for the synthesis of LIG, hygroelectric devices were integrated
24 into surgical masks.^[45] The devices consisted of films of LIG where the hydrophilicity (i.e., the
25 level of oxidation) changed gradually along the device. This generates a gradient of protons
26 in the presence of moisture that induces a measurable electric field. The performance of this
27 device decreases significantly with the accumulation of bacteria, and therefore it could be used
28 to evaluate the masks conditions.

29



1

2 **Figure 3.** Functionalization of GRMs with specific biomolecules for the sensing of SARS-CoV-
 3 2. GRMs derivatization can be done either before or after the deposition onto the substrates.
 4 Most of the sensing events described so far include the interaction of a specific biomolecule
 5 with functionalized GRMs. The first attempts mainly included sensors with electrical readout,
 6 yet other outputs (e.g., spectroscopic signal) are foreseen.

7

8 **Conclusion and outlooks**

9 Unexpected problems require unexpected solutions. The outbreak of the current pandemic is
 10 urging scientists to be creative in the development of novel strategies to fight against the
 11 spread of SARS-CoV-2. Such circumstances impose (more than ever) a strong collaboration
 12 between laboratories with different background. Here, we have provided some examples
 13 demonstrating that thanks to their chemical and physical properties, GRMs can be key
 14 components offering innovative solutions in antiviral research. When suitably functionalized
 15 GRMs are able to interfere with viral replication machinery, drop down viral survival on
 16 surfaces, and selectively and sensibly detect capsid proteins making them powerful allies
 17 against this and future pandemics.

18 On the short-term, by taking advantage of the industrial scaled production of 2D materials,
 19 antiviral surfaces and coatings as well as sensors based on GRMs can be realized by making
 20 use of the tools described above. Alongside, appropriate chemical modifications should be
 21 exploited to enhance the functional control over these materials. The use of cost-effective
 22 coating techniques may allow large-area GRMs deposition on surfaces of different nature.
 23 Fine control of the chemistry of GRMs, preferably through stable covalent bonding, should be
 24 directed to generate robust coatings to expand their utility life (e.g., resistance to washing or
 25 to ambient exposure). The ideal coatings should present broad-spectrum antimicrobial
 26 activities both for harmful viruses and bacteria. A thorough evaluation against real pathogens
 27 would therefore be required. However, access to these assays is generally limited and require
 28 a lot of resources. In addition, while the design of new GRMs can be developed in different

1 research groups, we must keep in mind that only few labs are equipped with access to the
2 pathogenic viruses and so can study the efficacy of GRMs as antiviral materials. Therefore,
3 calls for new collaborations, establishment of excellence specialized scientific clusters and
4 financial supports to innovative research initiatives are mandatory for an efficient fight against
5 new viral pandemics.

6 The remarkable results on the sensing of SARS-CoV-2 using GRMs point to a promising future.
7 Importantly, the approaches developed for graphene may be exploited with other 2D materials
8 (e.g., MoS₂) offering a broader choice of physical and chemical properties, towards the design
9 of new sensors beyond GRMs. Most of the proposed strategies involved the use of non-
10 covalent chemistry to immobilize large biomolecules on the flat surface of 2D materials.
11 Consequently, key aspects such as the reproducibility during the large-scale production and
12 the stability of the devices over time has to be assessed before bringing these sensors to the
13 market.

14 Application of GRMs and in general of hard nanomaterials in drug delivery has to be
15 considered in a long-term perspective. Multifunctionalization strategies and antiviral efficacy
16 must be still deepened. Additionally, there are different concerns about biocompatibility and
17 long-term toxicity of 2D materials. Although studies are already available,^[61] the toxicological
18 risks of GRMs are not completely cleared. In this context, we solicit the scientific community
19 work for the standardization of the tested GRMs in order to obtain robust answers for their
20 safe applications in any field. Different hard nanomaterials are now clinically approved for
21 human treatment,^[62] and we hope that some of the GRMs will be soon included into this list.
22 Despite the current extraordinary results, GRMs have been only marginally explored in viral
23 infections. In particular, their potential in blocking cell uptake, inhibiting viral replication and
24 alerting the immune system has not been unleashed yet. There is still a long way to go before
25 establishing GRMs as optimal solution to combat SARS-CoV-2. However, by considering the
26 huge potential held by such unique materials in view of their unique properties, which can be
27 further modulated via controlled functionalization, one can foresee a bright future for GRMs in
28 the fight against the current COVID-19 and future pandemics.

