# THE ENDOCANNABINOID-CB RECEPTOR SYSTEM: A NEW PLAYER IN THE BRAIN-GUT-ADIPOSE FIELD

#### Ester Fride

College of Judea and Samaria, Departments of Behavioral Sciences and Molecular Biology, Ariel, Israel

The 1990's have witnessed the discovery of the 'endocannabinoid-cannabinoid (CB) receptor (ECBR) system' consisting of specific receptors (CB<sub>1</sub> and CB<sub>2</sub>), at least 3 endogenous ligands (anandamide, 2-arachidonyl glycerol and noladin ether) and their enzymes. Subsequently, a series of discoveries were made on the involvement of the ECBR system in many physiological functions including immunity, inflammation, neurotoxicity and neurotrauma, epilepsy, depression and stress, appetite, food intake and energy homeostasis, cardiovascular regulation, reproduction, and bone remodeling. The brain-gut-adipose axis regulates digestive processes, food ingestion and energy balance and is closely associated with hormonal regulation by the hypothalamicpituitary-adrenal stress axis and by the mesolimbic reward system. It is proposed to call these interfacing systems the "alimentary control system". ECBR presence in brain, gastrointestinal as well as adipose tissue explains its role in food intake, digestion and the regulation of adipose tissue mass. Moreover, the ECBR system's involvement in stress and emotional processing, makes it eminently suited to be (one of) the principle players in the alimentary control system. The ECBR system is present during the early embryonal and postnatal stages and we have discovered endocannabinoids in maternal milk. The ECBR system seems to be of critical value for newborn milk ingestion and suckling. Ghrelin, an orexigenic gastric hormone, exerts many effects similar to those of endocannabinoids. Therefore this review will compare some of the functions and anatomy of ghrelin and endocannabinoids. It is concluded that (i) the ECBR system is a major mediator between the brain and the alimentary system, and possibly, the adipose tissue, (ii) the role of the ECBR system in adult regulation of food processing is a remnant of its critical role for the initiation of feeding in the newborn, and (iii) the pervasive influence of the ECBR system in alimentary control make it a highly suitable target for therapeutic developments for pathophysiological conditions such as inflammatory bowel disease, irritable bowel syndrome, gastric ulcers, nausea, anorexia and failure-to-thrive. Biomed Rev 2006; 17: 23-42.

Key words: adipose tissue, appetite, CB1, CB2, development, feeding, gastrointestinal tract

#### INTRODUCTION: THE "ALIMENTARY CONTROL SYSTEM"

Appetite, hunger and satiety are regulated by a number of hormonal signals emanating from the central nervous system (CNS), and upper gastrointestinal (GI) tract (GIT) and adipose tissue (1,2). Thus it is impossible to separate the appetite-hunger-satiety signals which regulate food intake, from the digestive processes in the GIT and the adipose tissue secretions such as leptin. Therefore in this review, regulation of food intake, appetite and the brain-gut-adipose system will be considered as one system regulating *ingestion* and *digestion*, and will be denoted the *"alimentary control system"*.

The rich bidirectional interactions between the brain and the local neuronal network (the enteric nervous system -ENS) in the intestinal system (the 'brain-gut axis'), have been extensively studied. Thus, for example, psychological stress as well as transcranial stimulation of the cortex in humans, have been shown to affect gut activities such as secretion and motility (3). Conversely, using brain imaging techniques such as functional magnetic resonance imaging (fMRI) or positron emission tomography (PET), have shown that visceral sensation resulting from stimulation of the oesophagus, stomach or rectum, resulted in activation of higher brain centers including the somatosensory cortex, anterior cingulate cortex, thalamus, insula and prefrontal cortex (1,3-5). Moreover patients suffering from irritable bowel syndrome (IBS), displayed different response patterns (4). Relatively few animal studies have investigated the brain-gut associations in an integrative (biobehavioral) way. The data that have been collected, however, are very intriguing. For example, traumatic brain injury caused intestinal dysfunction and affected concentrations of several brain/intestinal peptides such as vasoactive intestinal peptide (VIP) and cholecystokinin (CCK) in blood and intestinal tissue (jejunum) (6). Further, secretin, a secretory gut hormone, which has also been suggested to ameliorate autistic symptoms, activated a number of brain areas as measured by *c-fos* gene expression in rat brain. These brain structures included not only the dorsal vagal complex structures which are part of the brain-gut axis, but also the prefrontal cortex, which is thought to play a role in autism (7).

The brain-gut axis has been the subject of a number of recent reviews (3,5,8,9). Adipose tissue is now recognized as an endo- and paracrine secretory organ regulating a number of functions beyond lipid and energy homeostasis (reviewed in 10,11). Recently, the ECBR system has been shown to be fully represented and functional in adipocytes (12,13). In this

review, we will examine the role of the relatively recently discovered endocannabinoids and their cannabinoid CB receptors (the 'ECBR system') (14,15) in alimentary control and adipocyte-regulated energy balance *via* the brain-gut-adipose system. Thus first, the major aspects of the alimentary control system will be briefly outlined; next, the ECBR system will be reviewed, after which its multiple role in alimentary control will be discussed. Finally, a brief discussion of developmental aspects of the ECBR system in alimentary control will be offered followed by a comparison with the hormone ghrelin.

# THE BRAIN-GUT-ADIPOSE AXIS AND THE ALIMENTARY CONTROL SYSTEM

# Ingestion

Ingestion is controlled by a large number of interacting hormones originating from the brain, GIT and adipose tissue.

## Hormones from adipose tissue

Leptin is a major adipocyte-derived hormone, its plasma concentrations being proportionate to body fat content and it enter the brain according to its plasma levels. Inside the arcuate nucleus of the hypothalamus, it regulates a number of peptides involved in energy control and food intake (11,16,17) (see below). Recently more than 100 adipose-secreted signaling proteins, collectively named adipokines, have been isolated and reviewed; many of these are pleiotropic (10,11). The present review will concern itself only with those which are pimarily associated with energy balance, including leptin and adiponectin. Additional adipokines, like nerve growth factor (NGF), which have many additional functions and locations, will be discussed in the appropriate sections.

#### Hormones from the GIT

Cholecystokinins (CCK-8, CCK-33, CCK-39) activate CCK<sub>1</sub> receptors on vagal afferents to the nucleus of the solitary tract (NTS). Cholecystokinins also enters the bloodstream, reducing appetite directly in the hypothalamus (see below) (2,14,18,19). It is interesting to note that CCK has the ability to stimulate the synthesis of nerve growth factor (NGF), an adopokine (10) which is also found in additional peripheral tissue and in the brain, thus enhancing NGF-induced anti-inflammatory activity (20).

**Peptide tyrosine tyrosine (PYY)** activates vagal Y2 receptors, but also penetrates the brain through the median

eminence, reaching the hypothalamus, where it activates hypothalamic Y2 receptors. Plasma PYY levels rise almost immediately after the onset of a meal (2,21), thus suggesting a tight control mechanism on meal size.

Ghrelin is produced primarily by the stomach, but minor amounts are also made in the small intestine and hypothalamus (22). Endogenous ghrelin levels increase before a meal and fall almost immediately upon food intake, while exogenous ghrelin enhances food intake (2,21). Conversely, a ghrelin antagonist reduced food intake and body weight in mice (23). In addition to appetite and weight control, ghrelin enhances intestinal motility, acid secretion and conveys mucosal protection (2). It also fulfills other physiological roles such as cardiovascular and sleep regulation (22). In view of its rapid response to the commencement of food intake, ghrelin is thought to play a major role in the *initiation* of food intake (2) (see Table 2). Exogenous ghrelin almost tripled food intake and decreased plasma leptin by 30%. The suppression is mutual ('ghrelin-leptin tango') (2,24).

**Orexin A and B** are found in the gut and enhance food intake by inhibiting vagal afferent discharge, thus negatively interacting with the anorexic effects of CCK (2)

#### Hormones in the brain

Many of the GI hormones including those mentioned in the previous section are also synthesized in the CNS, where they directly influence appetite and also have additional functions (18,22,25,26). However, several hormones are particularly involved in the central neural network responsible for the regulation of food intake and these are briefly described below.

The adipocyte-secreted hormone leptin, entering the brain from the blood circulation, is a major regulator of a number of peptides located in the arcuate nucleus of the hypothalamus. There, leptin inhibits orexigenic peptides (neuropeptide tyrosine, NPY, and *agouti*-related potein, AgRP) and simultaneously, activates the anorexigenic precursor molecule proopiomelanocortin (POMC) (27). An increase in POMC in turn, produces enhanced levels of its cleavage product,  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH). Cocaine-and-amphetamine-regulated transcript (CART) which is colocalized with POMC in arcuate nucleus neurons, is also stimulated by leptin, eventually suppressing food intake (16,17,28). CCK, a satiety-inducing peptide previously found in the gut, directly

signals satiety through its receptors in several feeding centers of the hypothalamus (arcuate nucleus, lateral hypothalamus and paraventricular nucleus (PVN) (29,30).

Additional CNS molecules which have been implicated in alimentary control include the neurotransmitters serotonin (5), GABA (31), and the opioid neuropeptides (32). However, an exhaustive discussion of such molecules is beyond the scope of this review.

#### Digestion

The gastrointestinal tract has an independent nerve center (the enteric nervous system, ENS, or "gut brain") which maintains bilateral neural communication with the CNS along the nerves of the sympathetic and parasympathetic systems (2,5). Under normal conditions CNS control is minor. However, when homeostasis is perturbed such as in digestive disorders or during psychological stress, CNS control plays a major role (5).

Gastrointestinal tract activity (mainly secretory and motor functions) is regulated via a 3-tier loop system (see ref. 2,5): (i) short (local ENS) loop, relying on independent ENS activity, (ii) spinal cord-reflexes, via the autonomic (sympathetic and parasympathetic systems), and (iii) via the brain: a number of brain regions play a role and/or are influenced by the GIT: the first target region is the brainstem, where vagal afferents target the nucleus of the solitary tract (NTS), adjacent to the dorsal motor nucleus of the vagus (DMV). Via interneurons connecting the NTS with the DMV, a long vago-vagal neural reflex loop is established. In addition, part of the intestinal neural input to the NTS is transmitted further to the hypothalamus (the paraventricular nucleus [PVN] and the arcuate nucleus), and further to the amygdala, the bed nucleus of the stria terminalis (BNST) and ventral thalamus (2). Cortical areas, including the anterior cingulate cortex, somatosendory cortex, and prefrontal cortex, have been shown to represent GI sensations (1,3,5). Notably, several if these structures (amygdala, hypothalamus, cingulate and prefrontal cortices) are part of the limbic system for emotional processing (28).

