



Adipobiology  
ISSN 1313-3705  
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RESEARCH  
ARTICLE

# EFFECT OF GRAPE POLYPHENOL CONCENTRATE “ENOANT” ON ADIPOKINES SECRETION IN EXPERIMENTAL METABOLIC SYNDROME

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## Abstract

Recent studies demonstrate that adipokine secretion is altered in cardiometabolic diseases including atherosclerosis, hypertension, obesity, type 2 diabetes and the metabolic syndrome (MS). Polyphenols are the most important phytochemicals in grape because they possess many biological activities and health-promoting effects. The aim of the present study was to investigate the effect of grape polyphenol concentrate “Enoant” on secretion of adipokines in MS in golden hamster (*Mesocricetus auratus*). We found that the development of experimental MS is accompanied by a significant increase in plasma levels of visfatin and resistin (pro-inflammatory adipokines), whereas the level of adiponectin (anti-inflammatory and anti-diabetic adipokine) was decreased. However, the introduction of grape polyphenol concentrate “Enoant” to animal diet in MS led to a decrease in resistin and visfatin plasma levels, whereas the adiponectin level was significantly increased. To test whether the observed changes may be related to the direct action of grape polyphenols on adipocyte secretion, we used primary cultures of adipocytes isolated from adipose tissue of control animals and from animals with MS. It was found that incubation of cells for 48 h in the presence of grape polyphenol concentrate is accompanied by a significant decrease in the visfatin concentration in the incubation medium, compared to control. In cultured adipocytes isolated from the adipose tissue of animals with experimental MS, polyphenol concentrate induced a significant increase in adiponectin and decrease in visfatin secretion. These findings suggest salutogenic action of grape polyphenol concentrate “Enoant” in experimental MS. Further studies are required to evaluate whether “Enoant” might be beneficial for cardiometabolic health via the secretion of other metabotropic adipokines (e.g., IL-10, NGF, BDNF).

Adipobiology 2012; 4: 85-90

**Key words:** adipose tissue, adiponectin, cultured adipocytes, grape polyphenols, visfatin, resistin

Received 5 August 2012, revised 29 August 2012, accepted 30 August 2012.

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## Introduction

The metabolic syndrome (MS) is a relatively new definition, designed to help the health care practitioner to easily identify people at risk for the development of cardiometabolic diseases. According to the National Cholesterol Education Program – Adult Treatment Panel III (NCEP-ATP III) (*JAMA* 2001; 285: 2486-2497), a person, to be defined as having the MS, must have any three of five characteristics: abdominal obesity, blood pressure higher than 130/80 mmHg, plasma triglycerides higher than 150 mg/dL, HDL-cholesterol lower than 40 mg/dL (men) or 50 mg/dL (women), and blood glucose higher than 100 mg/dL.

White adipose tissue is a dynamic endocrine and paracrine organ secreting multiple adipokines (reviewed in 1,2) involved in the pathogenesis of obesity and its related diseases (3-9). As described by these authors, adiponectin is a multifunctional, salutogenic adipokine, which circulatory and/or local levels are significantly decreased in cardiometabolic diseases. Visfatin is nicotinamide phosphoribosyltransferase that is the rate-limiting enzyme catalyzing the first step in the

biosynthesis of nicotinamide adenine dinucleotide from nicotinamide. This protein was originally cloned as a putative pre-B cell colony-enhancing factor and recently, in the “era of adipobiology”, found to be a visceral fat-secreted adipokine, hence visfatin. It is involved in the control of glucose and inflammation homeostasis. Resistin is an adipokine exerting negative effects on insulin action and on inflammation and thus implicated in the pathogenesis of obesity and diabetes.

A grape is a fruiting berry of the deciduous woody vines of the botanical genus described as *Vitis vinifera*. Polyphenols are the most important phytochemicals in grape because they possess many biological activities and health-promoting benefits (10–12). The phenolic compounds mainly include anthocyanins, flavanols, stilbenes (resveratrol) and phenolic acids (13–15). The use of grape seed polyphenol extract at low doses protects against fat accumulation and improves the plasma lipid profile in hamsters (16). Grape seed polyphenols suppressed high-fat-diet-induced obesity, hyperlipidemia and non-alcoholic fatty liver disease in mice (17). Grape seed polyphenol extract reduces the expression of IL-6 and MCP-1 and enhances the production of the anti-inflammatory adipokine adiponectin suggesting that may have a beneficial effect on low-grade inflammatory diseases such atherosclerosis, obesity, type 2 diabetes and MS (18). Chronic consumption of procyanidin extract from Chardonnay grape seed is shown to reduce obesity and related metabolic pathways including adipokine secretion and oxidative stress (19). Grape seed extract ameliorates the defective insulin and adiponectin signaling pathways in the skeletal muscle, resulting in improved insulin resistance in fructose-fed rats (20).