29

30 **Acknowledgements**

31 This work was supported the Agence Nationale de la Recherche (ANR) through the LabEx
32 project Chemistry of Complex Systems (ANR-10-LABX-0026_CSC), the Centre National de
33 la Recherche Scientifique (CNRS), the International Center for Frontier Research in Chemistry

1 (icFRC) and the EC through the Graphene Flagship Core 3 project (GA- 881603) and the ERC
2 project SUPRA2DMAT (GA-833707).

3

4 **References**

- 5 [1] V. Palmieri, M. Papi, *Nano Today* **2020**, 33, DOI 10.1016/j.nantod.2020.100883.
- 6 [2] D. Almaghaslah, G. Kandasamy, M. Almanasef, R. Vasudevan, S. Chandramohan,
7 *Int. J. Clin. Pract.* **2020**, 74, DOI 10.1111/ijcp.13637.
- 8 [3] P. Wick, A. E. Louw-Gaume, M. Kucki, H. F. Krug, K. Kostarelos, B. Fadeel, K. A.
9 Dawson, A. Salvati, E. Vázquez, L. Ballerini, M. Tretiach, F. Benfenati, E. Flahaut, L.
10 Gauthier, M. Prato, A. Bianco, *Angew. Chemie - Int. Ed.* **2014**, 53, 7714.
- 11 [4] A. Bianco, H. M. Cheng, T. Enoki, Y. Gogotsi, R. H. Hurt, N. Koratkar, T. Kyotani, M.
12 Monthieux, C. R. Park, J. M. D. Tascon, J. Zhang, *Carbon* **2013**, 65, 1.
- 13 [5] G. Reina, J. M. González-Domínguez, A. Criado, E. Vázquez, A. Bianco, M. Prato,
14 *Chem. Soc. Rev.* **2017**, 46, 4400.
- 15 [6] V. Cagno, E. D. Tseligka, S. T. Jones, C. Tapparel, *Viruses* **2019**, 11, DOI
16 10.3390/v11070596.
- 17 [7] J. Liu, J. Li, K. Arnold, R. Pawlinski, N. S. Key, *Res. Pract. Thromb. Haemost.* **2020**,
18 4, 518.
- 19 [8] G. Reina, S. Peng, L. Jacquemin, A. F. Andrade, A. Bianco, *ACS Nano* **2020**, 14,
20 9364.
- 21 [9] Z. Qi, P. Bharate, C. H. Lai, B. Ziem, C. Böttcher, A. Schulz, F. Beckert, B. Hatting, R.
22 Mülhaupt, P. H. Seeberger, R. Haag, *Nano Lett.* **2015**, 15, 6051.
- 23 [10] J. Luczkowiak, A. Muñoz, M. Sánchez-Navarro, R. Ribeiro-Viana, A. Ginieis, B. M.
24 Illescas, N. Martín, R. Delgado, J. Rojo, *Biomacromolecules* **2013**, 14, 431.
- 25 [11] M. Sametband, I. Kalt, A. Gedanken, R. Sarid, *ACS Appl. Mater. Interfaces* **2014**, 6,
26 1228.
- 27 [12] B. Ziem, H. Thien, K. Achazi, C. Yue, D. Stern, K. Silberreis, M. F. Gholami, F.
28 Beckert, D. Gröger, R. Mülhaupt, J. P. Rabe, A. Nitsche, R. Haag, *Adv. Healthc.*
29 *Mater.* **2016**, 5, 2922.
- 30 [13] B. Ziem, J. Rahn, I. Donskyi, K. Silberreis, L. Cuellar, J. Dervede, G. Keil, T. C.
31 Mettenleiter, R. Haag, *Macromol. Biosci.* **2017**, 17, DOI 10.1002/mabi.201600499.
- 32 [14] I. S. Donskyi, W. Azab, J. L. Cuellar-Camacho, G. Guday, A. Lippitz, W. E. S. Unger,
33 K. Osterrieder, M. Adeli, R. Haag, *Nanoscale* **2019**, 11, 15804.
- 34 [15] J. B. GC, R. Pokhrel, N. Bhattarai, K. A. Johnson, B. S. Gerstman, R. V. Stahelin, P.
35 P. Chapagain, *Biochem. Biophys. Res. Commun.* **2017**, 493, 176.
- 36 [16] M. Zhang, X. Mao, C. Wang, W. Zeng, C. Zhang, Z. Li, Y. Fang, Y. Yang, W. Liang,
37 C. Wang, *Biomaterials* **2013**, 34, 1383.
- 38 [17] J. I. Paredes, S. Villar-Rodil, *Nanoscale* **2016**, 8, 15389.

