

INTERROGATION OF MICROARRAY DATASETS INDICATES THAT MACROPHAGE-SECRETED FACTORS STIMULATE THE EXPRESSION OF GENES ASSOCIATED WITH VITAMIN D METABOLISM (*VDR* AND *CYP27B1*) IN HUMAN ADIPOCYTES

Paul Trayhurn^{1,2}, Adrian O'Hara¹, and Chen Bing¹

¹Obesity Biology Research Unit, School of Clinical Sciences, University of Liverpool, Liverpool, UK and ²Clore Laboratory, University of Buckingham, Buckingham, UK

Abstract

Microarray datasets have been interrogated to determine whether the expression of vitamin D-related genes is modulated in adipocytes during inflammation. The datasets were from human adipocytes and preadipocytes incubated with macrophageconditioned medium (U937 cells). In adipocytes, exposure to the conditioned medium for 24 h resulted in a major increase (82.2-fold) in mRNA level of CYP27B1, the gene encoding the enzyme that converts 25-hydroxycholecalciferol to the active form of vitamin D₂, 1,25-dihydroxycholecalciferol; exposure for 4 h also raised CYP27B1 mRNA level (10.9-fold). The level of the mRNA encoding the vitamin D receptor (VDR) was increased after 24 h (7.7-fold), but there was no change at 4 h. In contrast, incubation with conditioned medium for either 4 or 24 h had no effect on the expression of the CYP24 and CYP2R1 genes, which encode enzymes that catalyse a 24-hydroxylation and the conversion of vitamin D₃ to 25-hydroxycholecalciferol, respectively. In preadipocytes, the only effect of the macrophage-conditioned medium was to stimulate CYP27B1 expression (5.7-fold) after 24 h. It is concluded that the capacity of adipocytes to produce active vitamin D, hormone and its nuclear receptor is strongly upregulated by secretory products from macrophages; this is consistent with a counter-regulatory effect of the vitamin D system to ameliorate inflammation.

Adipobiology 2011; 3:31-36

Keywords: 1,25-dihydroxycholecalciferol, 25-hydroxycholecalciferol, inflammation, vitamin D receptor

Correspondence: Professor Paul Trayhurn FRSE, Clore Laboratory, University of Buckingham, Hunter Street, Buckingham MK18 1EG, UK. Fax: +44 1280 820 135; E-mail: p.trayhurn@liverpool.ac.uk; paul.trayhurn@buckingham.ac.uk

Introduction

Microarray studies have examined global gene expression in adipose tissue and in adipocytes in particular, and the changes that take place in response to specific stimuli (1-5). Our own studies have focused on the effects of macrophage secretions on gene expression in human adipocytes and preadipocytes in culture (6-7). Bioinformatic analysis of microarray data allows major networks and pathways that are modulated in particular conditions, or in response to specific stimuli, to be identified. These pathways, together with the individual genes whose expression is altered most extensively, are normally the main focus of interest; this is certainly so during the initial analysis. For example, in our studies on the effect of macrophage secretions on adipocyte gene expression, a group of matrix metalloproteinases (MMP1, MMP3, MMP10) where the mRNA level increased by >1000-fold were of key interest, as were the major pathways

Received 18 December 2011, accepted 23 December 2011.

activated - which included those associated with inflammation and macrophage infiltration (6).

Microarrays are a rich source of data beyond the major pathways and highly differentially expressed genes, and interrogation of the datasets can provide important insight in unexpected areas. One of the emerging interests in adipobiology is the extent to which adipocytes are involved in vitamin D metabolism, both in relation to the storage of the hormone and as a target tissue. In this short article we describe the outcome of examination of our microarray data for genes encoding the vitamin D receptor (*VDR*) and enzymes involved in the synthesis of the active hormone (*CYP27B1* and *CYP2R1*).

Vitamin D

The vitamin D endocrine system has been linked historically to calcium homeostasis and bone metabolism, with rickets being the classical deficiency disease (8-10). However, it is increasingly evident that this hormonal system is involved in a range of physiological functions. The main route by which vitamin D is obtained is by endogenous synthesis in the skin, though provision through the diet is important for those populations who live in regions where sunlight is limited, or where exposure to the sun is restricted for cultural reasons or because of factors such as age. The main dietary sources of vitamin D are oily fishes (especially the liver), eggs, and full-fat milk.

