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1 **Journal of Applied Phycology**

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3 **Optimization of photosynthesis, growth, and biochemical composition of the microalgae**
4 ***Rhodomonas salina* – an established diet for live feed copepods in aquaculture**

5

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20

21

22 **Abstract**

23 The Cryptophyte *Rhodomonas salina* is widely used as feed for copepod cultures. However, the
24 culturing conditions to obtain high quality algae have not yet been efficiently optimized. Therefore,
25 we aimed to develop a cultivation protocol for *R. salina* to optimize its nutritional value and provide
26 technical recommendations for later large scale production in algal photobioreactors. We studied
27 photosynthesis, growth, pigments, fatty acids (FA) and free amino acids (FAA) composition of *R.*
28 *salina* cultured at different irradiances (10-300 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) and nutrient availability
29 (deficiency and excess). The optimal range of irradiance for photosynthesis and growth was 60-100
30 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. The content of chlorophylls *a* and *c* decreased with increasing irradiance
31 while phycoerythrin peaked at irradiances of 40-100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. The total FA content was
32 maximal at optimal irradiances for growth, especially under nutrient deficiency. However, highly-
33 unsaturated fatty acids, desired components for copepods, were higher under nutrient excess. The
34 total FAA content was highest at limited irradiances (10-40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) but a better
35 composition with higher fraction of essential amino acids was obtained at saturated irradiances (60-
36 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$). These results demonstrate that quality and quantity of FA and FAA of *R.*
37 *salina* can be optimized by manipulating the irradiance and nutrient conditions. We suggest that *R.*
38 *salina* should be cultivated in a range of irradiance 60-100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and nutrient excess
39 to obtain the algae with high production and a balanced biochemical composition as feed for
40 copepods.

41 Key words: algal production, amino acids, designer feed, fatty acids, phycoerythrin

42

43 **Introduction**

44 Microalgae are essential feeds for many cultured molluscs and larvae of marine fishes and
45 crustaceans (Brown et al. 1997; Muller-Feuga et al. 2003). Microalgae are also used in aquaculture
46 as food for other important live feeds such as for feeding or enriching rotifers, *Artemia* and
47 copepods (Støttrup 2003; Dhert et al. 2001; Sorgeloos et al. 2001; Muller-Feuga et al. 2003). As
48 live feeds in aquaculture, the optimization of their nutritional values, beside the biomass production,
49 is of crucial importance and of special interest.

50 In microalgae, low irradiance may limit photosynthesis (Dunstan 1973), but high irradiances may
51 cause photoinhibition (Neidhardt et al. 1998). Importantly, irradiance may influence the production
52 and composition of the fatty acids in microalgae (Renaud et al. 1991; Mortillaro et al. 2009). Hence,
53 it is possible to manipulate irradiance to optimize growth and the preferred biochemical quality of
54 microalgae.

55 Another important factor regarding algal culture is inorganic nutrients that not only affect
56 photosynthesis and productivity of cell biomass, but also influence the biochemical composition of
57 microalgae (Hu 2004; Juneja et al. 2013). For example, under nutrient deficiency, microalgae
58 exhibit low growth rates (Bi et al. 2014) and produce higher levels of total fatty acids (TFA), but
59 levels of unsaturated fatty acids, the desired components for live feed in aquaculture, are low
60 (Breteler et al. 2005).

61 Nutritional quality of microalgae species is associated with the level of highly unsaturated fatty
62 acids (HUFA), especially Eicosapentaenoic acid (EPA; 20:5 n-3) and Docosahexaenoic acid (DHA;
63 22:6 n-3) (Renaud et al. 1991). HUFAs (characterized by a carbon number ≥ 20 and double bonds
64 ≥ 3) are synthesized *de novo* only by photosynthetic organisms (Spector 1999), and are essential
65 dietary nutrients for marine copepods (Fraser et al. 1989). HUFA, in particular, DHA and EPA
66 appear to be very important in controlling reproduction, growth and metabolism in copepods
67 (reviewed in Rasdi and Qin 2014). High dietary DHA/EPA ratios in feed improve survival, reduce
68 time to maturity, increase maturation rate, female length of calanoid copepod species, egg
69 production, and hatching success (Jónasdóttir 1994; Jónasdóttir and Kiørboe 1996; Payne and
70 Rippingale 2000; Arendt et al. 2006; Rasdi and Qin 2014). Interestingly, copepods are carriers of
71 high DHA/EPA ratios from microalgae into fish larvae (Parrish 2009). A DHA/EPA ratio ≥ 2 is
72 regarded favorable for fish larval nutrition (Sargent et al. 1997). The enhance HUFA content in

73 algae prior to feeding to copepods is recommended as the nutrient content in copepods cannot be
74 manipulated through enrichment techniques due to their avoidance behavior (Rasdi and Qin 2014).

75 Amino acids (AA) constitute another group of important biochemical constituents determining the
76 nutritional quality of microalgae (Brown 1991). AAs are the building blocks for protein synthesis,
77 and are involved in numerous specific physiological functions (Aragão et al. 2004). Some AAs are
78 defined as essential amino acids (EAA) that either cannot be synthesized within the animal body or
79 at an insufficient rate to meet the physiological needs for the growth of animals. EAAs must
80 therefore be supplied from the diet. For copepods, microalgae are the only external source of the
81 EAA (Wu 2009). AA composition of the algal prey also affects the egg production of the copepods
82 (reviewed in Rasdi and Qin 2014).

83 Among the marine microalgae, species of the Cryptophyte genus *Rhodomonas*, such as *R. salina*, *R.*
84 *baltica* and *R. reticulate*, are commonly cultivated for use as live feeds for scallop larvae (Malzahn
85 and Boersma 2012), oyster larvae and spats (Brown et al. 1998; Muller-Feuga et al. 2003) and
86 Queen conch veliger larvae (Aldana-Aranda and Patiño Suárez 1998). Especially, *Rhodomonas*
87 species are excellent feeds for culturing copepods (Støttrup and Jensen 1990; Jónasdóttir 1994;
88 Marinho da Costa and Fernández 2002; Zhang et al. 2013; Broglio et al. 2003).

89 The overall purpose of the present study is to develop a cultivation protocol for applying *R. salina*
90 in large scale algal photobioreactors while optimizing their growth and nutritional value as algal
91 feed for live feed calanoid copepods. We measured the photosynthesis of *R. salina* under
92 irradiances from 10 to 300 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. In the next step, we conducted a full factorial
93 growth experiment in which *R. salina* was cultivated in a series of irradiances ranging from 10 to
94 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ under two inorganic nutrient levels: deficiency and excess. We quantified
95 algal growth rates, three important pigments (chlorophyll *a*, chlorophyll *c* and phycoerythrin,
96 Yaakob et al. 2014), fatty acids and free amino acids profiles for all irradiant levels and nutrient
97 treatments. We have the ambition to develop a simple method where the copepod's biochemical
98 profile is always optimal for the use as fish larvae feed by simply manipulating two algal growth
99 factors (irradiance and nutrient). This will enable us to develop an intensive setup for a nutritious
100 food chain delivering live feed on request for marine hatcheries.

101 **Materials and methods**

102 *Microalgal material and culturing conditions*

103 The studied Cryptophyte *Rhodomonas salina* (equivalent spherical diameter 8 µm) was originally
104 derived as SCCAP K-1487 of the Scandinavian Culture Collection of Algae and Protozoa
105 (University of Copenhagen, Denmark). Cultures of *R. salina* was grown in acid washed 3-6 L round
106 glass flasks containing autoclaved 0.2 µm filtered seawater (salinity 30 ‰) enriched with B1
107 medium (1 mL L⁻¹ of seawater, Hansen 1989). The cultures were maintained under a continuous
108 irradiance of 80 µmol photons m⁻² s⁻¹ Photosynthetic Active Radiation (PAR) in a climate room at
109 20°C. The flasks were gently aerated with atmospheric air through 0.45 µm filters to mix the
110 cultures to avoid temperature stratification, algal sedimentation, CO₂ depletion and O₂ build up.

111 *Photosynthesis measurements*

112 Measurements of photosynthesis-irradiance (P-I) curves were performed for 5 cell densities (0.1,
113 0.5, 1, 2, 10 × 10⁶ cells mL⁻¹) at 16 increasing irradiances (0, 10, 20, 40, 60, 80, 100, 120, 140, 160,
114 180, 200, 220, 240, 260, 300 µmol photons m⁻² s⁻¹ PAR), with n = 5 experimental replicates of each
115 irradiances from 0 to 140 µmol photons m⁻² s⁻¹ and n = 2-3 experimental replicates for irradiances
116 from 160 to 300 µmol photons m⁻² s⁻¹. The photosynthesis of *R. salina* was measured when the
117 algae was in the exponential phase for all treatments. A high concentration of algae was achieved
118 by centrifuging at 1000 rpm for 5 minutes at 20°C. The algae suspension was diluted to the required
119 cell concentration using fresh 0.2 µm filtered autoclaved seawater containing B1 medium. Net
120 oxygen exchange rates were measured with a Clark-type oxygen electrode (S1 Oxygen Electrode
121 Disc, Hansatech Instruments, Norfolk, UK) fitted in a stirred Hansatech DW3 chamber. Light was
122 provided by a red LED-lamp (Hansatech LC1). The measurement at each irradiance was completed
123 within 5 min when steady-state photosynthesis had been achieved.