- 1 [18] G. Reina, A. Ruiz, D. Murera, Y. Nishina, A. Bianco, *ACS Appl. Mater. Interfaces*
2 **2019**, *11*, 7695.
- 3 [19] Z. Song, X. Wang, G. Zhu, Q. Nian, H. Zhou, D. Yang, C. Qin, R. Tang, *Small* **2015**,
4 *11*, 1171.
- 5 [20] S. Ye, K. Shao, Z. Li, N. Guo, Y. Zuo, Q. Li, Z. Lu, L. Chen, Q. He, H. Han, **2015**, DOI
6 10.1021/acsami.5b06876.
- 7 [21] C. Liu, Q. Zhou, Y. Li, L. V. Garner, S. P. Watkins, L. J. Carter, J. Smoot, A. C. Gregg,
8 A. D. Daniels, S. Jervej, D. Albaiu, *ACS Cent. Sci.* **2020**, *6*, 315.
- 9 [22] X. Du, R. Xiao, H. Fu, Z. Yuan, W. Zhang, L. Yin, C. He, C. Li, J. Zhou, G. Liu, G.
10 Shu, Z. Chen, *Mater. Sci. Eng. C* **2019**, *105*, 110052.
- 11 [23] X. X. Yang, C. M. Li, Y. F. Li, J. Wang, C. Z. Huang, *Nanoscale* **2017**, *9*, 16086.
- 12 [24] A. R. Deokar, A. P. Nagvenkar, I. Kalt, L. Shani, Y. Yeshurun, A. Gedanken, R. Sarid,
13 *Bioconjug. Chem.* **2017**, *28*, 1115.
- 14 [25] N. van Doremalen, T. Bushmaker, D. H. Morris, M. G. Holbrook, A. Gamble, B. N.
15 Williamson, A. Tamin, J. L. Harcourt, N. J. Thornburg, S. I. Gerber, J. O. Lloyd-Smith,
16 E. de Wit, V. J. Munster, *N. Engl. J. Med.* **2020**, DOI 10.1056/nejmc2004973.
- 17 [26] S. J. Smither, L. S. Eastaugh, J. S. Findlay, M. S. Lever, S. J. Smither, L. S.
18 Eastaugh, J. S. Findlay, M. S. Lever, *Emerg. Microbes Infect.* **2020**, *9*, 1415.
- 19 [27] S. Riddell, S. Goldie, A. Hill, D. Eagles, T. W. Drew, *Viol. J.* **2020**, *17*, 145.
- 20 [28] L. Díaz-marug, M. García, C. Bort, M. Fern, V. Fanjul, *Environ. Res.* **2021**, *192*,
21 110293.
- 22 [29] X. Xue, J. K. Ball, C. Alexander, M. R. Alexander, *Matter* **2020**, *3*, 1433.
- 23 [30] H. Liu, I. Elkin, J. Chen, A. M. Klibanov, *Biomacromolecules* **2015**, *16*, 351.
- 24 [31] N. D. Q. Chau, G. Reina, J. Raya, I. A. Vacchi, C. Ménard-Moyon, Y. Nishina, A.
25 Bianco, *Carbon* **2017**, *122*, 643.
- 26 [32] D. Pakulski, W. Czepa, S. Witomska, A. Aliprandi, V. Patroniak, A. Ciesielski, P.
27 Samorì, *J. Mater. Chem. A* **2018**, *6*, 9384.
- 28 [33] S. Witomska, T. Leydecker, A. Ciesielski, P. Samorì, *Adv. Funct. Mater.* **2019**, *29*,
29 1901126.
- 30 [34] A. Aliprandi, T. Moreira, C. Anichini, M. A. Stoeckel, M. Eredia, U. Sassi, M. Bruna, C.
31 Pinheiro, C. A. T. Laia, S. Bonacchi, P. Samorì, *Adv. Mater.* **2017**, *29*, 1.
- 32 [35] N. Karim, S. Afroj, S. Tan, P. He, A. Fernando, C. Carr, K. S. Novoselov, *ACS Nano*
33 **2017**, *11*, 12266.
- 34 [36] Y. Guo, G. Xu, X. Yang, K. Ruan, T. Ma, Q. Zhang, J. Gu, Y. Wu, H. Liu, Z. Guo, *J.*
35 *Mater. Chem. C* **2018**, *6*, 3004.
- 36 [37] M. P. Arrieta, L. Peponi, J. Rodríguez-Hernández, in *Mater. Biomed. Eng. Bioact.*
37 *Mater. Antimicrob. Anticancer Gene Ther.*, **2019**, pp. 53–76.
- 38 [38] Y. Gao, Y. B. Truong, Y. Zhu, I. L. Kyratzis, *J. Appl. Polym. Sci.* **2014**, *131*, 40797.