# EMOTIONAL STRESS AND THE ALIMENTARY CONTROL SYSTEM

As stated above, gastrointestinal functions are regulated by both the local ENS as well as by the CNS, the latter becoming more dominant when homeostasis is perturbed (5). It is well

known that emotional processes including stress and depression influence food intake (33,34) as well as GI functioning, by for example, inhibiting gastric emptying and stimulating GI motility (35,36). Interestingly, it has now been shown with brain imaging methods, that the limbic (emotional) system structures, the anterior cingulate cortex and prefrontal cortex, are activated during visceral pain, while patients with irritable bowel disease display a different pattern of activation (4,37).

Corticotrophic releasing hormone (CRH), which is the hypothalamic hormone first in line in the series of hormones released by the hypothalamic-pituitary-adrenal (HPA) axis upon stress exposure, is an important mediator of stress-induced regulation of GI functioning (35). Thus the HPA and brain-gut-adipose axes are closely interrelated; it even has been suggested that the HPA axis is the hormonal arm of the brain-gut system (34). Indeed, traditionally, and still in various regions of the globe, food deprivation is a primary threat to survival and logically, food searching and the motivation for food ingestion, should be driven by hunger- and food deprivation-induced stress. Such integrative approach to feeding mechanisms (34) is supported by the neuranatomical and functional associations and overlap between the brain areas regulating stress responsivity and the alimentary system. Thus the limbic (emotional) system, including the hypothalamus, amygdala, anterior cingulate cortex and the prefrontal cortex, also contains the focal CNS structures which direct GIT activities such as GI motility and gastric emptying (1). Functionally, CRH induces an anxiogenic response, but also participates in the stress-induced alterations in upper GIT and gastric motility (35). Conversely, the gastrin-releasing peptide (GRP) and its receptors, appear to be necessary for the memory of traumatic memories through their location in the amygdala (38). Additionally, a number of molecules including unicortin, CCK, ghrelin and the melanocortin product of POMC, which all regulate appetite and/or GI function, also influence emotional processes (22,39,40).

#### **REWARD AND THE ALIMENTARY CONTROL SYSTEM**

As noted above, increased levels of the anorexigenic precursor molecule POMC (27) results in enhanced production of  $\alpha$ -MSH thus contributing to the suppression of food intake

(16,17,28). Additional products of POMC are the opioids, which are intimately involved in the regulation of reward in the dopaminergic mesolimbic system (a neural pathway projecting from the ventral tegmental area in the midbrain to the forebrain nucleus accumbens (28,34)). The dopamine system, in turn, has been associated with food-related behaviors in humans (41). In rats, POMC neurons were inhibited after the nucleus accumbens shell was manipulated to elicit feeding behavior in rats (42). Thus, POMC's involvement in both food intake (43) and reward may be part of the rewarding qualities of food ingestion. An additional important observation was made in a self-stimulation paradigm, where the stimulating electrode was placed in the lateral hypothalamus of food-restricted rats. Thus whereas, replicating previous data (44), food restriction enhanced self stimulation, administering leptin to such rats, decreased self stimulation. Together with the multitude of orexigenic and anorexigenic peptides which activate and/or reside in the nuclei of the lateral hypothalamus (including NPY, α-MSH, AgRP, CART, orexin, MCH) these observations strongly suggest that the reward/incentive value of food plays a major role in feeding and body weight regulation and has far reaching implications for addictive patterns of food intake (45). As outlined above, higher brain centers including the prefrontal cortex (PFC), have also been implicated in the neural circuits comprising the brain-gut axis, while the PFC also comprises an important component of the reward/addiction system (46,47). This anatomical overlap between alimentary control and incentive/reward processing further supports common ground for feeding behavior, metabolism, and emotional processing of reward and addiction. Thus all of these processes need to be taken into account when designing strategies to combat abnormal feeding behaviors such as seen in obesity and anorexia nervosa.

#### THE ENDOCANNABINOID-CB RECEPTOR SYSTEM

#### Cannabinoid CB receptors

Thus far two CB receptors,  $CB_1$  and  $CB_2$ , have been identified and cloned (48-50). In addition, 2 splice variants to the "classical"  $CB_1$  receptor have been identified (51-53). Widely different lines of evidence suggest the existence of an additional (or several additional) CB (" $CB_x$ ", or " $CB_3$ ") receptors (54-56). The  $CB_1$  and  $CB_2$  receptors both belong to the superfamily of G-protein-coupled receptors (57). The CNS

concentrations of the CB<sub>1</sub> receptor in general, reach levels similar to those of GABA- and glutamate-gated ion channel receptors (57). At the single cell resolution, high CB<sub>1</sub> receptor mRNA expressing neurons in mouse forebrain were found to be mostly GABAergic, while most low CB<sub>1</sub> expressing cells were not (58). In the same study, very high co-expression with the neuromodulator CCK was found in limbic as well as non-limbic cortical neurons. Since brain CCK and CBs act usually in opposite directions (59), the ECBR and CCK systems may conduct cross talk in their regulation of a number of physiological or pathophysiological functions including learning and memory (60), food intake (2) and schizophrenia (58,61). CB<sub>1</sub> receptor mRNA is generally found at about 10fold lower levels in peripheral tissues than in the CNS (53). The CB2 receptor is generally found in non-neural, mostly immune tissue (57,62,63) but also in the GIT (53,64,65) (also see below). Recently, CB<sub>2</sub> receptors have been discovered in neural cells throughout the brain. Perhaps the reason for their belated detection on neurons of the CNS, is their appearance in relatively low concentrations (66,67).

### Endocannabinoids and their enzymes

Several endocannabinoids (ECs) have been discovered thus

far, all derivatives of arachidonic acid (14). The apparently major and most thoroughly studied ECs are anandamide (AEA) (68) and 2-arachidonyl glycerol (2-AG) (69). Additional ECs include noladin ether (64), N-arachidonoylglycerol dopamine (NADA) (70,71); virodhamine is thought to be a partial agonist/antagonist on CB<sub>1</sub> receptors (72). Noladin ether, NADA and virodhamine have not been systematically studied for their presence in the alimentary control system except for our observation that noladin ether inhibits intestinal motility in mice (64).

Up-to-date information on the synthesis, release, reuptake and degradation mechanisms for the ECs has been reviewed (73). In short, ECs are synthesized "on demand". This concept is strongly supported by a positive relationship between EC production and intracellular calcium concentrations. Anandamide is synthesized mainly from N-arachidonylphosphatidyl-ethanolamine using the enzyme phospholipase-D. 2-AG is enzymatically formed with diacylglycerol lipases  $\alpha$  and  $\beta$ . The degradation of AEA is catalyzed by fatty acid amide hydrolase (FAAH). On the other hand, 2-AG can be broken down by several pathways: primarily by monoacylglycerol lipase (MAGL) but also by FAAH, by an esterification pathway

Table 1. Levels of alimentary control by the ECBR system

Location	Effect	Selected references
CNS-forebrain including hypothalamus	Rewarding/Stress effect on food intake and digestion; Orexigenic effect	74,76-78
CNS-hindbrain	Gastric secretion and Intestinal motility/secretion reduced; Antiemetic effect	99,101
Mouth	Salivation decreased	91
Lower esophageal sphincter	Relaxation	85,92
Stomach	Decreased gastric secretion	93
Intestinal tract	Inhibit motility Inhibit of secretion	100-102 104

and by Cox-2 together with PGE synthase, transforming it to PGE2-glyceryl ester. Uptake and release of ECs is probably regulated by a common transport system, although these mechanisms remain to be fully elucidated (73).

# THE ECBR SYSTEM AND THE ALIMENTARY CONTROL SYSTEM

The multifaceted involvement of the ECBR system in alimentary control will be discussed in the present section. Thus it appears that the ECBR system plays a role at every level of alimentary control including reward/addiction and stress. Therefore, the remainder of this article will be devoted to ECBR involvement in alimentary control in health and disease and in the neonatal organism. Finally, the extent of the ECBR involvement in *in*-and *digestion* will be compared to functions of the ghrelin peptide, another recently discovered and apparently multileveled player in alimentary control and other physiological systems.

# Cannabinoid CB<sub>1</sub> receptors in alimentary control

#### Forebrain, reward and stress

Components of the ECBR system are present in most structures throughout the brain (14,57), including the nucleus accumbens, the target area of the dopaminergic mesolimbic "reward" system (74). Importantly, CB<sub>1</sub> receptors are colocalized with dopamine receptors in the mouse forebrain (75). Not surprisingly then, 2-AG directly administered to the nucleus accumbens of the mesolimbic system, induced profound hyperphagia (76). These findings strongly suggest a role for the ECBR system in regulating food intake through the incentive value of food.

A role for the ECBR system in the ability to cope with stress has been studied using behavioral and biochemical (77-79) techniques. Further, endocrinologically, ECs influence the HPA stress axis at the pituitary and hypothalamic levels (77,79-81). As outlined above, the HPA stress axis strongly influences both feeding and digestion. Therefore, the ECBR system may be expected to exert part of its influence on alimentary control through the stress-regulating HPA axis.

The forebrain is also the interactive site for the ECBR system and other alimentary control hormones. Thus the orexigenic effect of local infusion of ghrelin into the hypothalamic PVN of rats, was abolished by a peripheral injection with rimonabant (82). These data suggest that hypothalamic ghrelin and CB<sub>1</sub> receptors interact as part of a finely tuned system aimed at maintaining balanced food intake. As mentioned above, the gut and brain peptide CCK induces a feeling of satiety mainly after its release in the gut by protein and fat digests (2). However, CCK is also highly coexpressed with CB<sub>1</sub> receptors in brain structures including the cerebral cortex, hippocampus (58) and the amygdala (83). As noted above, the limbic brain plays a role in emotional aspects of food intake and appetite. It has also been noted that CCK and the ECBR system often signal in opposite directions (59). Therefore, it is plausible to expect that the ECBR and CCK systems interact with respect to food intake, appetite and satiety through emotional regulation in the limbic system. Coexpression of CCK (29,30) and CB<sub>1</sub> receptors in feeding-related nuclei of the hypothalamus has not specifically been studied. Therefore, cross-talk between the CCK and ECBR systems at the hypothalamic level should also be examined.

#### Hindbrain

Evidence has accumulated that CB<sub>1</sub> receptors are located in the dorsal vagal complex in the brainstem, comprising the area postrema, nucleus of the solitary tract (NTS) and dorsal motor nucleus of the vagus (DMV). From these locations it is thought now that the CBs mediate their antiemetic effects and relaxation of the lower oesophagal sphincter (LOS) (84-86) and inhibition of gastric motility (87) and gastric acid secretion (84).