The term 'vitamin D' is used somewhat loosely and it is important to note that it is not strictly a vitamin in the sense of being an essential dietary nutrient, given the synthesis in the skin. Furthermore, vitamin D in practise refers essentially to D_3 (cholecalciferol), the only form of the vitamin that is found naturally in humans and other animals (8-9).

Vitamin D metabolism

Vitamin D_3 is synthesised in the skin non-enzymatically from 7-dehydrocholesterol through irradiation by UV light. The product of irradiation, precalciferol, isomerises to cholecalciferol. This is then transported to the liver bound to a plasma transport protein, Gc-globulin (also known as transcalciferin). Dietary-derived vitamin D_3 is absorbed and transferred to the liver in chylomicrons. Vitamin D_3 , whether from endogenous synthesis in the skin or from the diet, is then hydroxylated in the liver to form 25-hydroxycholecalciferol (or calcidiol). The released 25(OH) D_3 , which is the main circulating form of vitamin D_3 hormone, is further hydroxylated to form 1,25-dihydroxycholecalciferol (or calcitriol), and this occurs primarily in the kidney (8-9). The enzyme that catalyses this second hydroxylation is 25(OH) D_3 -1 α -hydroxylase, and is present in at least ten other tissues additional to the kidney (9). A 24-hydroxylation can also occur in the kidney (and other tissues) to form 24,25-dihydroxycholecalciferol. The main route for the degradation of $1,25(OH)_2D_3$ is also through a 24-hydroxylation.

There are two modes of action of $1,25(OH)_2D_3$ – genomic and non-genomic (11). The actions on gene transcription are mediated through a specific nuclear receptor, the vitamin D receptor (VDR). This receptor is widely expressed, expression being reported in a number of tissues (11). The transcription of as many as 500 genes may be regulated through VDR. There are also rapid responses to $1,25(OH)_2D_3$ which occur within minutes and which do not involve gene transcription, such as the stimulation of intestinal calcium transport. The current view is that VDR is present not only in the nucleus, but also in calveolae associated with the plasma membrane of target cells, and is the receptor for the non-genomic as well as genomic actions of $1,25(OH)_2D_3$ (11).

Functions of vitamin D

Vitamin D has long been recognised to play a central role in the stimulation of calcium absorption by the intestine and in the mineralisation and re-modelling of bone (8-10). Indeed, vitamin D deficiency is closely associated with the development of rickets in children and osteomalacia in adults. Other functions are now increasingly evident, particularly in relation to the immune system (10,12,13). Examples include an anti-inflammatory action in macrophages, with the down-regulation of the production of tumor necrosis factor-alpha (TNF- α) through a decrease in NFkB activity (14-15). Indeed, an anti-inflammatory action seems to be a characteristic of 1,25(OH)₂D₃. Other major actions of the hormone include the stimulation of insulin secretion by pancreatic β cells, vitamin D deficiency leading to the inhibition of secretion (9, 16).

A growing number of diseases have been linked to vitamin D status, apart from rickets and osteomalacia, and these include several types of cancer (breast, colon and prostate) (9). In addition, inflammatory bowel disease, hypertension, peridontal disease, multiple sclerosis and muscle weakness (especially in the elderly) have each been associated with vitamin D insufficiency (9-10,13). Of particular relevance to obesity is the proposed link with cardiometabolic diseases (atherosclerosis, hypertension, type 2 diabetes and the metabolic syndrome). Indeed, a recent editorial in the *Journal of Clinical Endocrinology and Metabolism* was provocatively titled "25-OH vitamin D: is it the universal panacea for metabolic syndrome and type 2 diabetes?" (17).

Vitamin D status is considered to be best assessed by the serum level of $25(OH)D_3$ (9). The circulating level of $1,25(OH)_2D_3$ is of the order of 1000 times lower than that of $25(OH)D_3$. Several epidemiological studies have demonstrated an inverse corre-

lation in adults between body mass index (BMI) and circulating $25(OH)D_3$ level (18-20). In practise, it is the amount of body fat that is important. The explanation for the relationship with obesity is not clear, but it is possible that the hormone is sequestered within adipose tissue by virtue of the high lipid solubility. If this is the case, then low serum $25(OH)D_3$ levels are not indicative of an insufficiency of vitamin D *per se*. An alternative explanation for low serum levels in obesity is that adipocytes, or other cells within adipose tissue, have the capacity to degrade the steroid hormone.

