AMYGDALA AND SUBCORTICAL VISION: RECOGNITION OF THREAT AND FEAR

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The amygdala (Am) is a relatively voluminous gray substance, located in the depth of the ventromedial temporal lobe. The Am has diverse afferent and efferent connections throughout the neuraxis, and is involved in the modulation of neuroendocrine functions, visceral effector mechanisms, and in complex patterns of behavior: learning and memory, aggression and defense, pain modulation, reproduction, food intake, etc. A recently revealed important function of the Am is that it acts as the brain ' lighthouse' which constantly monitors the environment for stimuli which signal a threat to the organism. The data from patients with extensive lesions of the striate cortex indicate that unseen fearful and fear-conditioned faces elicit increased Am responses. Thus, also extrageniculostriate pathways are involved. A multisynaptic pathway from the retina to the Am via the superior colliculus and several thalamic nuclei was recently suggested. We here present data based on retrograde neuronal labeling that the parabigeminal nucleus emits a substantial bilateral projection to the Am. This small cholinergic nucleus (Ch8 group) in the midbrain tegmentum is a subcortical relay visual center that is reciprocally connected with the superior colliculus. We suggest the existence of a second extrageniculostriate multisynaptic connection to Am: retina – superior colliculus – parabigeminal nucleus – Am. This pathway might be very effective since all tracts listed above are bilateral. The function of the Am by the rapid response to the sources of threat before conscious detection is significantly altered by various neuropsychiatric diseases. *Biomed Rev 2008; 19: 1-16.*

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INTRODUCTION

The amygdala (Am) is a relatively voluminous gray substance, located in the depth of the ventromedial temporal lobe, ventral to the caudolateral striatum and to the pallidum. It is a very complicated structure and consists of several nuclei, divided on the basis of cytoarchitectonic, hodological, histochemical and immunohistochemical studies (1-22). The Am has diverse afferent and efferent connections throughout the neuraxis, from the cerebral cortex to the spinal cord, and is involved in the modulation of neuroendocrine functions, visceral effector mechanisms, and in complex patterns of behavior: learning and memory, aggression and defense, pain modulation, reproduction, food intake, etc. (15,18,23-46).

Recently, a new function of Am was described by two independently working teams (47-50). They found out that Am is a part of the visual system, responsible for the recognition of threat signals. Adolphs *et al* (47,48) observed a patient with a rare, autosomal recessive, genetic disease – the Urbach-Wiethe disease see 51 for exhaustive description), which has resulted in bilateral calcification of her amygdala. Young *et al* (49,50) examined a female patient with a partial bilateral amygdalotomy. Both teams provided the intriguing evidence that the bilateral Am damage compromises the recognition of fear in facial expressions while leaving intact recognition of face identity.

Amygdala and subcortical vision

Am does not receive a monosynaptic retinal input (52), e.g. the visual information should follow a polysynaptic pathway. However, several retrograde tracing studies scanned throughout the thalamus and brainstem the cells of origin of pathways to the Am but have not encountered retrogradely labeled neurons in structures that receive a monosynaptic retinal input: the lateral geniculate body, the pretectal area, the superior colliculus and the terminal nuclei of the optic tract (22,53-58).

Visual information reaches the Am from association areas rather than primary cortex (8,11,19,59,60). The observations of Adolphs *et al* (47,48) and of Young *et al* (49,50) quoted above were immediately followed by a considerable body of investigations that confirmed and extended these significant data (28,33,34,60-87). The eyes may be a particularly salient stimulus for the Am (88). Kawashima *et al* (89) reported left Am activation when subjects interpreted gaze direction, whereas the right Am was activated during eye-to-eye contact. Whalen *et al* (90) found out that the eye region was sufficient to elicit Am responses during fMRI. More specifically, the white sclera surrounding the dark pupil in fearful eyes was a necessary component of the stimulus. Consistent with these findings, a patient with bilateral damage to the Am failed to use information from the eye region when viewing faces (91). More recently, Demos *et al* (87) established that the Am is sensitive to variations in the pupil size of others. There is increased bilateral Am activity for faces with relatively large pupils, e.g. there is also a function for the Am in the detection of changes in pupil size, an index of arousal and/or interest.

Importantly, the data from healthy volunteers when masking procedures were used, and in patients with extensive lesions of the striate cortex indicate that "unseen" fearful and fear-conditioned faces elicit increased Am responses (64,66,67,78,86,90,92-95). Apparently, extrageniculate pathways are involved (64,66-68,72,78,80,90,93,94). Morris et al (67) suggest that the retinal inputs reach the Am via a multisynaptic pathway: superior colliculus – pulvinar – Am. This multineuronal chain was already traced in the previous studies of Benevento and Fallon (96), Jones and Burton (97) and Grieve et al (98). A second multineuronal chain was described by Linke et al (99). They traced to the lateral Am nucleus axons from the suprageniculate nucleus, the medial division of the medial geniculate nucleus, and from the posterior intralaminar thalamic nuclei. All these structures receive an afferent input from the superior colliculus. According to Zald (36), the recent observations suggest that the Am may be the lynch-pin of an organism's ability to rapidly respond to sources of threat without explicit knowledge of the presence of stimulus, i. e. before conscious detection. Data are accumulating that the Am acts as the brain's "lighthouse", which constantly monitors the environment for stimuli which signal a threat or danger to the organism (33,82).

The Am receives a noradrenergic input from locus ceruleus (10,22,100; and references therein). Recently, Liddell *et al* (82) examined the direct brainstem-Am "alarm" system for subliminal signals of fear and pointed out that locus ceruleus also plays a significant role. Previous animal experiments also provided evidence that locus ceruleus is strongly involved in facilitating the rapid neural responses to fear-related signals (101).

Following the descriptions of two multineuronal chains conveying visual information to the Am (96-99), we described a third multisynaptic pathway in which the parabigeminal nucleus plays a significant role (22,102).

Parabigeminal nucleus: structure, transmitters, connections, and participation in extrageniculostriate visual pathways

The term "corpus parabigeminum" was introduced by von Bechterew (103), according to Flechsig's suggestion to avoid confusion with nucleus lemnisci lateralis. Presently, the term "parabigeminal nucleus" (Pbg) is used. It is a small structure, located subpially along the lateral border of the mesencephalon, dorsocaudolateral to the substantia nigra (Fig. 1). It contains densely arranged, small ovoid and elongated



Figure 1. The extent of the Pbg of the right hemisphere of a rat as shown by equidistant 5 μ m thick sections stained with an optimized Gallyas silver stain protocol (104). The Pbg can be followed over a rostro-caudal distance of approximately 600 μ m. The most rostral section containing the Pbg is signed with 0 μ m, the most caudal one with 580 μ m. Scale bar = 5 mm.

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neurons. Although with some discrepancies, Pbg was described in various mammalian species (reviewed in 105,106), while in non-mammalian species Pbg corresponds to the nucleus isthmi (107,108).

Mufson *et al* (109) first convincingly described that the Pbg neurons are cholinergic and designated them as Ch8 group. This was confirmed repeatedly (110-121; see also Fig. 2). However, no cholinergic cells are present in Pbg of the monotremes (122) and of bats (123).

The classical studies (reviewed in 124) considered Pbg to be one of the nuclear groups, associated with the lateral lemniscus, but the Nauta silver impregnation studies of van Noort (124) found no connections between Pbg and the inferior colliculus. On the other hand, numerous studies in the last decades pointed out that Pbg is interconnected with several subcortical visual centers: superior colliculus, lateral geniculate body, striaterecipient zone of the pulvinar, and suprachiasmatic nucleus (96,106,108,110,113,114,117-118,124-141). In particular, the reciprocal connections between the Pbg and the superior colliculus are so strong that Graybiel (127) designated the Pbg as a "satellite system of the superior colliculus".

The hodological studies quoted above have not described interconnections between Am and Pbg. Notably, several retrograde tracing studies described various Am afferents from the



Figure 2. Low magnification microphotograph of the caudolateral mesencephalic tegmentum. Medial is to the right. The cholinergic neurons are demonstrated with immunohistochemical staining of choline acetyltransferase. The small but sharply demarcated region containing densely arranged small cholinergic neurons is the Pbg (Ch8 group). Medial to it several scattered cholinergic neurons belong to pars dissipata of the pedunculopontine tegmental nucleus (Ch5 group). Scale bar = 250 µm.

brainstem as well (53-58) but have not mentioned a pathway from Pbg.