The dietary concentrate “Enoant” (hereafter to be written Enoant) contains high levels of grape polyphenols: flavonoid monomers and nonflavonoid polyphenols and their derivatives, including malvidin, cyanidin, delphinidin, peonidin, petunidin, quercetin, rutin, (+) catechin, (–) epicatechin, (–) epicatechin-gallate, gallic acid, syringic acid, caffeic acid, protocatechic acid, chlorogenic acid and trans-resveratrol. *In vivo* data from experiments in rats indicates stress-protective, hepatoprotective and antiatherogenic effects of Enoant. However, whether these effects could be mediated via Enoant action on the secretion of adipokines have been less studied.

In the present work we studied the effect of grape polyphenol concentrate Enoant on the secretion of the adipokines adiponectin, resistin and visfatin in golden hamsters under experimental MS.

## Material and Methods

4 month-old male Golden hamsters (approximately 90 g of body weight) were housed two per cage in a room with con-

trolled temperature (22–24°C), humidity and an inverse alternating light and dark cycle (12:12 hour light:dark cycle, lights on at 7 am and off at 7 pm). Animals were divided into: group 1 - controls (n=15) given standard diet and water; group 2 - animals with MS (n=15) kept for 4 weeks to a diet containing 29% vegetable oils and animal fats and fructose (2 g/100 g b.w./day); group 3 - MS + Enoant (n=15), animals with MS receiving Enoant (0,05 ml/100 g b.w., *per os*) contained approx. 20 g polyphenols per 1 L for the last 14 days; group 4 - Enoant group (n=17) received Enoant (0,05 ml/100 g b.w., *per os*) containing approximately 20 g polyphenols.

Enoant was developed in Institute of grape and wine Magarach (Yalta, Ukraine) and developed in Small Private Enterprise „Ressfud“ (Yalta, Ukraine).

Adipocytes were isolated as described previously (21). Visceral adipose tissue was removed always between 10.00 and 12.00 h to avoid chronobiological variations in adipokine profiles, and chopped with scissors into 2 µl Krebs–Ringer–HEPES (KRH) buffer (131.5 µM NaCl, 4.7 µM KCl, 2.5 µM CaCl<sub>2</sub>, 1.25 µM MgSO<sub>4</sub>, 2.5 µM NaH<sub>2</sub>PO<sub>4</sub>, 10.0 µM HEPES), supplemented with 1% BSA. Tissues were digested with collagenase type II (1 µmg/ml) for 1 h at 37 °C in a shaker. After 1 h of digestion, supernatant was removed and adipocytes were washed with fresh KRH buffer; his method of isolation effectively removed macrophages. The number of adipocytes was counted and diluted to 1×10<sup>6</sup> cells/ml with 10% fetal calf serum medium. Our experiments were performed using primary cultures of hamster adipocytes.

Cells were seeded at a concentration of 2×10<sup>6</sup> cells/well in a plate and incubated 48 hours at 37°C in the presence of “Enoant” aliquote, contained approx. 50 µM polyphenols.

Adipokines were measured in (i) blood plasma of fasting animals, and (ii) the incubation medium of cultured adipocytes. Adiponectin was measured using hamster adiponectin ELISA kit Species Hamster (Gentaur, Belgium); visfatin - mouse visfatin (VF) ELISA Kit, (Biocompare, USA); resistin - a kit from MyBioSource (LLC, USA), according to manufacturer’s protocol. Insulin level was measured using ELISA Kit (Biocompare, USA).

Lipids were extracted with chloroform and methanol (1:2 v/v) twice, as described by Bligh *et al* (22), and the supernatant was collected for determination of triacylglycerols (TG) and free fatty acids (FFA); these were determined by enzymatic colorimetric methods with commercial kits (Zhongsheng, Beijing, China). Plasma glucose was measured by a glucose oxidase method with commercial kits (AmplexR Red Glucose/Glucose Oxidase Assay Kit, Molecular Probes, USA).