124 Curves were fitted to the photosynthesis – irradiance data using the equation 1 (Platt et al. 1980):

125
$$P^B = P_s^B \left(1 - e^{\frac{-\alpha I}{P_s^B}} \right) e^{\frac{-\beta I}{P_s^B}} \quad (1)$$

126 Where P^B : photosynthetic rate at irradiance I

127 P_s^B : Maximum theoretical (irradiance-saturated) photosynthetic rate in the absence of
128 photoinhibition

129 α : Initial slope of the P-I curve (the quantum yield)

130 β : Negative slope at high irradiance (photoinhibition)

131 In addition, the realized maximum photosynthetic rate attained (P_m^B) and irradiance of maximum
132 photosynthesis (I_m) were calculated using the following equations 2 and 3 (Platt et al. 1980):

133
$$P_m^B = P_s^B \left(\frac{\alpha}{\alpha + \beta} \right) \left(\frac{\alpha}{\alpha + \beta} \right)^{\frac{\beta}{\alpha}} \quad (2)$$

134
$$I_m = \frac{P_s^B}{\alpha} \ln \left(\frac{\alpha + \beta}{\beta} \right) \quad (3)$$

135 *Experimental design for growth experiment*

136 In this experiment, microalgae were grown in a Multi-cultivator MC1000 OD (Photon Systems
137 Instruments, Drasov, Czech Republic) with eight 100 mL test tubes. The test tubes were immersed
138 in a 5L flat, rectangular glass container in which water was circulated by pump through an
139 additional Cooling Unit AC-88 to maintain a stable temperature of 20°C in all test tubes. Between
140 each slot, there was a plastic divider in the cultivation vessel to separate light regimes of individual
141 tubes. Tubes were illuminated by white LEDs that were independently adjustable at up to 500 μmol
142 photons $\text{m}^{-2} \text{s}^{-1}$. Each test tube was bubbled with atmospheric air.

143 To evaluate the effect of the irradiance and nutrients on growth and biochemical profile of *R. salina*,
144 a factorial design of 8 irradiant levels (10, 20, 40, 60, 80, 100, 120 and 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) \times
145 2 nutrient levels (deficiency and excess) was conducted (a total of 16 experimental treatments). All
146 treatments were performed in duplicates (a total of 32 experimental units). The nutrient deficiency
147 and excess treatments were not based on the initial nutrient concentrations but based on how many
148 pulses nutrients were added to the culture. In nutrient deficiency, the nutrients were added only
149 once at the start of the experiment (B1 medium, 1 mL L^{-1} of seawater, Hansen 1989), hence the
150 nutrient level reduced over time and almost depleted at the end of the experiment. In the nutrient
151 excess, the nutrients were added daily (v:v 1 mL B1 medium $\text{L}^{-1} \text{day}^{-1}$ of algae culture), hence no
152 depletion occurred. The initial density of *R. salina* was $0.18 \pm 0.03 \times 10^6$ cells mL^{-1} . In the growth

153 experiment, all of algal cultures were grown for 5-6 days at 20°C, salinity 30 ‰ with the same flow
154 of gas bubbling to ensure stirring and gas mass transfer.

155 *Growth rate*

156 The cell density and cell biovolume of *R. salina* was determined every day by taking 1 mL of algae
157 samples from each treatment and measuring by a Beckman Multisizer™3 Coulter Counter®
158 (Beckman Coulter Inc., USA). All particles with a diameter in the range of 5-12 µm were
159 considered as algal cells. The growth rate (day⁻¹) of *R. salina* was calculated for the first three days
160 of the experiment by fitting cell density increase during the exponential phase with an exponential
161 growth equation 4:

$$162 \quad N_t = N_0 \times e^{(\mu \times t)} \quad (4)$$

163 N_t is the cell density at time t (cells mL⁻¹)

164 N_0 is the cell density at time zero (cells mL⁻¹)

165 μ is the growth rate (day⁻¹)

166 t is the time (day)

167 The specific growth rate-irradiance curves was fitted using tangent hyperbola functions (Jassby and
168 Platt 1976), equation 5:

$$169 \quad \mu = \mu_{\max} \tanh\left(\frac{\alpha I}{\mu_{\max}}\right) \quad (5)$$

170 Where, μ : specific growth rate (day⁻¹)

171 μ_{\max} : Maximum growth rate (day⁻¹)

172 I : Irradiance (µmol m⁻² day⁻¹)

173 α : Initial slope of the curve (maximum quantum yield for growth, day⁻¹ [µmol photons m⁻² s⁻¹]⁻¹)

174 *Cell biovolume*

175 The cell biovolume of the specific sample was determined as the mean of biovolume of all particles
176 presented by the particle counter in the frequency diagram with a diameter in the range of 5-12 µm.

177 The presented cell biovolume of *R. salina* was the mean (\pm SDs) of cell biovolume of the algae in
178 the stationary phase of the algal growth of all treatments when the biochemical composition of
179 algae cell were analyzed.

180 *Sample preparation for quantification of inorganic nutrients of algae cultures and pigments, fatty* 181 *acids and amino acids of algae cell*

182 To minimize the loss of culture volume due to the sampling, inorganic nutrient level analyses were
183 carried out by measuring the samples taken from the first experimental replicate, whereas
184 biochemical analyses of the algae cell *R. salina* were carried out by utilizing the samples taken from
185 the second experimental replicate. During every second day, sample water was sampled from each
186 test tube for analyzing nitrate (4 mL), ammonium (3 mL) and phosphate (6 mL). Culture water was
187 filtered through 25 mm syringe filter (VWR International, USA) containing a Whatman GF/F glass
188 fiber filter to remove algae, and the sample water was then stored in -20°C for later analyses of
189 nutrient compositions. In the stationary phase of the algal growth, samples of *R. salina* were taken
190 on two different days from each treatment for analyzing chlorophyll *a* and chlorophyll *c* (chl *a* &
191 chl *c*, 5 mL \times 2 analytical replicates), phycoerythrin (PE, 5 mL \times 2 analytical replicates), fatty acids
192 (FA, 5 mL \times 2-3 analytical replicates), and free amino acids (FAA, 5 mL \times 2 analytical replicates).
193 Then, samples were filtered onto three 12.8 mm diameter GF/C glass fiber filter (Whatman) and
194 preserved in a biofreezer at -80°C for later analyses of pigments, fatty acids and free amino acids
195 compositions.

196 *Inorganic nutrient analysis of R. salina culture*

197 Nitrate, ammonium and phosphate in the filtered water from the algae culture were quantified using
198 colorimetric techniques. Nitrate concentration was determined by flow injection analysis using
199 QuickChem Method 31-107-04-1-A (Diamond 1999). Ammonium concentration was analyzed by
200 salicylate-hypochlorite method for determining ammonium in seawater described by Bower and
201 Holm-Hansen (2011). Phosphate concentration was quantified by a Spectrachrom UV-1601 UV-
202 Visible Spectrophotometer (Shimadzu, Kyoto, Japan) following the method described by
203 S ndergaard and Riemann (1979). The final concentration of inorganic nutrients (nitrate,
204 ammonium and phosphate) in the specific experimental treatment was defined as the average (\pm SD)
205 concentration between the last two sampling days (day 4 and 6) of the experiment.

206 *Analyses of algal pigments*

207 Chl *a* and chl *c* were extracted based on the methods described by Jeffrey and Humphrey (1975)
208 and Ritchie (2006). Filter samples were lyophilized before extraction. Each of these filters was
209 placed in a glass vial where 3.3 mL of 90% acetone was added. Samples were shaken in a whirly
210 mixer. Then, samples were placed in the dark for 24 hours at 5°C. The extraction solvent in each
211 vial was transferred into a quartz cuvette through a 0.2 µm pore size syringe filter in which the
212 absorbance of each sample was measured at 664 nm and 630 nm on a GENESYS™ 6
213 Spectrophotometer (ThermoSpectronic). The concentration of chl *a* and chl *c* is expressed as pg
214 cell⁻¹.

215 The PE was extracted based on the procedure described by Bennett and Bogorad (1973), Evans
216 (1988) and Zimba (2012). After lyophilization, each of the filter samples for PE extraction was
217 placed into a glass vial together with 3 mL of phosphate buffer (0.1 mol pH 7, 0.05 mol K₂HPO₄,
218 0.05 mol KH₂PO₄). Samples were sonicated in ice-water bath for 15 minutes and then were left
219 refrigerated for 12 hours. Extraction solvent was filtered through at 25 mm 0.2 µm pore size syringe
220 filter and placed into a cuvette for measuring the absorbance spectrophotometrically at 455nm,
221 564nm and 592nm. PE concentration was calculated as in Bennett and Bogorad (1973). The
222 concentration of PE is expressed as pg cell⁻¹.