While reviewing the multineuronal pain pathways that involve the Am (41) we decided to reinvestigate some connections (from spinal cord, spinal trigeminal nucleus, parabrachial nuclear complex, raphe nuclei, locus ceruleus, substantia nigra and associated dopaminergic neuronal groups) by means of retrograde transport of Fluoro-Gold, injected stereotaxically in the Am (Fig. 3). Numerous sources of afferent connections from the brainstem were described (22), and two of them, from the spinal trigeminal nucleus and from Pbg, were completely original.



Figure 3. Low magnification microphotograph of the injection focus involving the basomedial nucleus of the Am (AmBm) and to a lesser extent the ventral portion of the central nucleus (AmCe). AmMe – medial nucleus of Am, AmBl – basolateral nucleus of Am. Medial is to the left. Scale bar = 250 µm.

The main target of the Pbg axons is the central nucleus of Am (Fig. 4a, b). In the cases with selective infiltration of the central nucleus with this very effective retrograde tracer, the Pbg exhibits bilaterally strong fluorescence of its neurons (Fig. 4a), whereas in cases with only a partial infiltration of the central nucleus is present, the Pbg neurons display only a light fluorescence (Fig. 4b). When followed on serial sections (Fig. 5a-d) it is evident that almost all sectors of Pbg send axons to Am and only the caudal pole of the ipsilateral Pbg remains unlabeled.

Our results strongly suggest that the newly described efferent connection from the Pbg to the Am might be a component of a third disynaptic connection from the superior colliculus to the Am, since this nucleus receives a significant input from the superior colliculus (96,106,125-128,138), and according to our results, the Pbg also projects to the Am bilaterally.

A plausible explanation, why the Pbg is constantly missing from the numerous data reviewed above, is its small size also in the human brain (115,142,143). Let us recall that even the keen eyes of Olszewski and Baxter (144) failed to recognize the Pbg as a separate entity in the human brainstem, and it was included only in the atlas of Paxinos and Huang (145) with the help of acetylcholinesterase staining.

It is remarkable that this tiny nucleus, with small neurons, emits so prominent efferent connections that in the cases of the superior colliculus and Am are bilateral. Indeed, the neurons are densely arranged, and the comparison with Nissl-stained sections suggests that at least the great majority, if not all, of the Pbg cells are projection neurons (see also 112). Nevertheless, the comparison of the number of Pbg cells with their broad and diverse innervation territories, invites the speculation that the Pbg neurons are able to innervate more than one target by means of divergent axon collaterals.

To test this hypothesis we carried out a double labeling study (102). We injected two fluorescing retrograde tracers in two targets (Fig. 6a-f, Fig. 7a, b). In the central nucleus of Am was injected Fluoro-Gold (the labeled neurons exhibit a yellow fluorescence), and in the superior colliculus multiple injections of Fluoro-Emerald were placed; by this tracer the labeled neurons exhibit a green fluorescence. These two tracers are fluorescing at different wavelength, so that each field was observed and photographed with two filters. As described above, the connection of the Pbg to central nucleus of Am is bilateral. On the ipsilateral side, the Pbg-Am neurons are concentrated in the central portion of Pbg, leaving the dorsal and ventral parts of the nucleus unlabeled (Fig. 7c). The crossed connection is more prominent. On the contralateral side the labeled neurons are located throughout the Pbg (Fig. 7d). The projection of Pbg to the superior colliculus is shown in Figs. 7e, f. This connection is also bilateral. On the ipsilateral side, two sharply delineated groups in Pbg are present - dorsal and ventral (Fig. 7e). Contralaterally projecting Pbg cells are distributed throughout the dorsoventral extent of the nucleus but are mainly concentrated in the central portion (Fig. 7f). The simultaneous tracing is demonstrated in Figs. 7g, h. It is evident that on the ipsilateral side (Fig. 7g) Pbg-Am and Pbg-superior colliculus tracts arise largely (if not exclusively) from different cell populations, as already indicated in Figs 7c, e. On the contralateral side (Fig. 7h), the cells of origin of these two pathways are mixed. There are apparently three

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Figure 4. (a) Low power microphotograph of the caudal mesencephalic tegmentum with an injection of AmCe. Strong retrograde labeling in the Pbg ipsilateral (to the right) and contralateral (to the left) to the injection side. In the midline the labeled neurons of the dorsal raphe nucleus (DRN) are seen, and on the side ipsilateral to the Am injection labeled neurons also in the deep mesencephalic nucleus (DpM) can be observed. Scale bar = 500 µm. (b) The caudal mesencephalic tegmentum from the case with injection focus illustrated in Fig. 3. The cells in Pbg are only lightly fluorescing when compared with the intense labeling of the DRN, DpM, and the dorsocaudal part of the retrorubral area (RR). Aq – cerebral aqueduct. Scale bar = 250 µm.



Figure 5. Serial sections through the rostrocaudal extent of Pbg with retrogradely labeled neurons following an AmCe injection. In the rostral part of Pbg, retrogradely labeled neurons are found both contralaterally (**a**) and ipsilaterally (**b**). In the central sector of Pbg, labeled neurons are also found contralaterally (**c**) and ipsilaterally (**e**). Near the caudal pole of Pbg, retrogradely labeled neurons are only present on the contralateral part (**d**). Scale bars: a, c, $d = 400 \mu m$; b, $e = 300 \mu m$.



Figure 6. Schematic diagrams showing cores (black) and halos (gray) of injections of Fluoro-Gold in the Am (a-c) and fluoro-emerald in the superior colliculus (**d-f**) (schematic frontal planes posterior to bregma, modified after Paxinos and Watson, 1998). B – basal nucleus of Meynert; BL – basolateral nucleus of Am; BSTIA – bed nucleus of stria terminalis, intraamygdaloid division; CeC – central nucleus of *Am, capsular part; CeL – central nucleus of Am, lateral part;* CeM – central nucleus of Am, medial part; DpG – deep gray *layer of superior colliculus; InWh – intermediate white layer* of superior colliculus; LaVL – lateral nucleus of Am, ventrolateral part; LaVM – lateral nucleus of Am, ventromedial part; Me - medial nucleus of Am; MeAD - medial nucleus of Am, anterodorsal part; Op – optic nerve layer of superior colliculus; Pir – piriform cortex; SuG – superficial gray layer of superior colliculus.



Figure 7. (a) Injection of Fluoro-Gold in the Am. The central necrotic zone involves the medial (CeM) and the capsular (CeC) subnuclei of the central nucleus of Am. BL - basolateral nucleus of Am; BM – basomedial nucleus of Am; IM - intercalated cell masses of Am. Scale $bar = 100 \ \mu m.$ (b) By means of four small injections of Fluoro-Emerald the superficial layers of the superior colliculus are regularly infiltrated. Scale bar = $100 \ \mu m.$ (c) Retrogradely labeled neurons in the central portion of Pbg, ipsilateral to the injection of Fluoro-Gold in the Am. Scale bar = $100 \ \mu m.$ (d) Retrogradely labeled neurons in the Pbg, contralateral to the injection of Fluoro-Gold in the Am. Scale bar = $100 \ \mu m.$ (e) Two groups (dorsal and ventral) of labeled neurons in Pbg following injection of Fluoro-Emerald in the ipsilateral superior colliculus. Scale bar = $100 \ \mu m.$ (f) Retrogradely labeled neurons in Pbg, contralateral to the injection of Fluoro-Emerald in the superior colliculus. Scale bar = $100 \ \mu m.$ (g*h*) Double labeling experiment with injections of Fluoro-Gold in Am and Fluoro-Emerald in the superior colliculus. (g) The dorsal neuronal group of the ipsilateral Pbg exhibits green fluorescence after a superior colliculus injection. The yellow fluorescence of Fluoro-Gold marks the central Pbg neurons projecting to Am. There are no double-labeled neurons in the Pbg ipsilateral to the injection. Scale bar = $100 \ \mu m.$ (h) In the contralateral Pbg neurons projecting only to Am (yellow) can be seen, quite a few neurons project only to superior colliculus (green), and there is a substantial number of double-labeled neurons (red). Scale bar = 100 µm.

populations: neurons that project only to the Am, a smaller number of cells that project only to the superior colliculus, and double-labeled neurons that simultaneously innervate Am and superior colliculus by means of divergent axon collaterals (indicated with red color in Fig. 7h). The crossed and uncrossed connections of Pbg to the central nucleus of Am and to the superior colliculus are summarized in Fig. 8.