Vitamin D and adipose tissue

White adipose tissue and adipocytes are among the extra-renal sites of expression of the 1a-hydroxylase gene (CYP27B1), and the conversion of 25(OH)D, to 1,25(OH),D, has been directly demonstrated, including in preadipocytes and in human mammary adipocytes (21-22). The vitamin D receptor is also expressed in mouse preadipocytes and adipocytes (22-26). Treatment of 3T3-L1 cells with 25(OH)D, has been reported to induce expression of CYP24, a 1,25(OH), D, responsive gene (21). There is also evidence that 1,25(OH)₂D₃ may increase lipid synthesis in human adipocytes; however, contrastingly, the expression of Insig-2, a gene that encodes a protein that is involved in inhibiting fat synthesis, is stimulated by 1,25(OH)₂D₃ (27). The augmentation of glucocorticoid production in adipocytes through a (small) upregulation of 11β-hydroxysteroid dehydrogenase type 1 expression by $1,25(OH)_2D_3$ has been observed (28). The favouring of inflammatory cytokine production (TNF-a and IL-6) in adipocytes and the suppression of anti-inflammatory cytokine production by $1,25(OH)_2D_2$ has also been reported in 3T3-L1 adipocytes and in human adipose cells (29). However, a very recent study showed that 1,25(OH), D, reduced the release of monocyte chemotactic protein-1 (MCP-1), also known as chemokine (C-C motif) ligand 2 (CCL2), and adiponectin by human adipocytes (30). This suggests that vitamin D₂ may have both pro- and anti-inflammatory effects in human fat cells.

Several studies have investigated the effects of $1,25(OH)_2D_3$ on adipocyte differentiation and proliferation, and while both an augmentation and an inhibition have been reported the growing consensus is that the hormone has primarily an inhibitory action on fat cell recruitment (22,31-32).

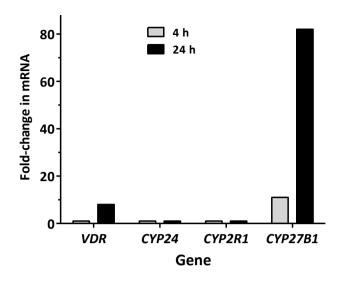
Interrogation of microarrays

In a preliminary exploration of the extent to which adipose tissue may be a significant site of vitamin D_3 metabolism and a target tissue of the hormone, we examined two different microarray datasets for the expression of genes involved in the vitamin D_3 endocrine system. The first microarrays were from a study on the effect of macrophage-conditioned (MC) medium on global gene expression in human adipocytes (Simpson-Golabi-Beymel Syndrome - SGBS) at short and longer time points – 4 and 24 h (6). The other was on the effects of MC medium on gene expression in human preadipocytes (SGBS), following a 24 h incubation with the medium (7). In both sets of studies, the comparison was with adipocytes/preadipocytes incubated in unconditioned macrophage (UC) medium – the medium used to culture macrophages, but which had not been exposed to cells (150 μ l of UC or MC medium added per ml of preadipocyte/ adipocyte medium). The macrophages were derived from U937 monocytes, which had been induced to differentiate through the addition of phorbol myristic acid.

The microarray data was interrogated for whether the treatments affected the expression of four genes associated with the metabolism and action of the vitamin D₂ system. These genes were: (i) CYP27B1 (encodes the 1a-hydroxylase), (ii) CYP2R1 (encodes the 25-hydroxylase), (iii) CYP24 (encodes the 24-hydroxylase that degrades 1,25(OH),D,), and (iv) VDR (encodes the vitamin D receptor). In human adipocytes exposed to MC medium, major changes in CYP27B1 expression were observed; the CYP27B1 mRNA level was increased 10.9 fold at 4 h and by as much as 82-fold at 24 h (Fig 1). At 24 h this gene was ranked number 40 out of 1307 genes whose expression was up-regulated by the conditioned medium, indicating that it is one of the most highly responsive genes to macrophage-secreted factors in human adipocytes. In contrast, there was no change in CYP24 expression, nor in CYP2R1. VDR mRNA level increased 7.7fold at 24 h, though there was no change at 4 h, indicating that expression of the vitamin D receptor is increased by prolonged exposure to MC medium (Fig 1).