223 *Analyses of fatty acids*

224 The FA composition of *R. salina* was determined by extraction of the lipids using a HPLC-grade
225 chloroform: methanol mixture (Folch et al. 1957) followed by trans esterification process by acetyl
226 chloride in methanol (see Drillet et al. 2006 for details). In brief, a chloroform: methanol mixture (3
227 mL, v:v = 2:1) was added to each of the algal sample. A volume of 20 µL of internal standard (1000
228 µg mL⁻¹ tricosanoic FA methyl ester [C23:0 FAME]) was also added for FA quantification.
229 Thereafter, samples underwent ultrasound extraction for 15 minutes in an ice bath to break the algal
230 cells. In the next step, samples were frozen at -20°C for 24 h for extraction. Subsequently, the
231 extraction solvent from each sample was transferred to GC vials and was placed into an aluminum
232 block at 60°C to evaporate the chloroform: methanol solvent by a flow of nitrogen. Thereafter,
233 AcOMe/HCl reagent in Toluene (1 mL) was added into the GC vials. The GC vials were covered by
234 aluminum caps and were placed in the aluminum block for 2 hours at 95°C. Next, the caps were
235 removed to add 500 µL of 5% of NaHCO₃, and two different phases appeared. The upper phase was
236 transferred into a new GC vial. Subsequently, 500 µL of heptane was added and the upper phase
237 was added to the new GC vial. The samples were evaporated at 65°C under a steady flow of

238 nitrogen. Thereafter, 0.5 ml of chloroform was added to each sample. Finally, all samples were
239 analyzed by an Agilent GC6890N gas chromatograph while connected to an Agilent MS 5975 mass
240 selective detector. The GC was equipped with a 60 m Agilent J&W DB23 column with 0.25 mm
241 internal diameter and film thickness of 0.25 μm . Splitless injection while running a positive electron
242 ionization at 70 eV was selected. ChemStation software was used for MS peak integration. MS
243 peaks were analyzed against a Supelco FAME standard mixture. Total fatty acid content (TFA) is
244 expressed as pg cell^{-1} , while content of each FA is expressed as percentage of TFA (% of TFA).

245 *Analyses of free amino acids*

246 The FAA of *R. salina* was analyzed based on the method also reported by Drillet et al. (2006). The
247 filter samples were lyophilized 24h prior to extraction. FAA was extracted in 1 mL Milli-Q water
248 by heating the filter samples to 95°C for 10 min. The extracts were filtered through 8 mm 0.2 μm
249 pore size GHP polypropylene membrane filters. The FAA was derivatized (Yu et al. 1994) using a
250 AccQFlour kit (Waters, MA, USA) and later separated on a Waters Alliance 2695 separation
251 module with a 3.9 \times 150 mm AccQTag column. The separated AA derivatives were quantified by
252 fluorescence (250 nm excitation and 395 nm emission) using an Alliance 2475 scanning
253 fluorescence detector. Due to the limitation of the number of FAA samples (only one sample for
254 each treatment), the FAA data was grouped into two categories according to the irradiance: i)
255 limited irradiance (irradiance from 10 to 40 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$, $n = 3$ for both nutrient deficiency
256 and nutrient excess treatments) and ii) saturated irradiance (irradiance from 60 to 140 $\mu\text{mol photons}$
257 $\text{m}^{-2} \text{ s}^{-1}$, $n = 5$ for nutrient deficiency and $n = 4$ for nutrient excess treatment). The saturated
258 irradiances were from 60 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ where no further increase in the microalgal growth
259 rate under the higher irradiance was observed (see the results). Total FAA is expressed as pg cell^{-1} ,
260 while content of each amino acid is expressed as percentage of total FAA (% of total FAA).

261 *Statistical analyses*

262 Response variables were subjected to two-way ANOVA with nutrient and irradiance as fixed
263 factors. Tukey tests were subsequently used to compare individual means across significantly
264 different treatment levels where relevant. For the results of Tukey tests for maximum cell density,
265 nutrients, algal pigments, total fatty acids content and the DHA/EPA ratio, it was so difficult to see
266 the different letters in bars as there are many bars in each figure that for clarity we prefer not adding
267 the letters above the bars to indicate the statistically difference. We therefore only provide the

268 results of the Tukey test in a letter code in the tables for fatty acids composition and free amino acid
269 composition results. Data were tested for homogeneity of variance (Cochran's test) and normal
270 distribution (Kolmogorov-Smirnoff goodness of fit test) before being analyzed by ANOVA. All
271 tests on data were carried out using SAS v. 9.3 with $\alpha = 0.05$.

272 Results

273 *Photosynthesis*

274 The realized photosynthesis rate (P^B_m) and the initial slope of the P-I curves (α) decreased with the
275 increase of cell density (Fig. 1 and Table 1). The P^B_m value was 22 times higher at the lowest cell
276 density (0.1×10^6 cells $mL^{-1} = 539.4 \times 10^{-15}$ mol O_2 cell $^{-1}$ h $^{-1}$) compared to the highest cell density
277 (10×10^6 cells $mL^{-1} = 20.6 \times 10^{-15}$ mol O_2 cell $^{-1}$ h $^{-1}$). In contrast, the irradiance of maximum
278 photosynthesis (I_m) increased with increasing cell density. The lowest I_m values were 57.5-59.2
279 μ mol photons m^{-2} s $^{-1}$ occurring at the two lowest cell densities of 0.1 - 0.5×10^6 cells mL^{-1} and the
280 highest I_m value was 103.0 μ mol photons m^{-2} s $^{-1}$ (ca. 1.5 times higher than the lowest I_m value)
281 occurring at the highest cell density of 10×10^6 cells mL^{-1} (Table 1). The photoinhibition (β)
282 decreased with increasing cell density (Fig. 1, Table 1). Dark respiration (R) decreased with
283 increasing cell density, indicated by the higher oxygen consumption per cell (Table 1). The highest
284 dark respiration was recorded at lowest cell density 0.1×10^6 cells mL^{-1} ($-319.1 \pm 19.8 \times 10^{-15}$ mol
285 O_2 cell $^{-1}$ h $^{-1} = 59.1\%$ P^B_m) which was ca. 18 times higher than the dark respiration of cells at highest
286 density 10×10^6 cells mL^{-1} ($-17.9 \pm 5.7 \times 10^{-15}$ mol O_2 cell $^{-1}$ h $^{-1} = 86.7\%$ P^B_m). The ratio of relative
287 dark respiration rate to maximum photosynthesis (R/P^B_m) increased slightly with increasing cell
288 density (Table 1) indicating that the cell culture became less autotrophic with increasing cell
289 density.

290 *Microalgae growth*

291 As the specific growth rate was measured during the exponential growth phase, where no nutrient
292 deficiency had yet set in, there is no significant difference in the specific growth rate between
293 nutrient deficiency and nutrient excess treatments. These were therefore pooled for fitting the same
294 specific growth rate-irradiance curve (Fig. 2a). The specific growth rate increased with the increase
295 of irradiance and reached a plateau from the irradiance of 60 μ mol photons m^{-2} s $^{-1}$ onward (Fig. 2a).
296 The maximal growth rate (μ_{max}) was 0.752 day $^{-1}$ with the maximal quantum yield for growth of
297 0.014 cells day $^{-1}$ [μ mol photons m^{-2} s $^{-1}$] $^{-1}$. Algae did not grow ($\mu \approx 0$) at the lowest irradiance (10
298 μ mol photons m^{-2} s $^{-1}$).

299 Maximum cell densities increased with increasing irradiance (Fig. 2b, Appendix 1). This pattern
300 was consistently stronger in nutrient excess cultures than in nutrient deficient cultures (Appendix 1).
301 The maximal cell density in nutrient excess (3.2 - 5.3×10^6 cells mL^{-1}) was about 2-3 times higher

302 than in nutrient deficiency ($1.7-1.9 \times 10^6$ cells mL⁻¹) at the irradiance from 80 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$
303 onwards, resulting in the overall higher maximal cell densities (Fig. 2b) in nutrient excess than in
304 nutrient deficiency (Appendix 1).

305 The cell biovolume of *R. salina* was found to be 200-300 μm^3 (Fig. 2c). Linear regression revealed
306 no significant relationship between specific growth rate and cell biovolume, $p = 0.633$ and $p =$
307 0.923 , for nutrient excess and deficiency, respectively, indicating that cell biovolume is independent
308 of growth rate in this study, regardless of treatment.

309 *Nutrient consumption*

310 The initial concentration of nitrate was $1092.9 \pm 108.3 \mu\text{mol L}^{-1}$ in all treatments. The final nitrate
311 concentrations (Fig. 3a) remaining in the algae culture decreased with increasing irradiance
312 (Appendix 1) and were higher in nutrient excess than in nutrient deficiency (Appendix 1). The
313 lowest final nitrate concentrations were recorded at the irradiances of 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (1.1
314 $\pm 1.0 \mu\text{mol L}^{-1}$) and 120 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ ($418.3 \pm 384.9 \mu\text{mol L}^{-1}$) in nutrient deficiency and
315 nutrient excess, respectively. In addition, the variation in the final nitrate concentrations was wider
316 under nutrient deficiency than under nutrient excess (Appendix 1).

317 At the start of the experiment, the ammonium concentration was under the detection limit (0.05
318 $\mu\text{mol L}^{-1}$) in all treatments. At the end of the experiment, the ammonium concentration was only
319 detectable in the treatments at irradiances of higher than 20 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and of higher than
320 40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ for nutrient deficiency and nutrient excess (Fig. 3b), respectively. The final
321 ammonium concentration increased with increasing irradiance (Appendix 1) and was considerably
322 higher in nutrient excess than nutrient deficiency (Appendix 1).

323 The initial concentration of phosphate was $160.9 \pm 29.0 \mu\text{mol L}^{-1}$ in all treatments. The final
324 phosphate concentration (Fig. 3a) remaining in the algae culture decreased with increasing
325 irradiance (Appendix 1) and was higher at nutrient excess than at nutrient deficiency (Appendix 1).
326 At the end of the experiment, the lowest phosphate concentration was obtained at the highest
327 irradiance 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in both nutrient deficiency ($86.7 \pm 35.0 \mu\text{mol L}^{-1}$) and nutrient
328 excess ($112 \pm 10.2 \mu\text{mol L}^{-1}$). The decrease in final phosphate concentration with irradiance was
329 more pronounced in nutrient deficiency than nutrient excess (Appendix 1).