All data were analyzed for statistical significance with SPSS

13.0 software. Data were presented as means  $\pm$  standard deviation. Statistical analysis used one-way ANOVA.  $P < 0.05$  was considered to be statistically significant.

## Results

We found that the development of experimental MS is accompanied by alterations in TG, FFA and glucose levels (Table 1). There was also an increase in insulin level as well as that of the adipokines visfatin and resistin, whereas the adiponectin content was significantly decreased (Table 2). Conversely, the introduction of grape polyphenol concentrate Enoant to the diet of animals with experimental MS led to a decrease insulin, resistin and visfatin levels in blood, whereas the adiponectin level was significantly increased (Table 2).

To test whether the observed changes may be related to the direct action of grape polyphenols on adipocyte secretion, we used primary cultures of adipocytes isolated from adipose tissue of control animals and from animals with MS. It was found that incubation of cells for 48 h in the presence of grape polyphenol concentrate was accompanied by a significant decrease in the visfatin concentration in the incubation medium, compared to control (Fig. 1). The concentrations of adiponectin and resistin in these conditions were not significantly changed.

Upon incubation of adipocytes isolated from adipose tissue of animals with experimental MS, there was a significant increase in the concentration of adiponectin and decreased visfatin in the cells incubation medium (Fig. 2). The resistin release in these conditions was not significantly changed.

## Discussion

At present, the MS is considered the major cardiometabolic risk factor, playing a leading role in the development of type 2 diabetes and atherosclerosis. It is known that MS is accompanied by (or, resulted from) low-grade inflammation and oxidative stress leading to changes in lipid and carbohydrate metabolism (25-28).

Effects of different plant polyphenols on metabolism in adipose tissue have been reported (23,24). The changes documented in the present study may be related to the direct action of grape polyphenols on the adipocytes and/or other adipose tissue cells. Plant polyphenols possess great antioxidant activity, which may also contribute to preventing the development of experimental MS (11,17,20). The increase in plasma FFA level may be one of the causes of the visfatin secretion impairment and subsequent insulin resistance (30). Our results are consistent with the literature data on the effects of plant polyphenols (quercetin

**Table 1.** The increase of glucose, triacylglycerols (TG) and free fatty acids (FFA) level in the blood of experimental animals with the metabolic syndrome (n = 6).

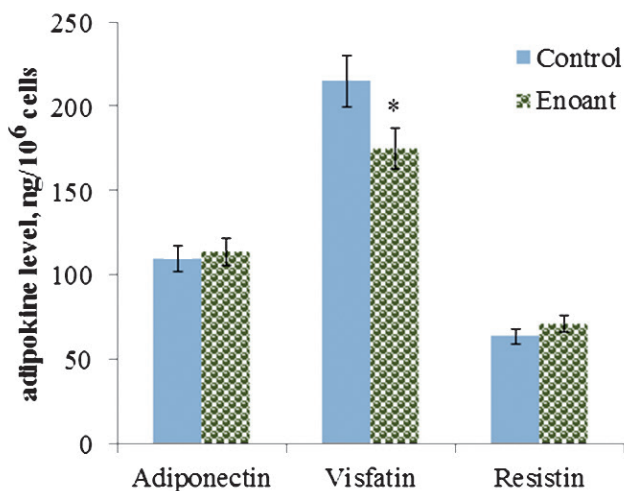
Group	FFA, mmol/l	TG, g/l	Glucose, mmol/ml
Control	0,85 $\pm$ 0,04	0.97 $\pm$ 0.03	4.65 $\pm$ 1.67
MS	1,20 $\pm$ 0,14*	2.14 $\pm$ 0.06*	12.7 $\pm$ 2.6*

The data presented as mean $\pm$ SD, \* - $p < 0.05$  versus control animals.