- 1 [39] A. F. De Faria, E. Shaulsky, L. H. A. Chavez, M. Elimelech, *ACS Appl. Mater.*
2 *Interfaces* **2015**, *7*, 12751.
- 3 [40] J. J. Klemeš, Y. Van Fan, P. Jiang, *Int. J. Energy Res.* **2020**, *1*.
- 4 [41] M. J. Pascoe, A. Robertson, A. Crayford, E. Durand, J. Steer, A. Castelli, R. Wesgate,
5 S. L. Evans, A. Porch, J. Y. Maillard, *J. Hosp. Infect.* **2020**, *106*, 10.
- 6 [42] H. Zhong, Z. Zhu, J. Lin, C. F. Cheung, V. L. Lu, F. Yan, C. Y. Chan, G. Li, *ACS Nano*
7 **2020**, *14*, 6213.
- 8 [43] J. Lin, Z. Peng, Y. Liu, F. Ruiz-Zepeda, R. Ye, E. L. G. Samuel, M. J. Yacaman, B. I.
9 Yakobson, J. M. Tour, *Nat. Commun.* **2014**, *5*, 5.
- 10 [44] R. Ye, D. K. James, J. M. Tour, *Acc. Chem. Res.* **2018**, *51*, 1609.
- 11 [45] L. Huang, S. Xu, Z. Wang, K. Xue, J. Su, Y. Song, S. Chen, C. Zhu, B. Z. Tang, R.
12 Ye, *ACS Nano* **2020**, *14*, 12045.
- 13 [46] M. K. Ahmed, M. Afifi, V. Uskoković, *J. Infect. Public Health* **2020**, *13*, 1243.
- 14 [47] A. J. Kucharski, P. Klepac, A. J. K. Conlan, S. M. Kissler, M. L. Tang, H. Fry, J. R.
15 Gog, W. J. Edmunds, J. C. Emery, G. Medley, J. D. Munday, T. W. Russell, Q. J.
16 Leclerc, C. Diamond, S. R. Procter, A. Gimma, F. Y. Sun, H. P. Gibbs, A. Rosello, K.
17 van Zandvoort, S. Hué, S. R. Meakin, A. K. Deol, G. Knight, T. Jombart, A. M. Foss,
18 N. I. Bosse, K. E. Atkins, B. J. Quilty, R. Lowe, K. Prem, S. Flasche, C. A. B. Pearson,
19 R. M. G. J. Houben, E. S. Nightingale, A. Endo, D. C. Tully, Y. Liu, J. Villabona-
20 Arenas, K. O'Reilly, S. Funk, R. M. Eggo, M. Jit, E. M. Rees, J. Hellewell, S. Clifford,
21 C. I. Jarvis, S. Abbott, M. Auzenbergs, N. G. Davies, D. Simons, *Lancet Infect. Dis.*
22 **2020**, *20*, 1151.
- 23 [48] C. Anichini, W. Czepa, D. Pakulski, A. Aliprandi, A. Ciesielski, P. Samorì, *Chem. Soc.*
24 *Rev.* **2018**, *47*, 4860.
- 25 [49] Cecilia Menard-Moyon, A. Bianco, K. Kalantar-Zadeh, *ACS Sensors* **2020**, in press.
- 26 [50] G. Seo, G. Lee, M. J. Kim, S. H. Baek, M. Choi, K. B. Ku, C. S. Lee, S. Jun, D. Park,
27 H. G. Kim, S. J. Kim, J. O. Lee, B. T. Kim, E. C. Park, S. Il Kim, *ACS Nano* **2020**, *14*,
28 5135.
- 29 [51] R. Arnaout, R. Lee, G. R. Lee, C. Callahan, C. Yen, K. Smith, R. Arora, J. Kirby,
30 *bioRxiv* **2020**, <https://doi.org/10.1101/2020.06.02.131144> .
- 31 [52] A. T. Lawal, *Biosens. Bioelectron.* **2018**, *106*, 149.
- 32 [53] E. T. S. G. da Silva, D. E. P. Souto, J. T. C. Barragan, J. de F. Giarola, A. C. M. de
33 Moraes, L. T. Kubota, *ChemElectroChem* **2017**, *4*, 778.
- 34 [54] H. Zhao, F. Liu, W. Xie, T. C. Zhou, J. OuYang, L. Jin, H. Li, C. Y. Zhao, L. Zhang, J.
35 Wei, Y. P. Zhang, C. P. Li, *Sensors Actuators, B Chem.* **2021**, *327*, 128899.
- 36 [55] C. Wu, X. Chen, Y. Cai, J. Xia, X. Zhou, S. Xu, H. Huang, L. Zhang, X. Zhou, C. Du,
37 Y. Zhang, J. Song, S. Wang, Y. Chao, Z. Yang, J. Xu, X. Zhou, D. Chen, W. Xiong, L.
38 Xu, F. Zhou, J. Jiang, C. Bai, J. Zheng, Y. Song, *JAMA Intern. Med.* **2020**, *180*, 934.
- 39 [56] R. M. Torrente-rodri, H. Lukas, J. Tu, C. Xu, H. B. Rossiter, H. Lukas, J. Tu, J. Min, Y.
40 Yang, *Matter* **2020**, *3*, 1.