330 *Algal pigments*

331 Overall, cellular chlorophyll *a* (chl *a*) levels decreased with increasing irradiance (Fig. 4a,
332 Appendix 1). The highest chl *a* (ca. 6.7 pg cell⁻¹) recorded at the irradiance of 20 μmol m⁻² s⁻¹ was
333 ca. 3 times higher than the lowest chl *a* (ca. 2.4 pg cell⁻¹) recorded at the highest irradiance (140
334 μmol m⁻² s⁻¹). Chl *a* levels did not differ between nutrient deficiency and nutrient excess (Appendix
335 1). There was a statistically significant interaction between the irradiance and nutrient on chl *a*
336 concentrations (Appendix 1).

337 Cellular chlorophyll *c* (chl *c*) levels decreased with increasing irradiance (Fig. 4b, Appendix 1). The
338 chl *c* levels were not affected by nutrient treatment (Appendix 1). However, there was an interaction
339 between irradiance and nutrient on chl *c* (Appendix 1).

340 The cellular phycoerythrin (PE) levels were influenced by both irradiance (Fig. 4c, Appendix 1) and
341 nutrients (Fig. 4c, Appendix 1). In nutrient deficiency, the PE rapidly increased at low irradiance
342 and peaked at 40 μmol photons m⁻² s⁻¹ (ca. 18 pg cell⁻¹). Thereafter, the PE decreased rapidly to the
343 lowest level at 140 μmol photons m⁻² s⁻¹ (ca. 3 pg cell⁻¹). In nutrient excess, the PE increased
344 rapidly with increasing irradiance at low irradiances, reaching the highest concentration at the
345 irradiance of 60 μmol photons m⁻² s⁻¹ (somewhat later compared to nutrient deficiency) and then
346 remained at this high level until 100 μmol photons m⁻² s⁻¹ before slightly decreasing at 120-140
347 μmol photons m⁻² s⁻¹. This difference generated a statistically significant Irradiance × Nutrient
348 interaction (Appendix 1). In term of nutrients, the PE level was higher in nutrient excess (11.1 ± 5.5
349 pg cell⁻¹) than nutrient deficiency (8.9 ± 6.5 pg cell⁻¹) but this pattern was driven by the fact that PE
350 levels were considerably higher in nutrient excess at the irradiance from 80 μmol photons m⁻² s⁻¹
351 onward.

352 The PE/chl *a* ratio (Fig. 4d) resembled the patterns of the PE. The PE/chl *a* ratio was affected by
353 both irradiance (Appendix 1) and nutrients (Appendix 1). Specifically in nutrient deficiency, the
354 PE/chl *a* ratio rapidly increased at low irradiances, peaking at an irradiance of 40 μmol photons m⁻²
355 s⁻¹ (4.2 pg cell⁻¹), then decreased steadily with increasing irradiances, attaining the lowest level at
356 140 μmol photons m⁻² s⁻¹ (1.2 pg cell⁻¹). In nutrient excess, the PE/chl *a* ratio increased with
357 increasing irradiance and reached a plateau (2.9-3.5 pg cell⁻¹) from the irradiance of 40 μmol
358 photons m⁻² s⁻¹ onwards (Fig. 4d). This difference also generated a statistically significant
359 interaction (Appendix 1) and resulted in an overall higher PE/chl *a* ratio in nutrient excess than in
360 nutrient deficiency.

361 The PE content tended to be negatively correlated to the cell density when cultivated in nutrient
362 deficiency ($p = 0.0579$, Fig. 4e). In nutrient excess, the PE content was not significantly related to
363 the cell density ($p = 0.3434$, Fig. 4f).

364 *Fatty acids*

365 Overall, the total fatty acids (TFA) content increased with increasing irradiance (Fig. 5a,
366 Appendix 1). The TFA was considerably higher (Appendix 1) in algae cultured in nutrient
367 deficiency (34.5 ± 22.5 pg cell⁻¹) than in nutrient excess (11.4 ± 4.2 pg cell⁻¹). Most notably, TFA
368 was ca. 3-4 times higher within the irradiance range of 60 to 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ in nutrient
369 deficient cultures in comparison to the nutrient excess, generating a statistically significant
370 Irradiance \times Nutrient interaction (Fig. 5a, Appendix 1).

371 Irradiance had statistically significant effects on the relative abundance of the mono-unsaturated
372 fatty acids (MUFA) (Fig. 5b&c, Appendix 1), especially in nutrient excess, generating an
373 interaction of Irradiance \times Nutrient on MUFA (Appendix 1). Algae cultured at lowest and highest
374 irradiance (10 and 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) had significant higher MUFA levels compared to other
375 irradiances (Fig. 5b&c).

376 Irradiance also affected the content of short chain polyunsaturated fatty acids (SC-PUFA =
377 characterizing by a carbon number < 20 and double bonds > 1) (Fig. 5b&c, Appendix 1). This
378 pattern was independent of nutrient levels (Appendix 1). SC-PUFA increased with increase in
379 irradiance. On the other hand, the changing in irradiance levels did not affect the concentration of
380 the saturated fatty acids (SFA) and highly unsaturated fatty acids (HUFA) (Fig. 5b&c, Appendix 1).

381 Nutrient had a statistically significant effect on the SFA (Appendix 1) and HUFA (Appendix 1) of
382 the algae (Fig. 5b&c). Relative abundance of SFA was considerably higher when algae were
383 cultured during nutrient deficiency (26.8 ± 8.3 % of TFA) compared to those in nutrient excess
384 (11.0 ± 3.1 % of TFA). In contrast, the relative abundance of HUFA was ca. 2 times higher in algae
385 cultured in nutrient excess than those cultured in nutrient deficiency (Appendix 1), accounting for
386 31.8 ± 2.5 % and 15.1 ± 5.7 % of TFA, respectively. Especially Eicosapentaenoic acid (EPA), the
387 most abundant HUFA in *R. salina*, was 22.1 % of TFA in nutrient excess, which was ca. 2 times
388 higher than in nutrient deficiency (10.3 % of TFA).

389 Overall, the ratio between the two essential fatty acids Docosahexaenoic acid and Eicosapentaenoic
390 acid (DHA/EPA) was lower at irradiance 10-20 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ than at irradiance from 40-140
391 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (Appendix 1). The nutrient had no effect on DHA/EPA ratio which varied
392 from 0-0.86 in nutrient deficiency and from 0.28-0.57 in nutrient excess. There was an Irradiance \times
393 Nutrient interaction (Appendix 1), but this interaction was driven by the fact that DHA was mostly
394 not synthesized at the two lowest irradiance level (Fig. 5d).

395 More details in fatty acids composition of *R. salina* cultured in nutrient deficiency and nutrient
396 excess under limited (irradiance from 10 to 40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) and saturated (irradiance from
397 60 to 140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) irradiances was summarized in Appendix 2. Noticeably, relative
398 abundance of SFA 18:0 was highest at saturated irradiance and under nutrient deficiency, whereas,
399 relative abundance of SC-PUFA 18:4, DHA and EPA was highest at saturated irradiance and in
400 nutrient excess.

401 *Free amino acids*

402 The total free amino acids (FAA) and specific essential amino acids (EAA, mean \pm SDs of total
403 FAA) for copepods (Claybrook 1983) and fish growth (Wilson 1985) that were present in *R. salina*
404 cells cultured at different irradiance and nutrient conditions are summarized in table 2. Total FAA
405 was affected by the irradiance (Table 2), but not by the nutrient levels (Table 2). The total FAA was
406 significantly higher when the algal cells were cultivated at limited irradiance ($8.3 \pm 2.5 \text{ pg cell}^{-1}$ at
407 10-40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) than in the saturated irradiance ($4.3 \pm 1.3 \text{ pg cell}^{-1}$ at 60-140 μmol
408 $\text{photons m}^{-2} \text{s}^{-1}$). There was no interaction between irradiance and nutrient levels on total FAA of
409 algae (Table 2).

410 The relative abundance of sub-total EAA (% of total FAA) was affected by the irradiance and the
411 interaction Irradiance \times Nutrient, but not the nutrient levels (Table 2). In contrast with total FAA,
412 the relative abundance of sub-total EAA was significantly higher at saturated irradiance, especially
413 in nutrient in excess (31.89 ± 4.24 % of total FAA, Table 2).

414 The difference in the irradiance and nutrient showed no consistent effect on relative abundance of
415 specific EAA (Table 2). The most abundant EAA in *R. salina*, arginine was higher in saturated
416 irradiance, especially under nutrient deficiency (Table 2). Seven out of ten EAA, including
417 isoleucine, leucine, lysine, methionine, phenylalanine, threonine, valine, were generally higher in
418 the algae cells cultured under saturated irradiance and in nutrient excess compared to those cultured

419 at limited irradiance and in nutrient deficiency (Table 2). In contrast, histidine was higher in algae
420 grown at limited irradiance and in nutrient deficiency (Table 2).

421

422 **Discussion**

423 In this study, we found strong effects of irradiance and/or nutrient levels on all measured variables
424 such as photosynthesis, growth rate, and biochemical composition of *Rhodomonas salina*.