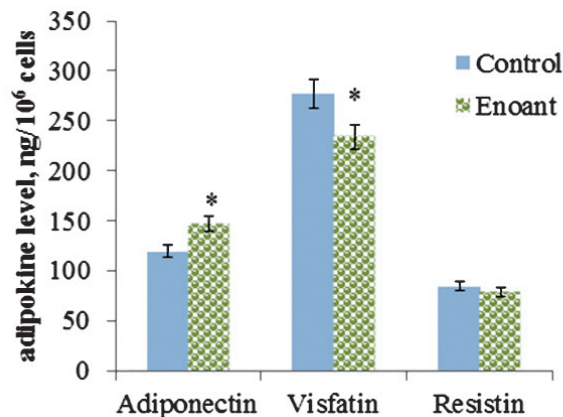
**Table 2.** Effect of grape polyphenol concentrate Enoant on the insulin, adiponectin, resistin and visfatin level in the blood plasma of gold hamsters with experimental metabolic syndrome (n = 6).

Molecules	MS	MS + Enoant	Control	Enoant
Insulin, ng/ml	1.46 $\pm$ 0.20*	0.94 $\pm$ 0.15**	0.85 $\pm$ 0.22	0.87 $\pm$ 0.13
Adiponectin, ng/ml	0.87 $\pm$ 0.09*	1.33 $\pm$ 0,49**	1.75 $\pm$ 0.38	1.85 $\pm$ 0.44
Visfatin, ng/ml	35,5 $\pm$ 5,4*	22,9 $\pm$ 4,2**	19,9 $\pm$ 1,4	19,7 $\pm$ 2,2
Resistin, ng/ml	7,38 $\pm$ 0,64*	5,01 $\pm$ 0,46**	4,01 $\pm$ 0,47	3,98 $\pm$ 0,67

The data presented as mean  $\pm$  SD; \* - $p < 0.05$  versus control animals, \*\* - $p < 0.05$  versus MS animals.



**Figure 1.** Effect of grape polyphenol concentrate Enoant treatment (48 hours) on adipokine secretion in cultured adipocytes isolated from control animals. Adipokine levels were measured in cell incubation medium. The data presented as mean  $\pm$  SD; \* – p0.05 versus control.



**Figure 2.** Effect of grape polyphenol concentrate Enoant treatment (48 hours) on adipokine secretion in cultured adipocytes isolated from animals with metabolic syndrome. Adipokine levels were measured in cell incubation medium. The data presented as mean  $\pm$  SD; \* – p 0.05 versus control.

and resveratrol) on the adipokine secretion in human Simpson Golabi Behmel Syndrome (SGBS) cells (31, also see the state-of-the-science review of Renes in this volume of *Adipobiology*). Grape polyphenols are involved in the regulation of signaling pathways which involve Akt/PKB system (32,33). We suppose that this may also be the case with the effects of Enoant's polyphenols.

In conclusion, the present study demonstrates that the grape polyphenol concentrate Enoant affects the secretion of adipokines both *in vivo* and *in vitro*. The administration of Enoant results in a normalizing the level of insulin, adiponectin, resistin, and visfatin in golden hamsters with experimental MS. Altogether, our results suggest a salutary action of grape polyphenol concentrate Enoant in MS. Further studies are required to evaluate whether Enoant (*i*) may also be beneficial for cardio-metabolic health in humans, (*ii*) may up-regulate the secretion of other (along adiponectin) metabotrophic adipokines, such as NGF, BDNF and IL-10 (34-36), and (*iii*) may like other natural polyphenols (e.g., resveratrol) mimic salutogenic effects of calorie restriction (37,38).

## References

1. Li FY, Lam KS, Xu A. Therapeutic perspectives for adiponectin: an update. *Curr Med Chem* 2012 Aug 9 (Epub ahead of print).

2. Chaldakov GN, Stankulov IS, Hristova MG, Ghenev PI. Adipobiology of disease: adipokines and adipokine-targeted pharmacology. *Curr Pharm Des* 2003; 9: 1023-1031.
3. Chaldakov GN, Fiore M, Stankulov IS, Manni L, Hristova MG, Antonelli A, *et al.* Neurotrophin presence in human coronary atherosclerosis and metabolic syndrome: a role for NGF and BDNF in cardiovascular disease? *Prog Brain Res* 2004; 146: 279-289.
4. Antuna-Puente B, Feve B, Fellahi S, Bastard JP. Adipokines: the missing link between insulin resistance and obesity. *Diab Metab* 2008; 34: 2-11.
5. Filippatos TD, Derdemezis CS, Kiortsis DN, Tselepis AD, Elisaf MS. Increased plasma levels of visfatin/pre-B cell colony-enhancing factor in obese and overweight patients with metabolic syndrome. *J Endocrinol Invest* 2007; 30: 323-326.
6. Filippatos TD, Randeva HS, Derdemezis CS, Elisaf MS, Mikhailidis DP. Visfatin/PBEF and atherosclerosis-related diseases. *Curr Vasc Pharmacol* 2010; 8: 12-28.
7. Catalan V, Gómez-Ambrosi J, Rodríguez A, Ramírez B, Silva C, Rotellar F, *et al.* Association of increased visfatin/PBEF/NAMPT circulating concentrations and gene expression levels in peripheral blood cells with lipid metabolism and fatty liver in human morbid obesity. *Nutr Metab Cardiovasc Dis* 2011; 21:245-253.

