

DANCE ROUND

WE DANCE ROUND IN A RING AND SUPPOSE, BUT
THE SECRET SITS IN THE MIDDLE AND KNOWS.

ROBERT FROST

NEUROLIPOLOGY: INTERACTIONS OF NERVES, NEUROTROPHIC FACTORS, AND LIPIDS

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Here we highlight current data of the involvement of cholesterol, lipoproteins and lipoprotein receptors in the neuronal development and of neurotrophic factors in the lipid and glucose metabolism. We term these interactions neuro-lipidology. And argue that in addition to its implication in neurodegenerative diseases such as Alzheimer's disease (1), neurolipidology may have wide-ranging potential within a variety of nonneuronal fields, including cardiovascular disease, particularly, atherosclerosis and related disorders.

Emerging evidence shows that neurotrophic factors, such as nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin-3 (NT-3), leukemia inhibitory factor (LIF), and hepatocyte growth factor are involved in the process of atherogenesis (2, this *volume of Biomedical Reviews*). Because altered plasma concentrations of low density lipoprotein (LDL), very low density lipoprotein (VLDL), and high density lipoprotein (HDL) are among the principle players in the process of atherogenesis, they have been considered by many authors to act mainly on various cells of the artery wall. However, it appears to be much more than that. Indeed, it is known that LDL receptors (LDLR) can bind not only lipoproteins but also other ligands, and thereby involved in processes not solely related to lipoprotein internalization and metabolism. More recently, the neuronal development becomes one remarkable example of that. Likewise, in addition to their effects on the lipid and glucose metabolism of neuronal cells, as first demonstrated for NGF by Levi-Montalcini, neurotrophic factors may also

exert such effects on nonneuronal cells, including adipocytes, hepatocytes, and pancreatic beta cells.

CHOLESTEROL, LIPOPROTEINS AND LIPOPROTEIN RECEPTORS AND NEURONAL DEVELOPMENT

Although cholesterol has long been known to be an essential component of plasma membrane, recent studies have suggested that cholesterol plays an essential role during the development of nervous system (3). Likewise, accumulating evidence suggests that membrane cholesterol and sphingolipids cluster to form functional rafts that move within the fluid bilayer and influence receptor-mediated signal transduction and membrane trafficking (4,5).

Since first demonstrated by Brown and Goldstein, the importance of the LDLR in the regulation of cholesterol homeostasis has been studied intensively by virtue of its malfunction being causally related to atherosclerosis. Other members of the mammalian LDLR gene family are: LDLR-related protein (LRP), very low density lipoprotein receptor (VLDLR), apolipoprotein E receptor 2 (apoER2), and megalin (6 and Refs therein). Further, cubilin, the intrinsic factor-B 12 receptor, also exerts an extraintestinal function, facilitating endocytosis of HDL (7). Most of these receptors bind and import not only lipoproteins but multiple ligands, such as amyloid precursor protein and various proteases and their inhibitors. Specifically, the extracellular matrix protein *reelin* binds the ectodomain of

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both VLDLR and apoER2, and thereby is critically involved in the neurotopogenesis in the brain (6,8,9). Furthermore, cubilin, megalin, and p39 receptor-associated protein also play important roles in the biology of neurons. In addition, cholesterol can also affect neuronal development (3,10) and, curiously enough, statins, a group of HMG-CoA inhibitors that are widely used in atherosclerosis therapy, exhibit neuroprotective properties (11).

While numerous studies demonstrated that perivascular sympathetic nerves are associated with spontaneously hypertensive rats, their role in atherosclerotic lesion formation is largely ignored despite several studies suggest an inverse relation between the density of these nerves and the development of atherosclerosis (2). Recently, gene knockout and targeted overexpression have been used to generate models of disordered lipoprotein metabolism and atherosclerosis. The most extensively characterized of these models are apoE and LDLR knockout mice. ApoE-deficient mice (12) and cholesterol-fed rabbits/rats (13,14) also develop neuronal abnormalities. Principally, the neuronal abnormalities related to deficiency of lipoprotein receptors, as well as neuropathies related to dyslipidemia, could also be associated with low levels of neurotrophic factors, as shown for NGF in diabetic neuropathy (15). Thus, the question remains as to whether lipoproteins and/or their receptors might affect the neuronal development through alterations in the secretion and/or signaling of neurotrophic factors. For example, caveolin, a coating protein of endocytic, plasma membrane cholesterol-dependent, organelle called caveolae, regulates neurotrophin signaling (16), whereas caveolin (17) and LRP (18) expressions are upregulated by NGF. And BDNF regulates *reelin* expression in neuronal development (19). Nonetheless, genetically-modified models of disordered lipoprotein metabolism could be further examined, in order to see whether nerves, including perivascular sympathetic nerves, are affected. These considerations taken together suggest that in cardiovascular research, the targets for lipid/glucose metabolism-associated molecules should no longer be considered only vascular wall cells but also perivascular nerves.