425 *Photosynthesis*

426 The photosynthesis rate of *R. salina* at all five cell densities increased with increasing irradiance
427 until reaching a saturating irradiance (I_m) of 60-100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. The range of I_m found in
428 this study confirmed what has been documented in previous studies ($I_m \geq 60 \mu\text{mol photons m}^{-2} \text{s}^{-1}$
429 Hammer et al. 2002; and $I_m \geq 200 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ Bartual et al. 2002). Photoinhibition,
430 indicated by $\beta > 0$ (Platt et al. 1980), occurred at all cell densities, but higher cell densities reduced
431 the photoinhibition as indicated by the rapid decrease of β with increasing cell densities. This
432 density-mediated decrease in photoinhibition is probably due to an increase in self-shading at higher
433 cell densities. This finding was in contrast with previous studies (e.g., Hammer et al. 2002; Bartual
434 et al. 2002) where *R. salina* showed no photoinhibition when being exposed to irradiances from 0-
435 1200 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$. This difference could be a result of methodical differences. In our study,
436 photosynthesis measurements were carried out using the culture maintained at 80 $\mu\text{mol photons m}^{-2}$
437 s^{-1} at a cell density of ca $1.5\text{-}2 \times 10^6 \text{ cells mL}^{-1}$, which was then diluted to different cell densities.
438 Therefore, the algae may already be acclimated to relatively low irradiances before the
439 measurements took place while in other studies the photosynthesis was measured at low algal cell
440 densities (although it was unclear which algal densities that these studies used) at different
441 irradiances, from 11-320 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (Bartual et al. 2002) or 10-150 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$
442 (Hammer et al. 2002).

443 In addition, the respiration rate in darkness was also higher at low cell density than at high cell
444 density, probably due to the higher cellular photosynthetic activity at low cell densities, leading to
445 higher metabolic maintenance cost (Kromkamp and Peen 2001).

446 *Algal growth*

447 Regarding the specific growth rate versus irradiance curve, the saturating irradiance (60-140 μmol
448 $\text{photons m}^{-2} \text{s}^{-1}$) for maximal growth and maximal growth rate (μ_{max}) obtained in our study are in
449 agreement with previous studies on *R. salina* (Bartual et al. 2002; Hammer et al. 2002; Lafarga-De
450 la Cruz et al. 2006). The maximal growth rate of *Pyrenomonas salina* (a taxonomic synonym of *R.*

451 *salina*) was somewhat higher (1.2 day^{-1}) at a saturating irradiance of ca. $100 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ in
452 the study of Lewitus and Caron (1990). The higher specific growth rate in the previous study in
453 comparison to our study is most likely due to the use of a more preferable nitrogen source-
454 ammonium for algal growth instead of nitrate in our study. It is well known that algae can take up
455 and assimilate ammonium directly while they have to reduce nitrate to ammonium before
456 assimilation (Rückert and Giani 2004), hence a higher energetic cost is associated with nitrate use.
457 Under low irradiance ($< 20 \mu\text{mol photons m}^{-2} \text{ s}^{-1}$), the growth rate was very low, as also observed in
458 other studies (Lewitus and Caron 1990; Bartual et al. 2002) that may be associated with the low
459 photosynthetic rate at low irradiance.

460 Until the end of the exponential phase (day 3), the specific growth rate of *R. salina* did not differ
461 between nutrient levels as nutrients in all treatments was still above the half saturation constants for
462 algal growth, namely nitrate: $0.4 \mu\text{mol L}^{-1}$ (Falkowski 1975) and phosphate: $0.51 \mu\text{mol L}^{-1}$ (Smith
463 and Kalff 1982). The nutrient deficiency only showed its effects on the algal growth from day 4 and
464 onwards where the nitrate concentration was almost depleted. Not surprisingly, the cell densities in
465 nutrient deficient cultures did not increase beyond this point. However, in the current study, the
466 maximum cell density in nutrient deficient cultures was still higher than the reported value $1.53 \times$
467 $10^6 \text{ cells mL}^{-1}$ by Lafarga-De la Cruz et al. (2006), which may be a result of the difference in
468 experimental set up. In Lafarga-De la Cruz et al. (2006), algae were cultivated in batch without
469 aeration in (250 mL) Erlenmeyer flasks whereas in our study, the algae was cultivated in a Multi-
470 cultivator with continuous bubbling of atmospheric air that may provide a better environment for
471 growth by enhancing CO_2 addition, avoiding light/temperature stratification, algal sedimentation
472 and O_2 build up. In contrast, the nitrate concentration in nutrient excess remained very high during
473 all 6 days of the experiment. As a result, cell densities increased continuously throughout this
474 period, especially under optimal range of irradiance and nutrient excess, where maximal cell
475 densities were 2-3 times higher than those cultured in nutrient deficiency of the same light regimes,
476 indicating a promising algal production.

477 As cell biovolume was not significantly affected by growth rate, and has been found to be constant
478 across experimental conditions and treatment, we have chosen to express all data on biochemical
479 composition on a per cell basis.

480 *Algal pigments*

481 In response to the decrease in irradiance, the phytoplankton typically increases chlorophyll *a* (chl *a*)
482 and other light harvesting pigments, such as chlorophyll *b* (chl *b*), chlorophyll *c* (chl *c*), and primary
483 carotenoids (Hu 2004). These results have been observed in Cryptophyte species (Faust and Gantt
484 1973; Lichtlé 1979), including *R. salina* (Bartual et al. 2002). In the present study, the chl *a* and chl
485 *c* concentrations were higher at lower irradiance as a response to low irradiance (Hammer et al.
486 2002; Hu 2004). When the algae are exposed to high irradiance, phycobiliproteins and carotenoids
487 act as protection mechanisms against the excess light (Pereira et al. 2012). Therefore, it was expected
488 that PE content would increase at high irradiance to increase the photoprotection, which indeed was
489 observed in our study under nutrient excess. In our experiment, the phycoerythrin (PE) and chl *a*
490 (PE/chl *a*) ratio of *R. salina* decreased with the increase of irradiance ($> 40 \mu\text{mol photons m}^{-2} \text{s}^{-1}$) in
491 nutrient deficiency, which has been also observed in the Cyanophyceae *Anacystis nidulans* (Halldal
492 1958) or the Cryptophyceae *Chroomonas* sp. (Faust and Gantt 1973). Moreover, the PE/chl *a* ratio
493 follows the pattern of total PE against irradiance, and can be explained by PE being more sensitive to
494 variations in irradiance than chl *a*. (Brown and Richardson 1968).

495 A common pattern in algae is that cells respond to nutrient (nitrogen) deficiency by decreasing
496 pigment content (Hu 2004). This pattern has been reported in several Cryptophyte species such as
497 *Cryptomonas rufescens* (Lichtlé 1979), *Cryptomonas maculata* (Rhiel et al. 1985) and
498 *Pyrenomonas salina* (Lewitus and Caron 1990). In our study, while the PE and PE/chl *a* ratio were
499 indeed lower in nutrient deficiency, we observed no effect of nutrients on the chl *a* and chl *c*
500 content. For the chl *a* content, the nitrate concentration in our experiment (the lowest nitrate
501 concentration was $1.1 \pm 1.0 \mu\text{mol}$) was probably not too depleted to induce a reduction in chl *a*
502 content like in the study of Bartual et al. (2002) in which *R. salina* suffered under nitrogen
503 concentrations below $0.5 \mu\text{mol}$ to total exhaustion from day 4 of the experiment. The drop of PE and
504 PE/chl *a* ratio under nutrient deficiency has been observed before in *R. salina* (Bartual et al. 2002).
505 This nutrient-induced reduction in PE and PE/chl *a* ratio is explained by mobilization of nitrogen
506 from PE (Bartual et al. 2002).

507 In our study, PE decreases with increasing cell density under nutrient deficiency, indicating that
508 this phycobillipigment is scavenged as a nitrogen source under nutrient deficiency (Bartual et al.
509 2002; Lewitus and Caron 1990; Eriksen and Iversen 1995) as photoprotection is decreasing with
510 increasing cell density and thus increasing self-shading.

511 *Fatty acids*

512 In line with previous studies, the higher total fatty acids (TFA) was obtained at higher irradiance
513 (Sharma et al. 2012; Dongre et al. 2014) and nutrient deficiency (Sriharan et al. 1991; Piorreck et al.
514 1984; Hu 2004; Shifrin and Chisholm 1981, but see Harrison et al. 1990). Strikingly, TFA levels
515 increased sharply to very high level at irradiance 60-140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ under nutrient
516 deficiency. The increase of TFA with increase of irradiance in nutrient deficiency could be
517 correlated to the increase of triacylglycerols-TAG (Sharma et al. 2012; Dongre et al. 2014). This is
518 likely associated with the synthesis of TAG (generally contain saturated fatty acids-SFA and mono-
519 unsaturated fatty acids-MUFA) that mostly occur under adequate light conditions, hence this
520 synthesis can be maximized when cultivated at light saturation (Dongre et al. 2014). Besides, in
521 nutrient deficiency condition (nitrogen starvation), many algal species accumulate lipids due to that
522 these constituents do not contain N (mostly TAG including SFA and MUFA, Shifrin and Chisholm
523 1981; Sharma et al. 2012). When nutrients are limited, the cell division rate decreases steadily,
524 hence the requirement for membrane compounds reduce or almost reach no requirement any more
525 (Sharma et al. 2012). However, active biosynthesis of FA is maintained (Sharma et al. 2012).
526 Consequently, the cells divert and deposit fatty acids into TAG (Sharma et al. 2012).