8. Wang P, Vanhoutte PM, Miao CY. Visfatin and cardio-cerebro-vascular disease. *J Cardiovasc Pharmacol* 2012; 59: 1-9.
9. Kusminski CM, McTernan PG, Kumar S. Role of resistin in obesity, insulin resistance and type II diabetes. *Clin Sci (Lond)* 2005; 109: 243–256.
10. Shrikhande AJ. Wine by-products with health benefits. *Food Res Int* 2000; 33: 469–474.
11. Silva RC, Rigaud J, Cheynier V, Chemina A. Procyanidin dimers and trimers from grape seeds. *Phytochemistry* 1991; 30: 1259–1264.
12. Wadaa M, Kidob H, Ohyamaa K, Ichibangasea T, Kishikawaa N, Ohbaa Y, et al. Chemiluminescent screening of quenching effects of natural colorants against reactive oxygen species: evaluation of grape seed, monascus, gardenia and red radish extracts as multi-functional food additives. *Food Chem* 2007; 101: 980–986.
13. Dopico-García MS, Figue A, Guerra L, Afonso JM, Pereira O, Valentão P, et al. Principal components of phenolics to characterize red VINO Verde grapes: anthocyanins or non-coloured compounds? *Talanta* 2008; 75: 1190–1202.
14. Novaka I, Janeiroa P, Serugab M, Oliveira-Brett AM. Ultrasound extracted flavonoids from four varieties of Portuguese red grape skins determined by reverse-phase high-performance liquid chromatography with electrochemical detection. *Anal Chim Acta* 2008; 630: 107–115.
15. Spacil Z, Novakova L, Solich P. Analysis of phenolic compounds by high performance liquid chromatography and ultra-performance liquid chromatography. *Talanta* 2008; 76: 189–199.
16. Baiges I, Palmfeldt J, Bladé C, Gregersen N, Arola L. Lipogenesis is decreased by grape seed proanthocyanidins according to liver proteomics of rats fed a high fat diet. *Mol Cell Proteomics* 2010; 9: 1499-513.
17. Kang JS, Lee WK, Yoon WK, Kim N, Park SK, Park HK, et al. A combination of grape extract, green tea extract and L-carnitine improves high-fat diet-induced obesity, hyperlipidemia and non-alcoholic fatty liver disease in mice. *Phytother Res* 2011; 25: 1789-1795.
18. Chacón MR, Ceperuelo-Mallafré V, Maymó-Masip E, Mateo-Sanz JM, Arola L, Guitierrez C, et al. Grape-seed procyanidins modulate inflammation on human differentiated adipocytes in vitro. *Cytokine* 2009; 47: 137-142.
19. Décordé K, Teissèdre PL, Sutra T, Ventura E, Cristol JP, Rouanet JM. Chardonnay grape seed procyanidin extract supplementation prevents high-fat diet-induced obesity in hamsters by improving adipokine imbalance and oxidative stress markers. *Mol Nutr Food Res* 2009; 53: 659-666.
20. Meepprom A, Sompong W, Suwannaphet W, Yibchok-anun S, Adisakwattana S. Grape seed extract supplementation prevents high-fructose diet-induced insulin resistance in rats by improving insulin and adiponectin signaling pathways. *Br J Nutr* 2011; 106: 1173-1181.
21. Vu V, Kim W, Fang X, Liu YT, Xu A, Sweeney G. Coculture with primary visceral rat adipocytes from control but not streptozotocin-induced diabetic animals increases glucose uptake in rat skeletal muscle cells: role of adiponectin. *Endocrinology* 2007; 148: 4411–4419.
22. Bligh EG, Dyer WJ. A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* 1959; 37: 911–917.
23. Harmon AW, Harp JB. Differential effects of flavonoids on 3T3-L1 adipogenesis and lipolysis. *Am J Physiol* 2001; 280: C807–C813.
24. Cho SY, Park PJ, Shin HJ, et al. (-)-Catechin suppresses expression of Kruppel-like factor 7 and increases expression and secretion of adiponectin protein in 3T3-L1 cells. *Am J Physiol* 2007; 292: E1166–E1172.
25. Haus JM, Solomon TP, Marchetti CM, Edmison JM, González F, Kirwan JP. Free fatty acid-induced hepatic insulin resistance is attenuated following lifestyle intervention in obese individuals with impaired glucose tolerance. *J Clin Endocrinol Metab* 2010; 95: 323-327.
26. Yuzefovych L, Wilson G, Rachek L. Different effects of oleate vs. palmitate on mitochondrial function, apoptosis, and insulin signaling in L6 skeletal muscle cells: role of oxidative stress. *Am J Physiol Endocrinol Metab* 2010; 299: E1096- E1105.
27. Haus JM, Solomon TP, Marchetti CM, Edmison JM, González F, Kirwan JP. Free fatty acid-induced hepatic insulin resistance is attenuated following lifestyle intervention in obese individuals with impaired glucose tolerance. *J Clin Endocrinol Metab* 2010; 95: 323-327.
28. Décordé K, Teissèdre PL, Sutra T, Ventura E, Cristol JP, Rouanet JM. Chardonnay grape seed procyanidin extract supplementation prevents high-fat diet-induced obesity in hamsters by improving adipokine imbalance and oxidative stress markers. *Mol Nutr Food Res* 2009; 53: 659-666.
29. Revollo JR, Körner A, Mills KF, Satoh A, Wang T, Garten A, Dasgupta B, Sasaki Y, Wolberger C, Townsend RR, Milbrandt J, Kiess W, Imai S. Nampt/PBEF/Visfatin regulates insulin secretion in beta cells as a systemic NAD biosynthetic enzyme. *Cell Metab* 2007; 6: 363-375.
30. Wen Y, Wang HW, Wu J, Lu HL, Hu XF, Cianflone K. Effects of fatty acid regulation on visfatin gene expression in adipocytes. *Chin Med J* 2006; 119: 1701-1708.
31. Derdemezis CS, Kiortsis DN, Tsimihodimos V, Petraki MP, Vezyraki P, Elisaf MS, et al. Effect of plant polyphenols on