The Fluoro-Emerald is not as effective as retrograde tracer as the Fluoro-Gold, yet it has a significant advantage: Fluoro-Emerald is transported also anterogradely. Axons arise from the injected superior colliculus that could be followed to the ipsilateral Pbg, where they form a very dense terminal field, in the central region, where the Pbg-Am cells are located.

Pbg emits bilateral projections to superior colliculus and Am, and also to the lateral geniculate body, which is bilateral in some species (133). The ipsilateral connection to the superior colliculus arises in the dorsal and ventral neuronal groups of Watanabe and Kawana (129) and the middle group projects to the Am. Exactly this group receives a dense terminal input from the superficial layers of the superior colliculus. Thus, there is a point-to-point multineuronal chain: superior colliculus - Pbg - Am. This characteristic labeling in Pbg was first demonstrated by Graybiel (127) in the cat (see her "band" on Fig. 6B in 127). Otherwise, the ipsilaterally projecting Pbg neurons in the cat have a broader distribution (127,130). The crossed connections are supplied by two neuronal types. Some cells project either to the Am or to the superior colliculus, and there are also neurons that simultaneously reciprocate the input from the superior colliculus, and project to the central nucleus of Am. Thus, a one and the same cell can project to visual and limbic structures.

The present results extend the previous observations that the superior colliculus conveys the retinal input to the Am via several extrageniculostriate diencephalic visual centers (Fig. 9). Despite its small size, the Pbg apparently has a strong contribution in the multineuronal subcortical pathways that transfer the retinal inputs to the Am, thus rapidly informing the "brain's lighthouse" about sources of threat before conscious detection (33,36,64,67,68,78,82).

Amygdala and subcortical vision in some neuropsychiatric diseases

As to be expected, the functions of the Am as a subcortical visual center are more or less altered by several neurological and psychiatric disorders. Although clinical and electrophysi-



Figure 8. Schematic representation of the crossed and uncrossed pathways from Pbg to the central nucleus of Am, and to the superior colliculus.



Figure 9. The retinal input to the superior colliculus is further transmitted to the Am via five disynaptic pathways: four of them – diencephalic, and probably the most important one – from the "satellite system of the superior colliculus", the Pbg.

ological evidence indicates that the Am plays an important role in the pathogenesis of temporal lobe epilepsy, there are few detailed data on histopathological changes of Am in epilepsy patients, in contrast to numerous investigations of experimental models. Wolf *et al* (146) examined the lateral nucleus of Am in 70 surgical specimens from patients with temporal lobe epilepsy and in 10 control specimens with respect to neuronal density and gliosis. They compared the results with the neuronal loss in the hippocampal formation. In epilepsy patients the neuronal density of the lateral nucleus of Am was significantly decreased as compared to normal controls. The mean volumetric density in epilepsy patients was reduced to 59% of that in normal individuals. The neuronal loss in the Am correlated well with the presence of fibrillary gliosis. Thus, the findings demonstrate that the Am is severely altered in most patients with temporal lobe epilepsy, and Wolf et al (146) suggest that these changes are independent of those in hippocampus. Pitkänen et al (147) compared the data on the Am damage in experimental and human temporal lobe epilepsy. The MRI studies of epileptic patients have shown that volume reduction of the Am ranges from 10 to 30%. In the human Am, neuronal loss and gliosis have been reported in lateral and basal nuclei. As to be expected, Pitkänen et al (147) encountered heavier alterations in rats with experimental status epilepticus. Recently, Daley et al (148) compared the Am volume in children with cryptogenic epilepsy who have complex partial seizures with that of age- and gender-matched normal children. There were no significant differences in the Am volume between epileptic and healthy children. However, within the group with complex partial seizures, the children with an affective/anxiety disorder had significantly larger left Am volumes, as well as greater Am asymmetry, compared with those with no psychopathology. Meletti et al (149) studied the ability of facial emotion recognition in patients with symptomatic epilepsy, evaluating whether medial temporal lobe damage is related to impairment in the recognition of specific emotions. They established that early-onset rightsided medial temporal lobe epilepsy is the key substrate determining a severe deficit in recognizing emotional facial expressions, especially fear. Yamada et al (150) examined the emotion recognition from facial expressions in a temporal lobe epileptic patient with ictal fear (affective aura). By this form of temporal lobe epilepsy the patients exhibit distinctly smaller Am volume (151). The patient of Yamada et al (150) underwent hippocampectomy which completely suppressed ictal fear. Before surgery, the patient tended to attach enhanced fear, sadness, and anger to various facial expressions. After surgery, such biases disappeared.

Schizophrenia was commonly regarded as a "functional psychosis", the implication being that the delusions, hallucinations and cognitive impairment characteristics of the disease have no organic basis. This view was due in no small way to the failure of neuropathologists to find convincing pathological changes associated with the disease in the first seven decades of the XXth century. Several studies of members of the school of Cecile and Oscar Vogt were neglected and forgotten (152,153). The first unequivocal evidences were summarized by Roberts and Bruton (152). Notably, already the first encouraging studies pointed out that the medial temporal lobe structures (parahippocampal gyrus, hippocampus and Am) are preferentially affected. These data were repeatedly confirmed (153-157; to cite only a few). According to Shenton et al (155) the brain abnormalities by schizophrenia are subtle and not all findings of certain teams are confirmed by other researchers. However, from 193 MRI studies, 100% of reviewed reports point out to changes in the Am. Recently, concrete and unequivocal data were presented by Kreczmanski et al (157). They compared several regions in brains of schizophrenics and controls and found that there is a reduced volume in the lateral nucleus of Am, and it is asymmetric: 12.1% on the left side and 17.6% on the right side. In conjunction, there is also a reduced mean total neuron number: 15.9% on the left side, and 16.2% on the right side. Already from classical clinical observations, it is generally agreed that schizophrenia patients show a markedly reduced ability to perceive and express facial emotions and they have difficulties in interpreting social information gathered from facial expression (158-160; and references therein). Several functional investigations of schizophrenic patients showed some alterations of the Am activity (80,161-167) but the results are not completely straightforward. Phillips et al (161) were the first to compare the responses to facial expressions in paranoid and non-paranoid patients. All patients were less accurate in identifying expressions, and showed less activation to these stimuli than normal subjects. Non-paranoids categorized disgust as either anger or fear more frequently than paranoids, and demonstrated in response to disgust expressions activation of the Am, that is associated with perception of fearful faces. Paranoids were more accurate in recognizing expressions, and demonstrated greater activation than non-paranoids to most stimuli. Paradiso et al (162) carried out a regional cerebral blood flow investigation by untreated schizophrenics and controls when viewing pleasant or unpleasant images. When patients evaluated the unpleasant images, they did not activate the fear-danger recognition circuit (e.g. the Am) used by healthy volunteers. Holt et al (163) carried out a functional MRI investigation in schizophrenic patients and healthy controls that passively viewed human faces displaying fearful, happy, and neutral emotional expressions. Relative to control subjects, the patients demonstrated significantly greater activation of the left hippocampus while observing all three facial expressions, and increased right Am activation during the initial presentation of fearful and neutral facial expressions. According to Das et al (80), in response to fearful facial expressions, schizophrenia patients displayed reduced Am activity, compared to controls. Williams et al (164) pointed out that schizophrenia patients show a disconnection

in Am-medial prefrontal cortex and autonomic arousal systems for processing fear and signals of danger. Comparing paranoid and nonparanoid patients, Williams et al (164) noticed that this disjunction is most apparent in patients with a profile of paranoia, coupled with poor social function and insight. Gur et al (166) encountered that patients showed reduced limbic activation compared with controls for the emotion discrimination task. Whereas in controls greater Am activation was associated with correct identifications of threat-related (anger and fear) expressions, patients showed the opposite effect of greater limbic activation, portending misidentifications. Furthermore, greater Am activation to the presentation of fearful faces was highly correlated with greater severity of so-called "flat effect". Finally, according to Hall et al (167), patients with schizophrenia showed a relative decrease in Am activation to fearful faces, compared with neutral faces. However, this difference resulted from an increase in Am activation to the neutral faces in schizophrenic patients, not from a decreased response to the fearful faces. Thus, Hall et al (167) think that the inappropriate activation of neural systems involved in fear to otherwise neutral stimuli may contribute to the development of psychotic symptoms in schizophrenia.