527 In our study, irradiance did not have a consistent effect on specific groups of fatty acids.
528 Specifically, irradiance had a positive correlation with the relative abundance of short chain-poly
529 unsaturated fatty acids (SC-PUFA), but no correlation with SFA and highly unsaturated fatty acids
530 (HUFA). We found that the irradiance of 20-140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ induced higher relative
531 content of SC-PUFA (a sub-group of poly unsaturated fatty acids-PUFA) than at 10 $\mu\text{mol photons}$
532 $\text{m}^{-2} \text{s}^{-1}$ whereas the HUFA (also sub-group of PUFA) was highest at 40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and
533 relative constant for all other irradiances. This result did not reflect the general rule reported by
534 Harwood (1998) when high irradiance usually leads to oxidative damage of PUFA (including SC-
535 PUFA and HUFA as defined in our study). The self-shading of the high algae density ($1.1-4.1 \times 10^6$
536 cells mL^{-1}) recorded at high irradiance cultures may reduce the effect of the high irradiance on
537 oxidative damage of the PUFA, indicating the potential to use density manipulation towards
538 designer feed. In detail, irradiance had positive correlation with the relative abundance of DHA but
539 not EPA. This result has also been observed in previous studies (e.g., Renaud et al. 1991; Harrison
540 et al. 1990; Thompson et al. 1990). Harrison et al. (1990) found that Docosahexaenoic acid (DHA)
541 increased as a function of irradiance for all three microalgae whereas Eicosapentaenoic acid (EPA)
542 was relatively constant over a range of irradiance for *Chaetoceros* and *Thalassiosira* but increased
543 significantly for *Isochrysis*. The increase in DHA and decrease in EPA at high irradiance result in

544 an increase of DHA/EPA ratio. In our study, the DHA/EPA of *R. salina* ratio was in the range of
545 0.51-0.70, except for the low irradiance (10-20 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$) where algae did not produce
546 DHA or at least below detection limit of our method. The similar range of DHA/EPA has been
547 reported before in this algal species (Mansour et al. 2005; Dunstan et al. 2005; Drillet et al. 2006).

548 In term of fatty acids composition, it is important to note that the higher TFA level under nutrient
549 deficiency comprised mainly of the higher relative contents of SFA 16:0, MUFA 18:1. On the other
550 hand, under nutrient deficiency, the relative contents of HUFA, including EPA-20:5n-3 and DHA-
551 22:6n-3, the most desired components for calanoid copepods (Arendt et al. 2006; Broglio et al.
552 2003), were lower. This phenomenon is common in many marine microalgae (Reitan et al. 1994).
553 The nutrient limitation probably reduced the synthesis of n-3 PUFA (Reitan et al. 1994). As we can
554 see in our study, the relative abundance of HUFA, especially DHA and EPA of *R. salina*, was
555 considerably higher in nutrient excess than in nutrient deficiency. Depending on the purpose of
556 aquaculture, with different desires of fatty acids, the high or low nutrient medium can be chosen to
557 generate a desired microalgae fatty acid profile.

558 In general the higher the level of relative abundance of PUFA (including SC-PUFA and HUFA)
559 content in the *R. salina*, the better feed for copepods (reviewed in Rasdi and Qin 2014). This is
560 obtained when the *R. salina* algae are cultured at saturated irradiances and in nutrient excess
561 condition. Previous studies showed that the increase in PUFA (including SC-PUFA and HUFA) in
562 algae diet would enhance the egg production and somatic growth of copepods (Rasdi and Qin
563 2014). Therefore, the relatively higher abundance of PUFA of *R. salina* cultured in saturated
564 irradiances and in nutrient excess will benefit the performance of copepods. Please note that the
565 nutrient content in copepods, unlike rotifer and *Artemia*, cannot be manipulated through enrichment
566 techniques due to their avoidance (Huntley et al. 1986; Rasdi and Qin 2014; Rasdi et al. 2015) and
567 nutrient content in copepods can only be enhanced by feeding on high quality algae (Rasdi and Qin
568 2014; Rasdi et al. 2015).

569 *Free amino acids*

570 The irradiance had significant effect on both the quantity and the composition of free amino acids
571 (FAA), whereas nutrient levels only affected the composition of the FAA. In particular, the limited
572 irradiance induced higher total FAA whereas relative abundance of essential amino acid was higher
573 at saturated irradiance. A more favorable essential amino acids (EAA) profile was obtained at

574 higher irradiance with higher contents of most of the specific EAAs, such as arginine, isoleucine,
575 leucine, lysine, methionine, phenylalanine, threonine and valine. This indicates a potential for
576 designer feed at high irradiance. While the changes in total FAA under different irradiances has not
577 been reported before, higher total FAA at lower cultured irradiance was observed in a seaweed
578 *Caulerpa prolifera* (Khaleafa et al. 1982). The effects of irradiance on FAA composition in our
579 study were in contrast with previous studies on different algae species *Isochrysis* sp., *Pavlova*
580 *lutheri* and *Nannochloropsis oculata* where the amino acid composition of the proteins of
581 microalgae has been shown relatively unaffected by the growth phase (normally nutrient is depleted
582 in the stationary phase) and light conditions (Brown et al. 1993a; Brown et al. 1993b; Brown et al.
583 1996). The difference in the effect of irradiance on FAA composition of our study and previous
584 studies could be a result of the species and/or strain specific responses. However, it is noted that the
585 changes in FAA composition of *R. salina* in different cultured irradiance and nutrient condition in
586 our study may not entirely reflect the protein bound AAs.

587

588 **Recommendations**

589 The microalgae *Rhodomonas salina* is a preferred feed item for e.g. copepods used as live feed in
590 hatcheries (Zhang et al. 2013). With our purpose to formulate designer feed for these promising
591 zooplankton live feed organisms, several scenarios are possible when cultivating *R. salina*. i) One
592 can either prioritize high microalga productivity where saturated irradiance and excess inorganic
593 nutrients are implicit. This condition will generate a relatively larger algal biomass with a better
594 fatty acids and free essential amino acids profiles. ii) Another strategy involves saturated irradiance
595 and nutrient deficiency, which generates algal biomass with high total fatty acids content and
596 remain an appropriate DHA/EPA ratio. Moreover, a relatively high content of highly unsaturated
597 fatty acids will occur as a result of nutrient excess, invariant of irradiance. Overall, the most durable
598 and recommended compromise for large scale production in algal photobioreactors for most
599 purposes is to cultivate the microalgae at 60-100 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ irradiance and in nutrient
600 excess.

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609

References

- 610
611
- 612 Aldana-Aranda D, Patiño Suárez V (1998) Overview of diets used in larviculture of three caribbean
613 conchs: Queen conch *Strombus gigas*, Milk conch *Strombus costatus* and Fighting conch
614 *Strombus pugilis*. *Aquaculture* 167 (3–4):163-178.
- 615 Aragão C, Conceição LEC, Dinis MT, Fyhn H-J (2004) Amino acid pools of rotifers and *Artemia*
616 under different conditions: nutritional implications for fish larvae. *Aquaculture* 234 (1–
617 4):429-445.
- 618 Arendt KE, Jónasdóttir SH, Hansen PJ, Gärtner S (2006) Effects of dietary fatty acids on the
619 reproductive success of the calanoid copepod *Temora longicornis*. *Mar Biol* 148 (6):1415-
620 1415.
- 621 Bartual A, Lubián LM, Gálvez JA, Niell FX (2002) Effect of irradiance on growth, photosynthesis,
622 pigment content and nutrient consumption in dense cultures of *Rhodomonas salina*
623 (Wislouch) (Cryptophyceae). *Cienc Mar* 28:381-392.
- 624 Bennett A, Bogorad L (1973) Complementary chromatic adaptation in a filamentous blue-green
625 alga. *J Cell Biol* 2:419-435.
- 626 Bi R, Arndt C, Sommer U (2014) Linking elements to biochemicals: Effects of nutrient supply
627 ratios and growth rates on fatty acid composition of phytoplankton species. *J Phycol* 50
628 (1):117-130.
- 629 Bower CE, Holm-Hansen T (2011) A salicylate–hypochlorite method for determining ammonia in
630 seawater. *Can J Fish Aquat Sci* 37 (5):794-798.
- 631 Breteler WCMK, Schogt N, Rampen S (2005) Effect of diatom nutrient limitation on copepod
632 development: Role of essential lipids. *Mar Ecol Prog Ser* 291:125-133.
- 633 Broglio E, Jónasdóttir SH, Calbet A, Jakobsen HH, Saiz E (2003) Effect of heterotrophic versus
634 autotrophic food on feeding and reproduction of the calanoid copepod *Acartia tonsa*:
635 Relationship with prey fatty acid composition. *Aquat Microb Ecol* 31:267-278.
- 636 Brown MR (1991) The amino-acid and sugar composition of 16 species of microalgae used in
637 mariculture. *J Exp Mar Biol Ecol* 145 (1):79-99.
- 638 Brown MR, Dunstan GA, Jeffrey SW, Volkman JK, Barrett SM, LeRoi J-M (1993a) The influence
639 of irradiance on the biochemical composition of the Prymnesiophyte *Isochrysis* sp. (Clone
640 T-ISO). *J Phycol* 29 (5):601-612.
- 641 Brown MR, Dunstan GA, Norwood SJ, Miller KA (1996) Effects of harvest stage and light on the
642 biochemical composition of the diatom *Thalassiosira pseudonana*. *J Phycol* 32 (1):64-73.

643 Brown MR, Garland CD, Jeffrey SW, Jameson ID, Leroi JM (1993b) The gross and amino acid
644 compositions of batch and semi-continuous cultures of *Isochrysis* sp. (clone T.ISO),
645 *Pavlova lutheri* and *Nannochloropsis oculata*. J Appl Phycol 5 (3):285-296.

646 Brown MR, Jeffrey SW, Volkman JK, Dunstan GA (1997) Nutritional properties of microalgae for
647 mariculture. Aquaculture 151:315-331.

648 Brown MR, McCausland MA, Kowalski K (1998) The nutritional value of four Australian
649 microalgal strains fed to Pacific oyster *Crassostrea gigas* spat. Aquaculture 165 (3-4):281-
650 293.