It was also suggested that CBs may enhance appetite by suppression of inhibitory synapses in the DMV (88). In support of this assertion, CB<sub>1</sub> receptors are coexpressed with CRH and CART in a significant proportion of neurons of the PVN and/or DMV/lateral hypothalamus of mice (89). Likewise, neuronal CB<sub>2</sub> receptors in the CNS have now been demonstrated in number of structures including cerebellum, cerebral cortex, hippocampus, brain stem and more (66,67). Their functional presence with respect to the brainstem, notably in the DMV, was investigated functionally. Thus evidence was found that CB<sub>2</sub> receptors participate together with CB<sub>1</sub>, in the suppression of the emetic response (66). Such cooperation with CB<sub>1</sub> receptors would explain how much lower levels of CB<sub>2</sub> receptors (1.5% of those present in spleen ref. 66,67), can be of

functional importance.

#### Vagus

CB<sub>1</sub> receptors are present in the peripheral vagal afferent nerve endings in the nodose ganglion to influence visceral perception (84,85). Also at this location CB<sub>1</sub> receptors are co-expressed with CCK receptors, thus suggesting that, in addition to central interactions, the ECBR system interacts with the CCK system to regulate food intake and satiety in the periphery (90).

#### Mouth

Very recently, functional CB<sub>1</sub> and CB<sub>2</sub> receptors have been identified in rat saliva. Thus activation of these CB receptors reduced saliva secretion (91). Therefore it is possible that CB receptors play a role in digestion from the first stage of the process.

# Lower oesophageal sphincter

 $CB_1$  receptors are located in the lower oesophageal sphincter (LOS) muscle (84). Cannabinoid ligands ( $\Delta 9$ -THC, WIN 55,212-2) inhibited its relaxation in ferrets and dogs, thereby counteracting gastroesophageal acid reflux (85,92). These studies also suggested however, that this effect is mediated by  $CB_1$  receptors in the dorsal vagal complex of the hindbrain.

### Stomach

CB<sub>1</sub> receptors are also present in gastric muscle and mucosa where they are localized in pre- and postganglionic cholinergic neurons (93). Izzo's group (94), investigating CB<sub>1</sub> receptor concentrations throughout the GIT, confirmed that concentrations are highest in the stomach and in the colon. CB<sub>2</sub> receptor mRNA has also been demonstrated in rat gastric tissue (95).

#### Intestines

CB<sub>1</sub> receptors are widely present in the GIT (96). The colon (together with the stomach as mentioned above) has the highest CB<sub>1</sub> receptor density in the GIT (94). CB<sub>1</sub> receptors are widely present along the GIT as studied in rats, mice and guinea pigs (see (97)). Thus CB<sub>1</sub> receptors were found on cell bodies and fibers in myenteric plexus of cholinergic sensory, interneuronal and motor neurons. In the submucosal plexus CB<sub>1</sub> receptors are colocalized with VIP and NPY (98). These findings are consistent with the influential role of the CBs on both secretory activity and motility of the gut (97). CB<sub>1</sub> receptors have also been shown to mediate the inhibition gastric emptying (99) and

intestinal transit/motility (100-102). Moreover, from the data collected thus far it seems that to a great extent intestinal CB1 receptors located on extrinsic neurons, by inhibiting the release of acetylcholine (Ach), are responsible for CB-directed regulation of secretory patterns and motility under normal conditions (103). However, it has been suggested that in pathophysiological states, CB<sub>1</sub> receptors located on intrinsic submucosal neurons are mobilized (84). Thus in an in vivo study, Izzo and colleagues demonstrated that cholera toxin-induced intestinal fluid accumulation, resulting in diarhoea, was inhibited by the CB<sub>1</sub> receptor agonist CP 55,940. This effect was not prevented by ganglionic blockade with chlorisondamine (104). The importance of CB<sub>1</sub> receptors in protecting the organism against inflammation of the GI system was demonstrated in a study where CB<sub>1</sub> receptor knockout mice were found to be more susceptible to experimentally induced colitis compared to controls; conversely, mice lacking the FAAH enzyme (thus presumably having higher levels of ECs), had greater protection against inflammatory agents (105).

A role for CB<sub>2</sub> receptors in gastrointestinal activity was only recently fully accepted. An earlier report on CB2-mediated effects (by the CB<sub>2</sub>-selective agonist HU-308) on defecation in mice (64), was followed by many negative reports on a role for CB<sub>2</sub> receptors in GI functions (96,97,106). For example croton oil-stimulated gastrointestinal transit was inhibited by WIN 55.212-2: this effect was reversed by rimonabant but not by the CB<sub>2</sub> receptor antagonist SR144528, indicating a CB<sub>1</sub> receptor-mediated inhibition of gastrointestinal transit (101). However, more recent reports indicated that while during resting conditions, CB2 receptor agonists did not affect gastrointestinal transit in rats, acceleration of intestinal motility was reduced by CB<sub>2</sub>, and not by CB<sub>1</sub> receptor activation after stimulation with lipopolysaccharide (LPS) (65). Although the issue needs further clarification, the accompanying commentary to this report was aptly entitled "Cannabinoids and intestinal motility: welcome to CB<sub>2</sub> receptor" (106).

# Adipose tissue

As indicated above, leptin is one of the first adipokine to be discovered. Several years ago, Di Marzo, Kunos and colleagues (107) showed for the first time a link between leptin and the ECBR system. Thus they reported that leptin injection to rats reduced concentrations of AEA and 2-AG. In the same study, elevated levels of ECs were found in mice (*ob/ob and db/db*)

and rats (Zucker) which display inherent impairments of the leptin system (107). Subsequently, CB<sub>1</sub> receptors have been detected in animal adipose tissue (89,108,109), while the full set of ECBR system components (CB<sub>1</sub> receptors, AEA and 2-AG, synthesizing and degrading enzymes) has recently been described in human adipose tissue (12). In a different study, CB<sub>2</sub> receptors were also detected on human adipose cells (110). Importantly, CB1 receptor knockout mice have less adipose tissue than wild type controls (89) and CB<sub>1</sub> receptors are dysregulated in human abdominal obesity (109).

The "classical", Rita Levi-Montalcini's neurotrophin NGF was recently introduced into the growing list of adipokines (10,11). Biology of NGF may be regulated by CB receptors, although thus far evidence for an interaction between NGF and the ECBR system has been shown only in nonadipose tissue. Thus, NGF receptor tropomyosin-related kinase A (TrkA) activation-induced inflammatory hyperalgesia was shown to be mediated by EC stimulation of CB<sub>1</sub> and CB<sub>2</sub> receptors on peripheral nerves (111). Further, interactions between (endo)CBs and NGF and/or brain-derived nerve growth factor (BDNF) have been proposed for the regulation of reward/addiction (112), pain processing (113) and cancer control (114). Recent evidence suggest that NGF/BDNF and the ECBR system interact in their modulation of brain plasticity, and emotional processes (112,115). Interestingly, NGF/BDNF have been associated with autism (116), while plasma NGF level were significantly raised in subjects classified as being in the first stage of romantic love (117). Thus it will be interesting to further investigate reciprocal relationships between these adipokines in the regulation of emotions/addiction, stress and social interaction. Furthermore, interactions between between NGF/BDNF and the ECBR system at the level of the adipose tissue should be addressed.

Adiponectin is another example of multifunctional adipokines (10,11,108,118-120). Plasma adiponectin levels, like those of NGF/BDNF (10), and differently from many other adipokines such as leptin, tumor necrosis factor- $\alpha$  and resistin (119), are reduced in patients with obesity and related cardiometabolic diseases (118,119). Interestingly, weight reducing effects of the CB<sub>1</sub> receptor antagonist/inverse agonist rimonabant was accompanied by enhanced levels of adiponectin in animal (108) as well as human (120) studies.

# Endocannabinoids in alimentary control

AEA-induced inhibition of defecation in mice was the first

report that ECs influence intestinal function (121). The ECs AEA and 2-AG have been detected in the small intestine and colon (97,100,122). Overall, the ECs are present in the GIT at several-fold higher concentrations than in the brain (100,123) and are physiologically active (122). Additionally, the degrading enzymes and uptake transporters for the ECs have been located in the GIT (100,122,124).

Two aspects of EC functioning in the alimentary system await final clarification.

- 1. Are the EC's constitutively active, or only when stimulated (such as in pathophysiological conditions)? It appears that both situations exist, depending on factors including genetic and species differences. For example, rimonabant induced emesis by itself in the shrew but not in the ferret. This species difference maybe explained by a higher constitutive level of ECs in shrews than in ferrets, in which the emetic stimulus may have to enhance EC concentrations before an emetic effect of rimonabant can be revealed (125,126). Further, cholera-toxin-induced fluid accumulation in the mural intestine, was accompanied by upregulated CB<sub>1</sub> receptor mRNA and elevated levels of AEA (104).
- 2. At least some of AEA's effects are mediated by non-CB receptors, some of which are known and some are not yet defined (55). For example, AEA activates TRPV1 vannilloid receptors (127). Thus, although exogenous CBs, acting *via* CB receptors, have anti-inflammatory effects in the GIT similarly to their effect in other systems and organs, application of the ECs AEA and 2-AG resulted in pro-inflammatory effects *via* TRPV1 receptors (128). Thus the 'duplicity' of the ECs as endovannilloids/ECs (129) suggests a complex role of the ECs in inflammatory states of the GIT. Furthermore, AEA inhibits, probably by allosteric modulation, serotonin 5-HT<sub>3</sub> receptor activation (130,131) and activates at least one nonCB<sub>1</sub>-nonCB<sub>2</sub> cannabinoid receptor ("CB3") (54-56).

Concluding this section, it clear that every level of the alimentary tract is affected by the ECBR system. Strikingly, ECBR influence on ingestion, digestion and emesis is always, at the organismic level, in harmony (see also Table 1). Thus (endo)CBs reduce intestinal and gastric motility, gastric acid secretion, emesis and nausea, are anti-diarrheal and enhance appetite. Conversely, CB<sub>1</sub> receptor antagonists cause emesis, anorexia and enhanced motility.

Future studies will have to address the relative importance of the various levels of ECBR involvement in alimentary

control, such as CNS-directed influence, compared to direct modulation by gastrointestinal/ENS sites. Izzo and colleagues have started to address this issue when they compared ED<sub>50</sub>values of the CB receptor agonist WIN55,212- and cannabinol-induced inhibition of intestinal transit after central (intracerebroventricular, icv) versus peripheral (intraperitoneal, ip) injections. ED<sub>50</sub> values were significantly, but not dramatically lower when injected icv. This prompted the authors to suggest that CB inhibit GI transit by both routes. However, after croton-oil-induced diarrhea, there were no differences in ED<sub>50</sub> values of *icv* or *ip* injected CBs. Moreover, ganglionic blockade with hexamethonium did not affect the inhibitory effect of ip injected CBs (101). In a subsequent study, the same group showed that after croton oil, intestinal CB<sub>1</sub> receptors were upregulated and that EC levels were not changed while the FAAH enzyme appeared at higher concentrations. These observations suggested that during inflammatory/diarhoea states, the local gut ECBR system is upregulated (100), apparently to reduce diarhoea and inflammation. Thus a dominantly peripheral response to pathological states by the ECBR system contrasts with the impression that in general in the brain-gut axis, CNS control plays a more major role when homeostasis is perturbed such as in digestive disorders or during psychological stress (5). Future studies will hopefully address the brain versus gut division of ECBR system labor in more detail.