Amygdala is involved in the neuropathology of autism (168), and the time course of brain development rather the final product is most disturbed (reviewed in 169). Shumann et al (170) checked the increase of Am volume in normally developing children and in children with autism. Interestingly, children with autism rapidly increase the Am volume (7.5-12.5 years of age), and it is initially larger than by normally developing children that exhibit increase up to 18.5 years. Thus, there is an abnormal program of early Am development in autism. In conjunction, Munson et al (171) encountered that larger right Am volume was associated with more severe impairments at ages 3 and 4 years, and was predictive of poorer social and communication abilities at age of 6 years. Lack of reciprocal eye contact is an early and striking manifestation of autism (reviewed in 88), and the increased Am activity by healthy persons while observing eyes is reviewed above. FMRI studies in adults have found abnormally low activation in the Am among persons with autism relative to controls for tasks involving emotion recognition (172,173). By adults with high-functioning autism (Asperger syndrome) the function of Am is also altered (174). Aschwin et al (175) observed that patients with autism spectrum conditions were less accurate on the emotion recognition task compared to controls, but only for the negative basic emotions. Notably, these authors

discuss their data in the light of similar findings from people with damage to the Am.

Recent data pointed out that by bipolar disorder abnormalities in prefrontal cortex, striatum and Am exist early in the course of the illness and predate illness onset (176). Am dysfunction during processing of facial expressions was encountered already in children with bipolar disorder (177). Compared with controls, patients perceived greater hostility in neutral faces and reported more fear when viewing them. Interestingly, patients had greater activation in the left Am, nucleus accumbens, putamen and ventral prefrontal cortex when rating face hostility, and greater activation in the left Am and bilateral accumbens when rating their fear of the face. Malhi et al (178) investigated euthymic bipolar disorder patients by emotion recognition task involving fear, disgust and neutral expressions. Patients were equally accurate in identifying facial expressions as healthy subjects but were slower to respond, especially with respect to fear and disgust. Responses to fear and disgust resulted in activation of Am and insula, respectively. Characteristically, euthymic bipolar patients responded largely to fear and healthy subjects responded more so to disgust.

Only recently it became apparent that by posttraumatic stress disorder (PTSD) the function of Am (including emotion recognition) is significantly altered (179-183). Shin et al (179) examined Vietnam male combat veterans and female nurse veterans with PTSD, studying the regional blood flow during the recollection of personal traumatic and neutral events. All veterans exhibited a decreased cerebral blood flow in the prefrontal cortex but only male veterans showed an increase in the left amygdala. Further, Shin et al (180,181) compared patients with PTSD with trauma-exposed men without this disorder. The PTSD group exhibited exaggerated Am responses and diminished medial prefrontal cortex to fearful vs. happy facial expressions. Comparable results were obtained by Bryant et al (183). They examined the responses to nonconscious processing of fear by PTSD patients. These patients display heightened Am activity but decreased medial prefrontal activity. However Bryant et al (183) established increased both Am and medial prefrontal cortex activity during nonconscious processing of fearful faces.

CONCLUSION

Until recently, it has been believed that the visual information is transferred to the Am by a multineuronal chain involving the superior colliculus and a variety of thalamic nuclei. Here we propose the existence of a putative alternative pathway, e.g. a second extrageniculostriate multisynaptic connection to Am: retina – superior colliculus – parabigeminal nucleus – Am. This pathway might be very effective since all tracts listed above are bilateral. It seems that the function of the Am by the rapid response to the sources of threat before conscious detection is significantly altered by various neuropsychiatric diseases.

REFERENCES

- Brockhaus H. Zur normalen und pathologischen Anatomie des Mandelkerngebietes. *J Psychol Neurol* 1938; 49: 1-136.
- 2. Brodal A. The amygdaloid nucleus of the rat. *J Comp Neurol* 1947; 87: 1-16.
- Koikegami H. Amygdala and other related structures. Experimental studies on the anatomy and function. I. Anatomical researches with some neurophysiological observations. *Acta Med Biol (Niigata)* 1963; 10: 161-277.
- Hall E. Some aspects of the structural organization of the amygdala. In: Eleftheriou B, editor. *The Neurobiology of the Amygdala* 1972; Plenum Press, New York, 95-121.
- Krettek JE, Price JL. A description of the amygdaloid complex in the rat and cat with observations on intraamygdaloid axonal connections. *J Comp Neurol* 1978; 178: 255-280.
- Price JL, Russchen FT, Amaral DG. The limbic region: II: The amygdaloid complex. In: Björklund A, Hökfelt T, Swanson LW, editors. *Handbook of chemical neuroanatomy, Vol 5, Integrated systems of the CNS, Part I.* Elsevier, Amsterdam, 1987; 289-381.
- de Olmos JS. Amygdala. In: Paxinos G, editor. *The human nervous system*. Academic Press, San Diego, 1990; 583-710.
- Amaral DG, Price JL, Pitkänen A, Carmichael ST. Anatomical organization of the primate amygdaloid complex. In: Aggleton JP, editor. *The Amygdala: Neurobiological aspects of Emotion, Memory, and Mental Dysfunction.* Wiley-Liss, New York, 1992; 1-66.
- 9. Swanson LW. *Brain Maps: Structure of the Rat Brain*. Elsevier, Amsterdam, 1992.
- 10. Asan E. The catecholaminergic innervation of the rat amygdala. *Adv Anat Embryol Cell Biol* 1998; 142: 1-118.
- 11. McDonald AJ. Cortical pathways to the mammalian amygdala. *Prog Neurobiol* 1998; 55: 257-332.
- Swanson LW, Petrovich GD. What is the amygdala? Trends Neurosci 1998; 21: 323-331.
- 13. Heimer L, de Olmos JS, Alheid GF, Pearson J, Sakamoto

N, Shinoda K *et al.* The human basal forebrain. Part II. In: Bloom FE, Björklund A, Hökfelt T, editors. *Handbook of Chemical Neuroanatomy*, Vol 15, *The Primate Nervous System, Part III.* Elsevier, Amsterdam, 1999; 57-226.

- Paxinos G, Kus L, Ashwell KWS, Watson C. Chemoarchitectonic Atlas of the Rat Forebrain. Academic Press, San Diego, 1999.
- Aggleton JP, Saunders RC. The amygdala what's happened in the last decade? In: Aggleton JP, editor. *The Amygdala. A Functional Analysis.* 2nd edition. Oxford University Press, Oxford, 2000; 1-30.
- Kemppainen S, Pitkänen A. Distribution of parvalbumin, calretinin, and calbindin-D(28k) immunoreactivity in the rat amygdaloid complex and colocalization with gammaaminobutyric acid. *J Comp Neurol* 2000; 426: 441-467.
- Pitkänen A. Connectivity of the rat amygdaloid complex. In: Aggleton JP, editor. *The Amygdala: A Functional Analysis*, 2nd edition. Oxford University Press, Oxford, 2000; 31-115.
- 18. Price JL. Comparative aspects of amygdala connectivity. *Ann NY Acad Sci* 2003; 985: 50-58.
- Sah P, Faber ES, Lopez De Armentia M, Power J. The amygdaloid complex: anatomy and physiology. *Physiol Rev* 2003; 83: 803-834.
- de Olmos JS, Beltramino CA, Alheid G. Amygdala and extended amygdala of the rat: a cytoarchitectonical, fibroarchitectonical, and chemoarchitectonical survey. In: Paxinos G, editor. *The Rat Nervous System*, 3rd edition. Academic Press, San Diego, 2004; 509-603.
- Schumann CM, Amaral DG. Stereological estimation of the number of neurons in the human amygdaloid complex. *J Comp Neurol* 2005; 491: 320-329.
- 22. Usunoff KG, Itzev DE, Rolfs A, Schmitt O, Wree A. Brain stem afferent connections of the amygdala in the rat with special references to a projection from the parabigeminal nucleus: a fluorescent retrograde tracing study. *Anat Embryol (Berl)* 2006; 211: 475-496.
- 23. Kaada BR. Stimulation and regional ablation of amygdaloid complex with reference to functional representations. In: Eleftherious BE, editor. *The Neurobiology of the Amygdala*. Plenum Press, New York, 1972; 205-281.
- 24. Ben-Ari Y (editor). *The Amygdaloid Complex*. Elsevier, Amsterdam, 1981.
- 25. Cechetto DF. Central representation of visceral function. *Fed Proc* 1987; 46: 17-23.
- 26. Cahill L, Babinsky R, Markowitsch H, McGaugh JL. The amygdala and emotional memory. *Nature* 1995; 377:

295-296.