In human preadipocytes exposed to MC medium for 24 h, no changes in VDR, CYP2R1, or CYP24 mRNA levels were observed (Fig 2). However, there was an increase in CYP27B1 mRNA level (5.7-fold) and this gene ranked at number 34 in the list of 401 genes that were up-regulated in preadipocytes by MC medium (Fig 2). The up-regulation of *CYP27B1* gene expression in preadipocytes induced by macrophage secretions indicates that the capacity of adipocytes to hydroxylate 25(OH)D₃ is not differentiation-dependent.

A further microarray dataset that was available to us on the effects of exposure to hypoxia $(1\% O_2)$ for 24 h on global gene expression in human adipocytes was also examined. No changes in expression were found for *CYP27B1*, *CYP24*, *CYP2R1*, or *VDR*, the mRNA levels for each of these genes being similar in adipocytes exposed to normoxia $(21\% O_2)$ or hypoxia. Thus vitamin D₃ system genes are not hypoxiasensitive in adipocytes.



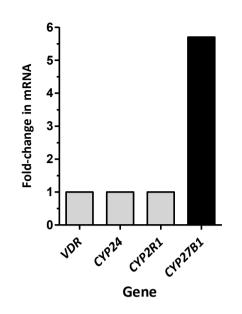


Figure 1. Effect of macrophage-conditioned medium on the expression of vitamin D₃-related genes in human adipocytes. Gene expression was assessed by microarrays following 4 and 24 h exposure to the conditioned medium. The fold-changes relate to the conditioned medium compared to unconditioned medium. The data may be reviewed through the following link:

http://www.ncbi.nlm.nih.gov/geo/query/acc.

cgi?token=rtmtvcmessiqutm&acc=

The GEO accession number is GSE14312.

Conclusion

This analysis of microarray datasets confirms that human adipocytes express the genes encoding the 1a-hydroxylase that converts 25(OH)D₃ to 1,25(OH)₂D₃, and the VDR, and demonstrates that the expression of both genes is markedly stimulated by MC medium. Thus the capacity of adipocytes to produce the active vitamin D₂ hormone and its nuclear receptor would appear to be strongly upregulated by secretory products released by macrophages (assuming that the changes in mRNA are mirrored at the protein level). Given the apparent anti-inflammatory actions of 1,25(OH), D₃, the upregulation of VDR and CYP27B1 expression in adipocytes on exposure to macrophage-secreted products would suggest a counter-regulatory response to ameliorate inflammation. The stimulation of VDR expression by inflammatory mediators is consistent with a local action within adipose tissue for increased 1,25(OH), D, produced during inflammation.

The strong upregulation of *CYP27B1* by MC medium raises the intriguing possibility that adipocytes are a significant site of

Figure 2. Effect of macrophage-conditioned medium on the expression of vitamin D_3 -related genes in human preadipocytes. Gene expression was assessed by microarrays following 24 h exposure to the conditioned medium. The fold-changes relate to the conditioned medium compared to unconditioned medium. The data may be reviewed through the following link:

http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?token = rfqfjamewuwmors&acc=

The GEO accession number is GSE27503.

the generation of $1,25(OH)_2D_3$ under inflammatory conditions. Obesity is, of course, characterised by chronic mild inflammation, and adipose tissue exhibits a substantial inflammatory response in the obese (33-35). This involves macrophage recruitment in the tissue, as well as the production of inflammation-related factors by adipocytes including pro-inflammatory cytokines and chemokines (36,37). The nature of the factors secreted from adipocytes that stimulate transcription of the *VDR* and *CYP27B1* genes in adipocytes is unclear. However, cytokines such as TNF- α and IL-1 β may well play a key role, similar to their action in stimulating matrix metalloproteinase expression and release (6).

In conclusion, the present report illustrates the information and insight that can be obtained through the mining of microarray datasets.