Overall, the unified activity makes the ECBR system in the alimentary control system exquisitely suitable for multi-leveled treatment of syndromes such as irritable bowel syndrome and cachexia/anorexia, using ECBR-activating agents (84) or obesity/metabolic syndrome, using ECBR-inhibitors (132). A discussion of CB-based treatment of the metabolic syndrome/obesity is beyond the scope of this review and the reader is referred to previous publications (132-134). However, some of the promising approaches to treating GIT discomfort and disease, are presented in the next section.

# BRAIN-GUT BIDIRECTIONAL CONTROL VIA THE ECBR SYSTEM IN PATHOPHYSIOLOGICAL AND PSYCHOSOMATIC CONDITIONS

The ECBR system, as described above, is strongly represented in both brain and gut, or, more precisely in the CNS, in the dorsal vagal complex and at almost all levels of the gastrointestinal tract. Especially its involvement in stress and emotional processing make the ECBR system an excellent candidate to

study both as an underlying cause for pathological and psychosomatic problems of the digestive system as well as, as a target for new therapeutic approaches. Only conditions with a stress/psychosomatic component are considered here.

Irritable bowel syndrome. There is little doubt that stress and the HPA axis play a major role in IBS symptomatology. Experimentally, upregulation of CRH receptors in the PVN and locus ceroeleus have been shown after colonic stimulation (135). The role of cortisol in IBS has also been demonstrated although its levels may be up- or down-regulated depending on perhaps, the type of patients and conditions (36,136). This is in agreement with our own findings that low levels of corticosterone may signify decreased stress coping capability in mice (77). Solid evidence exists for an (endo)CB-regulated reduction of intestinal motility (121,122,137,138) through CB<sub>1</sub> (100) and CB<sub>2</sub> (65,121,139) receptors. This property of ECBR has been proposed to be beneficial for IBS which is often characterized by enhanced intestinal motility and contractility (122,124).

Inflammatory bowel disease (IBD). Inflammatory bowel diseases, including Crohn's disease and ulcerative colitis, result from inflammatory processes in the intestine and is characterized by ulcers, rectal bleeding diarrhea, nausea and lack of appetite (124). Major life stresses are thought to influence disease activity in IBS, while a role for minor daily stressors in IBD expression needs further investigation (36). In an animal model for Crohn's disease (orally applied croton oil to mice which causes inflammation in the small intestine), a beneficial, slowing effect of CBs on intestinal motility has been shown (100,101). This effect was apparently peripherally mediated (in the gut). Therefore such treatment is promising when central/psychological treatment is not likely to be helpful. As stated above, the importance of CB<sub>1</sub> receptors in protecting the organism against inflammation of the GI system was demonstrated in a study where CB<sub>1</sub> receptor-deficient mice were more vulnerable, while FAAH-deficient mice were less vulnerable to experimentally-induced colitis (105).

Interestingly, adiponectin levels were increased in mesenteric adipose tissue of patients with Crohn's disease, but not in patients with ulcerative colitis (140). Thus similarly to obesity, at least in Crohn's pathology too, ECs and adiponectin are counterbalanced. The putative significance of this finding needs further investigation.

Gastroesophageal reflux disease (GORD). This disorder results from a weak lower oesophagal sphincter causing gastric content to flow upward, hence symptoms including heartburn and acid regurgitation appear (124,141). The influence of psychological stress, anxiety and depression on this condition has been shown (141-143). An CB-centered treatment may be developed either through central effects or through peripheral muscle relaxation. Indeed, cannabinoid CB<sub>1</sub> receptor agonists have been shown to relax the oesophageal sphincter in dogs and ferrets. This effects was mediated by vagal neural loops at peripheral and central (dorsal vagal complex) sites (85,92,97).

Secretory diarrhea. Fluid-induced increase in stool volume, that is, secretory diarrhea, is caused by abnormal secretion and/or absorption of water and electrolytes as well as dysregulated intestinal motility (144,145). Psychological stress may precipitate secretory diarhoea, probably mediated by corticotrophin releasing hormone (CRH1) receptors (145). Activation of the HPA stress axis results in stress-induced secretory diarrhea (144). Stimulation of the ECBR system by the uptake inhibitor VDM11 reduced intestinal fluid accumulation; conversely, the CB<sub>1</sub> receptor antagonist rimonabant increased fluid accumulation (97). These observations suggest that the ECs exert a tonic regulation of intestinal secretory activity which can be up-or down-regulated according to the (psycho)physiological environment. Further, cannabinoid CB<sub>1</sub> receptor activation resulted in decreased fluid accumulation in the small intestine of the rat, suggesting that cannabinoid-based treatment may be beneficial for diarhoea (146).

Gastric ulcer. Summarizing a vast body of literature on the association between psychosocial factors (such as stress and the perception of stress) and peptic ulcers (36), Jones conservatively concludes that psychosocial factors may play a role in ulcer formation (147). Cannabinoid-induced reduction in gastric secretions and in ulcer formation has been observed many years ago and more recently, several reports have replicated and extended such observations to include the mediation by CB<sub>1</sub> receptors in stomach and/or central locations (93,124,148). More specifically, Win 55,212 reduced stress-induced ulcers in rats (148). From this study it is not clear whether the anti-stress effect was mediated centrally or peripherally. Possibly either or both routes are used under such conditions, since central stress-reducing effects of CBs have been shown (149), while the presence of CB<sub>1</sub> receptors in (pre- and postganglionic) vagal pathways leading to the stomach (93) may have been activated as well.

Emesis. Antiemetic effects of (endo)cannabinod have been extensively demonstrated and several  $\Delta 9$ -THC-like cannabinoid drugs (the synthetic nabilone and dronabinol) are selectively available for oral clinical use (97,124,150,151). The antiemetic effects are mediated by CB<sub>1</sub> receptor-mediated responses on gastric vagal nerves or centrally via the area postrema and dorsal vagal complex (97). Taken together, it is widely agreed that CB-based drugs especially those which do not have central side effects, should be developed as antiemetic drugs for cancer and AIDS patients who receive chemotherapeutic treatment. Cannabinoids may be especially effective in combating anticipatory nausea and vomiting (152). Since the area postrema has receptors located outside the CNS, CB<sub>1</sub> receptor-specific cannabinoids which are not active within the CNS, should be particularly suitable (153,154).

#### **DEVELOPMENT**

It is evident from the data summarized thus far, that the ECBR system is abundantly present and functionally important at all stages of ingestion and digestion in the adult organism. Although perhaps one of the very major systems (see also Table 1), the ECBR system and its CB<sub>1</sub> and CB<sub>2</sub> receptors do not seem critical for survival, as most CB1 and/or CB2 knockout mice reach adulthood, be it with reduced body weights (155-157). Similarly, very long term treatment (4 months) with the CB<sub>1</sub> receptor antagonist rimonabant did not cause mortality or (overt) detrimental effects, except a robust reduction in body weight (about 8%) (158). In contrast, we have shown that a single injection with rimonabant administered within 24 h of birth in mice permanently interferes with milk ingestion, resulting in growth failure and death in many cases. The effect was dose dependent and was observed in several strains of mice (ICR, SABRA, C57BL/6). The growth stunting effect was not seen in CB<sub>1</sub> receptor knockout pups but mortality was induced by rimonabant even in these knockouts, suggesting that these pups had developed a compensatory CB<sub>1</sub>-like ("CB<sub>3</sub>") receptor (56,159,160). Further psychobiological analyses of rimonabant-treated neonates suggested that CB<sub>1</sub> receptor activation in neonates is required for oral-motor development required for sucking (161). Since the behavioral and physiological deficiencies of CB<sub>1</sub> receptor-blocked mouse pups resemble infants suffering from "non-organic failure-to-thrive" (NOFTT), we have suggested that CB<sub>1</sub> receptor-blocked neonates may be used as a model for NOFTT and form the basis for the devel-

Table 2. Comparison between the physiological roles of endocannabinoids and ghrelin

Physiological function/location	ECBR system	Ghrelin	Selected references
Localization in GIT	Stomach, small intestine, colon	Mainly stomach, much smaller amounts in small intestine	22,100,122,174,175
Appetite/food intake	Orexigenic	orexigenic	74,175
Central/peripheral mechanism of food intake	Central (DVC, PVN/lateral hypothalamus) and peripheral	Central (arcutae nucleus, PVN) and peripheral	82,124,175,176
Colon	Decreased secretion/motility	enhanced motility	177
lleum	Relaxation/ Decreased secretion	No effect/decreased intestinal transit	177,178
Stomach	Decreased acid secretion/delayed emptying	Main source of ghrelin production; Enhanced gastric emptying/ Decreased acid secretion	22,84,97,178
Lower esophageal sphincter	Inhibition of relaxation	?	85,92
Dorsal vagal complex	Anti-emetic, GIT relaxation, GIT decreased secretion	Transmits peripheral ghrelin-induced signaling to hypothalamus	84,179,180
Interaction with leptin	Inverse	Inverse	17,24,107
HPA(stress) axis	Corticosterone increased/decreased	Corticosterone increased	77,149,181
Hypothalamus	Hypothermia	Hyperthermia/ hypothermia	14,181,182
CNS Motor areas(?)	Decreased locomotion	Increased/decreased locomotion	14,181,182
Bone and <b>-</b> cartilage homeostais	Bone (increased/decreased density)	Bone and cartilage homeostasis	22,162,183
Immune system/ inflammation	Immune regulation, anti-inflammatory	Immune regulation/ antiinflammatory	22,164,168
anxiety	Incease/decrease	Increase	165,167
sleep	Increase	Increase/decrease	166,167

opment of CB-based treatments.

#### CANNABINOID VERSUS GHRELIN

As stated above the ECBR system is ubiquitous and involved at many, if not all, levels of alimentary control and in many other functions. Interestingly, the recently discovered appetitive hormone ghrelin which binds to GHS receptors and is largely synthesized in the stomach (as described above), also participates in alimentary control at a number of levels. In addition to their appetite stimulating qualities, ghrelin and the CBs have a number of functions in common, including modulation of bone homeostasis, anxiety, sleep regulation, anti-cancer potential and immune modulation (22,162-168). Moreover, direct interaction between the two systems in the hypothalamus mediating food intake, has been demonstrated (82). Thus the remarkable functional and anatomical overlap between ghrelin and the ECs is examined in Table 2. Inspection of this information reveals that despite the similarities between CBs and ghrelin, these systems also have opposite effects in a number of spheres including intestinal motility and gastric secretion and perhaps temperature control and motor activity (see Table 2 for references).