- McGaugh JL, Cahill L, Roozendaal B. Involvement of the amygdala in memory storage: interaction with other brain systems. *Proc Natl Acad Sci USA* 1996; 93: 13508-13514.
- Gorman JM, Kent JM, Sullivan GM, Coplan JD. Neuroanatomical hypothesis of panic disorder, revised. *Am J Psychiatry* 2000; 157: 493-505.
- 29. LeDoux JE. Emotion circuits in the brain. *Annu Rev Neurosci* 2000a; 23: 155-184.
- LeDoux JE. The amygdala and emotion: a view through fear. In: Aggleton JP, editor. *The Amygdala. A Functional Analysis*, 2nd edition. Oxford University Press, Oxford, 2000b; 289-310.
- 31. Rasia-Filho AA,Londero RG,Achaval M, Functional activities of the amygdala: an overview. *J Psychiatry Neurosci* 2000; 25: 14-23.
- 32. Rolls ET. Memory systems in the brain. *Ann Rev Psychol* 2000; 51: 599-630.
- Davis M, Whalen PJ. The amygdala: vigilance and emotion. *Mol Psychiatry* 2001; 6: 13-34.
- 34. Amaral DG. The amygdala, social behavior, and danger detection. *Ann NY Acad Sci* 2003; 1000: 337-347.
- 35. Petrovich GD, Callager M. Amygdala subsystems and control of feeding behavior by learned cues. *Ann N Y Sci* 2003; 985: 251-262.
- 36. Zald DH. The human amygdala and the evaluation of sensory stimuli. *Brain Res Rev* 2003; 41: 88-123.
- 37. Neugebauer V, Li W, Bird GC, Hans JS. The amygdala and persistent pain. *Neuroscientist* 2004; 10: 221-234.
- Yaniv D, Desmedt A, Jaffard R, Richter-Levin G. The amygdala and appraisal processes: stimulus and response complexity as an organizing factor. *Brain Res Rev* 2004; 44: 179-186.
- 39. Maren S. Building and burying fear memories in the brain. *Neuroscientist* 2005; 11: 89-99.
- 40. Peper M, Herpers M, Spreer J, Hennig J, Zentner J. Functional neuroimaging of emotional learning and autonomic reactions. *J Physiol (Paris)* 2006; 99: 342-354.
- 41. Usunoff KG, Popratiloff A, Schmitt O, Wree A. Functional neuroanatomy of pain. *Adv Anat Embryol Cell Biol* 2006; 184: I-IX, 1-119.
- 42. Hull EM, Dominguez JM. Sexual behavior in male rodents. *Horm Behav* 2007; 52: 45-55.
- 43. Petrovich GD, Gallagher M. Control of food consumption by learned cues: a forebrain-hypothalamic network. *Physiol Behav* 2007; 91: 397-403.

- 44. Talarovicova A, Krskova L, Kiss A. Some assessments of the amygdala role in suprahypothalamic neuroendocrine regulation: a minireview. *Endocr Regul* 2007; 41: 155-162.
- 45. Haas BW, Canli T. Emotional memory function, personality structure and psychopathology: a neural system approach to the identification of vulnerability markers. *Brain Res Rev* 2008; 58: 71-84.
- 46. Lanuza E, Novejarque A, Martinez-Ricos J, Martinez-Hernandez J, Augustin-Pavon C, Martinez-Garcia F. Sexual pheromones and the evolution of the reward system of the brain: the chemosensory function of the amygdala. *Brain Res Bull* 2008; 75: 460-466.
- Adolphs R, Tranel D, Damasio H, Damasio A. Impaired recognition of emotion in facial expressions following bilateral damage to the human amygdala. *Nature* 1994; 372: 669-672.
- 48. Adolphs R, Tranel D, Damasio H, Damasio AR. Fear and the human amygdala. *J Neurosci* 1995; 15: 5879-5891.
- 49. Young AW, Aggleton JP, Hellawell DJ, Johnson M, Broks P, Hanley JR. Face processing impairment after amygdalectomy. *Brain* 1995; 118: 15-24.
- Young AW, Hellawell DJ, van de Wal C., Johnson M. Facial expression processing after amygdalotomy. *Neuropsychologia* 1996; 34: 31-39.
- 51. Hofer P-A. Urbach-Wiethe disease: a review. *Acta Derm Venerol* 1973; 53: 5-52.
- 52. Parent A. *Carpenter's Human Neuroanatomy*, 9th edition. Baltimore, Williams and Wilkins, 1996.
- Mehler WR. Subcortical afferent connections of the amygdala in the monkey. *J Comp Neurology* 1980; 190: 733-762.
- Ottersen OP. Afferent connections to the amygdaloid complex of the rat with some observations in the cat. III. Afferents from the lower brain stem. *J Comp Neurol* 1981; 202: 335-356.
- Russchen FT. Amygdalopetal projections in the cat. II. Subcortical afferent connections. A study with retrograde tracing techniques. *J Comp Neurol* 1982; 207: 157-176.
- Volz HP, Rehbein G, Triepel J, Knuepfer MM, Stumpf H, Stock G. Afferent connections of the nucleus centralis amygdalae. A horseradish peroxidase study and literature survey. *Anat Embryol (Berl)* 1990; 181: 177-194.
- Canteras NS, Simerly RB, Swanson LW. Connections of the posterior nucleus of the amygdala. *J Comp Neurol* 1992; 324: 143-179.
- 58. Shammah-Lagnado SJ, Alheid GF, Heimer L. Afferent

connections of the interstitial nucleus of the posterior limb of the anterior commissure and adjacent amygdalostriatal transition area in the rat. *Neuroscience* 1999; 94: 1097-1123.

- Herzog AG, Van Hoesen GW. Temporal neocortical afferent connections to the amygdala in the rhesus monkey. *Brain Res* 1976; 115: 57-69.
- Shi C, Davis M. Visual pathways involved in fear conditioning measured with fear-potentiated startle: behavioral and anatomic studies. *J Neurosci* 2001; 21: 9844-9855.
- Breitner HC, Etcoff NL, Whalen PJ, Kennedy WA, Rauch SL, Strauss MM *et al.* Response and habituation of the human amygdala during visual processing of facial expression. *Neuron* 1996; 17: 875-887.
- 62. Morris JS, Frith CD, Perrett DI, Rowland D, Young AW, Calder AJ *et al.* A differential neural response in the human amygdala to fearful and happy facial expressions. *Nature* 1996; 383: 812-815.
- 63. Morris JS, Friston KJ, Buchel C, Frith CD, Young AW, Calder AJ *et al.* A neuromodulatory role for the human amygdala in processing emotional facial expressions. *Brain* 1998; 121: 47-57.
- Whalen PJ, Rauch SL, Etcoff NL, McInerney SC, Lee MB, Jenike MA. Masked presentations of emotional facial expressions modulate amygdala activity without explicit knowledge. *J Neurosci* 1998; 18: 411-418.
- Blair RJ, Morris JS, Frith CD, Perrett DI, Dolan RJ. Dissociable neural responses to facial expressions of sadness and anger. *Brain* 1999; 122: 883-893.
- 66. Morris JS, Ohman A, Dolan RJ. A subcortical pathway to the right amygdala mediating "unseen" fear. *Proc Natl Acad Sci USA* 1999; 96: 1680-1685.
- Morris JS, DeGelder B, Weiskrantz L, Dolan RJ. Differential extrageniculostriate and amygdala responses to presentation of emotional faces in a cortically blind field. *Brain* 2001; 124: 1241-1252.
- Adolphs R, Tranel D. Emotion, recognition, and the human amygdala. In: Aggleton JP, editor. *The amygdala*. *A functional analysis*. 2nd edition. Oxford University Press, Oxford, 2000; 587-630.
- 69. Adolphs R, Tranel D. Impaired judgment of sadness but not happiness following bilateral amygdala damage. J Cogn Neurosci 2004; 16: 453-462.
- Adolphs R, Tranel D, Damasio H. Emotion recognition from faces and prosody following temporal lobectomy. *Neuropsychology* 2001; 15: 396-404.
- 71. Iidaka T, Omori M, Murata T, Kosaka H, Yonekura Y,

Okada T *et al.* Neural interaction of the amygdala with the prefrontal and temporal cortices in the processing of facial expressions as revealed by fMRI. *J Cogn Neurosci* 2001; 13: 1035-1047.