Acknowledgments

AOH gratefully acknowledges the receipt of an Industrial CASE Studentship from the Biotechnology and Biological Research Council (UK). The support of Dr Dawn Mazzatti and Dr Fei-Lim Ling (Unilever plc) is gratefully acknowledged.

References

- 1. Burton GR, Guan Y, Nagarajan R, McGehee Jr RE. Microarray analysis of gene expression during early adipocyte differentiation. *Gene* 2002; 293: 21-31.
- 2. Gabrielsson BG *et al.* High expression of complement components in omental adipose tissue in obese men. *Obesity Res* 2003; 11: 699-708.
- 3. Lopez IP *et al.* DNA microarray analysis of genes differentially expressed in diet-induced (cafeteria) obese rats. *Obesity Res* 2003; 11: 188-194.
- 4. Sjoholm K *et al.* A microarray search for genes predominantly expressed in human omental adipocytes: Adipose tissue as a major production site of serum amyloid A. *J Clin Endocrinol Metab* 2005; 90: 2233-2239.
- Yamashita A *et al.* DNA microarray analyses of genes expressed differentially in 3T3-L1 adipocytes co-cultured with murine macrophage cell line RAW264.7 in the presence of the Toll-like receptor 4 ligand bacterial endotoxin. *Int J Obesity* 2008; 32: 1725-1729.
- O'Hara A, Lim F-L, Mazzatti D, Trayhurn P. Microarray analysis identifies matrix metalloproteinases (MMPs) as key genes whose expression is up-regulated in human adipocytes by macrophage-conditioned medium. *Pflügers Archiv Eur J Physiol* 2009; 458: 1103-1114.
- O'Hara A, Lim F-L, Mazzatti D, Trayhurn P. Stimulation of inflammatory gene expression in human preadipocytes by macrophage-conditioned medium: Upregulation of Il-6 production by macrophage-derived Il-1β. *Mol Cell Endocrinol* 2012; 349: 239-247.
- 8. Fraser DR. Regulation of the metabolism of vitamin D. *Physiol Rev* 1980; 60: 551-613.
- 9. Norman AW. From vitamin D to hormone D: Fundamentals of the vitamin D endocrine system essential for good health. *Am J Clin Nutr* 2008; 88: 491S-499S.
- 10. Plum LA, DeLuca HF. Vitamin D, disease and therapeutic opportunities. *Nat Rev Drug Discov* 2010; 9: 941-955.
- 11. Norman AW. Vitamin D receptor: New assignments for an already busy receptor. *Endocrinology* 2006; 147: 5542-5548.
- Hayes CE, Nashold FE, Spach KM, Pedersen LB. The immunological functions of the vitamin D endocrine system. *Cell Mol Biol (Noisy-le-Grand)* 2003; 49: 277-300.
- 13. Christakos S, DeLuca HF. Vitamin D: Is there a role in extraskeletal health? *Endocrinology* 2011; 152: 2930-2936.
- 14. Cohen-Lahav M, Douvdevani A, Chaimovitz C, Shany S. The anti-inflammatory activity of 1,25-dihydroxyvitamin

 $\mathrm{D_3}$ in macrophages. J Steroid Biochem Mol Biol 2007; 103: 558-562.