- 1 [57] S. A. Hashemi, N. G. G. Behbahan, S. Bahrani, S. M. Mousavi, A. Gholami, S.
2 Ramakrishna, M. Firoozsani, M. Moghadami, K. B. Lankarani, N. Omidifar, *Biosens.*
3 *Bioelectron.* **2021**, *171*, 112731.
- 4 [58] J. Laguarda, F. Hueto, B. Subirana, *IEEE Open J. Eng. Med. Biol.* **2020**,
5 10.1109/OJEMB.2020.3026928,.
- 6 [59] L. Tao, H. Tian, Y. Liu, Z. Ju, Y. Pang, Y. Chen, D. Wang, X. Tian, J. Yan, N. Deng, Y.
7 Yang, T. Ren, *Nat. Chem.* **2017**, *8*, 14579.
- 8 [60] C. B. Huang, S. Witomska, A. Aliprandi, M. A. Stoeckel, M. Bonini, A. Ciesielski, P.
9 Samorì, *Adv. Mater.* **2019**, *31*, 1804600.
- 10 [61] B. Fadeel, C. Bussy, S. Merino, E. Vázquez, E. Flahaut, F. Mouchet, L. Evariste, L.
11 Gauthier, A. J. Koivisto, U. Vogel, C. Martín, L. G. Delogu, T. Buerki-Thurnherr, P.
12 Wick, D. Beloin-Saint-Pierre, R. Hischer, M. Pelin, F. Candotto Carniel, M. Tretiach,
13 F. Cesca, F. Benfenati, D. Scaini, L. Ballerini, K. Kostarelos, M. Prato, A. Bianco, *ACS*
14 *Nano* **2018**, *12*, 10582.
- 15 [62] S. Nardecchia, P. Sánchez-Moreno, J. de Vicente, J. A. Marchal, H. Boulaiz,
16 *Nanomaterials* **2019**, *9*, 191.
- 17