- 72. Adolphs R. Recognizing emotion from facial expressions: psychological and neurological mechanisms. *Behav Cogn Neurosci Rev* 2002; 1: 21-62.
- Hariri AR, Tessitore A, Mattay VS, Fera F, Weinberger DR. The amygdala response to emotional stimuli: a comparison of faces and scenes. *Neuroimage* 2002; 17: 317-323.
- Morris JS, deBonis M, Dolan RJ. Human amygdala responses to fearful eyes. *Neuroimage* 2002; 17: 214-222.
- Oya H, Kawasaki H, Howard MA 3rd, Adolphs R. Electrophysiological responses in the human amygdala discriminate emotion categories of complex visual stimuli. *J Neurosci* 2002; 22: 9502-9512.
- Anderson AK, Christoff K, Panitz D, De Rose E, Gabrieli GD. Neural correlates of the automatic processing of threat facial signals. *J Neurosci* 2003; 23: 5627-5633.
- Kim H, Somerville LH, Johnstone T, Polis S, Alexander AL, Shin LM *et al.* Contextual modulation of amygdala responsivity to surprised faces. *J Cogn Neurosci* 2004; 16: 1730-1745.
- Williams MA, Morris AP, McGlone F, Abbott DF, Mattingley JB. Amygdala responses to fearful and happy facial expressions under conditions of binocular suppression. J Neurosci 2004; 24: 2898-2904.
- 79. Das P, Kemp AH, Liddell BJ, Brown KJ, Olivieri G, Peduto A *et al.* Pathways for fear perception: modulation of amygdala activity by thalamo-cortical systems. *Neuroimage* 2005; 26: 141-148.
- Das P, Kemp AH, Flynn G, Harris AW, Liddell BJ, Whitford TJ *et al.* Functional disconnections in the direct and indirect amygdala pathways for fear processing in schizophrenia. *Scizophr Res* 2007; 90: 284-294.
- Johnstone T, Somerville LH, Alexander AL, Oakes TR, Davidson RJ, Kalin NH *et al.* Stability of amygdala BOLD response to fearful faces over multiple scan sessions. *Neuroimage* 2005; 25: 1112-1123.
- Liddell BJ, Brown KJ, Kemp AH, Barton MJ, Das P, Peduto A *et al*. A direct brainstem-amygdala-cortical "alarm" system for subliminal signals of fear. *Neuroim-age* 2005; 24: 235-243.
- 83. Tabbert K, Stark R, Kirsch P, Vaitl D. Hemodynamic responses of the amygdala, the orbitofrontal cortex and the visual cortex during a fear conditioning paradigm. *Int*

J Psychophysiol 2005; 57: 15-23.

- Williams LM, Barton MJ, Kemp AH, Liddell BJ, Peduto A, Gordon E *et al.* Distinct amygdala-autonomic arousal profiles in response to fear signals in healthy males and females. *Neuroimage* 2005; 28: 618-626.
- Yoshimura N, Kawamura M, Masaika Y, Homma I. The amygdala of patients with Parkinson's disease is silent in response to fearful facial expressions. *Neuroscience* 2005; 131: 523-534.
- 86. Adolphs R. Fear, faces, and the human amygdala. *Curr Opin Neurobiol* 2008; 18: 166-172.
- Demos KE, Kelley WM, Ryan SL, Davis FC, Whalen PJ. Human amygdala sensitivity to the pupil size of others. *Cereb Cortex* 2008; 18: 2729-2734.
- Golarai G, Grill-Spector K, Reiss AL. Autism and development of face processing. *Clin Neurosci Res* 2006; 6: 145-160.
- Kawashima R, Sugiura M, Kato T, Nakamura A, Hatano K, Ito K *et al.* The human amygdala plays an important role in gaze monitoring. A PET study. *Brain* 1999; 122: 779-783.
- Whalen PJ, Kagan J, Cook RG, Davis FC, Kim H, Polis S *et al.* Human amygdala responsivity to masked fearful eye whites. *Science* 2004; 306: 2061.
- Adolphs R, Gosselin F, Buchanan TW, Tranel D, Schyns P, Damasio AR. A mechanism for impaired fear recognition after amygdala damage. *Nature* 2005; 433: 68-72.
- 92. Whalen PJ, Shin LM, McInerney SC, Fischer H, Wright CI, Rauch SL. A functional MRI study of human amygdala responses to facial expressions of fear versus anger. *Emotion* 2001; 1: 70-83.
- de Gelder B, Morris JS, Dolan RJ. Unconscious fear influences emotional awareness of faces and voices. *Proc Natl Acad Sci USA* 2005; 102: 18682-18687.
- Williams LM, Das P, Liddell BJ, Kemp AH, Rennie CJ, Gordon E. Mode of functional connectivity in amygdala pathways dissociates level of awareness for signals of fear. *J Neurosci* 2006; 26: 9264-9271.
- 95. Williams LM, Liddell BJ, Kemp AH, Bryant RA, Meares RA, Peduto AS *et al.* Amygdala-prefrontal dissociation of subliminal and supraliminal fear. *Hum Brain Mapp* 2006; 27: 652-661.
- 96. Benevento LA, Fallon JH. The ascending projections of the superior colliculus in the rhesus monkey (*Macaca mulatta*). J Comp Neurol 1975; 160: 339-362.
- 97. Jones EG, Burton H. A projection from the medial pulvinar to the amygdala in primates. *Brain Res* 1976; 104:

142-147.

- Grieve KL, Acuna C, Cudeiro J. The primate pulvinar nuclei: vision and action. *Trends Neurosci* 2000; 23: 35-39.
- 99. Linke R, De Lima AD, Schwegler H, Pape HC. Direct synaptic connections of axons from superior colliculus with identified thalamo-amygdaloid projection neurons in the rat: possible substrates of a subcortical visual pathway to the amygdala. *J Comp Neurol* 1999; 403: 158-170.
- 100. Aston-Jones G. Locus coeruleus, A5 and A7 noradrenergic cell groups. In: Paxinos G, editor. *The Rat Nervous System*, 3rd edition. San Diego, Elsevier Academic Press, 2004; 259-294.
- 101. Aston-Jones G, Rajkowski J, Kubiak P, Valentino RJ, Shipley MT. Role of the locus coeruleus in emotional Activation. *Prog Brain Res* 1996; 107: 379-402.
- 102. Usunoff KG, Schmitt O, Itzev DE, Rolfs A, Wree A. Efferent connections of the parabigeminal nucleus to the amygdala and the superior colliculus in the rat: a double-labeling fluorescent retrograde tracing study. *Brain Res* 2007; 1133: 87-91.
- 103. von Bechterew VM. *Die Leitungsbahnen in Gehirn und Ruckenmark*. Leipzig, Arthur Georgi, 1899.
- 104. Gallyas F, Hsu M, Buzsaki G. Four modified silver methods for thick sections of formaldehyde-fixed mammalian central nervous tissue: "dark" neurons, perikarya of all neurons, microglial cells and capillaries. *J Neurosci Methods* 1993; 50: 159-164.
- Tokunaga A, Otani K. Neuronal organization of the corpus parabigeminum in the rat. *Exp Neurol* 1978; 58: 361-375.
- 106. Baleydier C, Magnin M. Afferent and efferent connections of the parabigeminal nucleus in cat revealed by retrograde axonal transport of horseradish peroxidase. *Brain Res* 1979; 161: 187-198.
- 107. Wang SR. The nucleus isthmi and dual modulation of the receptive field of tectal neurons in non-mammals. *Brain Res Rev* 2003; 41: 13-25.
- 108. Dudkin EA, Sheffield JB, Gruberg ER. Combining visual information from the two eyes: the relationship between isthmotectal cells that project to ipsilateral and to contralateral optic tectum using fluorescent retrograde labels in the frog, *Rana pipiens*. *J Comp Neurol* 2007; 502: 38-54.
- Mufson EJ, Martin TL, Mash DC, Wainer BH, Mesulam MM. Cholinergic projections from the parabigeminal nucleus (Ch8) to the superior colliculus in the mouse: a

combined analysis of horseradish peroxidase transport and choline acetyltransferase immunohistochemistry. *Brain Res* 1986; 370: 144-148.