In conclusion, ghrelin and CBs share a number of areas of influence in alimentary control and in other functions. However, the overlap is not complete. Further investigations into the interactions between the two systems seem warranted.

#### **CONCLUSIONS AND FUTURE DIRECTIONS**

Three major conclusions emanate from the material reviewed here: (i) the ECBR system is one of the major mediators between brain, the alimentary system and the adipose tissue, (ii) it is suggested that the role of the ECBR system in adult regulation of digestion and ingestion is a remnant of its critical role for the initiation of feeding and survival in the newborn, and (iii) the pervasive influence of the ECBR system in alimentary control make it a highly suitable target for therapeutic developments aimed at alleviating pathophysiological conditions such as IBD, IBS, gastric ulcers, nausea and anorexia.

Especially manipulation of CB<sub>1</sub> receptor activation will have therapeutic value in a number of alimentary pathophysiological conditions. However CB<sub>1</sub> receptor agonists often have undesirable psychoactive effects, such as confusion and

anxiety. In view of the predominantly CB<sub>1</sub> receptor-mediated effects on the GIT, it is desirable that CB-based treatment of GI disorders be mediated by ligands which do not cross the blood brain barrier or are not psychoactive for other reasons. We have recently demonstrated that several (+)cannabidiol derivates [such as (+)cannabidiol-demethyl heptyl and (+)carboxy-cannabidiol-demethyl heptyl], displayed CB<sub>1</sub>-receptor mediated inhibition of intestinal motility in the absence of central effects (153,154). Further studies on such compounds should address peripheral mechanisms of energy control including their effect on adipose tissue.

Upregulating the ECBR system by the inhibition of enzymatic breakdown or reuptake inhibition is expected to yield more selective therapeutic effects. However, preventing EC degradation with oleamide, a fatty acid amide which is considered by most (169-172) not to bind CB<sub>1</sub> receptors, resulted in psychoactive effects similar to those of AEA itself (169). On the other hand, a highly selective FAAH inhibitor URB597 is thought to have anxiolytic and antidepressant potential without sedative and addictive properties (173).

#### **ACKNOWLEDGMENTS**

This work was supported in part by a grant from Sanofi-Aventis (France) and by intramural support from the College of Judea and Samaria.

### **REFERENCES**

- Jones MP, Dilley JB, Drossman D, Crowell MD. Braingut connections in functional GI disorders: anatomic and physiologic relationships. *Neurogastroenterol Motil* 2006; 18: 91-103.
- 2. Konturek SJ, Konturek JW, Pawlik T, Brzozowski T. Brain-gut axis and its role in the control of food intake. *J Physiol Pharmacol* 2004; 55: 137-154.
- 3. Ringel Y. New directions in brain imaging research in functional gastrointestinal disorders. *Dig Dis* 2006; 24: 278-285.
- 4. Silverman DH, Munakata JA, Ennes H, Mandelkern MA, Hoh CK, Mayer EA. Regional cerebral activity in normal and pathological perception of visceral pain. *Gastroenterology* 1997; 112: 64-72.
- Mulak A, Bonaz B. Irritable bowel syndrome: a model of the brain-gut interactions. *Med Sci Monit* 2004; 10: RA55-62.

- Hang CH, Shi JX, Li JS, Wu W, Li WQ, Yin HX. Levels
  of vasoactive intestinal peptide, cholecystokinin and
  calcitonin gene-related peptide in plasma and jejunum
  of rats following traumatic brain injury and underlying
  significance in gastrointestinal dysfunction. World J
  Gastroenterol 2004; 10: 875-880.
- Welch MG, Keune JD, Welch-Horan TB, Anwar N, Anwar M, Ruggiero DA. Secretin activates visceral brain regions in the rat including areas abnormal in autism. Cell Mol Neurobiol 2003; 23: 817-837.
- 8. Filaretova L. The hypothalamic-pituitary-adrenocortical system: Hormonal brain-gut interaction and gastroprotection. *Auton Neurosci* 2006; 125: 86-93.
- 9. Kellow JE, Azpiroz F, Delvaux M, Gebhart GF, Mertz HR, Quigley EM, *et al.* Applied principles of neurogastroenterology: physiology/motility sensation. *Gastroenterology* 2006; 130: 1412-1420.
- Chaldakov GN, Stankulov IS, Hristova M, Ghenev PI. Adipobiology of disease: adipokines and adipokine-targeted pharmacology. *Curr Pharm Des* 2003; 9: 1023-1031.
- 11. Trayhurn P. Endocrine and signalling role of adipose tissue: new perspectives on fat. *Acta Physiol Scand* 2005; 184: 285-293.
- Spoto B, Fezza F, Parlongo G, Battista N, Sgro E, Gasperi V, et al. Human adipose tissue binds and metabolizes the endocannabinoids anandamide and 2-arachidonoylglycerol. Biochimie 2006; In press.
- Matias I, Gonthier MP, Orlando P, Martiadis V, De Petrocellis L, Cervino C, et al. Regulation, function, and dysregulation of endocannabinoids in models of adipose and beta-pancreatic cells and in obesity and hyperglycemia. J Clin Endocrinol Metab 2006; 91: 3171-3180.
- 14. Fride E. Endocannabinoids in the central nervous system: from neuronal networks to behavior. *Curr Drug Targets CNS Neurol Disord* 2005; 4: 633-642.
- 15. Mechoulam R, Hanus L, Fride E. Towards cannabinoid drugs revisited. *Prog Med Chem* 1998; 35: 199-243.
- Schwartz MW, Woods SC, Porte D, Jr., Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature* 2000; 404: 661-671.
- 17. Mechoulam R, Fride E. Physiology. A hunger for cannabinoids. *Nature* 2001; 410: 763, 765.
- 18. Strader AD, Woods SC. Gastrointestinal hormones and food intake. *Gastroenterology* 2005; 128: 175-191.

- 19. Zhang DM, Bula W, Stellar E. Brain cholecystokinin as a satiety peptide. *Physiol Behav* 1986; 36: 1183-1186.
- 20. Manni L, Lundeberg T, Tirassa P, Aloe L. Role of cholecystokinin-8 in nerve growth factor and nerve growth factor mRNA expression in carrageenan-induced joint inflammation in adult rats. *Rheumatology (Oxford)* 2002; 41: 787-792.
- 21. Neary NM, Small CJ, Bloom SR. Gut and mind. *Gut* 2003; 52: 918-921.
- 22. Lago F, Gonzalez-Juanatey JR, Casanueva FF, Gomez-Reino J, Dieguez C, Gualillo O. Ghrelin, the same peptide for different functions: player or bystander? *Vitam Horm* 2005; 71: 405-432.
- 23. Asakawa A, Inui A, Kaga T, Katsuura G, Fujimiya M, Fujino MA, *et al.* Antagonism of ghrelin receptor reduces food intake and body weight gain in mice. *Gut* 2003; 52: 947-952.
- 24. Cummings DE, Foster KE. Ghrelin-leptin tango in body-weight regulation. *Gastroenterology* 2003; 124: 1532-1535.
- 25. Ehrstrom M, Gustafsson T, Finn A, Kirchgessner A, Gryback P, Jacobsson H, *et al.* Inhibitory effect of exogenous orexin a on gastric emptying, plasma leptin, and the distribution of orexin and orexin receptors in the gut and pancreas in man. *J Clin Endocrinol Metab* 2005; 90: 2370-2377.
- 26. Kirchgessner AL. Orexins in the brain-gut axis. *Endocr Rev* 2002; 23: 1-15.
- Cowley MA, Smart JL, Rubinstein M, Cerdan MG, Diano S, Horvath TL, et al. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 2001; 411: 480-484.
- 28. Carlson N. *Physiology of Behavior*. 8 ed. New York: Pearson, 2004.
- 29. Schick RR, Schusdziarra V, Yaksh TL, Go VL. Brain regions where cholecystokinin exerts its effect on satiety. *Ann N Y Acad Sci* 1994; 713: 242-254.
- 30. Lee IS, Nam YS, Lee CH, Chung DW, Yoon YS, Kim JS, *et al.* Expressional changes of neuropeptide Y and cholecystokinin in the arcuate and paraventricular nuclei after capsaicin administration. *J Nutr Sci Vitaminol* (*Tokyo*) 2004; 50: 144-148.
- 31. Blackshaw LA. Receptors and transmission in the braingut axis: potential for novel therapies. IV. GABA(B) receptors in the brain-gastroesophageal axis. *Am J Physiol*

- Gastrointest Liver Physiol 2001; 281: G311-315.
- 32. Sternini C. Receptors and transmission in the braingut axis: potential for novel therapies. III. Mu-opioid receptors in the enteric nervous system. *Am J Physiol Gastrointest Liver Physiol* 2001; 281: G8-15.
- Dallman MF, Pecoraro N, Akana SF, La Fleur SE, Gomez F, Houshyar H, *et al*. Chronic stress and obesity: a new view of "comfort food". *Proc Natl Acad Sci U S A* 2003; 100: 11696-11701.
- 34. Kral JG. The pathogenesis of obesity: Stress and the brain-gut axis. *Surg Obes Relat Dis* 2005; 1: 25-34.
- 35. Monnikes H, Tebbe JJ, Hildebrandt M, Arck P, Osmanoglou E, Rose M, *et al.* Role of stress in functional gastrointestinal disorders. Evidence for stress-induced alterations in gastrointestinal motility and sensitivity. *Dig Dis* 2001; 19: 201-211.
- 36. Bhatia V, Tandon RK. Stress and the gastrointestinal tract. *J Gastroenterol Hepatol* 2005; 20: 332-339.
- 37. Drossman DA, Ringel Y, Vogt BA, Leserman J, Lin W, Smith JK, *et al.* Alterations of brain activity associated with resolution of emotional distress and pain in a case of severe irritable bowel syndrome. *Gastroenterology* 2003; 124: 754-761.
- 38. Shumyatsky GP, Tsvetkov E, Malleret G, Vronskaya S, Hatton M, Hampton L, *et al*. Identification of a signaling network in lateral nucleus of amygdala important for inhibiting memory specifically related to learned fear. *Cell* 2002; 111: 905-918.
- 39. Herranz R. Cholecystokinin antagonists: pharmacological and therapeutic potential. *Med Res Rev* 2003; 23: 559-605.
- Valdez GR, Zorrilla EP, Rivier J, Vale WW, Koob GF. Locomotor suppressive and anxiolytic-like effects of urocortin 3, a highly selective type 2 corticotropin-releasing factor agonist. *Brain Res* 2003; 980: 206-212.
- Volkow ND, Wang GJ, Maynard L, Jayne M, Fowler JS, Zhu W, et al. Brain dopamine is associated with eating behaviors in humans. Int J Eat Disord 2003; 33: 136-142.
- 42. Zheng H, Corkern M, Stoyanova I, Patterson LM, Tian R, Berthoud HR. Peptides that regulate food intake: appetite-inducing accumbens manipulation activates hypothalamic orexin neurons and inhibits POMC neurons. *Am J Physiol Regul Integr Comp Physiol* 2003; 284: R1436-1444.