651 Brown TE, Richardson FL (1968) The effect of growth environment on the physiology of algae:
652 light intensity. J Phycol 4 (1):38-54.

653 Claybrook DL (1983) Nitrogen metabolism. In: Mantel LH (ed) The biology of Crustacea: 5.
654 Internal anatomy and physiological regulation. The biology of Crustacea. Academic Press,
655 New York, pp 162-213.

656 Dhert P, Rombaut G, Suantika G, Sorgeloos P (2001) Advancement of rotifer culture and
657 manipulation techniques in Europe. Aquaculture 200 (1-2):129-146.

658 Diamond D (1999) Determination of nitrate in brackish or seawater by flow injection analysis.
659 QuikChem Method 31-107-04-1-A. Millwaukee, WI 53218-1239 USA.

660 Dongre SK, Manglawat S, Singh P, Yadav M, Tiwari A (2014) Effect of environmental parameters
661 enhancing the micro algal lipid as a sustainable energy source for biodiesel production-A
662 review. Int J Biol Pharm Res 5 (4):327-335.

663 Drillet G, Jørgensen NOG, Sørensen TF, Ramløv H, Hansen BW (2006) Biochemical and technical
664 observations supporting the use of copepods as live feed organisms in marine larviculture.
665 Aquacult Res 37:756-772.

666 Dunstan GA, Brown MR, Volkman JK (2005) Cryptophyceae and rhodophyceae; chemotaxonomy,
667 phylogeny, and application. Phytochemistry 66 (21):2557-2570.

668 Dunstan WM (1973) A comparison of the photosynthesis - light intensity relationship in
669 phylogenetically different marine microalgae. J Exp Mar Biol Ecol 13:181-187.

670 Eriksen NT, Iversen JLL (1995) Photosynthetic pigments as nitrogen stores in the cryptophyte alga
671 *Rhodomonas* sp. J Mar Biotechnol 3:193-195.

672 Evans LV (1988) The effect of spectral composition and irradiance level on pigment levels in
673 seaweed. Experimental phycology: A laboratory manual. Cambridge University Press,
674 Cambridge.

675 Falkowski PG (1975) Nitrate uptake by marine phytoplankton: Comparison of half- saturation
676 constants for seven species. *Limnol Oceanogr* 20:412-417.

677 Faust MA, Gantt E (1973) Effect of light intensity and glycerol on the growth, pigment
678 composition, and ultrastructure of *Chroomonas* sp. *J Phycol* 9 (4):489-495.

679 Folch J, Lees M, Stanley GHS (1957) A simple method for isolation and purification of total lipids
680 from animal tissues. *J Biol Chem* 226:497-509.

681 Fraser AJ, Sargent JR, Gamble JC (1989) Lipid class and fatty acid composition of *Calanus*
682 *finmarchicus* (Gunnerus), *Pseudocalanus* sp. and *Temora longicornis* Muller from a
683 nutrient-enriched seawater enclosure. *J Exp Mar Biol Ecol* 130 (1):81-92.

684 Halldal P (1958) Pigment formation and growth in blue-green algae in crossed gradients of light
685 intensity and temperature. *Physiol Plant* 11:401-420.

686 Hammer A, Schumann R, Schubert H (2002) Light and temperature acclimation of *Rhodomonas*
687 *salina* (Cryptophyceae): Photosynthetic performance. *Aquat Microb Ecol* 29:287-286.

688 Hansen PJ (1989) The red tide dinoflagellate *Alexandrium tamarense*: Effects on behaviour and
689 growth of a tintinnid ciliate. *Mar Ecol Prog Ser* 53:105-116.

690 Harrison PJ, Thompson PA, Calderwood GS (1990) Effects of nutrient and light limitation on the
691 biochemical composition of phytoplankton. *J Appl Phycol* 2 (1):45-56.

692 Harwood J (1998) Involvement of Chloroplast Lipids in the Reaction of Plants Submitted to Stress.
693 In: Paul-André S, Norio M (eds) *Lipids in Photosynthesis: Structure, Function and Genetics*,
694 vol 6. *Advances in Photosynthesis and Respiration*. Springer Netherlands, pp 287-302.

695 Hu Q (2004) Environmental effects on cell composition. In: Richmond A (ed) *Handbook of*
696 *microalgal culture: Biotechnology and applied phycology*, vol XVIII, 566 s. Blackwell
697 Science, Oxford, pp 83-93.

698 Huntley M, Sykes P, Rohan S, Marin V (1986) Chemically-mediated rejection of dinoflagellate
699 prey by the copepods *Calanus pacificus* and *Paracalanus parvus*: mechanism, occurrence
700 and significance. *Mar Ecol Prog Ser* 28:105-120.

701 Jassby AD, Platt T (1976) Mathematical formulation of the relationship between photosynthesis and
702 light for phytoplankton. *Limnol Oceanogr* 21 (4):540-547.

703 Jeffrey SW, Humphrey GF (1975) New spectrophotometric equations for determining chlorophylls
704 a1, b1, c1 and c2 in higher plants, algae and natural phytoplankton. *Biochem Physiol*
705 *Pflanzen* 167:191-194.

- 706 Jónasdóttir SH (1994) Effects of food quality on the reproductive success of *Acartia tonsa* and
707 *Acartia hudsonica*: laboratory observations. *Mar Biol* 121 (1):67-81.
- 708 Jónasdóttir SH, Kiørboe T (1996) Copepod recruitment and food composition: do diatoms affect
709 hatching success? *Mar Biol* 125:743-750.
- 710 Juneja A, Ceballos RM, Murthy GS (2013) Effects of environmental factors and nutrient
711 availability on the biochemical composition of algae for biofuels production: A review.
712 *Energies* 6:4607-4638.
- 713 Khaleafa AF, Mohsen AF, Shaalan SH (1982) Effect of different light intensities on growth, amino-
714 acid, fat and sugar concentrations in *Caulerpa prolifera* (Foerskal) Lamouroux. *Hydrobiol*
715 *Bull* 16 (2-3):207-212.
- 716 Kromkamp JC, Peen J (2001) Oxygen consumption in the light by unicellular algae. In: PS2001
717 Proceedings 12th International Congress on Photosynthesis. CSIRO Publishing, Brisbane,
718 Australia, pp 1-6.
- 719 Lafarga-De la Cruz F, Valenzuela-Espinoza E, Millán-Núñez R, Trees CC, Santamaría-del-Ángel
720 E, Núñez-Cebrero F (2006) Nutrient uptake, chlorophyll a and carbon fixation by
721 *Rhodomonas sp.* (Cryptophyceae) cultured at different irradiance and nutrient
722 concentrations. *Aquacult Eng* 35:51-60.
- 723 Lewitus AJ, Caron DA (1990) Relative effects of nitrogen or phosphorus depletion and light
724 intensity on the pigmentation, chemical composition, and volume of *Pyrenomonas salina*
725 (Cryptophyceae) *Mar Ecol Prog Ser* 61:171-181.
- 726 Lichtlé C (1979) Effects of nitrogen deficiency and light of high intensity on *Cryptomonas*
727 *rufescens* (Cryptophyceae). *Protoplasma* 101 (4):283-299.
- 728 Malzahn AM, Boersma M (2012) Effects of poor food quality on copepod growth are dose
729 dependent and non-reversible. *Oikos* 121 (9):1408-1416.
- 730 Mansour M, Frampton DF, Nichols P, Volkman J, Blackburn S (2005) Lipid and fatty acid yield of
731 nine stationary-phase microalgae: Applications and unusual C24–C28 polyunsaturated fatty
732 acids. *J Appl Phycol* 17 (4):287-300.
- 733 Marinho da Costa Rr, Fernández F (2002) Feeding and survival rates of the copepods *Euterpina*
734 *acutifrons* Dana and *Acartia grani* Sars on the dinoflagellates *Alexandrium minutum* Balech
735 and *Gyrodinium corsicum* Paulmier and the Chryptophyta *Rhodomonas baltica* Karsten. *J*
736 *Exp Mar Biol Ecol* 273 (2):131-142.

737 Mortillaro JM, Pitt KA, Lee SY, Meziane T (2009) Light intensity influences the production and
738 translocation of fatty acids by zooxanthellae in the jellyfish *Cassiopea* sp. *J Exp Mar Biol*
739 *Ecol* 378 (1–2):22-30.

740 Muller-Feuga A, Moal J, Kaas R (2003) The microalgae of aquaculture. In: Støttrup JG, McEvoy
741 LA (eds) *Live feeds in Marine Aquaculture*. Blackwell Publishing, Oxford, pp 206-299.

742 Neidhardt J, Benemann J, Zhang L, Melis A (1998) Photosystem-II repair and chloroplast recovery
743 from irradiance stress: Relationship between chronic photoinhibition, light-harvesting
744 chlorophyll antenna size and photosynthetic productivity in *Dunaliella salina* (green algae).
745 *Photosynthesis Res* 56 (2):175-184.

746 Parrish CC (2009) Essential fatty acids in aquatic food webs. In: Arts MT, Brett MT, Kainz MJ
747 (eds) *Lipids in Aquatic Ecosystems*. Springer, New York, pp 309-326.

748 Payne MF, Rippingale RJ (2000) Evaluation of diets for culture of the calanoid copepod
749 *Gladioferens imparipes*. *Aquaculture* 187 (1-2):85-96.

750 Pereira DC, Trigueiro TG, Colepicolo P, Marinho-Soriano E (2012) Seasonal changes in the
751 pigment composition of natural population of *Gracilaria domingensis* (Gracilariales,
752 Rhodophyta). *Rev Bras Farmacogn* 22:874-880.