- Henderson Z. Cholinergic innervation of ferret visual system. *Neuroscience* 1987; 20: 503-518.
- 111. Jones BE, Beaudet A. Distribution of acetylcholine and catecholamine neurons in the cat brainstem: a choline acetyltransferase and tyrosine hydroxylase immunohistochemical study. *J Comp Neurol* 1987; 261: 15-32.
- 112. Vincent SR, Reiner PB. The immunohistochemical localization of choline acetyltransferase in the cat brain. *Brain Res Bull* 1987; 18: 371-415.
- 113. Fitzpatrick D, Conley M, Luppino G, Matelli M, Diamond IT. Cholinergic projections from the midbrain reticular formation and the parabigeminal nucleus to the lateral geniculate nucleus in the tree shrew. *J Comp Neurol* 1988; 272, 43-67.
- 114. Hall WC, Fitzpatrick D, Klatt LL, Raczkowski D. Cholinergic innervation of the superior colliculus in the cat. *J Comp Neurol* 1989; 287: 495-514.
- 115. Mesulam M-M, Geula C, Bothwell MA, Hersh LB. Human reticular formation: cholinergic neurons of the pedunculopontine and laterodorsal tegmental nuclei and some cytochemical comparisons to forebrain cholinergic neurons. *J Comp Neurol* 1989; 283: 611-633.
- 116. Usunoff KG. Cytoarchitectural, Ultrastructural and Histochemical Characterization of Substantia Nigra. DSc Thesis, Vols I-VI. Medical Academy, Sofia, 1990.
- 117. Bina KG, Rusak B, Semba K. Localization of cholinergic neurons in the forebrain and brainstem that project to the suprachiasmatic nucleus of the hypothalamus in rat. J Comp Neurol 1993; 335: 295-397.
- 118. Bickford ME, Ramcharan E, Godwin DW, Erisir A, Gnadt J, Sherman SM. Neurotransmitters contained in the subcortical extraretinal inputs to the monkey lateral geniculate nucleus. *J Comp Neurol* 2000; 424: 701-717.
- 119. Oda Y, Nakanishi T. The distribution of cholinergic neurons in the human central nervous system. *Histol Histopathol* 2000; 15: 825-834.
- 120. Usunoff KG, Lazarov NE, Itzev DE, Schmitt O, Rolfs A, Wree A. Cholinergic neuronal system of the amygdala in the rat. An immunocytochemical study. *Compt Rend Acad Bulg Sci* 2007; 60: 1215-1220.
- Motts SD, Slusarczyk AS, Sowick CS, Schofield BR. Distribution of cholinergic cells in guinea pig brainstem. *Neuroscience* 2008; 154: 186-195.
- 122. Manger PR, Fahringer HM, Pettigrew JD, Siegel JM.

The distribution and morphological characteristics of cholinergic cells in the brain of monotremes as revealed by ChAT immunohistochemistry. *Brain Behav Evol* 2002; 60: 275-297.

- 123. Maseko BC, Manger PR. Distribution and morphology of cholinergic, catecholaminergic and serotonergic neurons in the brain of Schreiber's long-fingered bat, *Miniopterus screibersii. J Chem Neuroanat* 2007; 34: 80-94.
- 124. van Noort J. *The Structure and Connections of the Inferior Colliculus*. Thesis. Van Gorcum, Assen, 1969.
- 125. Harting JK, Hall WC, Diamond IT, Martin GF. Anterograde degeneration study of the superior colliculus in tupaia glis: evidence for a subdivision between superficial and deep layers. *J Comp Neurol* 1973; 148: 361-386.
- 126. Harting JK. Descending pathways from the superior colliculus: an autoradiographic analysis in the rhesus monkey (*Macaca mulatta*). J Comp Neurol 1977; 583-612.
- 127. Graybiel AM. A satellite system of the superior colliculus: the parabigeminal nucleus and its projections to the superficial collicular layers. *Brain Res* 1978; 145: 365-374.
- 128. Sherk H. A comparison of visual-response properties in cat's parabigeminal nucleus and superior colliculus. J Neurophysiol 1979; 42: 1640-1655.
- 129. Watanabe K, Kawana E. Efferent projections of the parabigeminal nucleus in rats: a horseradish peroxidase (HRP) study. *Brain Res* 1979; 168: 1-11.
- Roldan M, Reinoso-Suarez F, Tortelly A. Parabigeminal projections to the superior colliculus in the cat. *Brain Res* 1983; 280: 1-13.
- 131. Hashikawa T, Van Lishout D, Harting JK. Projections from the parabigeminal nucleus to the dorsal lateral geniculate nucleus in the tree shrew *Tupaia glis*. *J Comp Neurol* 1986; 246: 382-394.
- 132. De Lima AD, Singer W. The brainstem projection to the lateral geniculate nucleus in the cat: identification of cholinergic and monoaminergic elements. *J Comp Neurol* 1987; 259: 92-121.
- 133. Harting JK, Van Lieshout DP, Hashikawa T, Weber JT. The parabigeminogeniculate projection: connectional studies in eight mammals. *J Comp Neurol* 1991; 305: 559-581.
- 134. Diamond IT, Fitzpatrick D, Conley M. A projection from the parabigeminal nucleus to the pulvinar nucleus in Galago. *J Comp Neurol* 1992; 316: 375-382.
- 135. Jeon CJ, Spencer RF, Mize RR. Organization and syn-

aptic connections of cholinergic fibers in the cat superior colliculus. *J Comp Neurol* 1993; 333: 363-380.

- 136. Feig S, Harting JK. Ultrastructural studies of the primate lateral geniculate nucleus: morphology and spatial relationships of axon terminals arising from the retina, visual cortex (area 17), superior colliculus, parabigeminal nucleus, and pretectum of *Galago crassicaudatus*. J Comp Neurol 1994; 343: 17-34.
- 137. Jiang ZD, King AJ, Moore DR. Topographic organization of projection from the parabigeminal nucleus to the superior colliculus in the ferret revealed with fluorescent latex microspheres. *Brain Res* 1996; 743: 217-232.
- 138. Künzle H. Connections of the superior colliculus with the tegmentum and the cerebellum in the hedgehog tenrec. *Neurosci Res* 1997; 28: 127-145.
- Lee PH, Schmidt M, Hall WC. Excitatory and inhibitory circuitry in the superficial gray layer of the superior colliculus. *J Neurosci* 2001; 21: 8145-8153.
- 140. Cui H, Malpeli JG. Activity in the parabigeminal nucleus during eye movements directed at moving and stationary targets. *J Neurophysiol* 2003; 89: 3128-3142.
- 141. Klop EM, Mouton LJ, Holstege G. Periparabigeminal and adjoining mesencephalic tegmental field projections to the dorsolateral periaqueductal grey in cat – a possible role for oculomotor input in the defensive system. *Eur J Neurosci* 2006; 23 2145-2157.
- 142. Paxinos G, Törk I, Halliday G, Mehler WR. Human homologs to brainstem nuclei identified in other animals as revealed by acetylcholinesterase activity. In: Paxinos G, editor. *The human nervous system*. Academic Press, San Diego, 1990; 149-202.
- 143. Saper CB. Cholinergic systems. In: Paxinos G, editor. *The Human Nervous System*. Academic Press, San Diego, 1990; 1095-1113.
- 144. Olszewski J, Baxter D. *Cytoarchitecture of the Human Brain Stem.* Karger, Basel, 1954.
- 145. Paxinos G, Huang XF. *Atlas of the Human Brainstem*. Academic Press, San Diego, 1995.
- 146. Wolf HK, Aliashkevich AF, Blumcke I, Wiestler OD, Zentner J. Neuronal loss and gliosis of the amygdaloid nucleus in temporal lobe epilepsy. A quantitative analysis of 70 surgical specimens. *Acta Neuropathol* 1997; 93: 606-610.
- 147. Pitkänen A, Tuunanen J, Kalviainen R, Partanen K, Salmenpera T. Amygdala damage in experimental and human temporal lobe epilepsy. *Epilepsy Res* 1998; 32: 233-253.