- adipokine secretion from human SGBS adipocytes. *Biochem Res Int* 2011; 285618: 1-12.
32. Anselm E, Chataigneau M, Ndiaye M, Chataigneau T, Schini-Kerth VB. Grape juice causes endothelium-dependent relaxation via a redox-sensitive Src- and Akt-dependent activation of eNOS. *Cardiovasc Res* 2007; 73: 404-413
  33. Pan W, Ciociola E, Saraf M, Tumurbaatar B, Tuvdendorj D, Prasad S, *et al.* Metabolic consequences of ENPP1 overexpression in adipose tissue. *Am J Physiol Endocrinol Metab* 2011; 301: E901- E911.
  34. Chalidakov GN, Tonchev AB, Aloe L. NGF and BDNF: from nerves to adipose tissue, from neurokinines to metabokines. *Riv Psichiatr* 2009; 44:79-87.
  35. Sornelli F, Fiore M, Chalidakov GN, Aloe L. Adipose tissue-derived nerve growth factor and brain-derived neurotrophic factor: results from experimental stress and diabetes. *Gen Physiol Biophys* 2009;28:179-183.
  36. De Nicolo S, Ceccanti M, Fiore M. Effects of olive polyphenols administration on nerve growth factor and brain-derived neurotrophic factor in a brain mouse model [abstract]. *Adipobiology* 2012; 4: 90.
  37. Rosenow A, Noben JP, Jocken J, Kallendrusch S, Fischer-Posovszky P, Mariman EC, Renes J. Resveratrol-induced changes of the human adipocyte secretion profile. *J Proteome Res* 2012 Aug 20 (Epub ahead of print).
  38. Chung JH, Manganiello V, Dyck JR. Resveratrol as a calorie restriction mimetic: therapeutic implications. *Trends Cell Biol* 2012 Aug 9 (Epub ahead of print).