- 15. Jeffery LE *et al.* 1,25-dihydroxyvitamin D_3 and Il-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. *J Immunol* 2009; 183: 5458-5467.
- 16. Teegarden D, Donkin SS. Vitamin D: Emerging new roles in insulin sensitivity. *Nutr Res Rev* 2009; 22: 82-92.
- 17. Osei K. 25-OH vitamin D: Is it the universal panacea for metabolic syndrome and type 2 diabetes? *J Clin Endocrinol Metab* 2010; 95: 4220-4222.
- Arunabh S, Pollack S, Yeh J, Aloia JF. Body fat content and 25-hydroxyvitamin D levels in healthy women. *J Clin Endocrinol Metab* 2003; 88: 157-161.
- 19. Parikh SJ, Edelman M, Uwaifo GI, Freedman RJ, Semega-Janneh M, Reynolds J, *et al.* The relationship between obesity and serum 1,25-dihydroxy vitamin D concentrations in healthy adults. *J Clin Endocrinol Metab* 2004; 89: 1196-1199.
- Cheng S, Massaro JM, Fox CS, Larson MG, Keyes MJ, Mc-Cabe EL, *et al.* Adiposity, cardiometabolic risk, and vitamin D status: The Framingham heart study. *Diabetes* 2010; 59: 242-248.
- Li J, Byrne ME, Chang E, Jiang Y, Donkin SS, Buhman KK, et al. 1α,25-dihydroxyvitamin D hydroxylase in adipocytes. J Steroid Biochem Mol Biol 2008; 112: 122-126.
- 22. Ching S, Kashinkunti S, Niehaus MD, Zinser GM. Mammary adipocytes bioactivate 25-hydroxyvitamin D and signal via vitamin D receptor, modulating mammary epithelial cell growth. *J Cell Biochem* 2011; 112: 3393-3405.
- Kamei Y, Kawada T, Kazuki R, Ono T, Kato S, Sugimoto E. Vitamin D receptor gene expression is up-regulated by 1,25-dihydroxyvitamin D₃ in 3T3-L1 preadipocytes. *Biochem Biophys Res Commun* 1993; 193: 948-955.
- 24. Querfeld U, Hoffmann MM, Klaus G, Eifinger F, Ackerschott M, Michalk D, *et al.* Antagonistic effects of vitamin D and parathyroid hormone on lipoprotein lipase in cultured adipocytes. *J Am Soc Nephrol* 1999; 10: 2158-2164.
- Sun X, Zemel MB. 1α,25-dihydroxyvitamin D and corticosteroid regulate adipocyte nuclear vitamin D receptor. *Int J Obesity* 2008; 32: 1305-1311.
- 26. Wong KE, Kong J, Zhang W, Szeto FL, Ye H, Deb DK, *et al.* Targeted expression of human vitamin D receptor in adipocytes decreases energy expenditure and induces obesity in mice. *J Biol Chem* 2011; 286: 33804-33810.
- 27. Lee S, Lee DK, Choi E, Lee JW. Identification of a functional vitamin D response element in the murine Insig-2 promoter and its potential role in the differentiation of 3T3-L1 preadipocytes. *Mol Endocrinol* 2005; 19: 399-408.

36 Vitamin D-related gene expression

- Morris KL, Zemel MB. 1,25-dihydroxyvitamin D₃ modulation of adipocyte glucocorticoid function. *Obesity Res* 2005; 13: 670-677.
- 29. Sun X, Zemel MB. Calcium and 1,25-dihydroxyvitamin D_3 regulation of adipokine expression. *Obesity* 2007; 15: 340-348.
- Lorente-Cebrián S, Eriksson A, Dunlop T, Mejhert N, Dahlman I, Aström G, et *al*. Differential effects of 1α,25dihydroxycholecalciferol on MCP-1 and adiponectin production in human white adipocytes. *Eur J Nutr* 2011; in press. doi: 10.1007/s00394-011-0218-z.
- Blumberg JM, Tzameli I, Astapova I, Lam FS, Flier JS, Hollenberg AN, *et al.* Complex role of the vitamin D receptor and its ligand in adipogenesis in 3T3-L1 cells. *J Biol Chem* 2006; 281: 11205-11213.
- 32. Kong J, Li YC. Molecular mechanism of 1,25-dihydroxyvi-

tamin D₃ inhibition of adipogenesis in 3T3-L1 cells. *Am J Physiol Endocrinol Metab* 2006; 290: E916-924.

- Trayhurn P. Endocrine and signalling role of adipose tissue: New perspectives on fat. *Acta Physiol Scand* 2005; 184: 285-293.
- 34. Hotamisligil GS. Inflammation and metabolic disorders. *Nature* 2006; 444: 860-867.
- Rosen ED, Spiegelman BM. Adipocytes as regulators of energy balance and glucose homeostasis. *Nature* 2006; 444: 847-853.
- Trayhurn P, Wang B, Wood IS. Hypoxia in adipose tissue: A basis for the dysregulation of tissue function in obesity? *Br J Nutr* 2008; 100: 227-235.
- Trayhurn P, de Heredia FP, Wang B, de Olivera C, González-Muniesa P, Wood IS. Cellular hypoxia: A key modulator of adipocyte function in obesity? *Adipobiology* 2001; 1: 19-26.