- 43. Shu IW, Lindenberg DL, Mizuno TM, Roberts JL, Mobbs CV. The fatty acid synthase inhibitor cerulenin and feeding, like leptin, activate hypothalamic pro-opiomelanocortin (POMC) neurons. *Brain Res* 2003; 985: 1-12.
- 44. Abrahamsen GC, Berman Y, Carr KD. Curve-shift analysis of self-stimulation in food-restricted rats: relationship between daily meal, plasma corticosterone and reward sensitization. *Brain Res* 1995; 695: 186-194.
- 45. Fulton S, Woodside B, Shizgal P. Modulation of brain reward circuitry by leptin. *Science* 2000; 287: 125-128.
- 46. Gardner EL. Endocannabinoid signaling system and brain reward: emphasis on dopamine. *Pharmacol Biochem Behav* 2005; 81: 263-284.
- Laviolette SR, Grace AA. The roles of cannabinoid and dopamine receptor systems in neural emotional learning circuits: implications for schizophrenia and addiction. *Cell Mol Life Sci* 2006; 63: 1597-1613.
- 48. Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for cannabinoids. *Nature* 1993; 365: 61-65.
- Devane WA, Dysarz FA, 3rd, Johnson MR, Melvin LS, Howlett AC. Determination and characterization of a cannabinoid receptor in rat brain. *Mol Pharmacol* 1988; 34: 605-613.
- Matsuda LA, Lolait SJ, Brownstein MJ, Young AC, Bonner TI. Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* 1990; 346: 561-564.
- 51. Ryberg E, Vu HK, Larsson N, Groblewski T, Hjorth S, Elebring T, *et al.* Identification and characterisation of a novel splice variant of the human CB1 receptor. *FEBS Lett* 2005; 579: 259-264.
- 52. Shire D, Carillon C, Kaghad M, Calandra B, Rinaldi-Carmona M, Le Fur G, *et al*. An amino-terminal variant of the central cannabinoid receptor resulting from alternative splicing. *J Biol Chem* 1995; 270: 3726-3731.
- 53. Pertwee RG. Pharmacology of cannabinoid CB1 and CB2 receptors. *Pharmacol Ther* 1997; 74: 129-180.
- 54. Breivogel CS, Griffin G, Di Marzo V, Martin BR. Evidence for a new G protein-coupled cannabinoid receptor in mouse brain. *Mol Pharmacol* 2001; 60: 155-163.
- 55. Fride E. Endocannabinoids in the central nervous system an overview. *Prostaglandins Leukot Essent Fatty Acids* 2002; 66: 221-233.

- Fride E, Foox A, Rosenberg E, Faigenboim M, Cohen V, Barda L, et al. Milk intake and survival in newborn cannabinoid CB1 receptor knockout mice: evidence for a "CB3" receptor. Eur J Pharmacol 2003; 461: 27-34.
- 57. Howlett AC, Breivogel CS, Childers SR, Deadwyler SA, Hampson RE, Porrino LJ. Cannabinoid physiology and pharmacology: 30 years of progress. *Neuropharmacology* 2004; 47 Suppl 1: 345-358.
- 58. Marsicano G, Lutz B. Expression of the cannabinoid receptor CB1 in distinct neuronal subpopulations in the adult mouse forebrain. *Eur J Neurosci* 1999; 11: 4213-4225.
- 59. Crawley JN, Corwin RL. Biological actions of cholecystokinin. *Peptides* 1994; 15: 731-755.
- Hadjiivanova C, Belcheva S, Belcheva I. Cholecystokinin and learning and memory processes. *Acta Physiol Pharmacol Bulg* 2003; 27: 83-88.
- 61. Bachus SE, Hyde TM, Herman MM, Egan MF, Kleinman JE. Abnormal cholecystokinin mRNA levels in entorhinal cortex of schizophrenics. *J Psychiatr Res* 1997; 31: 233-256.
- 62. Cabral GA, Dove Pettit DA. Drugs and immunity: cannabinoids and their role in decreased resistance to infectious disease. *J Neuroimmunol* 1998; 83: 116-123.
- 63. Nunez E, Benito C, Pazos MR, Barbachano A, Fajardo O, Gonzalez S, *et al.* Cannabinoid CB2 receptors are expressed by perivascular microglial cells in the human brain: an immunohistochemical study. *Synapse* 2004; 53: 208-213.
- 64. Hanus L, Abu-Lafi S, Fride E, Breuer A, Vogel Z, Shalev DE, *et al.* 2-arachidonyl glyceryl ether, an endogenous agonist of the cannabinoid CB1 receptor. *Proc Natl Acad Sci USA* 2001; 98: 3662-3665.
- Mathison R, Ho W, Pittman QJ, Davison JS, Sharkey KA. Effects of cannabinoid receptor-2 activation on accelerated gastrointestinal transit in lipopolysaccharide-treated rats. *Br J Pharmacol* 2004; 142: 1247-1254.
- Van Sickle MD, Duncan M, Kingsley PJ, Mouihate A, Urbani P, Mackie K, *et al.* Identification and functional characterization of brainstem cannabinoid CB2 receptors. *Science* 2005; 310: 329-332.
- 67. Gong JP, Onaivi ES, Ishiguro H, Liu QR, Tagliaferro PA, Brusco A, *et al.* Cannabinoid CB2 receptors: immunohistochemical localization in rat brain. *Brain Res* 2006; 1071: 10-23.

- 68. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, *et al.* Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* 1992; 258: 1946-1949.
- Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. Biochem Pharmacol 1995; 50: 83-90.
- Huang SM, Bisogno T, Trevisani M, Al-Hayani A, De Petrocellis L, Fezza F, et al. An endogenous capsaicinlike substance with high potency at recombinant and native vanilloid VR1 receptors. Proc Natl Acad Sci USA 2002; 99: 8400-8405.
- Huang SM, Walker JM. Enhancement of spontaneous and heat-evoked activity in spinal nociceptive neurons by the endovanilloid/endocannabinoid N-arachidonoyldopamine (NADA). *J Neurophysiol* 2006; 95: 1207-1212.
- Porter AC, Sauer JM, Knierman MD, Becker GW, Berna MJ, Bao J, et al. Characterization of a novel endocannabinoid, virodhamine, with antagonist activity at the CB1 receptor. J Pharmacol Exp Ther 2002; 301: 1020-1024.
- 73. Ligresti A, Cascio MG, Di Marzo V. Endocannabinoid metabolic pathways and enzymes. *Curr Drug Targets CNS Neurol Disord* 2005; 4: 615-623.
- 74. Fride E, Bregman T, Kirkham TC. Endocannabinoids and food intake: newborn suckling and appetite regulation in adulthood. *Exp Biol Med (Maywood)* 2005; 230: 225-234.
- Hermann H, Marsicano G, Lutz B. Coexpression of the cannabinoid receptor type 1 with dopamine and serotonin receptors in distinct neuronal subpopulations of the adult mouse forebrain. *Neuroscience* 2002; 109: 451-460.
- Kirkham TC, Williams CM, Fezza F, Di Marzo V. Endocannabinoid levels in rat limbic forebrain and hypothalamus in relation to fasting, feeding and satiation: stimulation of eating by 2-arachidonoyl glycerol. *Br J Pharmacol* 2002; 136: 550-557.
- 77. Fride E, Suris R, Weidenfeld J, Mechoulam R. Differential response to acute and repeated stress in cannabinoid CB1 receptor knockout newborn and adult mice. *Behav Pharmacol* 2005; 16: 431-440.
- 78. Patel S, Cravatt BF, Hillard CJ. Synergistic inter-

actions between cannabinoids and environmental stress in the activation of the central amygdala. *Neuropsychopharmacology* 2005; 30: 497-507.

- 79. Hill MN, Gorzalka BB. Increased sensitivity to restraint stress and novelty-induced emotionality following long-term, high dose cannabinoid exposure. *Psychoneuroendocrinology* 2006; 31: 526-536.
- Wenger T, Ledent C, Tramu G. The endogenous cannabinoid, anandamide, activates the hypothalamo-pituitary-adrenal axis in CB1 cannabinoid receptor knockout mice. *Neuroendocrinology* 2003; 78: 294-300.
- 81. Weidenfeld J, Feldman S, Mechoulam R. Effect of the brain constituent anandamide, a cannabinoid receptor agonist, on the hypothalamo-pituitary-adrenal axis in the rat. *Neuroendocrinology* 1994; 59: 110-112.
- 82. Tucci SA, Rogers EK, Korbonits M, Kirkham TC. The cannabinoid CB1 receptor antagonist SR141716 blocks the orexigenic effects of intrahypothalamic ghrelin. *Br J Pharmacol* 2004; 143: 520-523.
- 83. McDonald AJ, Mascagni F. Localization of the CB1 type cannabinoid receptor in the rat basolateral amygdala: high concentrations in a subpopulation of cholecysto-kinin-containing interneurons. *Neuroscience* 2001; 107: 641-652.
- 84. Hornby PJ, Prouty SM. Involvement of cannabinoid receptors in gut motility and visceral perception. *Br J Pharmacol* 2004; 141: 1335-1345.
- Partosoedarso ER, Abrahams TP, Scullion RT, Moerschbaecher JM, Hornby PJ. Cannabinoid 1 receptor in the dorsal vagal complex modulates lower oesophageal sphincter relaxation in ferrets. *J Physiol* 2003; 550: 149-158.
- 86. Van Sickle MD, Oland LD, Mackie K, Davison JS, Sharkey KA. Delta9-tetrahydrocannabinol selectively acts on CB1 receptors in specific regions of dorsal vagal complex to inhibit emesis in ferrets. *Am J Physiol Gastrointest Liver Physiol* 2003; 285: G566-576.
- Krowicki ZK, Moerschbaecher JM, Winsauer PJ, Digavalli SV, Hornby PJ. Delta9-tetrahydrocannabinol inhibits gastric motility in the rat through cannabinoid CB1 receptors. *Eur J Pharmacol* 1999; 371: 187-196.
- Derbenev AV, Stuart TC, Smith BN. Cannabinoids suppress synaptic input to neurones of the rat dorsal motor nucleus of the vagus nerve. *J Physiol* 2004; 559: 923-938.