753 Piorreck M, Baasch K-H, Pohl P (1984) Biomass production, total protein, chlorophylls, lipids and
754 fatty acids of freshwater green and blue-green algae under different nitrogen regimes.
755 *Phytochemistry* 23 (2):207-216.

756 Platt T, Gallegos C, Harrison W (1980) Photoinhibition of photosynthesis in natural assemblages of
757 marine phytoplankton. *J Mar Res* 38:687-701.

758 Rasdi NW, Qin JG (2014) Improvement of copepod nutritional quality as live food for aquaculture:
759 a review. *Aquacult Res* n/a-n/a: DOI: 10.1111/are.12471.

760 Rasdi NW, Qin JG, Li Y (2015) Effects of dietary microalgae on fatty acids and digestive enzymes
761 in copepod *Cyclopina kasignete*, a potential live food for fish larvae. *Aquacult Res* n/a-
762 n/a:DOI: 10.1111/are.12778.

763 Reitan KI, Rainuzzo JR, Olsen Y (1994) Effect of nutrient limitation on fatty acid and lipid content
764 of marine microalgae. *J Phycol* 30 (6):972-979.

765 Renaud SM, Parry DL, Thinh L-V, Kuo C, Padovan A, Sammy N (1991) Effect of light intensity on
766 the proximate biochemical and fatty acid composition of *Isochrysis* sp. and *Nannochloropsis*
767 *oculata* for use in tropical aquaculture. *J Appl Phycol* 3 (1):43-53.

768 Rhiel E, Mörschel E, Wehrmeyer W (1985) Correlation of pigment deprivation and ultrastructural
769 organization of thylakoid membranes in *Cryptomonas maculata* following nutrient
770 deficiency. *Protoplasma* 129 (1):62-73.

771 Ritchie R (2006) Consistent sets of spectrophotometric chlorophyll equations for acetone, methanol
772 and ethanol solvents. *Photosynthesis Res* 89 (1):27-41.

773 Rückert GV, Giani A (2004) Effect of nitrate and ammonium on the growth and protein
774 concentration of *Microcystis viridis* Lemmermann (Cyanobacteria). *Rev Bras Bot* 27:325-
775 331.

776 Sargent JR, McEvoy LA, Bell JG (1997) Requirements, presentation and sources of
777 polyunsaturated fatty acids in marine fish larval feeds. *Aquaculture* 155 (1-4):117-127.

778 Sharma KK, Schuhmann H, Schenk PM (2012) High lipid induction in microalgae for biodiesel
779 production. *Energies* 5 (5):1996-1073.

780 Shifrin NS, Chisholm SW (1981) Phytoplankton lipids: Interspecific differences and effects of
781 nitrate, silicate and light-dark cycles. *J Phycol* 17 (4):374-384.

782 Smith REH, Kalff J (1982) Size-dependent phosphorus uptake kinetics and cell quota in
783 phytoplankton. *J Phycol* 18 (2):275-284.

784 Søndergaard M, Riemann B (1979) *Ferskvandsbiologiske analysemetoder*. Akademisk Forlag,
785 Copenhagen.

786 Sorgeloos P, Dhert P, Candreva P (2001) Use of brine shrimp, *Artemia* spp., in larval crustacean
787 nutrition: a review. *Aquaculture* 200:147-159.

788 Spector A (1999) Essentiality of fatty acids. *Lipids* 34 (1):S1-S3.

789 Sriharan S, Bagga D, Nawaz M (1991) The effects of nutrients and temperature on biomass,
790 growth, lipid production, and fatty acid composition of *Cyclotella cryptica* Reimann, Lewin,
791 and Guillard. *Appl Biochem Biotechnol* 28-29 (1):317-326.

792 Støttrup JG (2003) Production and nutritional value of copepods. In: Støttrup JG, McEvoy LA (eds)
793 *Live feeds in Marine Aquaculture*. Blackwell Publishing, Oxford, pp 145-205.

794 Støttrup JG, Jensen J (1990) Influence of algal diet on feeding and egg-production of the calanoid
795 copepod *Acartia tonsa* Dana. *J Exp Mar Biol Ecol* 141 (2-3):87-105.

796 Thompson PA, Harrison PJ, Whyte JNC (1990) Influence of irradiance on the fatty acid
797 composition of phytoplankton. *J Phycol* 26 (2):278-288.

798 Wilson RP (1985) Amino acid and protein requirements of fish. In: Cowey CB, Mackie AM, Bell
799 JG (eds) *Nutrition and Feeding in Fish*. Academic Press, London, pp 1-16.

800 Wu G (2009) Amino acids: metabolism, functions, and nutrition. *Amino Acids* 37 (1):1-17.

801 Yaakob Z, Ali E, Zainal A, Mohamad M, Takriff MS (2014) An overview: Biomolecules from
802 microalgae for animal feed and aquaculture. *J Biol Res-Thessalon* 21:6 (6).

803 Yu J, Li G, Krull I, Cohen S (1994) Polymeric 6-aminoquinone, an activated carbamate reagent for
804 derivatization of amines and amino acids by high performance liquid chromatography. *J*
805 *Chromatogr B Biomed Appl* 658 (2):249-260.

806 Zhang J, Wu C, Pellegrini D, Romano G, Esposito F, Ianora A, Buttino I (2013) Effects of different
807 monoalgal diets on egg production, hatching success and apoptosis induction in a
808 Mediterranean population of the calanoid copepod *Acartia tonsa* (Dana). *Aquaculture* 400-
809 401 (0):65-72.

810 Zimba PV (2012) An improved phycobilin extraction method. *Harmful Algae* 17:35-39.

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814 **Caption for figures**

815 **Fig. 1** Net photosynthesis per cell of *R. salina* ($\times 10^{-15}$ mol O₂ cell⁻¹ h⁻¹) incubated at different cell
816 densities and irradiance levels. Note: the number of experimental replicates, n = 5 for irradiances
817 from 0-140 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ and n = 2-3 for irradiances from 160-300 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$

818 **Fig. 2** The specific growth rate (SGR) (a), maximum cell density (b) and the plot between SGR and
819 cell biovolume (c) of *Rhodomonas salina* in response to different irradiance and nutrient levels.
820 Data are indicated by means (\pm SDs) of specific growth rate of algae cultured in nutrient deficiency
821 and nutrient excess at the same levels of irradiance. Note: the number of experimental replicates, n
822 = 4 (2 replicates from nutrient deficiency and 2 replicates from nutrient excess treatments)

823 **Fig. 3** Nitrate, ammonium and phosphate concentrations in the culture media of *Rhodomonas salina*
824 with different irradiance and nutrient levels. Data are means (\pm SDs) of nutrient at day 4 and day 6,
825 the number of analytical replicates, n = 2

826 **Fig. 4** The chlorophyll *a* (a), chlorophyll *c* (b), phycoerythrin (PE) (c) and the
827 phycoerythrin/chlorophyll *a* (PE/chl *a*) ratio (d) and the relationship between PE and cell density of
828 *Rhodomonas salina* cultured in nutrient deficiency (e) and nutrient excess (f) under different
829 irradiance levels. The number of analytical replicates, n = 2 for all of presented parameters. In
830 figure 4 e and f, the solid lines are the regression lines between PE and cell density, the dashed lines
831 are the 95% confident interval of these regression lines

832 **Fig. 5** Total fatty acid (TFA) (a), the FA composition in nutrient deficiency (b) and nutrient excess
833 (c), and the DHA/EPA ratio (d) of *Rhodomonas salina* under different irradiance levels. SFA:
834 saturated fatty acids; MUFA: mono unsaturated fatty acids; HUFA: highly unsaturated fatty acids,
835 SC-PUFA: short chain-poly unsaturated fatty acids. The number of analytical replicates, n = 3 for
836 nutrient deficiency and n = 2 for nutrient excess treatment

837

838 **Caption for tables**

839 **Table 1** Summary of photosynthetic-irradiance parameters for five algae densities of *Rhodomonas*
840 *salina* exposed to 16 increasing irradiances from 0 to 300 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ PAR, with the
841 number of experimental replicates, $n = 5$ for irradiances from 0-140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ and $n = 2$ -
842 3 for irradiances from 160-300 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$

843 Note: α = Initial slope of the photosynthesis-irradiance curve; β = Negative slope at high irradiance;
844 I_m = irradiance of maximum photosynthesis, R = Dark respiration = mean \pm SDs (plus/minus
845 standard deviation) of dark respiration at 0 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ of 5 experimental replicates at a
846 specific cell density. Units: $\alpha, \beta = 10^{-15} \text{ mol O}_2 \text{ cell}^{-1} \text{ h}^{-1} [\mu\text{mol photons m}^{-2} \text{s}^{-1}]^{-1}$; $P_m^B, R = 10^{-15} \text{ mol}$
847 $\text{O}_2 \text{ cell}^{-1} \text{ h}^{-1}$ and $I_m = \mu\text{mol photons m}^{-2} \text{s}^{-1}$

848 **Table 2** Total free amino acids (FAA) and essential amino acids (EAA) in *Rhodomonas salina*
849 cultured in different irradiance and nutrient levels

850 Note: Units of total FAA = pg cell^{-1} ; sub-total EAA, specific EAA = % of total FAA. Values for
851 limited irradiance = mean \pm SDs (plus/minus standard deviation) of FAA/EAA at irradiance from
852 10-40 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$; values for saturated irradiance = mean \pm SDs of FAA/EAA at
853 irradiance from 60-140 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$; n is the number of experimental replicates. Different
854 letters in the same row denote the significant differences in the same specific EAA between the
855 different treatments