NEUROTROPHIC FACTORS AND LIPID AND GLUCOSE METABOLISM

Recent years witnessed an increased knowledge of the roles played by neurotrophic factors in the biology of a large number of nonneuronal cells. The potential role of neurotrophic factors in the lipid and glucose metabolism of nonneuronal cells has just recently emerged. For example, NGF, which shares a striking structural homology with proinsulin (20), and LIF exert effects on the lipid metabolism in both adipocytes and hepatocytes (21,22). Similarly, LIF and ciliary neurotrophic factor stimulate hepatic triglyceride secretion (23), and LIF increases LDLR expression in liver cells and decreases serum cholesterol and

LDL levels (24), and thereby inhibits atherosclerosis development (25). It is also noteworthy that (i) BDNF (26) and mast cells (27), known to secrete multiple neurotrophic factors, are implicated in the glucose and lipid metabolism, and (ii) pancreatic beta cells secrete and respond to NGF (28). These findings suggest neurotrophin-mediated metabotropic functions, thus further implicate these molecules in cardiovascular disease and related disorders, such as metabolic syndrome.

As outlined above, there are certain interactions between NGF and caveolin (16,17). An additional comment about caveolin needs a special attention. First, both the structure and the function of caveolae are crucially dependent on plasma membrane cholesterol (4,5). Second, oxidation of cholesterol leads to migration of caveolin, from the plasma membrane to the Golgi complex, and hence inhibition of caveolae-mediated process called potocytosis (4,5). Third, folate is one of the principle molecules that is imported by caveolae (4,5). Fourth, high plasma levels of homocysteine are among the known risk factors for atherosclerosis (29); since homocysteine metabolism is closely linked to that of vitamin B12, one may keep in mind that an important function of cubilin (7) is the import of vitamin B12. All these possibilities could be further explored, using multiple transgenic animal models of neurotrophic factors/receptors, in order to see whether the lipid, glucose, and homocysteine metabolism are also affected.

„SHALL WE DANCE“: A CHALLENGE OF FIELD MIXING

The findings presented here suggest a complex arrangement between cholesterol, lipoproteins/receptors, nerves, and neurotrophic factors and the pathobiology of cardiovascular disease. Mixing fields is a challenge that promises high rewards, for both scientists and science alike. Metaphorically, as in the song "Shall we dance", from the American movie "Anna and the King", our hypothesis suggests an invitation for vascular biology and neurobiology to *dance round in a ring* of neurolipidology, in the study of atherosclerosis and related disorders.