- 89. Cota D, Marsicano G, Tschop M, Grubler Y, Flachskamm C, Schubert M, *et al.* The endogenous cannabinoid system affects energy balance via central orexigenic drive and peripheral lipogenesis. *J Clin Invest* 2003; 112: 423-431.
- Burdyga G, Lal S, Varro A, Dimaline R, Thompson DG, Dockray GJ. Expression of cannabinoid CB1 receptors by vagal afferent neurons is inhibited by cholecystokinin. *J Neurosci* 2004; 24: 2708-2715.
- 91. Prestifilippo JP, Fernandez-Solari J, de la Cal C, Iribarne M, Suburo AM, Rettori V, *et al.* Inhibition of salivary secretion by activation of cannabinoid receptors. *Exp Biol Med (Maywood)* 2006; 231: 1421-1429.
- 92. Lehmann A, Blackshaw LA, Branden L, Carlsson A, Jensen J, Nygren E, *et al.* Cannabinoid receptor agonism inhibits transient lower esophageal sphincter relaxations and reflux in dogs. *Gastroenterology* 2002; 123: 1129-1134.
- 93. Adami M, Frati P, Bertini S, Kulkarni-Narla A, Brown DR, de Caro G, *et al.* Gastric antisecretory role and immunohistochemical localization of cannabinoid receptors in the rat stomach. *Br J Pharmacol* 2002; 135: 1598-1606.
- 94. Casu MA, Porcella A, Ruiu S, Saba P, Marchese G, Carai MA, *et al.* Differential distribution of functional cannabinoid CB1 receptors in the mouse gastroenteric tract. *Eur J Pharmacol* 2003; 459: 97-105.
- 95. Storr M, Gaffal E, Saur D, Schusdziarra V, Allescher HD. Effect of cannabinoids on neural transmission in rat gastric fundus. *Can J Physiol Pharmacol* 2002; 80: 67-76.
- 96. Pertwee RG. Cannabinoids and the gastrointestinal tract. *Gut* 2001; 48: 859-867.
- 97. Coutts AA, Izzo AA. The gastrointestinal pharmacology of cannabinoids: an update. *Curr Opin Pharmacol* 2004; 4: 572-579.
- 98. Coutts AA, Irving AJ, Mackie K, Pertwee RG, Anavi-Goffer S. Localisation of cannabinoid CB(1) receptor immunoreactivity in the guinea pig and rat myenteric plexus. *J Comp Neurol* 2002; 448: 410-422.
- 99. Landi M, Croci T, Rinaldi-Carmona M, Maffrand JP, Le Fur G, Manara L. Modulation of gastric emptying and gastrointestinal transit in rats through intestinal cannabinoid CB(1) receptors. *Eur J Pharmacol* 2002; 450: 77-83.

- 100. Izzo AA, Fezza F, Capasso R, Bisogno T, Pinto L, Iuvone T, et al. Cannabinoid CB1-receptor mediated regulation of gastrointestinal motility in mice in a model of intestinal inflammation. Br J Pharmacol 2001; 134: 563-570.
- 101. Izzo AA, Pinto L, Borrelli F, Capasso R, Mascolo N, Capasso F. Central and peripheral cannabinoid modulation of gastrointestinal transit in physiological states or during the diarrhoea induced by croton oil. *Br J Pharmacol* 2000; 129: 1627-1632.
- 102. Manara L, Croci T, Guagnini F, Rinaldi-Carmona M, Maffrand JP, Le Fur G, et al. Functional assessment of neuronal cannabinoid receptors in the muscular layers of human ileum and colon. Dig Liver Dis 2002; 34: 262-269.
- 103. MacNaughton WK, Van Sickle MD, Keenan CM, Cushing K, Mackie K, Sharkey KA. Distribution and function of the cannabinoid-1 receptor in the modulation of ion transport in the guinea pig ileum: relationship to capsaicin-sensitive nerves. *Am J Physiol Gastrointest Liver Physiol* 2004; 286: G863-871.
- 104. Izzo AA, Capasso F, Costagliola A, Bisogno T, Marsicano G, Ligresti A, et al. An endogenous cannabinoid tone attenuates cholera toxin-induced fluid accumulation in mice. Gastroenterology 2003; 125: 765-774.
- 105. Massa F, Marsicano G, Hermann H, Cannich A, Monory K, Cravatt BF, *et al.* The endogenous cannabinoid system protects against colonic inflammation. *J Clin Invest* 2004; 113: 1202-1209.
- Izzo AA. Cannabinoids and intestinal motility: welcome to CB2 receptors. *Br J Pharmacol* 2004; 142: 1201-1202.
- Di Marzo V, Goparaju SK, Wang L, Liu J, Batkai S, Jarai Z, et al. Leptin-regulated endocannabinoids are involved in maintaining food intake. *Nature* 2001; 410: 822-825.
- 108. Bensaid M, Gary-Bobo M, Esclangon A, Maffrand JP, Le Fur G, Oury-Donat F, et al. The cannabinoid CB1 receptor antagonist SR141716 increases Acrp30 mRNA expression in adipose tissue of obese fa/fa rats and in cultured adipocyte cells. Mol Pharmacol 2003; 63: 908-914.
- 109. Bluher M, Engeli S, Kloting N, Berndt J, Fasshauer M, Batkai S, et al. Dysregulation of the peripheral and adipose tissue endocannabinoid system in human abdominal

- obesity. Diabetes 2006; 55: 3053-3060.
- 110. Roche R, Hoareau L, Bes-Houtmann S, Gonthier MP, Laborde C, Baron JF, et al. Presence of the cannabinoid receptors, CB1 and CB2, in human omental and subcutaneous adipocytes. *Histochem Cell Biol* 2006; 126: 177-187.
- 111. Farquhar-Smith WP, Jaggar SI, Rice AS. Attenuation of nerve growth factor-induced visceral hyperalgesia via cannabinoid CB(1) and CB(2)-like receptors. *Pain* 2002; 97: 11-21.
- 112. Butovsky E, Juknat A, Goncharov I, Elbaz J, Eilam R, Zangen A, et al. In vivo up-regulation of brain-derived neurotrophic factor in specific brain areas by chronic exposure to Delta-tetrahydrocannabinol. *Journal of Neurochemistry* 2005; 93: 802-811.
- 113. Rice AS, Farquhar-Smith WP, Nagy I. Endocannabinoids and pain: spinal and peripheral analgesia in inflammation and neuropathy. *Prostaglandins Leukot Essent Fatty Acids* 2002; 66: 243-256.
- 114. Velasco L, Ruiz L, Sanchez MG, Diaz-Laviada I. delta(9)-Tetrahydrocannabinol increases nerve growth factor production by prostate PC-3 cells. Involvement of CB1 cannabinoid receptor and Raf-1. Eur J Biochem 2001; 268: 531-535.
- 115. Berghuis P, Dobszay MB, Wang X, Spano S, Ledda F, Sousa KM, *et al.* Endocannabinoids regulate interneuron migration and morphogenesis by transactivating the TrkB receptor. *Proc Natl Acad Sci USA* 2005; 102: 19115-19120.
- 116. Tsaj S. Is autism caused by early hyperactivity of brainderived neurotrophic factor? *Med Hypotheses* 2006; 65: 79-82.
- 117. Emanuele E, Politi P, Bianchi M, Minoretti P, Bertona M, Geroldi D. Raised plasma nerve growth factor levels associated with early-stage romantic love. *Psychoneuroendocrinology* 2006; 31: 288-294.
- 118. Whitehead JP, Richards AA, Hickman IJ, Macdonald GA, Prins JB. Adiponectin a key adipokine in the metabolic syndrome. *Diabetes Obes Metab* 2006; 8: 264-280.
- Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Invest* 2006; 116: 1784-1792.
- 120. Despres JP, Golay A, Sjostrom L. Effects of rimonabant

on metabolic risk factors in overweight patients with dyslipidemia. *N Engl J Med* 2005; 353: 2121-2134.

- 121. Fride E. Anandamides: tolerance and cross-tolerance to delta 9-tetrahydrocannabinol. *Brain Res* 1995; 697: 83-90.
- 122. Pinto L, Izzo AA, Cascio MG, Bisogno T, Hospodar-Scott K, Brown DR, *et al.* Endocannabinoids as physiological regulators of colonic propulsion in mice. *Gastroenterology* 2002; 123: 227-234.
- 123. Pinto L, Capasso R, Di Carlo G, Izzo AA. Endocannabinoids and the gut. *Prostaglandins Leukot Essent Fatty Acids* 2002; 66: 333-341.
- 124. Di Carlo G, Izzo AA. Cannabinoids for gastrointestinal diseases: potential therapeutic applications. *Expert Opin Investig Drugs* 2003; 12: 39-49.
- 125. Van Sickle MD, Oland LD, Ho W, Hillard CJ, Mackie K, Davison JS, *et al*. Cannabinoids inhibit emesis through CB1 receptors in the brainstem of the ferret. *Gastroenterology* 2001; 121: 767-774.
- 126. Darmani NA, Janoyan JJ, Kumar N, Crim JL. Behaviorally active doses of the CB1 receptor antagonist SR 141716A increase brain serotonin and dopamine levels and turnover. *Pharmacol Biochem Behav* 2003; 75: 777-787.
- 127. Di Marzo V, Bisogno T, De Petrocellis L. Anandamide: some like it hot. *Trends Pharmacol Sci* 2001; 22: 346-349.
- 128. McVey DC, Schmid PC, Schmid HH, Vigna SR. Endocannabinoids induce ileitis in rats via the capsaicin receptor (VR1). J Pharmacol Exp Ther 2003; 304: 713-722.
- 129. Di Marzo V, Blumberg PM, Szallasi A. Endovanilloid signaling in pain. *Curr Opin Neurobiol* 2002; 12: 372-379.
- 130. Fride E, Ben-Shabat S, Mechoulam R. Pharmacology of anandamide: Interactions with serotonin systems? *International Cannabinoid Research Society Meeting* 1998.
- 131. Morales M, Wang SD, Diaz-Ruiz O, Jho DH. Cannabinoid CB1 receptor and serotonin 3 receptor subunit A (5-HT3A) are co-expressed in GABA neurons in the rat telencephalon. *J Comp Neurol* 2004; 468: 205-216.
- 132. Wierzbicki AS. Rimonabant: endocannabinoid inhibition for the metabolic syndrome. *Int J Clin Pract* 2006; 60: 1697-1706.