- 148. Daley M, Siddarth P, Levitt J, Gurbani S, Schields WD, Sankar R *et al.* Amygdala volume and psychopathology in childhood complex partial seizures. *Epilepsy Behav* 2008; 13: 212-217.
- 149. Meletti S, Benuzzi F, Rubboli G, Cantalupo G, Stanzani Maserati M, Niuchelli P *et al.* Impaired facial emotion recognition in early-onset right medial temporal lobe epilepsy. *Neurology* 2003; 60: 426-431.
- 150. Yamada M, Murai T, Sato W, Namiki C, Miyamoto T, Ohigashi Y. Emotion recognition from facial expressions in a temporal lobe epileptic patient with ictal fear. *Neuropsychologia* 2005; 43: 434-441.
- 151. Cendes F, Andermann F, Gloor P, Gambardella A, Lopes-Cendes I, Watson C *et al.* Relationship between atrophy of the amygdala and ictal fear in temporal lobe epilepsy. *Brain* 1994; 117: 739-746.
- 152. Roberts GW, Bruton CJ. Notes from the graveyard: neuropathology and schizophrenia. *Neuropathol Appl Neurobiol* 1990; 16: 3-16.
- 153. Bogerts B. Recent advances in the neuropathology of schizophrenia. *Schizophr Bull*, 1993; 19: 431-445.
- 154. Altschuler LL, Bartzokis G, Grieder T, Curran J, Jimenez T, Leight K *et al.* An MRI study of temporal lobe structures in men with bipolar disorder or schizophrenia. *Biol Psychiatry* 2000; 48: 147-162.
- 155. Shenton ME, Dickey CC, Frumin M, McCarley RW. A review of MRI findings in schizophrenia. *Schizophr Res* 2001; 49: 1-52.
- 156. Kucharska-Pietura K, Russell T, Masiak M. Perception of negative affect in schizophrenia – functional and structural changes in the amygdala. *Ann Univ Mariae Curie Sklodowska (Med)* 2003: 58: 453-458.
- 157. Kreczmanski P, Heinsen H, Mantua V, Woltersdorf F, Masson T, Ulfig N *et al.* Volume, neuron density and total neuron number in five subcortical regions in schizophrenia. *Brain* 2007; 130: 678-692.
- 158. Mandal MK, Pandey R, Prasad AB. Facial expressions of emotions and schizophrenia: a review. *Schizophr Bull* 1998;24:399-412.
- 159. Leitman DI, Loughead J, Wolf DH, Ruparel K, Kohler CG, Elliott MA *et al.* Abnormal superior temporal connectivity during fear perception in schizophrenia. *Schizophr Bull* 2008; 34: 673-678.
- Marwick K, Hall J. Social cognition in schizophrenia: a review of face processing. *Br Med Bull* 2008, doi:10.1093/bmb/ldn035.
- 161. Phillips ML, Williams L, Senior C, Bullmore ET, Bram-

mer MJ, Andrew C *et al.* A differential neural response to threatening and non-threatening negative facial expressions in paranoid and non-paranoid schizophrenics. *Psychiatry Res* 1999; 92: 11-31.

- 162. Paradiso S, Andreasen NC, Crespo-Facorro B, O'Leary DS, Watkins GL *et al.* Emotions in unmedicated patients with schizophrenia during evaluation with positron emission tomography. *Am J Psychiatry* 2003; 160: 1775-1783.
- 163. Holt DJ, Kunkel L, Weiss AP, Goff DC, Wright CI, Shin LM *et al.* Increased medial temporal lobe activation during the passive viewing of emotional and neutral facial expressions in schizophrenia. *Scizophr Res* 2006; 82: 153-162.
- 164. Williams LM, Das P, Liddell BJ, Olivieri G, Peduto AS, David AS *et al.* fronto-limbic and autonomic disjunctions to negative emotion distinguish schizophrenia subtypes. *Psychiatry Res* 2007; 155: 29-44.
- 165. Gur RE, McGrath C, Chan RM, Scroeder L, Turner T, Turetsky BI *et al.* An fMRI study of facial emotion processing in patients with schizophrenia. *Am J Psychiatry* 2002; 159: 1992-1999.
- 166. Gur RE, Loughead J, Kohler CG, Elliott MA, Lesko K, Ruparel K *et al.* Limbic activation associated with misidentification of fearful faces and flat affect in schizophrenia. *Arch Gen Psychiatry* 2007; 64: 1356-1366.
- Hall J, Whalley HC, McKirdy JW, Romaniuk L, McGonigle D, McIntosh AM *et al*. Overactivation of fear systems to neutral faces in schizophrenia. *Biol Psychiatry* 2008; 64: 70-73.
- Kemper TL, Bauman ML (1993) The contribution of neuropathologic studies to the understanding of autism. *Neurol Clin* 1993; 11: 175-187.
- Amaral DG, Schumann CM, Nordahl CW. Neuroanatomy of autism. *Trends Neurosci* 2008; 31: 137-145.
- 170. Schumann CM, Hamstra J, Goodlin-Jones BL, Lotspeich LJ, Kwon H, Buonocore MH *et al.* The amygdala is enlarged in children but not adolescents with autism; the hippocampus is enlarged at all ages. *J Neurosci* 2004; 24: 6392-6401.
- 171. Munson J, Dawson G, Abbott R, Faja S, Webb SJ, FriedmanSD *et al.* Amygdala volume and behavioral development in autism. *Arch Gen Psychiatry* 2006; 63: 686-693.
- 172. Baron-Cohen S, Ring HA, Bullmore ET, Wheelwright S, Aschwin C, Williams SC. The amygdala theory of autism. *Neurosci Biobehav Rev* 2000; 24: 355-364.
- 173. Critchley HD, Daly EM, Bullmore ET, Williams SC,

Van Amelsvoort T, Robertson DM *et al.* The functional neuroanatomy of social behaviour: changes in cerebral blood flow when people with autistic disorder process facial expressions. *Brain* 2000; 123: 2203-2212.

- 174. Ashwin C, Baron-Cohen S, Wheelwright S, O'Riordan M, Bullmore ET. Differential activation of the amygdala and the "social brain" during fearful face-processing in Asperger Syndrome. *Neuropsychologia* 2007; 45: 2-14.
- 175. Aschwin C, Chapman E, Colle L, Baron-Cohen S. Impaired recognition of negative basic emotions in autism: a test of the amygdala theory. *Soc Neurosci* 2006; 1: 349-363.
- 176. Strakowski SM, Delbello MP, Adler CM. The functional neuroanatomy of bipolar disorder: a review of neuroimaging findings. *Mol Psychiatry* 2005; 10: 105-116.
- 177. Rich BA, Vinton DT, Roberson-Nay R, Hommer RE, Berghorst LH, McClure EB *et al.* Limbic hyperactivation during processing of neutral facial expressions in children with bipolar disorder. *Proc Natl Acad Sci USA* 2006; 193: 8900-8905.
- 178. Malhi GS, Lagopoulos J, Sachdev PS, Ivanovski B, Schnier R, Ketter T. Is a lack of disgust something to fear? A functional magnetic resonance imaging facial emotion recognition study in euthymic bipolar disorder patients. *Bipolar Disord* 2007; 9: 345-357.
- 179. Shin LM, Orr SP, Carson MA, Rauch SL, Macklin ML, Lasko NB *et al.* Regional cerebral blood flow in the amygdala and medial prefrontal cortex during traumatic imagery in male and female Vietnam veterans with PTSD. *Arch Gen Psychiatry* 2004; 61: 168-176.
- 180. Shin LM, Wright CI, Cannistraro PA, Wedig MM, Mc-Mullin K, Martis B *et al.* A functional magnetic resonance imaging study of amygdala and medial prefrontal cortex responses to overtly presented fearful faces in posttraumatic stress disorder. *Arch Gen Psychiatry* 2005; 62: 273-281.
- 181. Shin LM, Rauch SL, Pitman RK. Amygdala, medial prefrontal cortex, and hippocampal function in PTSD. *Ann NY Acad Sci* 2006; 1071: 67-79.
- 182. Williams LM, Kemp AH, Felmingham K, Barton M, Olivieri G, Peduto A *et al*. Trauma modulates amygdala and medial prefrontal responses to consciously attended fear. *Neuroimage* 2006c; 29: 347-357.
- 183. Bryant RA, Kemp AH, Felmingham KL, Liddell B, Olivieri G, Peduto A *et al*. Enhanced amygdala and medial prefrontal activation during nonconscious processing of fear in posttraumatic stress disorder: an fMRI study. *Hum Brain Mapp* 2008; 29: 517-523.