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REFERENCES

1. Van Uden E, Carlson G, St George-Hyslop P, Westaway D, Orlando R, Mallory *Metal*. Aberrant presenilin-1 expression

- downregulates LDL receptor-related protein (LRP): is LRP central to Alzheimer's disease pathogenesis? *Mol Cell Neurosci* 1999; 14:129-140.
2. Chaldakov GN, Fiore M, Stankulov IS, Triaca V, Ghenev PI, Aloe L. Neuroimmune hypothesis of atherosclerosis. *Biomed Rev* 1999; 10: 37-44.
 3. Farese RV Jr, Herz J. Cholesterol metabolism and embryogenesis. *Trends Genet* 1998; 14:115-120.
 4. Simons K, Ikonen E. Functional rafts in cell membranes. *Nature* 1997; 387:569-572.
 5. Gimpl G, Burger K, Fahrenholz F. Cholesterol as modulator of receptor function. *Biochemistry* 1997; 36:10959-10974.
 6. Trommsdorff M, Gotthardt M, Hiesberger T, Shelton J, Stockinger W, Nimpf J *et al.* Reeler/Disabled-like disruption of neuronal migration in knockout mice lacking the VLDL receptor and ApoE receptor 2. *Cell* 1999; 97: 689-701.
 7. Kozyraki R, Fyfe J, Kristiansen M, Gerdes C, Jacobsen C, Cui S *et al.* The intrinsic factor-vitamine B₁₂ receptor, cubilin, is a high-affinity apolipoprotein A-I receptor facilitating endocytosis of high-density lipoprotein. *Nat Med* 1999; 5: 656-661.
 8. Cooper JA, Howell B W. Lipoprotein receptors: signaling functions in the brain? *Cell* 1999; 97: 671-674.
 9. D'Arcangelo G, Homayouni R, Keshvara L, Rice DS, Sheldon M, Curran T. Reelin is a ligand for lipoprotein receptors. *Neuron* 1999; 24:471-479.
 10. Chang JY, Liu LZ. Toxicity of cholesterol oxides on cultured neuroretinal cells. *Curr Eye Res* 1998; 17: 95-103.
 11. Vaughan CJ, Delanty N. Neuroprotective properties of statins in cerebral ischemia and stroke. *Stroke* 1999; 30:1969-1973.
 12. Fullerton SM, Strittmatter WJ, Mathew WD. Peripheral sensory nerve defects in apolipoprotein E knockout mice. *Exp Neurol* 1998; 153: 156-163.
 13. Verbenren TJ, Simonet S, Herman AG. Diet-induced atherosclerosis inhibits release of noradrenaline from sympathetic nerves in rabbit arteries. *Eur J Pharmacol* 1994; 270:27-34.
 14. Karoon P, Bumstock G. Reduced sympathetic noradrenergic neurotransmission in the tail artery of Donryu rats fed with high cholesterol-supplemented diet. *Br J Pharmacol* 1998; 123:1016-1021.
 15. Apfel SC. Neurotrophic factors in the therapy. *Am J Med* 1999; 107(2B):34S-42S.
 16. Bilderback TR, Gazula V-R, Lisanti MP, Dobrowsky RT. Caveolin interacts with Trk A and p75^{NTR} and regulates neurotrophin signaling pathways. *J Biol Chem* 1999; 274: 257-263.
 17. Galbiati F, Volonte D, Gil O, Zanazzi G, Salzer JL, Sargiacomo M *et al.* Expression of cavin-1 and -2 in differentiating PC12 cells and dorsal root ganglion neurons: caveolin-2 is up-regulated in response to cell injury. *Proc Natl Acad Sci USA* 1998; 95:10257-10262.
 18. Bu G, Sun Y, Schwartz AL, Holtzman DM. Nerve growth factor induces rapid increases in functional cell surface low density lipoprotein receptor-related protein. *J Biol Chem* 1998; 273:13359-13365.
 19. Ringstedt T, Linnarsson S, Wagner J, Lendahl U, Kokaia Z, Arenas E *et al.* BDNF regulates reelin expression and Cajal-Retzius cell development in the cerebral cortex. *Neuron* 1998; 21:305-315.
 20. Mukherjee SP, Mukherjee C. Similar activities of nerve growth factor and its homologue proinsulin in intracellular hydrogen peroxide production and metabolism in adipocytes. Transmembrane signalling relative to insulin-mimicking effects. *Biochem Pharmacol* 1982; 31: 3163-3172.
 21. Ng TB, Wong CM. Epidermal and nerve growth factors manifest antilipolytic and lipogenic activities in isolated rat adipocytes. *Comp Biochem Physiol (B)* 1985; 81: 687-689.
 22. Nonogaki K, Moser AH, Shigenaga J, Feingold KR, Grunfeld C. Beta-nerve growth factor as a mediator of the acute phase response in vivo. *Biochem Biophys Res Commun* 1996; 219: 956-961.
 23. Nonogaki K, Pan XM, Moser AH, Shigenaga J, Staprans I, Sakamoto N *et al.* LIF and CNTF, which share the gp130 transduction system, stimulate hepatic lipid metabolism in rats. *Am J Physiol* 1996; 271 (3 Pt 1): E521-E528.
 24. Moran CS, Campbell JH, Campbell GR. Human leukemia inhibitory factor upregulates LDL receptors on liver cells and decreases serum cholesterol in the cholesterol-fed rabbit. *Arterioscler Thromb Vasc Biol* 1997; 17:1267-1273.
 25. Moran CS, Campbell JH, Simmons DL, Campbell GR. Human leukemia inhibitory factor inhibits development of experimental atherosclerosis. *Arterioscler Thromb* 1994; 14: 1356-1363.
 26. Ono M, Ichihara J, Nonomura T, Itakura Y, Taiji M, Nakayama C *et al.* Brain-derived neurotrophic factor reduces blood glucose level in obese diabetic mice but not in normal mice. *Biochem Biophys Res Commun* 1997; 238:633-637.
 27. Nunes MT, Curi R. Hyperthyroid-like metabolic changes induced by chronic administration of compound BW 487 80. *Braz J Med Biol Res* 1989; 22:417-420.
 28. Rosenbaum T, Vidaltamayo R, Sanchez-Soto MC, Zentella A, Hiriart M. Pancreatic p1 cells synthesize and secrete nerve growth factor. *Proc Natl Acad Sci USA* 1998; 95:7784-7788.
 29. Ross R. Mechanisms of disease: atherosclerosis - an inflammatory disease. *N Engl J Med* 1999; 340: 115-126.