- 133. Carai MA, Colombo G, Maccioni P, Gessa GL. Efficacy of rimonabant and other cannabinoid CB1 receptor antagonists in reducing food intake and body weight: preclinical and clinical data. CNS Drug Rev 2006; 12: 91-99.
- 134. Curioni C, Andre C. Rimonabant for overweight or obesity. *Cochrane Database Syst Rev* 2006: CD006162.
- 135. Valentino RJ, Miselis RR, Pavcovich LA. Pontine regulation of pelvic viscera: pharmacological target for pelvic visceral dysfunctions. *Trends Pharmacol Sci* 1999; 20: 253-260.
- 136. Heitkemper M, Jarrett M, Cain K, Shaver J, Bond E, Woods NF, et al. Increased urine catecholamines and cortisol in women with irritable bowel syndrome. Am J Gastroenterol 1996; 91: 906-913.
- 137. Fride E, Mechoulam R. Pharmacological activity of the cannabinoid receptor agonist, anandamide, a brain constituent. *Eur J Pharmacol* 1993; 231: 313-314.
- 138. Pertwee RG, Fernando SR, Nash JE, Coutts AA. Further evidence for the presence of cannabinoid CB1 receptors in guinea-pig small intestine. *Br J Pharmacol* 1996; 118: 2199-2205.
- 139. Hanus L, Breuer A, Tchilibon S, Shiloah S, Goldenberg D, Horowitz M, et al. HU-308: a specific agonist for CB(2), a peripheral cannabinoid receptor. Proc Natl Acad Sci USA 1999; 96: 14228-14233.
- 140. Yamamoto K, Kiyohara T, Murayama Y, Kihara S, Okamoto Y, Funahashi T, *et al.* Production of adiponectin, an anti-inflammatory protein, in mesenteric adipose tissue in Crohn's disease. *Gut* 2005; 54: 789-796.
- 141. Fass R, Ofman JJ. Gastroesophageal reflux disease - should we adopt a new conceptual framework? Am J Gastroenterol 2002; 97: 1901-1909.
- 142. Wright CE, Ebrecht M, Mitchell R, Anggiansah A, Weinman J. The effect of psychological stress on symptom severity and perception in patients with gastro-oesophageal reflux. *J Psychosom Res* 2005; 59: 415-424.
- 143. Richter JE, Bradley LC. Psychophysiological interactions in esophageal diseases. *Semin Gastrointest Dis* 1996; 7: 169-184.
- 144. Sanger GJ, Yoshida M, Yahyah M, Kitazumi K. Increased defecation during stress or after 5-hydroxytryptophan: selective inhibition by the 5-HT(4) receptor antagonist, SB-207266. *Br J Pharmacol* 2000; 130: 706-712.

- 145. Saunders PR, Maillot C, Million M, Tache Y. Peripheral corticotropin-releasing factor induces diarrhea in rats: role of CRF1 receptor in fecal watery excretion. *Eur J Pharmacol* 2002; 435: 231-235.
- 146. Tyler K, Hillard CJ, Greenwood-Van Meerveld B. Inhibition of small intestinal secretion by cannabinoids is CB1 receptor-mediated in rats. *Eur J Pharmacol* 2000; 409: 207-211.
- 147. Jones MP. The role of psychosocial factors in peptic ulcer disease: beyond *Helicobacter pylori* and NSAIDs. *J Psychosom Res* 2006; 60: 407-412.
- 148. Germano MP, D'Angelo V, Mondello MR, Pergolizzi S, Capasso F, Capasso R, et al. Cannabinoid CB1-mediated inhibition of stress-induced gastric ulcers in rats. Naunyn Schmiedebergs Arch Pharmacol 2001; 363: 241-244.
- 149. Patel S, Roelke CT, Rademacher DJ, Cullinan WE, Hillard CJ. Endocannabinoid signaling negatively modulates stress-induced activation of the hypothalamic-pituitary-adrenal axis. *Endocrinology* 2004; 145: 5431-5438.
- Darmani NA. Methods evaluating cannabinoid and endocannabinoid effects on gastrointestinal functions. *Methods Mol Med* 2006; 123: 169-189.
- 151. Parker LA, Kwiatkowska M, Burton P, Mechoulam R. Effect of cannabinoids on lithium-induced vomiting in the Suncus murinus (house musk shrew). Psychopharmacology (Berl) 2004; 171: 156-161.
- 152. Parker LA, Kemp SW. Tetrahydrocannabinol (THC) interferes with conditioned retching in Suncus murinus: an animal model of anticipatory nausea and vomiting (ANV). *Neuroreport* 2001; 12: 749-751.
- 153. Fride E, Feigin C, Ponde DE, Breuer A, Hanus L, Arshavsky N, *et al.* (+)-Cannabidiol analogues which bind cannabinoid receptors but exert peripheral activity only. *Eur J Pharmacol* 2004; 506: 179-188.
- 154. Fride E, Ponde D, Breuer A, Hanus L. Peripheral, but not central effects of cannabidiol derivatives: Mediation by CB(1) and unidentified receptors. *Neuropharmacology* 2005; 48: 1117-1129.
- 155. Zimmer A, Zimmer AM, Hohmann AG, Herkenham M, Bonner TI. Increased mortality, hypoactivity, and hypoalgesia in cannabinoid CB1 receptor knockout mice. *Proc Natl Acad Sci USA* 1999; 96: 5780-5785.
- 156. Ledent C, Valverde O, Cossu G, Petitet F, Aubert JF, Beslot F, *et al.* Unresponsiveness to cannabinoids and

- reduced addictive effects of opiates in CB1 receptor knockout mice. *Science* 1999; 283: 401-404.
- 157. Paria BC, Song H, Wang X, Schmid PC, Krebsbach RJ, Schmid HH, *et al.* Dysregulated cannabinoid signaling disrupts uterine receptivity for embryo implantation. *J Biol Chem* 2001; 276: 20523-20528.
- 158. Gobshtis N, Ben-Shabat S, Fride E. Antidepressant-induced undesirable weight gain: Prevention with rimonabant without interference with behavioral effectiveness. *Eur J Pharmacol* 2007; 554: 155-63.
- 159. Fride E, Ginzburg Y, Breuer A, Bisogno T, Di Marzo V, Mechoulam R. Critical role of the endogenous cannabinoid system in mouse pup suckling and growth. *Eur J Pharmacol* 2001; 419: 207-214.
- 160. Fride E. The endocannabinoid-CB(1) receptor system in pre- and postnatal life. *Eur J Pharmacol* 2004; 500: 289-297
- 161. Fride E, Peretz-Ezra D, Arshavsky N, H D, Weller A. The CB<sub>1</sub> receptor and "non-organic failure-to thrive" in infants: the first animal (mouse) model. 33rd Annual Meeting of the Society for Neuroscience Washington DC, USA 2005.
- 162. Tam J, Ofek O, Fride E, Ledent C, Gabet Y, Muller R, et al. Involvement of neuronal cannabinoid receptor CB1 in regulation of bone mass and bone remodeling. Mol Pharmacol 2006; 70: 786-792.
- 163. Onaivi ES, Sugiura T, Di Marzo V. *Endocannabinoids*. Boca Raton: CRC Taylor & Francis, 2005.
- 164. Cabral GA, Staab A. Effects on the immune system. *Handb Exp Pharmacol* 2005: 385-423.
- 165. Valverde O, Karsak M, Zimmer A. Analysis of the endocannabinoid system by using CB1 cannabinoid receptor knockout mice. *Handb Exp Pharmacol* 2005: 117-145.
- 166. Weikel JC, Wichniak A, Ising M, Brunner H, Friess E, Held K, et al. Ghrelin promotes slow-wave sleep in humans. Am J Physiol Endocrinol Metab 2003; 284: E407-415.
- 167. Carlini VP, Monzon ME, Varas MM, Cragnolini AB, Schioth HB, Scimonelli TN, *et al*. Ghrelin increases anxiety-like behavior and memory retention in rats. *Biochem Biophys Res Commun* 2002; 299: 739-743.
- Carrier EJ, Patel S, Hillard CJ. Endocannabinoids in neuroimmunology and stress. *Curr Drug Targets CNS Neurol Disord* 2005; 4: 657-665.

169. Mechoulam R, Fride E, Hanus L, Sheskin T, Bisogno T, Di Marzo V, et al. Anandamide may mediate sleep induction. *Nature* 1997; 389: 25-26.

- 170. Sheskin T, Hanus L, Slager J, Vogel Z, Mechoulam R. Structural requirements for binding of anandamide-type compounds to the brain cannabinoid receptor. *J Med Chem* 1997; 40: 659-667.
- 171. Fowler CJ. Oleamide: a member of the endocannabinoid family? *Br J Pharmacol* 2004; 141: 195-196.
- 172. Leggett JD, Aspley S, Beckett SR, D'Antona AM, Kendall DA, Kendall DA. Oleamide is a selective endogenous agonist of rat and human CB1 cannabinoid receptors. *Br J Pharmacol* 2004; 141: 253-262.
- 173. Piomelli D, Tarzia G, Duranti A, Tontini A, Mor M, Compton TR, *et al.* Pharmacological profile of the selective FAAH inhibitor KDS-4103 (URB597). *CNS Drug Rev* 2006; 12: 21-38.
- 174. Matias I, McPartland JM, Di Marzo V. Occurrence and possible biological role of the endocannabinoid system in the sea squirt Ciona intestinalis. *J Neurochem* 2005; 93: 1141-1156.
- 175. Cummings DE, Shannon MH. Roles for ghrelin in the regulation of appetite and body weight. *Arch Surg* 2003; 138: 389-396.
- 176. Cowley MA, Grove KL. Ghrelin satisfying a hunger for the mechanism. *Endocrinology* 2004; 145: 2604-2606.
- 177. Shimizu Y, Chang EC, Shafton AD, Ferens DM, Sanger

- GJ, Witherington J, *et al.* Evidence that stimulation of ghrelin receptors in the spinal cord initiates propulsive activity in the colon of the rat. *J Physiol* 2006; 576: 329-338.
- 178. Sallam H, Oliveira HM, Gan HT, Herndon D, Chen J. Ghrelin improves burn-induced delayed gastrointestinal transit in rats. *Am J Physiol Regul Integr Comp Physiol* 2006. *in press*
- 179. Solomon A, De Fanti BA, Martinez JA. The nucleus tractus solitari (NTS) participates in peripheral ghrelin glucostatic hunger signalling mediated by insulin. *Neuropeptides* 2006; 40: 169-175.
- 180. Zigman JM, Jones JE, Lee CE, Saper CB, Elmquist JK. Expression of ghrelin receptor mRNA in the rat and the mouse brain. *J Comp Neurol* 2006; 494: 528-548.
- 181. Jaszberenyi M, Bujdoso E, Bagosi Z, Telegdy G. Mediation of the behavioral, endocrine and thermoregulatory actions of ghrelin. *Horm Behav* 2006; 50: 266-273.
- 182. Tang-Christensen M, Vrang N, Ortmann S, Bidlingmaier M, Horvath TL, Tschop M. Central administration of ghrelin and agouti-related protein (83-132) increases food intake and decreases spontaneous locomotor activity in rats. *Endocrinology* 2004; 145: 4645-4652.
- 183. Idris AI, van 't Hof RJ, Greig IR, Ridge SA, Baker D, Ross RA, *et al*. Regulation of bone mass, bone loss and osteoclast activity by cannabinoid receptors. *Nat Med* 2005; 11: 774-779.